

Supporting Information

Aptamer-tagged DNA Origami for Spatially Addressable Detection of Aflatoxin B1

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1. Materials and methods

1.1. Materials and reagents

Single-stranded *M13mp18* DNA was purchased from New England Biolabs (Catalog number: #N4040S). All unmodified staple strands and thiol-modified complementary ssDNA were synthesized and purified by Shanghai Sangon Biotech Inc. Magnesium acetate tetrahydrate ($C_4H_6MgO_4 \cdot 4H_2O$), sodium citrate tribasic dehydrate ($C_6H_5Na_3O_7 \cdot 2H_2O$), Tris (2-carboxyethyl) phosphine hydrochloride (TCEP), chloroauric acid ($HAuCl_4$) were purchased from Aladdin. Bis (p-sulfonatophenyl) phenylphosphine dihydrate dipotassium salt (BSPP), sodium chloride, aflatoxin B1 were bought from Sigma-Aldrich. All other reagents are of analytical grade and obtained from commercial sources.

1.2. Instruments

A PCR machine (SelectCycler II Thermal Cycler) was used to prepare DNA origami. The images of the DNA origami were obtained with an atomic force microscope (AFM) (DIMENSION ICON, Bruker Inc. USA) in ambient conditions. The morphologies of the AuNPs were characterized with field emission scanning electron microscope (SM-7800F, JEOL, Tokyo, Japan). The UV-vis spectra, hydrodynamic sizes and zeta potentials of AuNPs before/after modifications were measured using UV-VIS spectrophotometer (UV-2550, SHIMADZU) and Zetasizer Nano (Malvern Instruments Ltd), respectively.

1.3. DNA sequences

DNA origami substrates were prepared according to the method reported by Rothmund^[1] with minor modifications.

The sequences of the staple DNA strands are listed as below (From 5' end to 3' end):

t1s18h,D1, AATACTGCGGAATCGTAGGGGGTAATAGTAAAATGTTTACT

t1s28h,E1, TCTTTGATTAGTAATAGTCTGTCCATCACGCAAATTAACCGTT

t1s8h,F1, CAGAAGGAAACCGAGGTTTTTAAGAAAAGTAAGCAGATAGCCG

t1s10g,B2, GACGGGAGAATTAACCTCGGAATAAGTTTATTTCCAGCGCC

t1s12i,C2, TCATATGTGTAATCGTAAACTAGTCATTTTC

t1s14i,D2, GTGAGAAAATGTGTAGGTAAAGATACAACCTT

t1s16i,E2, GGCATCAAATTTGGGGCGCGAGCTAGTTAAAG

t1s18i,F2, TTCGAGCTAAGACTTCAAATATCGGGAACGAG

t1s20g,G2, GAATACCACATTCAACTTAAGAGGAAGCCCGATCAAAGCG
t1s22i,H2, TCGGGAGATATACAGTAACAGTACAAATAATT
t1s24i,A3, CCTGATTAAAGGAGCGGAATTATCTCGGCCTC
t1s26i,B3, GCAAATCACCTCAATCAATATCTGCAGGTCGA
t1s28i,C3, CGACCAGTACATTGGCAGATTCACCTGATTGC
t1s2i,D3, CGGGGTTTCCTCAAGAGAAGGATTTTGAATTA
t1s30g,E3, TTGACGAGCACGTATACTGAAATGGATTATTTAATAAAAAG
t1s4i,F3, AGCGTCATGTCTCTGAATTTACCGACTACCTT
t1s6i,G3, TTCATAATCCCCTTATTAGCGTTTTTCTTACC
t1s8i,H3, ATGGTTTATGTCACAATCAATAGATATTTAAAC
t2s11g,A4, AGAAAAGCCCCAAAAAGAGTCTGGAGCAAACAATCACCAT
t2s13g,B4, ACAGTCAAAGAGAATCGATGAACGACCCCGGTTGATAATC
t2s15f,C4, ATAGTAGTATGCAATGCCTGAGTAGGCCGGAG
t2s17f,D4, AACCAGACGTTTAGCTATATTTTCTTCTACTA
t2s1g,E4, GATAAGTGCCGTCGAGCTGAAACATGAAAGTATACAGGAG
t2s21g,F4, CCTGATTGCTTTGAATTGCGTAGATTTTCAGGCATCAATA
t2s23g,G4, TGGCAATTTTTAACGTCAGATGAAAACAATAACGGATTTCG
t2s25f,H4, AAGGAATTACAAAGAAACCACCAGTCAGATGA
t2s27f,A5, GGACATTCACCTCAAATATCAAACACAGTTGA
t2s3g,B5, TTTGATGATTAAGAGGCTGAGACTTGCTCAGTACCAGGCG
t2s5f,C5, CCGGAACCCAGAATGGAAAGCGCAACATGGCT
t2s7f,D5, AAAGACAACATTTTCGGTCATAGCCAAAATCA
t3s10g,E5, GTCAGAGGGTAATTGATGGCAACATATAAAAGCGATTGAG
t3s14e,F5, CAATATGACCCTCATATATTTTAAAGCATTAA
t3s16e,G5, CATCCAATAAATGGTCAATAACCTCGGAAGCA
t3s18g,H5, AACTCCAAGATTGCATCAAAAAGATAATGCAGATACATAA
t3s20g,A6, CGCCAAAAGGAATTACAGTCAGAAGCAAAGCGCAGGTCAG
t3s24e,B6, TAATCCTGATTATCATTTTGGCGGAGAGGAAGG
t3s26e,C6, TTATCTAAAGCATCACCTTGCTGATGGCCAAC
t3s28g,D6, AGAGATAGTTTGACGCTCAATCGTACGTGCTTTCCTCGTT

t3s30g,E6, AGAATCAGAGCGGGAGATGGAAATACCTACATAACCCTTC
t3s4e,F6, TGTACTGGAAATCCTCATTAAAGCAGAGCCAC
t3s6e,G6, CACCGGAAAGCGCGTTTTTCATCGGAAGGGCGA
t3s8g,H6, CATTCAACAAACGCAAAGACACCAGAACACCCTGAACAAA
t4s11g,A7, GCAAATATTTAAATTGAGATCTACAAAGGCTACTGATAAA
t4s13g,B7, CGTTCTAGTCAGGTCATTGCCTGACAGGAAGATTGTATAA
t4s15f,C7, CAGGCAAGATAAAAATTTTTAGAAATATTCAAC
t4s17f,D7, GATTAGAGATTAGATACATTTTCGCAAATCATA
t4s1g,E7, TAGCCCGGAATAGGTGAATGCCCCCTGCCTATGGTCAGTG
t4s21g,F7, GCGCAGAGGCGAATTAATTATTTGCACGTAAATTCTGAAT
t4s23g,G7, GATTATACACAGAAATAAAGAAATACCAAGTTACAAAATC
t4s25f,H7, TAGGAGCATAAAAAGTTTGAGTAACATTGTTTG
t4s27f,A8, TGACCTGACAAATGAAAAATCTAAAATATCTT
t4s3g,B8, TTTAACGGTTCGGAACCTATTATTAGGGTTGATATAAGTA
t4s5f,C8, CTCAGAGCATATTCACAAACAAATTAATAAGT
t4s7f,D8, GGAGGGAATTTAGCGTCAGACTGTCCGCCTCC
t5s10g,E8, GATAACCCACAAGAATGTTAGCAAACGTAGAAAATTATTC
t5s14e,F8, TTAATGCCTTATTTCAACGCAAGGGCAAAGAA
t5s16e,G8, TTAGCAAATAGATTTAGTTTGACCAGTACCTT
t5s18g,H8, TAATTGCTTTACCCTGACTATTATGAGGCATAGTAAGAGC
t5s20g,A9, AACACTATCATAACCCATCAAAAATCAGGTCTCCTTTTGA
t5s24e,B9, AATGGAAGCGAACGTTATTAATTTCTAACAAAC
t5s26e,C9, TAATAGATCGCTGAGAGCCAGCAGAAGCGTAA
t5s28g,D9, GAATACGTAACAGGAAAAACGCTCCTAACAGGAGGCCGA
t5s30g,E9, TTAAAGGGATTTTAGATACCGCCAGCCATTGCGGCACAGA
t5s4e,F9, CCTTGAGTCAGACGATTGGCCTTGCGCCACCC
t5s6e,G9, TCAGAACCCAGAATCAAGTTTGCCGGTAAATA
t5s8g,H9, TTGACGGAAATACATACATAAAGGGCGCTAATATCAGAGA
t6s15g,B10, ATAAAGCCTTTGCGGGAGAAGCCTGGAGAGGGTAG
t6s17f,C10, TAAGAGGTCAATTCTGCGAACGAGATTAAGCA

t6s25g,E10, TCAATAGATATTAAATCCTTTGCCGGTTAGAACCT
t6s27f,F10, CAATATTTGCCTGCAACAGTGCCATAGAGCCG
t6s5g,H10, CAGAGCCAGGAGGTTGAGGCAGGTAACAGTGCCCG
t6s7f,A11, ATTAAAGGCCGTAATCAGTAGCGAGCCACCCT
t7s10g,B11, ATAAGAGCAAGAAACATGGCATGATTAAGACTCCGACTTG
t7s14e,C11, ATGACCCTGTAATACTTCAGAGCA
t7s16e,D11, TAAAGCTATATAACAGTTGATTCCCATTTTTG
t7s18g,E11, CGGATGGCACGAGAATGACCATAATCGTTTACCAGACGAC
t7s20g,F11, GATAAAAACCAAAATATTAAACAGTTCAGAAATTAGAGCT
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t7s8g,E12, CACCGTCACCTTATTACGCAGTATTGAGTTAAGCCCAATA
t8s17g,G12, TAATTGCTTGGAAGTTTCATTCCAAATCGGTTGTA
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t9s18g,F1, TGCTGTAGATCCCCCTCAAATGCTGCGAGAGGCTTTTGCA
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t-10s27h,F2, AACTCACATTATTGAGTGTTGTTCCAGAAACCGTCTATCAGGG

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t-12s19h,C3, CCTGACGAGAAACACCAGAACGAGTAGGCTGCTCATTAGTGA
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t-2s13g,G6, AGACGTTACCATGTACCGTAACACCCCTCAGAACCGCCAC

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t-2s17f,A7, ATTGTGTCTCAGCAGCGAAAGACACCATCGCC
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t-4s1g,B10, GAGCAAAAAGAAGATGAGTGAATAACCTTGCTTATAGCTTA
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t-4s23g,D10, GGATAGGTACCCGTCGGATTCTCCTAAACGTTAATATTTT
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t-6s15c,G12, CGAGGTGAGGCTCCAAAAGGAGCC
t-6s17f,H12, ACCCCCAGACTTTTTTCATGAGGAACTTGCTTT
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t-6s25c,B1, TGGCGAAATGTTGGGAAGGGCGAT
t-6s27f,C1, TGTCGTGCACACAACATACGAGCCACGCCAGC
t-6s3f,D1, TCCCTTAGAATAACGCGAGAAAATTTTACCGACC
t-6s5c,E1, GTTTGAAATTCAAATATATTTTAG
t-6s7f,F1, AATAGATAGAGCCAGTAATAAGAGATTTAATG
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t-7s8g,G2, GCGCCTGTTATTCTAAGAACGCGATTCCAGAGCCTAATTT
t-7s8g,G2, GCGCCTGTTATTCTAAGAACGCGATTCCAGAGCCTAATTT
t-8s15f,H2, CGGTTTATCAGGTTTCCATTAAACGGGAATACACT

t-8s17c,A3, GGCAAAAGTAAAATACGTAATGCC
t-8s25f,B3, TCTTCGCTATTGGAAGCATAAAGTGTATGCCCCGCT
t-8s27c,C3, GCGCTCACAAGCCTGGGGTGCCTA
t-8s5f,D3, TTCTGACCTAAAATATAAAGTACCGACTGCAGAAC
t-8s7c,E3, TCAGCTAAAAAAGGTAAAGTAATT
t-9s10g,F3, ACGCTAACGAGCGTCTGGCGTTTTAGCGAACCCAACATGT
t-9s20g,H3, TGGTTTAATTTCAACTCGGATATTCATTACCCACGAAAGA
t-9s30g,B4, CGATGGCCCCACTACGTATAGCCCCGAGATAGGGATTGCGTT
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t-5s2e-t6s23c-3T,A6, TTAATTAATTTTTTACCATATCAAA
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t-9s6e-t10s27c-1T,C6, CTGTCCAGACGTATACCGAACGA
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t-9s16e-t10s7c-1T,G6, ACTACGAAGGCTTAGCACCATTA
t-11s18e-t12s9c-0T,H6, ATAAGGCTTGCAACAAAGTTAC
t-5s22e-t6s13c-3T,A7, GTGGGAACAAATTTCTATTTTTGAG
t-7s24e-t8s15c-2T,B7, CGGTGCGGGCCTTCCAAAAACATT
t-9s26e-t10s17c-1T,C7, ATGAGTGAGCTTTTAAATATGCA
t-11s28e-t12s19c-0T,D7, ACTATTAAGAGGATAGCGTCC

The blue colored staple strands (t1s14i, D2) were replaced by the AFB₁ probe in the designed origami.

Sequences of the AFB₁ capture probe:

GTTGGGCACGTGTTGTCTCTCTGTGTCTCGTGCCCTTCGCTAGG
CCCACTTTTTGTGAGAAAATGTGTAGGTAAAGATACAACCTTT^{[1][2]}

DNA stand complements with the AFB₁ probe:

AGAGACAACACGTGCCCAACAAAAA-(CH₂)₃-SH

2. Preparation of DNA origami substrate

The single-strand *M13mp18* DNA and the staple strands (including AFB₁ probe) were mixed at a molar ratio of 1:10 in a 1 × TAE~Mg²⁺ buffer (40 mM Tris, 20 mM acetic acid, 1 mM EDTA, 12.5 mM magnesium acetate). The mixture was annealed from 95°C to 20°C in PCR (1°C /100s).^[1] The DNA origami was washed with centrifugal filters for three times to remove excess staple strands (washing buffer: 1 × TAE~Mg²⁺ buffer).^[3]

3. Synthesis of AuNPs

AuNPs were synthesized by citrate reduction method^[4]. Briefly, 2 mL HAuCl₄ (1 wt%) was mixed with 198 mL deionized water in a round-bottom flask equipped with a reflux condenser. When the solution was heated to a boil, 4 mL sodium citrate (1%) was rapidly injected under stirring. The mixture was boiled for another 30 min until its color turned to deep red.

4. Preparation of the AuNP-complementary ssDNA conjugates

AuNP-DNA conjugates were synthesized according to Ding's route^[5]. Briefly, 15 mg BSPP was added into 50 mL AuNPs suspension (1.0 µg/mL), stirring overnight at room temperature. Solid sodium chloride was gradually added until the color of the solution changes from wine red to lilac. The suspension was centrifuged at 10,000 rpm for 15 min to pellet the AuNPs. After washing with a solution containing 1 mL BSPP (2.5 mM) and 1 mL CH₃OH, the pellet was dispersed in 1 mL BSPP (2.5mM). After activated by TCEP (10 mM) for 2 hours, thiol-modified DNA (100 µM) was incubated with AuNPs at a molar ratio of 200:1 in 0.5 × TBE~50 mM NaCl buffer for 40 hours at room temperature. After that, the products were collected and washed three-time with 0.5 × TBE~50 mM NaCl buffer by centrifugation at a speed of 13,000 rpm. The concentration of AuNPs-DNA conjugates was estimated according to the Wolfgang Haiss's approach.

5. AFB₁ detection with origami substrate

The AFB₁ probe-tagged DNA origami was incubated with AFB₁ at different molar ratio (1:100, 1:10, 1:1, 1:0) for 30 min at room temperature^[6]. The corresponding final concentrations are 80, 8, 0.8 and 0 ng/mL, respectively. Then, AuNPs-DNA conjugates were added into the mixture and slowly cooled down from 43°C to 20°C at a rate of 0.5°C per second in a PCR machine^[5]. During this process, all un-occupied probes can react with the AuNP-DNA conjugates.

6. AFM measurement

A 10- μL aqueous sample was dropped onto freshly cleaved mica, which was pre-treated with Mg^{2+} ions to enable the easy adsorption of DNA origami, drying at ambient conditions for 1 hour [7]. The dried samples were scanned with AFM at ScanAsyst-mode in air.

7. Gel electrophoresis

The samples were pre-mixed with NAS and then loaded in 1.5% agarose gels. After running for three hours at 80 V, [8, 9] the gels were photographed using a digital camera system (Chemi XR5).

8. Characterization of AuNPs

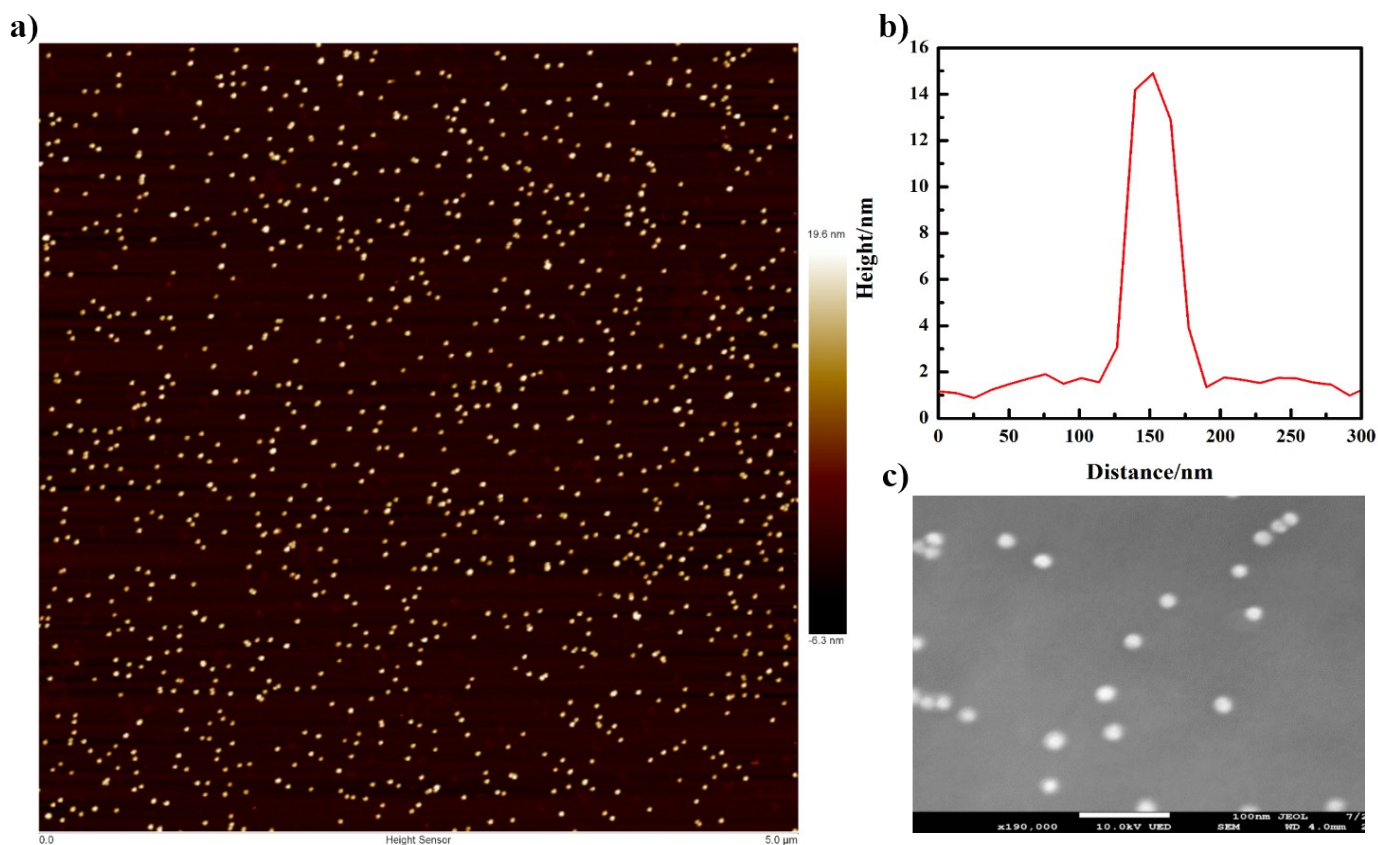


Figure S1. a) AFM and c) FE-SEM images of AuNPs; b) is an AFM section curve showing the size of a AuNP.

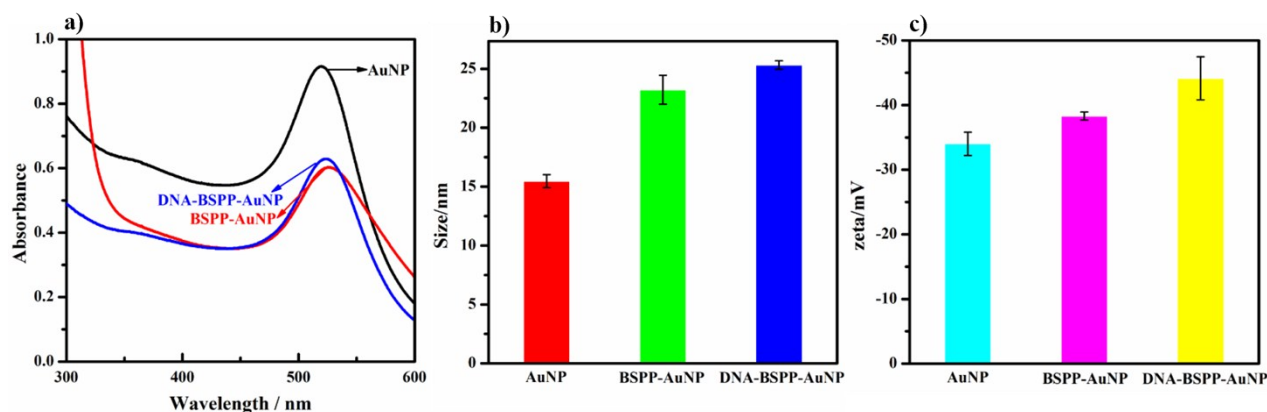


Figure S2. UV-vis (a), hydrodynamic size (b) and zeta potential (c) of AuNPs before and after modified with BSPP and SH-DNA. Data in b) and c) are presented as the average \pm standard deviation from three independent measurements.

As shown in the AFM image and the FESEM micrograph (Fig. S1), the as-prepared AuNPs are uniformly dispersed on the substrates and the average size of them is around 15 nm. The surface coating of BSPP and DNA causes the red-shift and significant decrease of the absorption peak (Fig. S2a). Both hydrodynamic size and zeta potential of AuNPs increase after surface modification with BSPP and DNA (Fig. S2b and S2c). The results clearly indicate the successful surface coating of BSPP and DNA on the AuNPs.

9. Relative sites between the origami and the anchored AuNP

Figure S3 illustrates the AFM images of the pure DNA origami and the DNA origami-AuNP complex. The attached AuNP could be located at the angle, side and center of the triangle DNA origami substrate. Since DNA origami is a highly addressable substrate, the possible resting site of the AuNP could be estimated based on the sizes of the origami and AuNP, as well as the lengths of the probe and complementary strands. As shown in Scheme S1, A is the designed site of the AFB1 probe. After hybridized with the AuNP-conjugated ssDNA, the AuNP could freely move in a circle with the A site as the center. The radius of the circle should be the length of linking strand plus the diameter of the AuNP. Thus, the relative sites observed in the AFM images could be well explained by the above calculation.

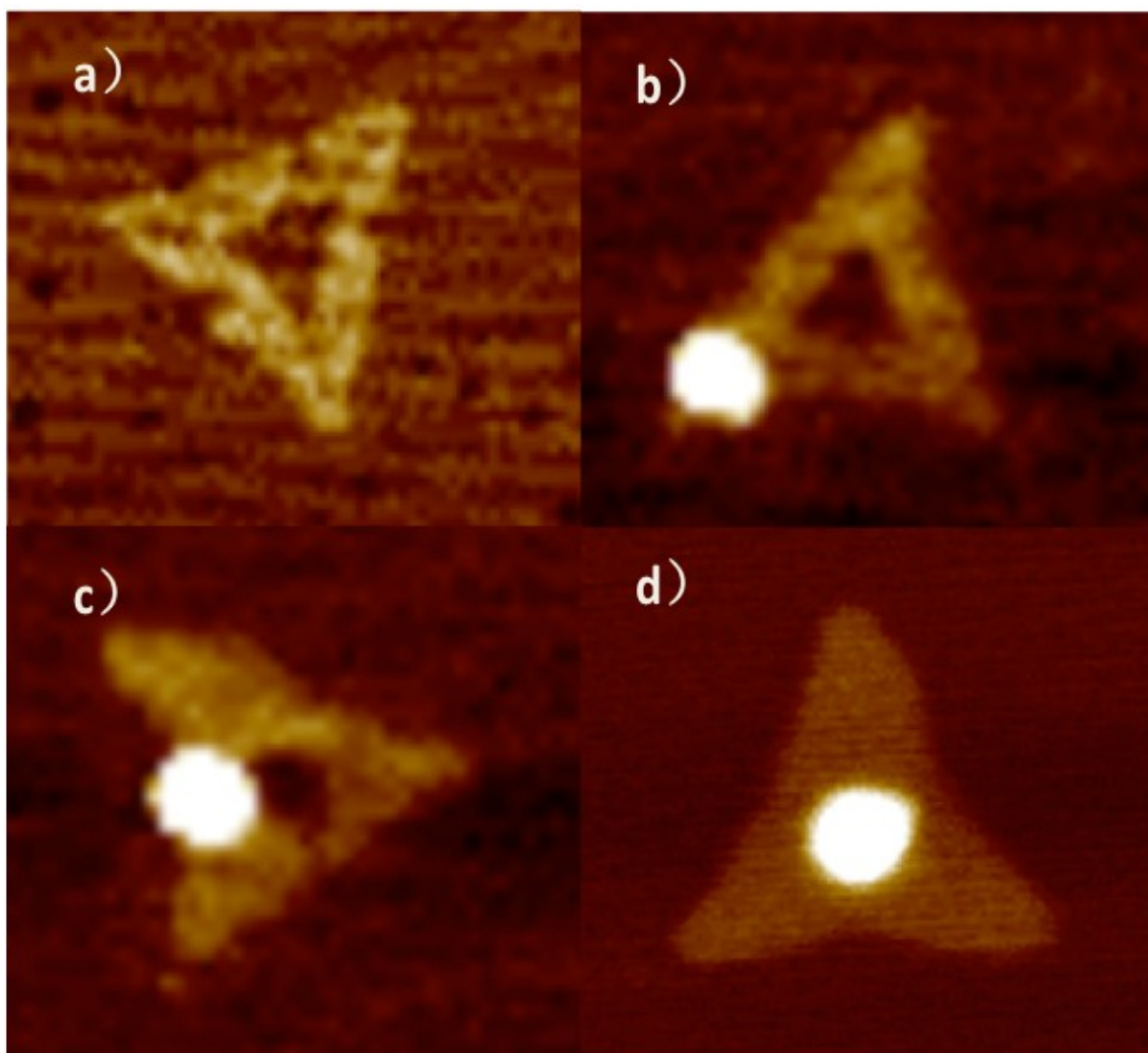
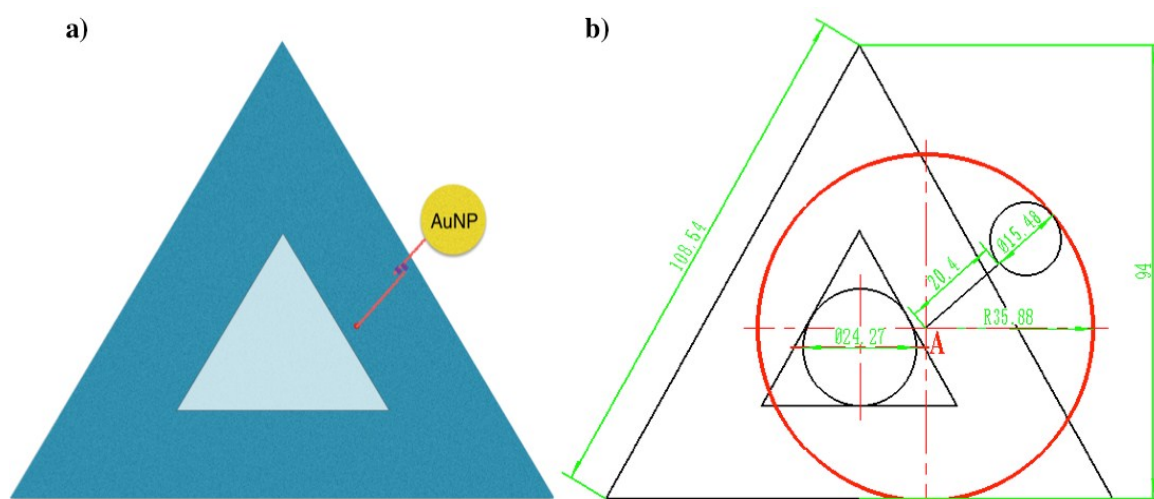


Figure S3. AFM images of the DNA origami and the DNA origami-AuNP complex. a) DNA origami with an AFB1 probe; DNA origami substrates with an angle- (b), side- (c) and center-attached (d) AuNP, respectively.



Scheme S1. Schematic drawing illustrating the possible sites of the origami-anchored AuNP.

Table S1. Binding rates of ssDNA-AuNPs conjugates on DNA origami templates in the detection systems containing different concentrations of AFB1.

Column	AFB1 Concentration	Mean	SD
a	0 ng / mL	64.65%	0.87
b	0.8 ng / mL	52.13%	1.93
c	8.0 ng / mL	47.43%	1.52
d	80.0 ng / mL	37.88%	2.45

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