Electronic Supplementary Material (ESI) for Chemical Communications. This journal is © The Royal Society of Chemistry 2016

Supplementary Information

Boronate based fluorescence (ESIPT) probe for peroxynitrite

Adam C. Sedgwick, a Xiaolong Sun, a Gyoungmi Kim, b Juyoung Yoon, *b Steven D. Bull*a and Tony D. James*a

Content

- 1. Preparation of ROS/RNS
- 2. Fluorescence analysis
- 3. UV analysis
- 4. Cell images and viability
- 5. Experimental
- 6. NMR

^a Department of Chemistry, University of Bath, BA2 7AY, UK. Email: S.D.Bull@bath.ac.uk, T.D.James@bath.ac.uk

^b Department of Chemistry and Nano Science, Ewha Womans University, Seoul 120-750, Korea. E-mail: jyoon@ewha.ac.kr.

1. Preparation of ROS/RNS

NO.

Nitric oxide (NO) was prepared by treating sulfuric acid (3.6 M) solution with sodium nitrite solution (7.3M) and its stock solution (2.0 mM) was prepared by bubbling NO into deoxygenated deionized water for 30 min

ROO•

ROO was generated from 2,2'-azobis (2-amidinopropane) dihydrochloride. AAPH (2, 2' azobis (2-amidinopropane) dihydrochloride,1 M) was added into deionizer water, and then stirred at 37 °C for 30 min;

$^{-}O_{2}$

Superoxide was generated from KO_2 . KO_2 and 18-crown-6 ether (2.5 eq) was dissolved in DMSO to afford a 0.25 M solution.

·HO

Hydroxyl radical was generated by the Fenton reaction. To prepare •OH solution, hydrogen peroxide (H₂O₂, 10 eq) was added to Fe(ClO₄)₂ in deionised water.

ONOO.

Simultaneously, 0.6 M KNO₂, 0.6 M in HC1, 0.7 M in H₂O₂ was added at to a 3 M NaOH solution at 0 °C. The concentration of peroxynitrite was estimated by using extinction co-efficient of 1670 cm⁻¹M⁻¹ at 302 nm in 0.1 M sodium hydroxide aqueous solutions

OCI

The concentration of ${}^{-}$ OCl was determined from the absorption at 292 nm ($\varepsilon = 350 \text{ M}^{-1}\text{cm}^{-1}$).

H_2O_2

The concentration of H_2O_2 was determined from the absorption at 240 nm ($\varepsilon = 43.6$ M-1cm-1).

2. Fluorescence analysis

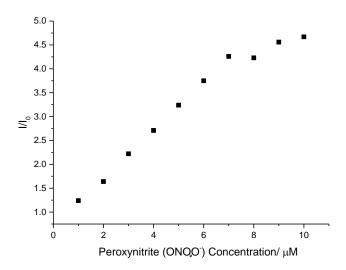


Fig S1: Fluorescence intensity changes (I/I₀) for probe 3 (0.25 μ M) with increasing concentrations of peroxynitrite (ONOO⁻) λ_{ex} 400 nm/ λ_{em} 461 nm in pH 8.2 buffer solution [52 wt% methanol]. each measurement was made after 1 minute.

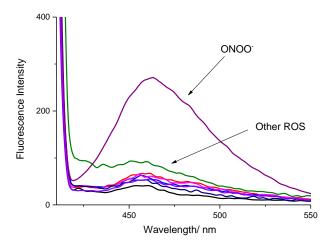


Fig S2: Fluorescence spectra of probe **3** (0.25 μM) in the presence of various ROS/RNS: ONOO (10 μM, 1 min), OCI (100 μM, 30 min), H₂O₂ (100 μM, 30 min), ROO (100 μM, 30 min), NO (100 μM, 1 min) O₂ (100 μM, 1 min), OH (100 μM, 1 min) λ_{ex} 400 nm in pH 8.2 buffer solution [52 wt% methanol].

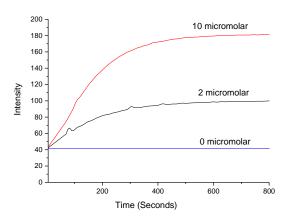


Fig S3: Fluorescence intensity changes for probe **3** (0.25 μ M) as a function of time (Seconds) with increasing concentrations of ONOO (0, 2, 10 μ M), λ_{ex} 400nm, λ_{em} 460 nm in pH 8.2 buffer solution [52 wt% methanol].

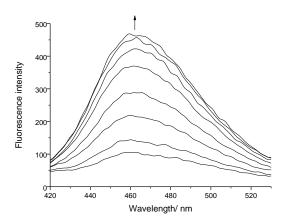


Fig S4: Fluorescence spectra of probe 3 (0.5 μ M) with the addition of hydrogen peroxide (0-14 mM), λ_{ex} 400 nm in pH 8.2 buffer solution [52 wt% methanol] each measurement was made after 30 minutes.

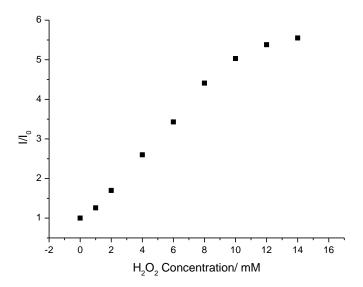


Fig S5: Fluorescence intensity changes (I/I₀) for probe 3 (0.5 μ M) with increasing concentrations of hydrogen peroxide (H₂O₂) λ_{ex} 400 nm/ λ_{em} 461 nm in pH 8.2 buffer solution [52 wt% methanol] each measurement was made after 30 minutes.

3. UV analysis

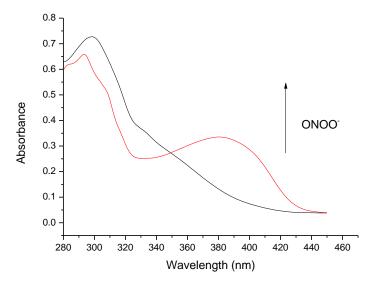


Fig S6: UV spectra of probe 3 (10 μ M), with and without ONOO in pH 8.2 buffer solution [52 wt% methanol].

4. Cell images and viability

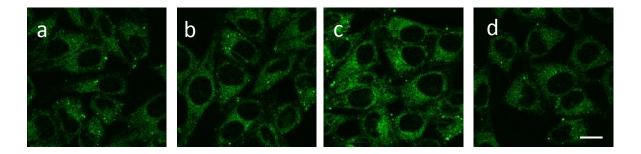


Fig S7: Fluorescence property in the live cell. HeLa cells were incubated with 20 μ M probe 3 for 30 min and washed with DPBS and added (a) 0, (b) 30, (c) 100 μ M ONOO- and (d) after pretreatment with 100 μ M ebselen, add 100 μ M ONOO- for 30 min. Fluorescence images were acquired by confocal microscopy. λ_{ex} . 473 nm/ λ_{em} 490 – 590 nm. Scale bar: 10 μ M

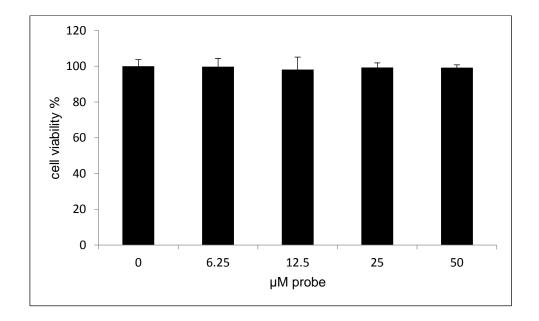


Fig S8: HeLa cells were incubated with each concentration of probe for 24hr. Cell viability was assayed by MTT test. Results are expressed as mean \pm standard deviation of three independent experiments.

5. Experimental

Scheme S1: (i) Isatoic anyhydride, NaOAc, AcOH, 1.5 h, 72 % yield (ii) 4, 4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)benzaldehyde, NaBH(OAc)₃, AcOH, DCE, 16 h, 78 % yield (iii) 5, NaCNBH₃, Formaldehyde – 35 % in H_2O , AcOH, MeCN, 17 h, 14 % yield.

2-(benzo[d]thiazol-2-yl)aniline (4)

$$S$$
 H_2N

Isatoic anhydride (2.93 g, 15.27 mmol), 2-Aminothiophenol (2.55 mL, 20.37 mmol), Sodium acetate (1.2 g, 12.43 mmol) in AcOH (120 mL) was refluxed for 1.5 h under Argon. The reaction was cooled to rt and EtOAc (100 mL) was added and the solution was quenched with solid NaHCO₃. The organic layer was washed with H₂O (2 x 50 mL), brine (50 mL), dried (MgSO₄) and concentrated *in vacuo* to afford the crude material. The crude product was purified via column chromatography 5:95 (EtOAc:Pet ether) to afford a cream solid (3.33 g, 14.7 mmol, 72 %). M.p. 123-127 °C. 1 H NMR (500MHz, CDCl₃) $\delta_{\rm H}$ 7.97 (d, J = 8.8 Hz, 1 H, ArH), 7.88 (d, J = 9.3 Hz, 1 H, ArH), 7.71 (dd, J = 1.5, 7.8 Hz, 1 H, ArH), 7.50 - 7.42 (m, 1 H, ArH), 7.40 - 7.33 (m, 1 H, ArH), 7.23 (ddd, J = 1.5, 7.2, 8.4 Hz, 1 H, ArH), 6.79 (dd, J = 1.0, 8.3 Hz, 1 H, ArH), 6.76 - 6.73 (m, 1 H, ArH), 6.40 (br. s., 2 H, Ar-NH₂); 13 C NMR (125.75 MHz, CDCl₃) $\delta_{\rm C}$ 169.2, 153.7, 146.7, 133.3, 131.5, 130.3, 126.0, 124.8, 122.4, 121.2, 116.9, 116.8, 115.3; I.R (thin film) v max (cm $^{-1}$): 3461.97 (N-H); HRMS (ESI): m/z calculated for C₁₃H₁₀N₂S: requires 227.0643 for [M+H] $^+$, found 227.0637; requires 249.0462 for [M+Na] $^+$, found 249.0443

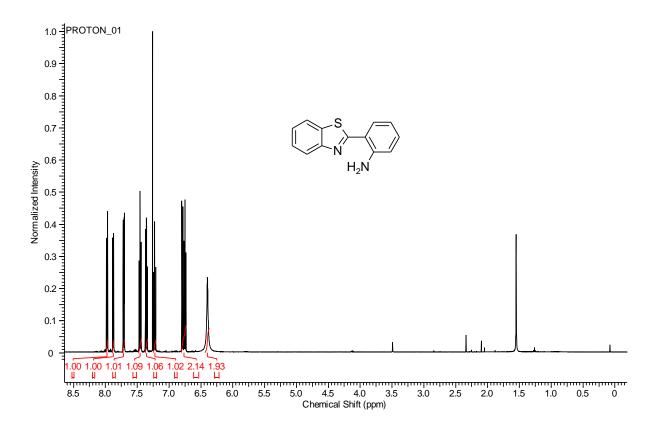
2-(benzo[d]thiazol-2-yl)-N-(4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)benzyl)aniline (5)

4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)benzaldehyde (1.18 g, 5.08 mmol) was added to a solution of **4** (0.72 g, 3.18 mmol) in DCE (10 mL) and AcOH (1.11 mL). The reaction mixture was stirred for 1 h before the addition of NaBH(OAc)₃ (1.75 g, 8.26 mmol). The reaction mixture was stirred for 24 h before being quenched with saturated NaHCO₃ solution. The organic layer was washed with H₂O (3 x 50 mL), brine (50 mL), dried (MgSO₄) and concentrated *in vacuo* to afford the crude product. The product was purified via column chromatography 10:90 (EtOAc:Pet ether) to afford a yellow oil (1.10 g, 2.49 mmol, 78 %). ¹H NMR (500MHz, CDCl₃) $\delta_{\rm H}$ 9.52 (t, J = 5.4 Hz, 1 H, N-H), 7.93 (d, J = 7.8 Hz, 1 H, ArH), 7.89 (d, J = 7.3 Hz, 1 H, ArH), 7.83 (d, J = 7.8 Hz, 2 H, ArH), 7.78 (dd, J = 1.5, 8.3 Hz, 1 H, ArH), 7.27 - 7.20 (m, 1 H, ArH), 6.74 - 6.68 (m, 2 H, ArH), 4.65 (d, J = 5.4 Hz, 2 H, NHCH₂Ar), 1.36 (s, 12 H, BPin); ¹³C NMR (125.75 MHz, CDCl₃) $\delta_{\rm C}$ 169.5, 153.5, 147.3, 142.5, 135.1, 133.1, 132.0, 130.6, 126.2, 126.0, 124.8, 122.3, 121.1, 115.6, 115.0, 111.9, 83.7, 47.3, 24.9; I.R (thinfilm) v max (cm⁻¹): 3282.94 (N-H); HRMS (ESI): m/z calculated for C₂₆H₂₇BN₂O₂S: requires 443.1965 for [M+H]⁺, found 443.2007

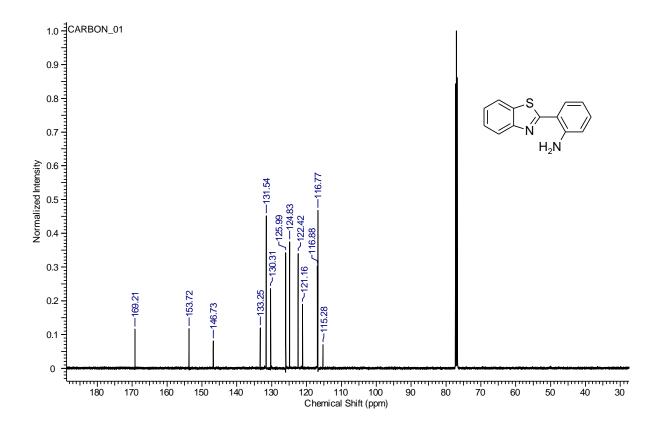
2-(benzo[d]thiazol-2-yl)-N-methyl-N-(4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)benzyl)aniline (3)

A solution of **5** (0.82 g, 1.85 mmol), formaldehyde -37 % in H₂O (1.3 mL, 18.54 mmol), AcOH (1.6 mL, 27.75 mmol) in MeCN (20 mL) was cooled to 0 °C before the portion wise addition of NaBH₃CN (0.3 g, 4.81 mmol). The reaction was stirred for 24 h before being quenched with saturated NaHCO₃ solution. Ether (100 mL) was then added and the organic layer was washed with H₂O (3 x 50 mL), brine (50 mL), dried (MgSO₄) and concentrated *in vacuo* to afford the crude material. The product was purified *via* column chromatography 5:95 (EtOAc:Pet ether) to afford a pale green foam (0.12 g, 0.263 mmol, 14 %). ¹H NMR (500MHz, CDCl₃) δ = 8.39 (dd, J = 2.0, 7.8 Hz, 1 H, ArH), 8.10 (d, J = 8.3 Hz, 1 H, ArH), 7.93 (d, J = 7.3 Hz, 1 H, ArH), 7.79 (d, J = 7.8 Hz, 2 H, ArH), 7.53 - 7.46 (m, 1 H, ArH), 7.42 - 7.33 (m, 4 H, ArH), 7.26 - 7.20 (m, 1 H, ArH), 7.16 (d, J = 7.8 Hz, 1 H, ArH), 4.20 (s, 2 H, CH₃NCH₂Ar), 2.66 (s, 3 H, CH₃NCH₂Ar), 1.36 (s, 12 H, BPin); ¹³C NMR (125.75 MHz, CDCl₃) δ = 165.0, 152.2, 151.4, 140.0, 136.7, 134.6, 130.9, 130.4, 129.0, 125.8, 124.9, 124.3, 122.9, 122.1, 121.4, 83.8, 61.1, 43.0, 24.9; HRMS (ESI): m/z calculated for C₂₇H₂₉BN₂O₂S: requires 479.1940 for [M+Na]⁺, found 479.1963

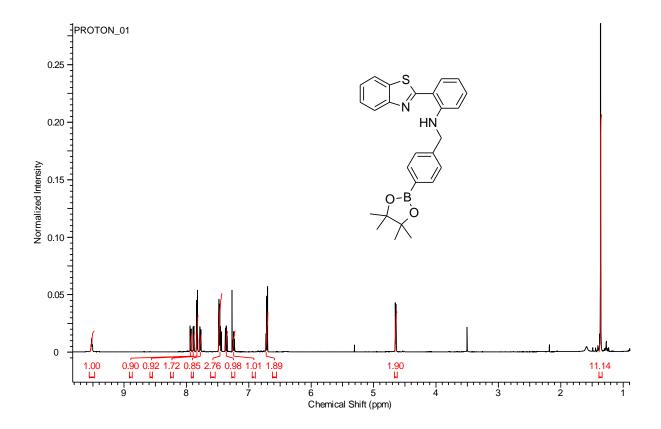
2-(benzo[d]thiazol-2-yl)aniline (4) (500 MHz, CDCl₃)



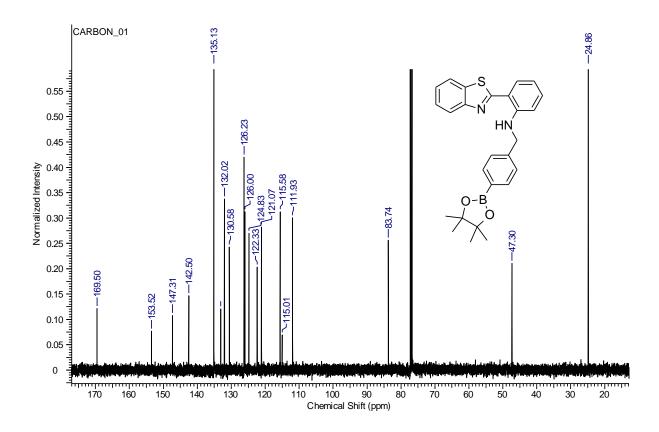
2-(benzo[d]thiazol-2-yl)aniline (4) (125.75 MHz, CDCl₃)



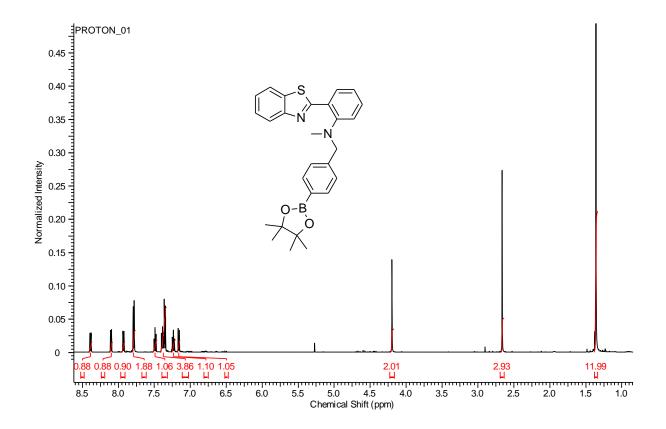
$\begin{tabular}{ll} 2-(benzo[d]thiazol-2-yl)-N-(4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)benzyl) aniline (5) (500 MHz, CDCl_3) \end{tabular}$



$\begin{tabular}{ll} 2-(benzo[d]thiazol-2-yl)-N-(4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)benzyl) aniline (5) (125.75 MHz, CDCl_3) \end{tabular}$



$\begin{tabular}{ll} 2-(benzo[d]thiazol-2-yl)-N-methyl-N-(4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)benzyl) aniline (3) (500 MHz, CDCl_3) \\ \end{tabular}$



$\begin{tabular}{ll} 2-(benzo[d]thiazol-2-yl)-N-methyl-N-(4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)benzyl) aniline (3) (125.75~MHz, CDCl_3) \\ \end{tabular}$

