## **Supporting Information for**

# Targeted fluorescence imaging enhanced by 2D materials: A

### comparison between 2D MoS<sub>2</sub> and graphene oxide

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### **Additional figures**



**Figure S1.** Dynamic light scattering (DLS) of graphene oxide (50  $\mu$ g mL<sup>-1</sup>) and 2D MoS<sub>2</sub> (50  $\mu$ g mL<sup>-1</sup>) in Tris-HCl (0.01 M, pH 7.4) carried out on a Horiba LB-550 DLS Nano-Analyzer.



**Figure S2.** Fluorescence spectroscopy of graphene oxide (50  $\mu$ g mL<sup>-1</sup>) and 2D MoS<sub>2</sub> (50  $\mu$ g mL<sup>-1</sup>) in Tris-HCl (0.01 M, pH 7.4) carried out on a Varian Cary Eclipsefluorescence spectrophotometer with excitation of 400 nm (use of other excitation wavelengths including 325, 350 and 460 nm did not produce any emission of the two materials; data not shown).

### **Experimental section**

Preparation of 2D thin-layer  $MoS_2$ . Layered  $MoS_2$  was obtained through sonicationassisted exfoliation of bulk  $MoS_2$  crystals in the mixed aqueous solution of EtOH and water.  $MoS_2$  (100 mg, Aladdin Reagent Inc.) was added to a 25 mL vial filled with 20 mL of EtOH/water (1:1, v/v). The sealed vial was sonicated for 8 h, and then the dispersion was centrifuged at 3200 rpm for 10 mins to remove aggregates. After collecting the supernatant and drying at 60 °C in a drying oven, an army green powder was obtained. The obtained podwer was dissolved in Milli-Q ultrapure water and sonicated for 0.5 h to provide a homogeneous stock solution.

*Scanning electron microscope (SEM)*. A droplet of 2D MoS<sub>2</sub> (0.1 mg mL<sup>-1</sup>) or GO (0.1 mg mL<sup>-1</sup>) was cast onto a freshly cleaved mica surface, followed by drying at room temperature. Then, SEM of the materials was carried out with S-3400N (HITACHI, Japan).

*High-Resolution Transmission electron microscope (HRTEM).* A droplet of MoS<sub>2</sub> (0.1 mg mL<sup>-1</sup>) or GO (0.1 mg mL<sup>-1</sup>) was cast onto 200 mesh holey carbon copper grids for HRTEM characterizations. JEOL 2100 equipped with a Gatan Orius charged-coupled device camera and Tridiem energy filter operating at 200 kV was used for the images. The obtained data were processed using Image J software.

*Raman spectroscopy.* Raman spectra were obtained using a Renishaw InVia Reflex Raman system (Renishaw plc, Wotton-under-Edge, UK) employing a grating spectrometer with a Peltier-cooled charge-coupled device (CCD) detector coupled to a confocal microscope. The data obtained were processed with Renishaw WiRE 3.2 software. The Raman scattering was excited by an argon ion laser (I = 514.5 nm).

*Real-time quantitative PCR.* Total RNA was isolated from cells and tissues using TRIzol Reagent (Invitrogen) according to the manufacturer's protocol. Complementary DNA generated using a PrimeScript® RT reagent kit (TaKaRa, Dalian, China) was analyzed by quantitative PCR using SYBR® Premix Ex TaqTM. Real-time PCR was performed using a 7300 Real-Time PCR system (Applied Biosystems, CA, USA). GAPDH was detected as the housekeeping gene. Primers for qPCR were as follows:

GAPDH forward, 5'-ATCACTGCCACCCAGAAGAC-3' and reverse, 5'-ATGAGGTCCACCACCCTGTT-3' ASGPR1 forward, 5'-CTGGACAATGAGGAGAGTGAC-3' and reverse, 5'-TTGAAGCCCGTCTCGTAGTC-3' *Fluorescence imaging of cells*. Cells  $(15 \times 10^4)$  were seeded on a black 96-well microplate with optically clear bottom (Greiner bio-one, Germany) overnight. Then the cells were incubated with **DK1** in the absence and presence of a material (GO or 2D MoS<sub>2</sub>) for 15 min. Then, cells were gently washed with PBS three times, fixed using 4% paraformaldehyde and stained with Hoechst (5 µg mL<sup>-1</sup>). The fluorescence images were recorded using an Operetta high content imaging system and quantified by the Columbus image data analysis system (Perkinelmer, US).

*Fluorescence imaging of tissues.* The xenograft, spleen and lung sections removed from Hep-G2-bearing xenograft mice (see: *J. Mater. Chem. B*, 2015, **3**, 9182) were fixed in 4% paraformaldehyde in PBS overnight. Sections (12  $\mu$ m) were cut using a rotary microtome, and then treated with PBS for 10 min. Then, the sections were incubated with **DK1** (10  $\mu$ M) with or without 2D MoS<sub>2</sub> and GO (100, 250 or 500  $\mu$ g mL<sup>-1</sup>) for 15 min, and were gently washed with PBS three times and mounted by glycerin. The fluorescence images were recorded using a fluorescence microscope (Olympus, Japan).