

Accelerated hydration reaction of an unsymmetrical tolan evidenced by a Hg(II)-trapped macrocycle and its application as a Hg(II)-selective indicator

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Contents

General Experimental	S2
Structural supporting data for 2b	S2
Stacked ¹ H NMR spectra of unreactive 1	S3
Fluorescence studies of tolans	S4
LC-MS studies	S6
Stacked ¹ H NMR spectra of 2 , 2a , 2b , and crude sample (2 + Hg(II))	S7
X-ray structural data of 2b	S8
Synthetic Experimental	S11
NMR Spectra	S13

General Experiments

Reagents were purchased at the highest commercial quality and used without further purification, unless otherwise stated. Yields of synthesized compounds were measured after chromatographic purification. Stock solutions of all of the compounds studied were made up in CH₃CN with the final concentrations being between 2×10^{-5} M and 2×10^{-6} M. ACS grade solvents were purchased and used without purification. The stock solutions were appropriately diluted with HPLC water (or HEPES buffer) for the ensuing studies.

Structural supporting data for **2b**

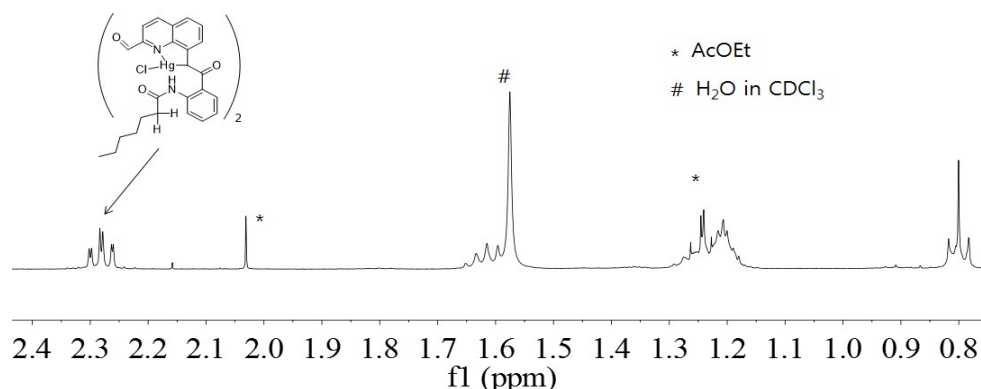


Figure S1. Partial ¹H NMR spectrum (400 MHz) of **2b** recorded in CDCl₃.

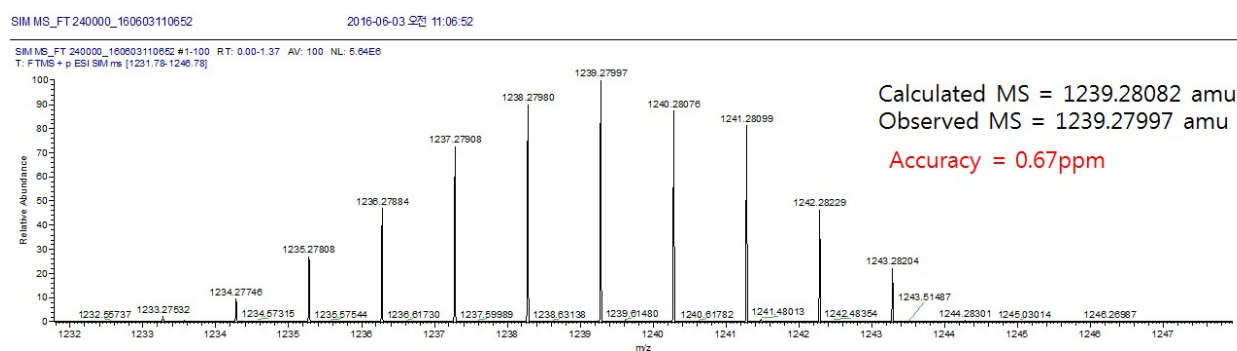


Figure S2. Observed high mass spectrum of **2b** (m/z , $[\mathbf{2b} - \text{Cl}]^+ = 1239.2800$). The value was obtained by high-resolution ESI mass spectrometry.

Stacked ^1H NMR spectra of unreactive **1**

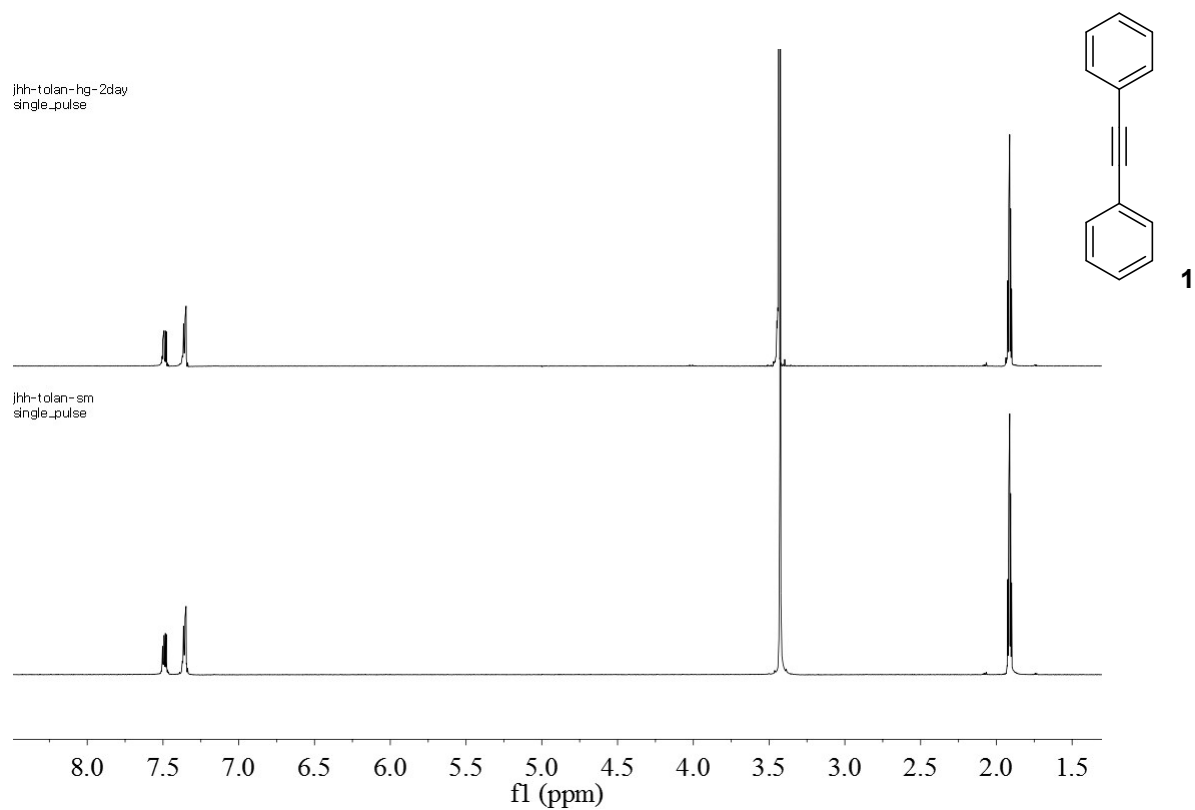


Figure S3. ^1H NMR spectra of **1** (2 mM in 20% D_2O containing $\text{CH}_3\text{CN-d}_3$) recorded before and 48 h after the addition of 2 equiv of Hg(II) .

Fluorescence studies of tolans

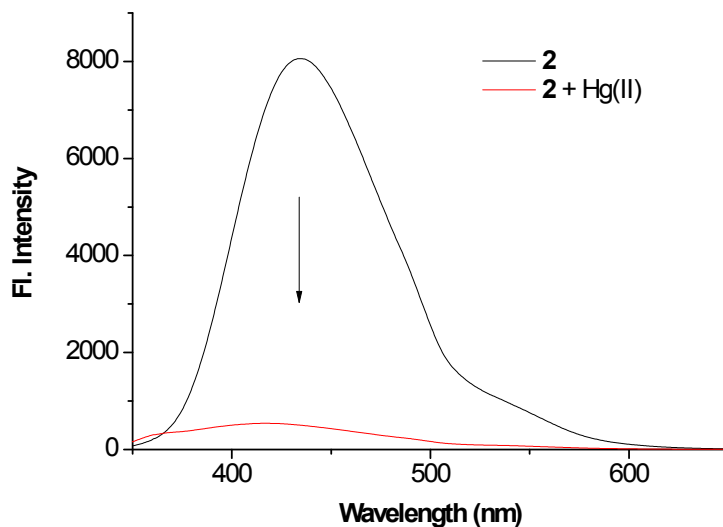


Figure S4. Fluorescence spectra of **2** (2 μ M) and final diluted solution obtained from reaction depicted in Figure 1, excited at 323 nm at 25 $^{\circ}$ C.

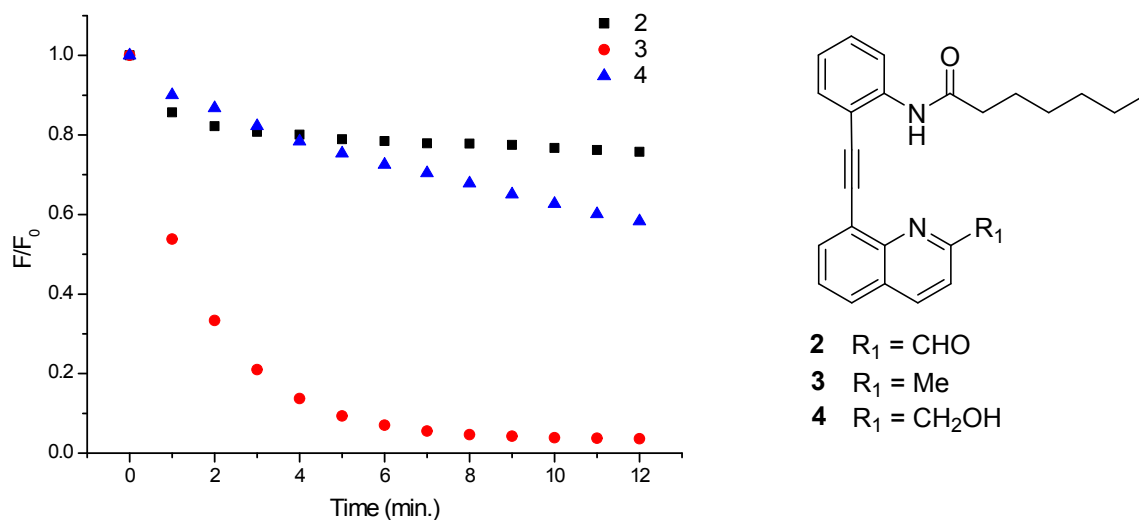


Figure S5. Plot of observed fluorescence ratio (F/F_0) of each tolan (2 μ M) in $\text{CH}_3\text{CN}/\text{water}$ (1:1, v/v) at 428 nm as a function of time (F_0 : the emission intensity of each tolan, F : the emission intensity of each tolan in the presence of 2 equiv of $\text{Hg}(\text{II})$).

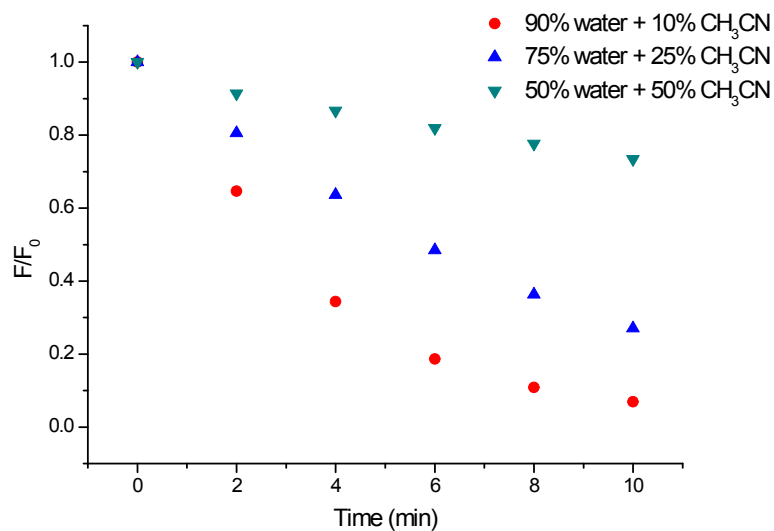


Figure S6. Plots of observed fluorescence ratio (F/F_0) of **3** (0.2 μM) at 428 nm as a function of time in various percentages of water and CH_3CN (v/v) (F_0 : the emission intensity of each tolan, F : the emission intensity of **3** in the presence of 2 equiv of $\text{Hg}(\text{II})$).

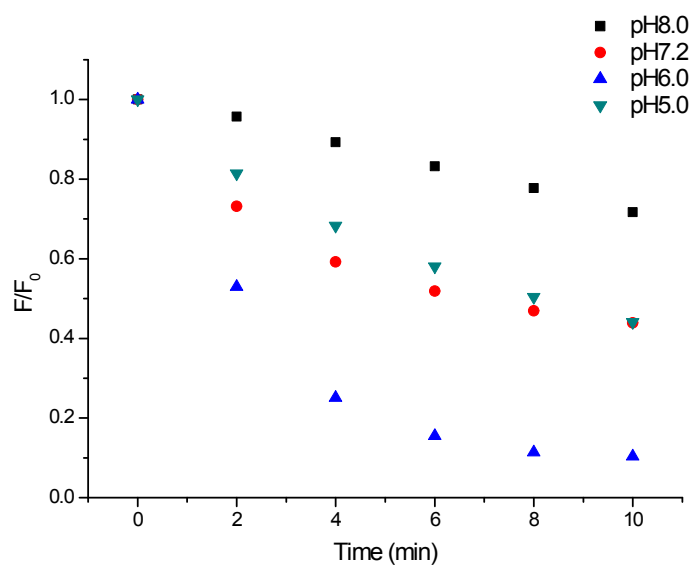
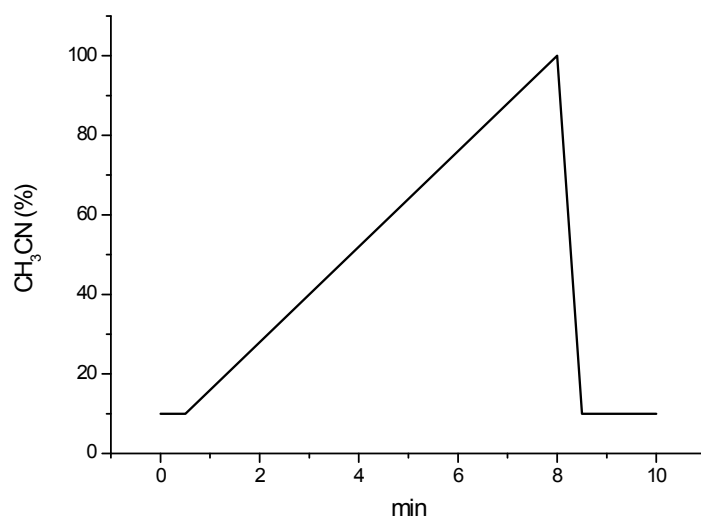


Figure S7. Plots of observed fluorescence ratio of **3** (0.2 μM) at 423nm as a function of time in various pH in 10 mM 90% HEPES buffer containing CH_3CN (F_0 : the emission intensity of each tolan, F : the emission intensity of **3** in the presence of 2 equiv of $\text{Hg}(\text{II})$).

LC-MS Spectroscopy



All LC-MS spectra were generated by Shimadzu LCMS-2020 (column; Sim-pack GIS C18, CN (100 × 3.0 mm, 3 μm); mobile phase flow; 0.8000 mL/min; mobile phase conditions (gradient A:B = water (0.1%TFA):acetonitrile) as shown above).

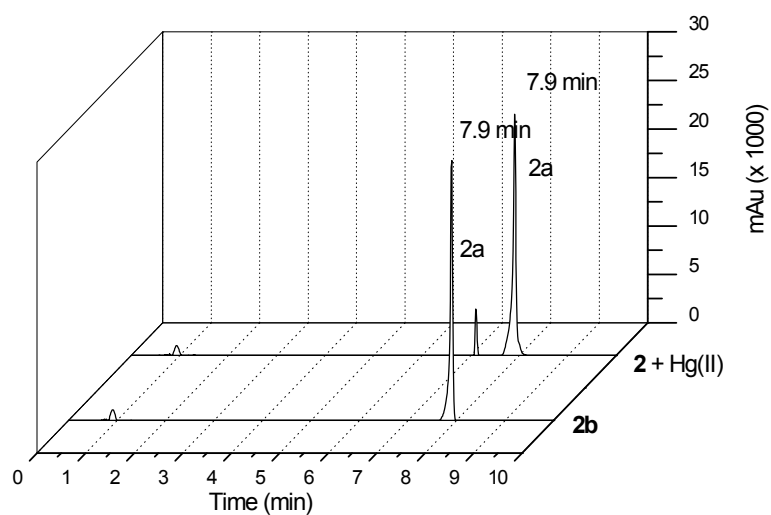


Figure S8. Obtained peak in the liquid chromatograms that undergo change upon the addition of Hg(II) (2 equiv) to a solution of indicator **2** (20 μM in CH₃CN-water (80:20, v/v) and resulted peak from pure **2b** (20 μM in CH₃CN-water (80:20, v/v) without Hg(II).

Stacked ^1H NMR spectra of **2, **2a**, **2b**, and crude sample (**2** + Hg(II))**

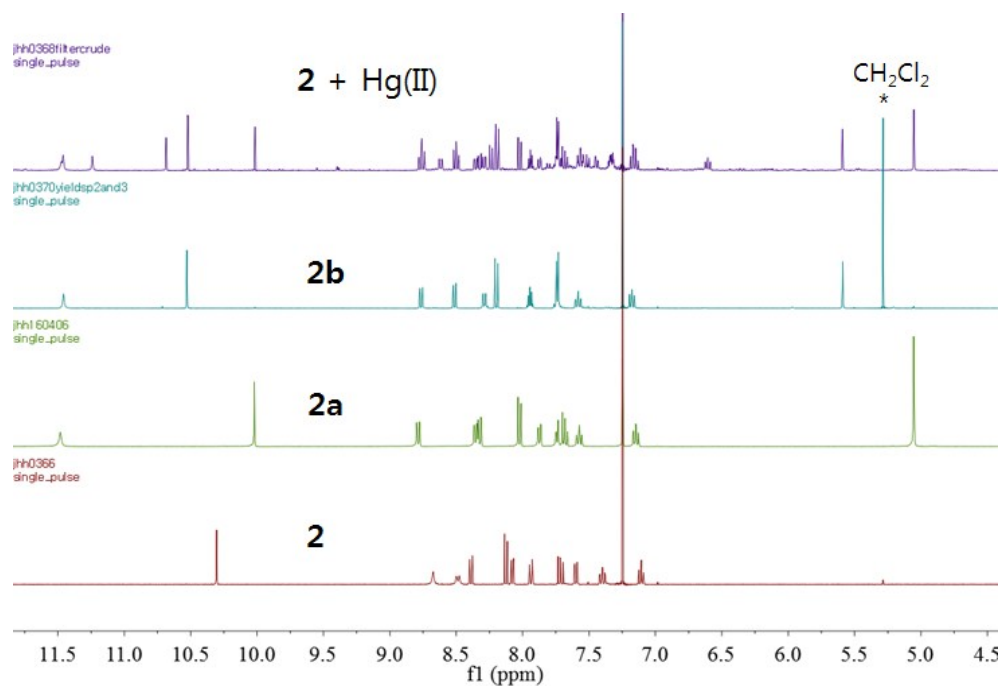


Figure S9. Stacked ^1H NMR spectra of **2**, **2a**, **2b**, and crude sample (**2** + Hg(II)) obtained after the hydration reaction for 10 min and extraction with CH_2Cl_2 in CDCl_3

X-ray structural data of 2b

Table S1. Crystal data and structure refinement for **2b**.

Identification code	2b
Empirical formula	C ₂₅ H ₂₅ ClHgN ₂ O ₃
Formula weight	637.51
Temperature/K	296.15
Crystal system	triclinic
Space group	P-1
a/Å	9.4629(2)
b/Å	11.3500(2)
c/Å	12.3473(3)
α/°	103.8430(10)
β/°	109.6160(10)
γ/°	99.7450(10)
Volume/Å ³	1166.86(4)
Z	2
ρ _{calc} /g/cm ³	1.814
μ/mm ⁻¹	6.739
F(000)	620.0
Crystal size/mm ³	0.435 × 0.283 × 0.22
Radiation	MoKα (λ = 0.71073)
2θ range for data collection/°	3.7 to 56.54
Index ranges	-12 ≤ h ≤ 12, -15 ≤ k ≤ 15, -16 ≤ l ≤ 16
Reflections collected	21104
Independent reflections	5748 [R _{int} = 0.0418, R _{sigma} = 0.0363]
Data/restraints/parameters	5748/0/291
Goodness-of-fit on F ²	1.065
Final R indexes [I ≥ 2σ (I)]	R ₁ = 0.0250, wR ₂ = 0.0589
Final R indexes [all data]	R ₁ = 0.0259, wR ₂ = 0.0593
Largest diff. peak/hole / e Å ⁻³	2.25/-1.39

Table S2. Fractional Atomic Coordinates ($\times 10^4$) and Equivalent Isotropic Displacement Parameters ($\text{\AA}^2 \times 10^3$) for y2. U_{eq} is defined as 1/3 of the trace of the orthogonalised U_{ij} tensor.

Atom	<i>x</i>	<i>y</i>	<i>z</i>	<i>U</i> (eq)
Hg1	1149.55(12)	1721.74(9)	6479.58(9)	13.04(5)
Cl1	2120.4(10)	515.7(8)	5225.4(8)	23.80(17)
O3	801(3)	4798(2)	7761(2)	18.2(4)
O1	6961(3)	2476(3)	8154(2)	34.4(7)
O2	-1503(3)	6517(2)	4577(2)	22.9(5)
N2	-729(3)	5657(3)	6097(2)	17.5(5)
N1	3560(3)	2890(3)	8534(2)	14.5(5)
C21	1004(6)	8350(5)	7551(4)	50.4(14)
C19	-576(4)	6573(3)	5574(3)	17.9(6)
C1	5644(4)	2553(4)	7941(3)	23.4(7)
C14	-2823(4)	2438(3)	5733(3)	15.5(6)
C7	1322(4)	3125(3)	8982(3)	14.2(6)
C12	-147(3)	3748(3)	7233(3)	12.6(5)
C5	3931(4)	3754(3)	10635(3)	16.1(6)
C6	2958(4)	3257(3)	9376(3)	14.0(6)
C8	732(4)	3462(3)	9850(3)	19.4(6)
C22	2489(5)	9396(4)	8297(3)	31.4(8)
C17	-3371(4)	4351(3)	4751(3)	22.1(7)
C23	2617(9)	10171(6)	9502(5)	83(2)
C15	-4252(4)	2265(3)	4821(3)	21.4(7)
C3	6122(4)	3494(3)	10129(3)	19.9(6)
C13	-1636(3)	3562(3)	6189(3)	12.5(5)
C9	1692(4)	3954(3)	11095(3)	22.3(7)
C10	3267(4)	4115(3)	11488(3)	20.3(7)
C11	210(4)	2667(3)	7658(3)	13.1(6)
C16	-4525(4)	3230(3)	4345(3)	25.4(7)
C20	879(5)	7650(4)	6331(3)	30.7(9)
C4	5545(4)	3886(3)	10988(3)	19.5(6)
C18	-1916(4)	4529(3)	5661(3)	14.5(6)
C2	5077(4)	2999(3)	8908(3)	17.4(6)
C25	-2596(9)	-490(5)	8421(5)	67.0(18)
C24	-2305(10)	355(7)	-332(5)	81(2)

Synthetic experimental

Synthetic procedures for Unsymmetrical tolans

N-(2-((2-formylquinolin-8-yl)ethynyl)phenyl)heptanamide (2)

N-(2-ethynylphenyl)heptanamide (81 mg, 0.35 mmol), PPh_3 (4.6 mg, 0.175 mmol), $\text{Pd}(\text{PPh}_3)_2\text{Cl}$ (12.4 mg, 0.018 mmol), and $\text{Cu}(\text{OAc})_2$ (7 mg, 0.035 mmol) were added to a well-stirred solution of 8-iodoquinoline-2-carbaldehyde (100 mg, 0.35 mmol) in dry DMF (4 mL) and DIEA (0.24 mL, 1.40 mmol). The resulting mixture was heated at 65 °C for 3 h. After cooling to room temperature, the reaction mixture was extracted with EtOAc/water. The combined organic layers were dried over anhydrous Na_2SO_4 and then filtered. The filtrate was concentrated under reduced pressure to obtain the residue. The residue was purified over silica gel to afford **2** (68 mg, 53%).

^1H NMR (400 MHz, Chloroform-*d*) δ 10.52 (d, J = 0.9 Hz, 1H), 8.89 (s, 1H), 8.71 (d, J = 8.4 Hz, 1H), 8.61 (d, J = 8.4 Hz, 1H), 8.34 (d, J = 8.4 Hz, 1H), 8.30 (dd, J = 7.2, 1.4 Hz, 1H), 8.16 (dd, J = 8.3, 1.4 Hz, 1H), 7.93 (dd, J = 8.3, 7.2 Hz, 1H), 7.82 (dd, J = 7.6, 1.5 Hz, 1H), 7.62 (ddd, J = 8.8, 7.6, 1.5 Hz, 1H), 7.33 (ddd, J = 7.6, 7.6, 1.2 Hz, 1H), 2.60 (t, J = 7.6 Hz, 2H), 1.89 (tt, J = 7.7, 7.7 Hz, 2H), 1.50 – 1.39 (m, 2H), 1.39 – 1.24 (m, 4H), 0.96 (t, J = 7.0 Hz, 3H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 193.03, 171.68, 152.89, 147.59, 139.79, 138.31, 134.30, 131.47, 130.40, 130.28, 128.96, 128.71, 124.01, 123.38, 119.83, 118.32, 112.05, 93.57, 92.18, 38.33, 31.49, 28.85, 25.56, 22.43, 13.99. HRMS–EI: m/z $[\text{M}]^+$ calcd for $\text{C}_{25}\text{H}_{24}\text{N}_2\text{O}_2$: 384.1838; found: 384.1837.

N-(2-((2-methylquinolin-8-yl)ethynyl)phenyl)heptanamide (3)

By following the general procedure, the reaction of 8-iodo-2-methylquinoline (100 mg, 0.37 mmol) yielded **3** (37 mg, 27%). To obtain the product, CuI was used instead of $\text{Cu}(\text{OAc})_2$ due to Glazer reaction.

^1H NMR (400 MHz, Chloroform-*d*) δ 9.72 (s, 1H), 9.25 (d, J = 8.3 Hz, 1H), 8.88 (d, J = 8.8 Hz, 1H), 8.73 (d, J = 7.5 Hz, 1H), 8.59 (d, J = 7.9 Hz, 1H), 8.35 (d, J = 7.7 Hz, 1H), 8.28 (dd, J = 7.6, 7.6 Hz, 1H), 8.19 – 8.11 (m, 2H), 8.05 (d, J = 1.0 Hz, 1H), 7.87 (dd, J = 7.8 Hz, 1H), 3.60 (s, 3H), 3.25 (t, J = 7.6 Hz, 2H), 2.47 (tt, J = 7.6, 7.6 Hz, 2H), 2.16 – 1.98 (m, 2H), 1.98 – 1.84 (m, 4H), 1.54 (t, J = 7.1 Hz, 3H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 172.83, 160.88, 148.56, 140.64, 137.55, 133.98, 131.90, 130.50, 129.42, 127.54, 126.21, 124.04, 123.64, 122.78, 120.74, 113.62, 95.56, 91.55, 38.91, 32.30, 29.69, 26.89, 26.43, 23.24, 14.79. HRMS–EI: m/z $[\text{M}]^+$ calcd for $\text{C}_{25}\text{H}_{26}\text{N}_2\text{O}$: 370.2045; found: 370.2042.

N-(2-((2-(hydroxymethyl)quinolin-8-yl)ethynyl)phenyl) heptanamide (4)

To a well-stirred solution of **3** (52 mg, 0.135 mmol) in MeOH (3.5 mL) was added NaBH_4 (7.7 mg, 0.20 mmol). The reaction mixture was stirred at 0 °C for 0.5 h. After the reaction mixture was quenched by adding one drop of water, MeOH was removed. The residue was extracted with Et₂O/EtOAc. The combined organic layers were dried over anhydrous Na_2SO_4 and then filtered. The filtrate was concentrated under reduced pressure to obtain the residue. The residue was purified over silica gel to afford **4** (45 mg, 87%).

¹H NMR (400 MHz, Acetonitrile-*d*₃) δ 8.61 (s, 1H), 8.31 (d, *J* = 8.6 Hz, 1H), 8.27 (d, *J* = 8.4 Hz, 1H), 8.00 (dd, *J* = 7.1, 1.5 Hz, 1H), 7.95 (dd, *J* = 8.2, 1.4 Hz, 1H), 7.61 (d, *J* = 8.6 Hz, 1H), 7.60 – 7.53 (m, 2H), 7.39 (ddd, *J* = 8.6, 7.7, 1.3 Hz, 1H), 7.13 (ddd, *J* = 7.7, 7.7, 0.9 Hz, 1H), 4.88 (d, *J* = 5.3 Hz, 2H), 4.07 (t, *J* = 5.4 Hz, 1H), 2.39 (t, *J* = 7.5 Hz, 2H), 1.59 (tt, *J* = 7.6 Hz, 2H), 1.25 – 1.77 (m, 2H), 1.17 – 1.01 (m, 4H), 0.72 (t, *J* = 6.9 Hz, 3H). ¹³C NMR (100 MHz, Acetonitrile-*d*₃) δ 172.89, 163.28, 147.59, 140.74, 138.47, 134.60, 132.43, 130.63, 130.22, 128.73, 126.96, 124.60, 122.63, 121.40, 120.81, 114.17, 95.25, 91.16, 65.96, 38.38, 32.26, 29.55, 26.34, 23.18, 14.31. HRMS–EI: *m/z* [M]⁺ calcd for C₂₅H₂₆N₂O₂: 386.1994; found: 386.1997.

Synthetic procedures for ketones with Hg(II)

N-(2-(2-(2-formylquinolin-8-yl)acetyl)phenyl)heptanamide (2a)

To a well-stirred solution of **2** (20 mg, 0.052 mmol) in CH₃CN/water (20 mL/4 mL) was added HgCl₂ (28 mg, 0.104 mmol). The reaction mixture was stirred at room temperature for 10 min. The reaction mixture was poured into H₂O, and the product was extracted with DCM. The combined organic layers were dried over anhydrous Na₂SO₄ and then filtered. The filtrate was concentrated under reduced pressure to obtain the residue. To obtain **2a** without **2b**, the same crude was dissolved in DCM (4 mL) and then 0.42 mL of HCl (1.25 M in MeOH, 0.52 mmol) was added to the solution. The reaction was stirred for 10 min. The residue was purified over preparative TLC to afford **2a** (17 mg, 82%).

¹H NMR (400 MHz, Chloroform-*d*) δ 11.48 (s, 1H), 10.02 (d, *J* = 0.9 Hz, 1H), 8.79 (dd, *J* = 8.5, 1.2 Hz, 1H), 8.35 (dd, *J* = 8.1, 1.6 Hz, 1H), 8.32 (dd, *J* = 8.4, 0.9 Hz, 1H), 8.02 (d, *J* = 8.4 Hz, 1H), 7.87 (dd, *J* = 8.1, 1.6 Hz, 1H), 7.74 (dd, *J* = 7.1, 1.6 Hz, 1H), 7.68 (dd, *J* = 8.1, 7.1 Hz, 1H), 7.57 (ddd, *J* = 8.7, 7.3, 1.6 Hz, 1H), 7.15 (ddd, *J* = 8.3, 7.3, 1.2 Hz, 1H), 5.05 (s, 2H), 2.31 (t, *J* = 7.5 Hz, 2H), 1.64 (tt, *J* = 7.6, 7.6 Hz, 2H), 1.35 – 1.15 (m, 4H), 0.82 (t, *J* = 6.7 Hz, 3H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 202.71, 193.68, 172.82, 151.96, 146.16, 141.41, 137.91, 135.76, 135.22, 131.82, 131.39, 130.41, 129.13, 127.54, 122.35, 121.84, 121.07, 117.49, 42.31, 38.74, 31.54, 28.89, 25.51, 22.53, 14.07. HRMS–FAB: *m/z* [M + H]⁺ calcd for C₂₅H₂₇N₂O₃: 403.2022; found: 403.2024.

(1-(2-formylquinolin-8-yl)-2-(2-heptanamidophenyl)-2-oxoethyl)mercury(II) chloride (2b)

The residue obtained from the reaction of **2** with HgCl₂ was directly purified by preparative TLC, which afforded **2a** (2.7 mg, 13%) and **2b** (21.8 mg 66%).

¹H NMR (400 MHz, Chloroform-*d*) δ 11.46 (s, 1H), 10.53 (d, *J* = 0.9 Hz, 1H), 8.76 (dd, *J* = 8.5, 1.2 Hz, 1H), 8.51 (dd, *J* = 8.4, 0.9 Hz, 1H), 8.29 (dd, *J* = 8.2, 1.6 Hz, 1H), 8.20 (d, *J* = 8.4 Hz, 1H), 7.94 (dd, *J* = 5.3, 4.4 Hz, 1H), 7.78 – 7.69 (m, 2H), 7.58 (ddd, *J* = 8.7, 7.4, 1.5 Hz, 1H), 7.18 (ddd, *J* = 8.1, 7.3, 1.3 Hz, 1H), 5.59 (d, *J* = 0.5 Hz, 1H), 2.28 (td, *J* = 7.5, 1.5 Hz, 2H), 1.63 (tt, *J* = 7.5, 7.5 Hz, 2H), 1.32 – 1.13 (m, 6H), 0.80 (t, *J* = 6.8 Hz, 3H). HRMS–ESI: *m/z* [M – Cl]¹⁺ calcd for C₅₀H₅₀ClHg₂N₄O₆: 1239.2808; found: 1239.2800.

N-(2-(2-(2-methylquinolin-8-yl)acetyl)phenyl)heptanamide (3a)

By following the procedure of **2a**, the reaction of **3** (20 mg, 0.054 mmol) yielded **3a** (11.1 mg, 53%).

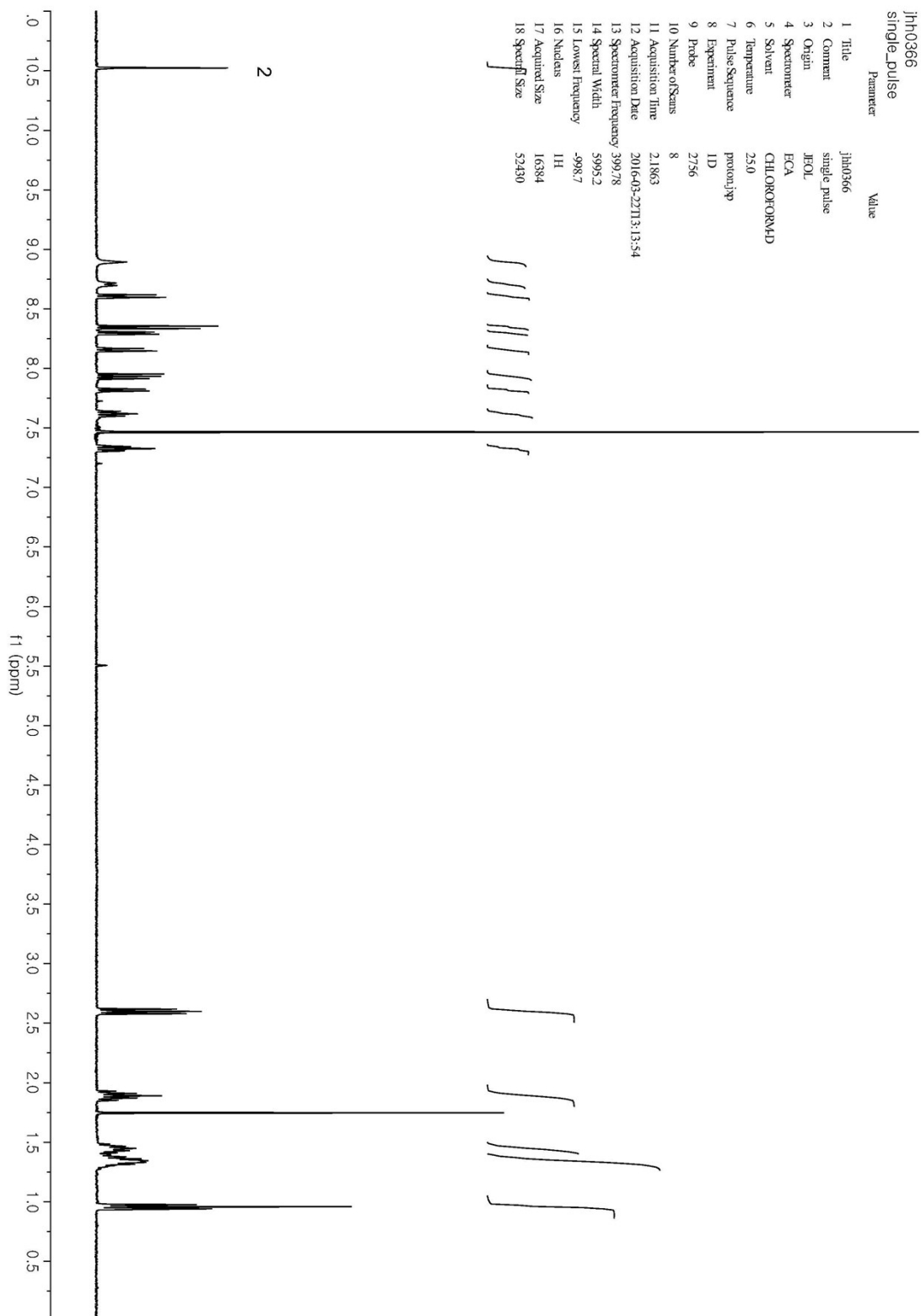
^1H NMR (400 MHz, Chloroform-*d*) δ 11.55 (s, 1H), 8.74 (d, J = 8.7 Hz, 1H), 8.35 (d, J = 9.6 Hz, 1H), 8.02 (d, J = 8.6 Hz, 1H), 7.72 (d, J = 9.2 Hz, 1H), 7.60 (d, J = 7.0 Hz, 1H), 7.52 (ddd, J = 8.6, 7.8, 1.5 Hz, 1H), 7.45 (dd, J = 8.2, 7.2 Hz, 1H), 7.25 (d, J = 8.4 Hz, 1H), 7.09 (ddd, J = 8.3, 7.4, 1.3 Hz, 1H), 4.91 (s, 2H), 2.60 (s, 3H), 2.33 (t, J = 7.6 Hz, 2H), 1.65 (tt, J = 7.6 Hz, 2H), 1.33 – 1.16 (m, 6H), 0.84 (t, J = 6.7 Hz, 3H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 203.86, 172.92, 158.55, 145.89, 141.22, 136.54, 134.78, 134.02, 131.80, 130.60, 127.32, 126.84, 125.58, 122.68, 122.36, 122.24, 120.99, 42.70, 38.88, 31.71, 29.07, 25.71, 25.52, 22.69, 14.23. HRMS–EI: m/z $[\text{M}]^+$ calcd for $\text{C}_{25}\text{H}_{28}\text{N}_2\text{O}_2$: 388.2151; found: 388.2153.

N-(2-(2-(2-(hydroxymethyl)quinolin-8-yl)acetyl)phenyl)heptanamid (**4a**)

By following the procedure of **2a**, the reaction of **4** (20 mg, 0.052 mmol) yielded **4a** (9.8 mg, 47%).

^1H NMR (400 MHz, Chloroform-*d*) δ 11.50 (s, 1H), 8.76 (dd, J = 8.6, 1.2 Hz, 1H), 8.22 (dd, J = 8.0, 1.6 Hz, 1H), 8.14 (d, J = 8.5 Hz, 1H), 7.79 (dd, J = 8.1, 1.5 Hz, 1H), 7.66 (dd, J = 7.2, 1.4 Hz, 1H), 7.54 (d, J = 7.6 Hz, 1H), 7.53 (d, J = 8.0 Hz, 1H), 7.26 (d, J = 8.5 Hz, 1H), 7.10 (ddd, J = 8.1, 7.6, 1.1 Hz, 1H), 4.92 (s, 2H), 4.82 (s, 2H), 3.98 (s, 1H), 2.34 (t, J = 7.5 Hz, 2H), 1.65 (tt, J = 7.6 Hz, 2H), 1.36 – 1.17 (m, 6H), 0.83 (t, J = 6.9 Hz, 3H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 203.04, 172.90, 158.62, 144.81, 141.43, 137.41, 135.18, 133.74, 131.34, 131.04, 127.86, 127.53, 126.32, 122.37, 121.71, 121.17, 118.58, 64.29, 42.97, 38.71, 31.58, 28.92, 25.50, 22.55, 14.09. HRMS–EI: m/z $[\text{M}]^+$ calcd for $\text{C}_{25}\text{H}_{28}\text{N}_2\text{O}_3$: 404.2100; found: 404.2098.

NMR Spectra

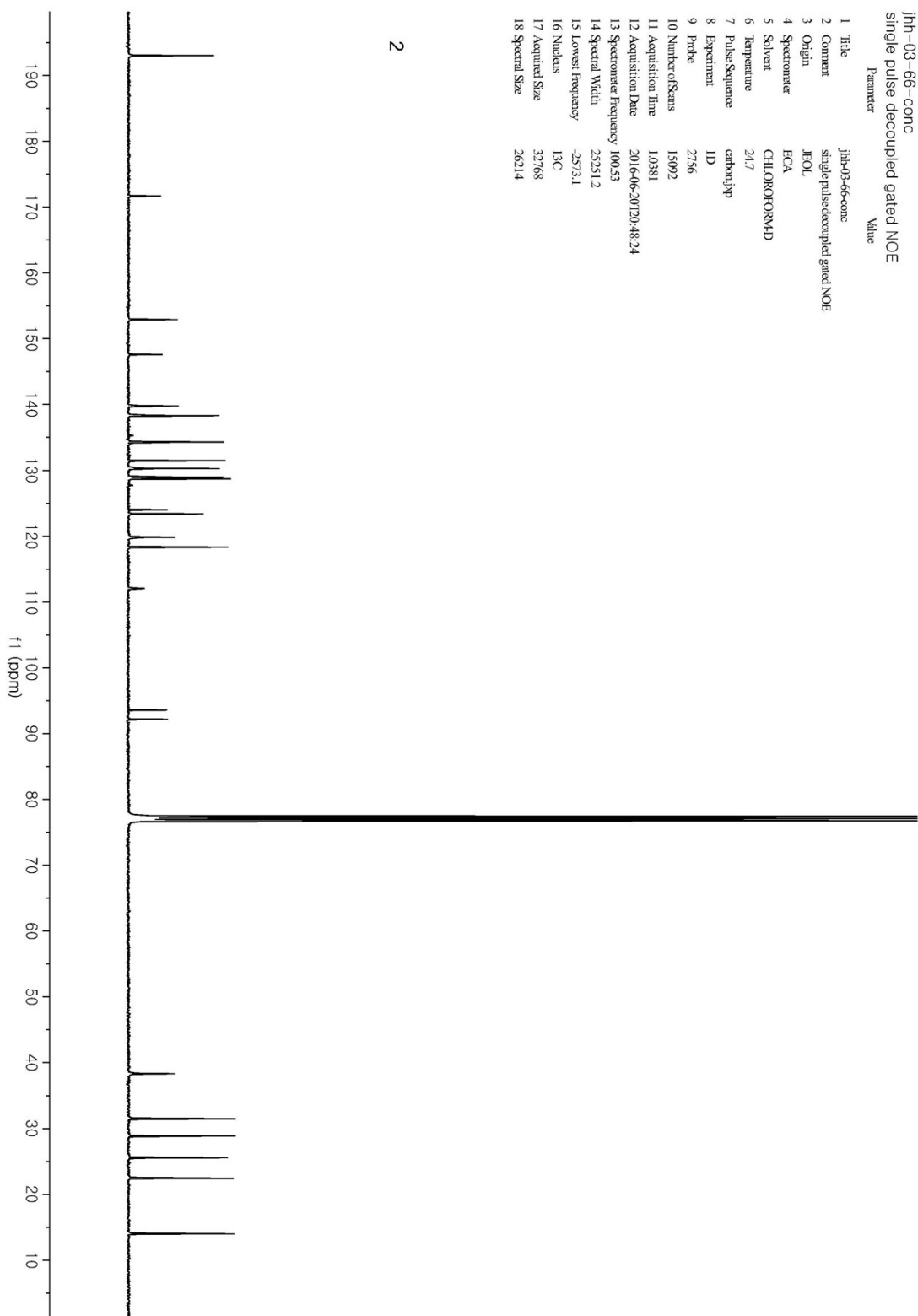


¹H NMR spectrum of **2** recorded in CDCl₃

jth-03-66-conc
single pulse decoupled gated NOE

Parameter	Value
1 Title	jth-03-66-conc
2 Comment	single pulse decoupled gated NOE
3 Origin	JFOL
4 Spectrometer	ECA
5 Solvent	CHLOROFORM-D
6 Temperature	24.7
7 Pulse Sequence	zgpg30
8 Experiment	1D
9 Probe	2756
10 Number of Scans	15092
11 Acquisition Time	1.0381
12 Acquisition Date	2016-06-20T20:48:24
13 Spectrometer Frequency	100.53
14 Spectral Width	22551.2
15 Lowest Frequency	-2573.1
16 Nucleus	¹³ C
17 Acquired Size	32768
18 Spectral Size	26214

2

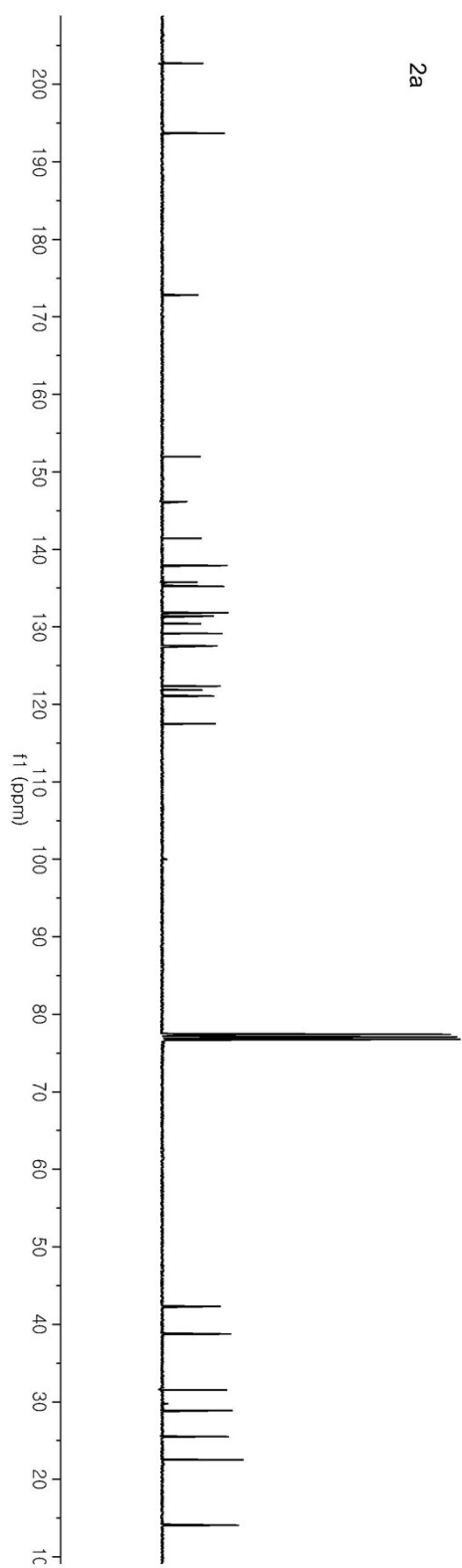


¹³C NMR spectrum of **2** recorded in CDCl₃

jhh160406
single pulse decoupled gated NOE

Parameter	Value
1 Title	jhh160406
2 Comment	single pulse decoupled gated NOE
3 Origin	JEOL
4 Spectrometer	ECA
5 Solvent	CHLOROFORMD
6 Temperature	25.2
7 Pulse Sequence	carbonjnp
8 Experiment	1D
9 Probe	2756
10 Number of Scans	12895
11 Acquisition Time	1.0381
12 Acquisition Date	2016-04-06T22:01:09
13 Spectrometer Frequency	100.53
14 Spectral Width	25252.5
15 Lowest Frequency	-2573.7
16 Nucleus	¹³ C
17 Acquired Size	32768
18 Spectral Size	52430

2a

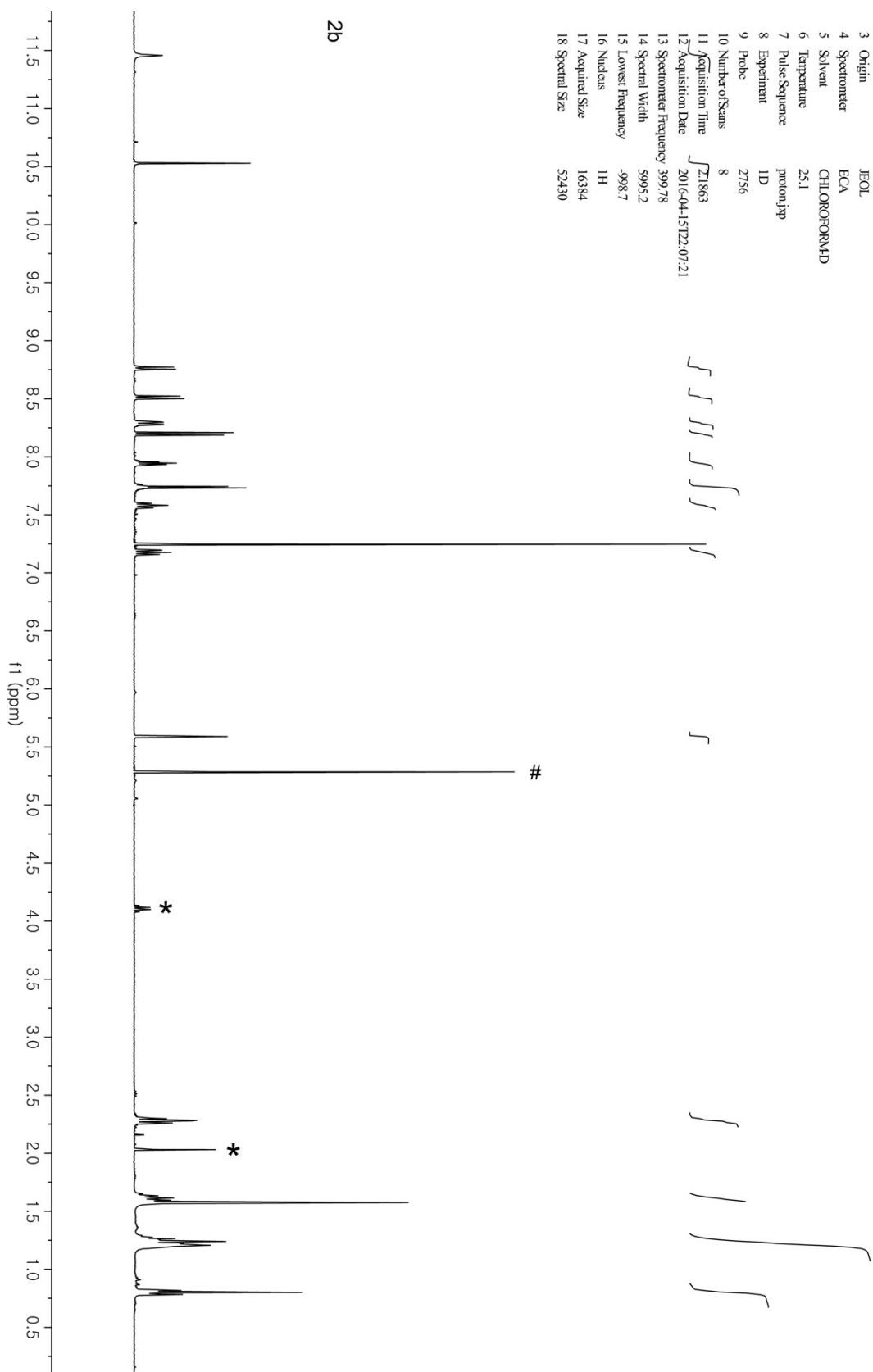


¹³C NMR spectrum of **2a** recorded in CDCl₃

jhm0370yieldsp2and3
single_pulse

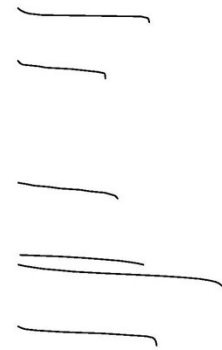
Parameter	Value
1 Title	jhm0370yieldsp2and3
2 Comment	single_pulse
3 Origin	JEOL
4 Spectrometer	ECA
5 Solvent	CHLOROFORMD
6 Temperature	25.1
7 Pulse Sequence	protonjyp
8 Experiment	1D
9 Probe	2756
10 Number of Scans	8
11 Acquisition Time	21.863
12 Acquisition Date	2016-04-15T22:07:21
13 Spectrometer Frequency	399.78
14 Spectral Width	5995.2
15 Lowest Frequency	-998.7
16 Nucleus	¹ H
17 Acquired Size	16384
18 Spectral Size	52430

2b



¹H NMR spectrum of **2b** recorded in CDCl₃ (#:CH₂Cl₂, *:EtOAc)

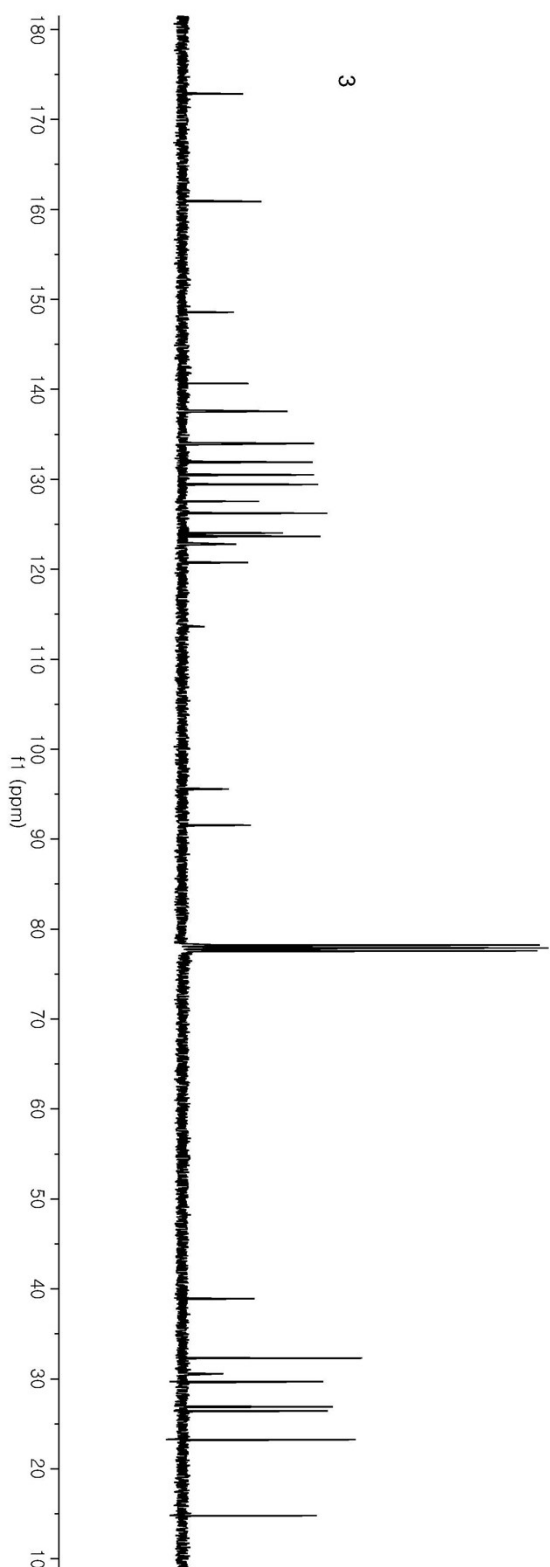
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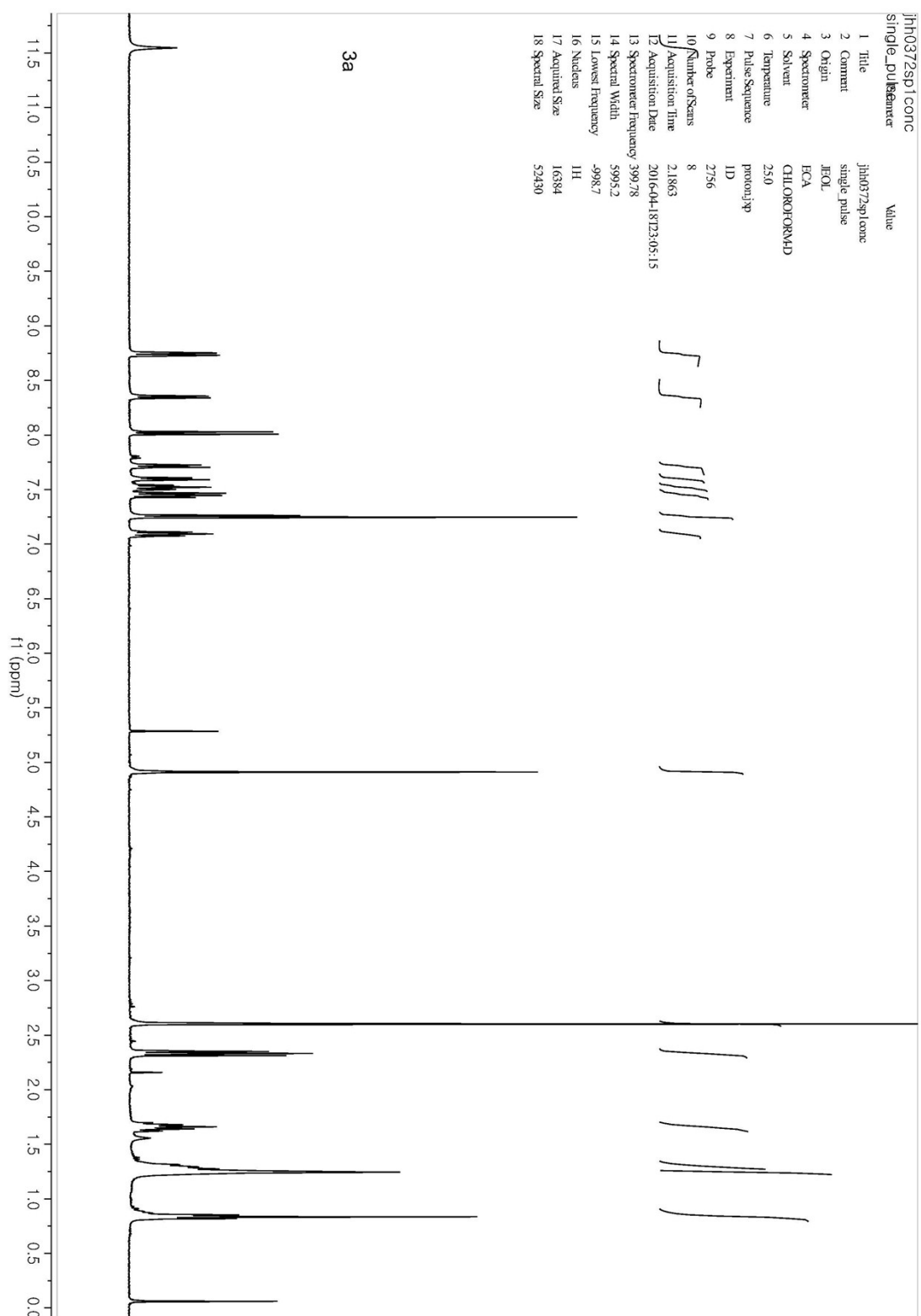
11

sat-methyl
single pulse decoupled gated NOE

Parameter	Value
1 Title	sat-methyl
2 Comment	single pulse decoupled gated NOE
3 Origin	JEOL
4 Spectrometer	ECA
5 Solvent	CHLOROFORM-D
6 Temperature	27.3
7 Pulse Sequence	carbon_jnp
8 Experiment	1D
9 Probe	2756
10 Number of Scans	528
11 Acquisition Time	1.0381
12 Acquisition Date	2016-03-19T14:50:48
13 Spectrometer Frequency	100.53
14 Spectral Width	25252.5
15 Lowest Frequency	-2573.7
16 Nucleus	¹³ C
17 Acquired Size	32768
18 Spectral Size	52430



¹³C NMR spectrum of **3** recorded in CDCl₃

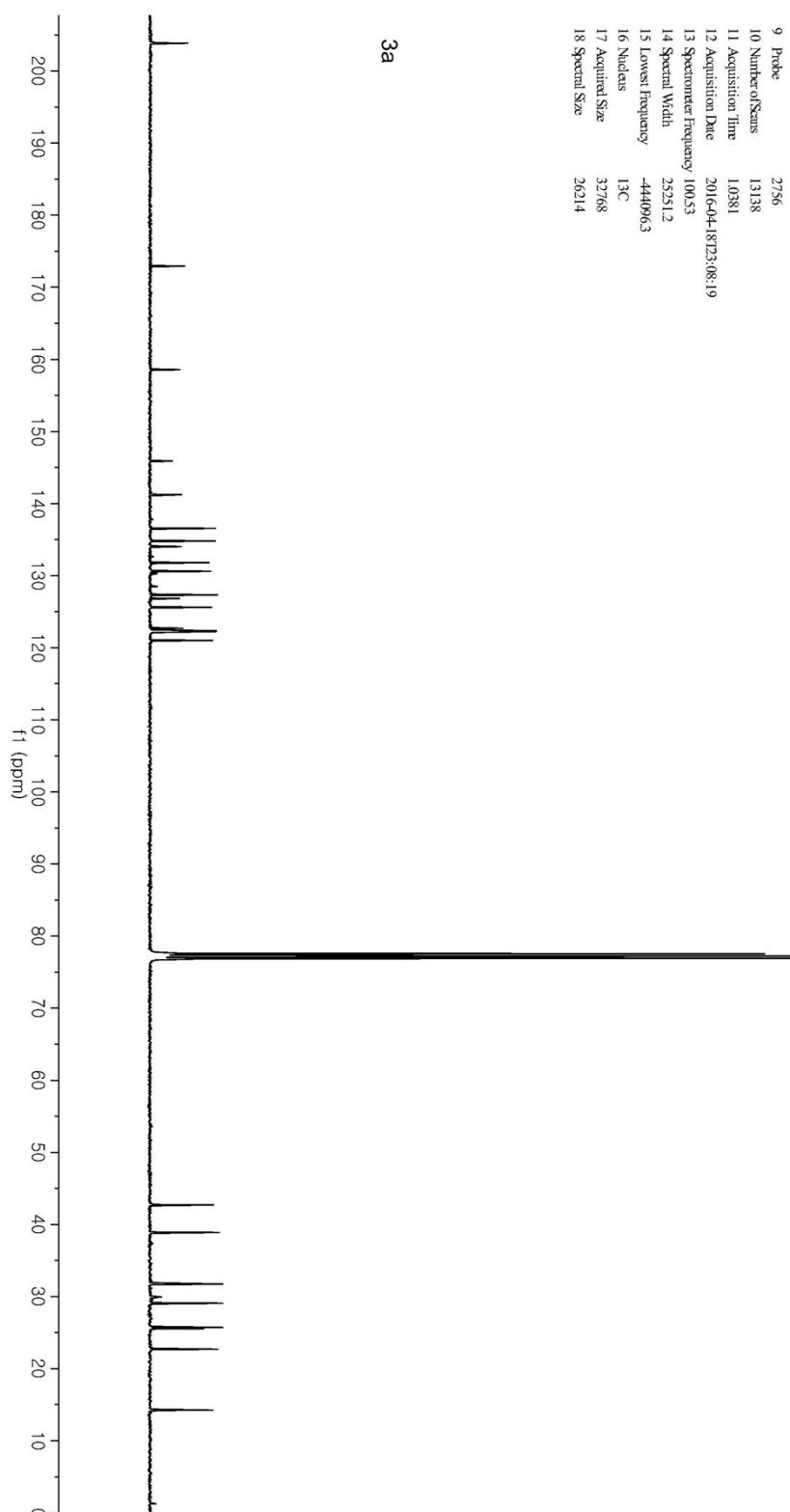


¹H NMR spectrum of **3a** recorded in CDCl₃

jhh0372sp1 conc
single pulse decoupled gated NOE

Parameter	Value
1 Title	jhh0372sp1 conc
2 Comment	single pulse decoupled gated NOE
3 Origin	JEOL
4 Spectrometer	ECA
5 Solvent	Chloroform-d
6 Temperature	25.0
7 Pulse Sequence	carbonjnp
8 Experiment	1D
9 Probe	2756
10 Number of Scans	1338
11 Acquisition Time	1.0381
12 Acquisition Date	2016-04-18T23:08:19
13 Spectrometer Frequency	100.53
14 Spectral Width	25251.2
15 Lowest Frequency	-444096.3
16 Nucleus	¹³ C
17 Acquired Size	32768
18 Spectral Size	26214

3a



¹³C NMR spectrum of **3a** recorded in CDCl₃

Parameter	Value
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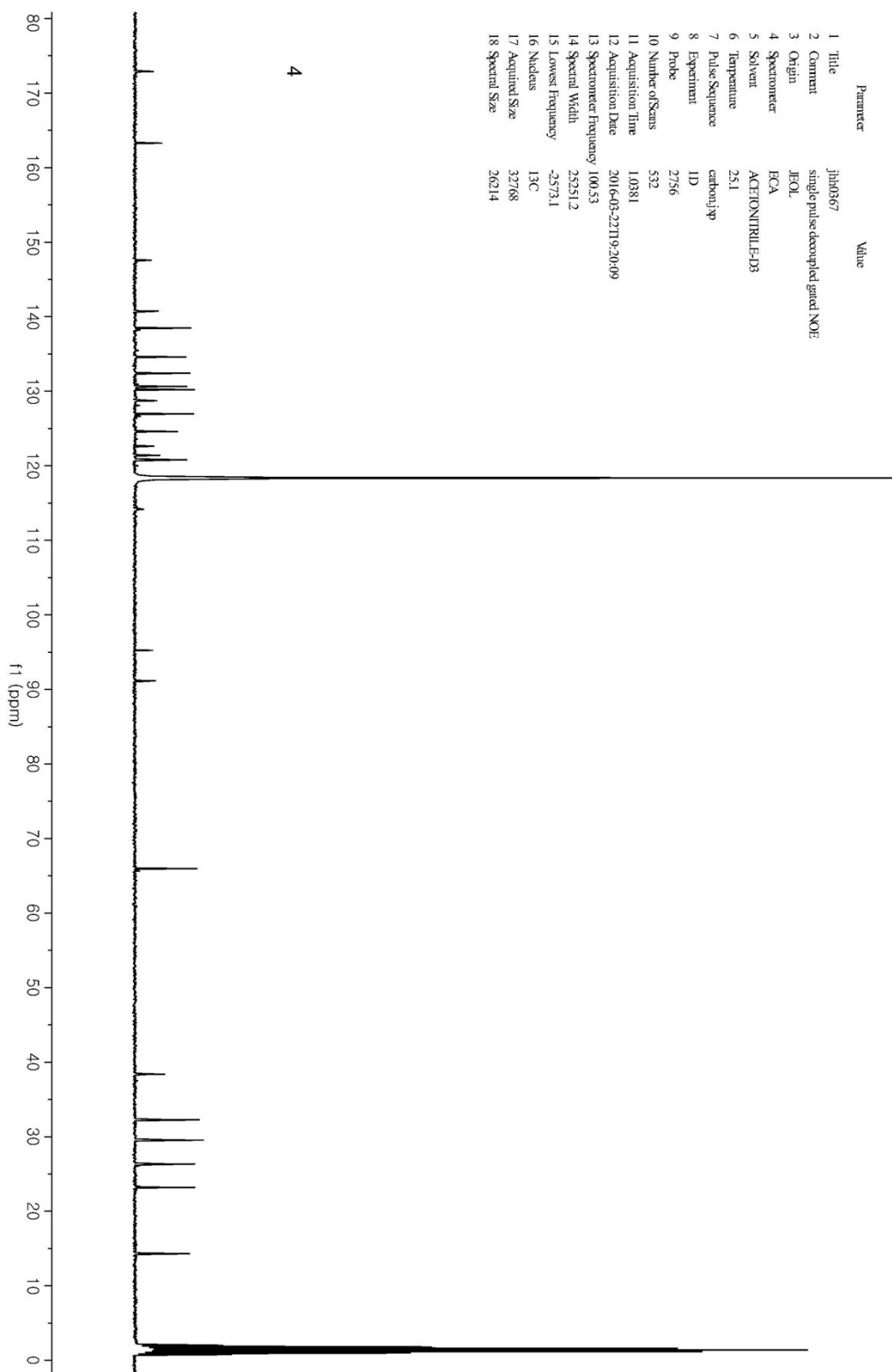
	single_pulse
2. Comment	
3. Origin	JFOL
4. Spectrometer	ECA
5. Solvent	ACETONITRILE-D3
6. Temperature	25.7
7. Pulse Sequence	proton_jmp
8. Experiment	1D
9. Probe	2746
10. Number of Scans	8
11. Acquisition Time	2.1863



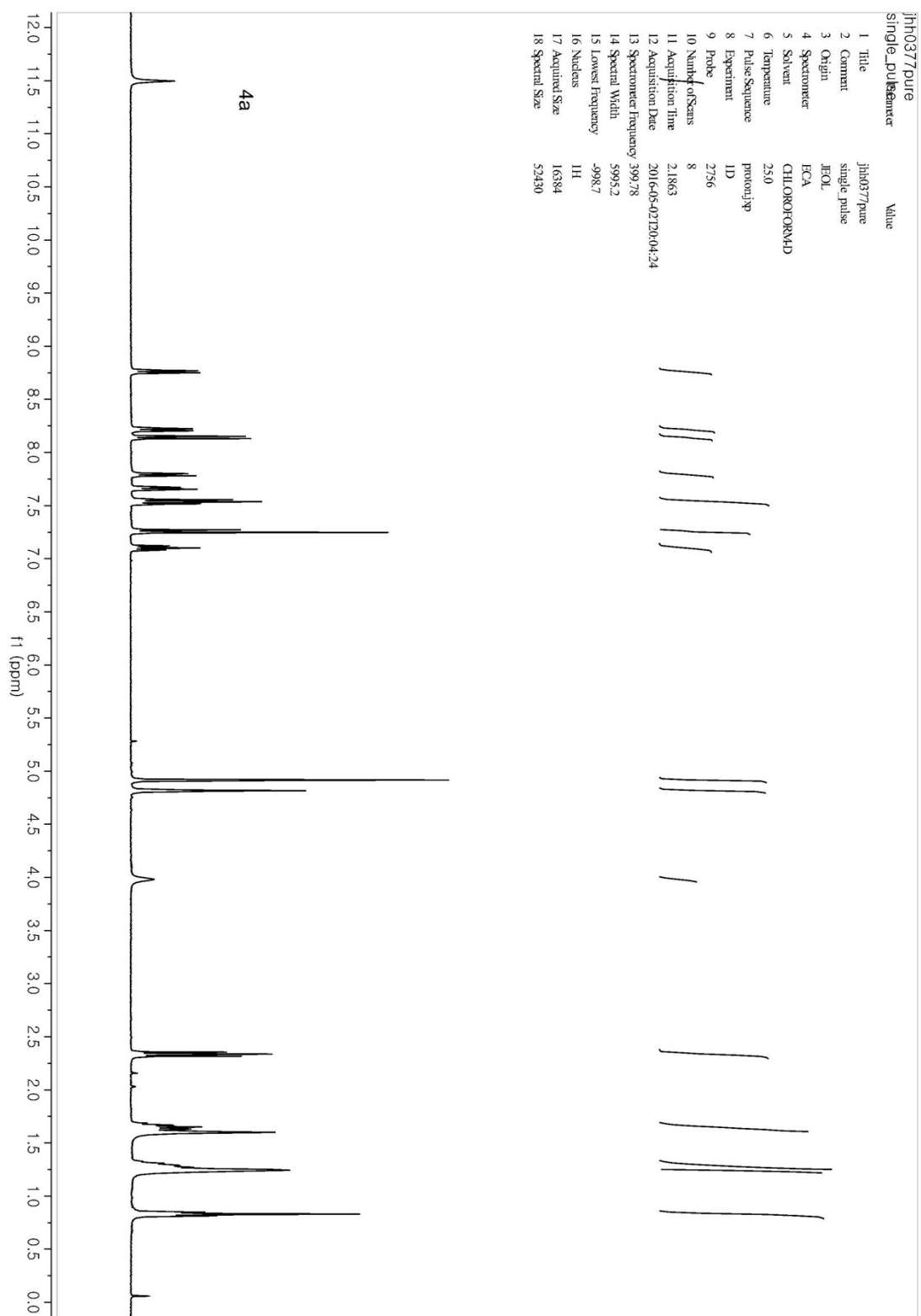
S22

jfh0367
single pulse decoupled gated NOE

Parameter	Value
1 Title	jfh0367
2 Comment	single pulse decoupled gated NOE
3 Origin	JFOL
4 Spectrometer	ECA
5 Solvent	ACETONITRILE-D3
6 Temperature	25.1
7 Pulse Sequence	carbou.jp
8 Experiment	1D
9 Probe	2756
10 Number of Scans	532
11 Acquisition Time	1.0381
12 Acquisition Date	2016-03-22T19:20:09
13 Spectrometer Frequency	100.53
14 Spectral Width	25251.2
15 Lowest Frequency	-2573.1
16 Nucleus	¹³ C
17 Acquired Size	32768
18 Spectral Size	26214



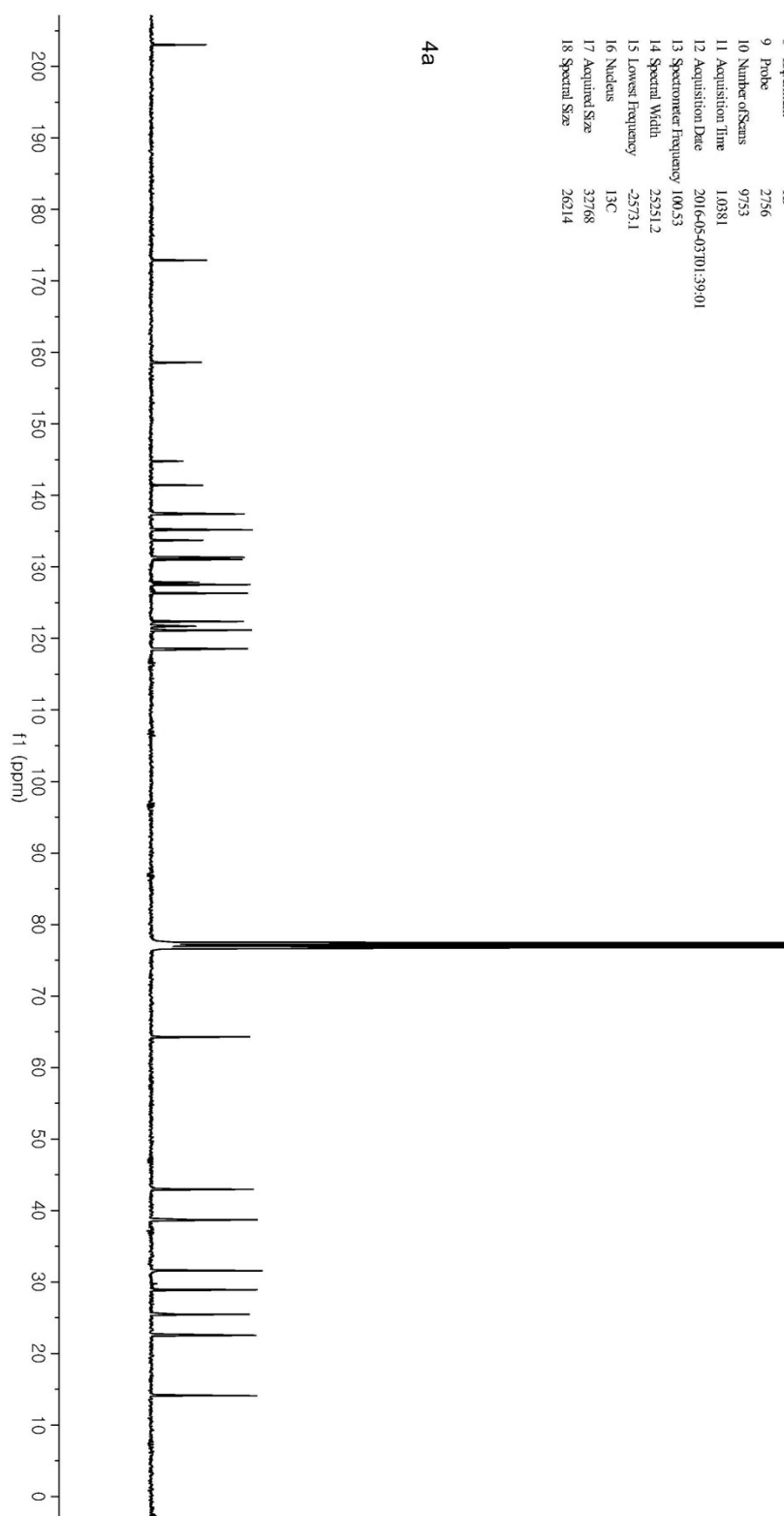
¹³C NMR spectrum of **4** recorded in CD₃CN



¹H NMR spectrum of **4a** recorded in CDCl₃

9	Probe	2756
10	Number of Scans	9753
11	Acquisition Time	1.0381
12	Acquisition Date	2016-05-03 10:13:9.01
13	Spectrometer Frequency	100.53
14	Spectral Width	25251.2
15	Lowest Frequency	-2573.1
16	Nucleus	¹³ C
17	Acquired Size	32768
18	Spectral Size	26214

4a



¹³C NMR spectrum of **4a** recorded in CDCl₃