Electronic supporting information

Preparation of glass slides with immobilised MIPs for SEM analysis

In order to prepare the surface for the immobilisation of nanoparticles the glass slide was incubated with 2 M NaOH for 15 min. The slide was then rinsed with water, neutralised with HCl and thoroughly washed with water. After the slide was dried at 100 °C for 30 minutes, the solution of 4% 3-(triethoxysilyl)propyl isocyanate in dry toluene was added and kept overnight at a room temperature completely covered. The slide was then removed and the mixture of amine-coated nanoparticles were added to cover and left for 2 hours. This slide was then washed with water and left to dry at room temperature.

The nanoMIPs have been coated with amino-groups using 7.5 mg of the amine 3-AMPA (N-(3aminopropyl) methacrylamide) in 15 mL of acetonitrile mixed with 11 drops of water solution. This mixture was purged with nitrogen for 5 min then poured over the solid phase containing MIPs formed around the immobilised trisaccharide before MIP elution. The beads were then exposed to UV light under conditions identical to those used for initial polymerisation for 35 seconds, before elution steps were performed as described in the paper.

Live subject statement

All blood tests described in the project were performed in the laboratory of Professor of Haemato-Oncology Martin Dyer under supervision of Dr Sandrine Jayne, Department of Cancer Studies, University of Leicester. I would like to acknowledge that Department of Cancer Studies, University of Leicester is holding the licence for the blood testing and that all procedures were performed in compliance with the relevant laws and institutional guidelines and that informed consent was obtained for the experimentation with human subjects.