

Supporting Information

Table of Contents

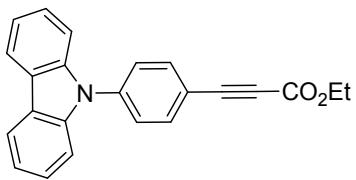
General Procedures	S2
Experimental Section	S3
Mechanistic Study	S19
References	S25
NMR Spectra	S27

General Procedures.

All reactions were carried out in oven-dried glassware under an atmosphere of dry N₂ with the rigid exclusion of air and moisture using standard Schlenk techniques or in a glovebox. All organic solvents were freshly distilled from sodium benzophenone ketyl immediately prior to use. Compounds **5**,¹ **6**,² ethyl 4-tolylpropiolate,³ ethyl 3-tolylpropiolate,⁴ ethyl 4-*tert*-butylphenyl propiolate,⁵ ethyl (4-methoxyphenyl)propiolate,³ ethyl 4-dimethyl aminophenylpropiolate,⁶ ethyl 4-fluorophenylpropiolate,⁴ ethyl 4-chlorophenylpropiolate,⁴ ethyl 4-bromophenylpropiolate,⁷ ethyl 3-([1,1'-biphenyl]-4-yl)propiolate,⁸ ethyl 3-(naphthalen-2-yl)propiolate,⁸ ethyl 3-(6-methoxy-2-naphthalenyl)propiolate,⁹ ethyl 2-furylpropiolate,¹⁰ ethyl 3-furylpropiolate,¹¹ ethyl 2-thienylpropiolate,¹² ethyl 3-thienylpropiolate,³ and ethyl 2-pyridinylpropiolate,³ were prepared according to literature procedures. All other chemicals were purchased from either Aldrich or Acros Chemical Co. and used as received unless otherwise specified. ¹H, ¹³C, and ¹¹B NMR spectra were recorded on a Bruker DPX 400 spectrometer at 400 MHz, 100 MHz and 128 MHz, respectively. All signals were reported in δ unit with references to the residual solvent resonances of the deuterated solvents for proton and carbon chemical shifts, to external BF₃·OEt₂ (0.00) for boron chemical shifts. Mass spectra were collected on an Agilent GC/MS 5975C system, or a MALDI Micro MX mass spectrometer, or an API QSTAR XL System.

Experimental Section.

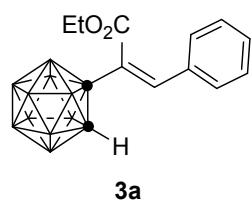
Preparation of ethyl 3-[4-(*N*-carbazolyl)phenyl]propiolate (2t**).** To a diethyl ether solution (20 mL) of *N*-(4-ethynylphenyl)carbazole (500 mg, 1.87 mmol) was added *n*-BuLi (0.75 mL, 2.5M in *n*-hexane, 1.87 mmol) at -78 °C and stirred for 0.5 h. Ethyl chloroformate (0.27 mL, 2.84 mmol) was then added at -78 °C. The reaction mixture was allowed to warm to room temperature and stirred overnight. After water (20 mL) was added to reaction mixture, the aqueous layer was extracted with ether (15 × 3 mL). The combined organic layer was then dried over anhydrous Na₂SO₄. After filtration and removal of solvent in vacuo, the residue was subjected to flash column chromatography on silica gel (230–400 mesh) using *n*-hexane/diethyl ether (8/1 in v/v) as eluent to give **2t** (420 mg, 66%, 1.24 mmol).



2t: light yellow crystals. ¹H NMR (C₆D₆, 400 MHz) δ 8.00 (m, 2H) (aromatic H), 7.24 (m, 6H) (aromatic H), 7.15 (m, 2H) (aromatic H), 6.85 (m, 2H) (aromatic H), 4.00 (q, *J* = 7.1 Hz, 2H) (OCH₂CH₃), 0.95 (t, *J* = 7.1 Hz, 3H) (OCH₂CH₃). ¹³C{¹H} NMR (C₆D₆, 100 MHz): δ 153.93 (COOEt), 140.57, 139.85, 134.65, 126.81, 126.46, 124.26, 120.94, 120.74, 118.32, 110.06 (aromatic C), 85.20, 82.41 (acytylenic C), 62.01 (OCH₂CH₃), 13.99 (OCH₂CH₃). HRMS: *m/z* calcd for C₂₃H₁₇NO₂ [M]⁺: 339.1259. Found: 339.1259.

Preparation of 1-Alkenyl-*o*-Carboranes (3**). A Representative Procedure.** *o*-Carborane (**1**, 29 mg, 0.20 mmol), alkyne (**2**, 0.40 mmol) and tris(4-methoxy

phenyl)phosphine (7 mg, 0.02 mmol) were mixed in dioxane (0.6 mL). The resulting mixture was heated at 110 °C for 72 h. Then the reaction solution was concentrated to dryness in vacuo. The residue was subjected to flash column chromatography on silica gel (230–400 mesh) using *n*-hexane/dichloromethane (20/1 in v/v) as eluent to give 1-alkenyl-*o*-carboranes (**3**).



3a: Colorless crystals. Yield: 82%. ^1H NMR (CDCl_3 , 400 MHz): δ 7.34 (m, 3H) (phenyl *H*), 7.23 (m, 2H) (phenyl *H*), 7.19 (s, 1H) (olefinic *H*), 4.23 (s, 1H) (cage *H*), 4.14 (q, *J* = 6.4 Hz, 2H) (OCH_2CH_3), 1.11 (t, *J* = 6.8 Hz, 3H) (OCH_2CH_3). $^{13}\text{C}\{\text{H}\}$ NMR (CDCl_3 , 100 MHz): δ 166.5 (COOEt), 139.1, 133.7, 129.7, 128.8, 128.3, 127.7 (phenyl & olefinic *C*), 72.9 (cage C-C), 62.5 (OCH_2CH_3), 59.4 (cage C-H), 13.7 (OCH_2CH_3). $^{11}\text{B}\{\text{H}\}$ NMR (CDCl_3 , 128 MHz): δ -0.8 (1B), -2.4 (1B), -7.6 (2B), -9.2 (2B), -10.0 (2B), -11.4 (2B). HRMS: *m/z* calcd for $\text{C}_{13}\text{H}_{21}^{11}\text{B}_8^{10}\text{B}_2\text{O}_2$ [M – H] $^+$: 317.2545. Found: 317.2552.

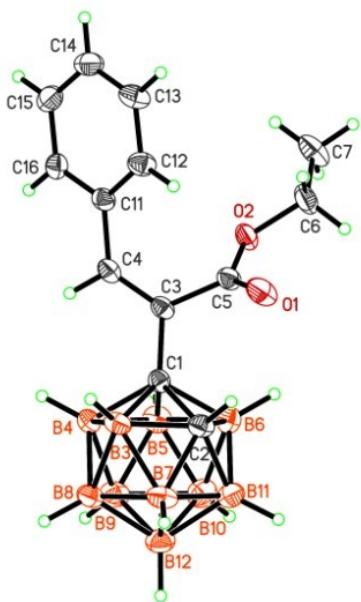


Figure S1. Molecular structure of **3a**

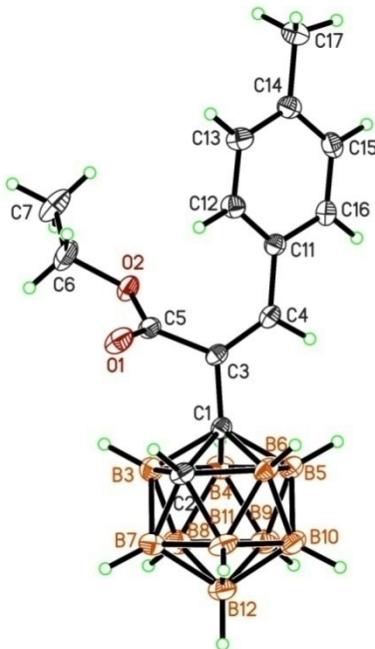
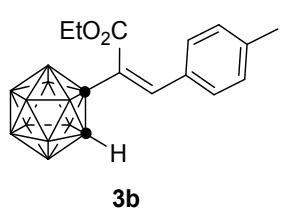
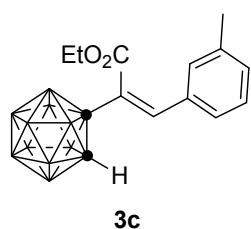


Figure S2. Molecular structure of **3b**

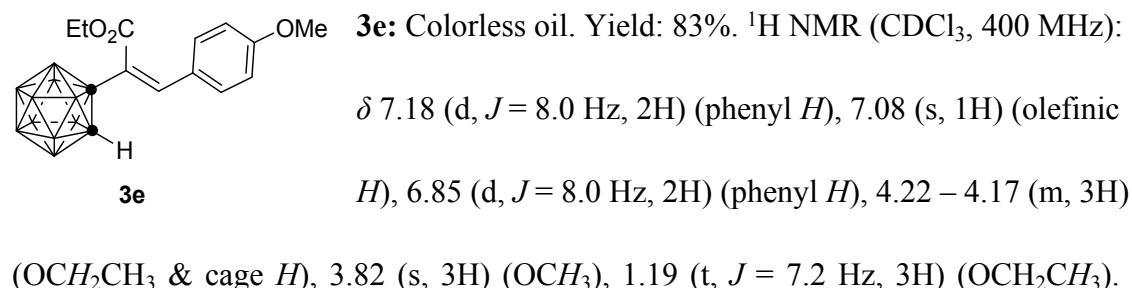
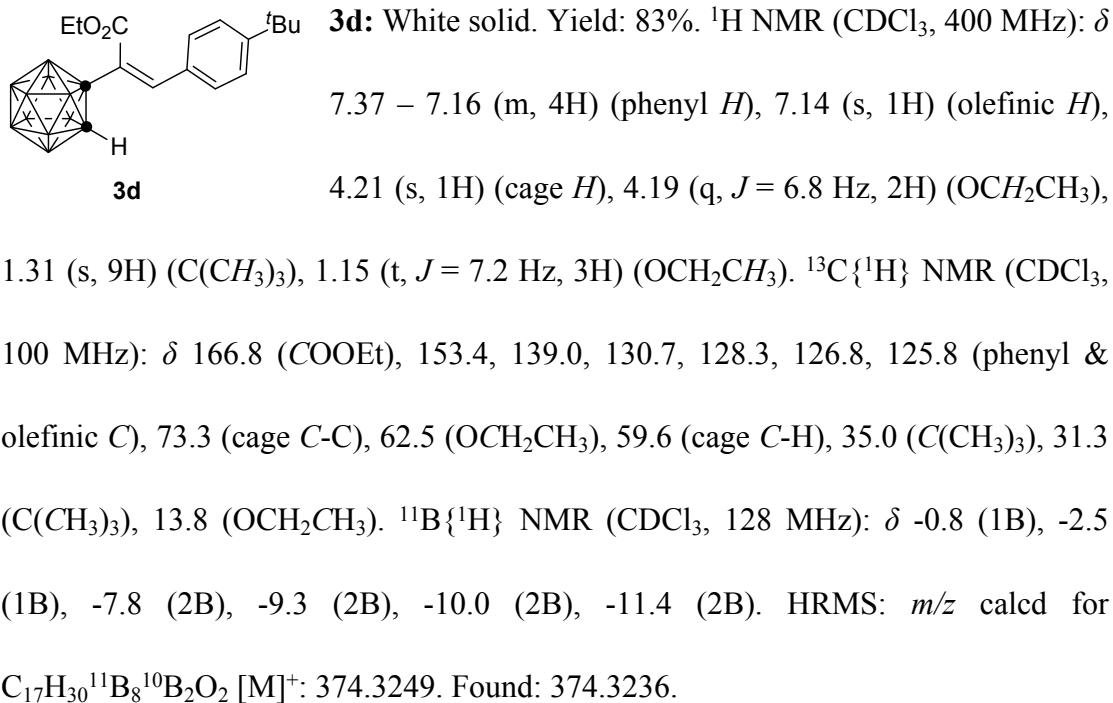


3b: Colorless crystals. Yield: 85%. ^1H NMR (CDCl_3 , 400 MHz): δ 7.14 (m, 5H) (phenyl & olefinic H), 4.21 (s, 1H) (cage H), 4.18 (q, $J = 7.2$ Hz, 2H) (OCH_2CH_3), 2.34 (s, 3H) ($\text{C}_6\text{H}_4\text{CH}_3$), 1.16 (t, $J = 6.8$ Hz, 3H) (OCH_2CH_3). $^{13}\text{C}\{\text{H}\}$ NMR (CDCl_3 , 100 MHz): δ 166.8 (COOEt), 140.2, 139.1, 130.7, 129.5, 128.4, 126.6 (phenyl & olefinic C), 73.3 (cage C-C), 62.5 (OCH_2CH_3), 59.6 (cage C-H), 21.6 ($\text{C}_6\text{H}_4\text{CH}_3$), 13.8 (OCH_2CH_3). $^{11}\text{B}\{\text{H}\}$ NMR (CDCl_3 , 128 MHz): δ -0.8 (1B), -2.5 (1B), -7.8 (2B), -9.3 (2B), -10.0 (2B), -11.4 (2B). HRMS: m/z calcd for $\text{C}_{14}\text{H}_{23}^{11}\text{B}_8^{10}\text{B}_2\text{O}_2$ [M - H] $^+$: 331.2701. Found: 331.2695.



3c: White solid. Yield: 89%. ^1H NMR (CDCl_3 , 400 MHz): δ S5

7.25 – 7.03 (m, 5H) (phenyl & olefinic *H*), 4.24 (s, 1H) (cage *H*), 4.16 (q, *J* = 7.2 Hz, 2H) (OCH₂CH₃), 2.34 (s, 3H) (C₆H₄CH₃), 1.13 (t, *J* = 7.2 Hz, 3H) (OCH₂CH₃). ¹³C{¹H} NMR (CDCl₃, 100 MHz): δ 166.7 (COOEt), 139.3, 138.5, 133.6, 130.6, 129.1, 128.7, 127.4, 125.4 (phenyl & olefinic *C*), 73.1 (cage C-C), 62.5 (OCH₂CH₃), 59.5 (cage C-H), 21.5 (C₆H₄CH₃), 13.7 (OCH₂CH₃). ¹¹B{¹H} NMR (CDCl₃, 128 MHz): δ -0.8 (1B), -2.4 (1B), -7.7 (2B), -9.3 (2B), -10.1 (2B), -11.4 (2B). HRMS: *m/z* calcd for C₁₄H₂₃¹¹B₈¹⁰B₂O₂ [M – H]⁺: 331.2701. Found: 331.2703.



125.3, 114.3 (phenyl & olefinic C), 73.5 (cage C-C), 62.5 (OCH_2CH_3), 59.7 (cage C-H), 55.6 (OCH_3), 13.9 (OCH_2CH_3). $^{11}\text{B}\{\text{H}\}$ NMR (CDCl_3 , 128 MHz): δ -0.8 (1B), -2.6 (1B), -7.8 (2B), -9.3 (2B), -10.0 (2B), -11.4 (2B). HRMS: m/z calcd for $\text{C}_{14}\text{H}_{23}^{11}\text{B}_8^{10}\text{B}_2\text{O}_3 [\text{M} - \text{H}]^+$: 347.2650. Found: 347.2648.

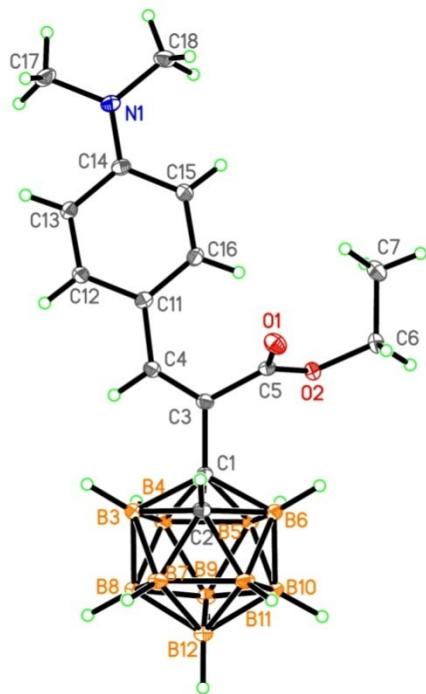
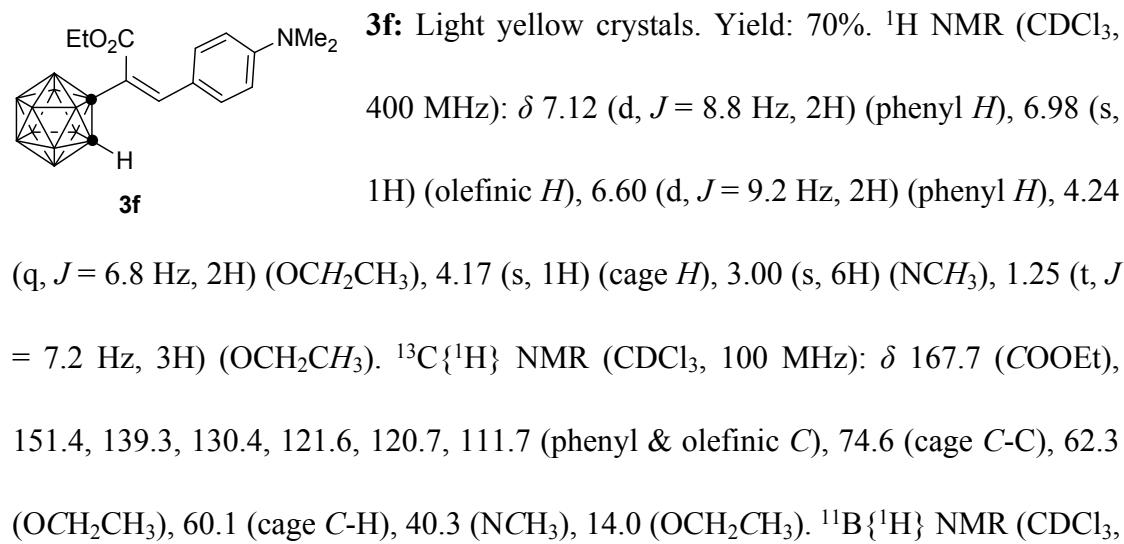
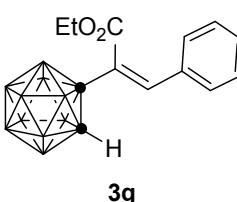


Figure S3. Molecular structure of **3f**



128 MHz): δ -0.8 (1B), -2.9 (1B), -8.0 (2B), -9.2 (2B), -9.9 (2B), -11.5 (2B). HRMS: m/z calcd for $C_{15}H_{27}^{11}B_8^{10}B_2NO_2 [M]^+$: 361.3045. Found: 361.3040.


3g: White solid. Yield: 32%. 1H NMR ($CDCl_3$, 400 MHz): δ 7.24 – 7.21 (m, 2H) (phenyl H), 7.13 (s, 1H) (olefinic H), 7.06 – 7.01 (m, 2H) (phenyl H), 4.21 (s, 1H) (cage H), 4.15 (q, J = 7.2 Hz, 2H) (OCH_2CH_3), 1.14 (t, J = 7.2 Hz, 3H) (OCH_2CH_3). $^{13}C\{^1H\}$ NMR ($CDCl_3$, 100 MHz): δ 166.5 (COOEt), 164.7, 162.2, 137.9, 130.5, 130.4, 129.9, 129.8, 127.8, 116.1, 115.9 (phenyl & olefinic C), 72.8 (cage C-C), 62.7 (OCH_2CH_3), 59.5 (cage C-H), 13.8 (OCH_2CH_3). $^{11}B\{^1H\}$ NMR ($CDCl_3$, 128 MHz): δ -0.8 (1B), -2.4 (1B), -7.7 (2B), -9.3 (2B), -10.1 (2B), -11.4 (2B). HRMS: m/z calcd for $C_{13}H_{20}^{11}B_8^{10}B_2FO_2 [M - H]^+$: 335.2451. Found: 335.2435.

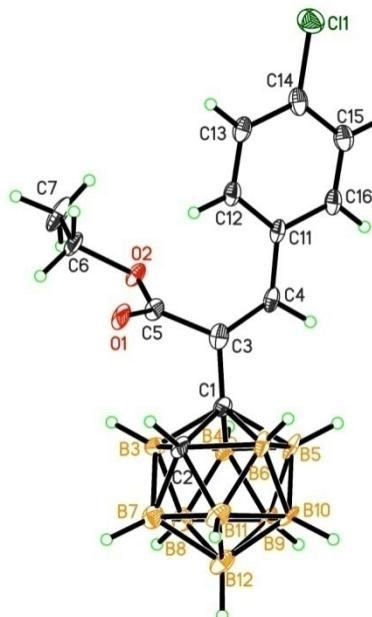
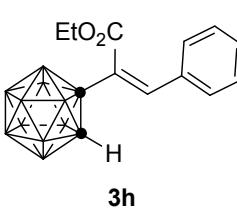


Figure S4. Molecular structure of **3h**


3h: Colorless crystals. Yield: 40%. 1H NMR ($CDCl_3$, 400 MHz): δ 7.24 – 7.21 (m, 2H) (phenyl H), 7.13 (s, 1H) (olefinic H), 7.06 – 7.01 (m, 2H) (phenyl H), 4.21 (s, 1H) (cage H), 4.15 (q, J = 7.2 Hz, 2H) (OCH_2CH_3), 1.14 (t, J = 7.2 Hz, 3H) (OCH_2CH_3). $^{13}C\{^1H\}$ NMR ($CDCl_3$, 100 MHz): δ 166.5 (COOEt), 164.7, 162.2, 137.9, 130.5, 130.4, 129.9, 129.8, 127.8, 116.1, 115.9 (phenyl & olefinic C), 72.8 (cage C-C), 62.7 (OCH_2CH_3), 59.5 (cage C-H), 13.8 (OCH_2CH_3). $^{11}B\{^1H\}$ NMR ($CDCl_3$, 128 MHz): δ -0.8 (1B), -2.4 (1B), -7.7 (2B), -9.3 (2B), -10.1 (2B), -11.4 (2B). HRMS: m/z calcd for $C_{13}H_{20}^{11}B_8^{10}B_2FO_2 [M - H]^+$: 335.2451. Found: 335.2435.

MHz): δ 7.34 – 7.15 (m, 4H) (phenyl *H*), 7.12 (s, 1H) (olefinic *H*), 4.21 (s, 1H) (cage *H*), 4.15 (q, J = 7.2 Hz, 2H) (OCH_2CH_3), 1.14 (t, J = 7.2 Hz, 3H) (OCH_2CH_3). $^{13}C\{^1H\}$ NMR ($CDCl_3$, 100 MHz): δ 166.3 (COOEt), 137.7, 135.8, 132.2, 129.7, 129.1, 128.5 (phenyl & olefinic *C*), 72.7 (cage *C-C*), 62.7 (OCH_2CH_3), 59.5 (cage *C-H*), 13.8 (OCH_2CH_3). $^{11}B\{^1H\}$ NMR ($CDCl_3$, 128 MHz): δ -0.8 (1B), -2.3 (1B), -7.7 (2B), -9.3 (2B), -10.1 (2B), -11.4 (2B). HRMS: *m/z* calcd for $C_{13}H_{20}^{11}B_9^{10}BClO_2$ [M – H] $^+$: 352.2119. Found: 352.2128.

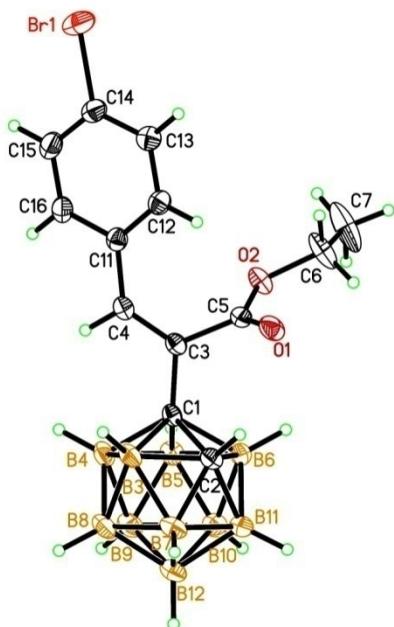
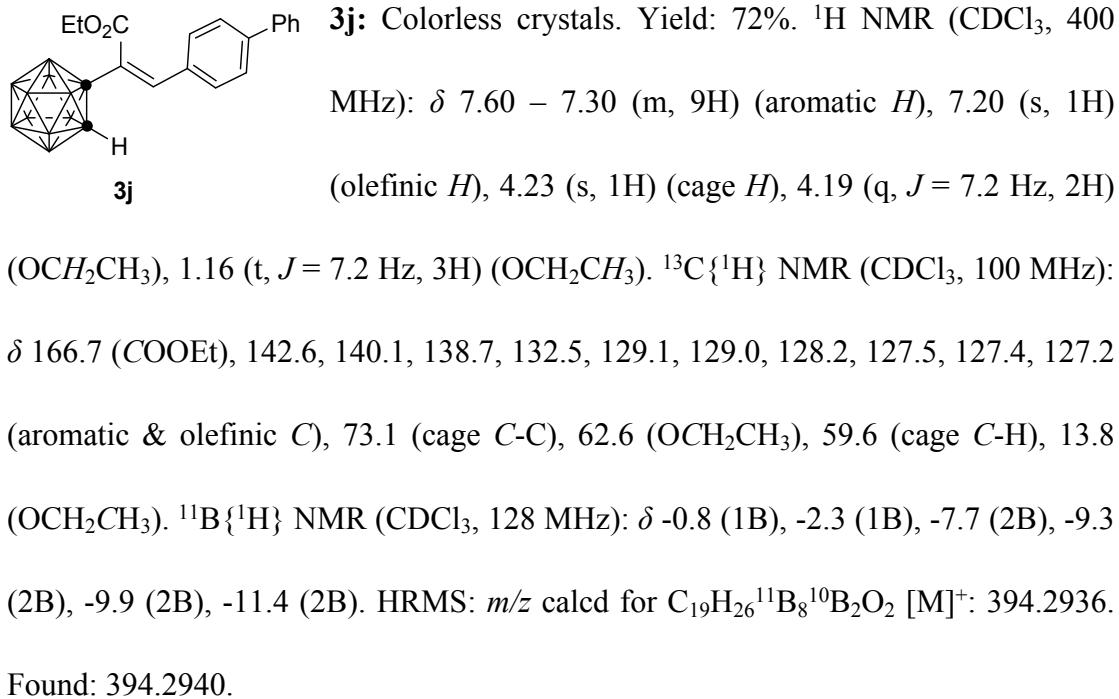


Figure S5. Molecular structure of **3i**

3i: Colorless crystals. Yield: 46%. 1H NMR ($CDCl_3$, 400 MHz): δ 7.49 – 7.10 (m, 4H) (phenyl *H*), 7.09 (s, 1H) (olefinic *H*), 4.20 (s, 1H) (cage *H*), 4.15 (q, J = 7.2 Hz, 2H) (OCH_2CH_3), 1.14 (t, J = 7.2 Hz, 3H) (OCH_2CH_3). $^{13}C\{^1H\}$ NMR ($CDCl_3$, 100 MHz): δ 166.3 (COOEt), 137.8, 132.6, 132.1, 129.9, 128.6, 124.1 (phenyl & olefinic *C*), 72.7

(cage C-C), 62.8 (OCH₂CH₃), 59.5 (cage C-H), 13.8 (OCH₂CH₃). ¹¹B{¹H} NMR (CDCl₃, 128 MHz): δ -0.8 (1B), -2.2 (1B), -7.6 (2B), -9.3 (2B), -10.1 (2B), -11.4 (2B). HRMS: *m/z* calcd for C₁₃H₂₁¹¹B₉¹⁰BBrO₂ [M]⁺: 397.1692. Found: 397.1686.



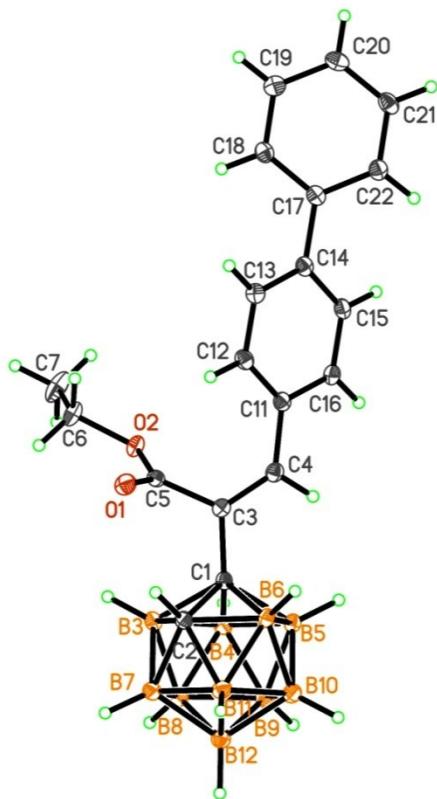
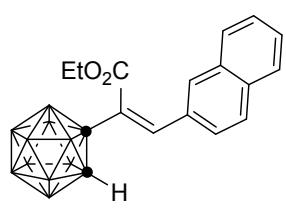


Figure S6. Molecular structure of **3j**



3k: Colorless crystals. Yield: 90%. ^1H NMR (CDCl_3 , 400 MHz): δ 7.81 – 7.29 (m, 8H) (aromatic & olefinic H), 4.27 (s, 1H) (cage H), 4.16 (q, $J = 7.2$ Hz, 2H) (OCH_2CH_3), 1.08 (t, $J = 7.2$ Hz, 3H) (OCH_2CH_3). $^{13}\text{C}\{\text{H}\}$ NMR (CDCl_3 , 100 MHz): δ 166.8 (COOEt), 139.2, 133.7, 133.2, 131.1, 129.0, 128.6, 127.9, 127.8, 127.5, 127.1, 125.0 (aromatic & olefinic C), 73.1 (cage $C\text{-}C$), 62.6 (OCH_2CH_3), 59.6 (cage $C\text{-}H$), 13.8 (OCH_2CH_3). $^{11}\text{B}\{\text{H}\}$ NMR (CDCl_3 , 128 MHz): δ -0.8 (1B), -2.3 (1B), -7.6 (2B), -9.2 (2B), -10.0 (2B), -11.4 (2B). HRMS: m/z calcd for $\text{C}_{17}\text{H}_{24}{^{11}\text{B}}_8{^{10}\text{B}}_2\text{O}_2$ [M] $^+$: 368.2779. Found: 368.2761.

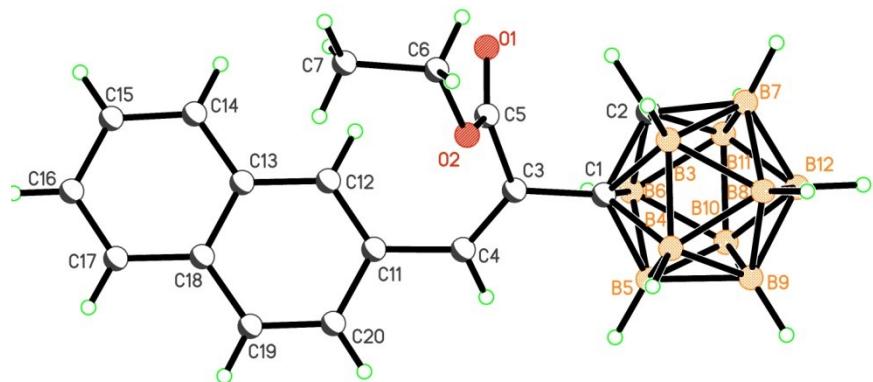


Figure S7. Molecular structure of **3k**

3l: Colorless crystals. Yield: 86%. ¹H NMR ($CDCl_3$, 400 MHz): δ 7.71 – 7.11 (m, 7H) (aromatic & olefinic H), 4.27 (s, 1H) (cage H), 4.19 (q, $J = 7.2$ Hz, 2H) (OCH_2CH_3), 3.93 (s, 3H) (OCH_3), 1.11 (t, $J = 7.2$ Hz, 3H) (OCH_2CH_3). ¹³C{¹H} NMR ($CDCl_3$, 100 MHz): δ 167.0 (COOEt), 159.0, 139.3, 135.1, 130.1, 129.0, 128.8, 128.5, 127.3, 126.7, 125.6, 119.9, 105.9 (aromatic & olefinic C), 73.4 (cage C-C), 62.5 (OCH_2CH_3), 59.7 (cage C-H), 55.6 (OCH_3), 13.8 (OCH_2CH_3). ¹¹B{¹H} NMR ($CDCl_3$, 128 MHz): δ -0.8 (1B), -2.4 (1B), -7.7 (2B), -9.3 (2B), -10.0 (2B), -11.4 (2B). HRMS: *m/z* calcd for $C_{18}H_{26}^{11}B_8^{10}B_2O_3$ [M]⁺: 398.2885. Found: 398.2878.

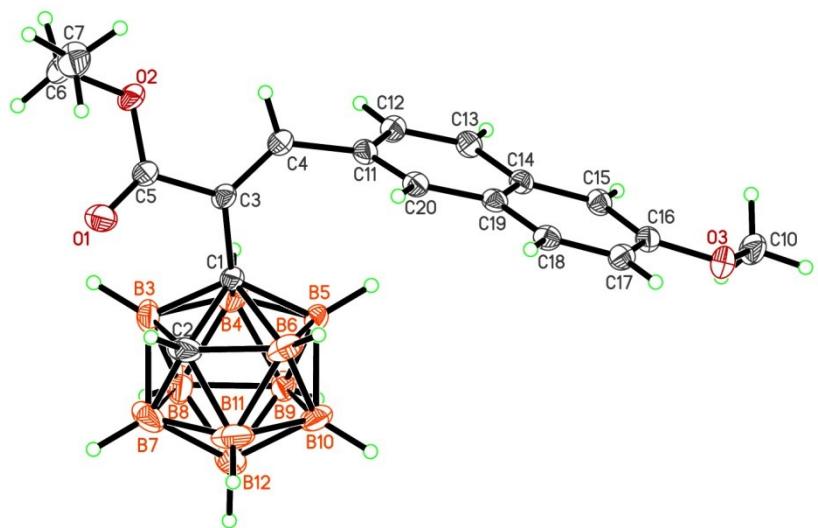


Figure S8. Molecular structure of **3l**

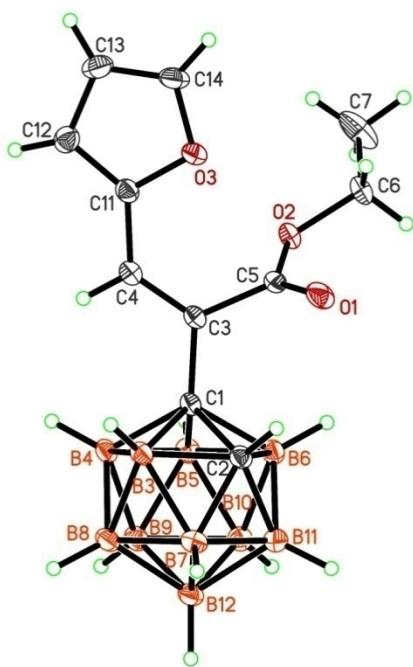
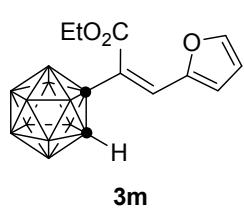
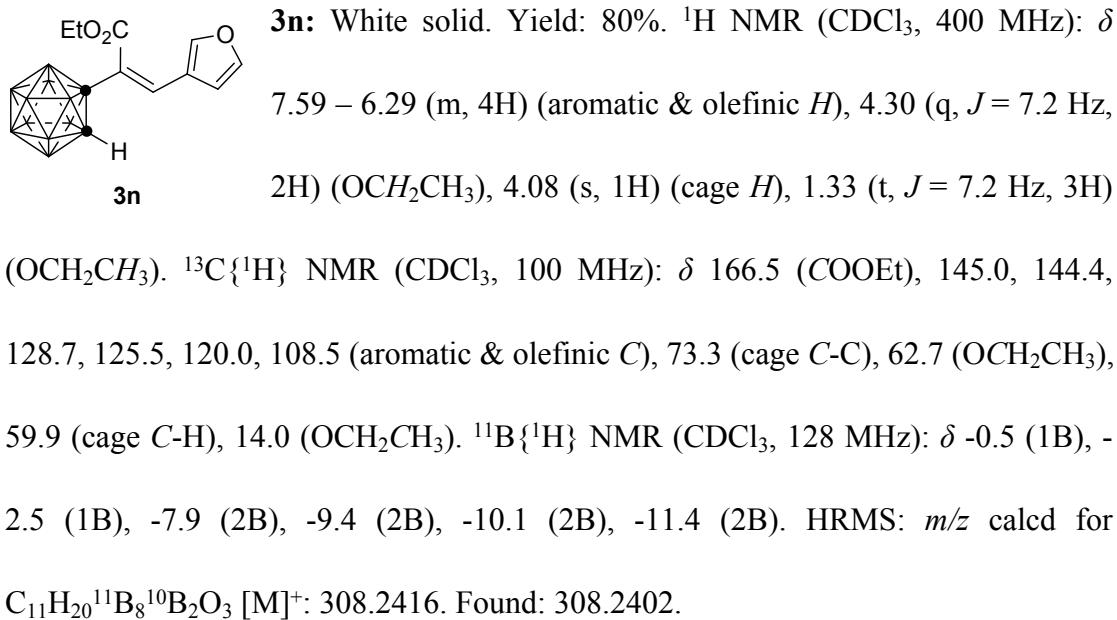


Figure S9. Molecular structure of **3m**



3m: Colorless crystals. Yield: 74%. ^1H NMR (CDCl_3 , 400 MHz): δ 7.41 – 6.44 (m, 4H) (aromatic & olefinic H), 4.34 (q, $J = 7.2$ Hz, 2H) (OCH_2CH_3), 4.07 (s, 1H) (cage H), 1.35 (t, $J = 7.2$ Hz,

3H) (OCH₂CH₃). ¹³C{¹H} NMR (CDCl₃, 100 MHz): δ 166.4 (COOEt), 148.6, 145.1, 124.6, 123.5, 115.8, 112.4 (aromatic & olefinic C), 73.2 (cage C-C), 62.6 (OCH₂CH₃), 59.9 (cage C-H), 14.2 (OCH₂CH₃). ¹¹B{¹H} NMR (CDCl₃, 128 MHz): δ -0.5 (1B), -2.5 (1B), -7.9 (2B), -9.4 (2B), -10.0 (2B), -11.3 (2B). HRMS: *m/z* calcd for C₁₁H₁₉¹¹B₈¹⁰B₂O₃ [M - H]⁺: 307.2337. Found: 307.2349.



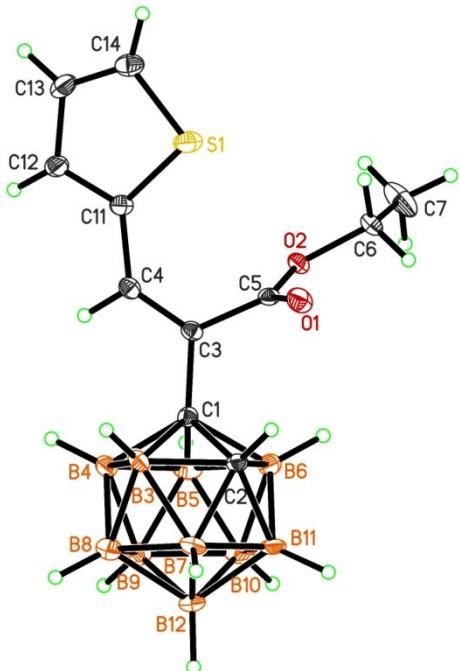
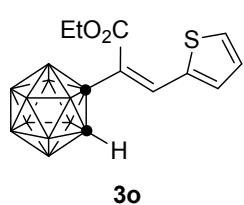


Figure S10. Molecular structure of **3o**



3o: Colorless crystals. Yield: 36%. ^1H NMR (CDCl_3 , 400 MHz): δ 7.43 – 7.03 (m, 4H) (aromatic & olefinic H), 4.35 (q, $J = 7.2$ Hz, 2H) (OCH_2CH_3), 4.13 (s, 1H) (cage H), 1.36 (t, $J = 7.2$ Hz, 3H) (OCH_2CH_3). $^{13}\text{C}\{\text{H}\}$ NMR (CDCl_3 , 100 MHz): δ 166.2 (COOEt), 135.9, 132.7, 131.3, 130.0, 127.7, 123.9 (aromatic & olefinic C), 73.5 (cage C-C), 63.0 (OCH_2CH_3), 60.0 (cage C-H), 14.0 (OCH_2CH_3). $^{11}\text{B}\{\text{H}\}$ NMR (CDCl_3 , 128 MHz): δ -0.5 (1B), -2.5 (1B), -7.9 (2B), -9.4 (2B), -10.0 (2B), -11.4 (2B). HRMS: m/z calcd for $\text{C}_{11}\text{H}_{20}^{11}\text{B}_8^{10}\text{B}_2\text{O}_2\text{S} [\text{M}]^+$: 324.2187. Found: 324.2181.

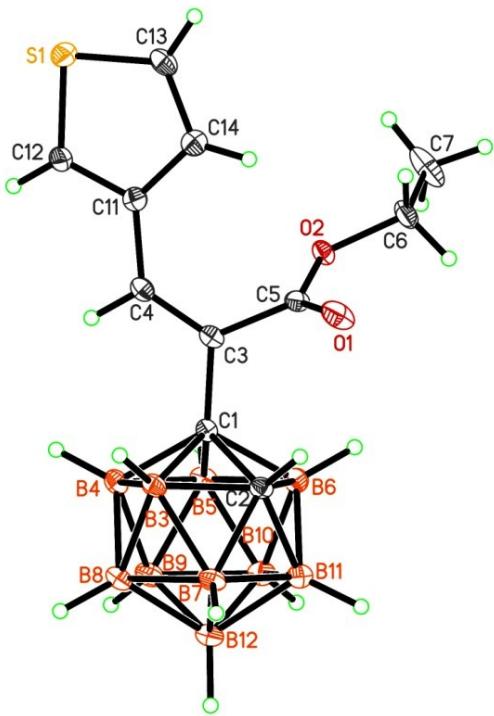
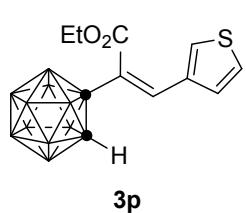
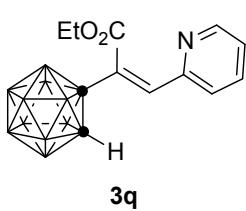


Figure S11. Molecular structure of 3p



Colorless crystals. Yield: 97%. ^1H NMR (CDCl_3 , 400 MHz): δ 7.37 – 6.97 (m, 4H) (aromatic & olefinic H), 4.26 (q, $J = 7.2$ Hz, 2H) (OCH_2CH_3), 4.11 (s, 1H) (cage H), 1.27 (t, $J = 7.2$ Hz, 3H) (OCH_2CH_3). $^{13}\text{C}\{\text{H}\}$ NMR (CDCl_3 , 100 MHz): δ 166.8 (COOEt), 134.7, 132.3, 128.1, 126.9, 126.6, 126.0 (aromatic & olefinic C), 73.2 (cage C -C), 62.7 (OCH_2CH_3), 59.9 (cage C -H), 14.0 (OCH_2CH_3). $^{11}\text{B}\{\text{H}\}$ NMR (CDCl_3 , 128 MHz): δ -0.6 (1B), -2.5 (1B), -7.8 (2B), -9.4 (2B), -10.0 (2B), -11.3 (2B). HRMS: m/z calcd for $\text{C}_{11}\text{H}_{20}^{11}\text{B}_8^{10}\text{B}_2\text{O}_2\text{S} [\text{M}]^+$: 324.2187. Found: 324.2176.



Colorless crystals. Yield: 88%. ^1H NMR (CDCl_3 , 400 MHz): δ 8.51 – 7.21 (m, 4H) (aromatic H), 7.00 (s, 1H) (olefinic H), 4.30 (q, $J = 7.2$ Hz, 2H) (OCH_2CH_3), 4.13 (s, 1H) (cage H), 1.28

(t, $J = 7.2$ Hz, 3H) (OCH_2CH_3). $^{13}\text{C}\{\text{H}\}$ NMR (CDCl_3 , 100 MHz): δ 166.6 (COOEt), 151.2, 149.5, 137.0, 135.7, 130.5, 125.3, 124.2 (aromatic & olefinic C), 72.7 (cage C-C), 62.3 (OCH_2CH_3), 59.4 (cage C-H), 14.0 (OCH_2CH_3). $^{11}\text{B}\{\text{H}\}$ NMR (CDCl_3 , 128 MHz): δ -0.7 (1B), -2.3 (1B), -7.7 (2B), -9.4 (2B), -9.9 (2B), -11.4 (2B). HRMS: m/z calcd for $\text{C}_{12}\text{H}_{20}^{11}\text{B}_8^{10}\text{B}_2\text{NO}_2$ [M - H] $^+$: 318.2497. Found: 318.2505.

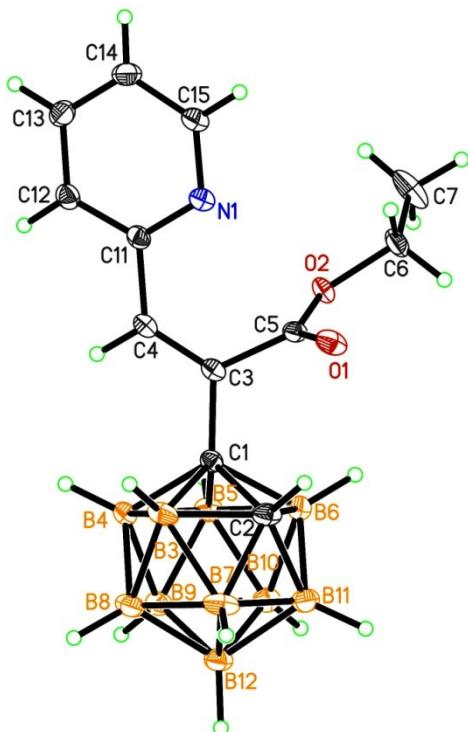
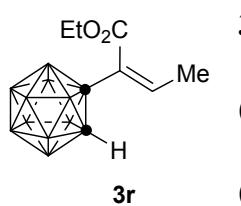
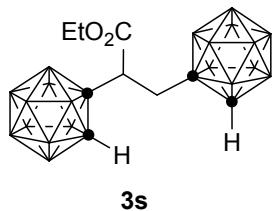


Figure S12. Molecular structure of **3q**

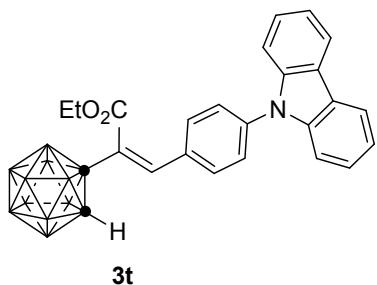


3r: Colorless oil. Yield: 87%. ^1H NMR (CDCl_3 , 400 MHz): δ 6.44 (q, $J = 7.2$ Hz, 1H) (olefinic H), 4.27 (q, $J = 7.2$ Hz, 2H) (OCH_2CH_3), 4.25 (s, 1H) (cage H), 1.81 (d, $J = 7.2$ Hz, 3H) ($=\text{CHCH}_3$), 1.25 (t, $J = 7.2$ Hz, 3H) (OCH_2CH_3). $^{13}\text{C}\{\text{H}\}$ NMR (CDCl_3 , 100 MHz): δ 165.8 (COOEt), 138.8, 129.0 (olefinic C), 72.8 (cage C-C), 62.2 (OCH_2CH_3), 59.4 (cage C-H), 16.2 ($=\text{CHCH}_3$), 14.0 (OCH_2CH_3). $^{11}\text{B}\{\text{H}\}$ NMR (CDCl_3 , 128 MHz): δ -1.1 (1B), -2.7 (1B), -7.9 (2B), -9.3 (2B), -10.2 (2B), -11.6 (2B). HRMS: m/z calcd for

$C_8H_{19}^{11}B_8^{10}B_2O_2 [M - H]^+$: 255.2388. Found: 255.2388.

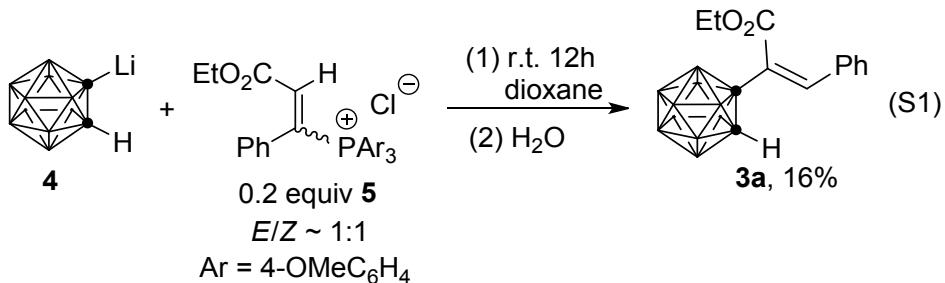


3s: White solid. Yield: 10%. 1H NMR ($CDCl_3$, 400 MHz): δ 4.28 (m, 2H) (OCH_2CH_3), 3.93 (s, 1H) (cage H), 3.63 (s, 1H) (cage H), 3.33 (d, $J = 11.6$ Hz, 1H) ($CHCH_2$), 2.97 (t, $J = 12.0$ Hz, 1H) ($CHCH_2$), 2.68 (d, $J = 15.2$ Hz, 1H) ($CHCH_2$), 1.37 (t, $J = 7.2$ Hz, 3H) (OCH_2CH_3). $^{13}C\{^1H\}$ NMR ($CDCl_3$, 100 MHz): δ 168.7 (COOEt), 72.2, 72.0 (cage C-C), 63.6 (OCH_2CH_3), 61.9, 59.7 (cage C-H), 50.9 ($CHCH_2$), 40.1 ($CHCH_2$), 14.1 (OCH_2CH_3). $^{11}B\{^1H\}$ NMR ($CDCl_3$, 128 MHz): δ -0.2 (2B), -2.1 (1B), -3.3 (1B), -7.8 (4B), -9.5 (2B), -10.1 (2B), -11.3 (8B). HRMS: m/z calcd for $C_9H_{29}^{11}B_{16}^{10}B_4O_2 [M - H]^+$: 385.4174. Found: 385.4185.



3t: Light yellow solid. Yield: 33%. 1H NMR ($CDCl_3$, 400 MHz): δ 8.15 – 7.28 (m, 13H) (aromatic & olefinic H), 4.27 (s, 1H) (cage H), 4.25 (q, $J = 7.2$ Hz, 2H) (OCH_2CH_3), 1.22 (t, $J = 7.2$ Hz, 3H) (OCH_2CH_3). $^{13}C\{^1H\}$ NMR ($CDCl_3$, 100 MHz): δ 166.5 (COOEt), 140.5, 139.1, 138.0, 132.4, 130.1, 128.4, 127.0, 126.3, 123.8, 120.7, 120.6, 109.7 (aromatic & olefinic C), 72.9 (cage C-C), 62.8 (OCH_2CH_3), 59.5 (cage C-H), 13.9 (OCH_2CH_3). $^{11}B\{^1H\}$ NMR ($CDCl_3$, 128 MHz): δ -0.7 (1B), -2.1 (1B), -7.6 (2B), -9.3 (2B), -9.9 (2B), -11.3 (2B). HRMS: m/z calcd for $C_{25}H_{29}^{11}B_9^{10}BNO_2 [M]^+$: 484.3165. Found: 484.3156.

Mechanistic study.

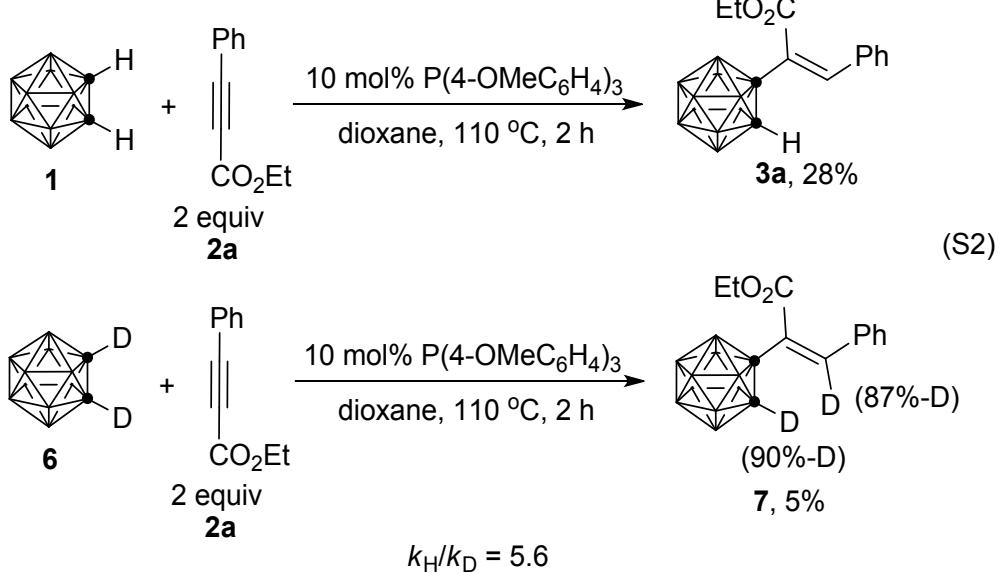


Reaction of 1-Li-1,2-C₂B₁₀H₁₁ (4) with Vinylphosphonium Salt (5). To a dioxane solution (10 mL) of *o*-carborane (72 mg, 0.50 mmol) was added ⁷BuLi (0.2 mL, 2.5 M in hexane, 0.50 mmol) at 0 °C. After stirring for 1 h at room temperature, a solution of vinylphosphonium salt (**5**)¹ (57 mg, 0.10 mmol) in dioxane (10 mL) was added dropwise to the reaction suspension at room temperature. The yellow suspension was stirred overnight. After water (10 mL) was added to reaction mixture, the aqueous layer was extracted with ethyl acetate (10 × 3 mL). The combined organic layer was then dried over anhydrous Na₂SO₄. After filtration and removal of solvent in vacuo, the residue was subjected to flash column chromatography on silica gel (230–400 mesh) using *n*-hexane/dichloromethane (20/1 in v/v) as eluent to give **3a** (26 mg, 16%, 0.08 mmol). See eq S1.

Kinetic Isotope Effect Experiments.

Reaction of *o*-Carborane (1) with Ethyl Phenylpropiolate (2a). *o*-Carborane (29 mg, 0.20 mmol), ethyl phenylpropiolate (70 mg, 0.40 mmol) and tris(4-methoxy phenyl)phosphine (7 mg, 0.02 mmol) were mixed in dioxane (0.6 mL). The resulting mixture was heated at 100 °C for 2 h. GC-MS yield: 28%. See eq S2.

$1,2\text{-C}_2\text{B}_{10}\text{H}_{10}$ ² (29 mg, 0.20 mmol), ethyl phenylpropiolate (70 mg, 0.40 mmol) and tris(4-methoxy phenyl)phosphine (7 mg, 0.02 mmol) were mixed in dioxane (0.6 mL). The resulting mixture was heated at 110 °C for 2 h. GC-MS yield: 5%. See eq S2.



Calculations based on the GC-MS showed a $k_{\text{H}}/k_{\text{D}}$ value of 5.6 (eq S2). Moreover, product 7 was obtained with 87% deuterium incorporation by flash column chromatography on silica gel (230–400 mesh) using *n*-hexane/dichloromethane (20/1 in v/v) as eluent. See eq S2 and Figures S13 and S14.

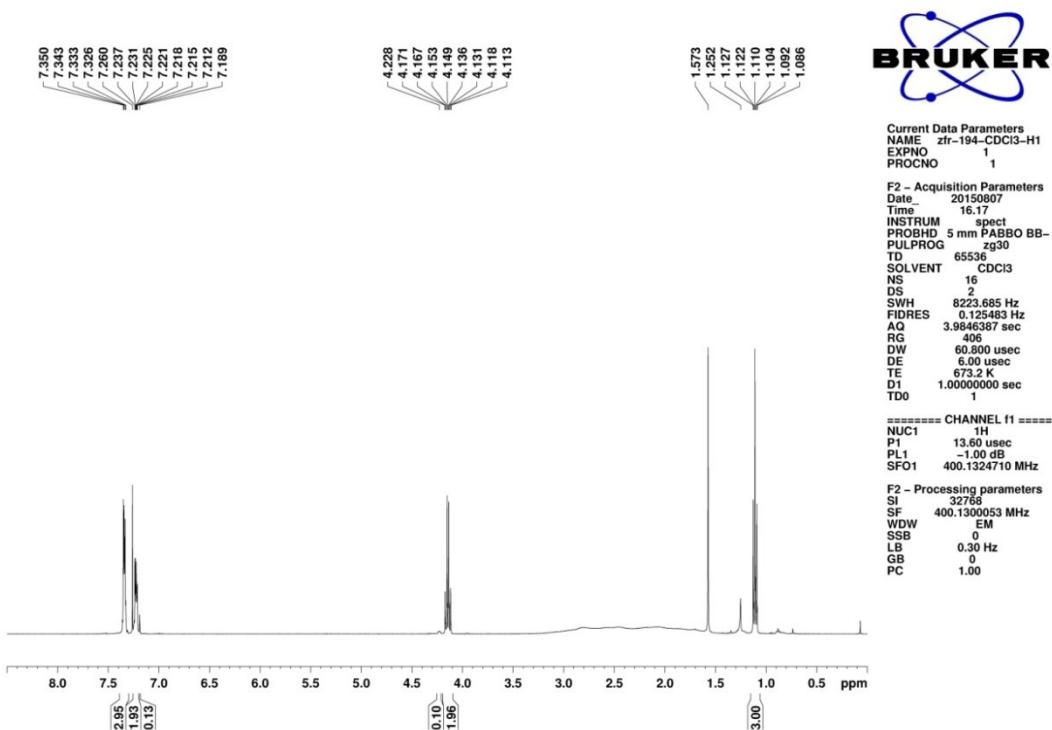


Figure S13. ^1H NMR spectrum of 7

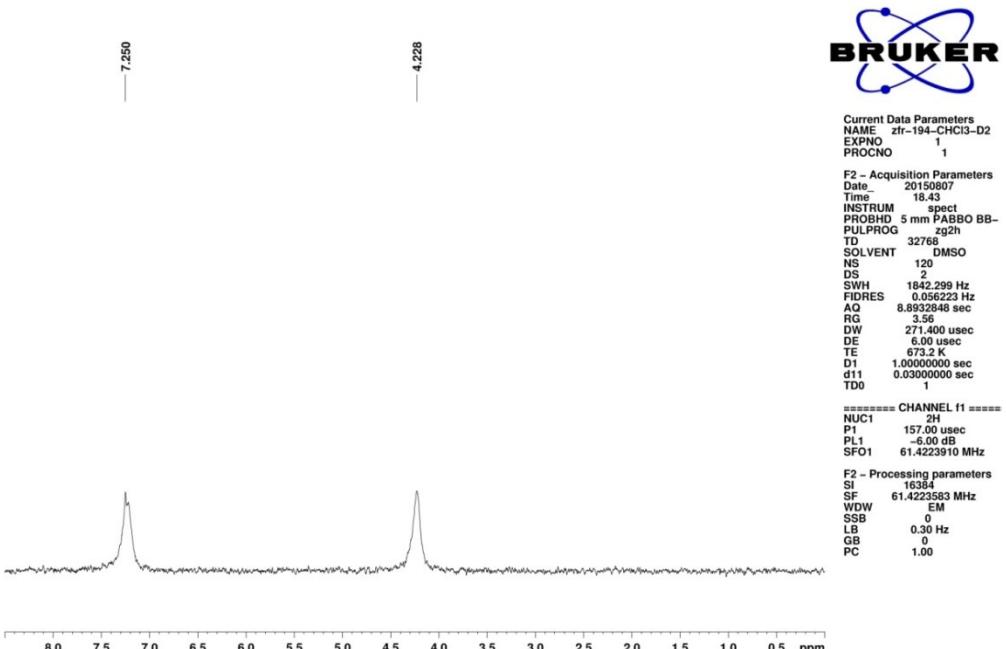
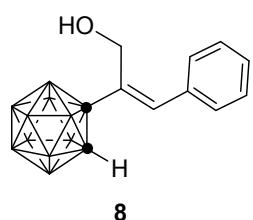


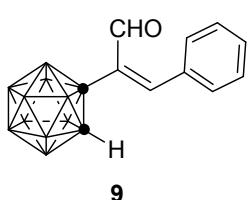
Figure S14. ^2H NMR spectrum of 7

Preparation of 8: To a solution of **3a** (64 mg, 0.20 mmol) in dry dichloromethane (5 mL) at – 78 °C was added DIBAL-H (0.4 mL of 1.0 M in *n*-hexane, 0.40 mmol) dropwise. The resulting mixture was further stirred at this temperature for another 2 h and then quenched by addition of potassium sodium tartrate solution (5 mL, 1.0 M in H₂O). The organic layer was separated from the aqueous layer which was further extracted with dichloromethane (10 x 3 mL). The combined organic layer was dried with anhydrous Na₂SO₄. After filtration and removal of solvent in vacuo, the residue was subjected to flash column chromatography on silica gel (230–400 mesh) using *n*-hexane/ethyl acetate (10/1 in v/v) as eluent to give **8** (32 mg, 58%, 0.12 mmol).



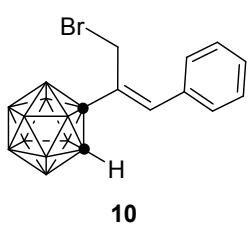
8: White solid. ¹H NMR (CDCl₃, 400 MHz): δ 7.30 (m, 3H) (phenyl *H*), 7.19 (m, 2H) (phenyl *H*), 7.07 (s, 1H) (olefinic *H*), 4.23 (s, 2H) (CH₂OH), 4.19 (s, 1H) (cage *H*). ¹³C{¹H} NMR (CDCl₃, 100 MHz): δ 137.7, 133.7, 131.6, 127.7, 127.6, 127.5 (phenyl & olefinic *C*), 59.1 (CH₂OH), 58.3 (cage *C-H*), cage *C-C* atom was not observed. ¹¹B{¹H} NMR (CDCl₃, 128 MHz): δ -1.3 (1B), -2.7 (1B), -7.7 (2B), -9.0 (2B), -10.2 (2B), -11.6 (2B). HRMS: *m/z* calcd for C₁₁H₂₀¹¹B₈¹⁰B₂O [M]⁺: 276.2517. Found: 276.2522.

Preparation of 9: To a solution of **8** (55 mg, 0.20 mmol) in dry dichloromethane (5 mL) at room temperature was added activated MnO₂ (348 mg, 4.0 mmol) in one portion. The resulting mixture was further stirred at this temperature for 48 h. After filtration through celite and removal of solvent in vacuo, the residue was subjected to flash column chromatography on silica gel (230–400 mesh) using *n*-hexane/ethyl acetate (15/1 in v/v) as eluent to give **9** (46 mg, 83%, 0.17 mmol).



9: White solid. ^1H NMR (CDCl_3 , 400 MHz): δ 9.59 (s, 1H) (CHO), 8.25 (s, 1H) (olefinic H), 7.47 (m, 3H) (phenyl H), 7.29 (m, 2H) (phenyl H), 5.49 (s, 1H) (cage H). $^{13}\text{C}\{\text{H}\}$ NMR (CDCl_3 , 100 MHz): δ 189.7 (CHO), 155.0, 131.9, 131.8, 131.3, 130.7, 129.1 (phenyl & olefinic C), 72.4 (cage C-C), 57.5 (cage C-H). $^{11}\text{B}\{\text{H}\}$ NMR (CDCl_3 , 128 MHz): δ -2.3 (2B), -7.2 (2B), -8.4 (2B), -10.6 (2B), -11.8 (2B). HRMS: m/z calcd for $\text{C}_{11}\text{H}_{18}^{11}\text{B}_8^{10}\text{B}_2\text{O} [\text{M}]^+$: 274.2361. Found: 274.2363.

Preparation of 10: To a solution of **8** (55 mg, 0.20 mmol) and tetrabromomethane (133mg, 0.40 mmol) in dry dichloromethane (2 mL) at 0 °C was added triphenylphosphine (157 mg, 0.60 mmol) in one portion. The resulting mixture was allowed to warm to room temperature gradually. A mixture of *n*-hexane/diethyl ether (10 mL, 20/1 in v/v) was added. After filtration through celite and removal of solvents in vacuo, the residue was subjected to flash column chromatography on silica gel (230–400 mesh) using *n*-hexane as eluent to give **10** (54 mg, 80%, 0.16 mmol).



10: White solid. ^1H NMR (CDCl_3 , 400 MHz): δ 7.42 (m, 5H) (phenyl H), 7.18 (s, 1H) (olefinic H), 4.16 (br, 3H) (CH_2Br & cage H). $^{13}\text{C}\{\text{H}\}$ NMR (CDCl_3 , 100 MHz): δ 141.0, 134.5, 129.6, 129.4, 129.1, 128.9 (phenyl & olefinic C), 77.4 (cage C-C), 59.8 (cage C-H), 30.7 (CH_2Br). $^{11}\text{B}\{\text{H}\}$ NMR (CDCl_3 , 128 MHz): δ -0.8 (1B), -2.4 (1B), -7.9 (2B), -8.9 (2B), -10.4 (2B), -11.4 (2B). HRMS: m/z calcd for $\text{C}_{11}\text{H}_{19}^{11}\text{B}_8^{10}\text{B}_2\text{Br} [\text{M}]^+$: 338.1673. Found: 338.1689.

X-ray Structure Determination. All data were collected with an Oxford

Diffraction Gemini™ S Ultra X-ray Diffractometer with monochromatized Cu-K α radiation ($\lambda = 1.54178 \text{ \AA}$) at 173 K or 100 K. Diffraction data were collected and processed using the CrysAlisPro software (version 1.171.35.19). Empirical absorption corrections were performed using spherical harmonics, implemented in SCALE3 ABSPACK scaling algorithm in the CrysAlisPro software suite. Structure solution and refinement for all compounds were performed using the Olex2 software package¹³ (which embedded SHELXTL).¹⁴ All the structures were solved by direct methods, expanded by difference Fourier syntheses and refined by full matrix least-squares on F². All non-hydrogen atoms were refined anisotropically with a riding model for the hydrogen atoms except noted separately.

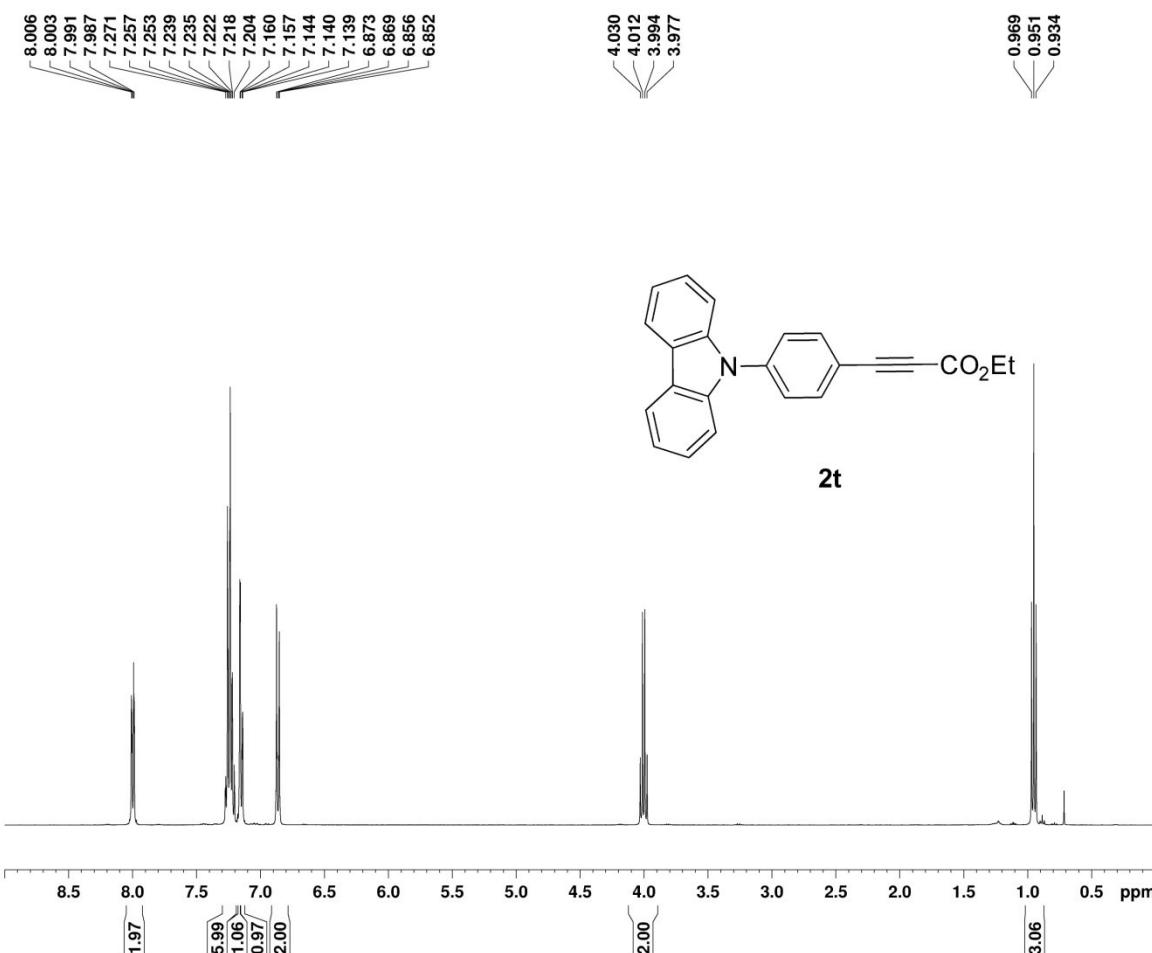
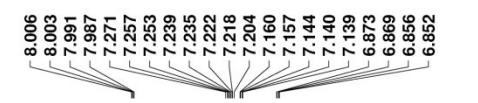
CCDC 1432244–1432255 (**3a**, **3b**, **3f**, **3h–3m** and **3o–3q**) contains the supplementary crystallographic data for this paper. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.

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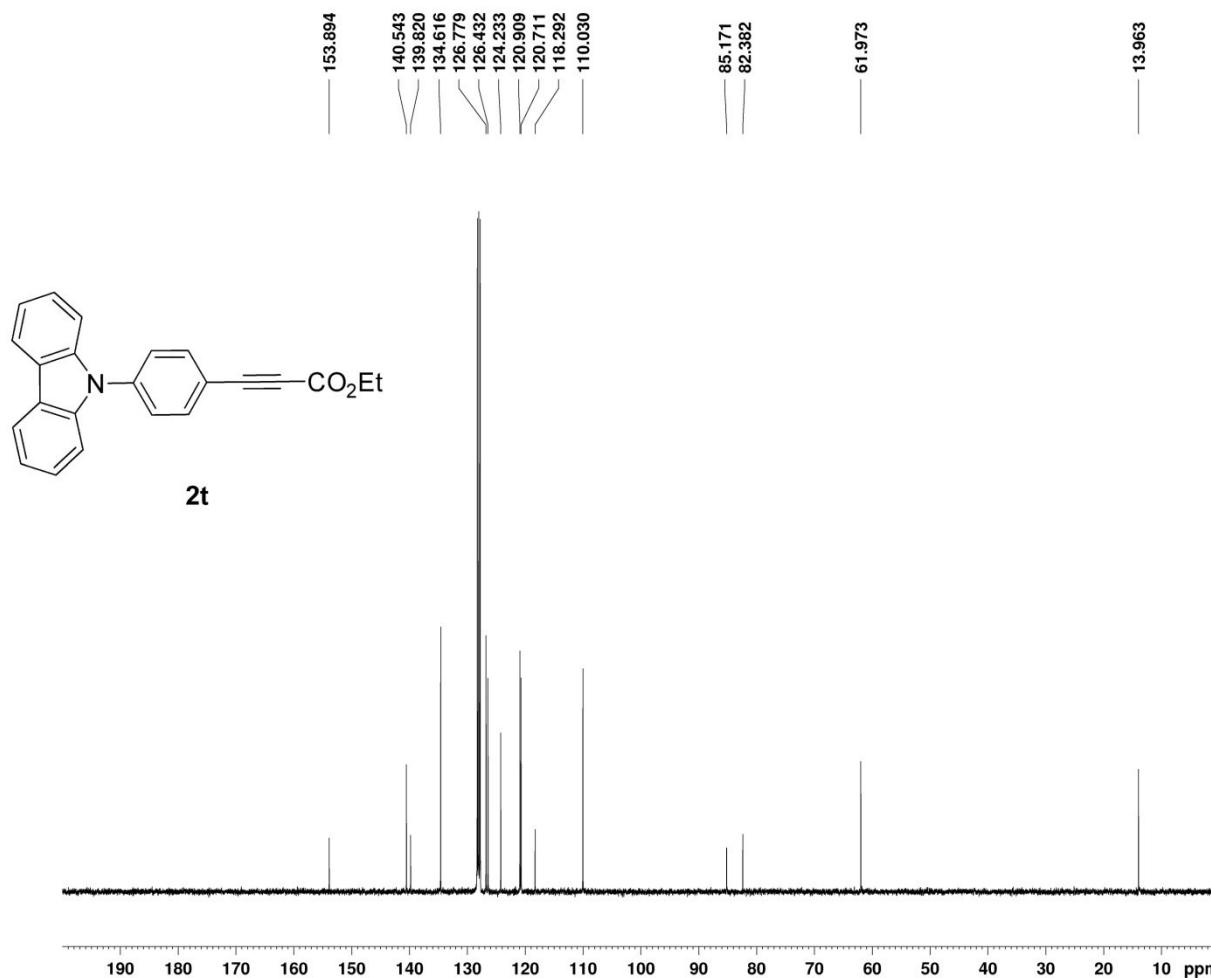


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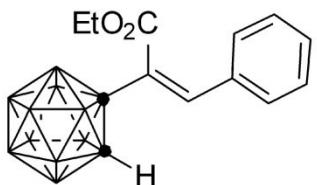
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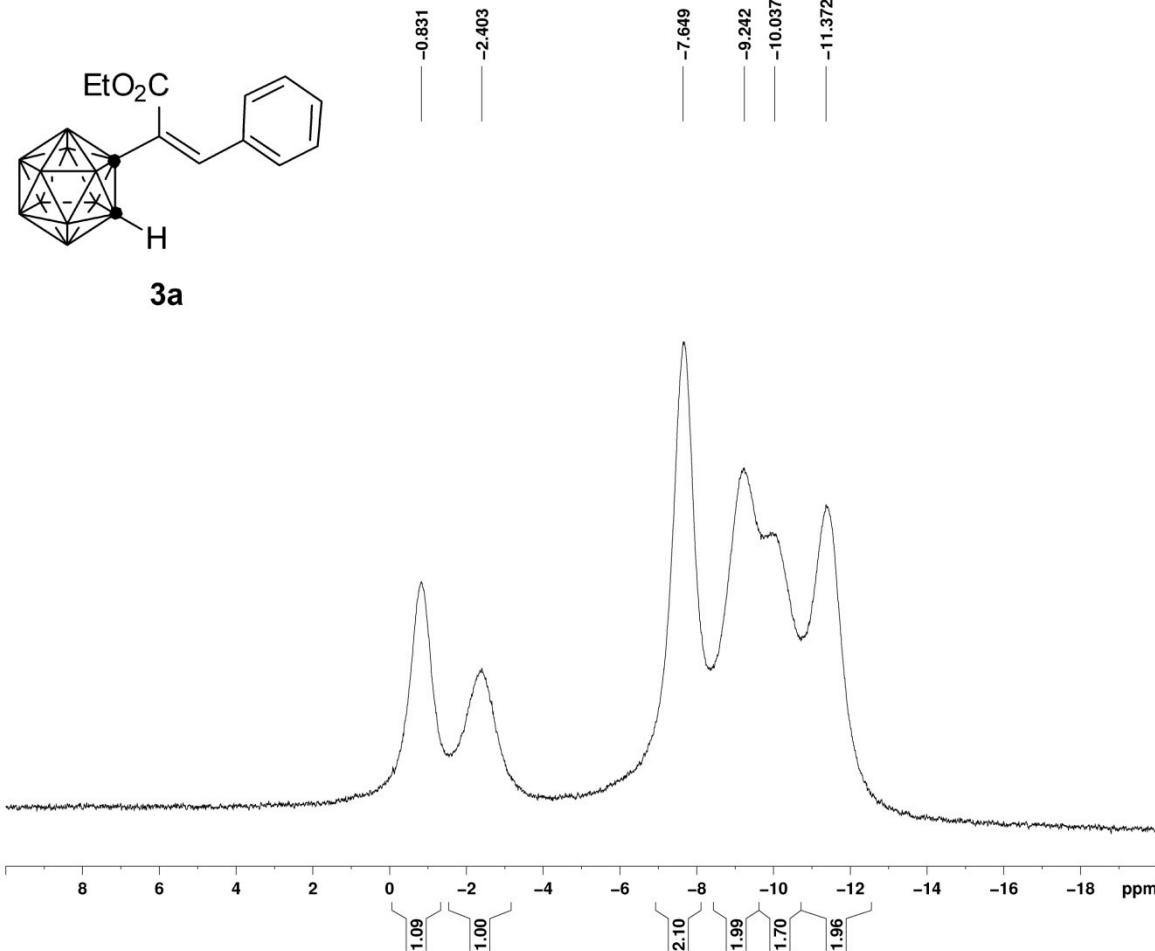
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3a



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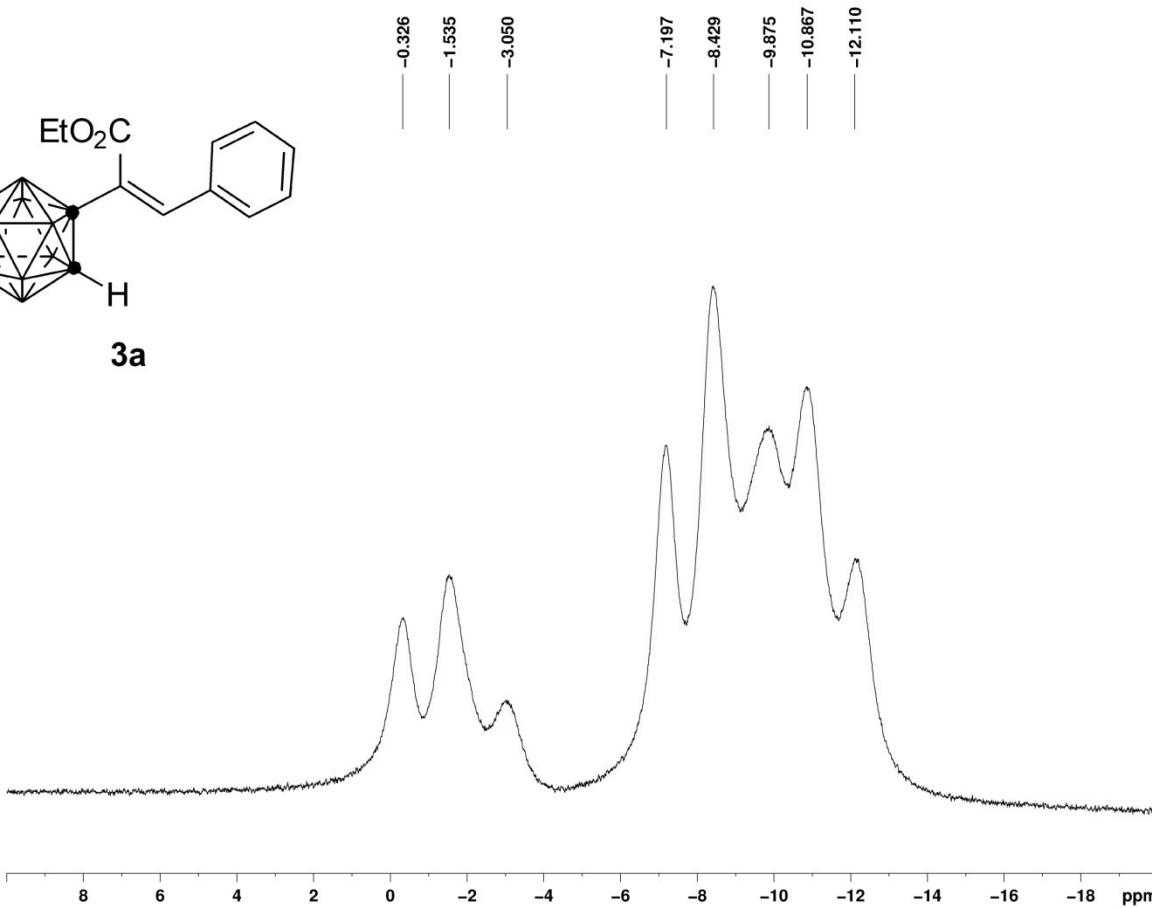
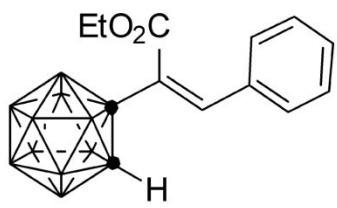
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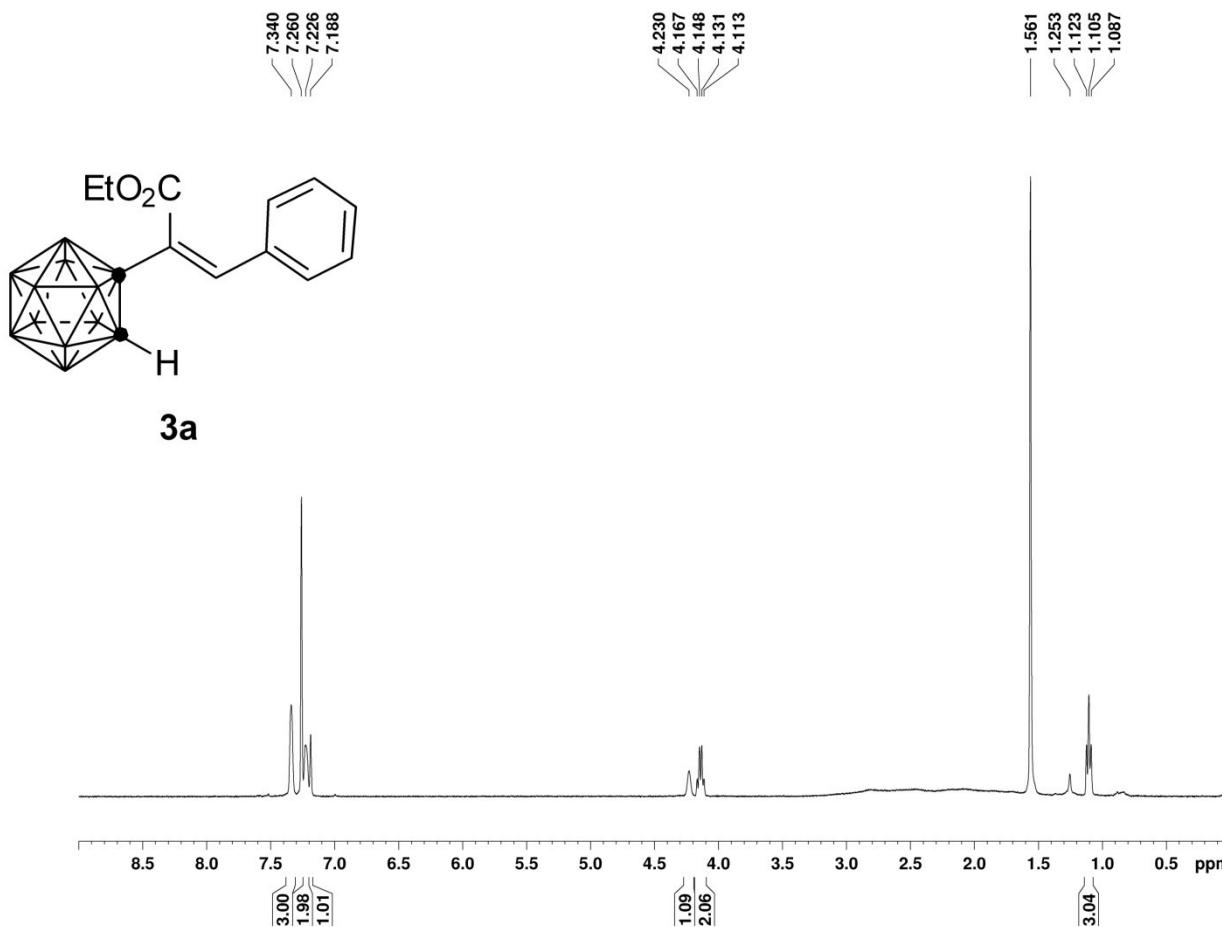


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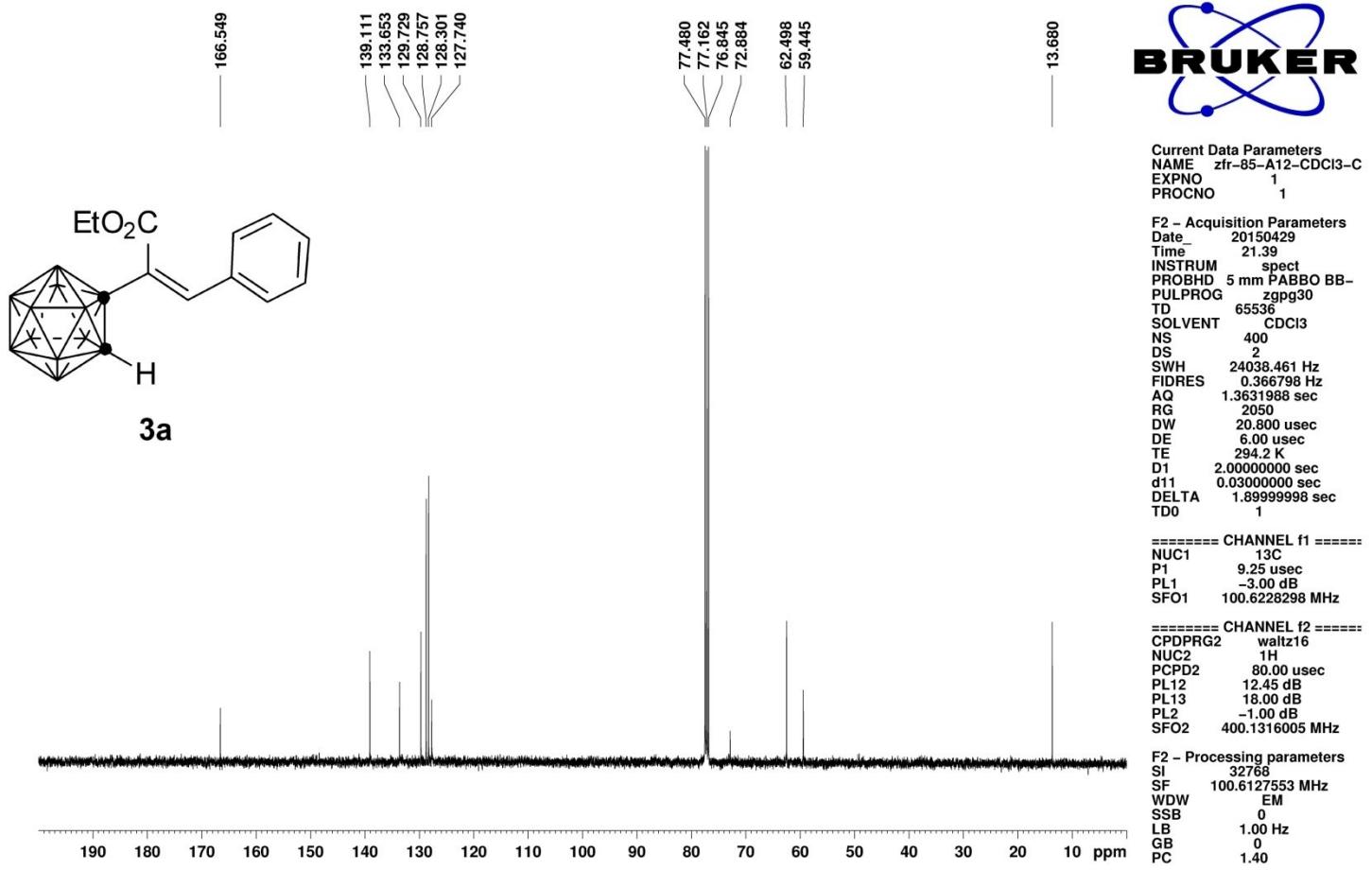


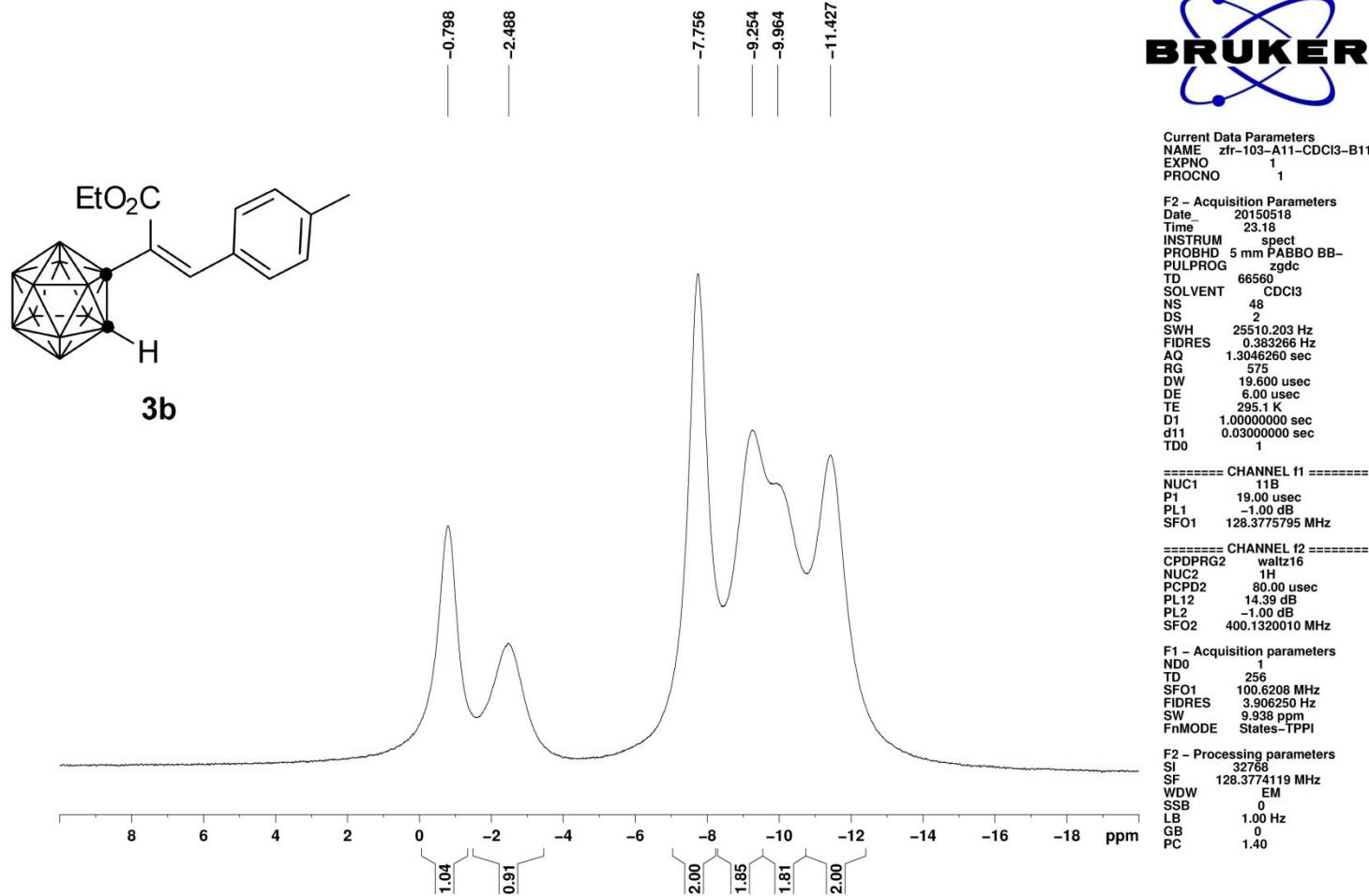
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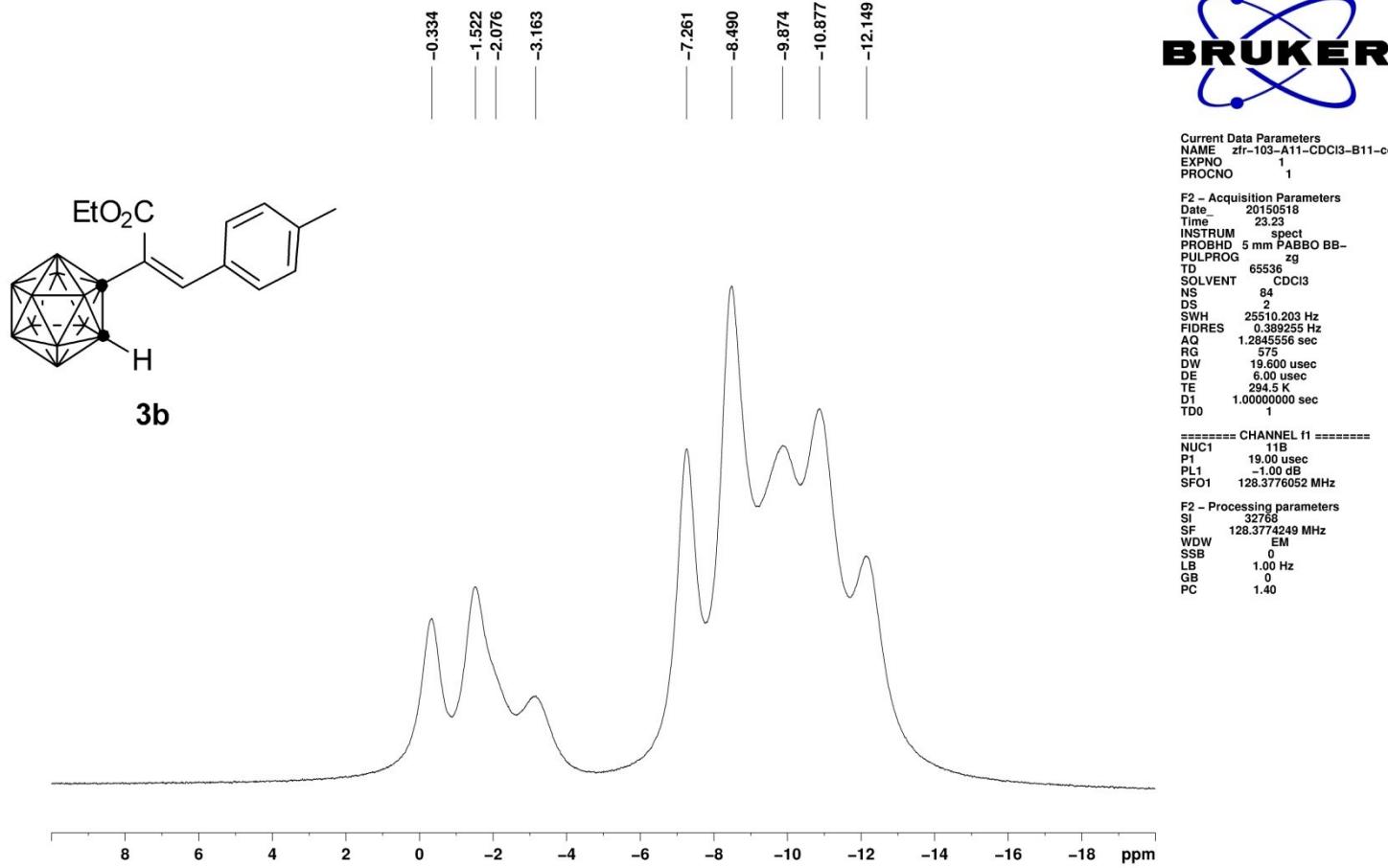
F2 - Acquisition Parameters
 Date 20150421
 Time 9.34
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 8223.685 Hz
 FIDRES 0.125483 Hz
 AQ 3.9846387 sec
 RG 724
 DW 60.800 usec
 DE 6.00 usec
 TE 294.2 K
 D1 1.0000000 sec
 TD0 1

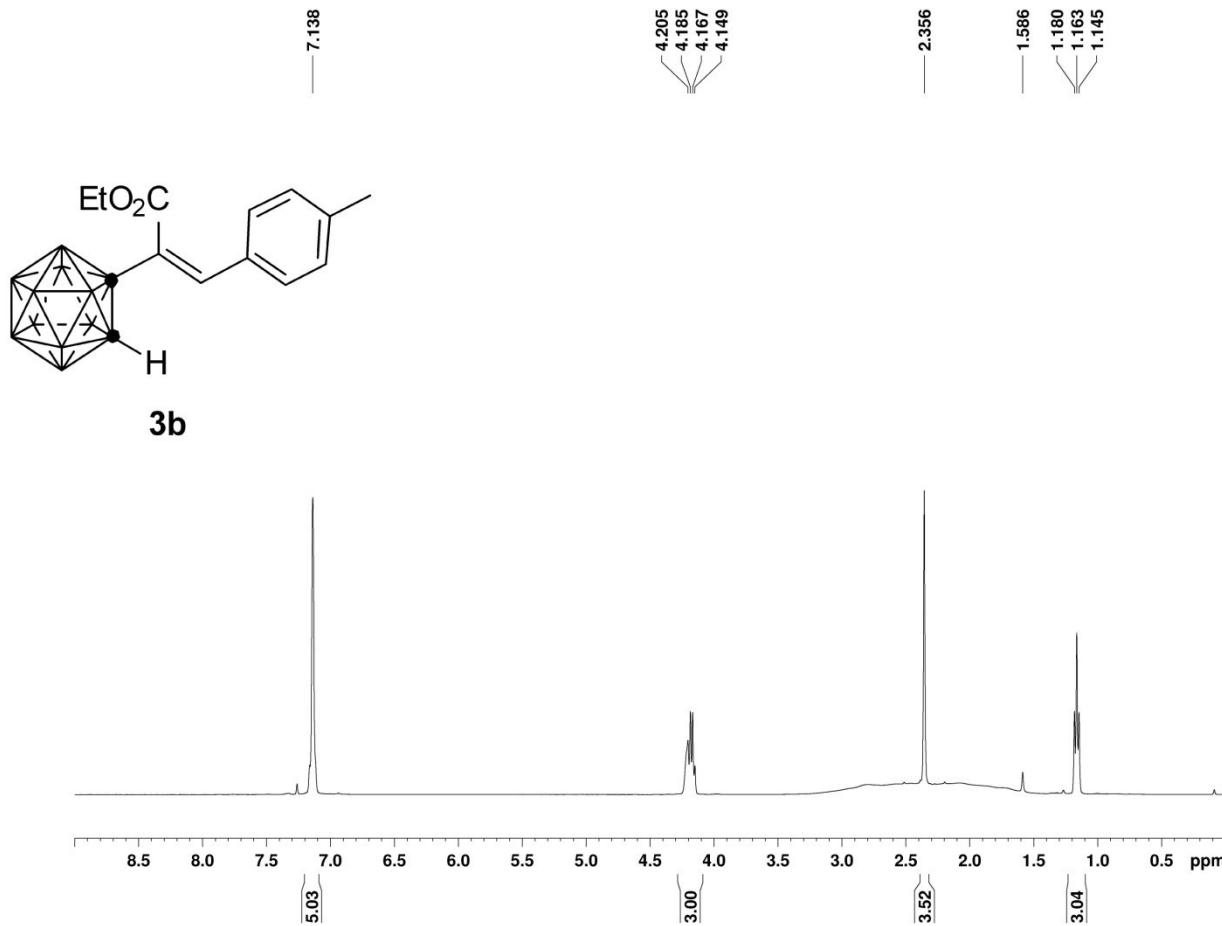
===== CHANNEL f1 =====
 NUC1 1H
 P1 13.60 usec
 PL1 -1.00 dB
 SFO1 400.1324710 MHz

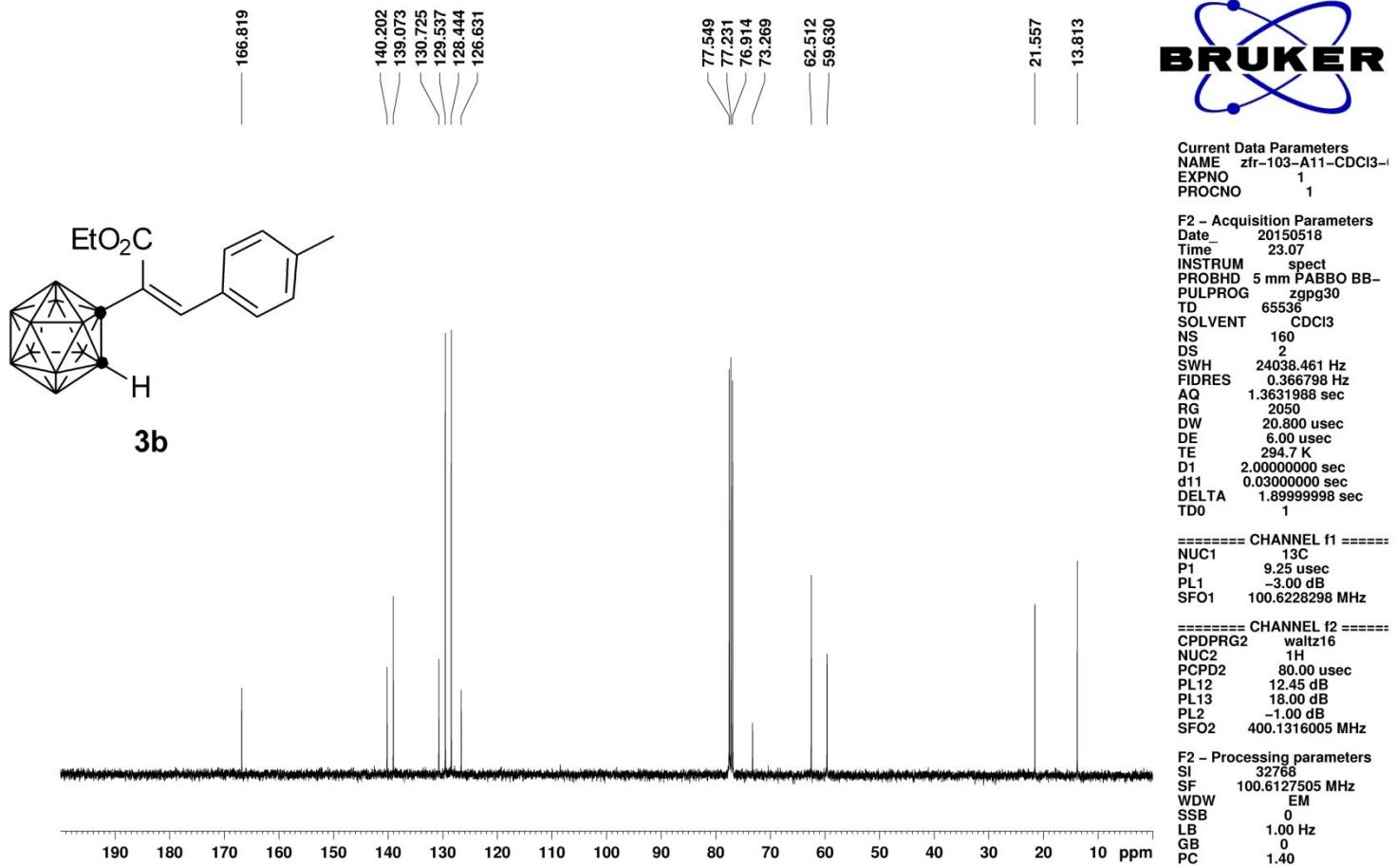
F2 - Processing parameters
 SI 32768
 SF 400.1300051 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

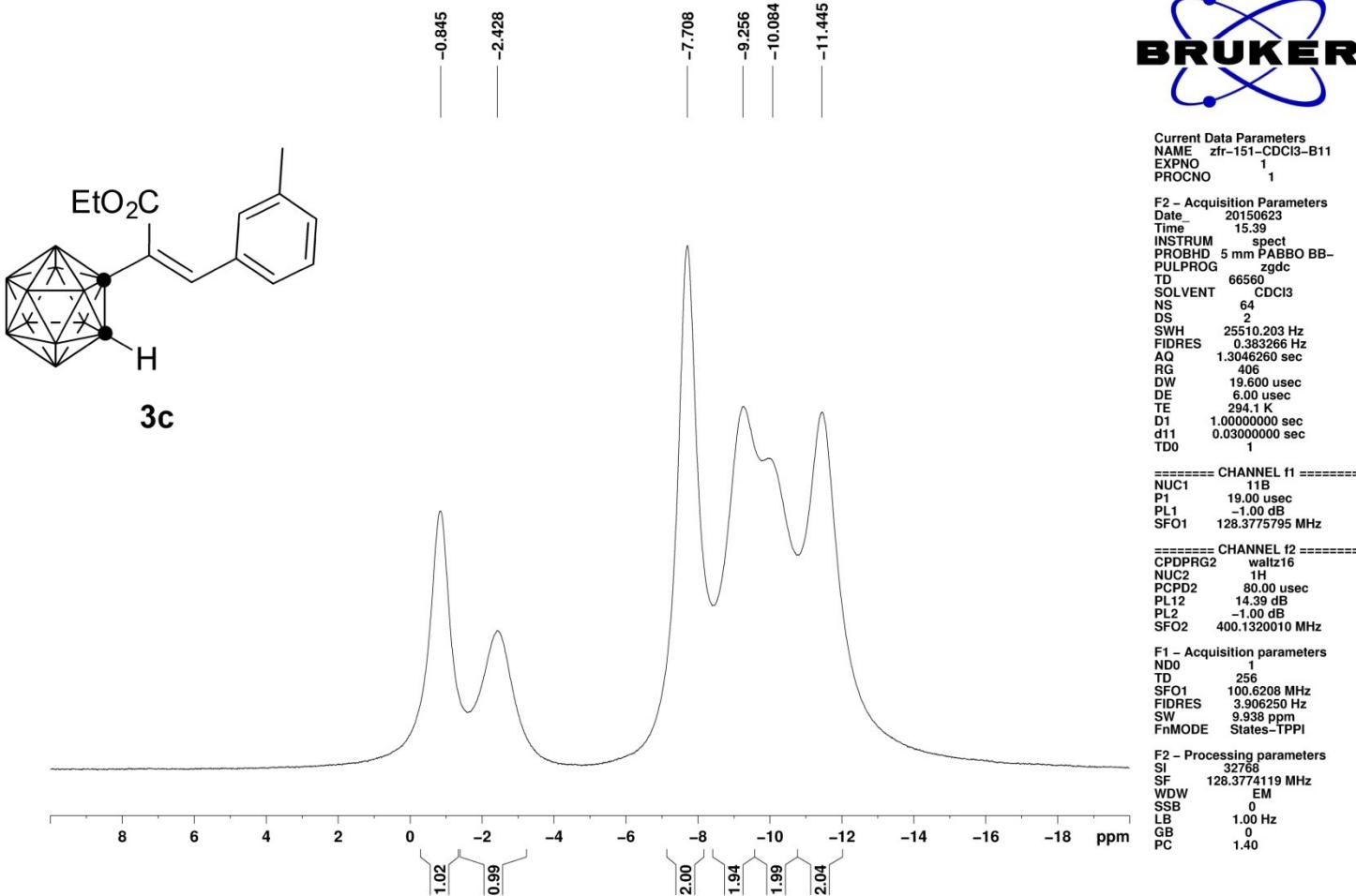


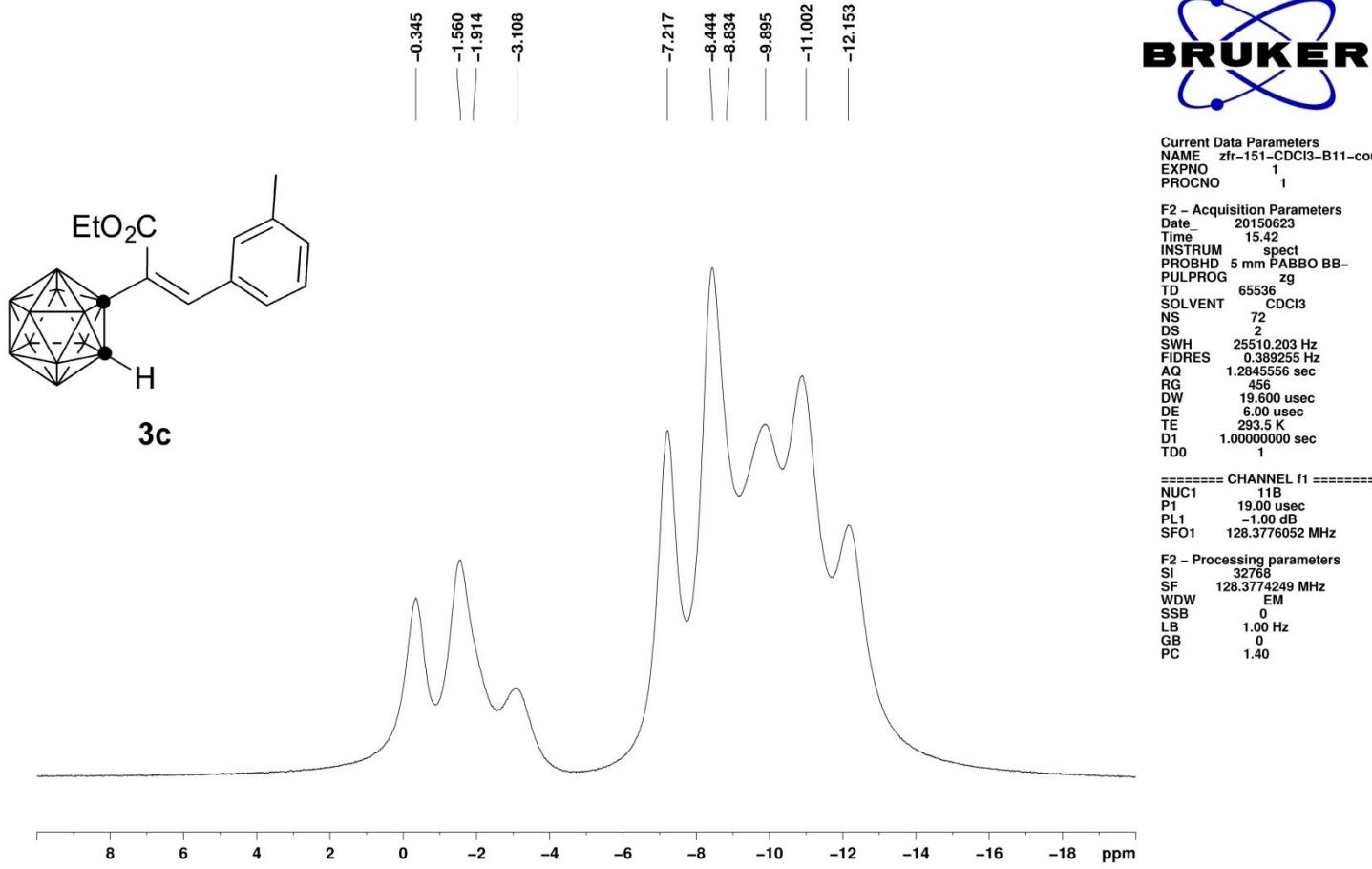


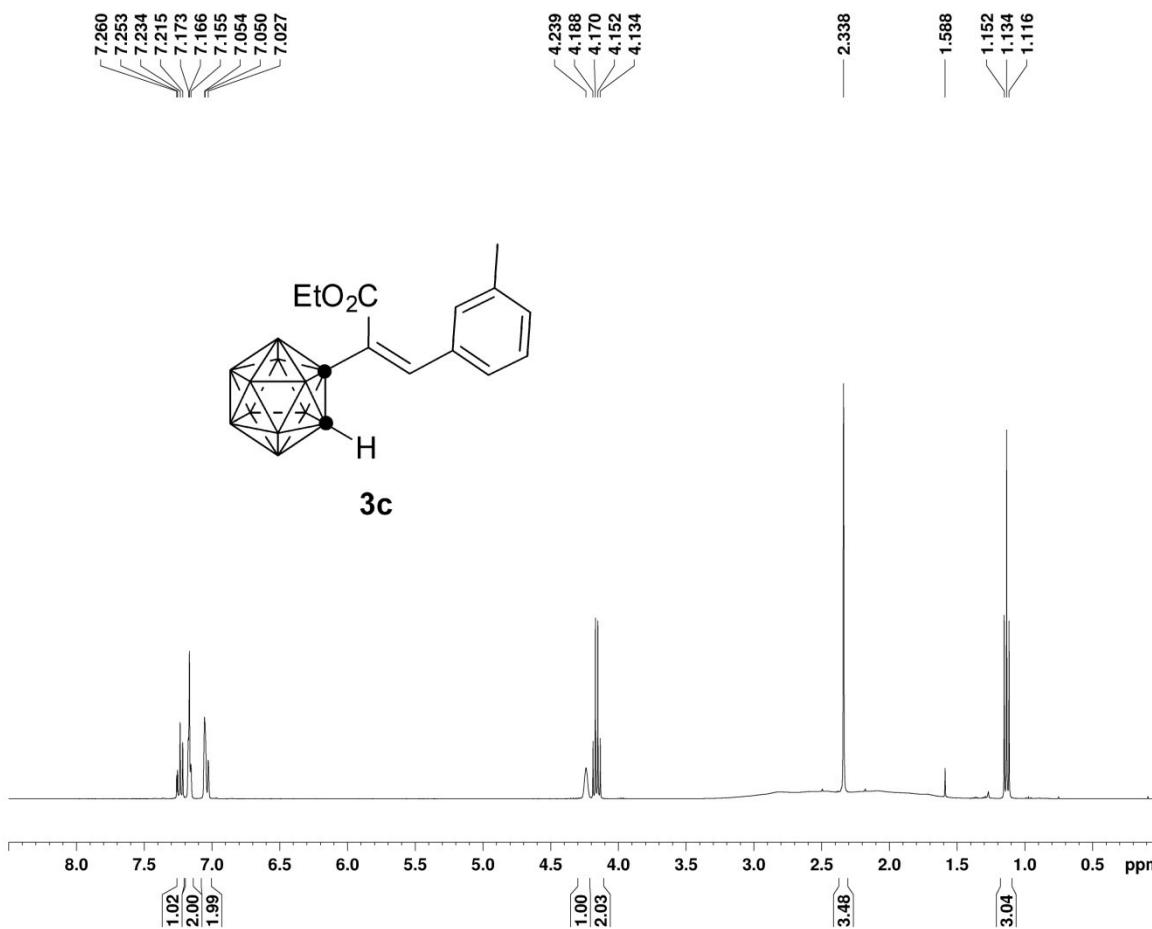










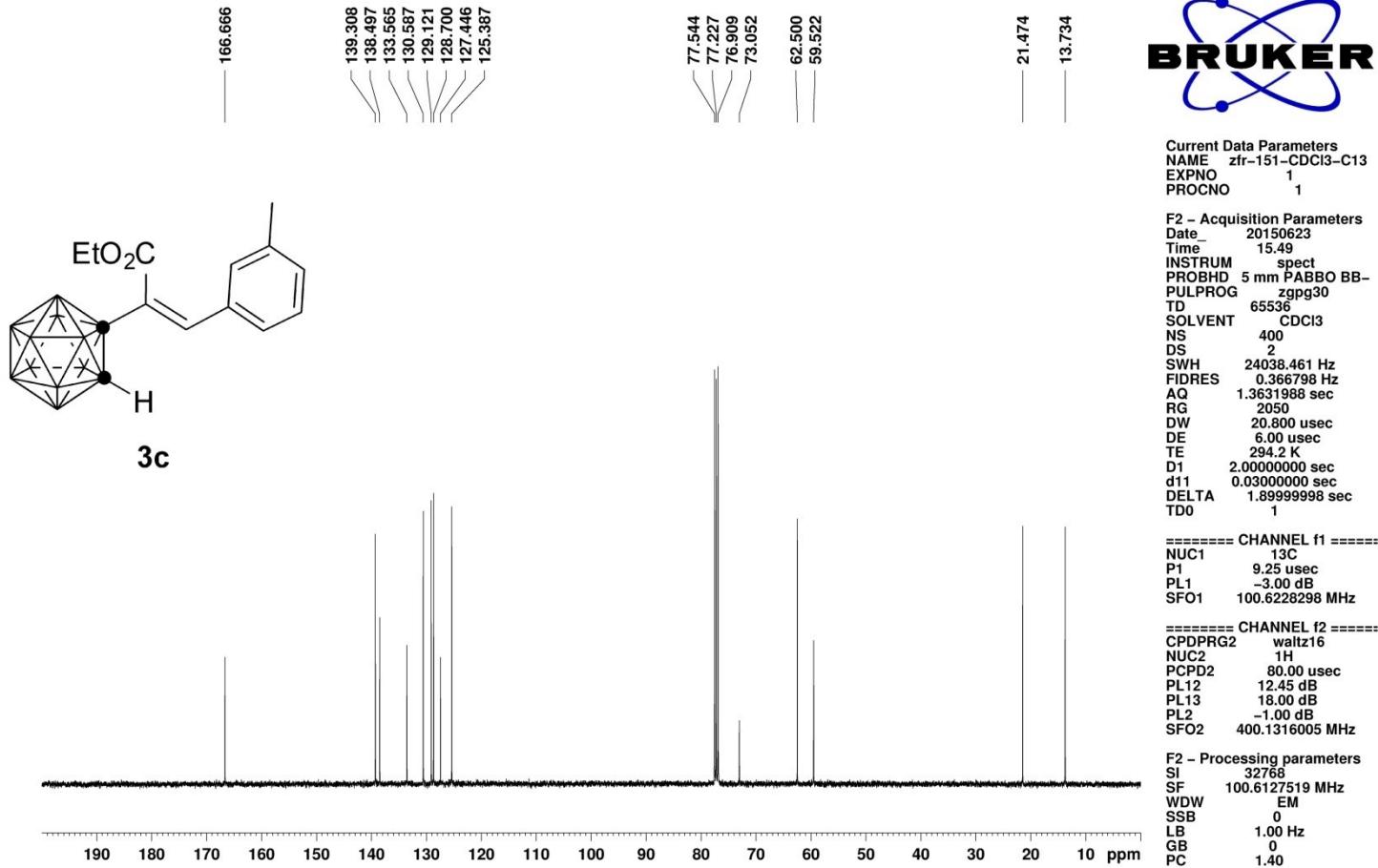


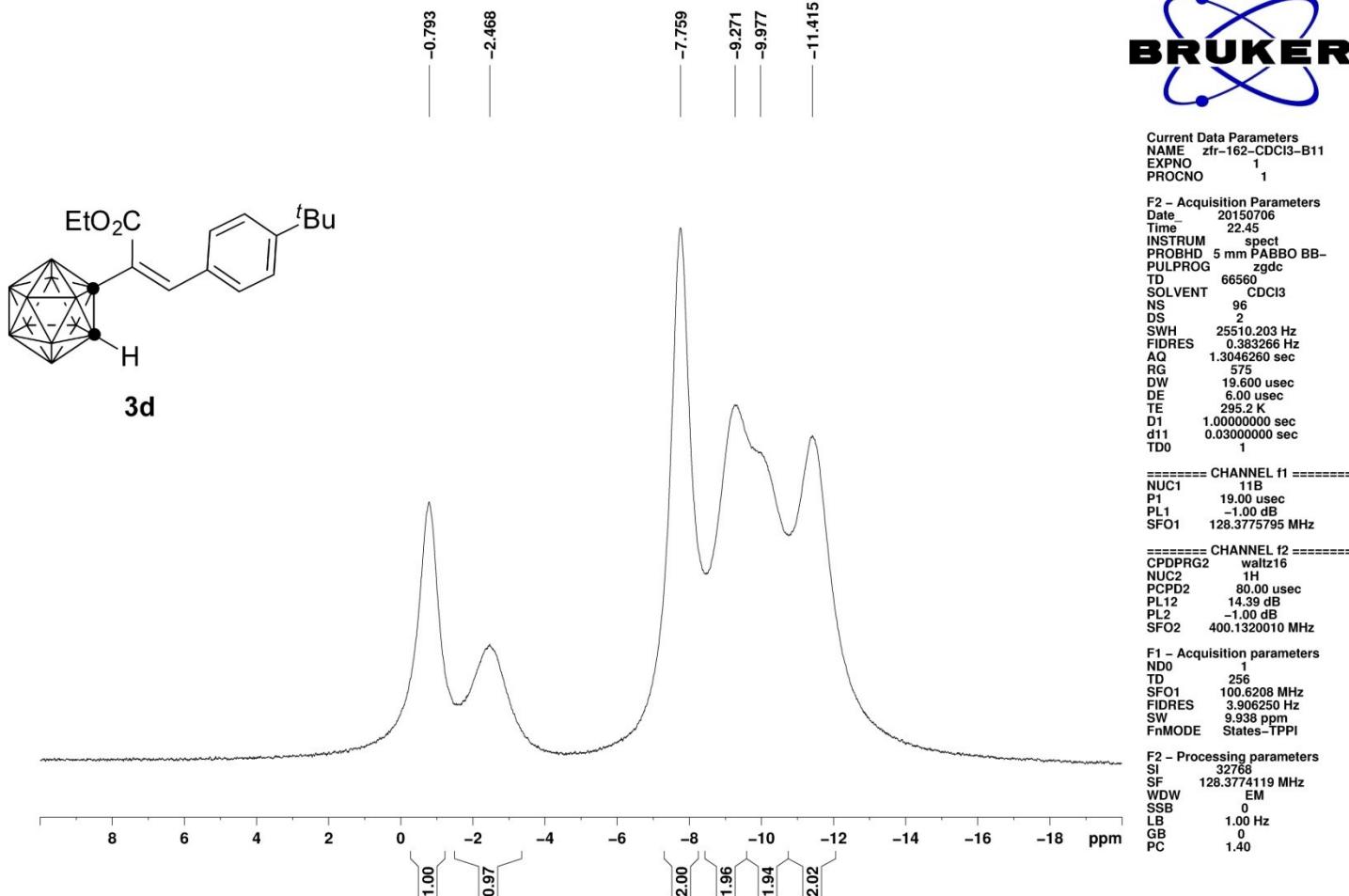
Current Data Parameters
NAME zfr-151CDCl3-H1
EXPNO 1
PROCNO 1

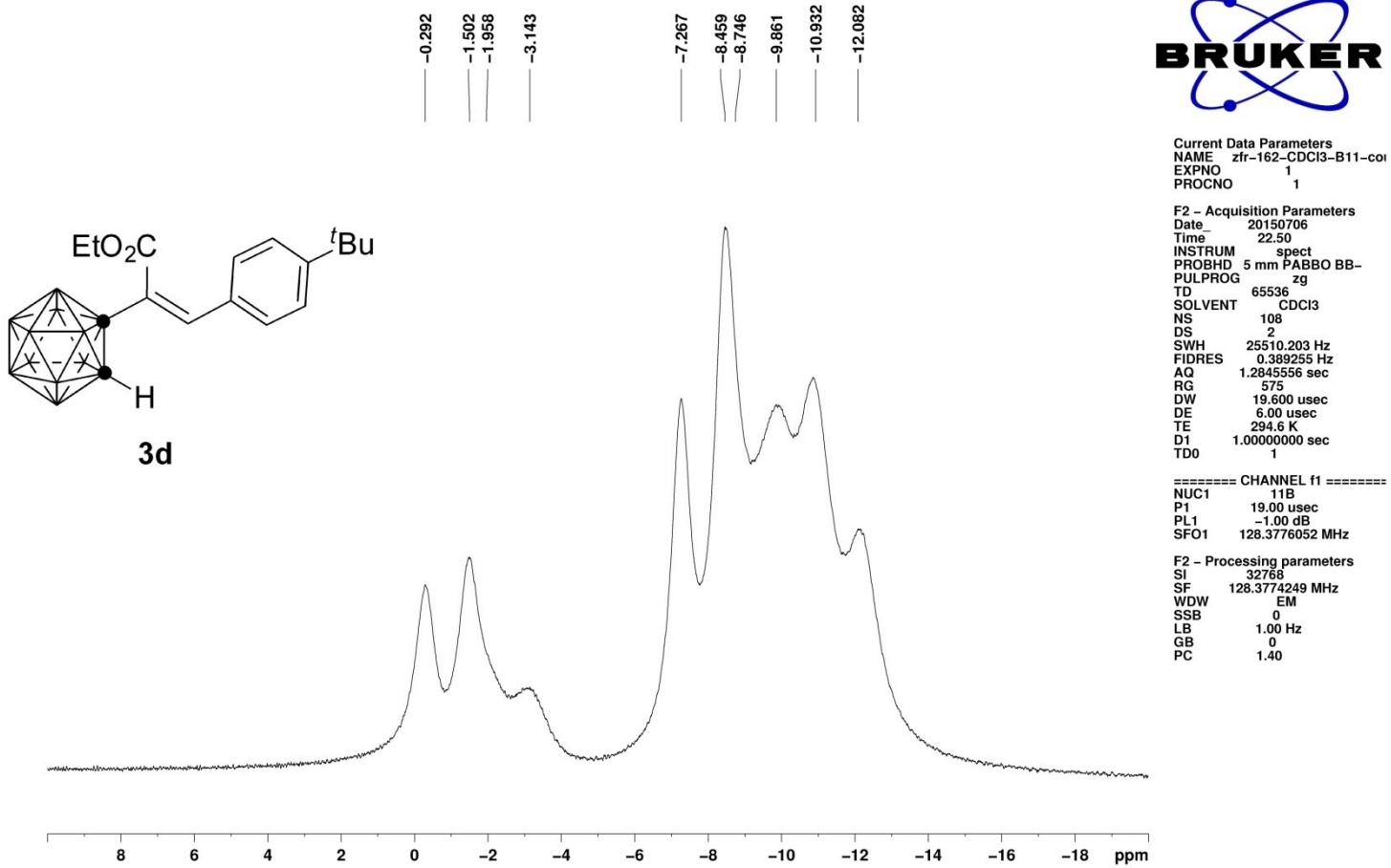
F2 – Acquisition Parameters
Date_ 20150623
Time 15.34
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9846387 sec
RG 64
DW 60.800 usec
DE 6.00 usec
TE 293.4 K
D1 1.0000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 13.60 usec
PL1 -1.00 dB
SFO1 400.1324710 MHz

F2 – Processing parameters
SI 32768
SF 400.1300053 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00







7.370
7.365
7.353
7.348
7.344
7.260
7.184
7.163
7.138

4.213
4.195
4.177
4.160

1.581
1.305
1.164
1.146
1.128

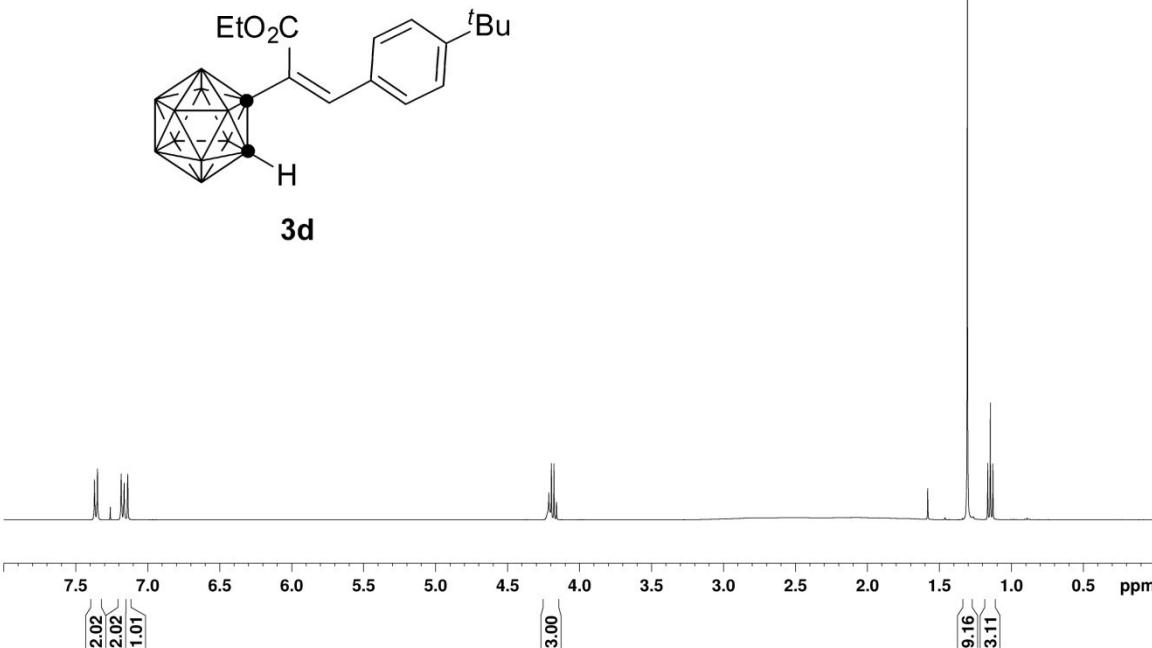


Current Data Parameters
NAME zfr-162-CDCl3-H1
EXPNO 1
PROCNO 1

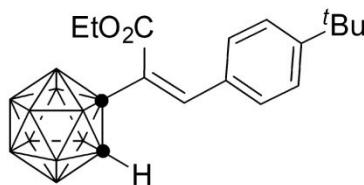
F2 – Acquisition Parameters
Date 20150706
Time 22.39
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9846387 sec
RG 128
DW 60.800 usec
DE 6.00 usec
TE 294.4 K
D1 1.0000000 sec
TD0 1

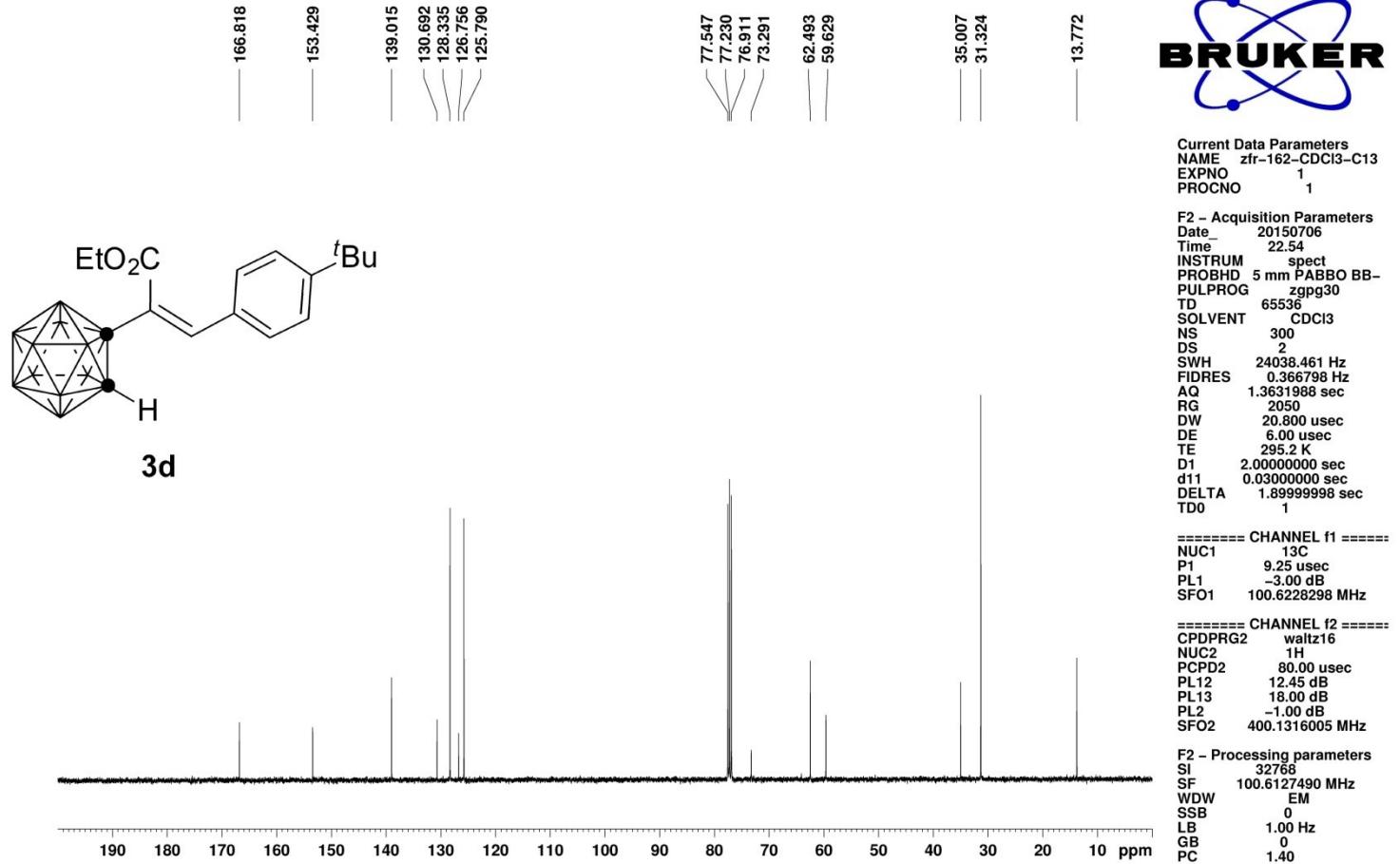
===== CHANNEL f1 =====
NUC1 1H
P1 13.60 usec
PL1 -1.00 dB
SFO1 400.1324710 MHz

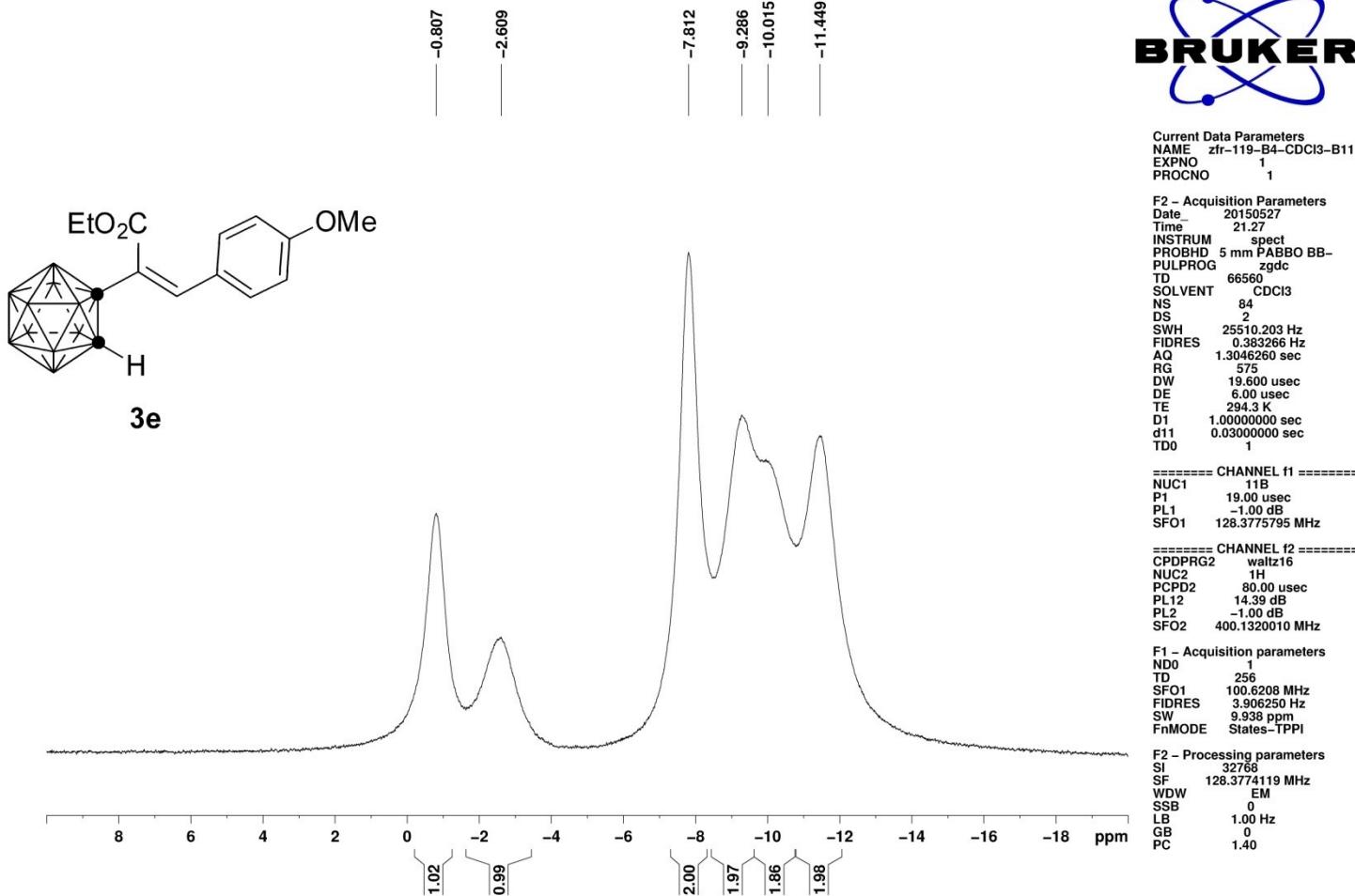
F2 – Processing parameters
SI 32768
SF 400.1300053 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

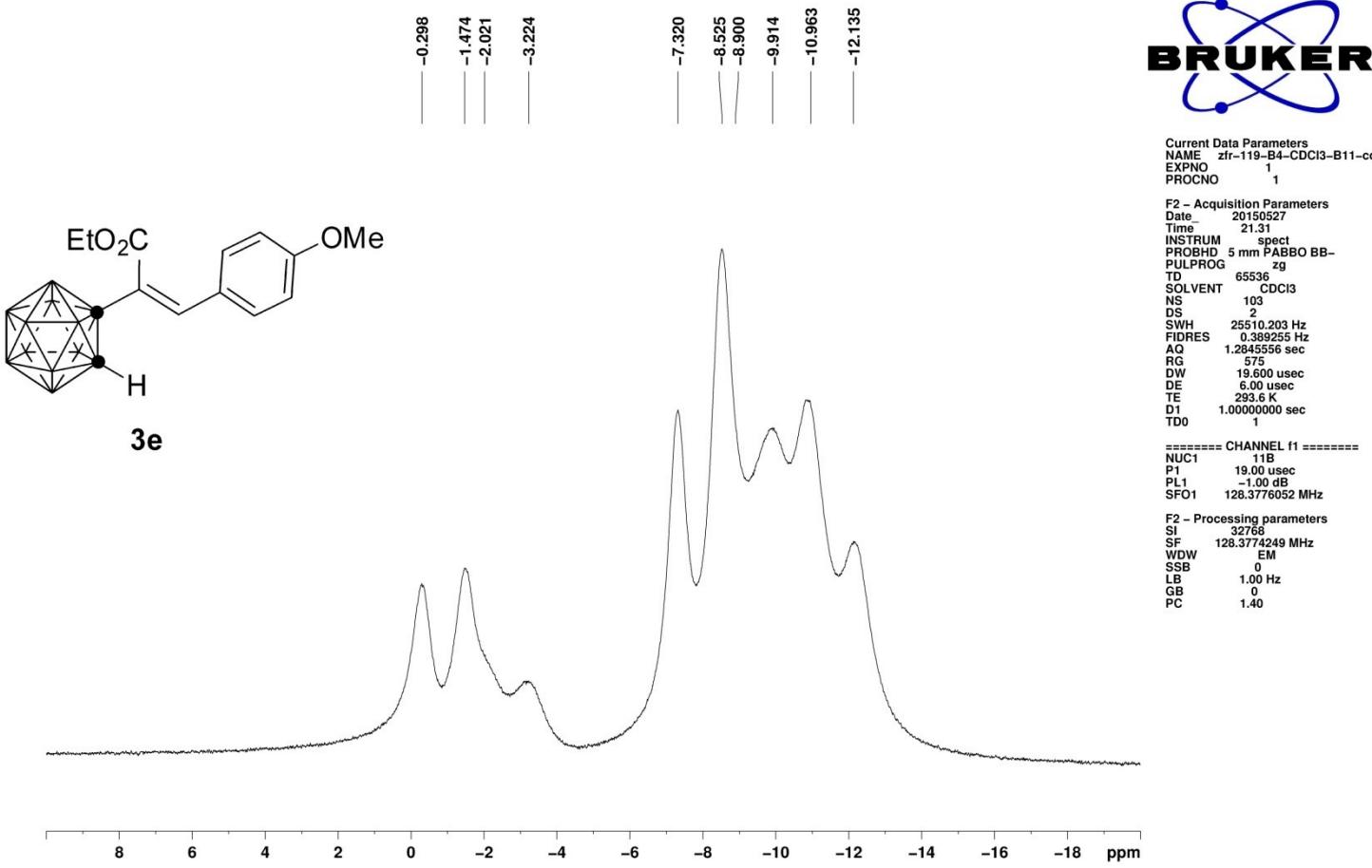


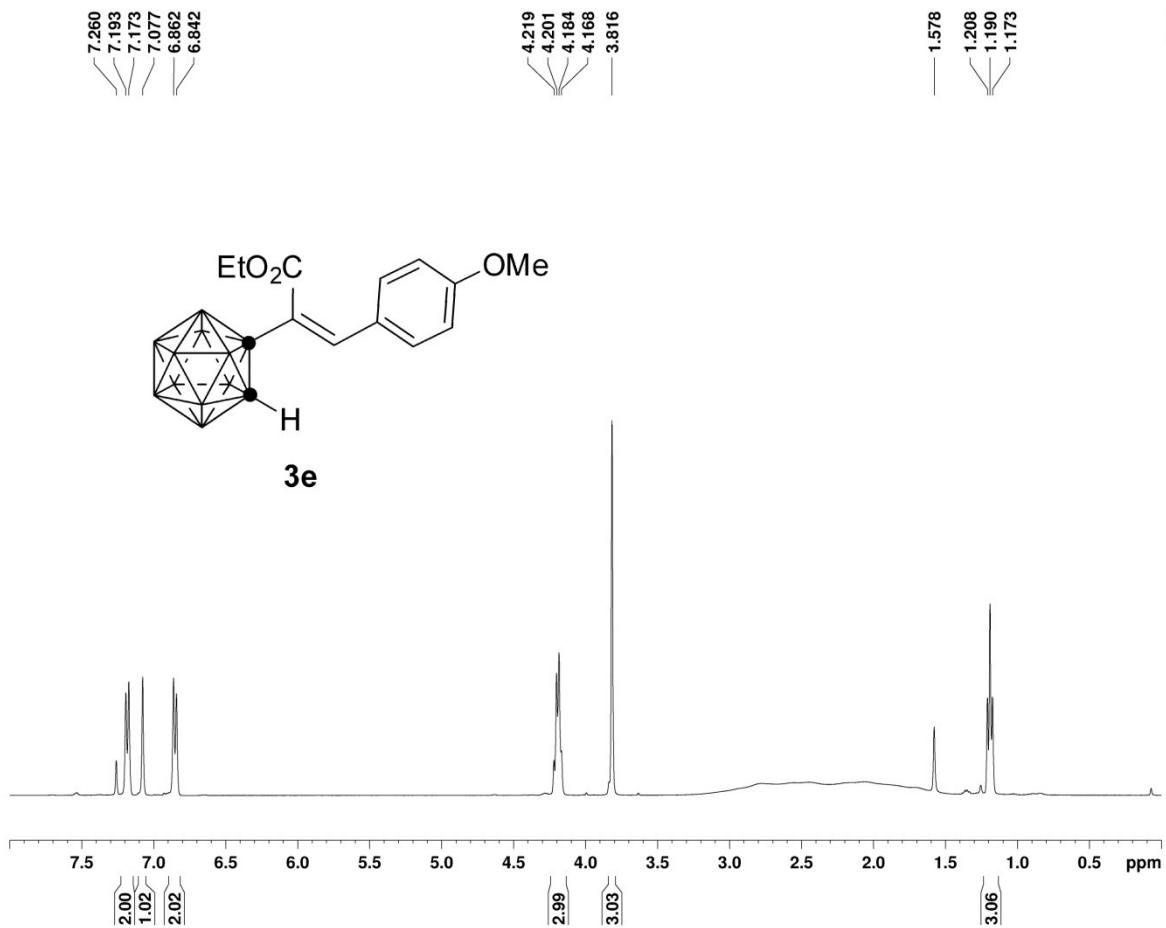
3d

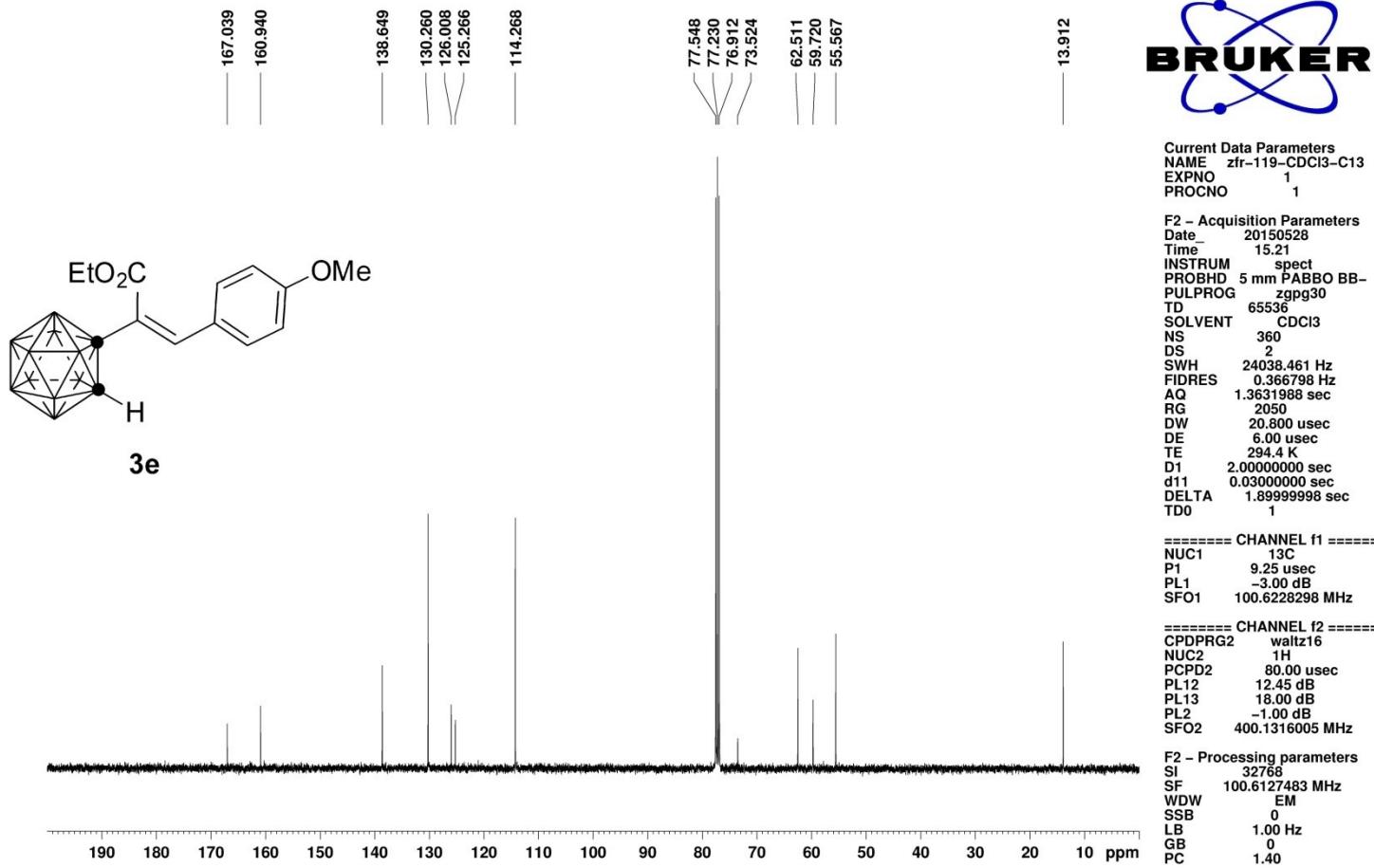


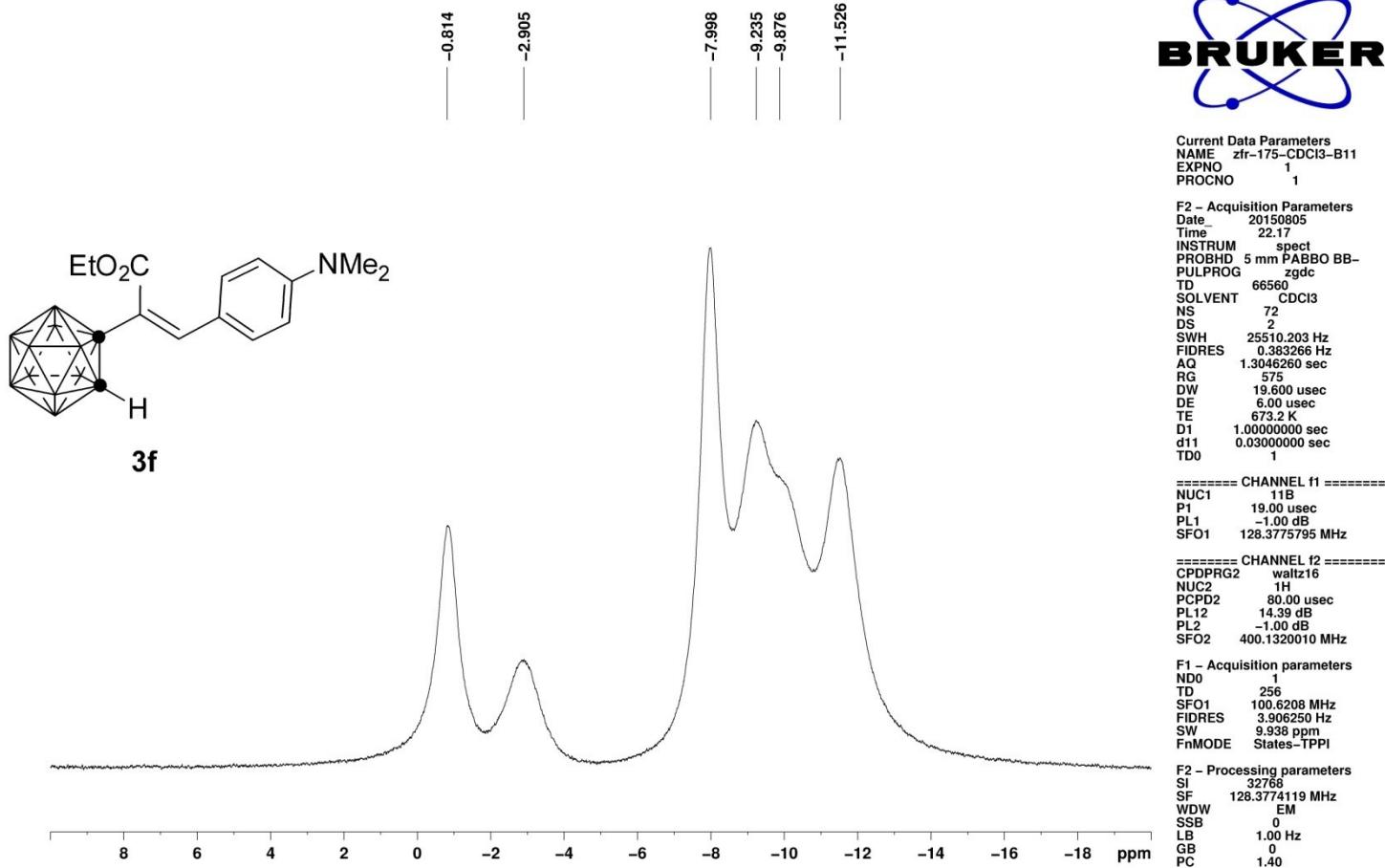












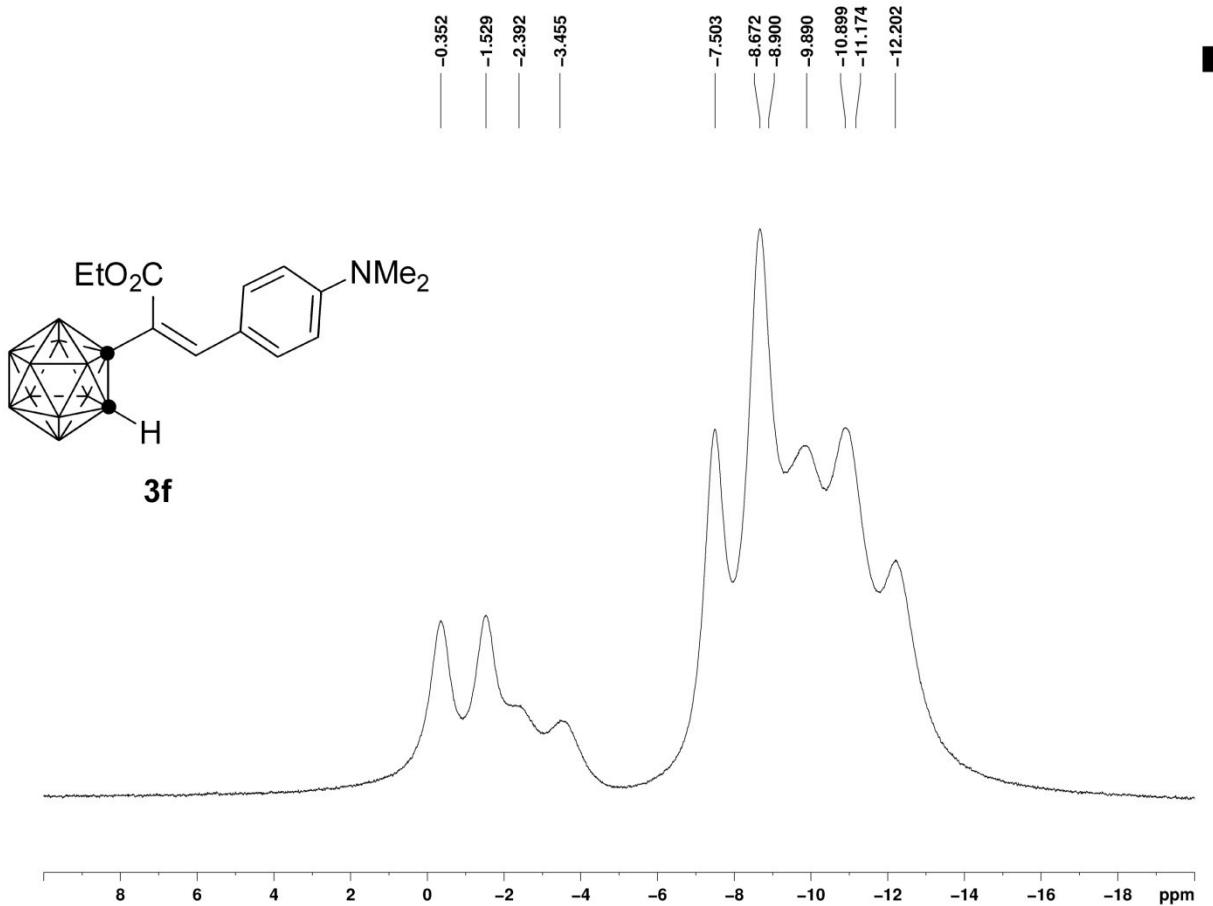


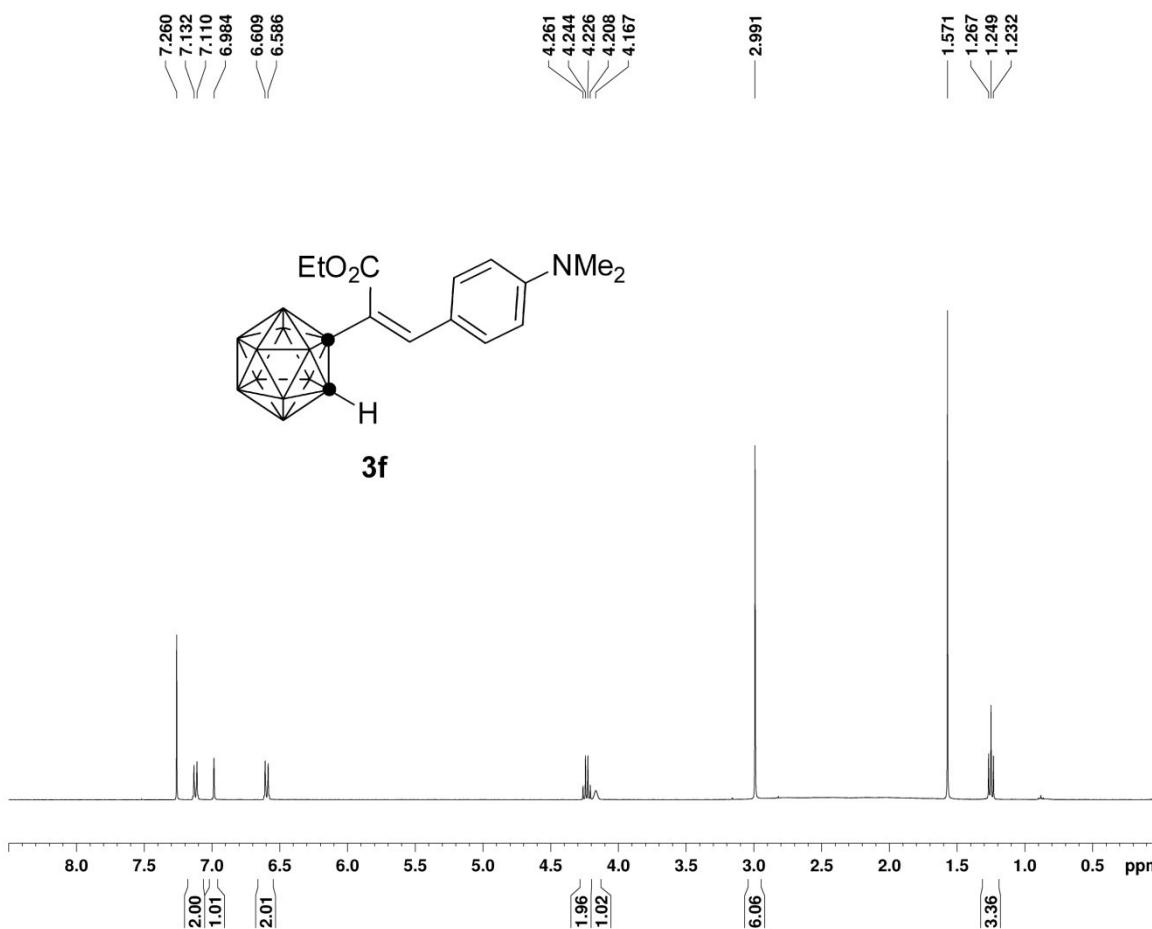
Current Data Parameters
NAME zfr-175-CDCl3-B11-co1
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date 20150805
Time 22:24
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg
TD 65536
SOLVENT CDCl3
NS 160
DS 2
SWH 25510.203 Hz
FIDRES 0.389255 Hz
AQ 1.2845556 sec
RG 575
DW 19.600 usec
DE 6.00 usec
TE 673.2 K
D1 1.0000000 sec
TD0 1

===== CHANNEL f1 ======
NUC1 11B
P1 19.00 usec
PL1 -1.00 dB
SFO1 128.3776052 MHz

F2 - Processing parameters
SI 32768
SF 128.3774249 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



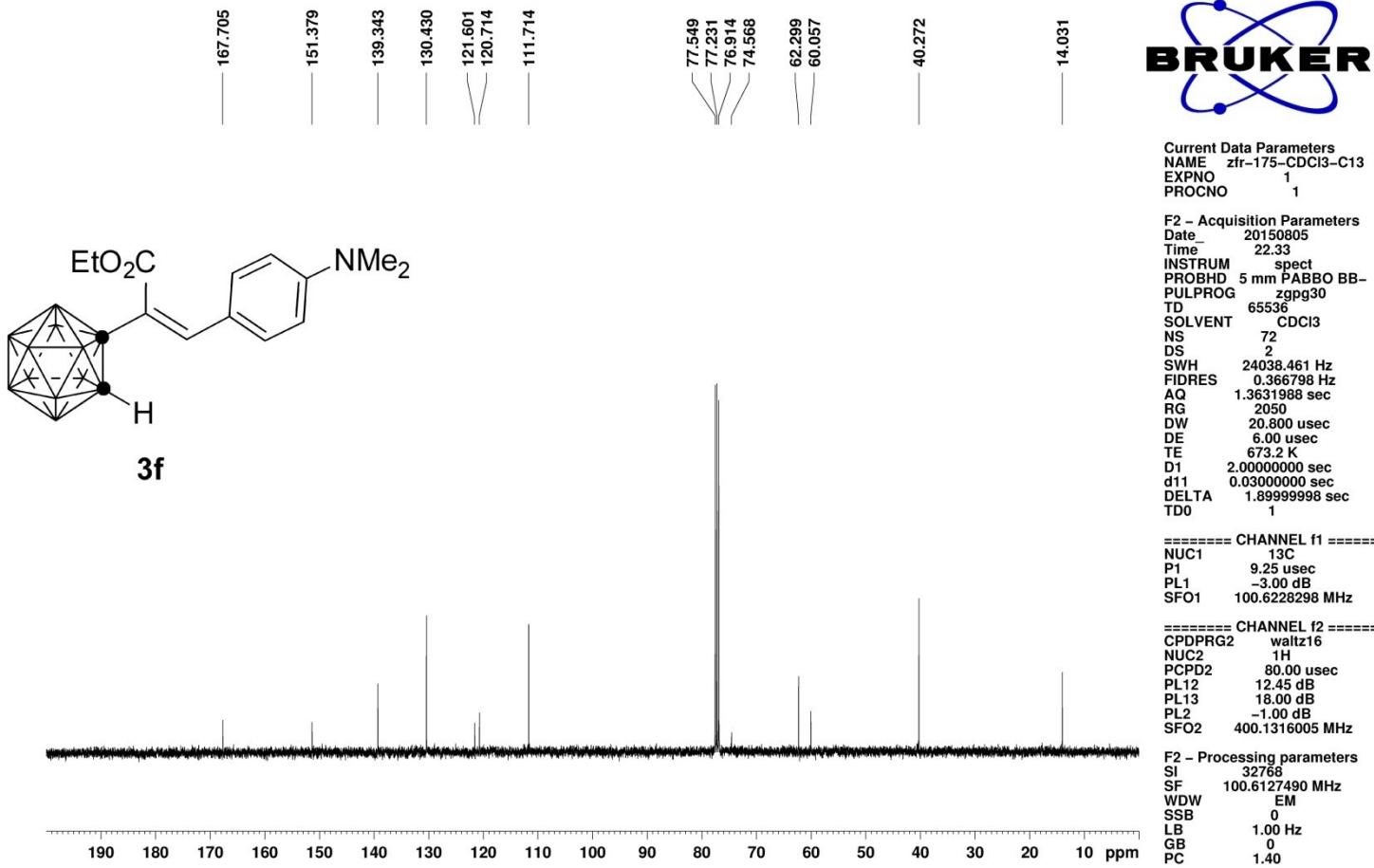


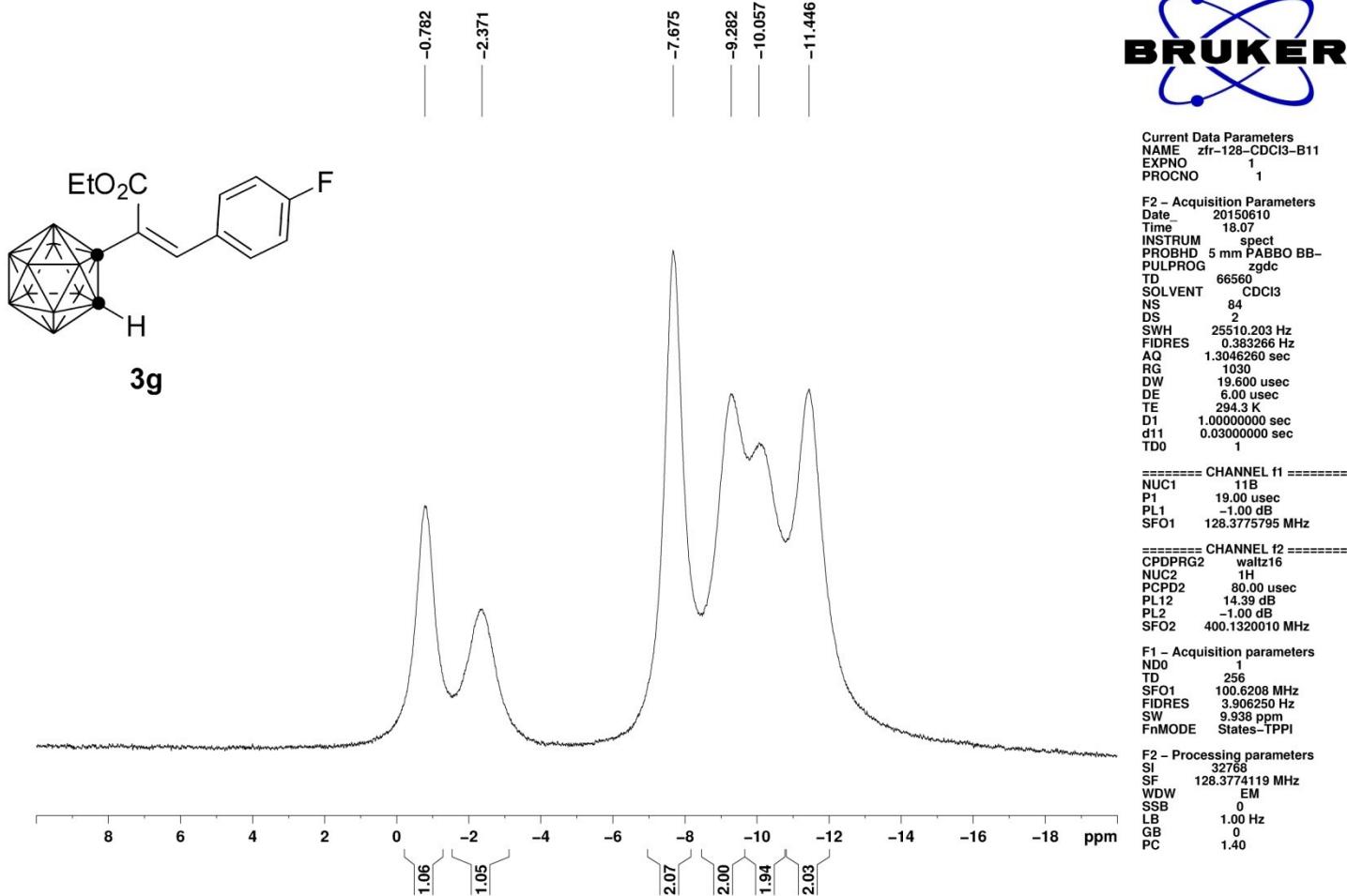
Current Data Parameters
 NAME zfr-192-CDCl3-H1
 EXPNO 2
 PROCNO 1

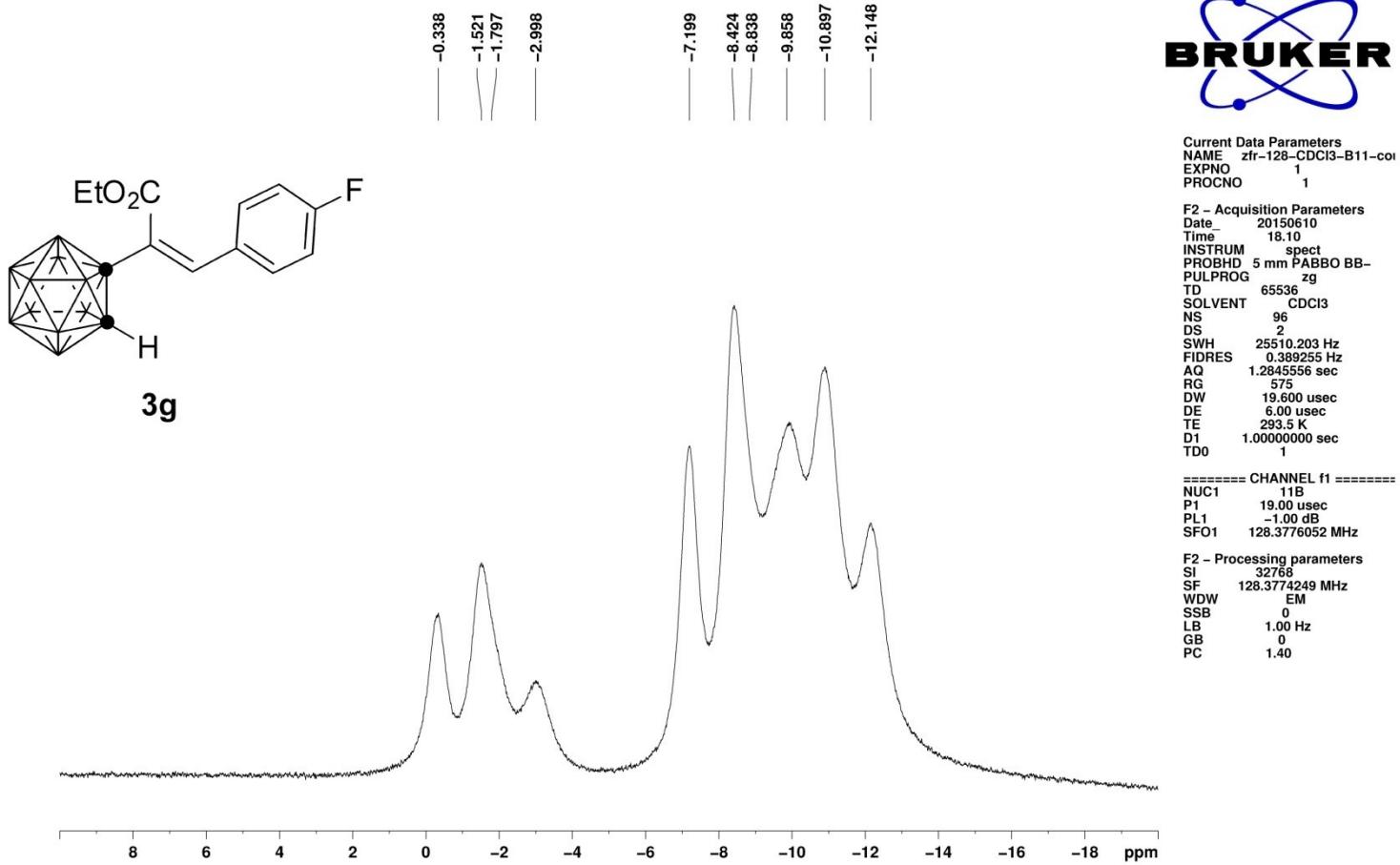
F2 - Acquisition Parameters
 Date 20150818
 Time 12.11
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 8223.685 Hz
 FIDRES 0.125483 Hz
 AQ 3.9846387 sec
 RG 575
 DW 60.800 usec
 DE 6.00 usec
 TE 294.0 K
 D1 1.0000000 sec
 TD0 1

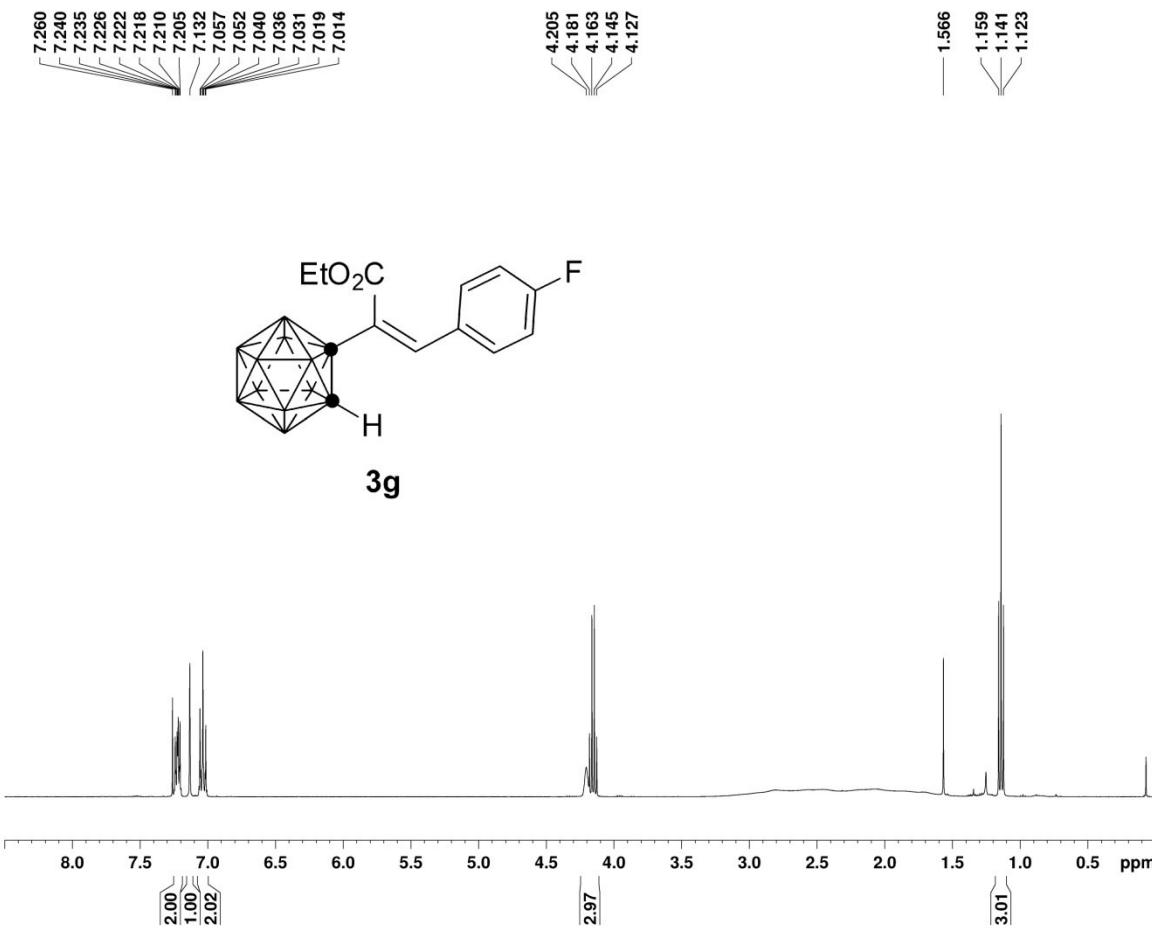
===== CHANNEL f1 =====
 NUC1 1H
 P1 13.60 usec
 PL1 -1.00 dB
 SFO1 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300053 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00







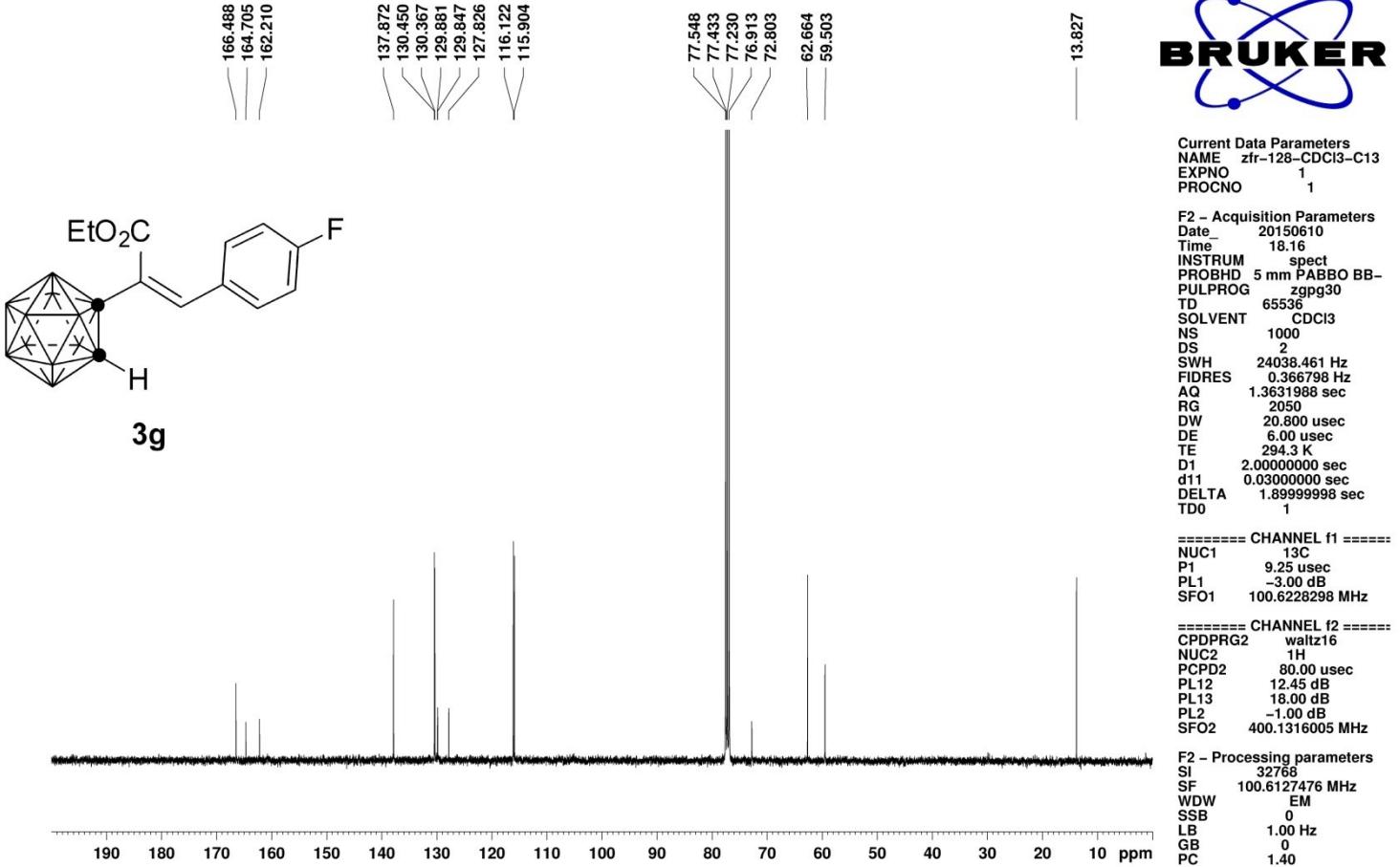


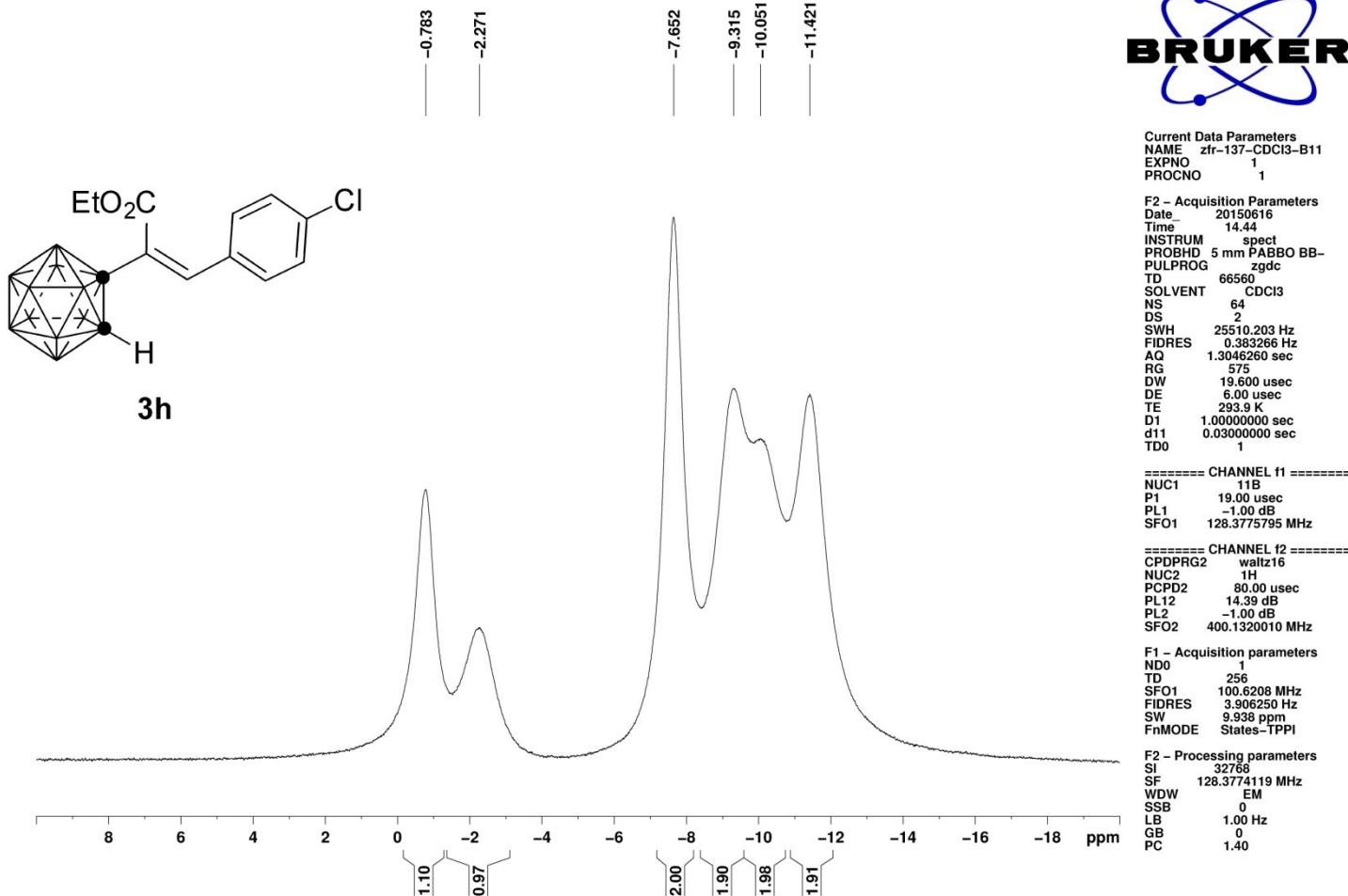
Current Data Parameters
 NAME zfr-128-CDCl3-H1
 EXPNO 2
 PROCNO 1

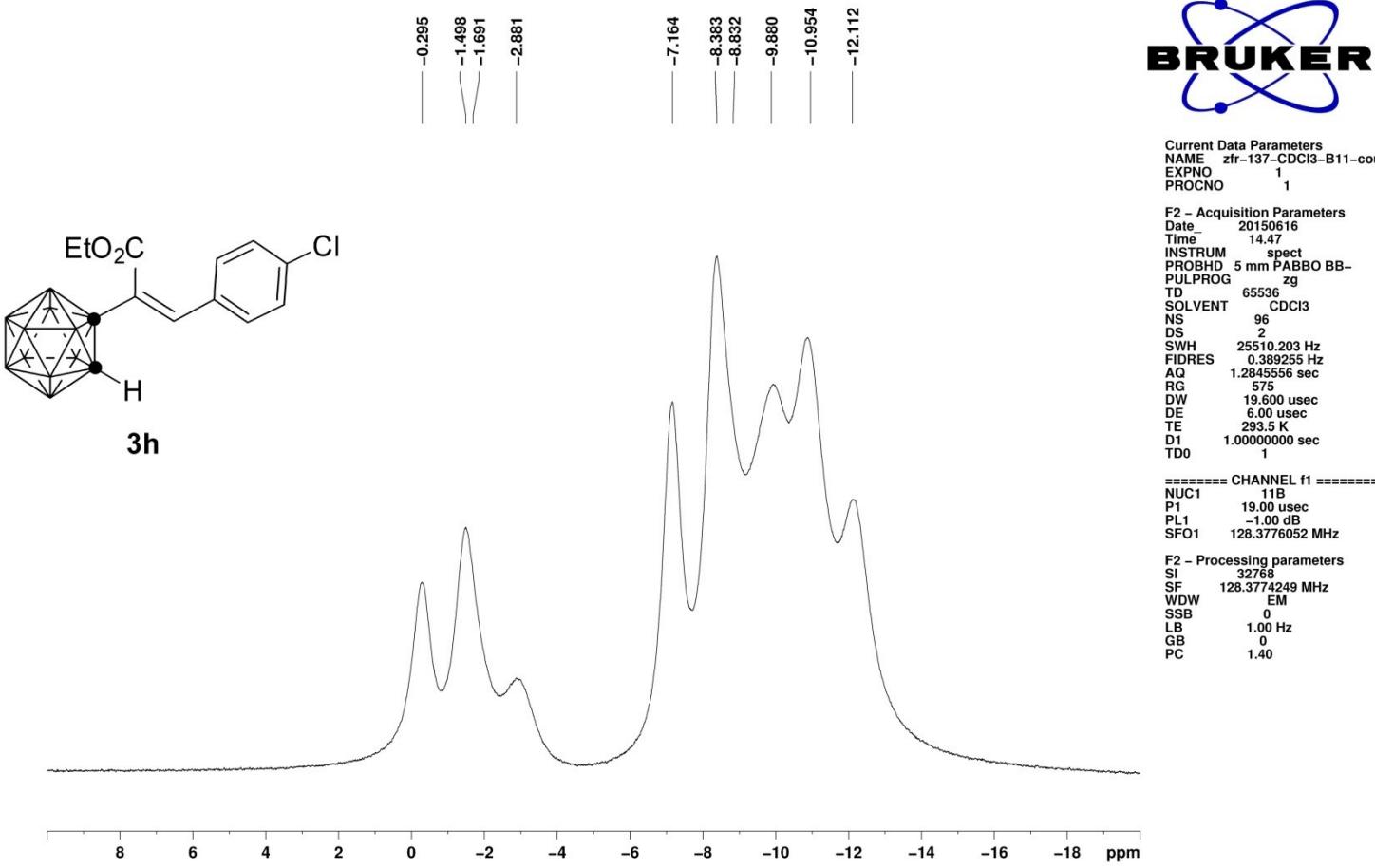
F2 – Acquisition Parameters
 Date 20150610
 Time 18.02
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 8223.685 Hz
 FIDRES 0.125483 Hz
 AQ 3.9846387 sec
 RG 287
 DW 60.800 usec
 DE 6.00 usec
 TE 293.5 K
 D1 1.0000000 sec
 T00 1

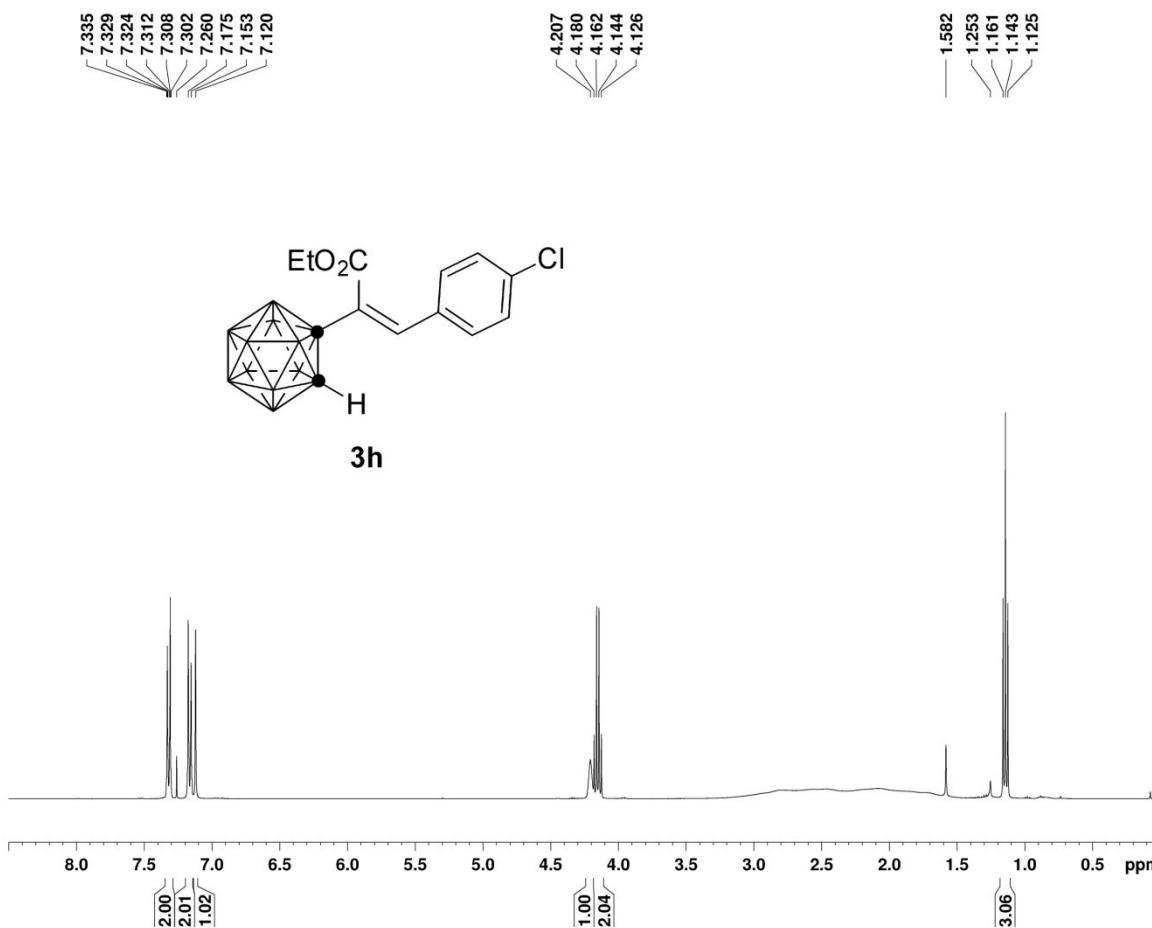
===== CHANNEL f1 =====
 NUC1 1H
 P1 13.60 usec
 PL1 -1.00 dB
 SFO1 400.1324710 MHz

F2 – Processing parameters
 SI 32768
 SF 400.1300053 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00







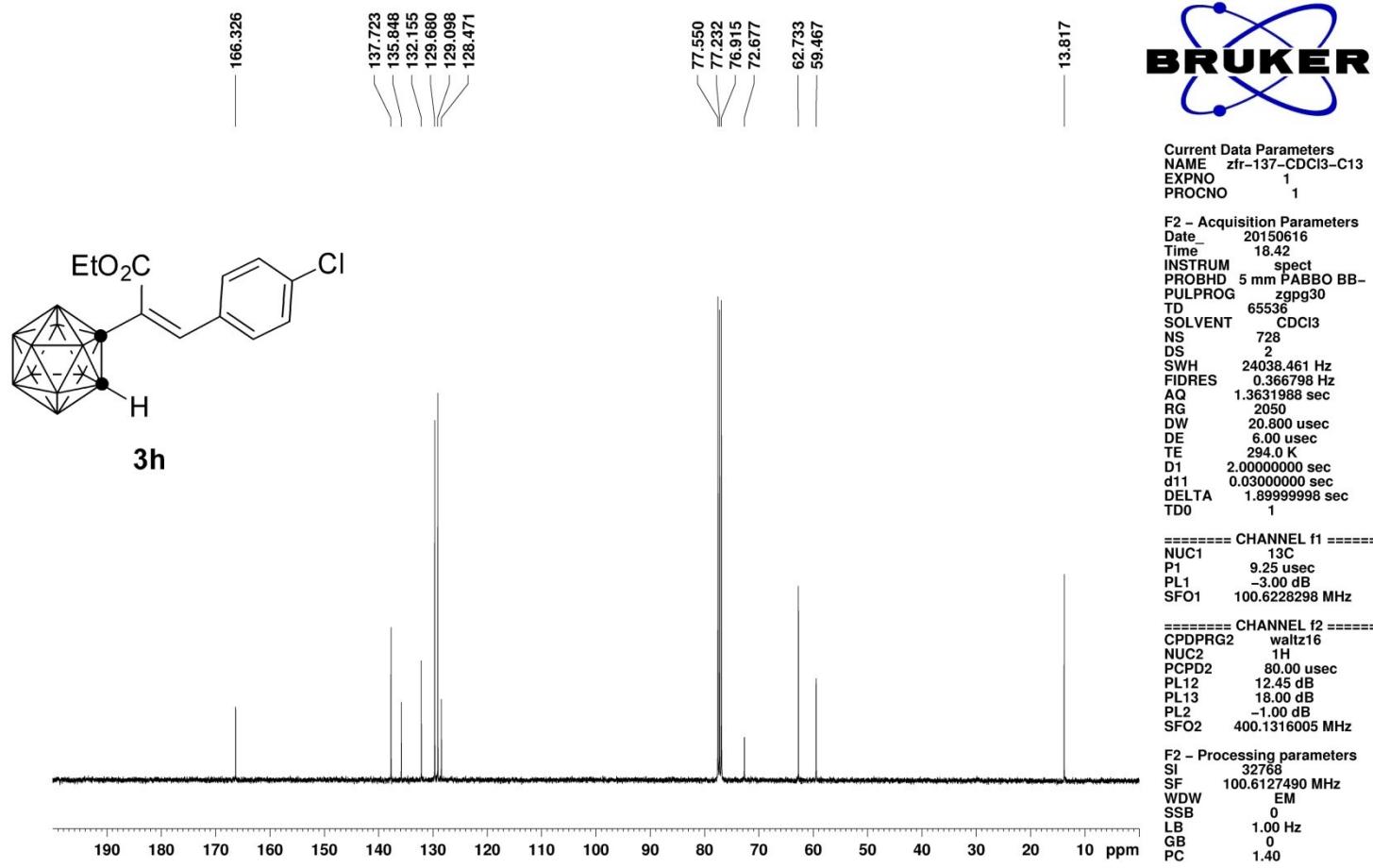


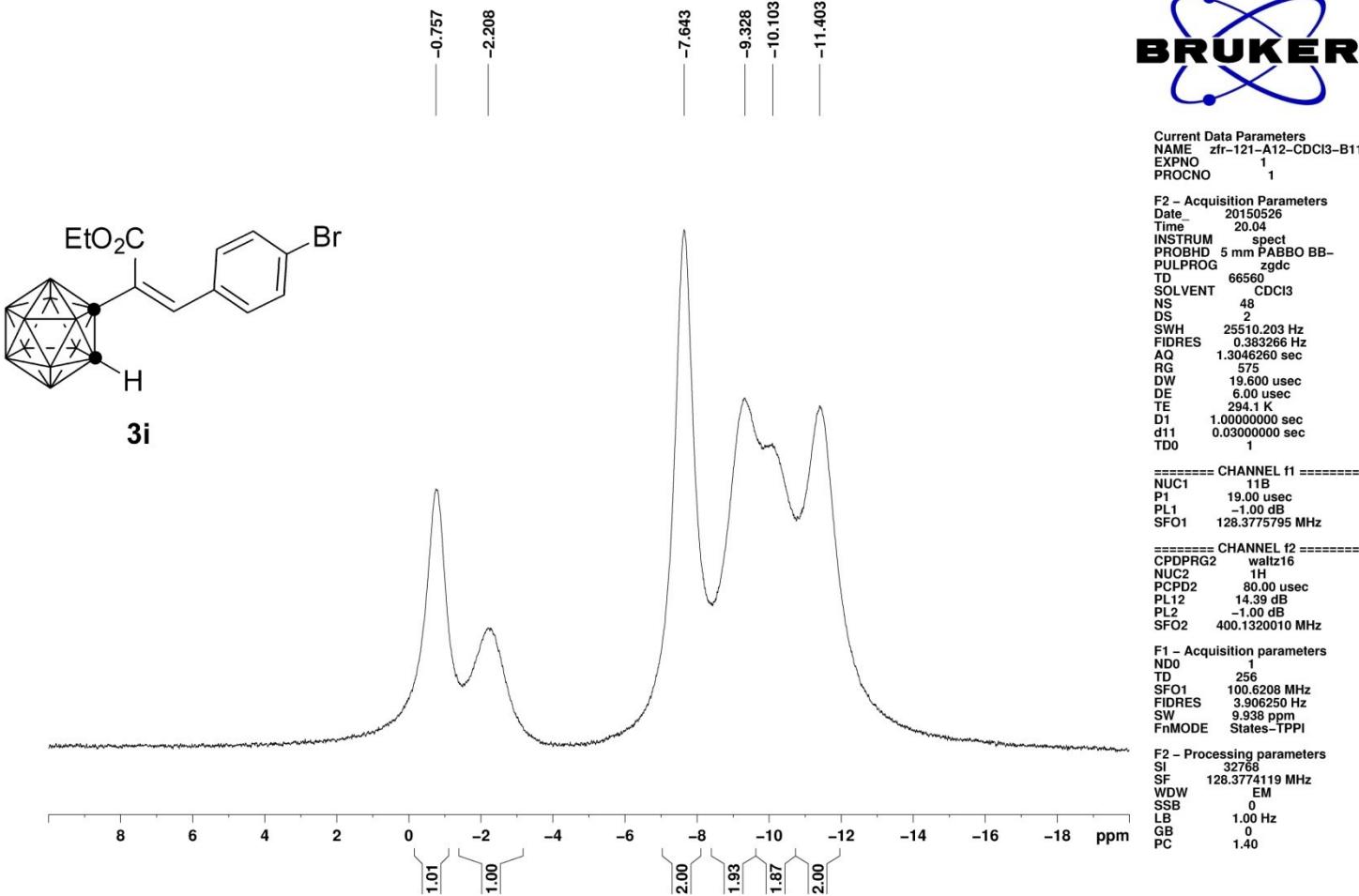
Current Data Parameters
NAME zfr-137-CDCl₃-H1
EXPNO 3
PROCNO 1

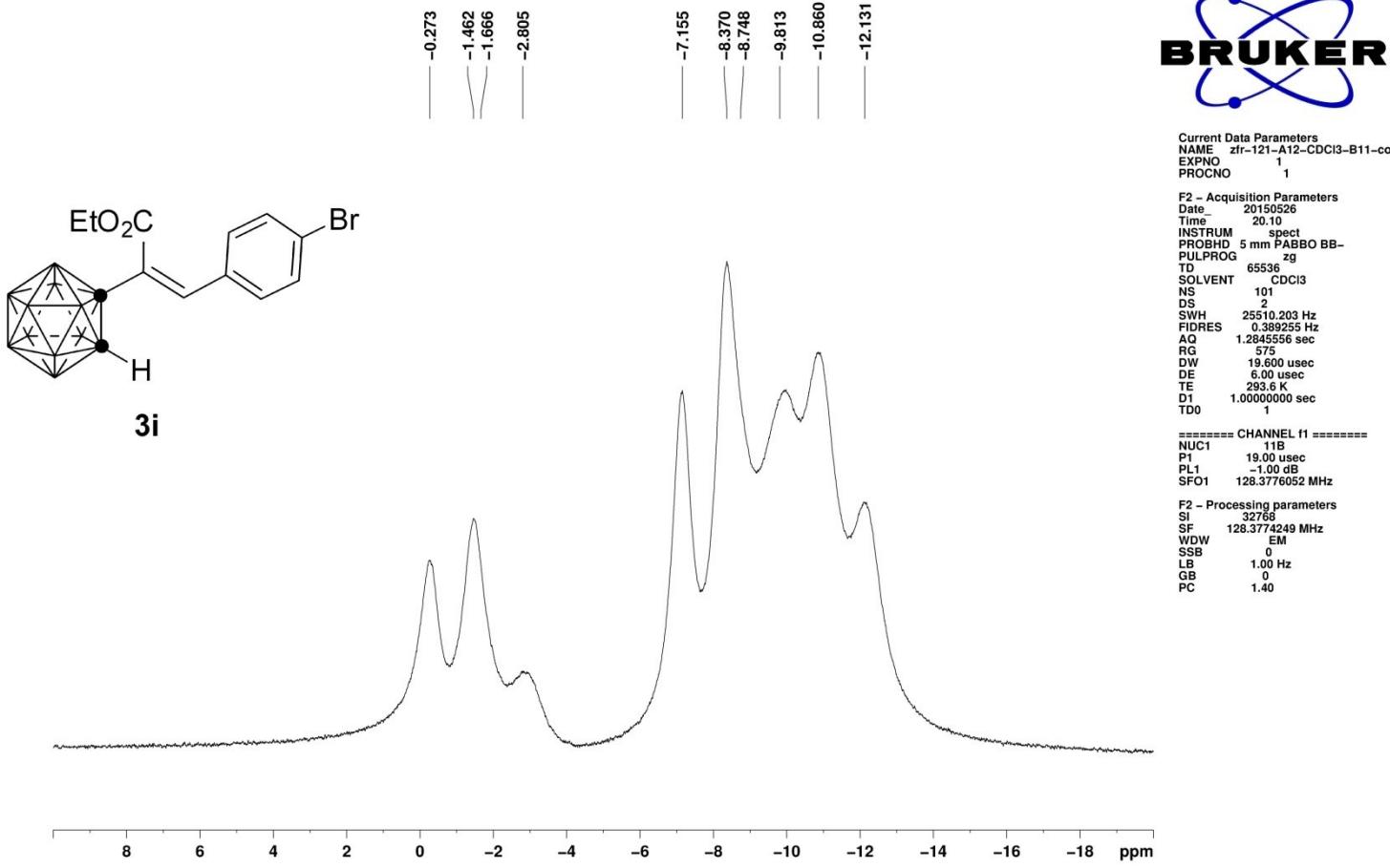
F2 – Acquisition Parameters
Date_ 20150616
Time 18.37
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl₃
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9846387 sec
RG 161
DW 60.800 usec
DE 6.00 usec
TE 293.3 K
D1 1.0000000 sec
TD0 1

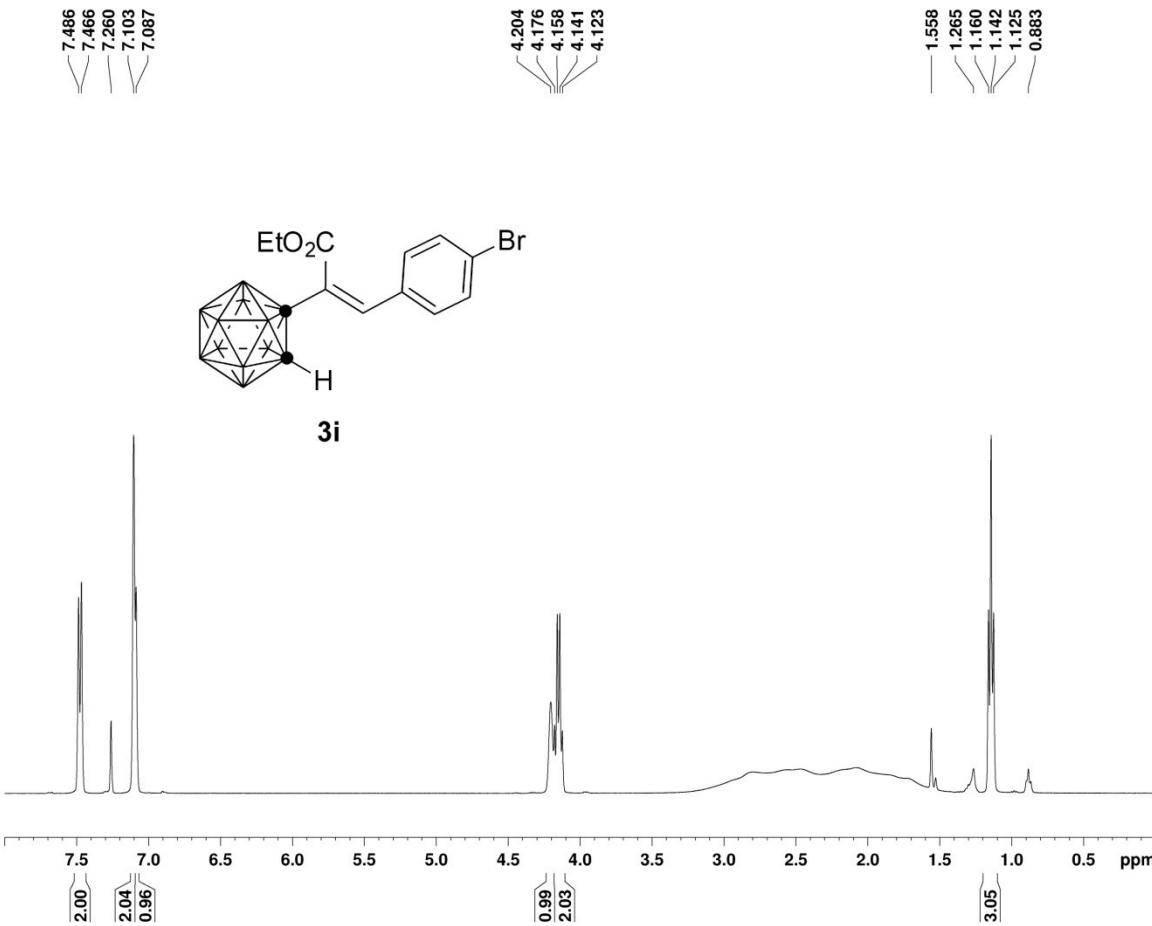
===== CHANNEL f1 =====
NUC1 1H
P1 13.60 usec
PL1 -1.00 dB
SFO1 400.1324710 MHz

F2 – Processing parameters
SI 32768
SF 400.1300053 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00







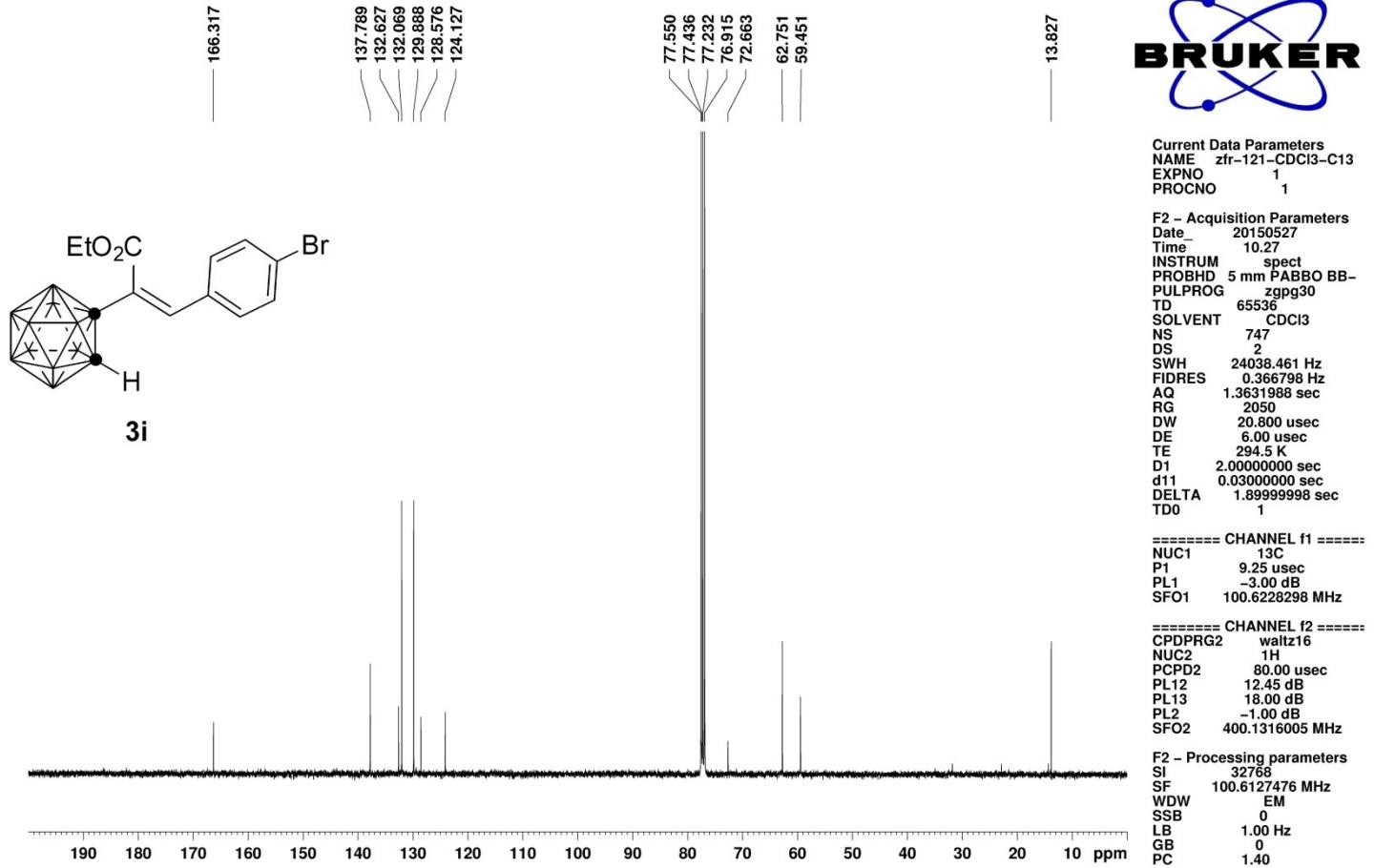


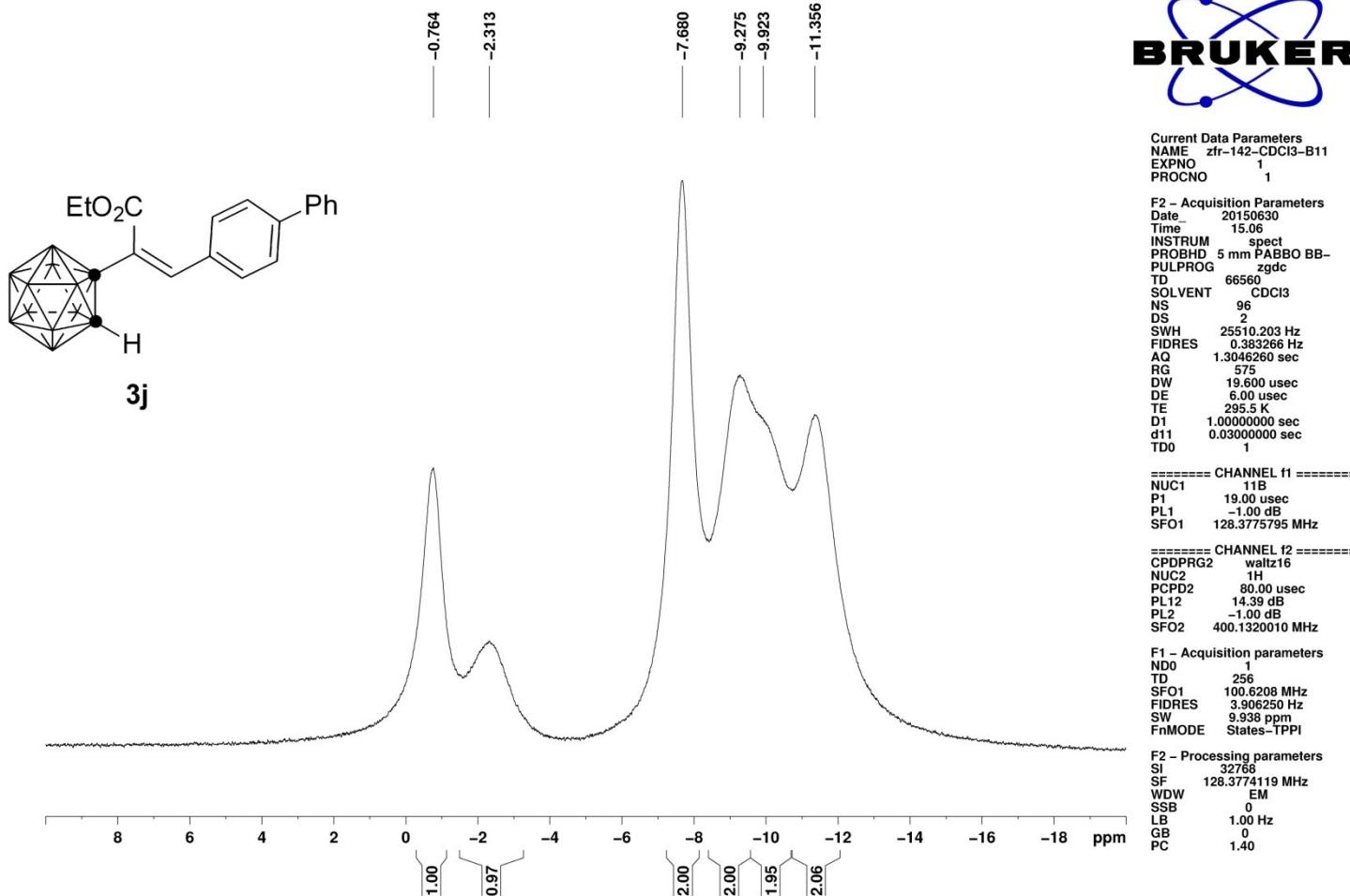
Current Data Parameters
 NAME zfr-121-CDCl3-H1
 EXPNO 1
 PROCNO 1

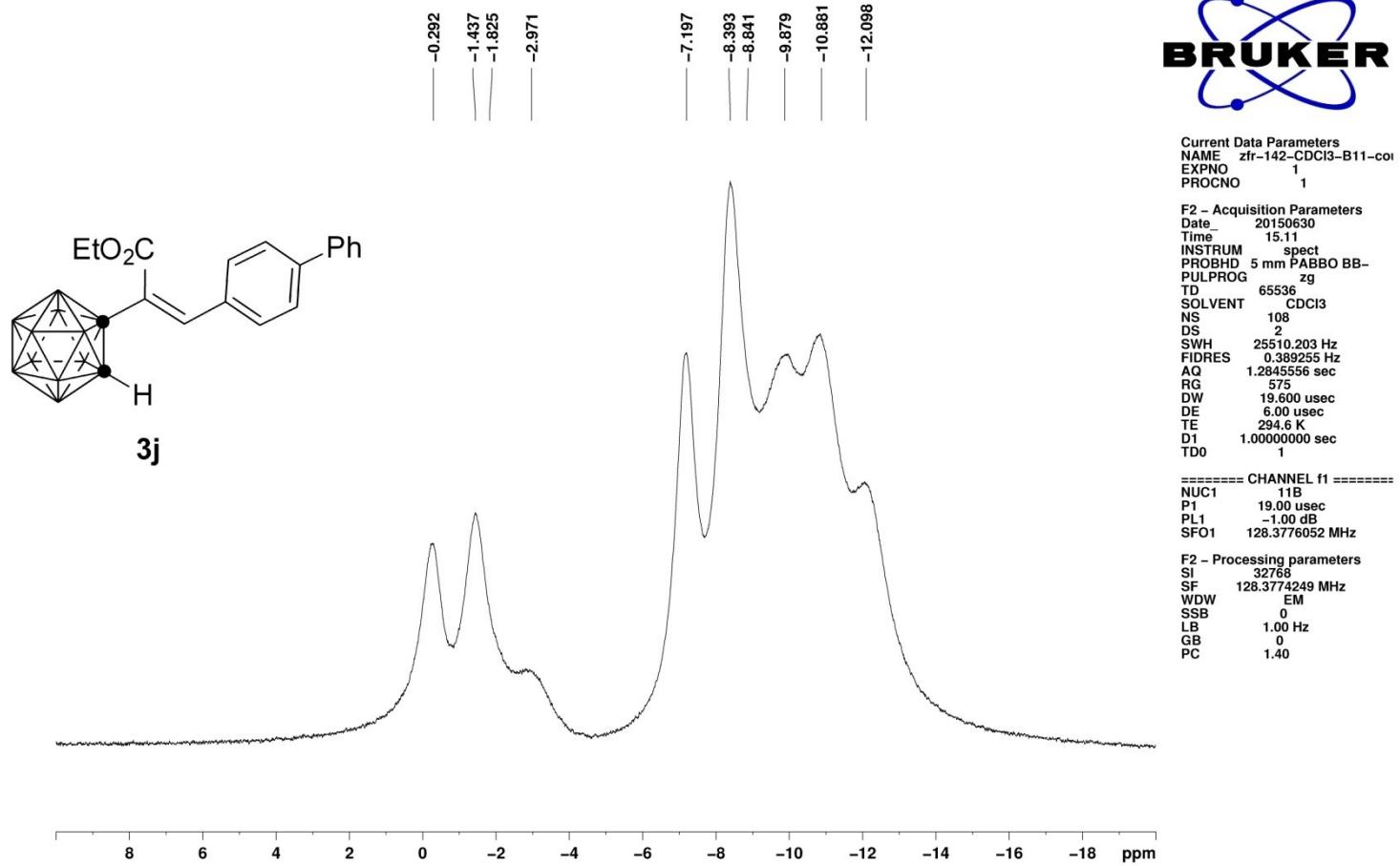
F2 – Acquisition Parameters
 Date 20150527
 Time 10.22
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 8223.685 Hz
 FIDRES 0.125483 Hz
 AQ 3.9846387 sec
 RG 256
 DW 60.800 usec
 DE 6.00 usec
 TE 293.8 K
 D1 1.0000000 sec
 TDO 1

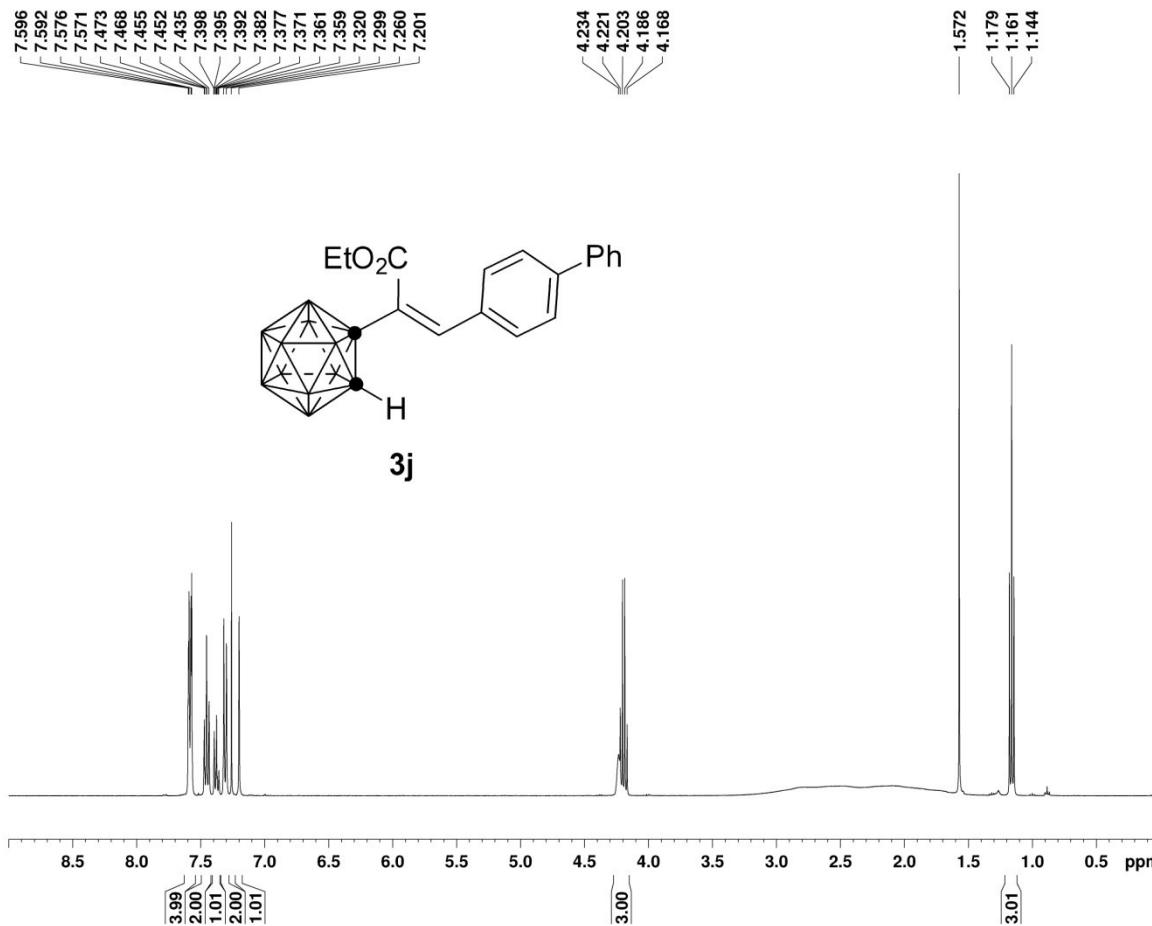
===== CHANNEL f1 =====
 NUC1 1H
 P1 13.60 usec
 PL1 -1.00 dB
 SFO1 400.1324710 MHz

F2 – Processing parameters
 SI 32768
 SF 400.1300051 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00







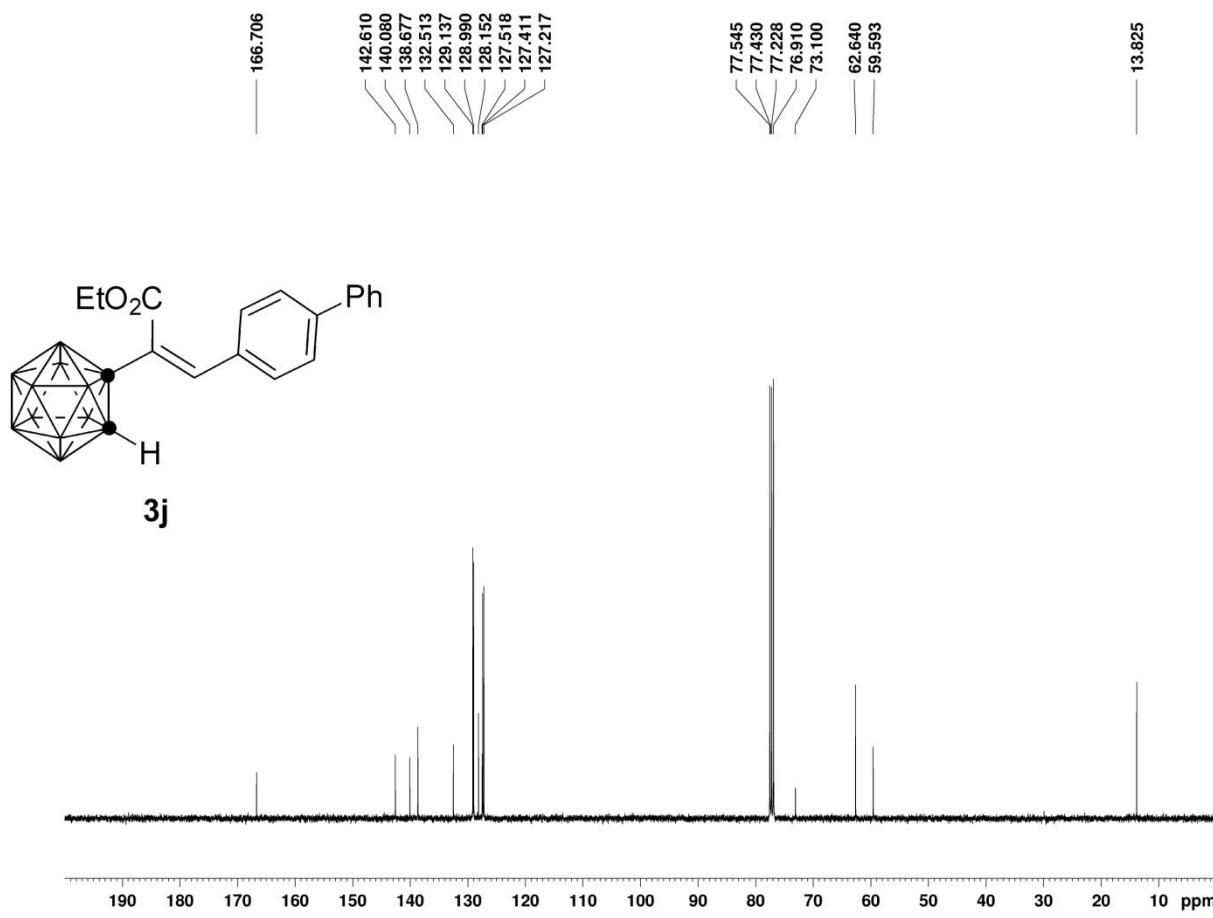


Current Data Parameters
 NAME zfr-142-CDCl3-H1
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date 20150630
 Time 20.24
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 8223.685 Hz
 FIDRES 0.125483 Hz
 AQ 3.9846387 sec
 RG 406
 DW 60.800 usec
 DE 6.00 usec
 TE 293.2 K
 D1 1.0000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 13.60 usec
 PL1 -1.00 dB
 SFO1 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300053 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



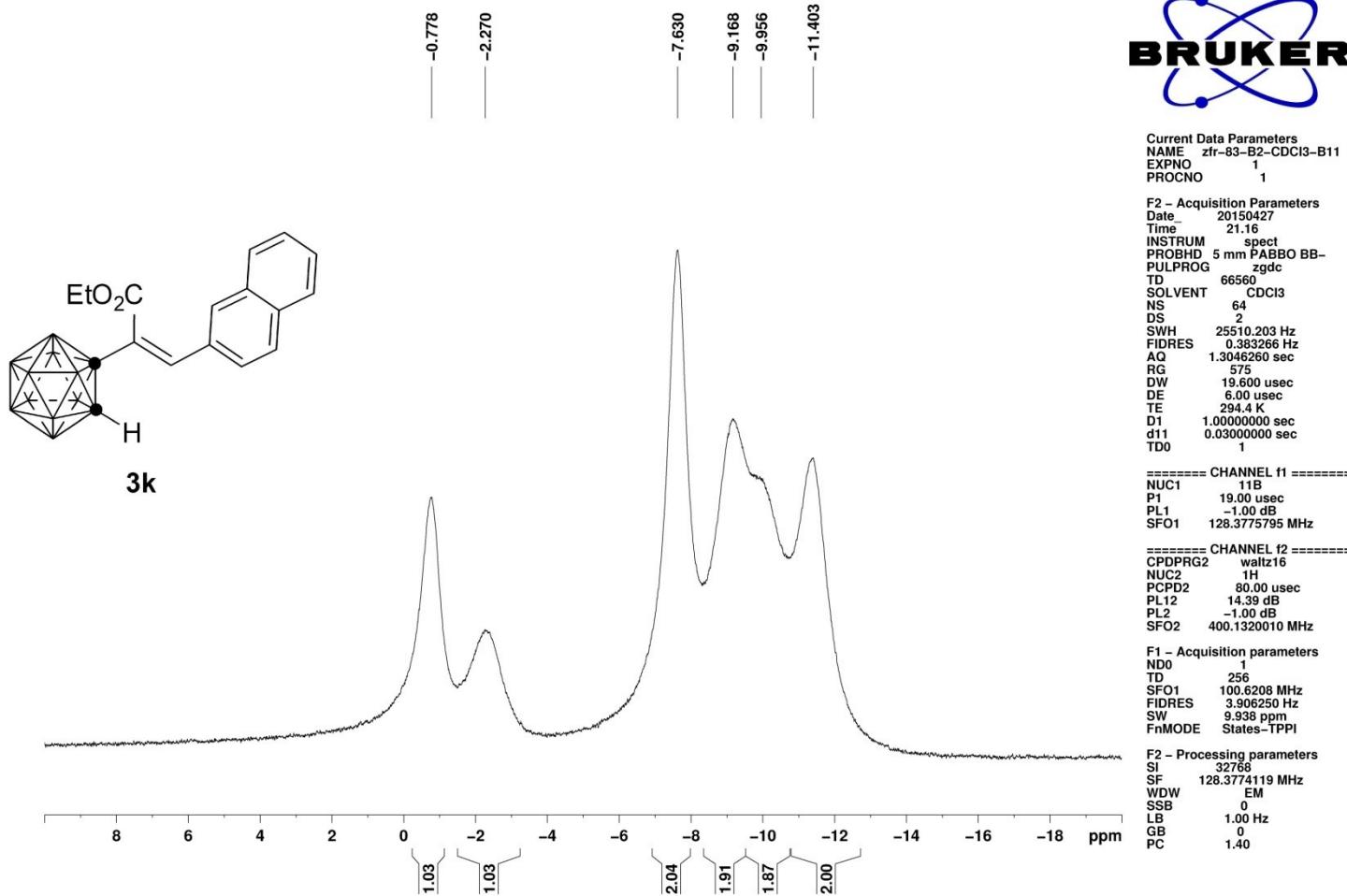
Current Data Parameters
 NAME zfr-142-CDCl3-C13
 EXPNO 1
 PROCNO 1

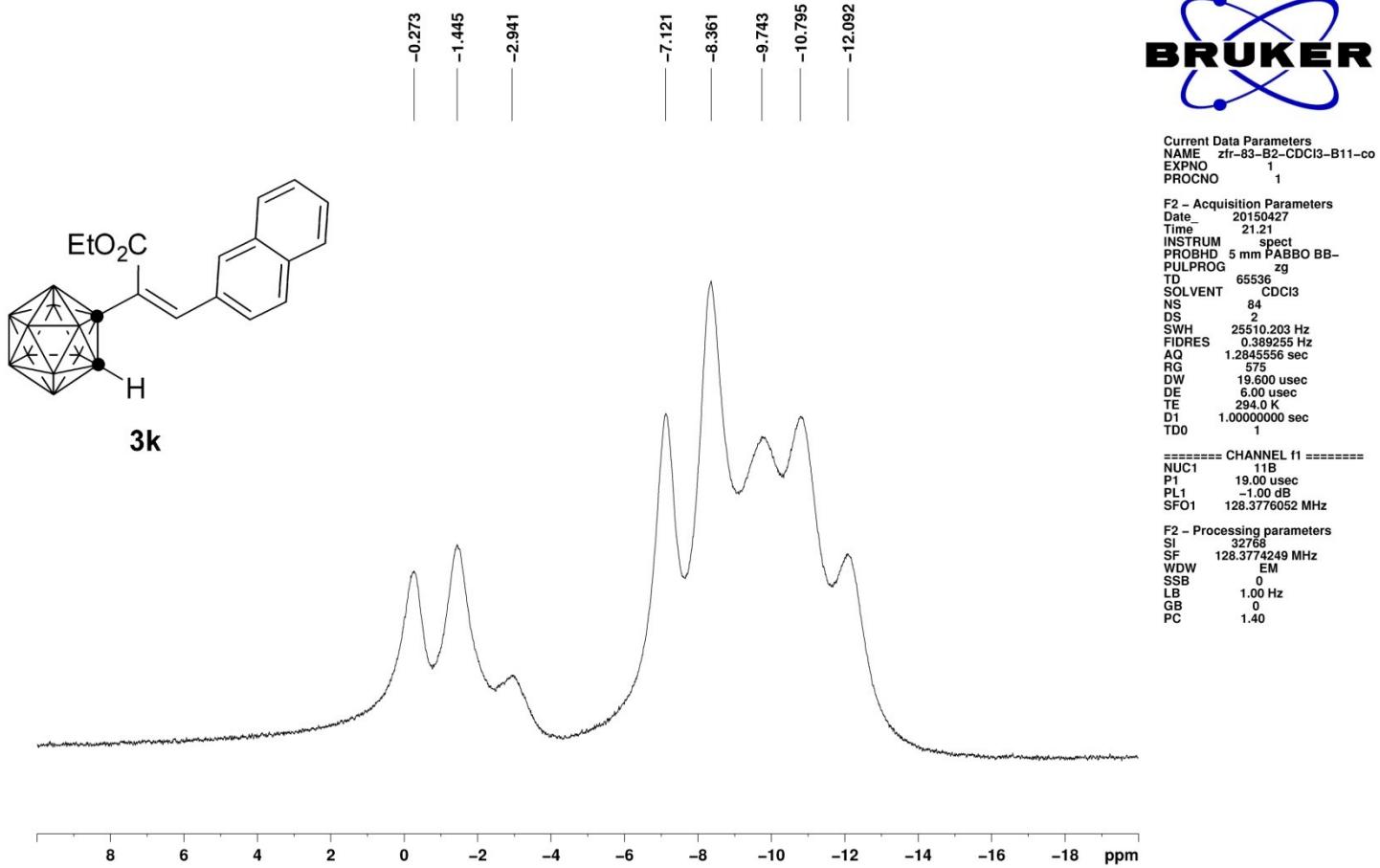
F2 - Acquisition Parameters
 Date 20150630
 Time 15.16
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 300
 DS 2
 SWH 24038.461 Hz
 FIDRES 0.366798 Hz
 AQ 1.3631988 sec
 RG 2050
 DW 20.800 usec
 DE 6.00 usec
 TE 295.3 K
 D1 2.0000000 sec
 d11 0.03000000 sec
 DELTA 1.8999998 sec
 TDO 1

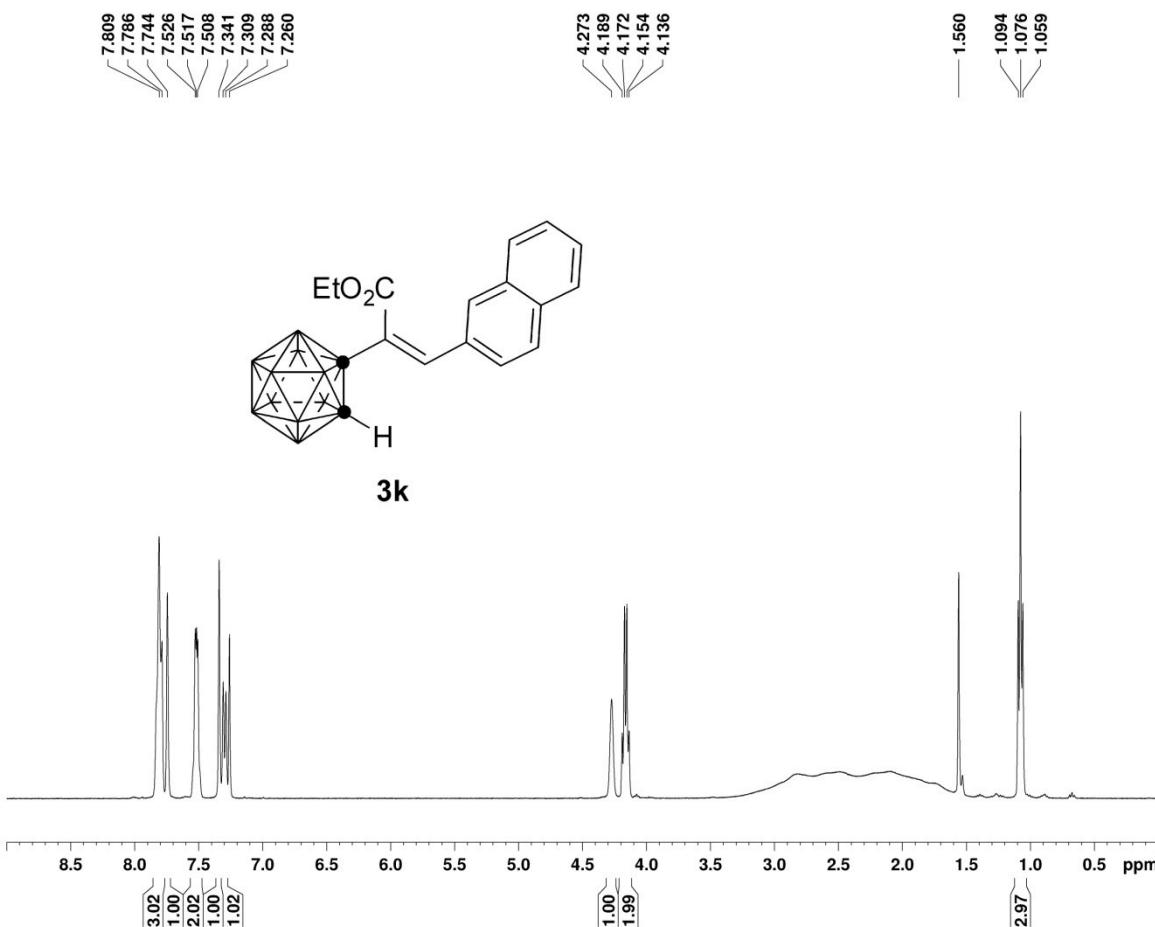
===== CHANNEL f1 =====
 NUC1 13C
 P1 9.25 usec
 PL1 -3.00 dB
 SFO1 100.6228298 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL12 12.45 dB
 PL13 18.00 dB
 PL2 -1.00 dB
 SFO2 400.1316005 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6127497 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40





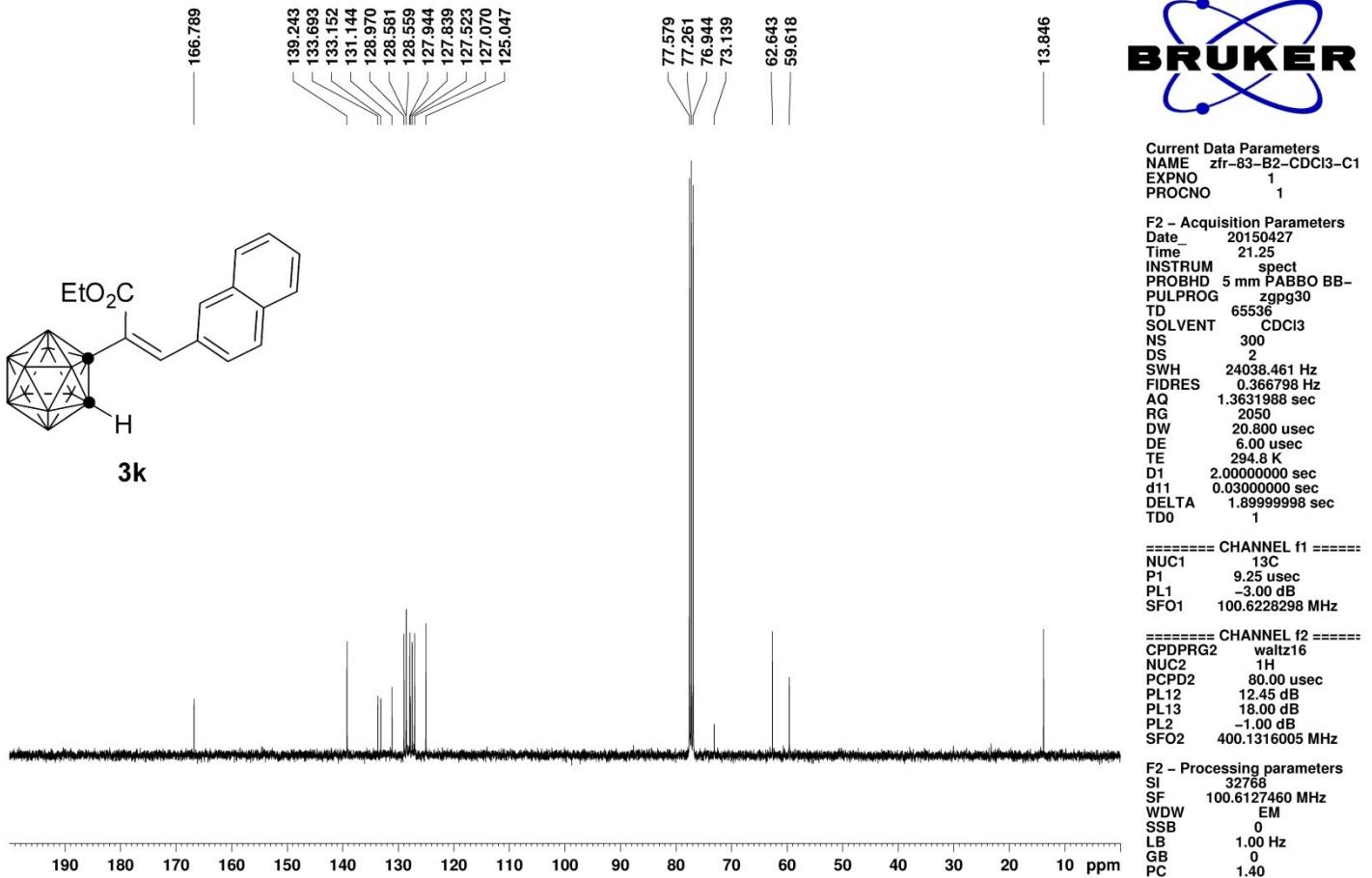


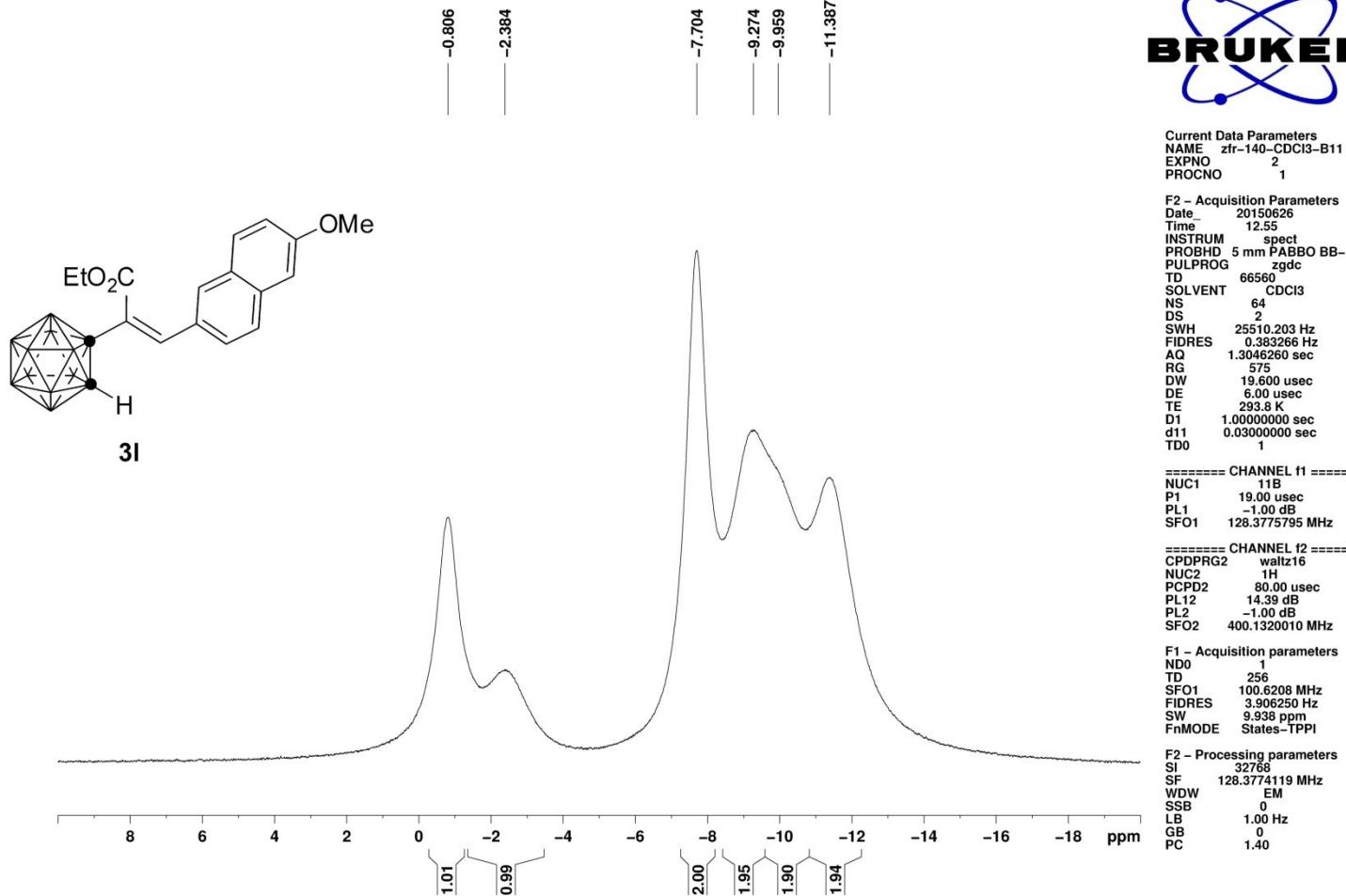
Current Data Parameters
 NAME zfr-83-CDCl3-H1
 EXPNO 1
 PROCNO 1

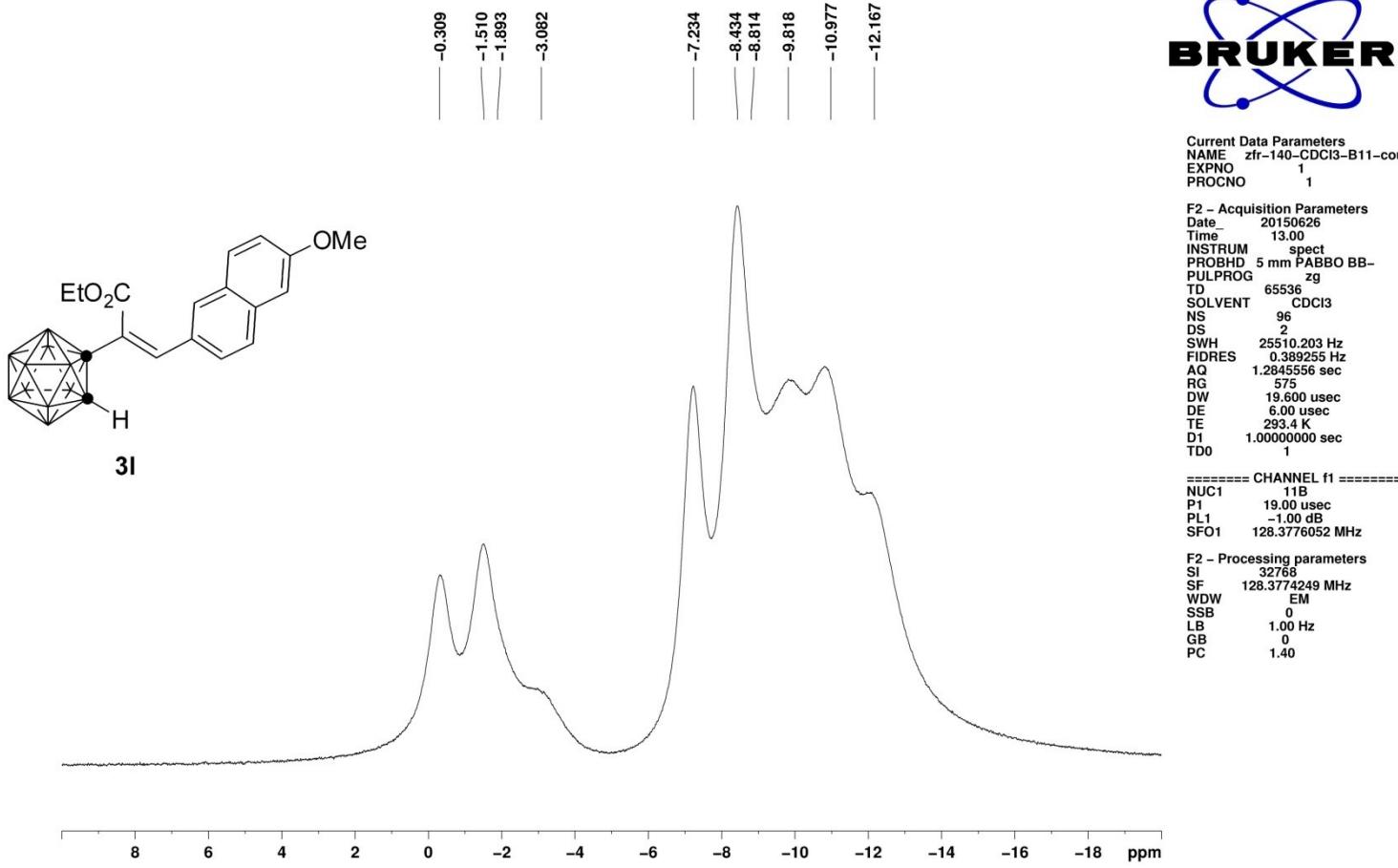
F2 - Acquisition Parameters
 Date 20150430
 Time 16.28
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 8223.685 Hz
 FIDRES 0.125483 Hz
 AQ 3.9846387 sec
 RG 406
 DW 60.800 usec
 DE 6.00 usec
 TE 293.6 K
 D1 1.0000000 sec
 TD0 1

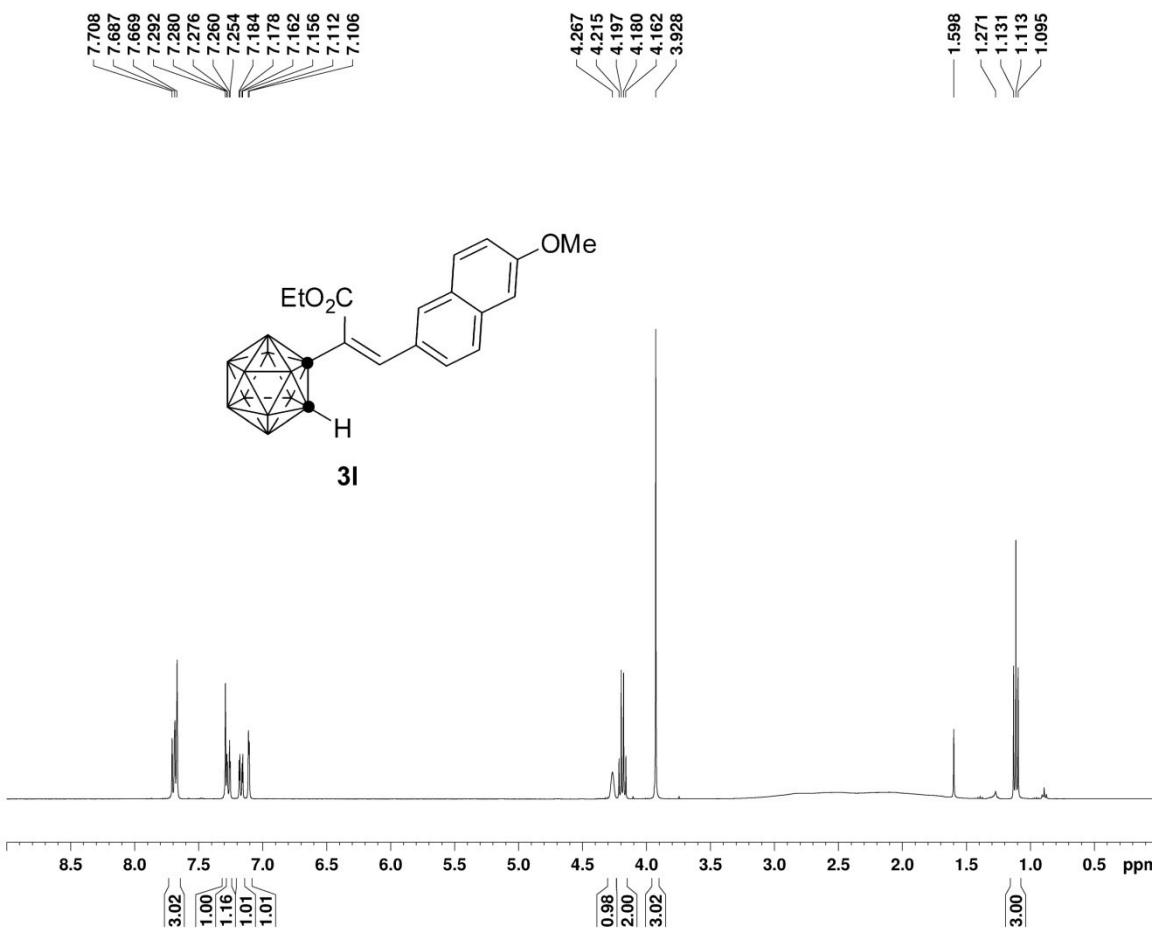
===== CHANNEL f1 =====
 NUC1 1H
 P1 13.60 usec
 PL1 -1.00 dB
 SFO1 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300051 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00







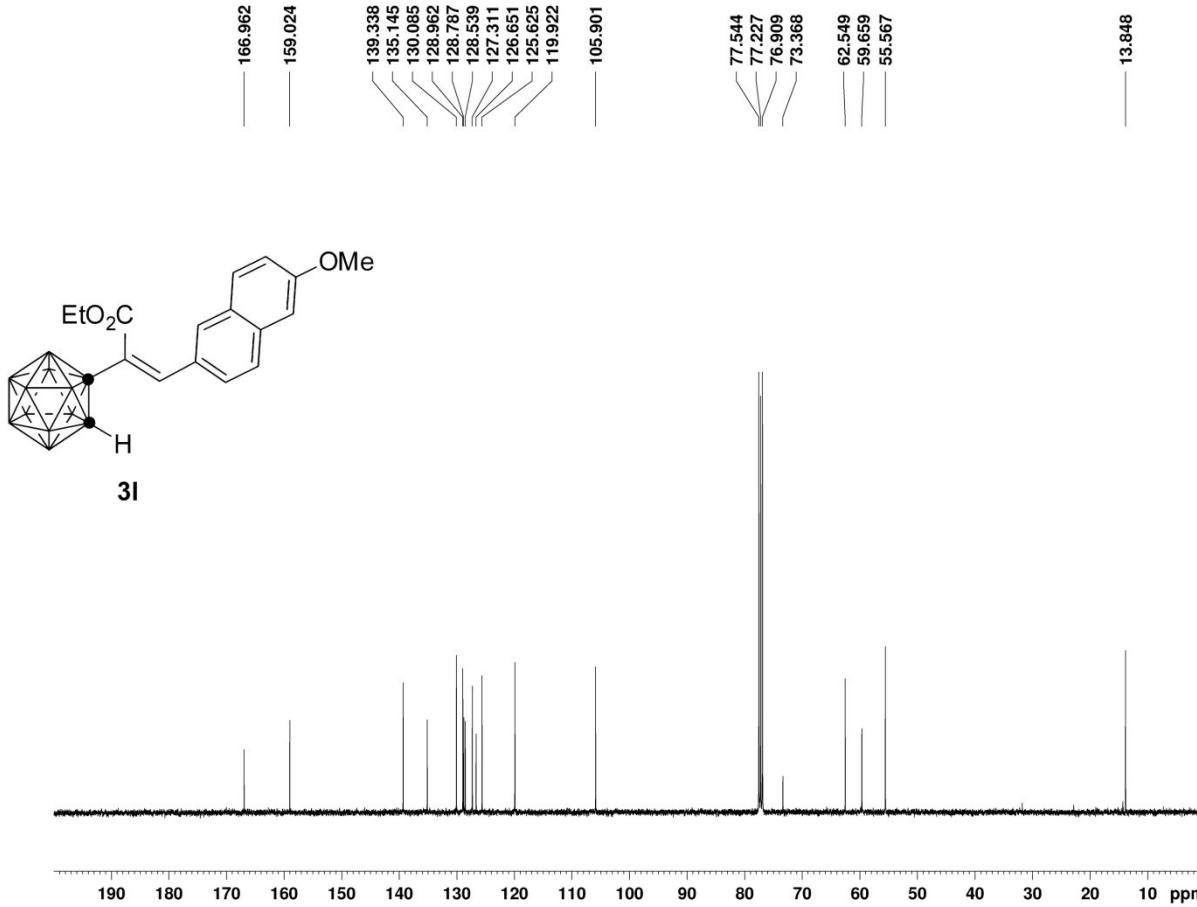


Current Data Parameters
NAME zfr-140-CDCl3-H1
EXPNO 3
PROCNO 1

F2 – Acquisition Parameters
Date_ 20150626
Time 21.34
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9846387 sec
RG 144
DW 60.800 usec
DE 6.00 usec
TE 294.2 K
D1 1.0000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 13.60 usec
PL1 -1.00 dB
SFO1 400.1324710 MHz

F2 – Processing parameters
SI 32768
SF 400.1300051 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



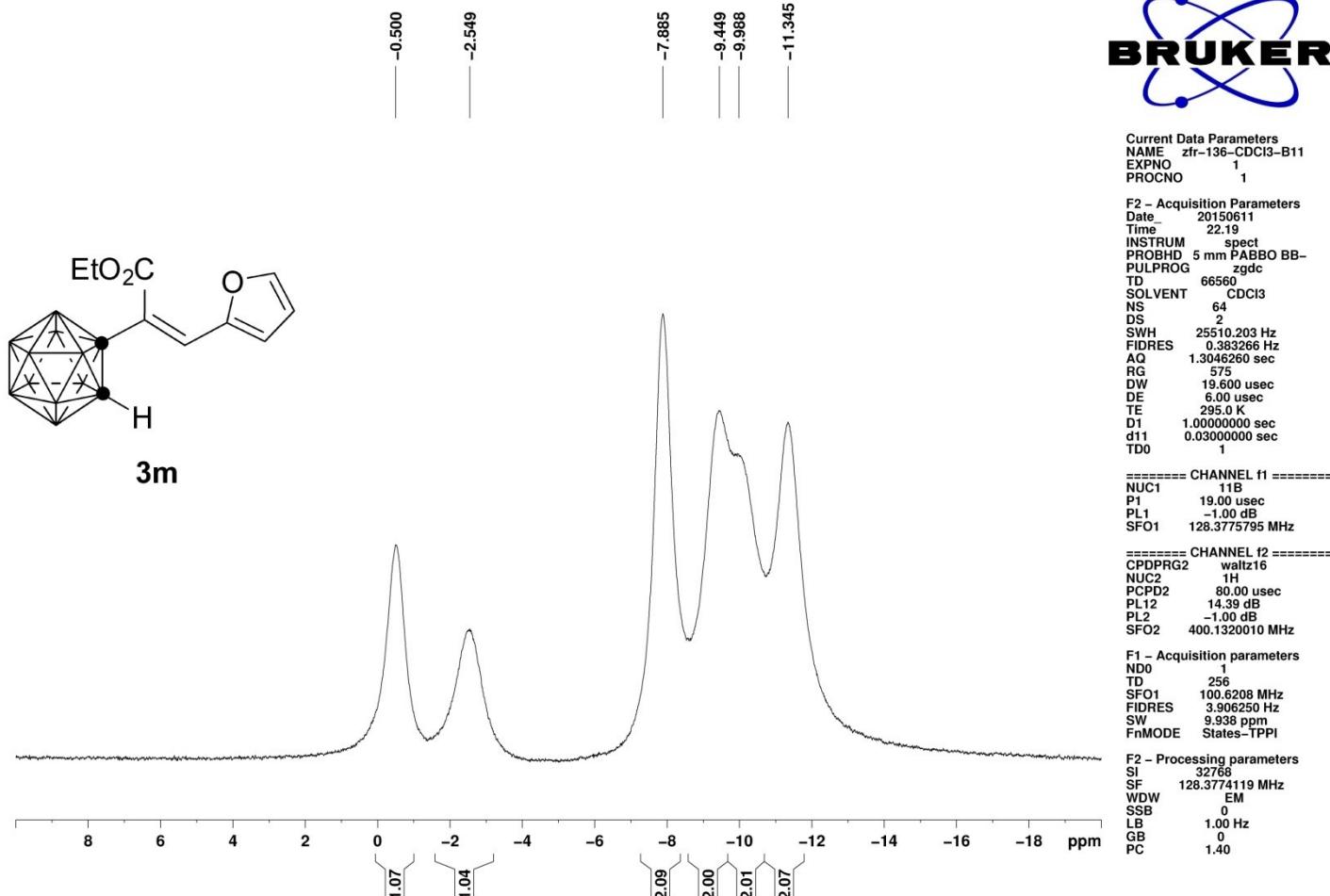
Current Data Parameters
NAME zfr-140-CDCl3-C13
EXPNO 1
PROCNO 1

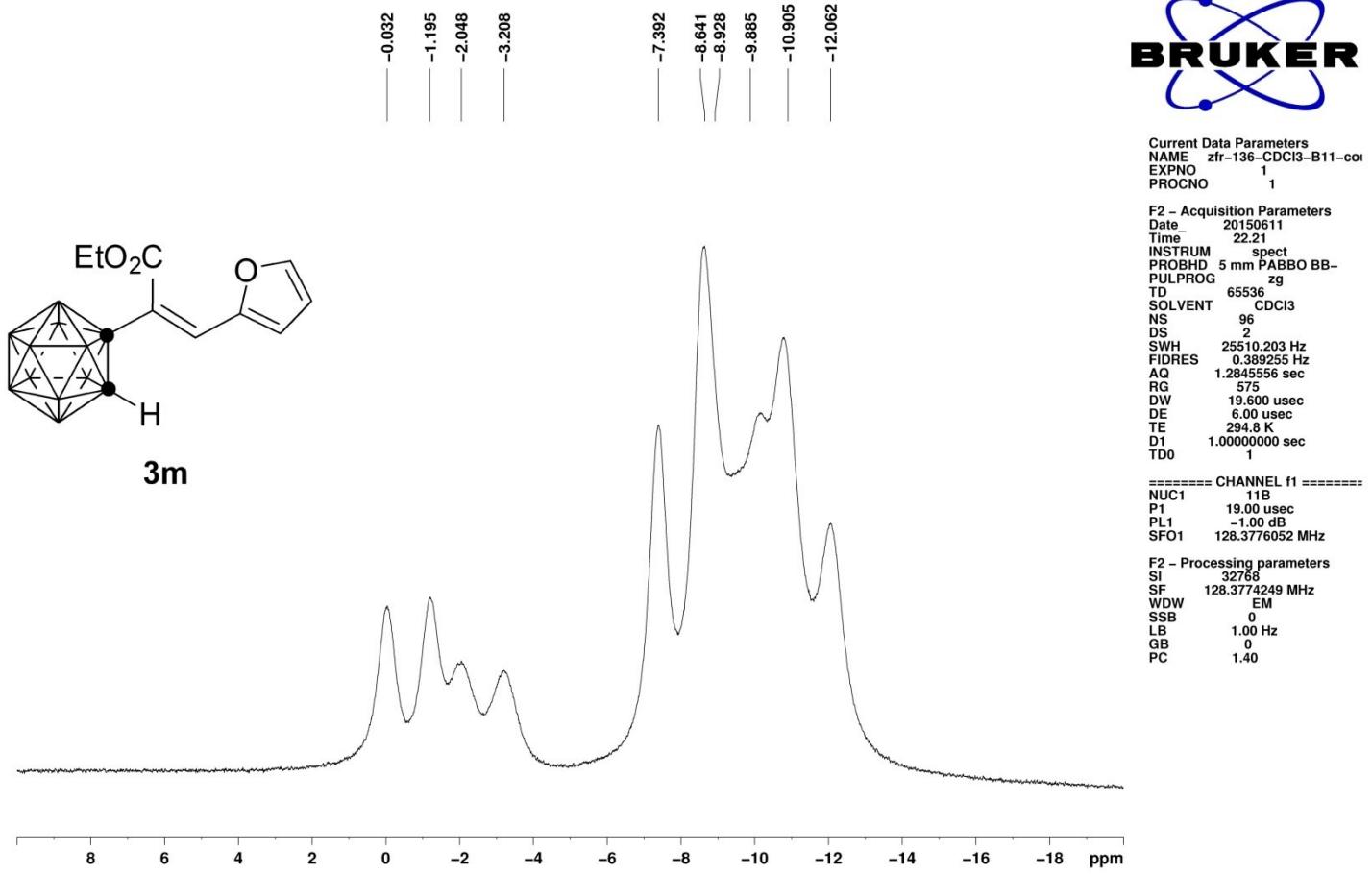
F2 - Acquisition Parameters
Date_ 20150626
Time 21.40
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 463
DS 2
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631988 sec
RG 2050
DW 20.800 usec
DE 6.00 usec
TE 294.9 K
D1 2.0000000 sec
d11 0.03000000 sec
DELTA 1.8999998 sec
TD0 1

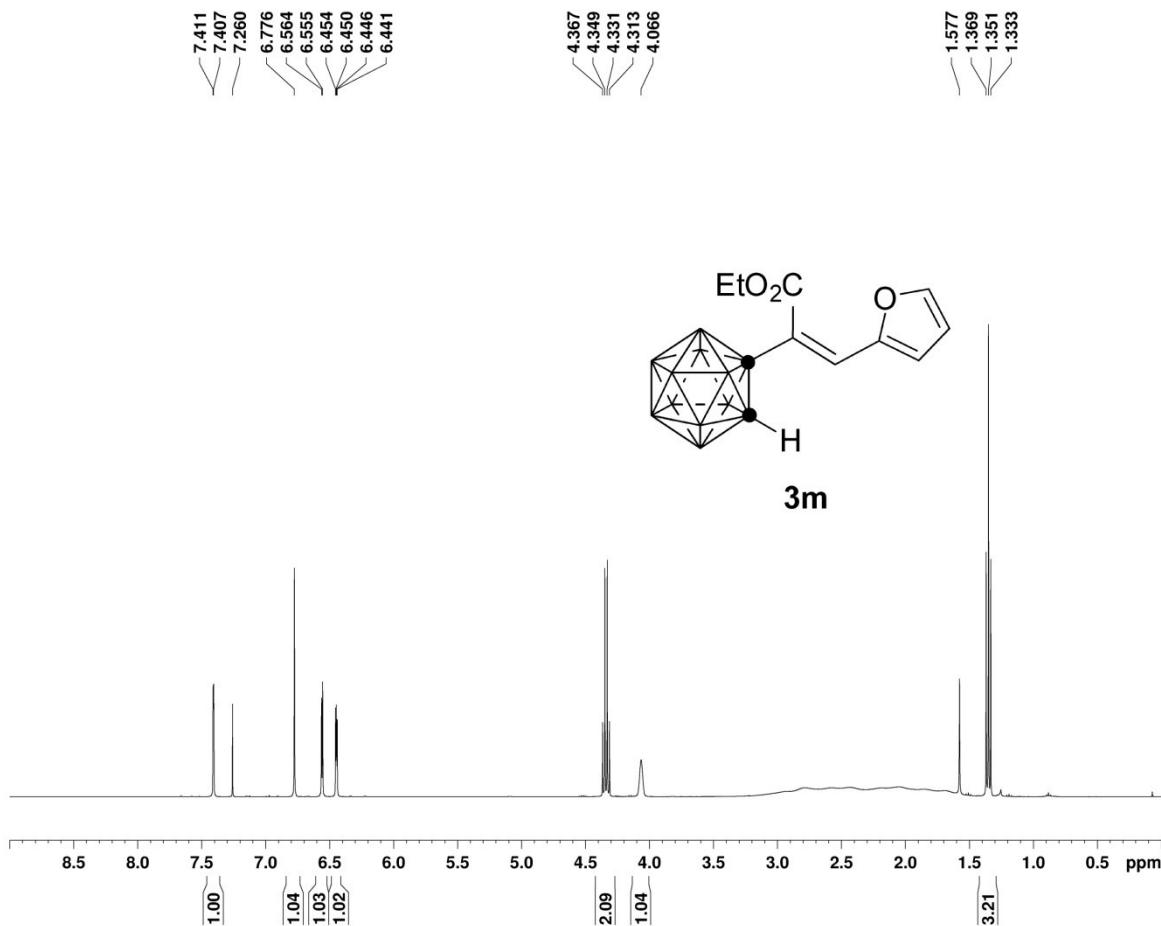
===== CHANNEL f1 =====
NUC1 13C
P1 9.25 usec
PL1 -3.00 dB
SFO1 100.6228298 MHz

===== CHANNEL f2 =====
CPDPGR2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL12 12.45 dB
PL13 18.00 dB
PL2 -1.00 dB
SFO2 400.1316005 MHz

F2 - Processing parameters
SI 32768
SF 100.6127504 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40





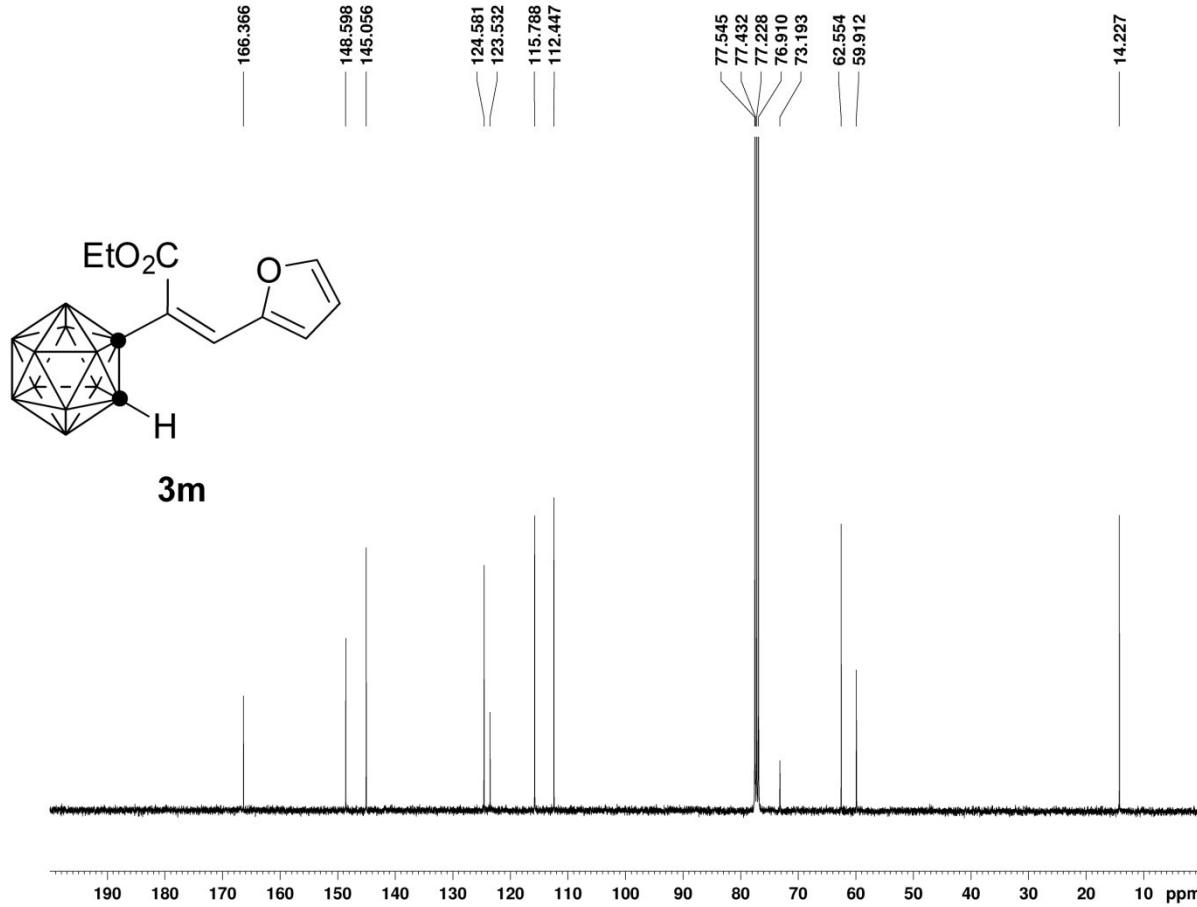


Current Data Parameters
 NAME zfr-136-CDCl₃-H1
 EXPNO 1
 PROCNO 1

F2 – Acquisition Parameters
 Date 20150611
 Time 22.14
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 65536
 SOLVENT CDCl₃
 NS 16
 DS 2
 SWH 8223.685 Hz
 FIDRES 0.125483 Hz
 AQ 3.9846387 sec
 RG 64
 DW 60.800 usec
 DE 6.00 usec
 TE 294.4 K
 D1 1.0000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 13.60 usec
 PL1 -1.00 dB
 SFO1 400.1324710 MHz

F2 – Processing parameters
 SI 32768
 SF 400.1300053 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



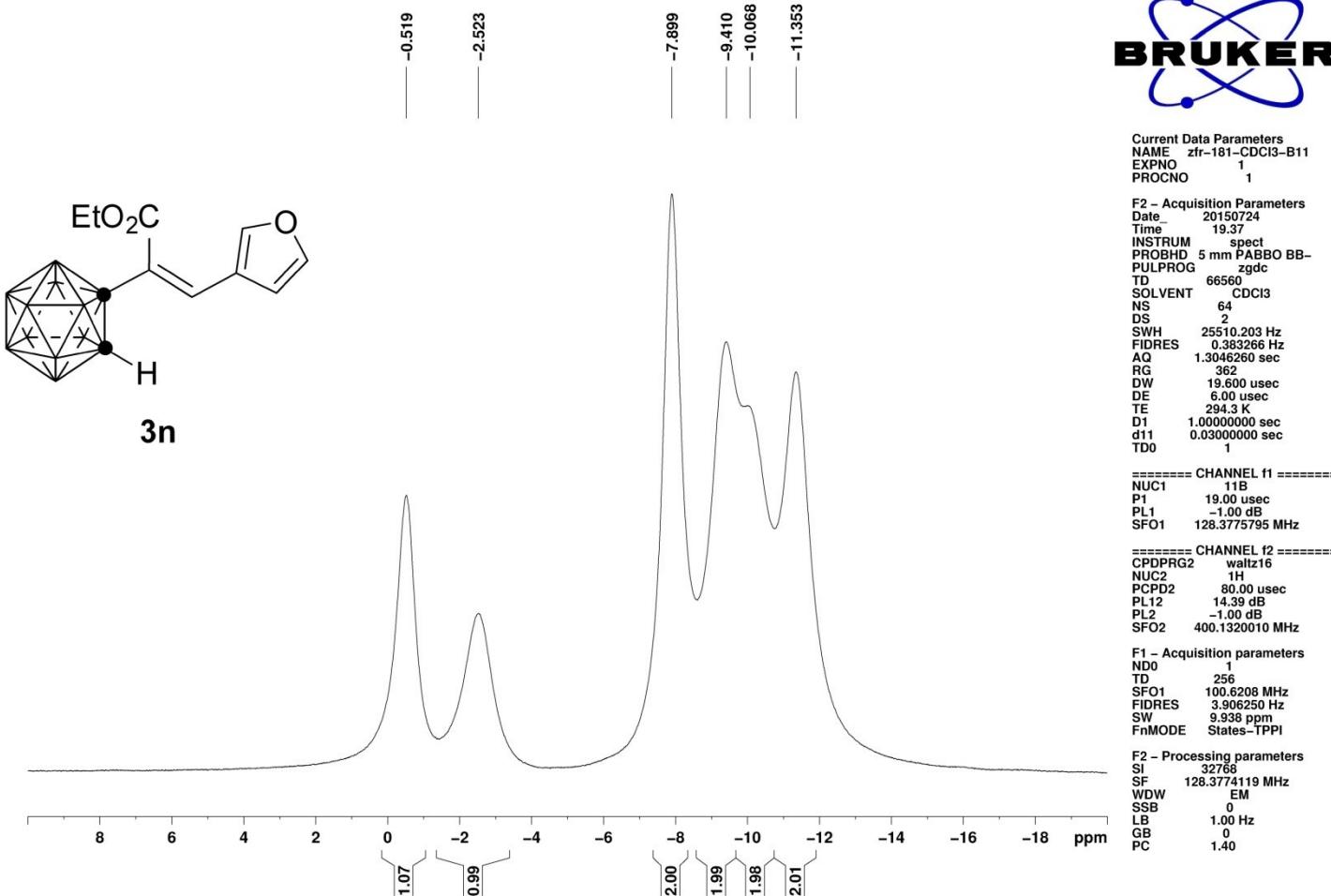
Current Data Parameters
 NAME zfr-136-CDCl3-C13
 EXPNO 1
 PROCNO 1

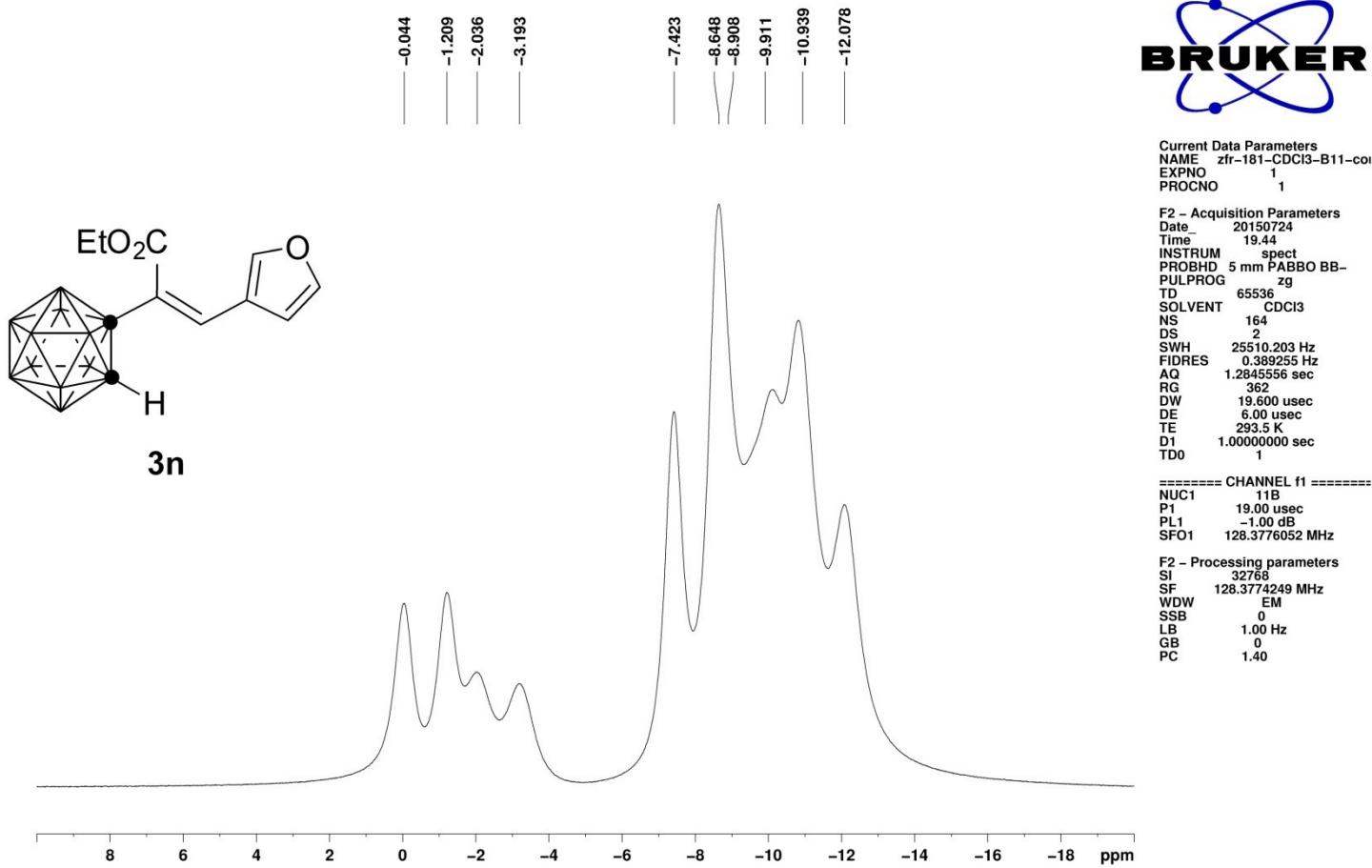
F2 - Acquisition Parameters
 Date 20150612
 Time 9.54
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 1336
 DS 2
 SWH 24038.461 Hz
 FIDRES 0.366798 Hz
 AQ 1.3631988 sec
 RG 2050
 DW 20.800 usec
 DE 6.00 usec
 TE 294.6 K
 D1 2.0000000 sec
 d11 0.03000000 sec
 DELTA 1.8999998 sec
 TDO 1

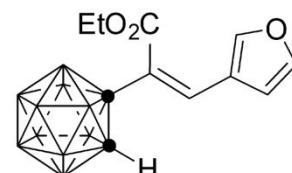
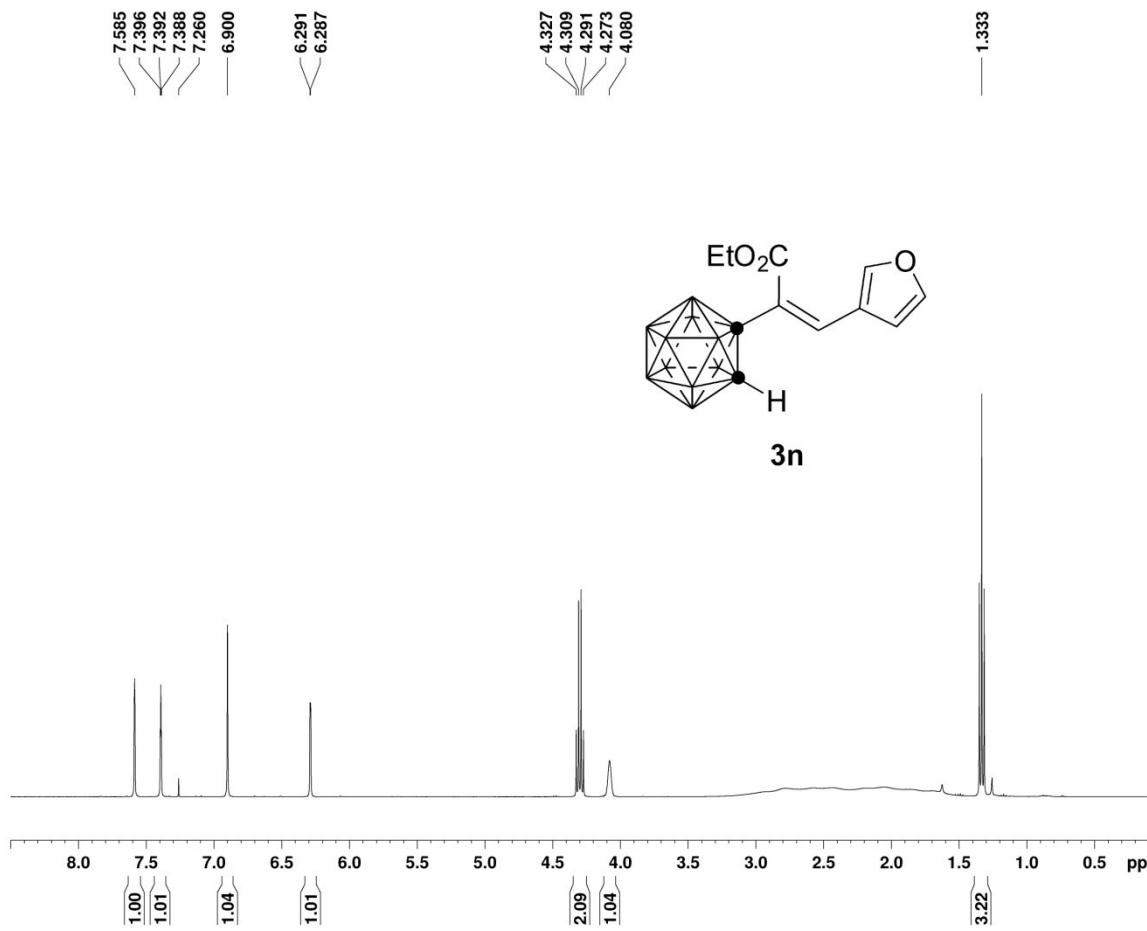
===== CHANNEL f1 =====
 NUC1 13C
 P1 9.25 usec
 PL1 -3.00 dB
 SF01 100.6228298 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL12 12.45 dB
 PL13 18.00 dB
 PL2 -1.00 dB
 SF02 400.1316005 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6127483 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40







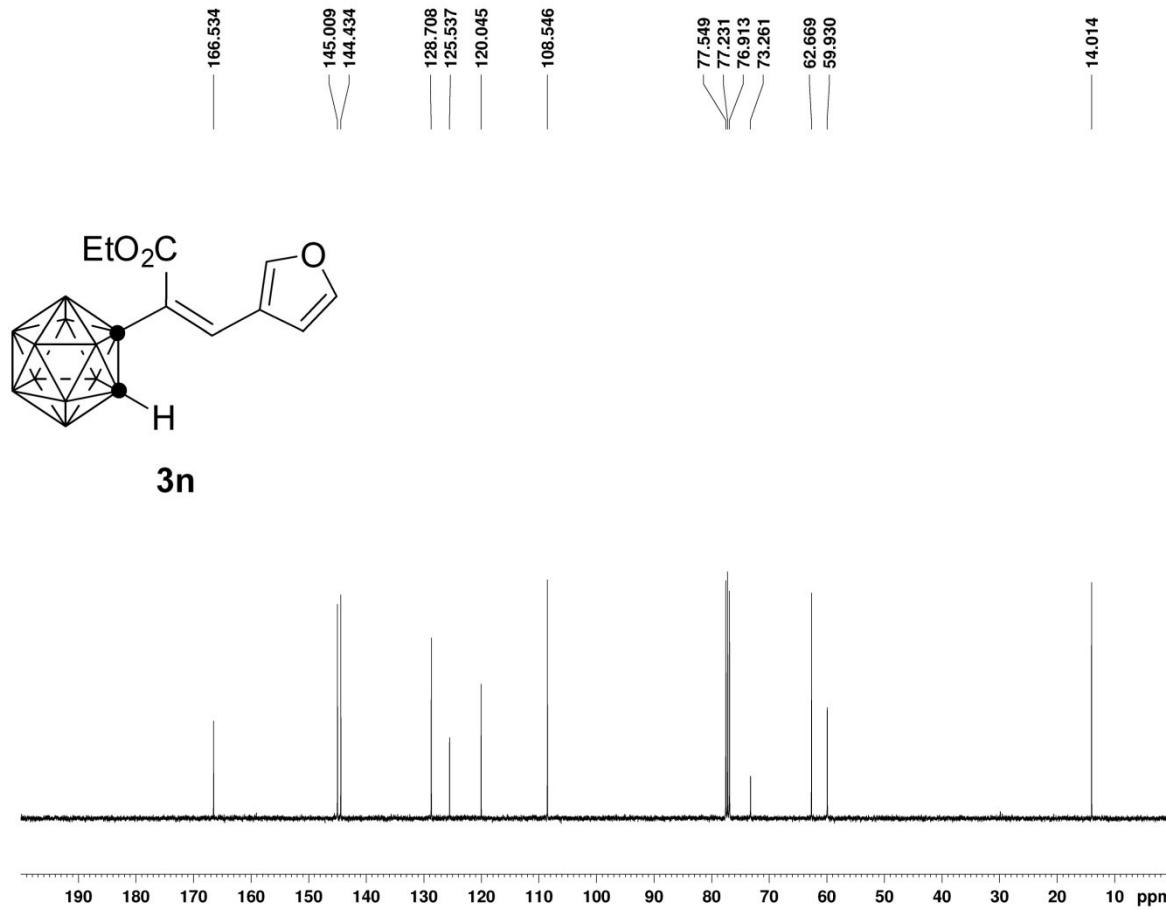
3n

Current Data Parameters
NAME zfr-181-CDCI3-H1
EXPNO 1
PROCNO 1

F2 – Acquisition Parameters
 Date 20150724
 Time 19.34
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9846387 sec
RG 64
DW 60.800 usec
DE 6.00 usec
TE 293.5 K
D1 1.0000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 13.60 usec
PL1 -1.00 dB
SFO1 400.1324710 MHz

F2 – Processing parameters
 SI 32768
 SF 400.1300053 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



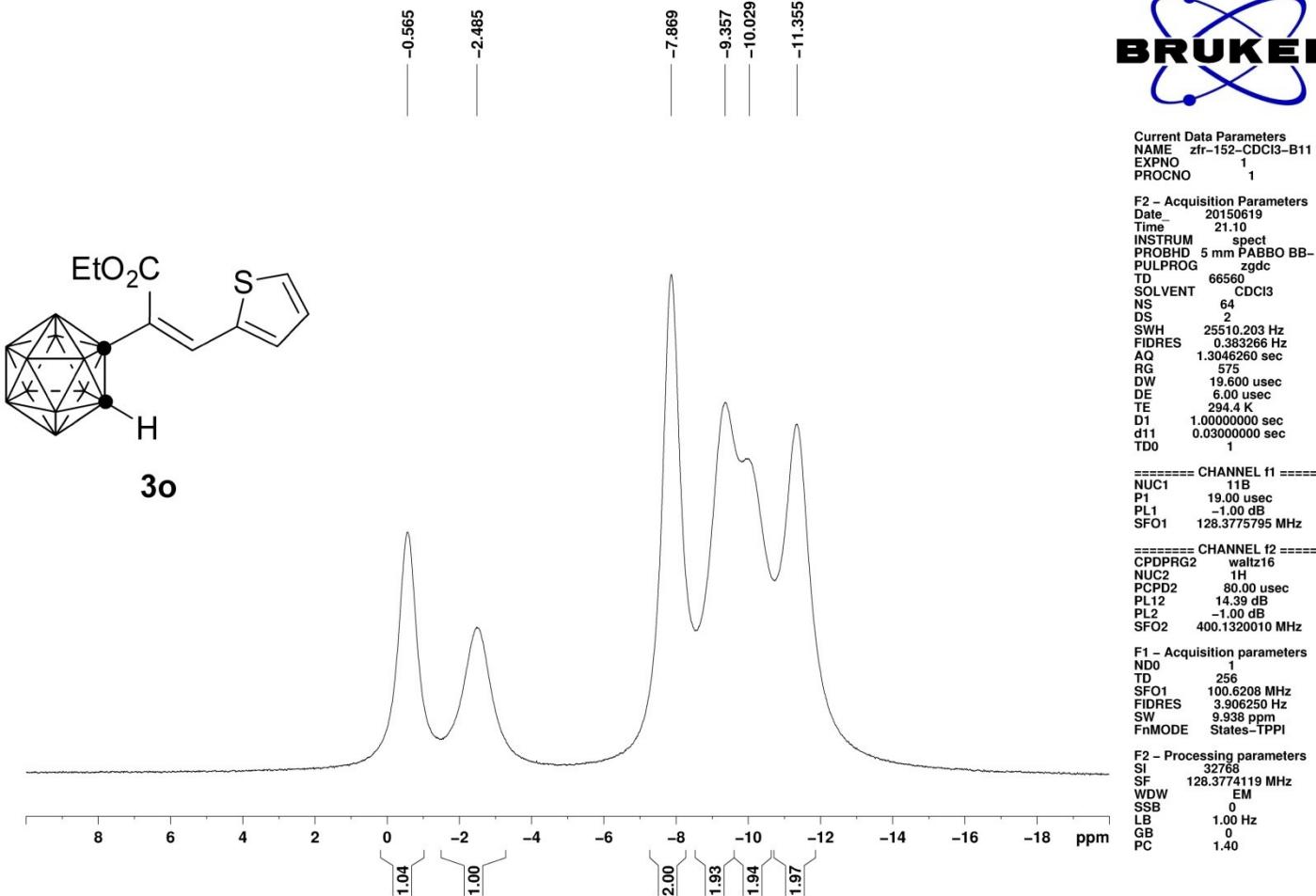
Current Data Parameters
 NAME zfr-181-CDCl₃-C13
 EXPNO 1
 PROCNO 1

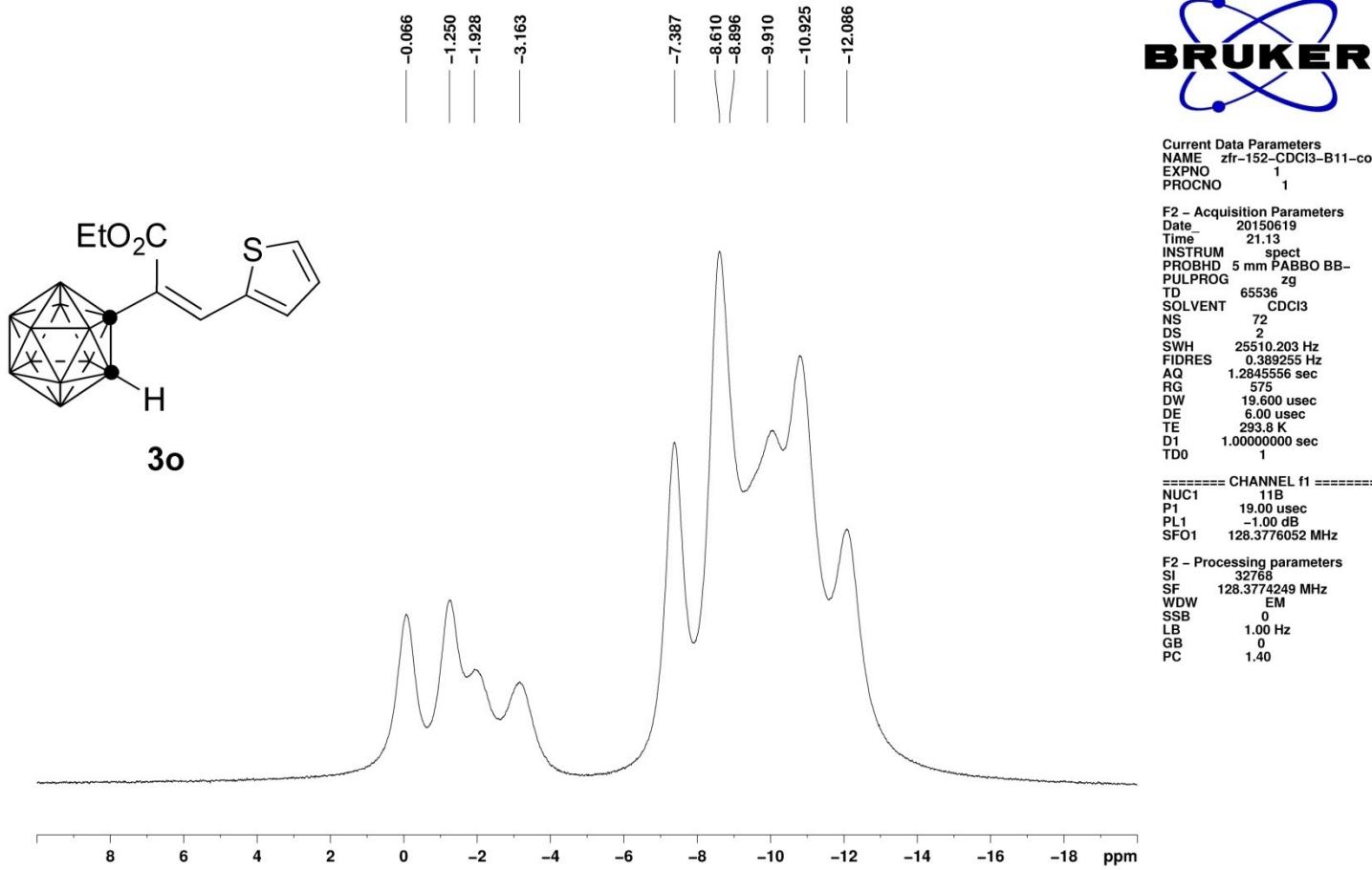
F2 – Acquisition Parameters
 Date 20150724
 Time 19.49
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl₃
 NS 160
 DS 2
 SWH 24038.461 Hz
 FIDRES 0.366798 Hz
 AQ 1.3631988 sec
 RG 2050
 DW 20.800 usec
 DE 6.00 usec
 TE 294.3 K
 D1 2.0000000 sec
 d11 0.03000000 sec
 DELTA 1.8999999 sec
 TD0 1

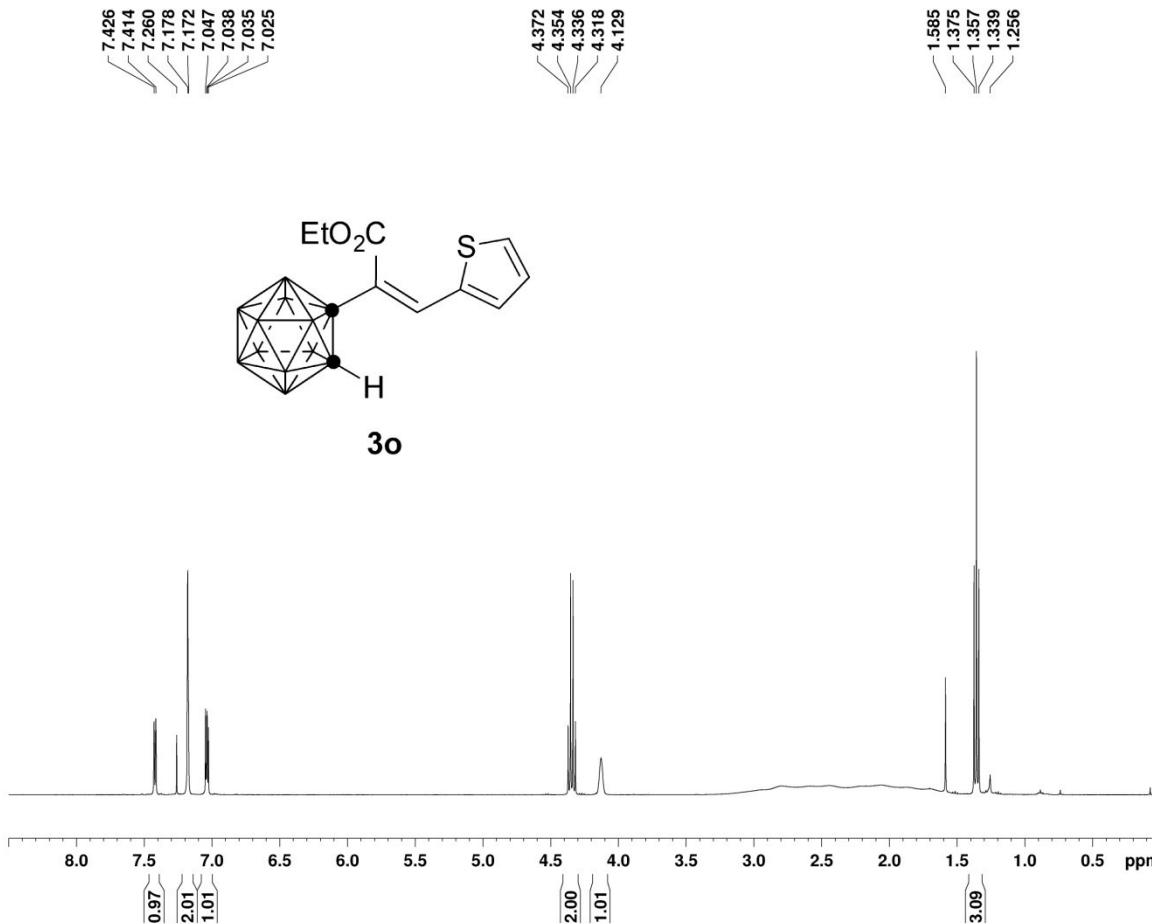
===== CHANNEL f1 =====
 NUC1 ¹³C
 P1 9.25 usec
 PL1 -3.00 dB
 SFO1 100.6228298 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 ¹H
 PCPD2 80.00 usec
 PL12 12.45 dB
 PL13 18.00 dB
 PL2 -1.00 dB
 SFO2 400.1316005 MHz

F2 – Processing parameters
 SI 32768
 SF 100.6127520 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40





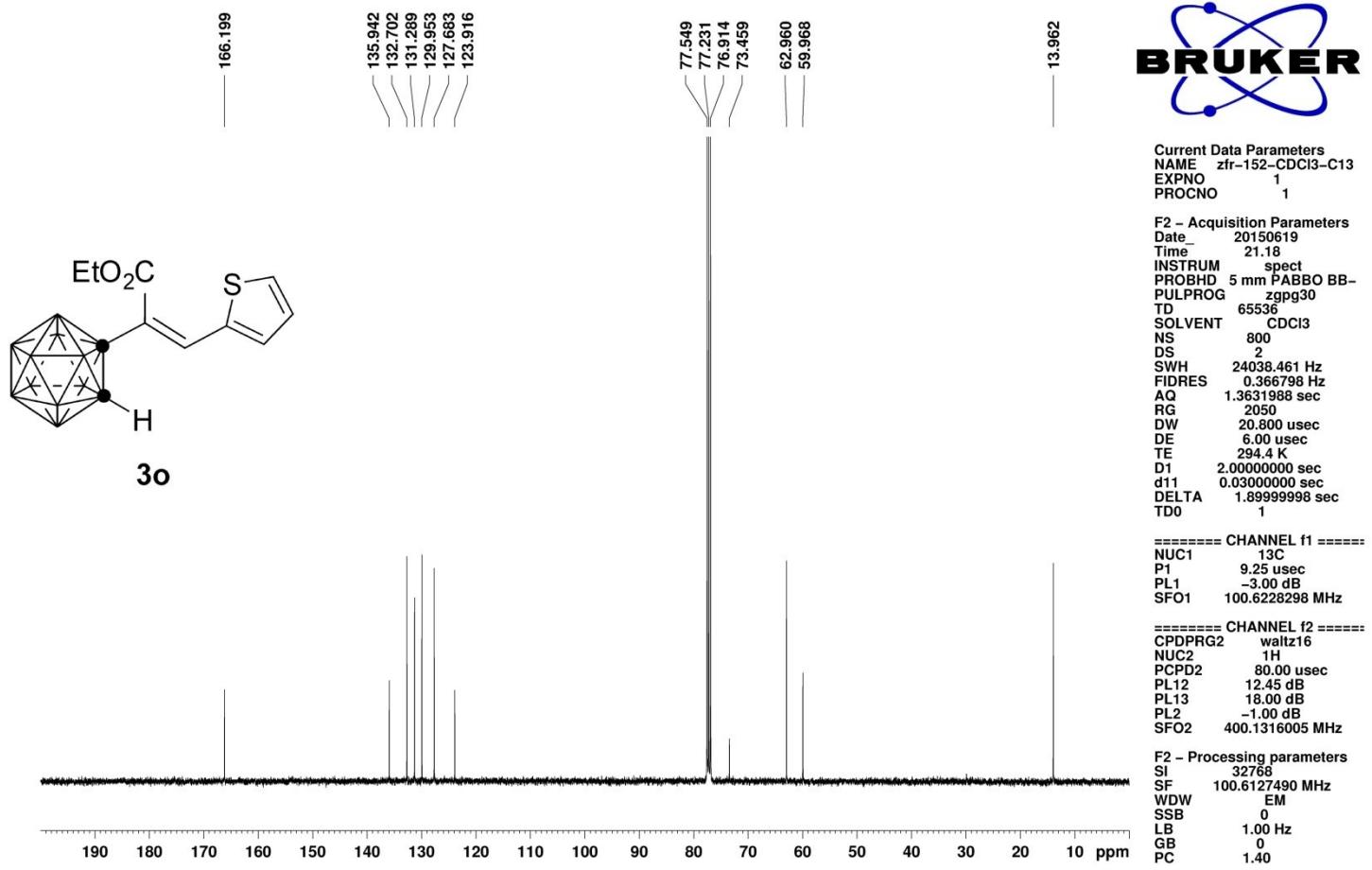


Current Data Parameters
NAME zfr-152-CDCl3-H1
EXPNO 1
PROCNO 1

F2 – Acquisition Parameters
Date_ 20150619
Time 21.06
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9846387 sec
RG 181
DW 60.800 usec
DE 6.00 usec
TE 293.6 K
D1 1.0000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 13.60 usec
PL1 -1.00 dB
SFO1 400.1324710 MHz

F2 – Processing parameters
SI 32768
SF 400.1300053 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00





Current Data Parameters
NAME zfr-125-CDCl3-B11
EXPNO 1
PROCNO 1

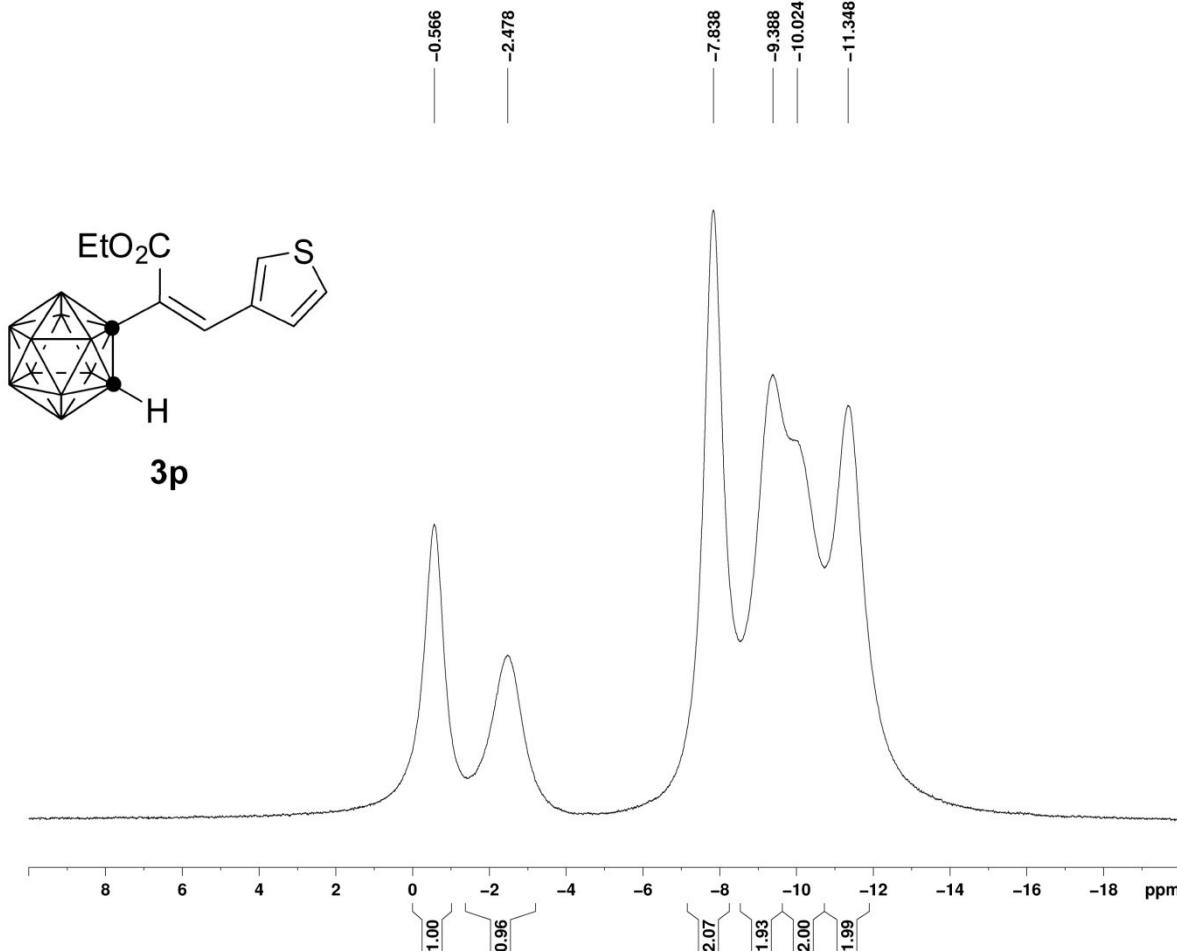
F2 - Acquisition Parameters
Date 20150605
Time 21.27
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgdc
TD 66560
SOLVENT CDCl3
NS 16
DS 2
SWH 25510.203 Hz
FIDRES 0.383266 Hz
AQ 1.3046260 sec
RG 406
DW 19.600 usec
DE 6.00 usec
TE 294.3 K
D1 1.0000000 sec
d11 0.0300000 sec
TD0 1

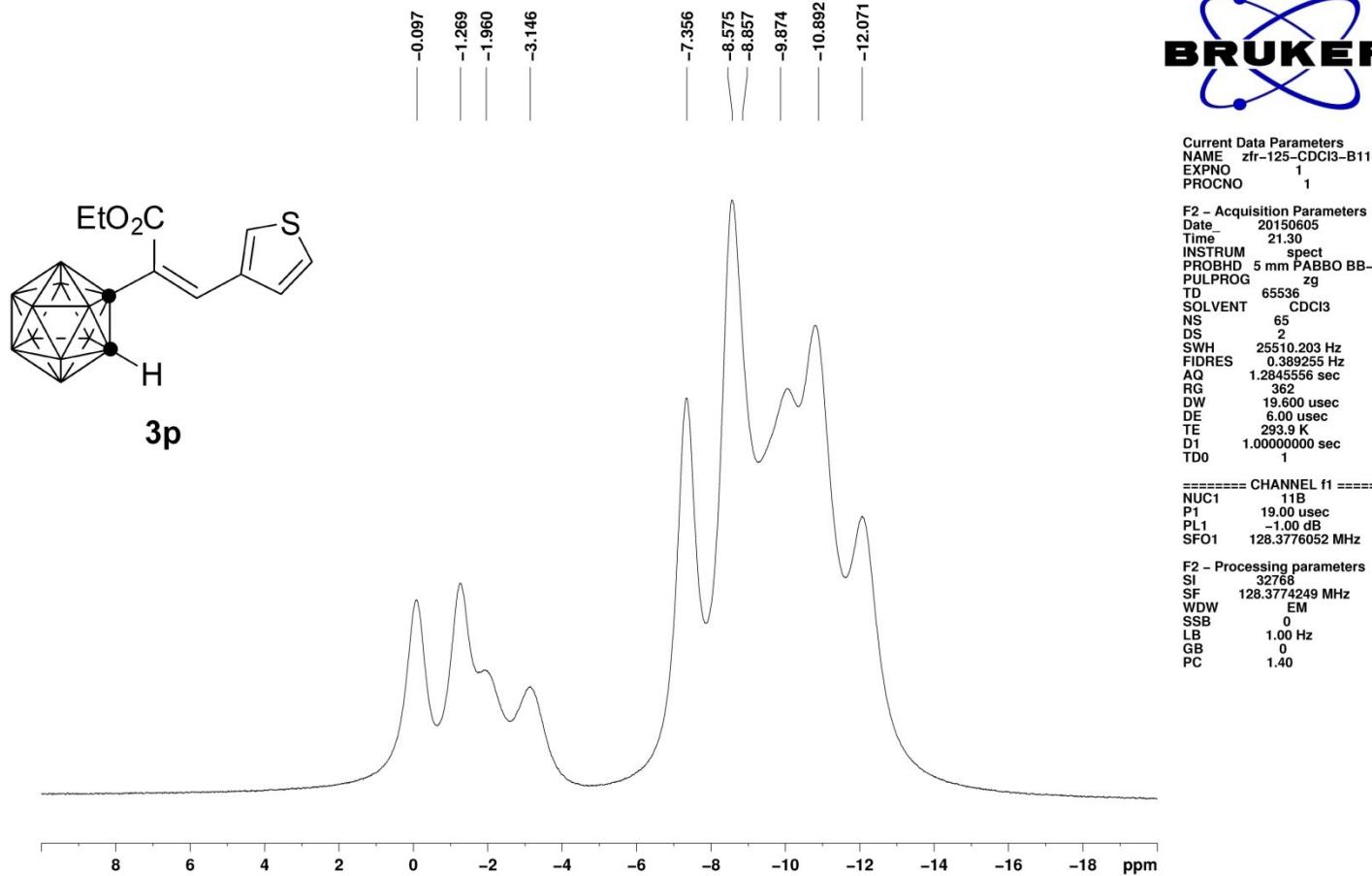
===== CHANNEL f1 ======
NUC1 11B
P1 19.00 usec
PL1 -1.00 dB
SFO1 128.377419 MHz

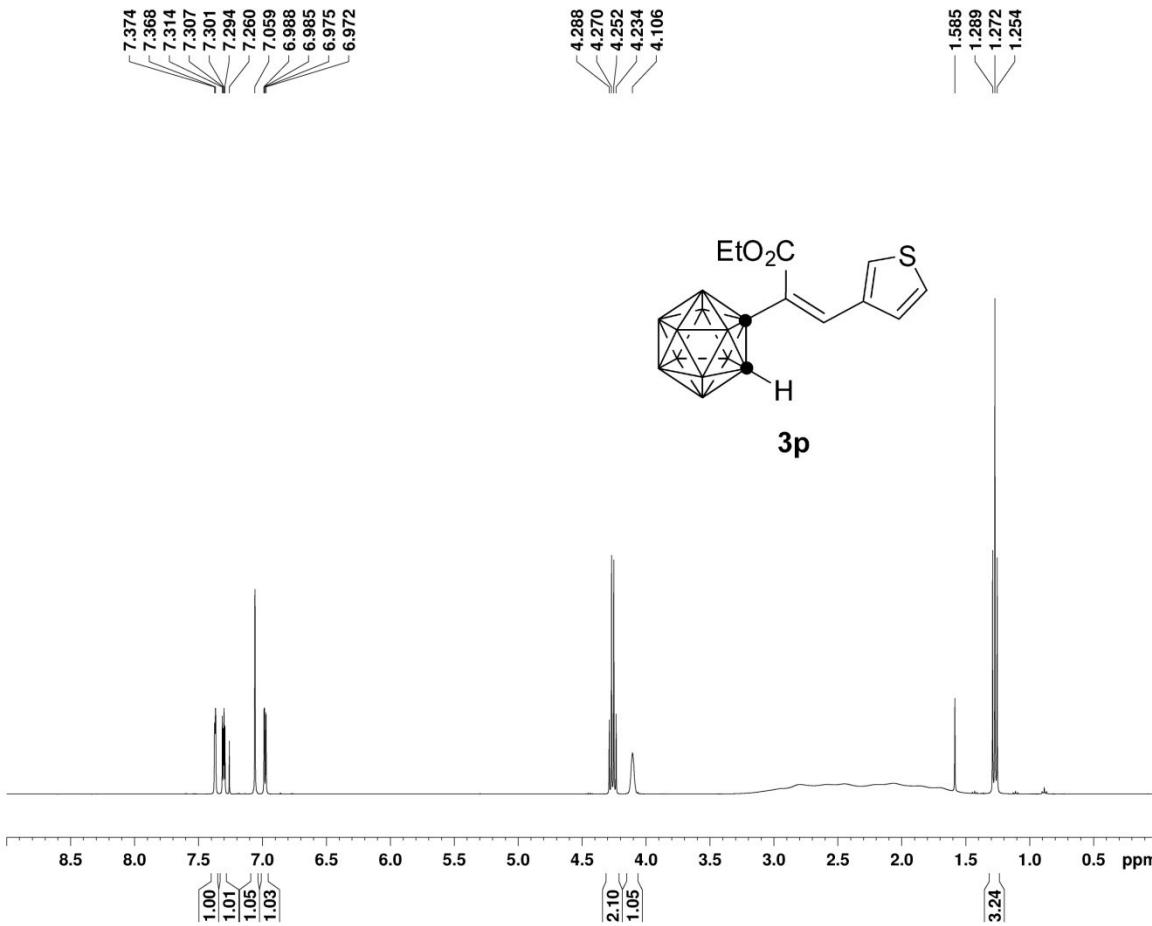
===== CHANNEL f2 ======
CPDPGR2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL12 14.39 dB
PL2 -1.00 dB
SFO2 400.1320010 MHz

F1 - Acquisition parameters
ND0 1
TD 256
SFO1 100.6208 MHz
FIDRES 3.906250 Hz
SW 9.938 ppm
FnMODE States-TPPI

F2 - Processing parameters
SI 32768
SF 128.3774119 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40





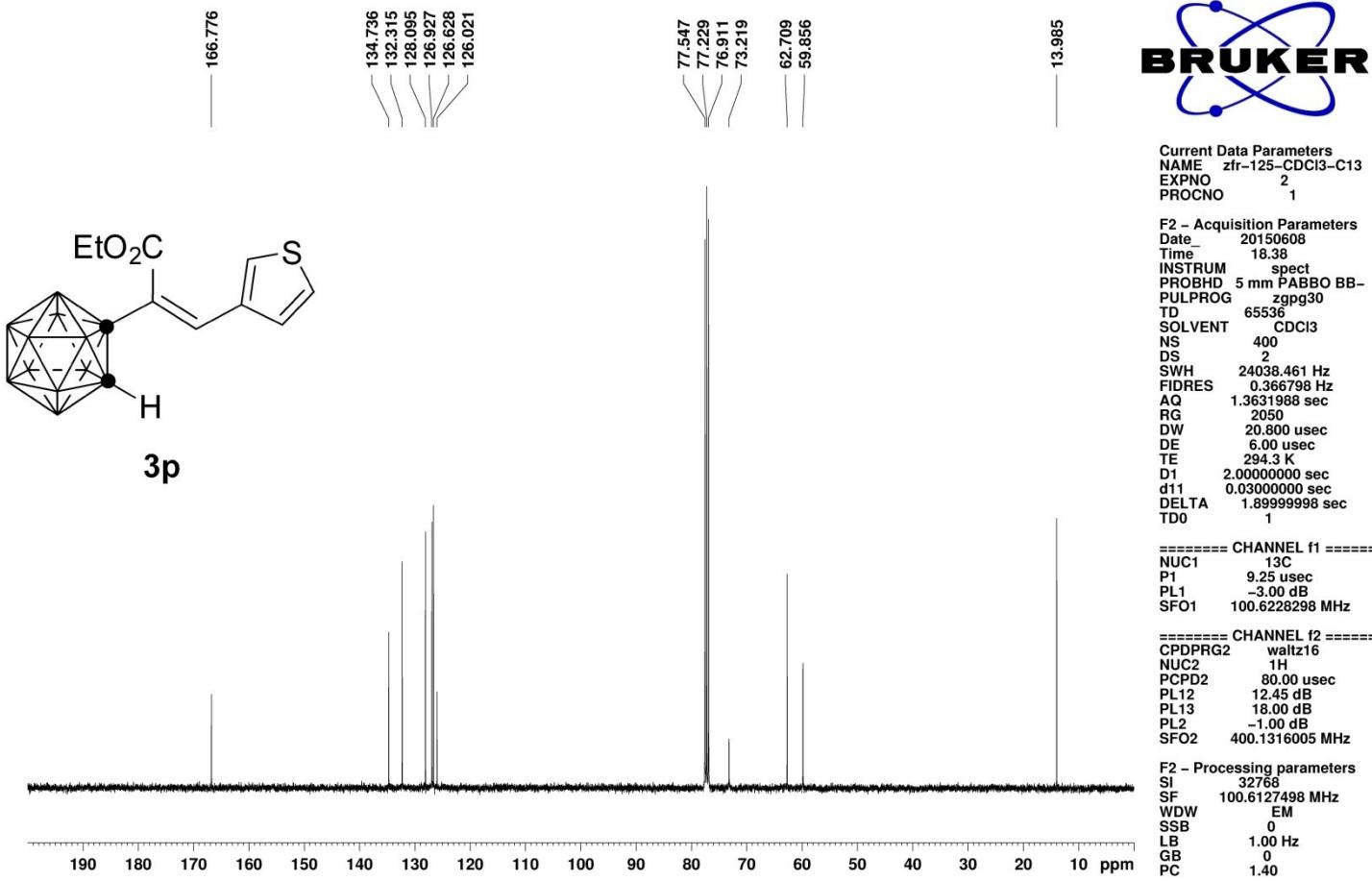


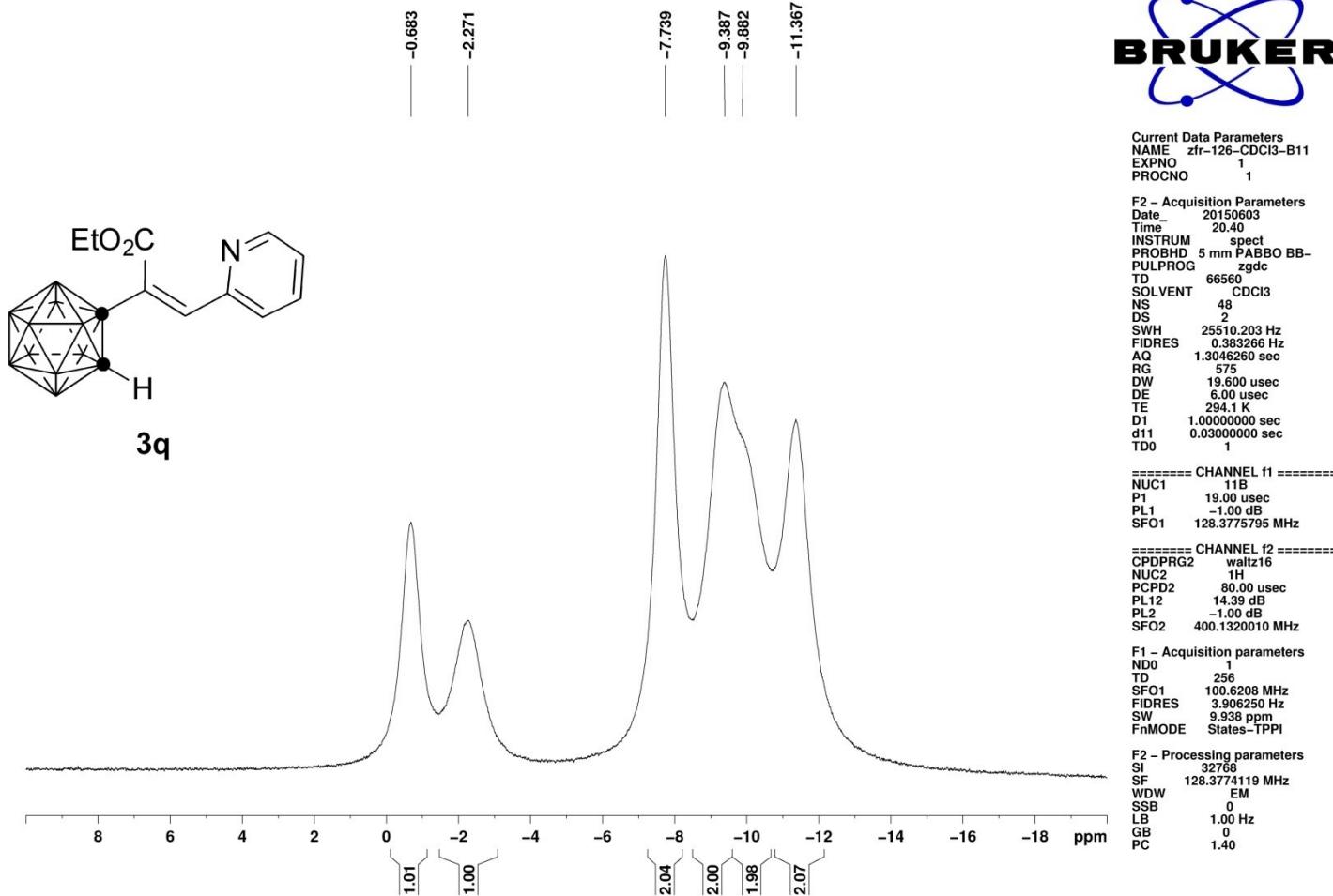
Current Data Parameters
NAME zfr-125-CDCl3-H1
EXPNO 2
PROCNO 1

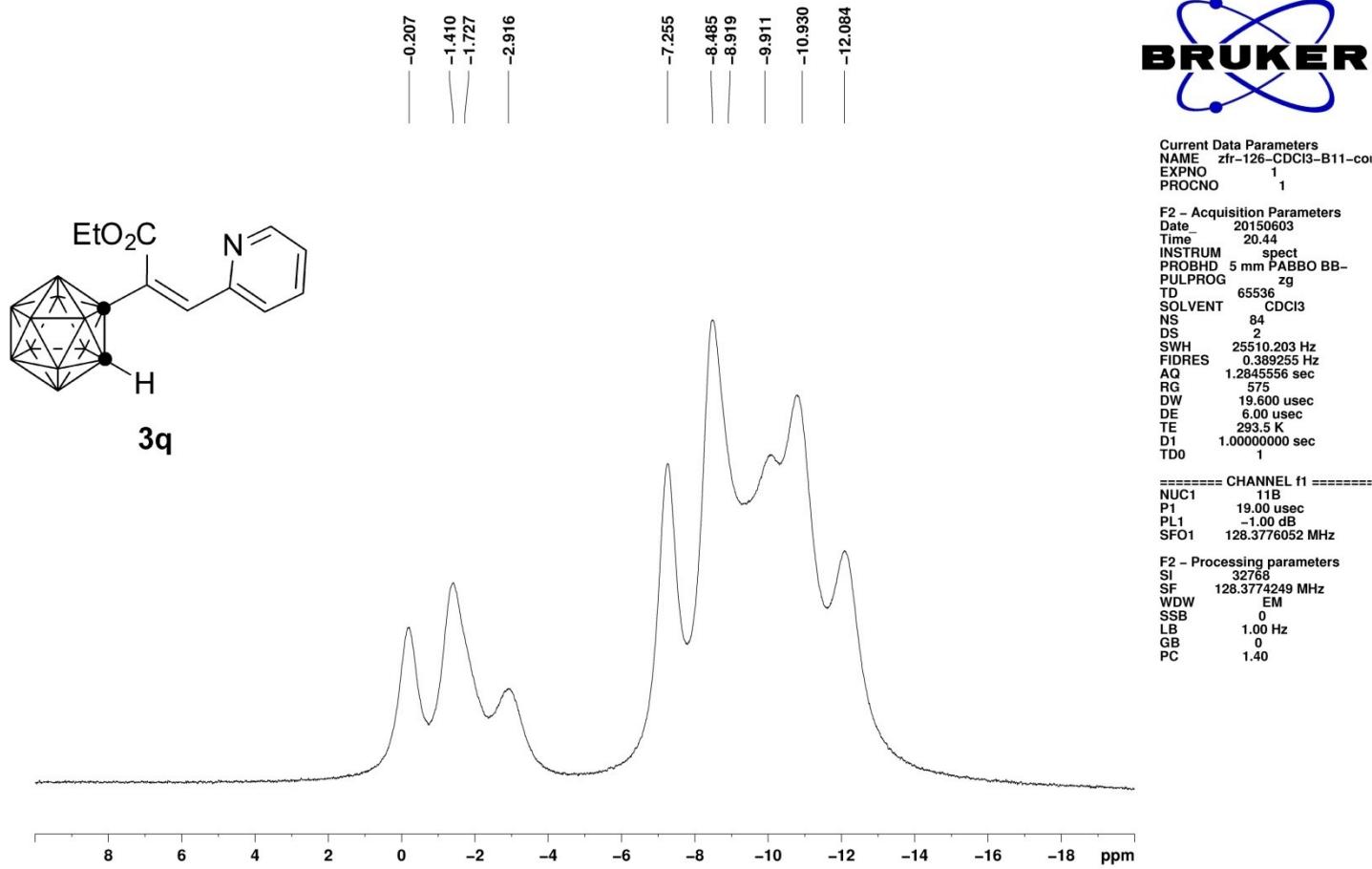
F2 – Acquisition Parameters
Date 20150608
Time 18.34
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9846387 sec
RG 161
DW 60.800 usec
DE 6.00 usec
TE 293.7 K
D1 1.0000000 sec
TD0 1

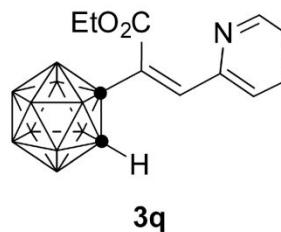
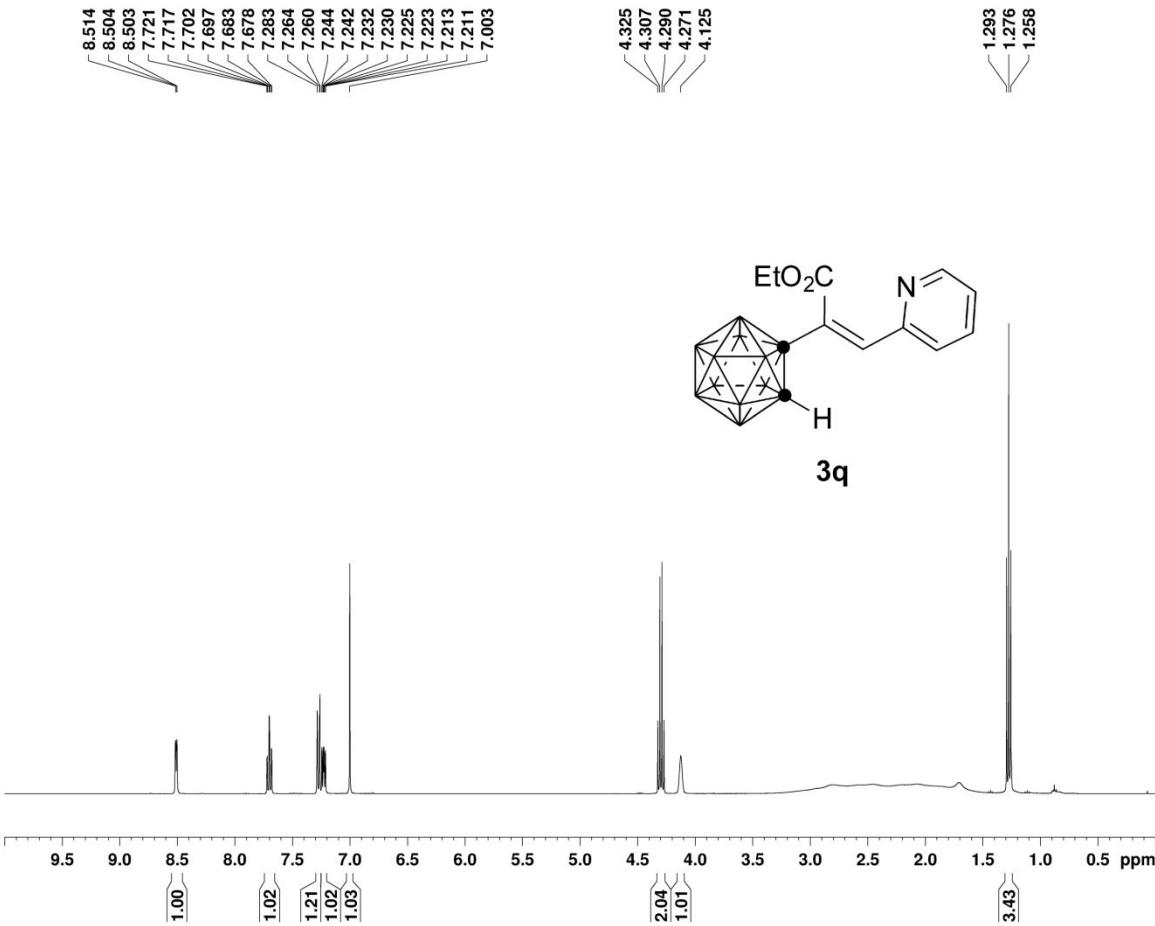
===== CHANNEL f1 =====
NUC1 1H
P1 13.60 usec
PL1 -1.00 dB
SFO1 400.1324710 MHz

F2 – Processing parameters
SI 32768
SF 400.1300053 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00







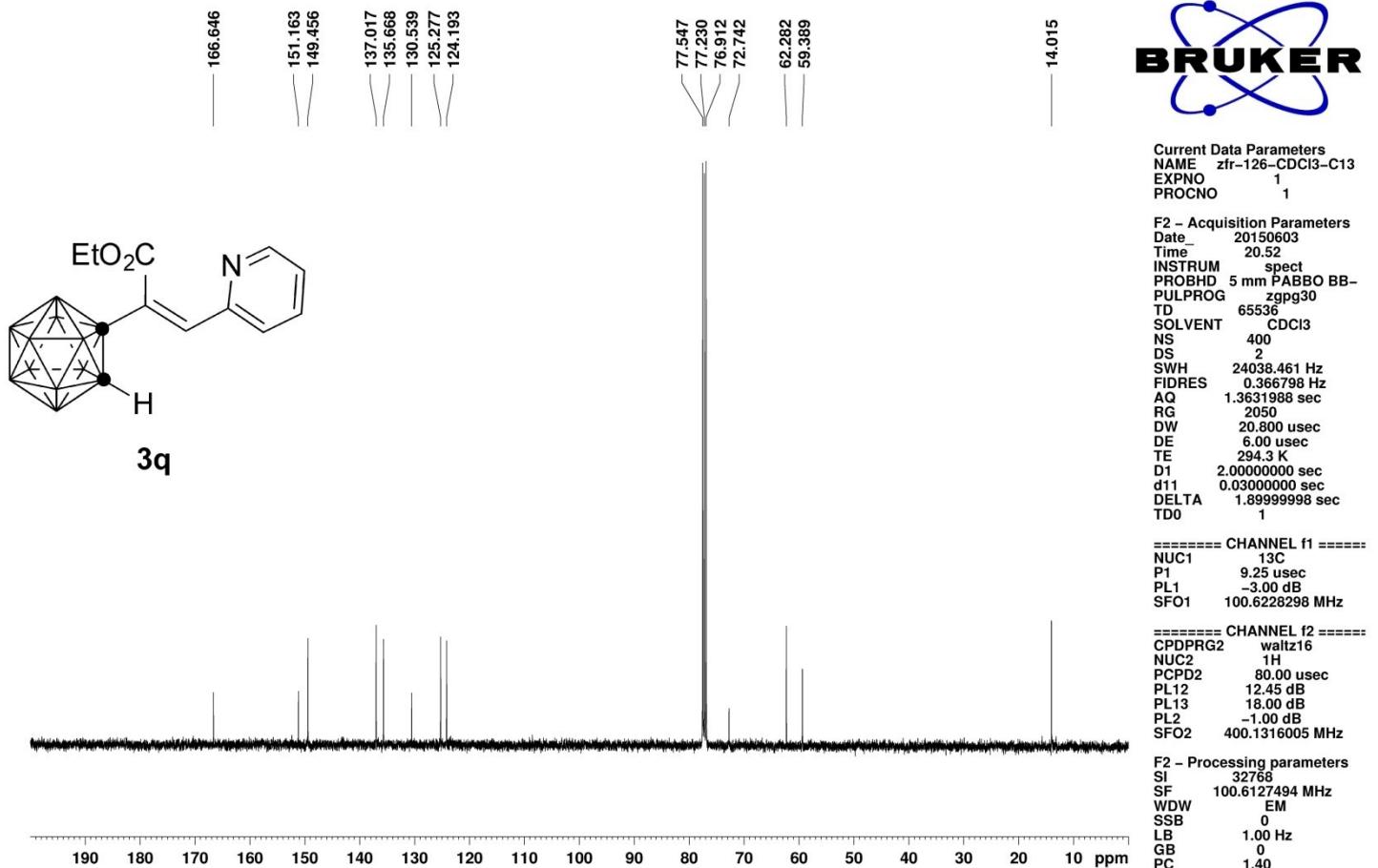


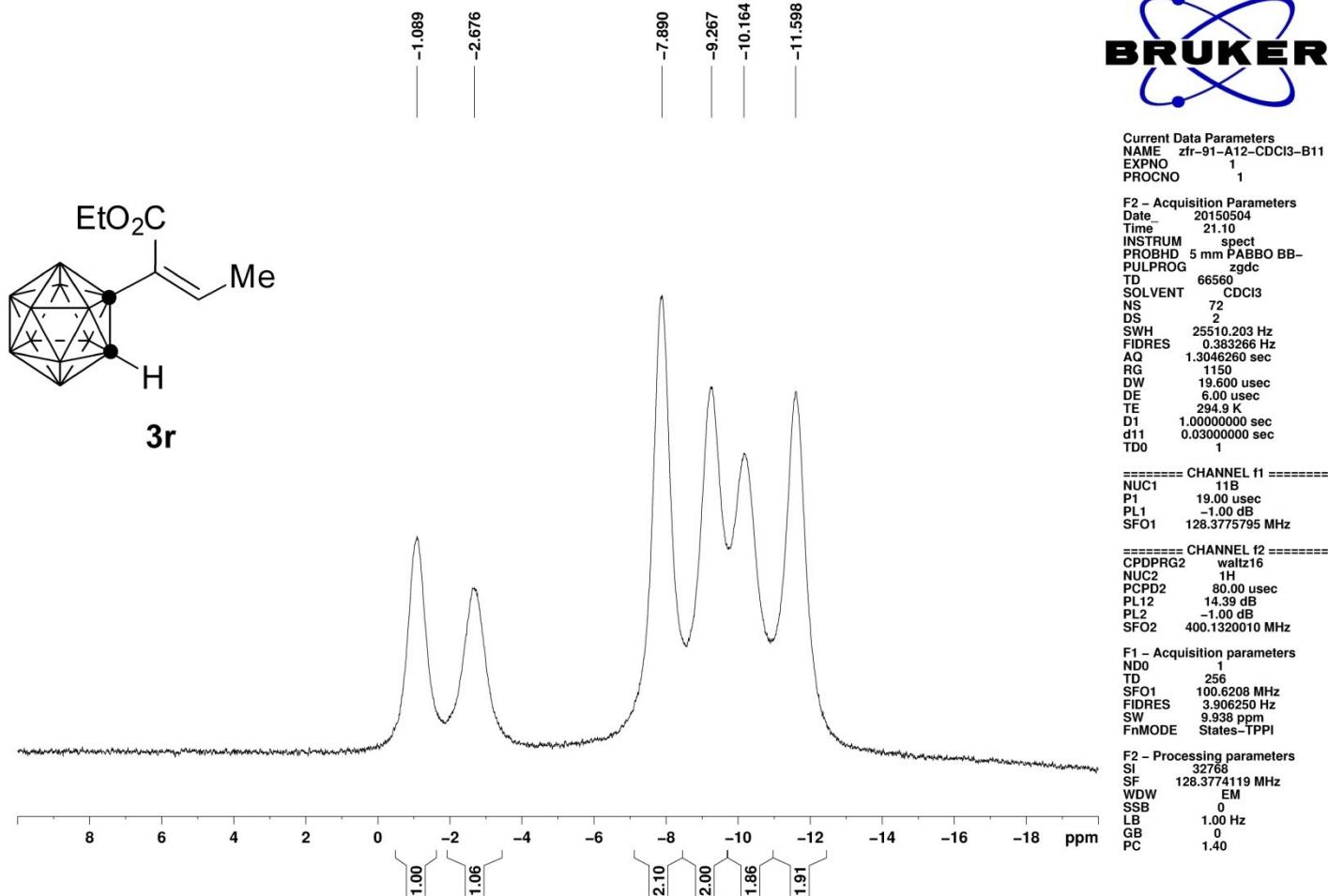
Current Data Parameters
 NAME zfr-126-CDCl3-H1
 EXPNO 2
 PROCNO 1

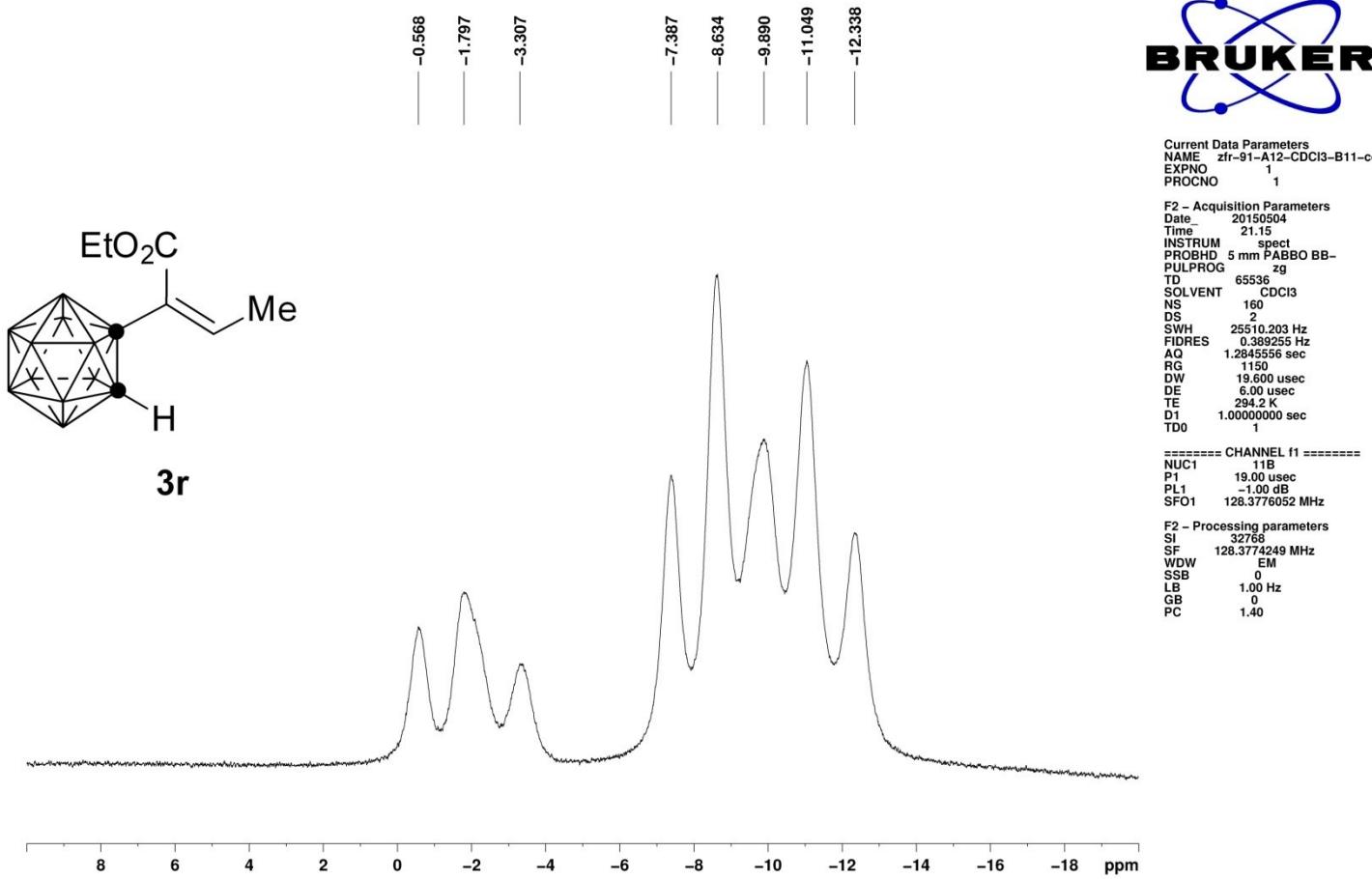
F2 – Acquisition Parameters
 Date 20150604
 Time 13.04
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 8223.685 Hz
 FIDRES 0.125483 Hz
 AQ 3.9846387 sec
 RG 203
 DW 60.800 usec
 DE 6.00 usec
 TE 293.5 K
 D1 1.0000000 sec
 TD0 1

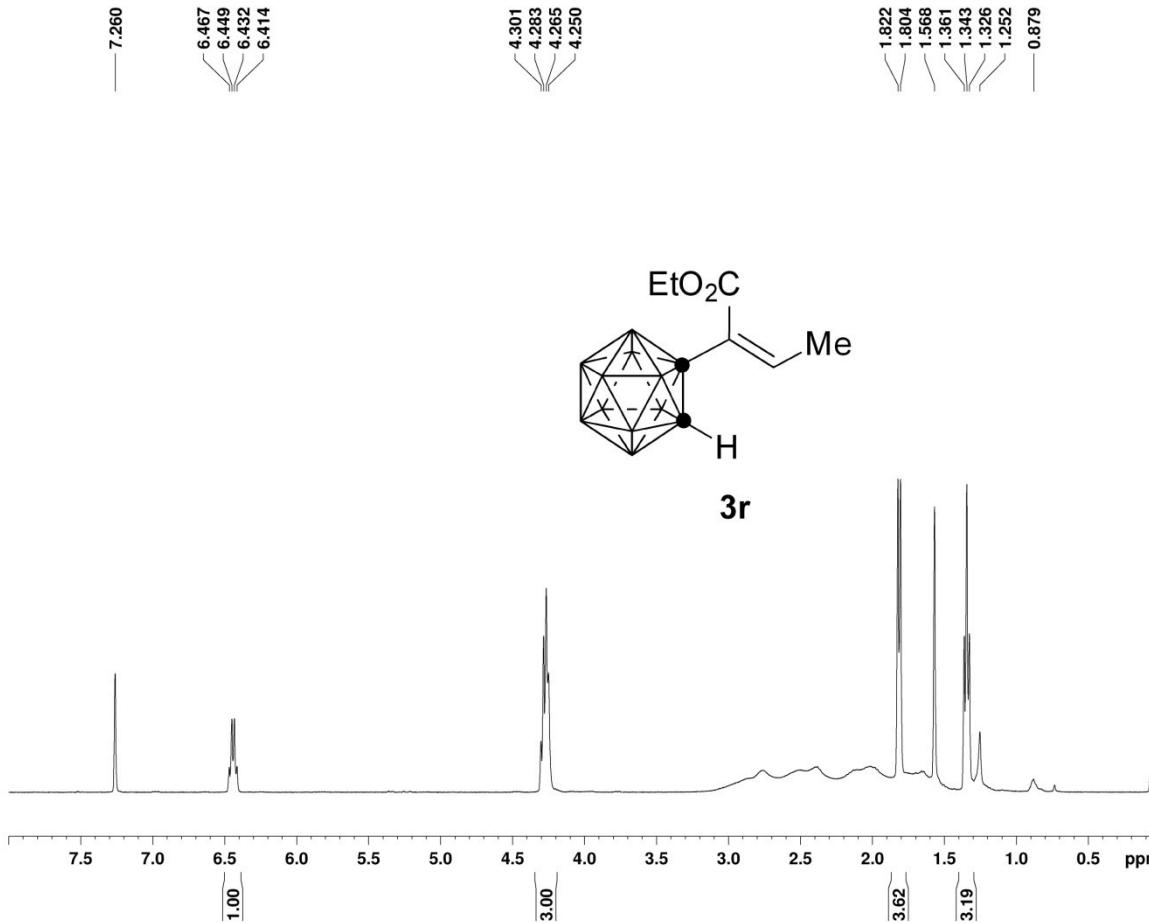
===== CHANNEL f1 =====
 NUC1 1H
 P1 13.60 usec
 PL1 -1.00 dB
 SFO1 400.1324710 MHz

F2 – Processing parameters
 SI 32768
 SF 400.1300053 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00







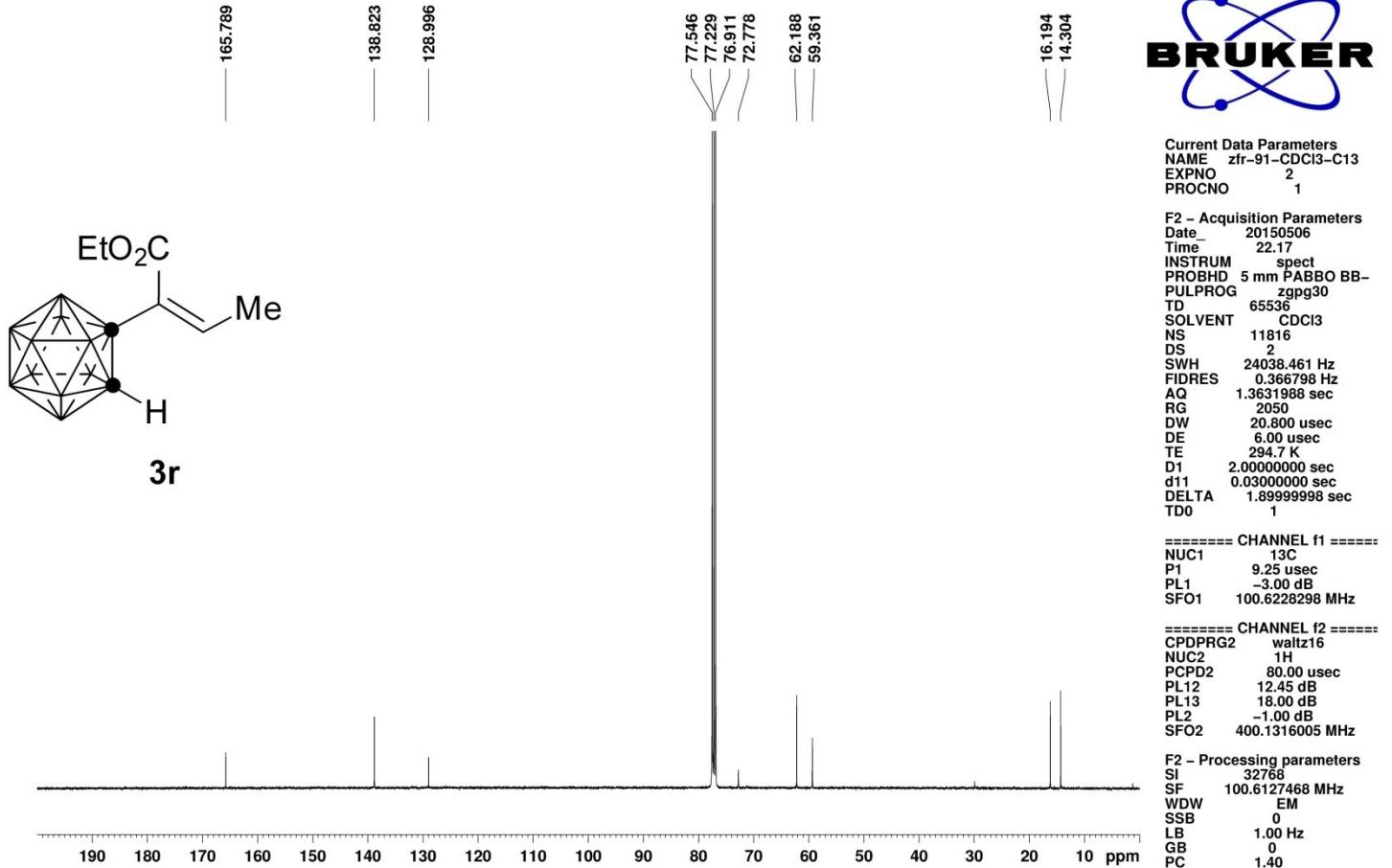


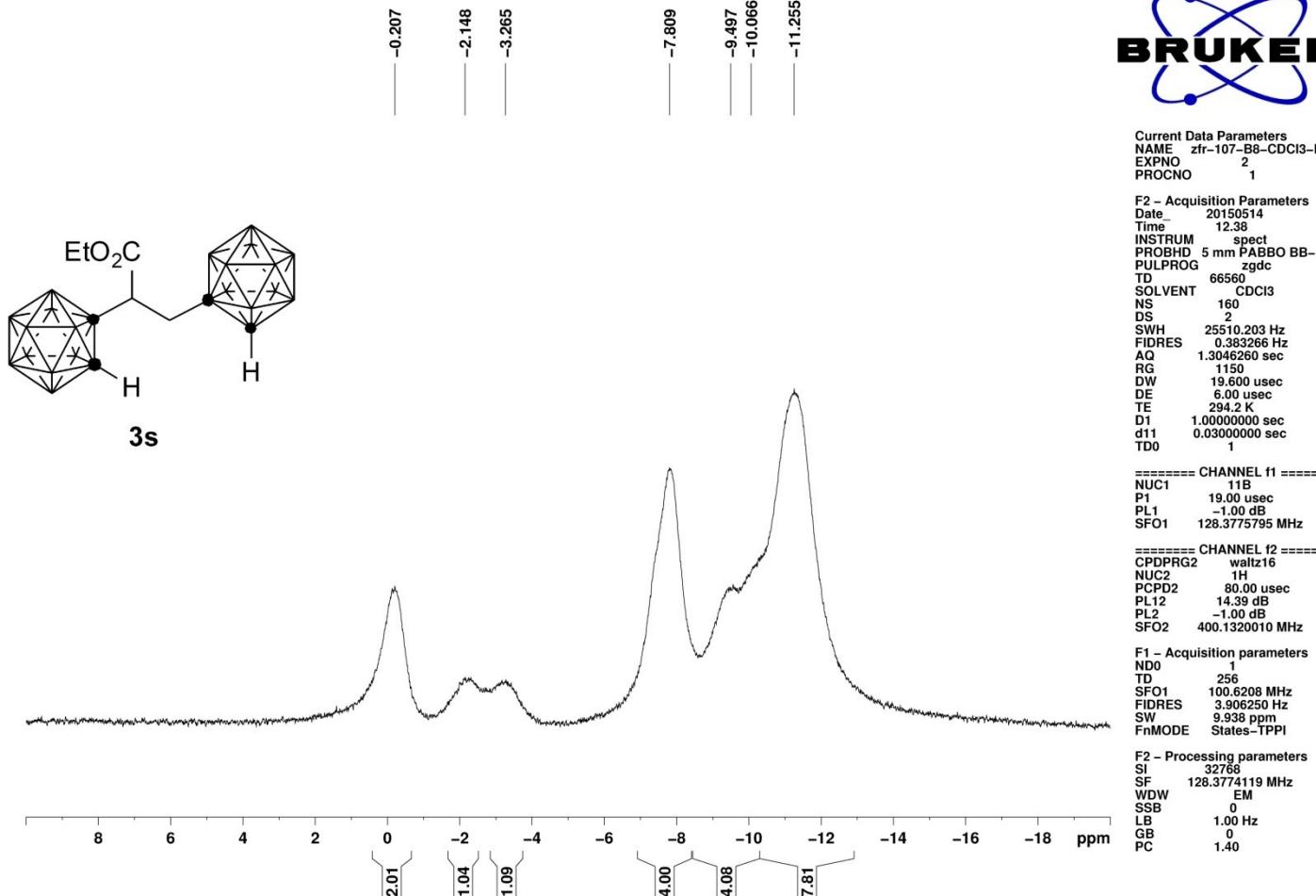
Current Data Parameters
 NAME zfr-91-A12-CDCl3-f
 EXPNO 1
 PROCNO 1

F2 – Acquisition Parameters
 Date_ 20150504
 Time 21.04
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 8223.685 Hz
 FIDRES 0.125483 Hz
 AQ 3.9846387 sec
 RG 456
 DW 60.800 usec
 DE 6.00 usec
 TE 294.1 K
 D1 1.0000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 13.60 usec
 PL1 -1.00 dB
 SFO1 400.1324710 MHz

F2 – Processing parameters
 SI 32768
 SF 400.1300050 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00





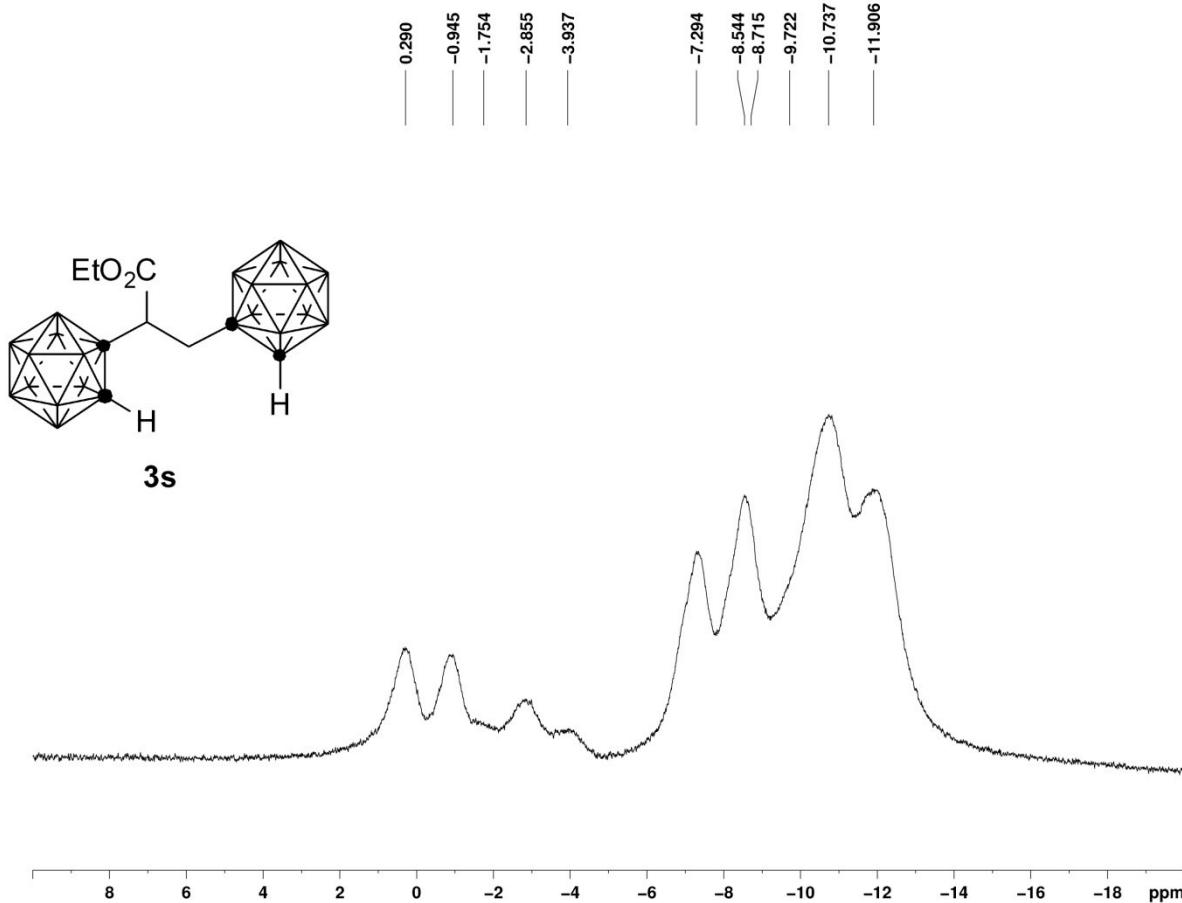


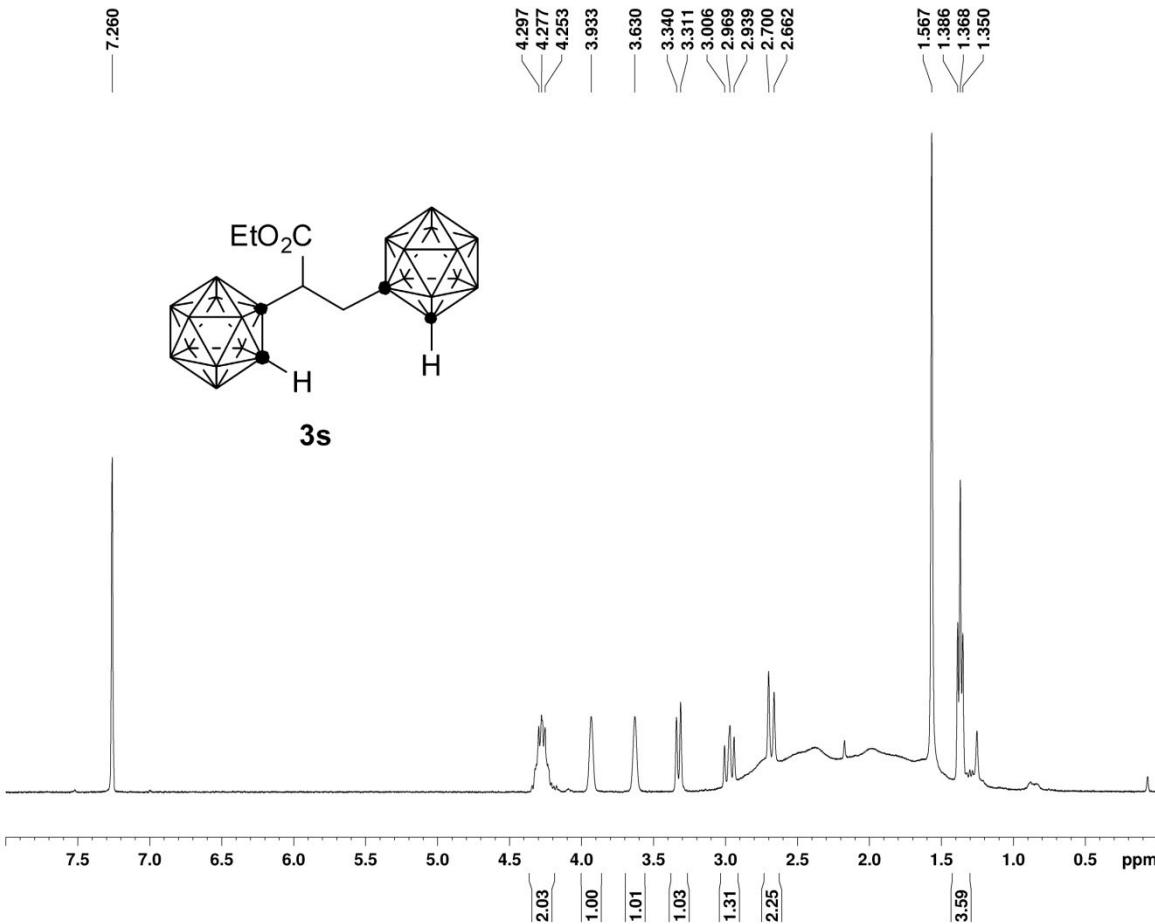
Current Data Parameters
NAME zfr-107-B8-CDCl3-B11-co
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date 20150514
Time 12.50
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg
TD 65536
SOLVENT CDCl3
NS 328
DS 2
SWH 25510.203 Hz
FIDRES 0.389255 Hz
AQ 1.2845556 sec
RG 1150
DW 19.600 usec
DE 6.00 usec
TE 293.5 K
D1 1.0000000 sec
TDO 1

===== CHANNEL f1 ======
NUC1 11B
P1 19.00 usec
PL1 -1.00 dB
SFO1 128.3776052 MHz

F2 - Processing parameters
SI 32768
SF 128.3774249 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



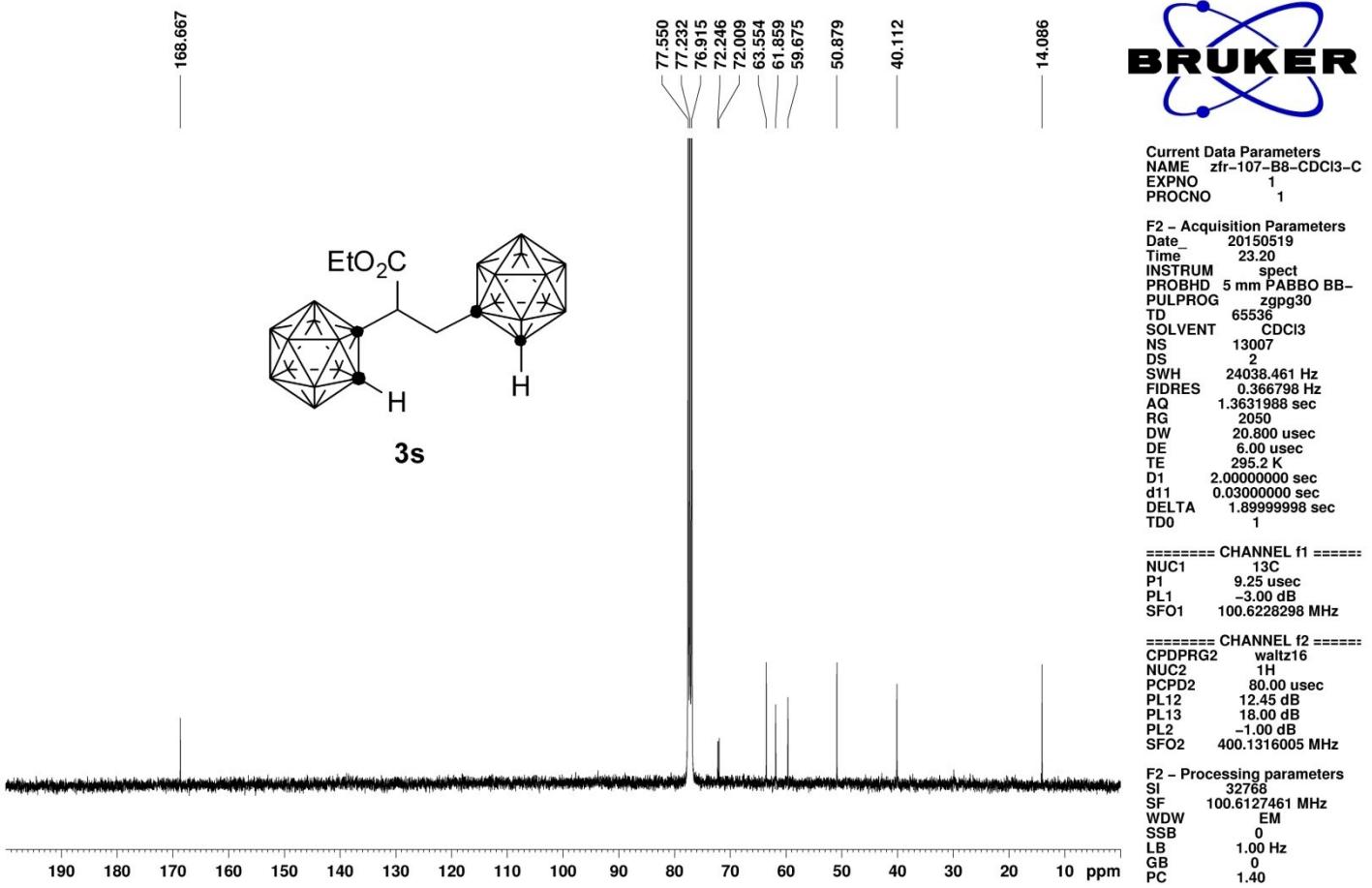


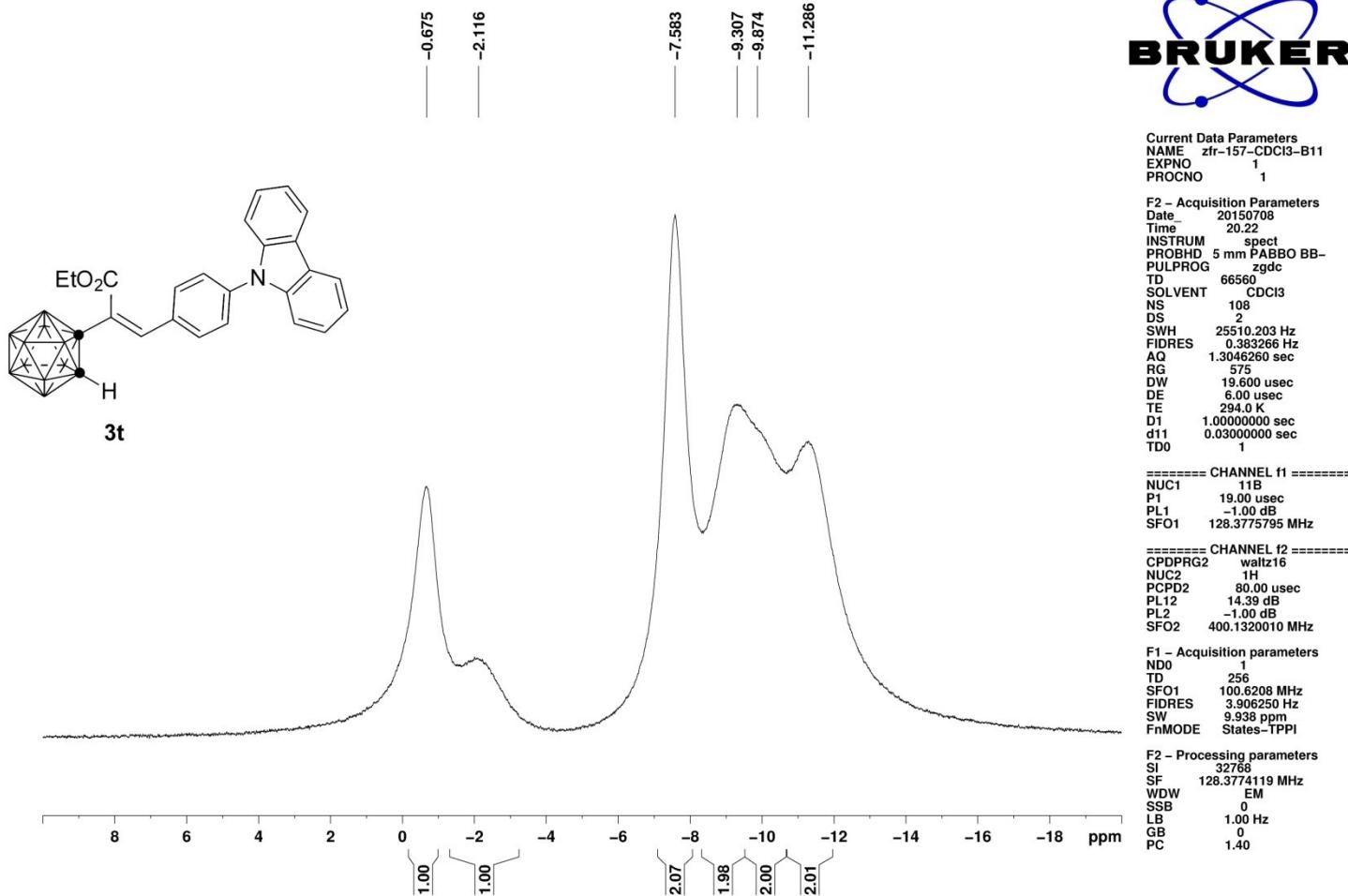
Current Data Parameters
 NAME zfr-107-B8-CDCl3-f
 EXPNO 2
 PROCNO 1

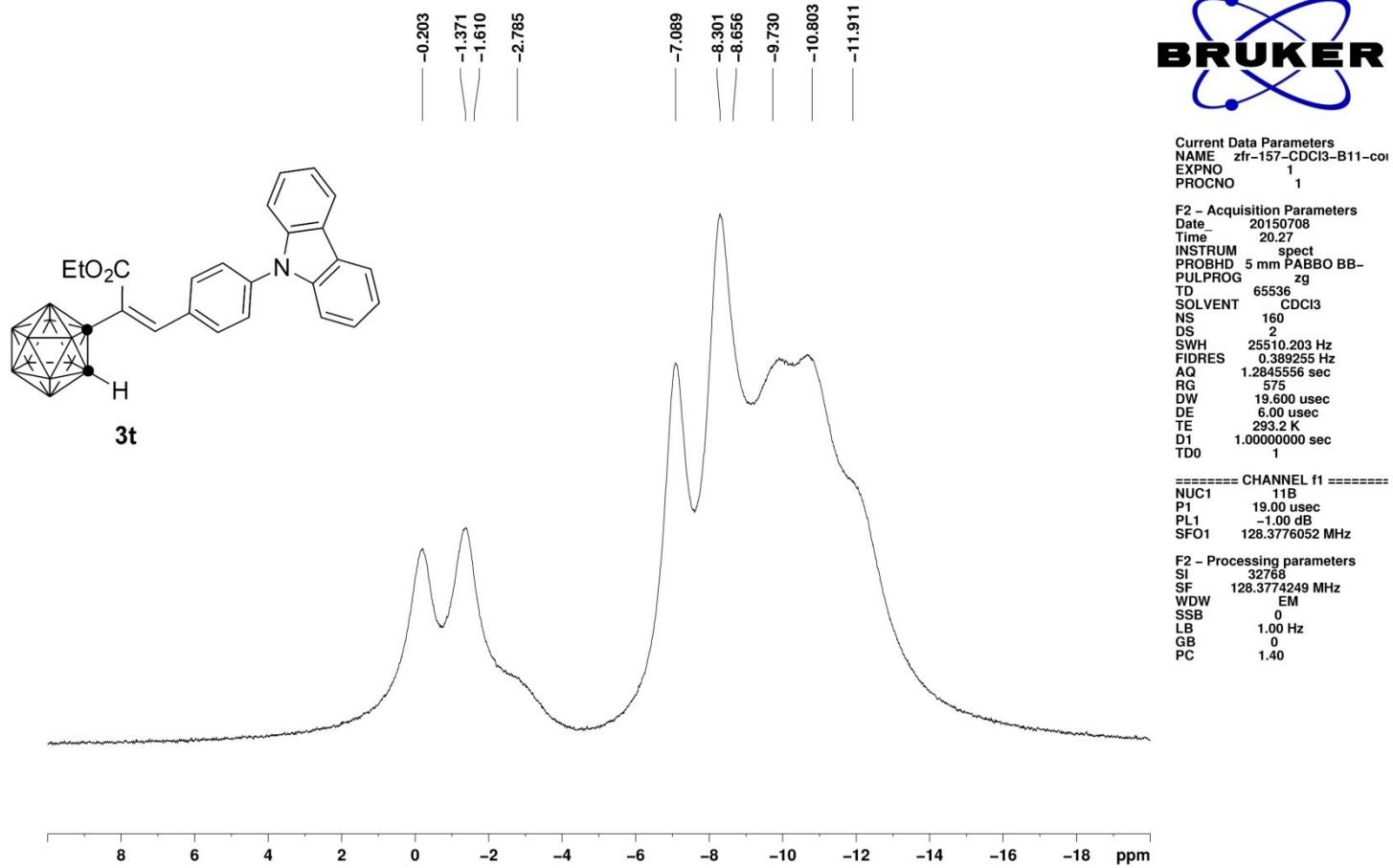
F2 - Acquisition Parameters
 Date 20150514
 Time 12.32
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 8223.685 Hz
 FIDRES 0.125483 Hz
 AQ 3.9846387 sec
 RG 512
 DW 60.800 usec
 DE 6.00 usec
 TE 293.4 K
 D1 1.0000000 sec
 TD0 1

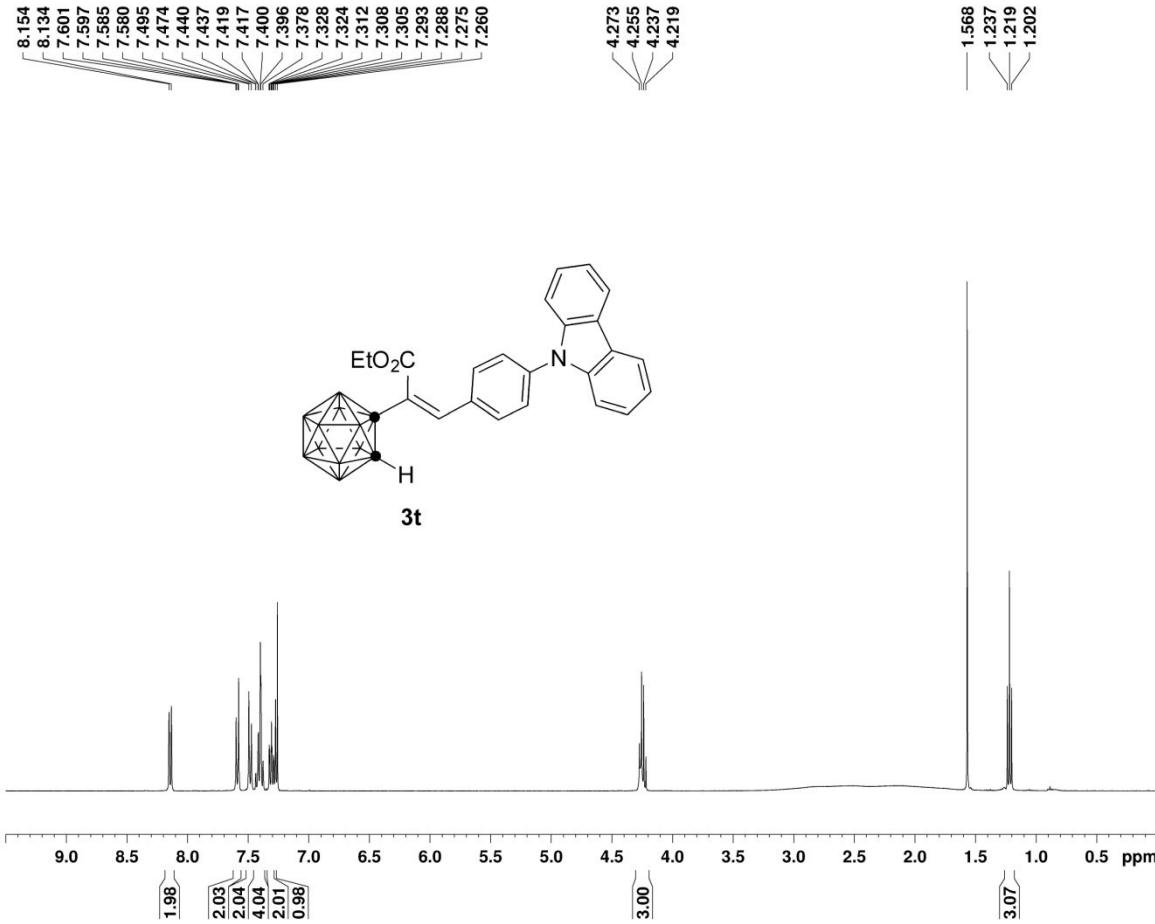
===== CHANNEL f1 =====
 NUC1 1H
 P1 13.60 usec
 PL1 -1.00 dB
 SFO1 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300051 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00







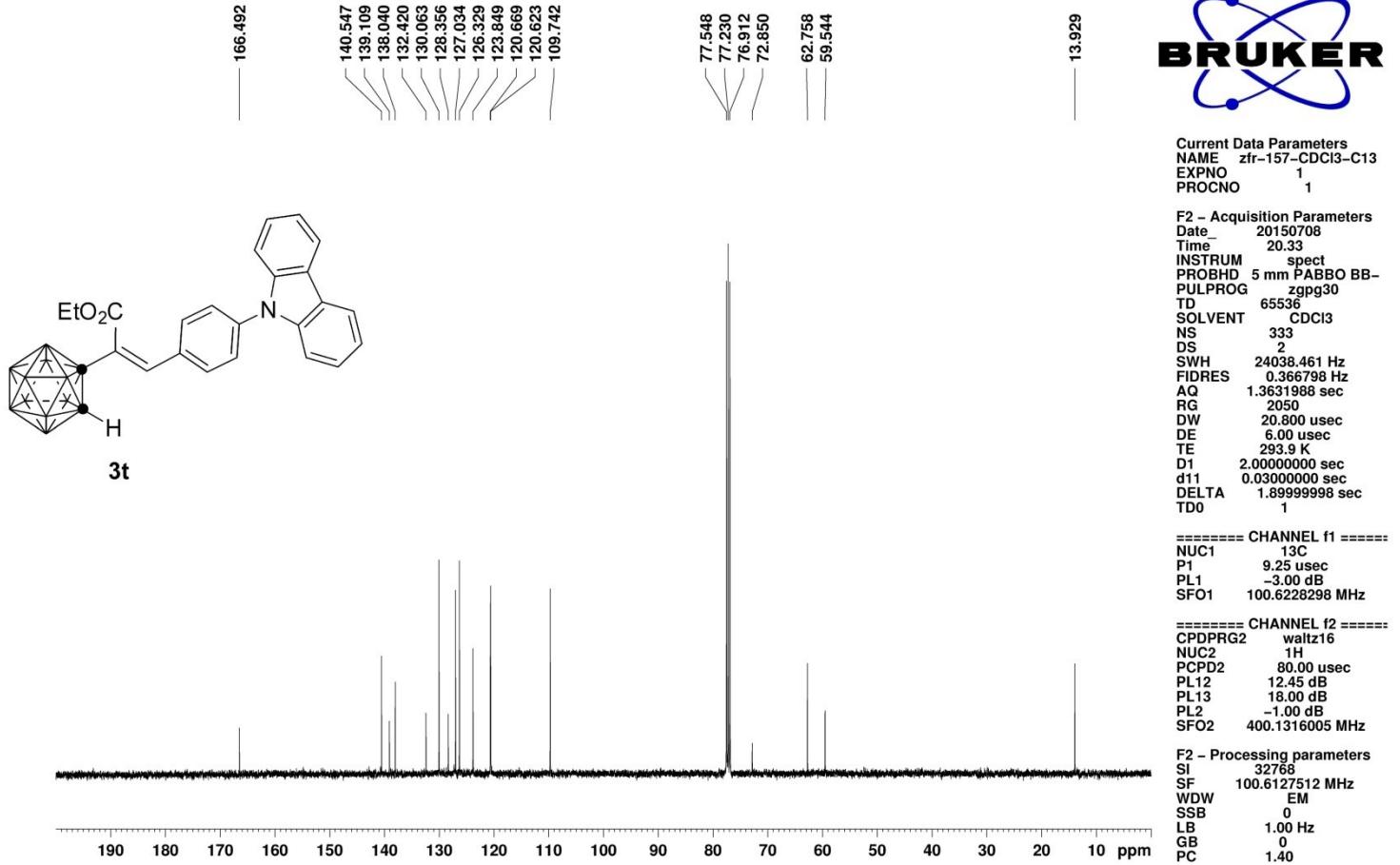


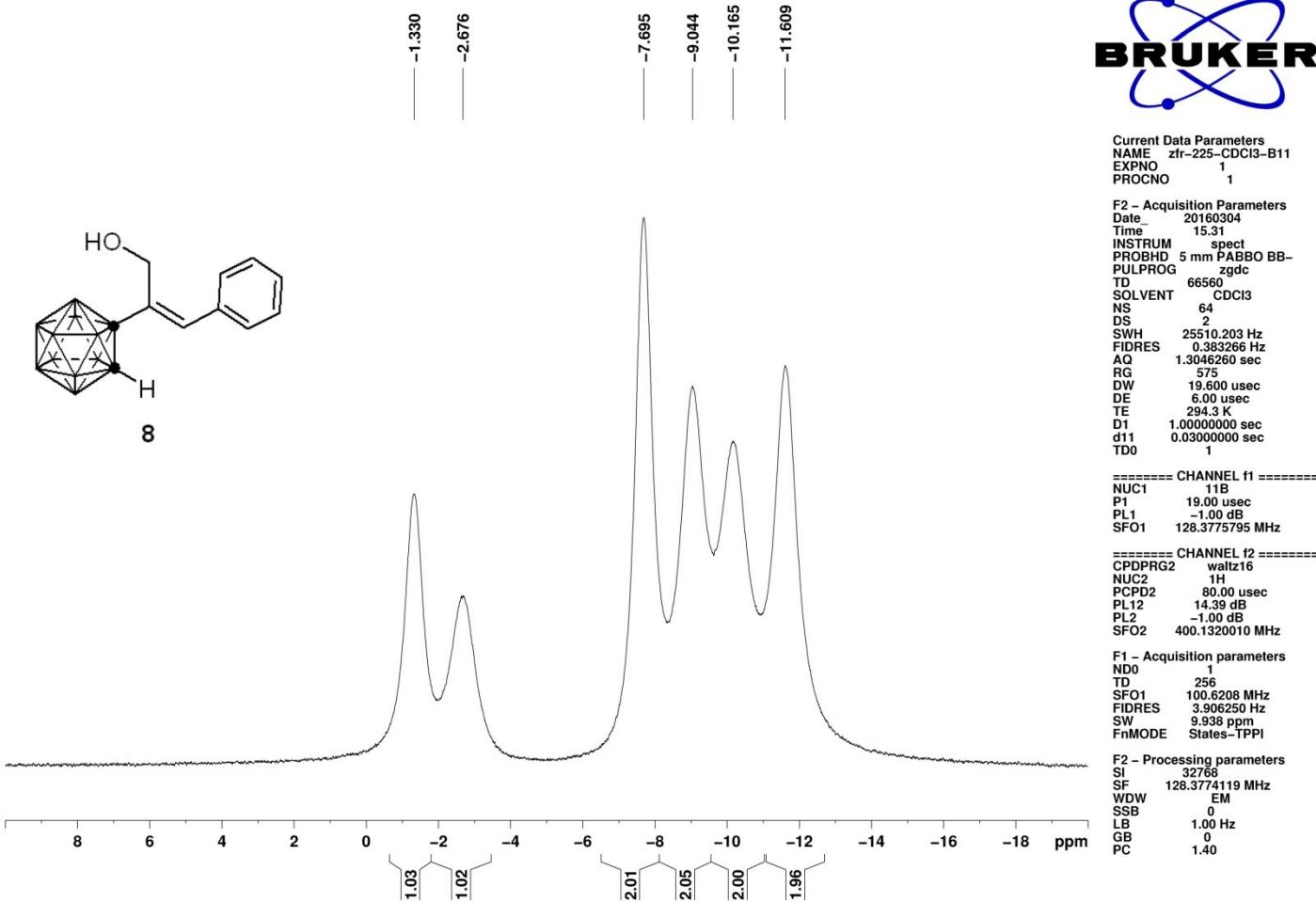
Current Data Parameters
 NAME zfr-157-CDCl3-H1
 EXPNO 2
 PROCNO 1

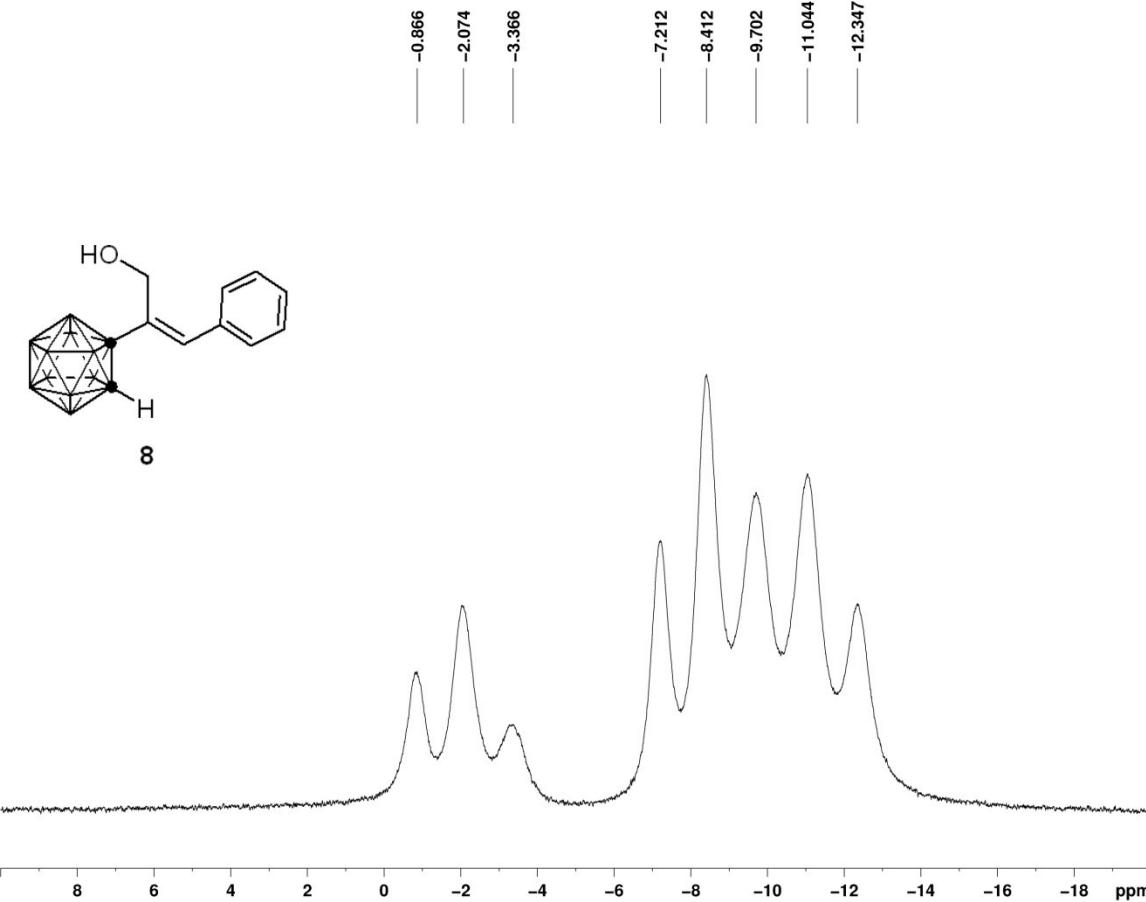
F2 – Acquisition Parameters
 Date 20150709
 Time 10.27
 INSTRUM spect
 PROBHD 5 mm PABBO BB-PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 8223.685 Hz
 FIDRES 0.125483 Hz
 AQ 3.9846387 sec
 RG 456
 DW 60.800 usec
 DE 6.00 usec
 TE 294.1 K
 D1 1.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 13.60 usec
 PL1 -1.00 dB
 SF01 400.1324710 MHz

F2 – Processing parameters
 SI 32768
 SF 400.1300053 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00





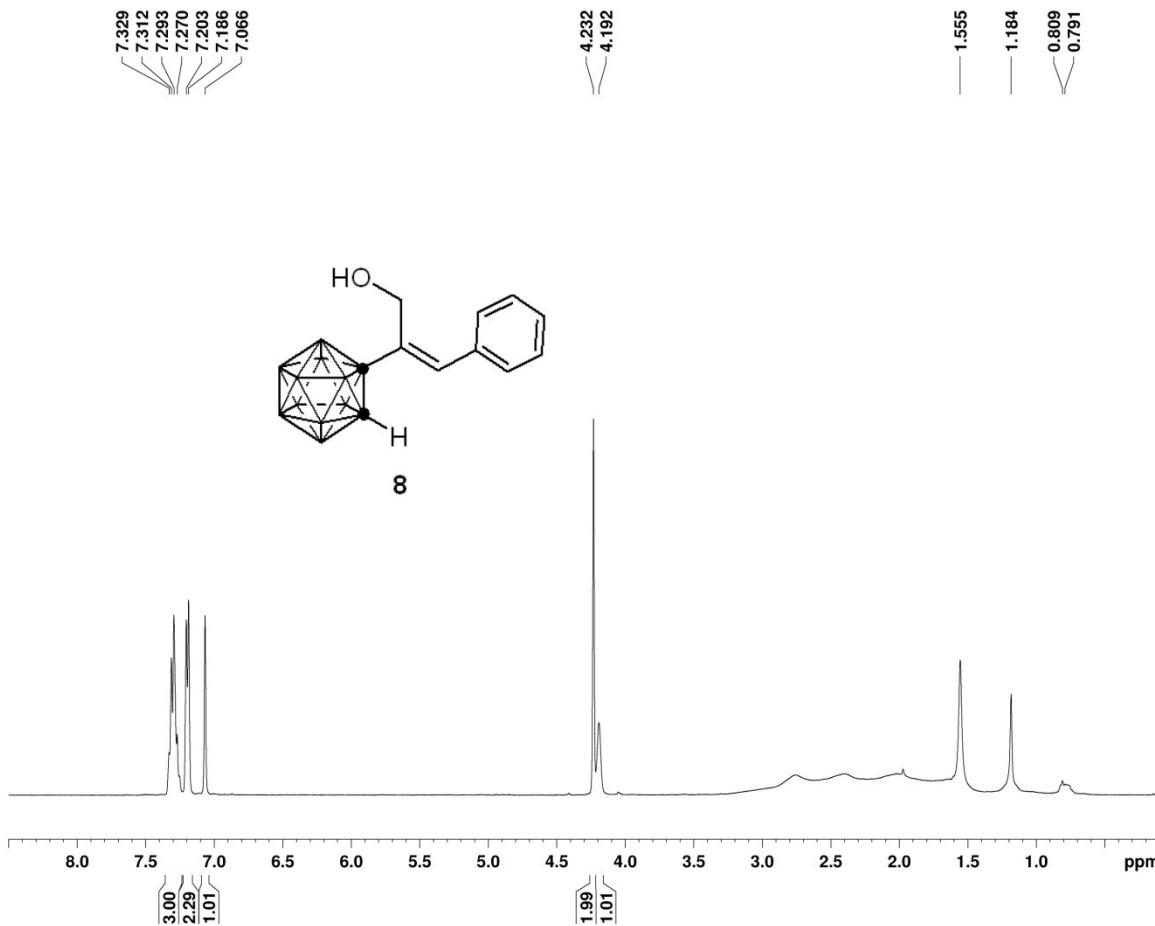


Current Data Parameters
 NAME TF-zfr-225
 EXPNO 3
 PROCNO 1

F2 – Acquisition Parameters
 Date 20160304
 Time 19.47
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg
 TD 65536
 SOLVENT CDCl₃
 NS 25
 DS 2
 SWH 25510.203 Hz
 FIDRES 0.389255 Hz
 AQ 1.2845556 sec
 RG 575
 DW 19.600 usec
 DE 6.00 usec
 TE 293.9 K
 D1 1.0000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 11B
 P1 19.00 usec
 PL1 -1.00 dB
 SFO1 128.3776052 MHz

F2 – Processing parameters
 SI 32768
 SF 128.3774249 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

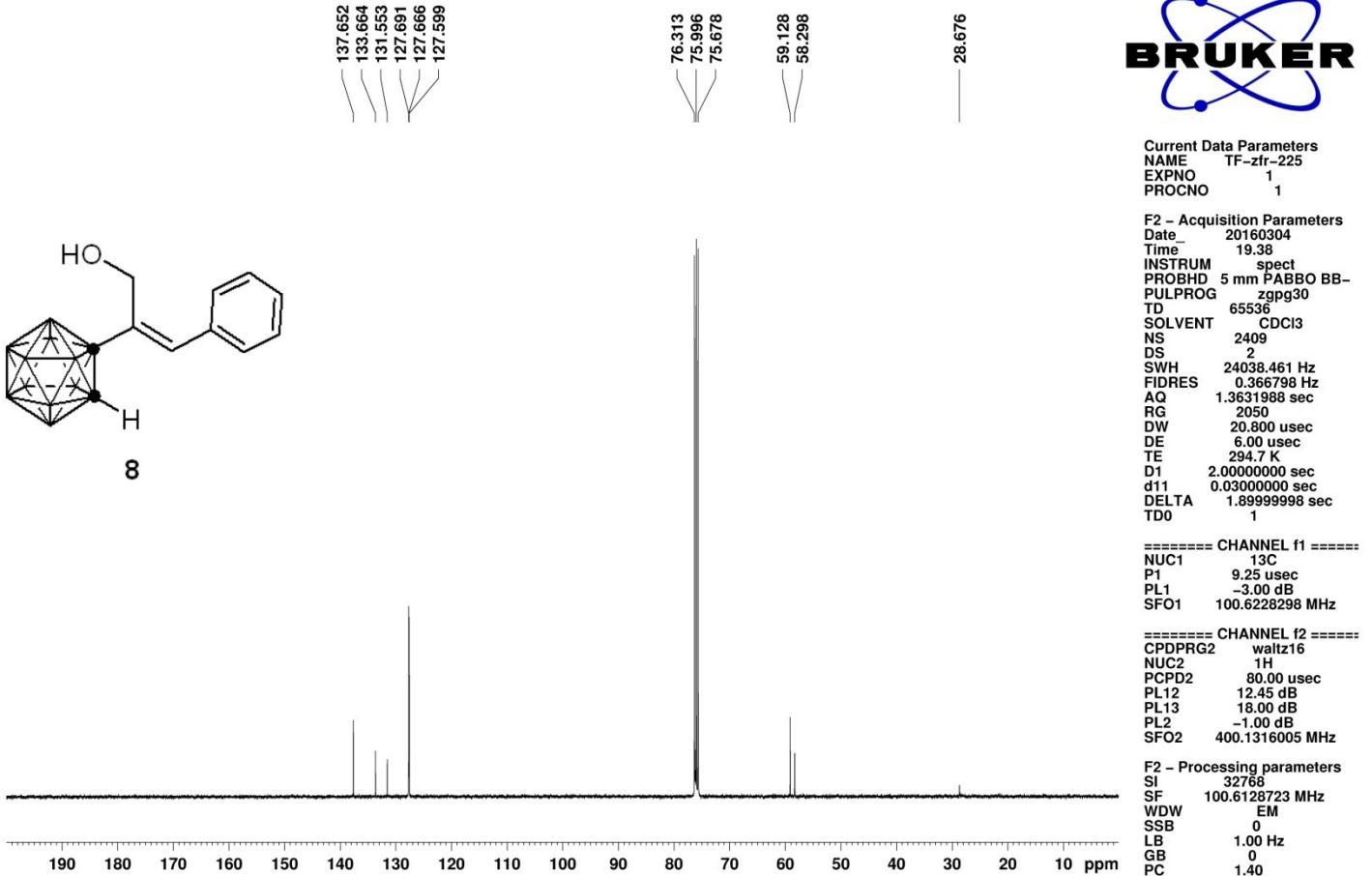


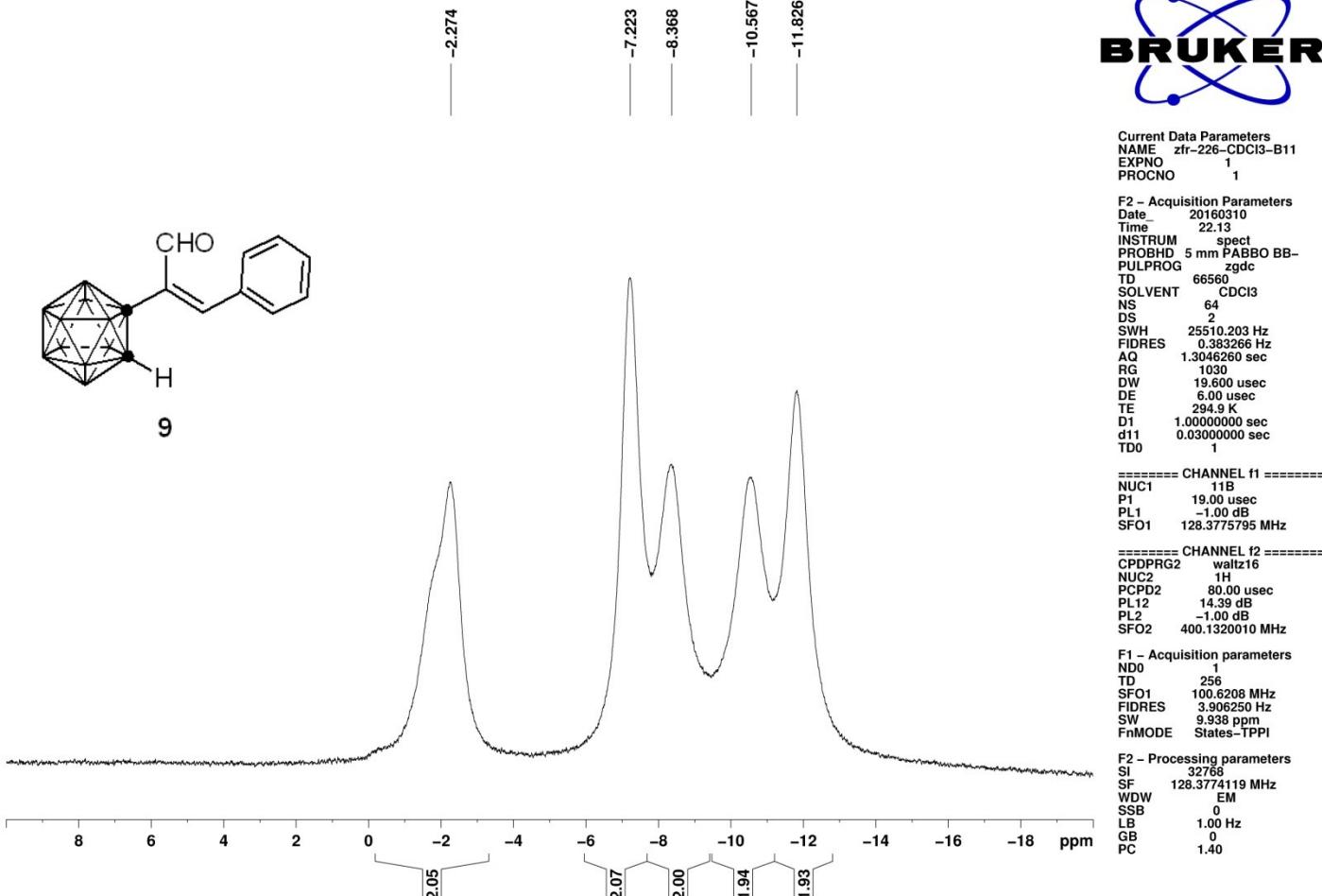
Current Data Parameters
 NAME zfr-225-CDCl₃-H1
 EXPNO 1
 PROCNO 1

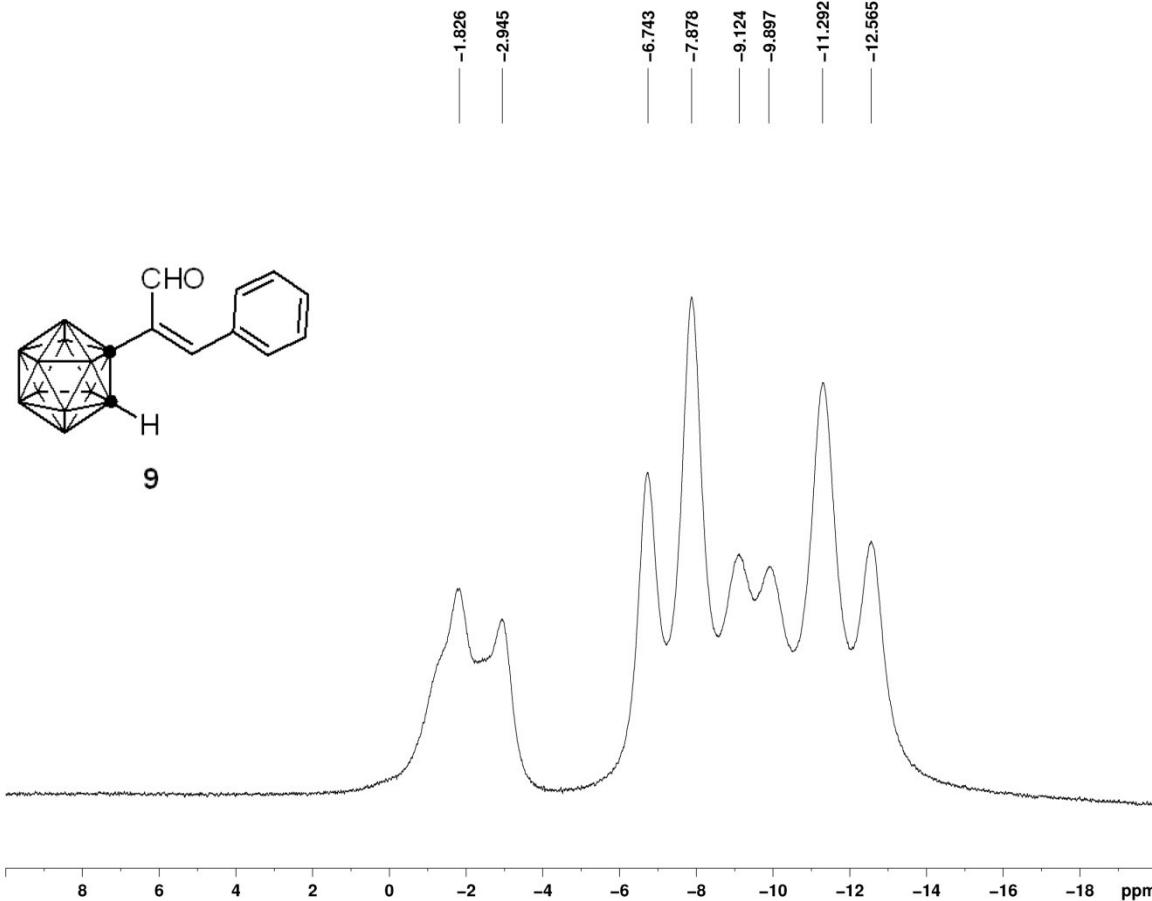
F2 – Acquisition Parameters
 Date 20160304
 Time 15.19
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 65536
 SOLVENT CDCl₃
 NS 16
 DS 2
 SWH 8223.665 Hz
 FIDRES 0.125483 Hz
 AQ 3.9846387 sec
 RG 256
 DW 60.800 usec
 DE 6.00 usec
 TE 293.7 K
 D1 1.0000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 13.60 usec
 PL1 -1.00 dB
 SFO1 400.1324710 MHz

F2 – Processing parameters
 SI 32768
 SF 400.1300345 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00





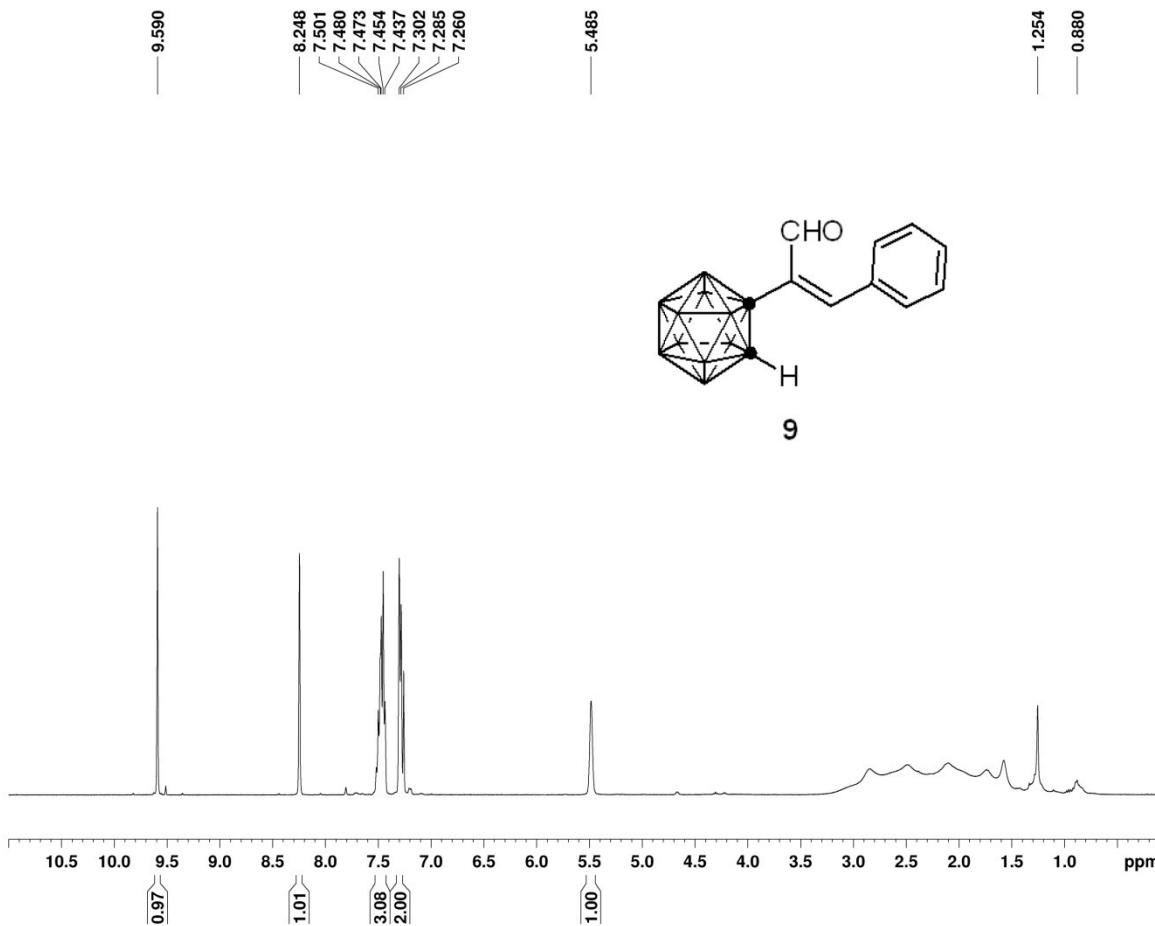


Current Data Parameters
 NAME zfr-226-CDCI3-B11-co1
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date 20160310
 Time 22.18
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg
 TD 65536
 SOLVENT CDCl3
 NS 160
 DS 2
 SWH 25510.203 Hz
 FIDRES 0.389255 Hz
 AQ 1.2845556 sec
 RG 1030
 DW 19.600 usec
 DE 6.00 usec
 TE 294.2 K
 D1 1.0000000 sec
 TD0 1

===== CHANNEL f1 ======
 NUC1 11B
 P1 19.00 usec
 PL1 -1.00 dB
 SF01 128.3776052 MHz

F2 - Processing parameters
 SI 32768
 SF 128.3774249 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

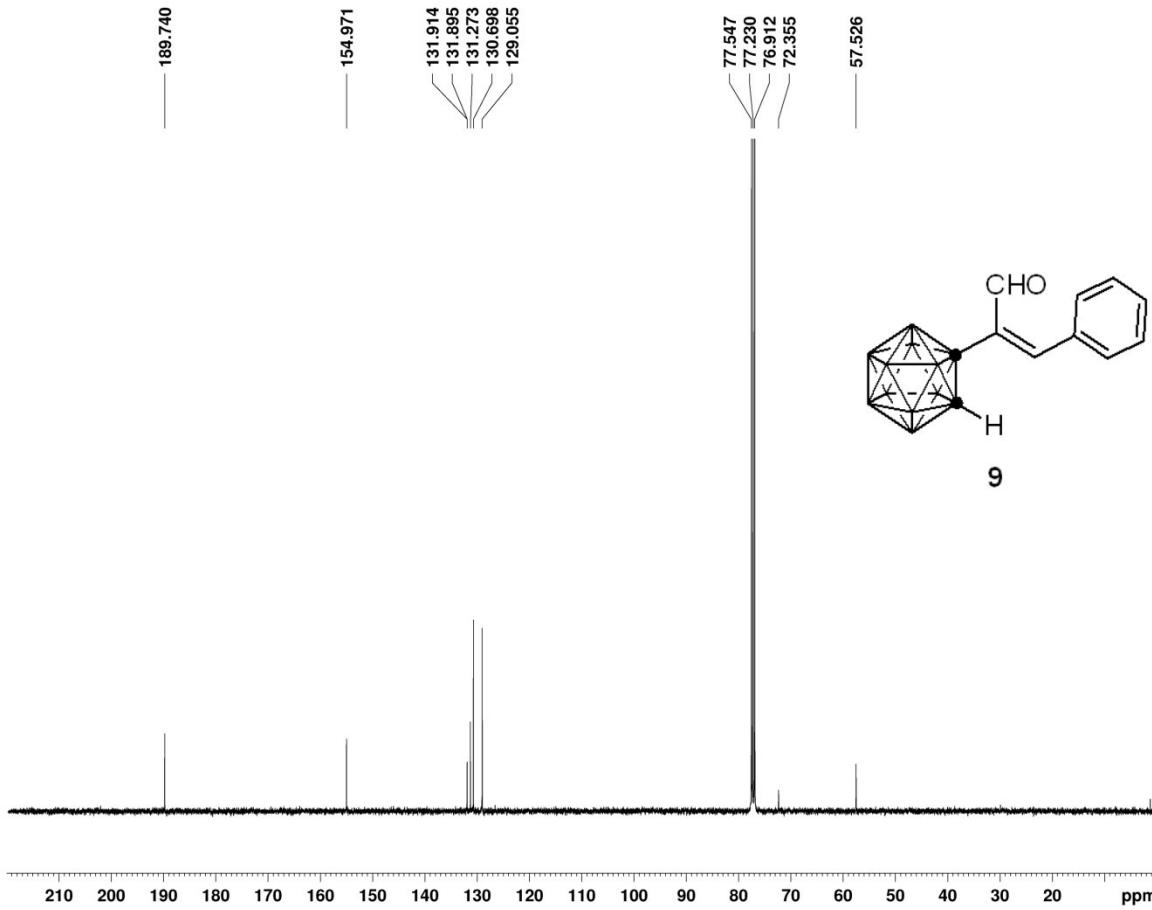


Current Data Parameters
 NAME zfr-226-CDCl₃-H1
 EXPNO 1
 PROCNO 1

F2 – Acquisition Parameters
 Date 20160310
 Time 22.08
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 65536
 SOLVENT CDCl₃
 NS 16
 DS 2
 SWH 8223.665 Hz
 FIDRES 0.125483 Hz
 AQ 3.9846387 sec
 RG 362
 DW 60.800 usec
 DE 6.00 usec
 TE 294.1 K
 D1 1.0000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 13.60 usec
 PL1 -1.00 dB
 SFO1 400.1324710 MHz

F2 – Processing parameters
 SI 32768
 SF 400.1300053 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



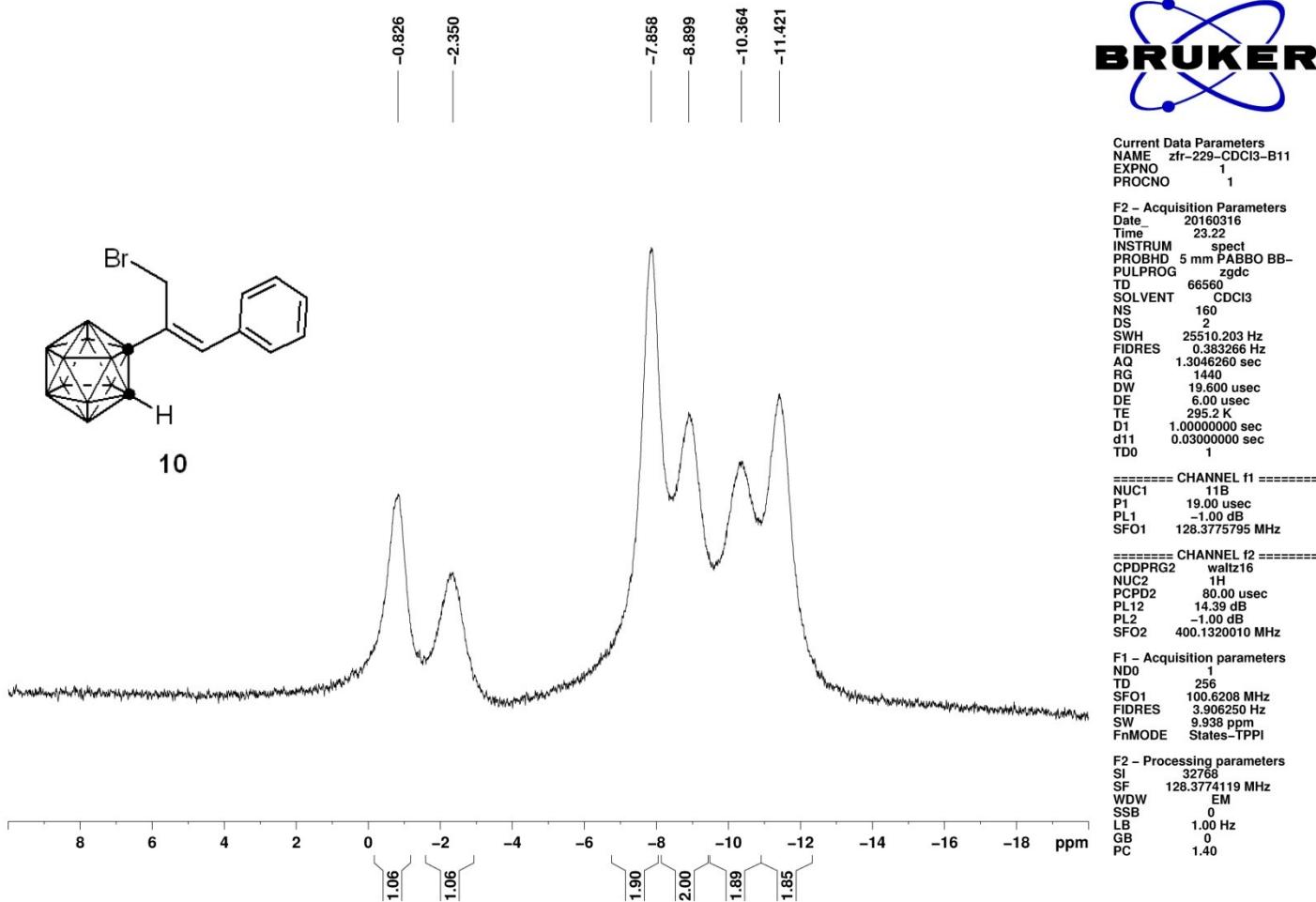
Current Data Parameters
 NAME zfr-226-CDCl₃-C13
 EXPNO 1
 PROCNO 1

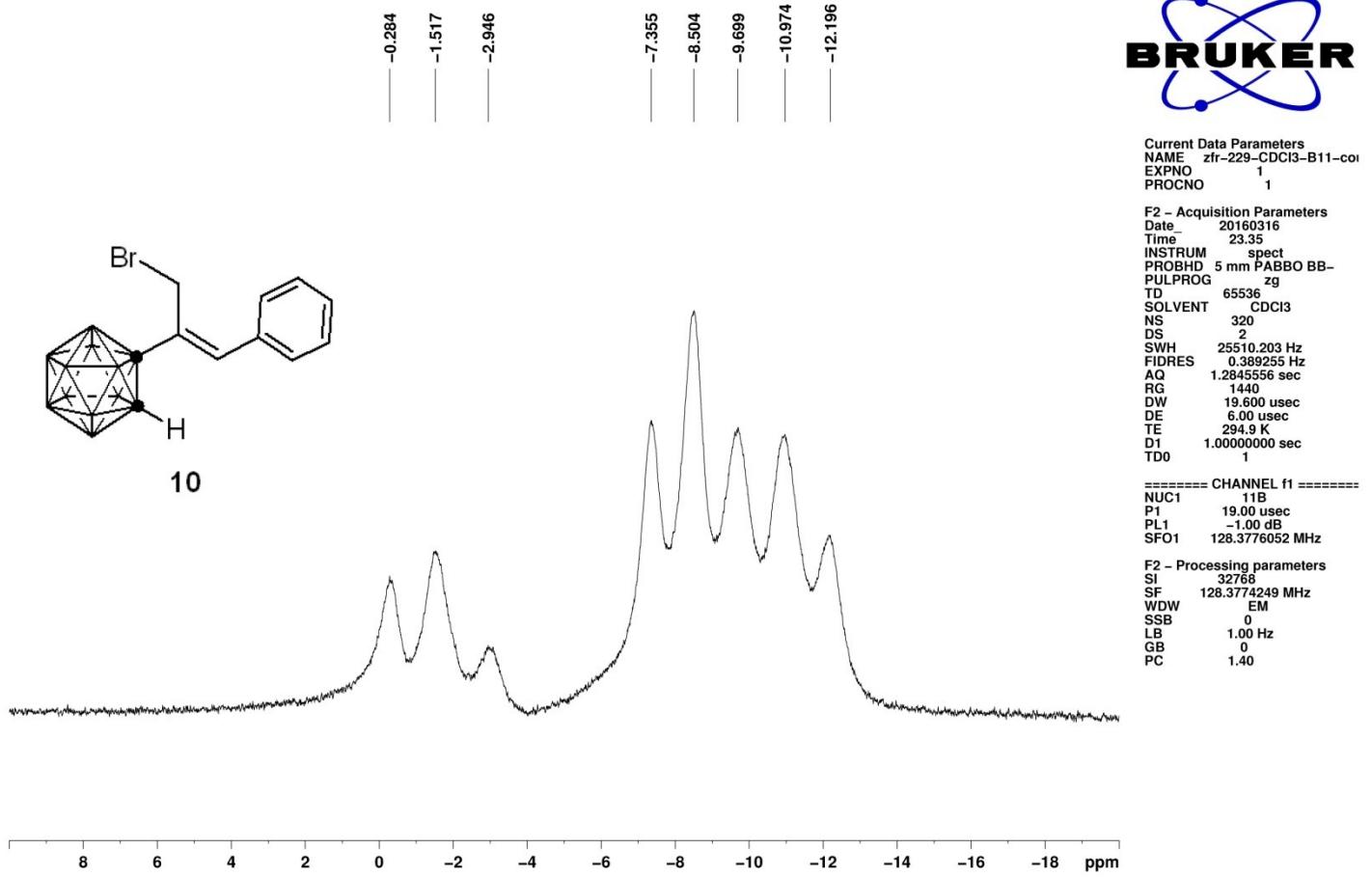
F2 - Acquisition Parameters
 Date 20160310
 Time 22:25
 INSTRUM spect
 PROBHD 5 mm PABBO BB-PULPROG zgpg30
 TD 65536
 SOLVENT CDCl₃
 NS 1194
 DS 2
 SWH 24038.461 Hz
 FIDRES 0.366798 Hz
 AQ 1.3631988 sec
 RG 2050
 DW 20.800 usec
 DE 6.00 usec
 TE 295.1 K
 D1 2.0000000 sec
 d11 0.03000000 sec
 DELTA 1.8999998 sec
 TD0 1

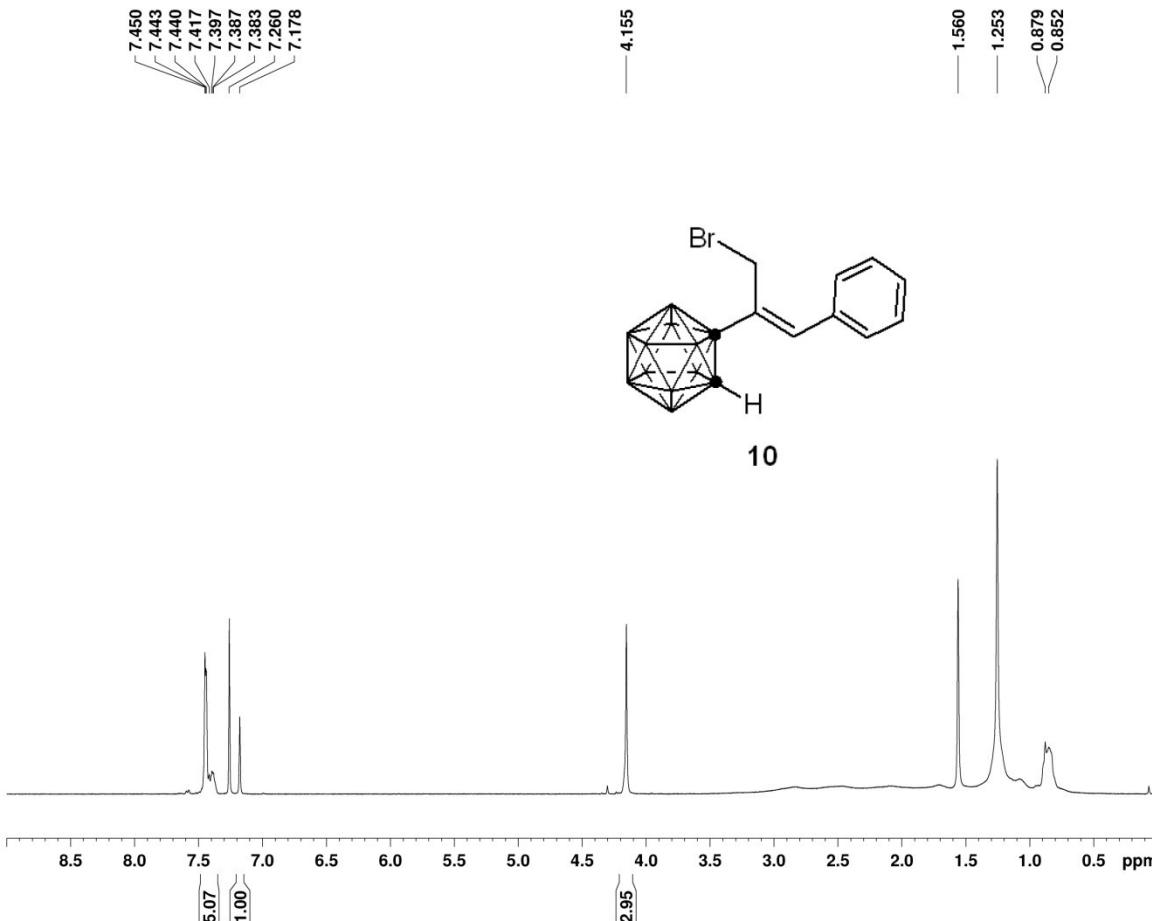
===== CHANNEL f1 =====
 NUC1 ¹³C
 P1 9.25 usec
 PL1 -3.00 dB
 SF01 100.6228298 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 ¹H
 PCPD2 80.00 usec
 PL12 12.45 dB
 PL13 18.00 dB
 PL2 -1.00 dB
 SF02 400.1316005 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6127476 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40







Current Data Parameters
 NAME zfr-229-CDCl3-H1
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date 20160317
 Time 19.01
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 8223.685 Hz
 FIDRES 0.125483 Hz
 AQ 3.9846387 sec
 RG 512
 DW 60.800 usec
 DE 6.00 usec
 TE 293.6 K
 D1 1.0000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 13.60 usec
 PL1 -1.00 dB
 SFO1 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300053 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

