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Supporting information for:

A Chiroptical Molecular Sensor for Ferrocene

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1) Synthetic procedures and characterization

General Experimental. All commercially available reagents and solvents were purchased from Sigma-Aldrich, Fluka and Alfa Aesar, and used as received. THF (Na, benzophenone) and CH₂Cl₂ (CaH₂) were dried and distilled before use. Analytical thin layer chromatography was performed on chromophore loaded, commercially available silica gel plates. Flash chromatography was carried out using silica gel (pore size 60 Å, 230-400 Mesh). ¹H- and ¹³C-NMR spectra were recorded from solutions in CDCl₃, d₆-DMSO or CD₃CN on 200, 300 or 500 MHz spectrometers using the solvent residual proton signal or tetramethylsilane as the internal standard. 2D ROESY NMR experiments were performed applying the following parameters: pulse programme: roesyph from Bruker library, T= 300 K, 1024x256 data points, NS= 128, D1= 1 s, spin-lock (mixing) time= 300 ms. Spectra were processed with Topspin 2.1.Samples for the mass spectrometry were analysed as FIA-ESI/MS with an ion trap mass spectrometer ThermoScientific LCQ FLEETin the following conditions: spray voltage=4.5 kV, capillar temperature=220°C, capillar voltage=19 V, tube lens=120 V, mass interval=50-2000 Da, mode=positive ions. The UV/Vis spectroscopic studies were recorded using commercially-available spectrophotometers, with a scanning speed of 400 nm/min at room temperature. CD spectroscopy was performed using a commercially-available spectropolarimeter. CD spectra were recorded at 25 °C and with the following parameters: scanning speed= 50 nm/min, data pitch= 0.5 nm, response= 4 seconds, band width= 1 nm. The spectra were then background corrected. Cyclic voltammetry were recorded on BASi PWR-3 power module and EF-1085 C-3 cell stand, equipped with a glassy carbon electrode (diameter 2.0 mm) as working electrode; an Ag/AgCl/NaCl (3 M NaCl, saturated with AgCl) reference electrode and a platinum wire as auxiliary electrode, both obtained from BASi, were used.

The overall synthesis for the compounds of interest is shown in Scheme S1. Compound (*R*)-**3** was synthesized according to ref. S1. $Pd(en)(NO_3)_2$ was prepared according to reference S2.



Figure S1. Scheme for the synthesis of compound $[Pd(en)(NO_3)_2]$ (a) and metal-linked macrocycles (*RR*)-6 and (b) (*RR*)-7.

Compound (*R*)-4. Cs₂CO₃ (0.403 g, 1.24 mmol, 4 eq) and 3-pyridineboronic acid pinacol ester (0.151 g, 0.74 mmol, 2.5 eq) were added to a solution of compound (*R*)-3 (0.139 g, 0.3 mmol, 1 eq) in dry THF (40 mL), keeping N₂ atmosphere and continuous stirring. After 15 min, Pd(PPh₃)₄ (0.075 g, 0.06 mmol, 0.2 eq) was added to the round-bottom flask and the dark yellow solution was stirred overnight under reflux. The reaction was monitored by TLC (AcOEt:Hexane:Et₃N 6:3:1) and the mixture was quenched with H₂O (150 mL), extracted with AcOEt (3x 150 mL) and the yellow organic phase dried (Na₂SO₄). The solution was

filtered and concentrated *in vacuo*, and the crude product was purified by flash chromatography (AcOEt:hexanes:Et₃N 6:3:1) to yield compound (*R*)-4 (0.14 g, 0.29 mmol, 96%) as a yellow oil. ¹H-NMR (CDCl₃, 200 MHz, 25 °C) δ = 8.95 (s, 2H; pyridine), 8.59 (m, 2H; pyridine), 8.10 (d, 2H; binaphthyl), 8.08 (d, 2H; binaphthyl), 7.95 (s, 2H; pyridine), 7.54 (d, 2H; binaphthyl), 7.48 (m, 2H; pyridine), 7.37 (dd, 2H; binaphthyl), 7.26 (d, 2H; binaphthyl), 3.82 (s, 6H; -OCH₃).¹³C-NMR (CDCl₃, 75 MHz, 25 °C) δ = 155.4 (C_{quat}), 148.3 (CH), 148.1 (CH), 136.5 (C_{quat}), 134.2 (CH), 133.4 (C_{quat}), 132.7 (C_{quat}), 129.9 (CH), 129.2 (C_{quat}), 126.2 (CH), 126.05 125.5 (CH), 123.5 (CH), 119.1 (C_{quat}), 114.7 (CH), 56.7 (CH₃).

Compound (*R*)-**5.** Cs₂CO₃ (0.389 g, 1.2 mmol, 4 eq) and 4-pyridineboronic acid pinacol ester (0.154 g, 0.75 mmol, 2.5 eq) were added to a solution of compound (*R*)-**3** (0.137 g, 0.3 mmol, 1 eq) in dry THF (40 mL), keeping N₂ atmosphere and continuous stirring. After 15 min, Pd(PPh₃)₄ (0.077 g, 0.07 mmol, 0.2 eq) was added to the round-bottom flask and the dark yellow solution was stirred overnight under reflux. The end of the reaction was monitored by TLC (AcOEt:Hexane:Et₃N 6:3:1) and the mixture was quenched with H₂O (150 mL), extracted with AcOEt (3x 150 mL) and the yellow organic phase dried (Na₂SO₄). The solution was filtered and concentrated in vacuo, and the crude product was purified by flash chromatography (AcOEt:Hexane:Et₃N 6:3:1) to yield compound (*R*)-**5** (0.13 g, 0.27 mmol, 90%) as a yellow solid. ¹H-NMR (CDCl₃, 200 MHz, 25 °C) δ = 8.67 (d, 4H; pyridine), 8.19 (s, 2H; binaphthyl), 8.11 (d, 2H; binaphthyl), 7.61 (d, 4H; pyridine), 7.57-7.50 (m, 4H; binaphthyl), 7.27 (d, 2H; binaphthyl), 3.83 (s, 6H; -OCH₃). ¹³C-NMR (CDCl₃, 75 MHz, 25 °C) δ = 155.7 (C_{quat}), 150.0 (CH), 148.2 (C_{quat}), 134.0 (C_{quat}), 132.9 (C_{quat}), 130.2 (CH), 129.0 (C_{quat}), 126.5 (CH), 126.0 (CH), 125.0 (CH), 121.5 (CH), 119.0 (C_{quat}), 114.6 (CH), 56.7 (CH₃).

Compound (*RR*)-**6.** A suspension of Pd(en)(NO₃)₂(0.034g, 0.12 mmol, 1 eq) in H₂O (10 mL) was stirred at room temperature. A solution of compound (*R*)-**4** (0.051 g, 0.11 mmol, 1 eq) in THF (5 mL) was added and the reaction mixture was then heated at 70°C for 30 minutes. The solution is clear and yellow. An aqueous solution of NH₄PF₆ (1M, 2.5 mL) was added dropwise and the solution became turbid. A dark agglomerate was formed. After additional 15 min stirring, the solution was filtered to yield compound (*RR*)-**6** (0.05 g, 0.03 mmol, 56%), as a light brown solid. ¹H-NMR (*d*₆-DMSO, 200 MHz, 25 °C) δ = 9.49 (s, 4H; pyridine), 8.95 (s, 4H; pyridine), 8.45 (s, 4H; binaphthyl), 8.39 (d, 4H; pyridine), 8.15 (d, 4H; binaphthyl), 7.79-7.72 (m, 8H; binaphthyl), 7.50 (d, 4H; pyridine), 6.98 (d, 2H; binaphthyl),

5.60 (broad peak, 8H; -NH₂), 3.78 (s, 12H; -OCH₃), 2.71 (broad singlet, 8H; -CH₂-). ¹³C-NMR (CD₃CN, 75 MHz, 25 °C) δ = 156.0 (C_{quat}), 149.8 (CH), 149.5 (CH), 139.2 (C_{quat}), 138.3 (CH), 133.6 (C_{quat}), 130.4 (CH), 129.7 (C_{quat}), 129.0 (C_{quat}), 127.2 (CH), 126.8 (CH), 125.6 (CH), 124.7 (CH), 118.3 (C_{quat}), 115.0 (CH), 56.1 (CH₃), 46.7 (CH₂). ESI-MS *m*/*z* = 1704.11 [M-PF₆]⁺.

Compound (*RR*)-7. A suspension of compound Pd(en)(NO₃)₂(0.034g, 0.12 mmol, 1 eq) in water (10 mL) was stirred at room temperature. A solution of compound (*R*)-**5** (0.052 g, 0.11 mmol, 1 eq) in THF (5 mL) was added and the reaction mixture was then heated at 70 °C for 30 minutes. The solution is clear and yellow. An aqueous solution of NH₄PF₆ (1M, 2.5 mL) was added dropwise and the solution became turbid. A dark agglomerate was formed. After additional 15 minutes stirring, the solution was filtered to yield compound (*RR*)-7 (0.07 g, 0.04mmol, 72%), as a dark green solid. ¹H-NMR (*d*₆-DMSO, 200 MHz, 25 °C) δ = 8.76 (d, 8H; pyridine), 8.67 (s, 4H; binaphthyl), 8.23 (d, 4H; binaphthyl), 8.02 (d, 8H; pyridine), 7.72 (d, 4H; binaphthyl), 7.55 (d, 2H; binaphthyl), 6.79 (d, 4H; binaphthyl), 5.65 (broad peak, 8H; -NH₂), 3.76 (s, 12H; -OCH₃), 2.71 (broad singlet, 8H; -CH₂-). ¹³C-NMR (CD₃CN, 75 MHz, 25 °C) δ = 156.7 (C_{quat}), 151.5 (CH), 151.4 (C_{quat}), 134.6 (C_{quat}), 131.0 (CH), 129.5 (C_{quat}), 128.8 (C_{quat}), 127.9 (CH), 125.7 (CH), 124.4 (CH), 123.6 (CH), 118.3 (C_{quat}), 115.1 (CH), 56.1 (CH₃), 46.7 (CH₂). ESI-MS (m/z)=1704.31 ([*M*-PF₆]⁺).



Figure S2.a) Comparison between the ¹H-NMR spectra of the ligand (*R*)-4 and the complex (*RR*)-6 in DMSO- d_6 ; b) comparison between the ¹H-NMR spectra of the ligand (*R*)-5 and the complex (*RR*)-7 in DMSO- d_6 .



Figure S3.Experimental (top) and theoretical (bottom) isotopic assets for the quasi molecular ions $[M-PF_6]^+$ of compounds (*RR*)-6 on the left and (*RR*)-7 on the right.

2) Additional titrations and procedures

General procedure for the NMR, UV, CD and CV titration experiments. The titration experiments (UV/Vis, Circular Dichroism (CD) and ¹H-NMR) were conducted as follows. To a stock solution of the host molecule (solution A) in pure CH₃CN or CH₃CN:H₂O mixtures (UV/Vis spectroscopic grade or HPLC grade; CD₃CN and D₂O in the case of ¹H-NMR) were added several aliquots of the guest compound (solution B). Solution B is formed by the guest molecule at higher concentration dissolved in solution A, in order to maintain the ligand always at the same, constant concentration. *Cyclic voltammetry (CV) experiments* were performed in a mixture of CH₃CN:H₂O 7:3 with tetrabutylammonium hexafluorophosphate (0.1 M) as the supporting electrolyte, and scan speed of 100 mV/s; potential range investigated was from -200mV to +900 mV (vs Ag/AgCl/3M NaCl). Solutions were deaerated with a nitrogen stream (5 min) before use. In this case, incrementing aliquots of concentrated *guest* solution were added to a diluted solution of *host* (1.3x10⁻⁴ M in the voltammetric cell) in order to avoid diluting effect.



Figure S4. Left: UV/Vis titration in MeCN of compound (*RR*)-6 ($1x10^{-5}$ M) with increasing amounts of ferrocene. Right: CD titration in MeCN of compound (*RR*)-6 ($1x10^{-5}$ M) with increasing amounts of ferrocene.



Figure S5. Top: UV-Vis titration spectra in different solvents mixtures of compound (*RR*)-7 with increasing amounts of ferrocene: left) $1x10^{-5}$ M of macrocycle in MeCN:H₂O 7:3; right) 2,5x10⁻⁵ M in MeCN:H₂O 9:1; and bottom:CD titration of (*RR*)-7(1x10⁻⁵ M)in MeCN:H₂O 7:3.

Entry	Solvent	٤ ^[a]	ΔG_{II}° (kJM ⁻¹)	$\frac{\Delta G_{l2}^{\circ}}{(\text{kJ M}^{-1})}$	ΔG_{tot} (kJM ⁻¹)	$\frac{\log K_{11}}{(\text{kcal M}^{-1})}$	<i>Log K</i> ₁₂ (kcal M ⁻¹)		
1	MeCN	35,9	-28,7	-19	-47,7	4,41	2,71		
2	MeCN:H ₂ O 9:1	39,4	-30,9	-16,2	-47,1	4,79	2,22		
3	MeCN:H ₂ O 7:3	46,8	-35,4	-17,2	-52,2	5,58	2,39		
[a] Dielectric constant. Data taken form ref. S3.									

Table S1. Binding energies and association constants at 298 K for the multiple equilibriums found in the titration of (RR)-7 with ferrocene obtained using SIVVU (see Table S2-S4).

Table S2. Fit of titration in entry 1 Table S1 (24.9 μ M Macrocycle (*RR*)-7 in solvent of 100% MeCN) was titrated with 0 –5 equivalents of ferrocene at 298 K. A 1.3 mM solution of ferrocene alone is also included.



Optimization Summary:

Data at 298 K

Non-negativity was enforced with optimization (not truncation).

Activity Coefficients Model: None.

Species with Fixed Molar Absorptivity Curves: None.

Solutions ignored: None.

Optimized Values (kJ/mol): $\Delta G_1^\circ = -28.7(\pm 0.3)$; $\Delta G_2^\circ = -19(\pm 2)$;

Equilibrium Restricted RMS Residual (4 chemical factors): 0.00083444

Unrestricted RMS Residual (4 mathematical factors): 0.00015618

Restricted Data Reconstruction (4 chemical factors): 99.9376%

Unrestricted Data Reconstruction (4 mathematical factors): 99.9423%

Remaining Error Imbedded in Absorbance Values: 0.00050319

R²: 99.9998%

The standard deviations of the ΔG° values are estimated by re-optimizing the model 40 times, each with half of the absorbance data randomly ignored (30% of the solutions and 30% percent of the wavelengths).

Table S3. Fit of titration in entry 2 Table S1 (26 μ M Macrocycle (*RR*)-7 in solvent of 90% MeCN and 10% water) with 0 – 15 equivalents of ferrocene at 298 K. A 0.511 mM solution of ferrocene alone is also included.



Optimization Summary:

Data at 298 K

Non-negativity was enforced with optimization (not truncation).

Activity Coefficients Model: None.

Species with Fixed Molar Absorptivity Curves: None.

Solutions ignored: None.

Optimized Values (kJ/mol): $\Delta G^{\circ}_{1} = -30.9(\pm 0.4)$; $\Delta G^{\circ}_{2} = -16.2(\pm 0.3)$;

Equilibrium Restricted RMS Residual (4 chemical factors): 0.0015441

Unrestricted RMS Residual (4 mathematical factors): 0.00022314

Restricted Data Reconstruction (4 chemical factors): 99.9106%

Unrestricted Data Reconstruction (4 mathematical factors): 99.917%

Remaining Error Imbedded in Absorbance Values: 0.0007279

R²: 99.9993%

The standard deviations of the ΔG° values are estimated by re-optimizing the model 40 times, each with half of the absorbance data randomly ignored (30% of the solutions and 30% percent of the wavelengths).

Table S4. Fit of titration in entry 3Table S1 (9.95 μ M Macrocycle (*RR*)-7in solvent of 70% MeCN and 30% water) with 0 – 18 equivalents of ferrocene at 298 K. A 0.538 mM solution of ferrocene alone is also included.



Optimization Summary:

Data at 298 K

Non-negativity was enforced with optimization (not truncation).

Activity Coefficients Model: None.

Species with Fixed Molar Absorptivity Curves: None.

Solutions ignored: None.

Optimized Values (kJ/mol): $\Delta G^{\circ}_1 = -35.4(\pm 0.6)$; $\Delta G^{\circ}_2 = -17.2(\pm 0.1)$; Equilibrium Restricted RMS Residual (4 chemical factors): 0.00075745

Unrestricted RMS Residual (4 mathematical factors): 0.00014401

Restricted Data Reconstruction (4 chemical factors): 99.8901%

Unrestricted Data Reconstruction (4 mathematical factors): 99.909%

Remaining Error Imbedded in Absorbance Values: 0.00050497

R²: 99.9989%

The standard deviations of the ΔG° values are estimated by re-optimizing the model 40 times, each with half of the absorbance data randomly ignored (30% of the solutions and 30% percent of the wavelengths).



Table S5. Fit of titration of a 10.3 μ M solution of Macrocycle (*RR*)-7in MeCN with 0 – 5.5 equivalents of ferrocene at 298 K.

Optimization Summary: Data at 298 K Non-negativity was not enforced. Activity Coefficients Model: None. Species with Fixed Molar Absorptivity Curves: None. Solutions ignored: None. Optimized Values (kJ/mol): $\Delta G^{\circ}_1 = -41(\pm 3)$; $\Delta G^{\circ}_2 = -31.9(\pm 0.6)$; Equilibrium Restricted RMS Residual (3 chemical factors): 25.1372 Unrestricted RMS Residual (3 mathematical factors): 19.9598 Restricted Data Reconstruction (3 chemical factors): 95.2171% Unrestricted Data Reconstruction (3 mathematical factors): 95.4065% Remaining Error Imbedded in Absorbance Values: 19.4712 R²: 99.8998%

The standard deviations of the ΔG° values are estimated by re-optimizing the model 40 times, each with half of the absorbance data randomly ignored (30% of the solutions and 30% of the wavelengths).



Figure S6.¹H-NMR titration (200 MHz, CD₃CN:D₂O 7:3) of compound (*RR*)-7 1x10⁻³ M.



Figure S7.¹H-NMR titration (200 MHz, CD₃CN) of compound (*RR*)-7 1x10⁻³ M.



Figure S8.¹H NMR spectra (500 MHz, CD₃CN) of compound (*RR*)-7 alone (a, black trace, 5.5 mM) and in the presence of ferrocene (b, red trace, 16 mM).



Figure S9.Plots of the shifts of the ¹H NMR peaks of ferrocene (a) and of hydrogens 8 and 5 of compound (*RR*)-7 (b) for the titration shown in Figure 3 main text.

Table S5. Outcome of the fitting of data from the ¹H NMR peaks of ferrocene (Figure S9) using the open access program BindFit (<u>www.supramolecular.org</u>). Model 2:1 binding (species at constant concentration:species at variable concentration, ferrocene:macrocycle, respectively)

K ₁₁	K ₂₁	K ₁₁ error (%)	K ₂₁ error (%)	SSR			
0,142173937	64585,6346	4,11042066	3,24260059	4,1554E-07			
Datapoints fitted	Params fitted	H coeffs	HG coeffs	H2G coeffs	Raw coeffs 1	Raw coeffs 2	Raw coeffs 3
17	4	4,15653	-40,2809993	4,26520904	4,15653	-40,2809993	4,26520904



Figure S10. 2D ROESY NMR spectrum of a mixture of ferrocene (2.4 mM) and (*RR*)-7 (14.7 mM) in CD₃CN.



Figure S11.2D ROESY NMR spectrum of a mixture of ferrocene (16.1 mM) and (*RR*)-7 (5.6 mM) in CD₃CN.



Figure S12. Modelling of the 1:2 host guest complex between (*RR*)-7 and ferrocene.



Figure S13. Comparisons between the Cyclic Voltammetry curves of ferrocene alone and in the presence of compound (*RR*)-7 ($1.3x10^{-4}$ M)in CH₃CN:H₂O 7:3.

Copies of NMR and Mass Spectra of Newly Synthesized Compounds Compound (R)-4.

¹H NMR (CDCl₃, 200 MHz)

s _ { r } / r

230 2.10 ę 101 9.5 9.0 7.0

¹³C NMR (CDCl₃, 75 MHz)



6.0

5.5

3.82 3.82 3.82

3.18-

3.5

4.0

4.5

5.0

¹³C NMR DEPT (CDCl₃, 75 MHz)



Compound (R)-**5**. ¹H NMR (CDCl₃, 200 MHz)



¹³C NMR DEPT (CDCl₃, 75 MHz)



Compound (RR)-6. ¹H NMR (DMSO-*d*₆, 200 MHz)





¹³C NMR DEPT (CD₃CN, 75 MHz)



ESI-MS



*Compound (RR)-***7**. ¹H NMR (DMSO-*d*₆, 200 MHz)



¹³C NMR (CD₃CN, 75 MHz)



¹³C NMR DEPT (CD₃CN, 75 MHz)



ESI-MS



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