

Supplementary Information: Why do Proton Conducting Polybenzimidazole Phosphoric Acid Membranes perform well in High-temperature PEM Fuel Cells?

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1 Experimental

Thermogravimetric Analysis (TGA) - Humidification A humidified TGA set up[9] was used to determine the equilibrium water uptake of phosphoric acid and phosphoric acid benzimidazole mixtures. In all cases nominally dry H_3PO_4 was used as a reference for $\lambda = 3$. A constant N_2 gas flow is led through water in a temperature controlled vessel (see figure 1) to ensure saturation of the gas. The water saturated gas is then led to the *sample chamber*, where the sample is placed in a teflon crucible magnetically coupled to a balance. Spatial separation of humidifier and sample chamber by the temperature controlled *transfer zone* ensures independent temperature control in the two compartments. With the temperature (in Celsius) in both humidifier T_W and sample chamber T_S the saturation partial water pressure in the respective compartments e_W and e_S are calculated according to the empirical equations:

$$\begin{aligned} e_{W,S}(T_{W,S}(^{\circ}\text{C})) &= 6.1078 \cdot 10^{7.5 \cdot T_i / (237.3 + T_i)} \text{ hPa} ; & T < 70^{\circ}\text{C} \\ e_{W,S}(T_{W,S}(^{\circ}\text{C})) &= 5.94062 \cdot 10^{7.28829 \cdot T_i / (226.531 + T_i)} \text{ hPa} ; & T > 70^{\circ}\text{C} \end{aligned} \quad (1)$$

obtained from a fit to literature data.[10] The relative humidity (RH) is calculated from the ratio of saturation water pressure in both chambers:

$$RH = \frac{e_W}{e_S} \cdot 100\% \quad (2)$$

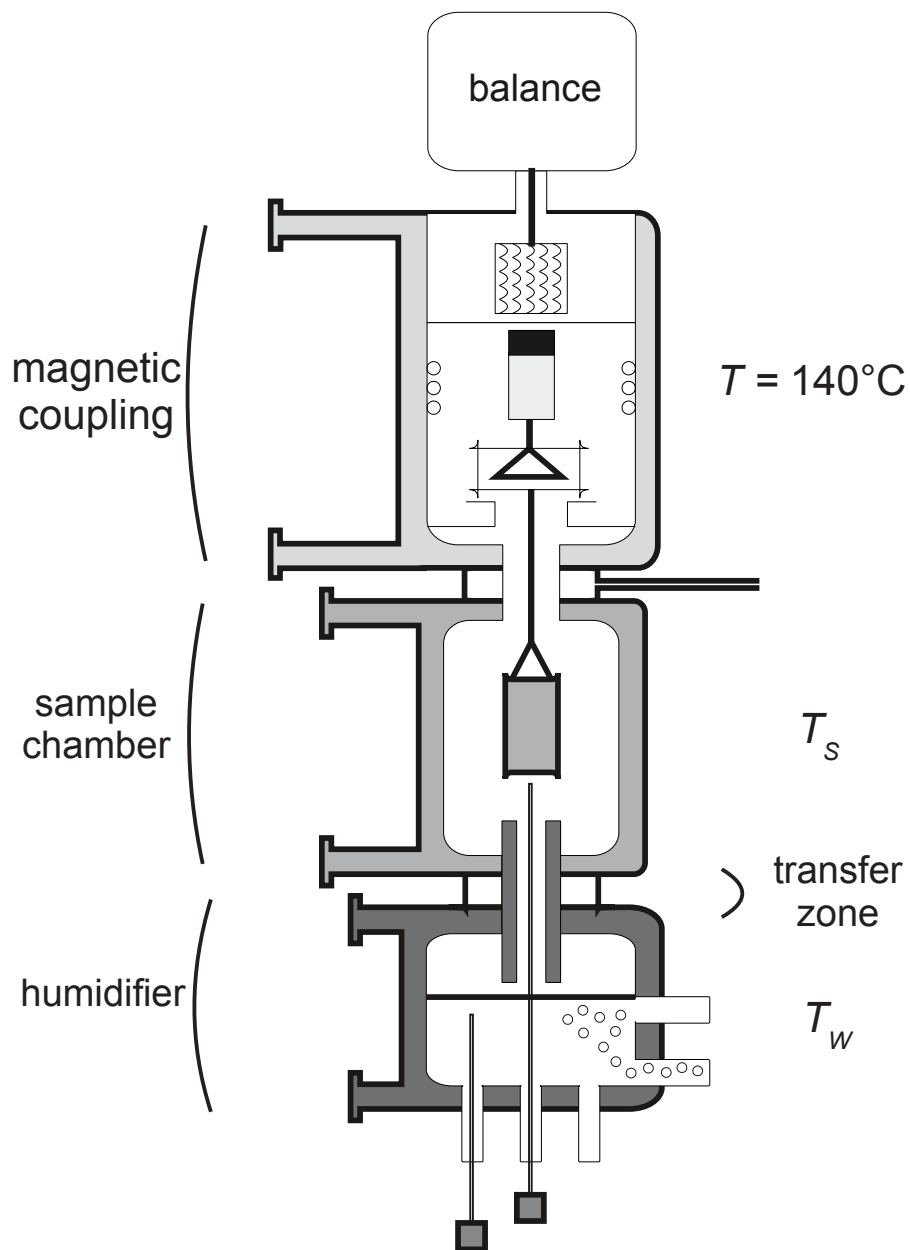


Figure 1: Schematic of inhouse built TGA device (see also [9]) with balance magnetically coupled to a crucible containing the sample in the temperature controlled sample chamber which is humidified through a constant gas stream N_2 bubbling through the temperature controlled humidifier containing water. The compartments are separated by the transferzone to ensure independent temperature control in the compartments. All flanges connecting the different parts are temperature controlled as to prevent condensation of water.

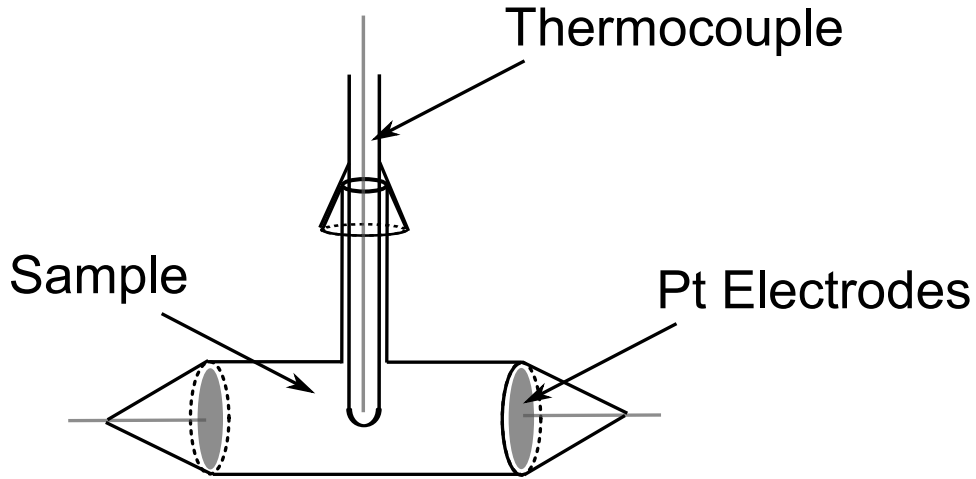


Figure 2: Schematic of the T-shaped impedance cell including a type K thermocouple and Pt electrodes to either side.

Impedance Cell Impedance spectroscopy experiments are conducted in a pseudo-four-point setting in a T-shaped Duran glass cell (figure 2) with two circular platinum electrodes. The sample chamber ($V \sim 3\text{ml}$) is filled through a central tapping socket which is sealed gas tight during the temperature dependent measurement by a glass insert containing a type K thermocouple through which temperature is controlled. Temperatures above $T = 60^\circ\text{C}$ have been set in a Memmert ULE 400 oven and temperatures below $T = 60^\circ\text{C}$ in a Lauda RE207 thermostat.

Equilibrium at low water contents The multitude of condensation and dissociation equilibria in phosphoric acid make extended equilibration times necessary. For neat phosphoric acid complete compositional equilibrium is reached after weeks at its melting point.[12] This estimate has been confirmed by measuring the ^{17}O exchange between H_2O and H_3PO_4 which takes place through condensation reactions. For lower water contents even longer equilibration times can be expected and equilibrium compositions of higher temperatures are “frozen in” by fast cooling of the sample. As already pointed out by Munson in 1964 this might be the reason for the severe difference between compositional data for neat H_3PO_4 obtain by Munson [12] ($\sim 2\%\text{H}_2\text{O}$) and by Huhiti et. al.[3] and Jameson[4] ($\sim 6\%\text{H}_2\text{O}$). In this work compositions have therefore only be analyzed by ^{31}P -NMR at elevated temperatures ($T > 70^\circ\text{C}$). Extrapolation towards lower temperatures reproduces the data of Munson. On the other hand, stable conductivity, diffusion and T_1 relaxation times are reached at much lower equilibration times and despite non-equilibrium compositions.

Calculations including phosphoric acid’s density In calculating $\sigma_{\text{vehicle}}^D$ and $\sigma_{\text{structure}}^D$ temperature dependent density values for different water contents are used. Summaries of physical properties of H_3PO_4 (aq) can be found in the book of Slack [14], the DOE report of Sarangapani et al. [13], or more recently the article by Korte [8]. Of special importance for this study are the empirical equations for density as a function of temperature and water content. MacDonald and Boyack report an empirical equation; (W = weight percentage of H_3PO_4 (aq); T = Temperature in $^\circ\text{C}$; valid in the range 86-102wt% H_3PO_4 (aq); 25-170 $^\circ\text{C}$):

$$\rho = 0.68235 + 1.20811 \cdot 10^{-2}W - \left(1.2379 \cdot 10^{-3} - 3.7938 \cdot 10^{-6}W\right) T \frac{\text{g}}{\text{cm}^3} \quad (3)$$

The data of Egan and Luff [2] and Christensen and Reed [1] (15-100wt% H_3PO_4 (aq); 25-60 $^\circ\text{C}$) have been joined and fitted with the following equation:

$$\rho = 0.959185 + 5.42201 \cdot 10^{-3}W + 2.264 \cdot 10^{-5}W^2 + 1.2172 \cdot 10^{-7}W^3 - (2.33111 \cdot 10^{-4} + 4.75028 \cdot 10^{-6}W)T + 2.43866 \cdot 10^{-6}T^2 \frac{\text{g}}{\text{cm}^3} \quad (4)$$

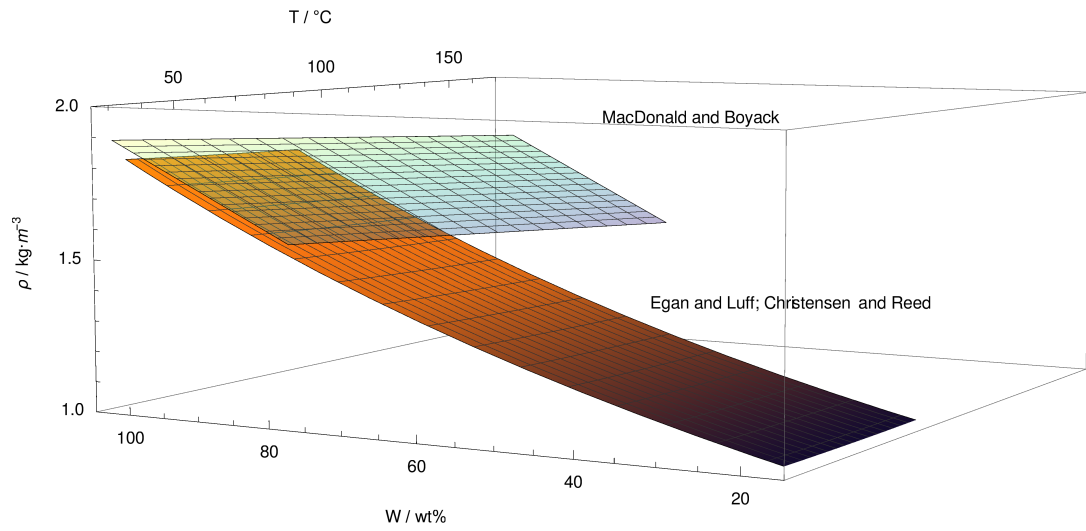


Figure 3: Visualization of empirical density equations by MacDonald and Boyack[11] (blue) and according to data of Egan and Luff [2]; Christensen and Reed [1].

Densities for the mixtures with benzimidazole and imidazole have been calculated through weighted summation of the temperature dependent H_3PO_4 density and (benz)imidazole densities ($\rho_{BI} = 1.23 \text{ g} \cdot \text{cm}^{-3}$; $\rho_{Imi} = 1.03 \text{ g} \cdot \text{cm}^{-3}$). [10]

2 Results

2.1 Composition

As pointed out in the main text frustration of the hydrogen bond network in nominally dry phosphoric acid and the investigated mixtures relaxes through condensation and protonation of the inherent water, or the protonation of the added Bronsted bases imidazole or benzimidazole. Therefore addition of imidazole and benzimidazole actually decreases the degree of condensation of phosphoric acid, i.e., the molar fraction of phosphoric acid condensates di-phosphoric acid, tri-phosphoric acid, etc. (X_{P_2} , X_{P_3} , etc.) and water X_{aq} is reduced upon addition of (benz)imidazole and the molar fraction of ortho-phosphoric acid X_{P_1} is increased. We like to stress here that the molar fraction of ortho-phosphoric acid X_{P_1} is virtually identical in mixtures with the respective same amounts of benzimidazole or imidazole (see figure 16), i.e., the amount of additional proton acceptors determines the molar fraction of ortho-phosphoric acid X_{P_1} . This also holds true for addition of bisbenzimidazole¹ (see figure 4) which contains twice as many additional acceptor sites per molecule as (benz)imidazole and therefore mixtures with 6 mole phosphoric acid per bisbenzimidazole have the same effective molar fraction of ortho-phosphoric acid X_{P_1} than mixtures with 3 mole phosphoric acid per (benz)imidazole (see figure 16).

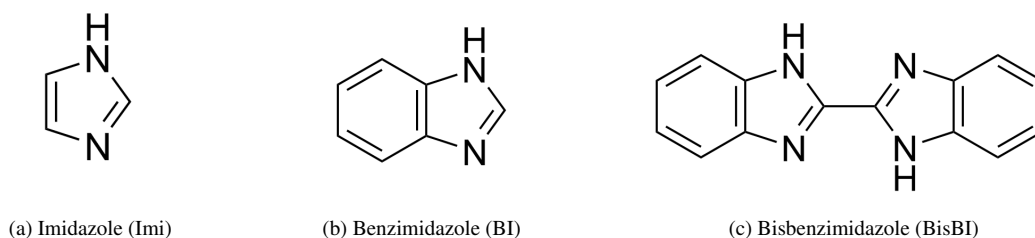


Figure 4: Basic additives used in mixtures with phosphoric acid and their respective abbreviations.

¹We thank Alexander Gullledge (University of South Carolina) for providing the bisbenzimidazole.

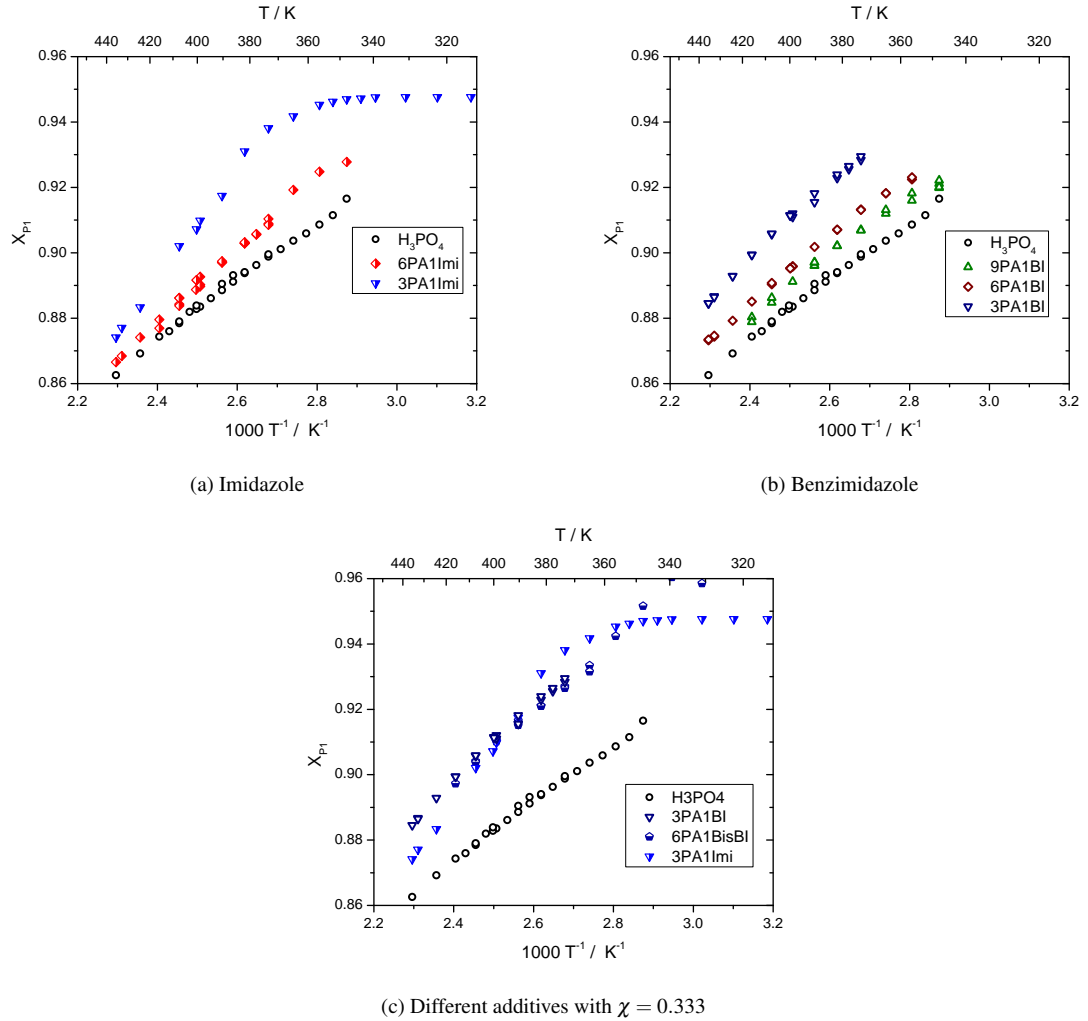


Figure 5: Molar fraction of ortho-phosphoric acid X_{P1} in the PA-part of nominally dry phosphoric acid mixed with a) imidazole and b) benzimidazole at different additive content and c) for different additives with the same ratio of imidazole units to phosphoric acid χ . Addition of basic groups increases X_{P1} in the PA-part and decreases the water content.

2.2 Conductivity

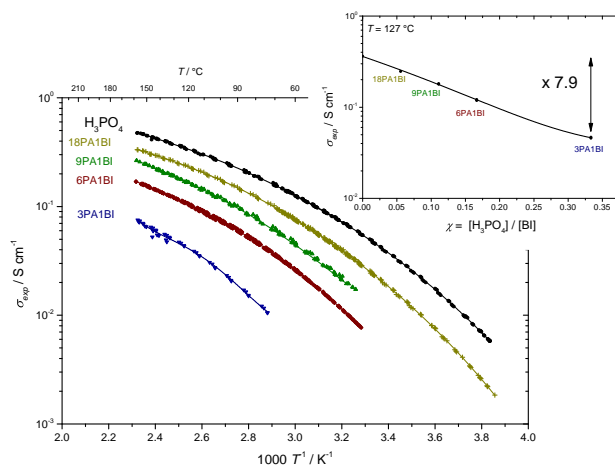


Figure 6: Conductivity measured for different benzimidazole content for nominally dry H_3PO_4 .

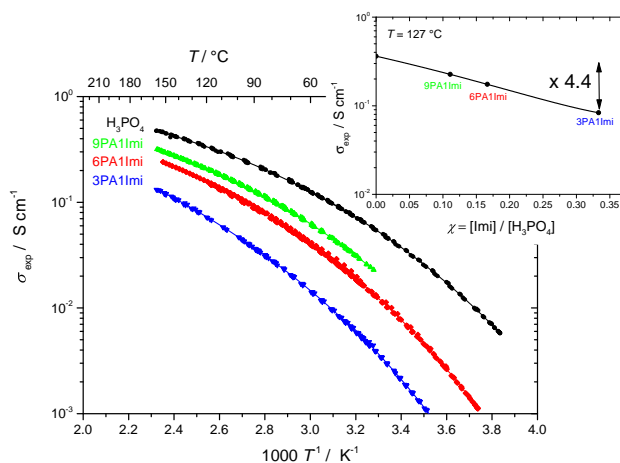


Figure 7: Conductivity measured for different benzimidazole content for nominally dry H_3PO_4 .

2.3 Diffusion

The diffusion coefficients for the different samples nominally dry H_3PO_4 (H_3PO_4), 9 Phosphoric Acid · Benzimidazole (9PA1BI), 6 Phosphoric Acid · Benzimidazole (6PA1BI), 3 Phosphoric Acid · Benzimidazole (3PA1BI), 6 Phosphoric Acid · Imidazole (6PA1Imi), and 3 Phosphoric Acid · Imidazole (3PA1Imi) are shown in the following figures. Proton diffusion coefficients in the mixtures have been obtained through the Kärger equations as described in the main part of the text.

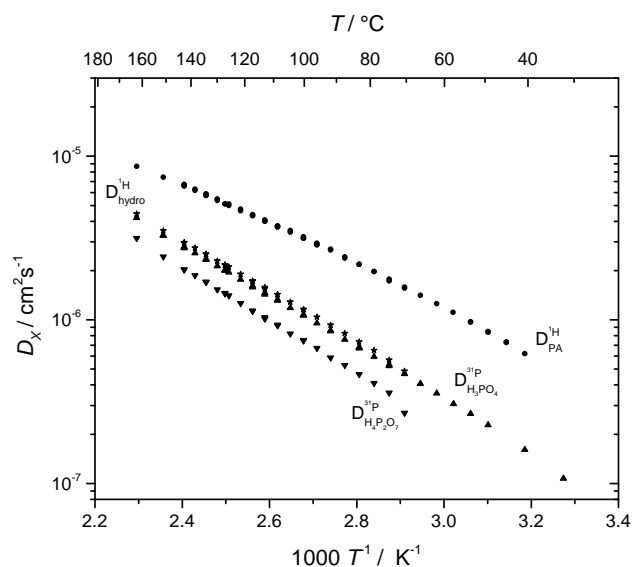


Figure 8: Diffusion coefficients for the different phosphoric acid species and protons for H_3PO_4 .

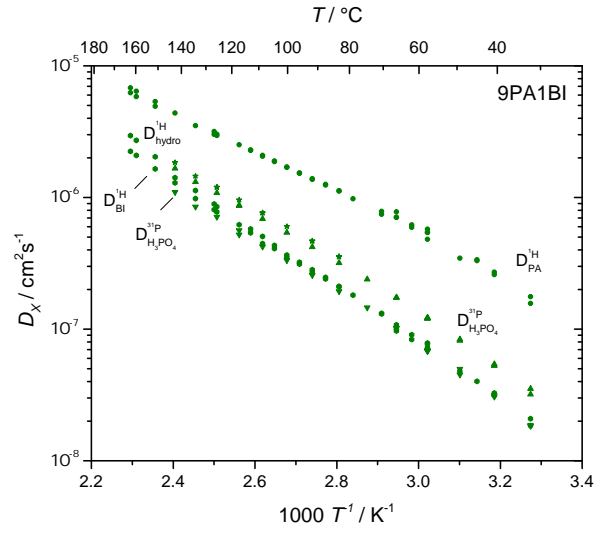


Figure 9: Diffusion coefficients for the different phosphoric acid species, benzimidazole, and protons for 9PA1BI.

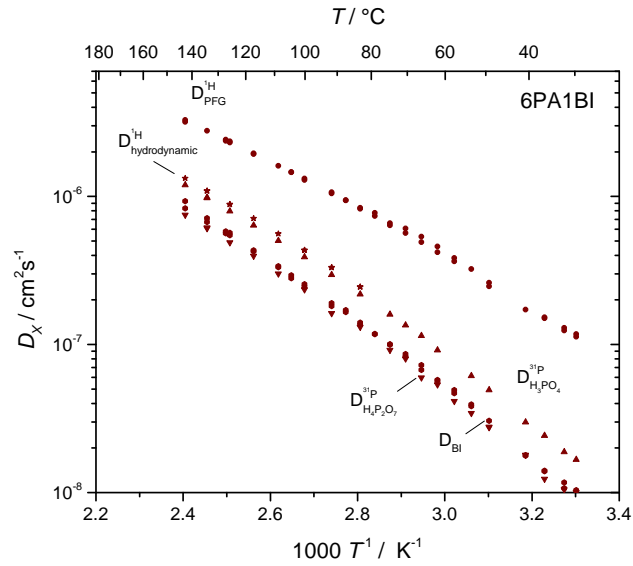


Figure 10: Diffusion coefficients for the different phosphoric acid species, benzimidazole, and protons for 6PA1BI.

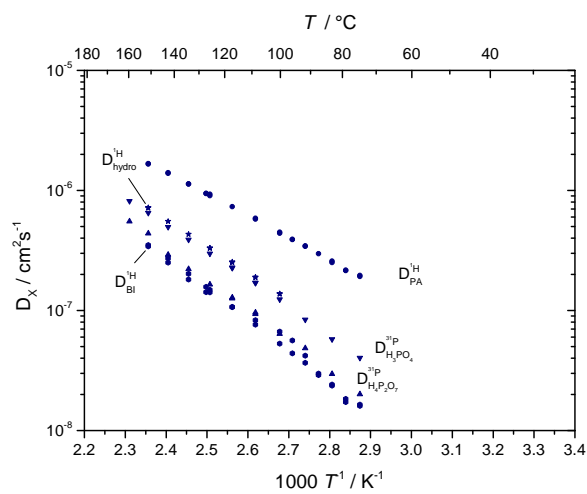


Figure 11: Diffusion coefficients for the different phosphoric acid species, benzimidazole, and protons for 3PA1BI.

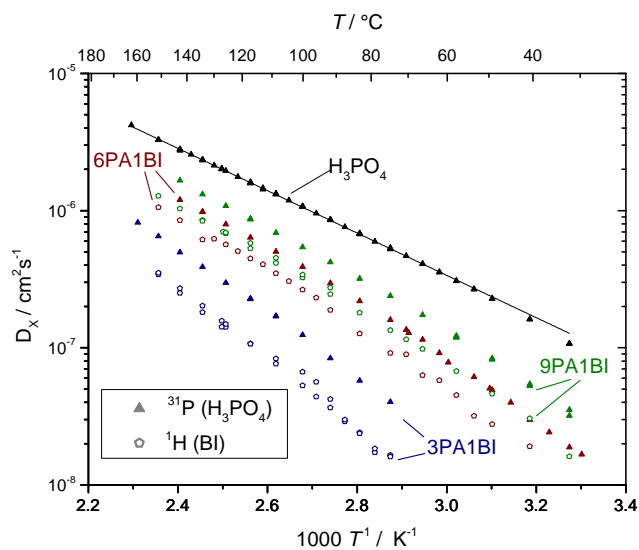


Figure 12: Diffusion coefficients for benzimidazole and orthophosphoric acid for 3PA1BI, 6PA1BI, and 9PA1BI.

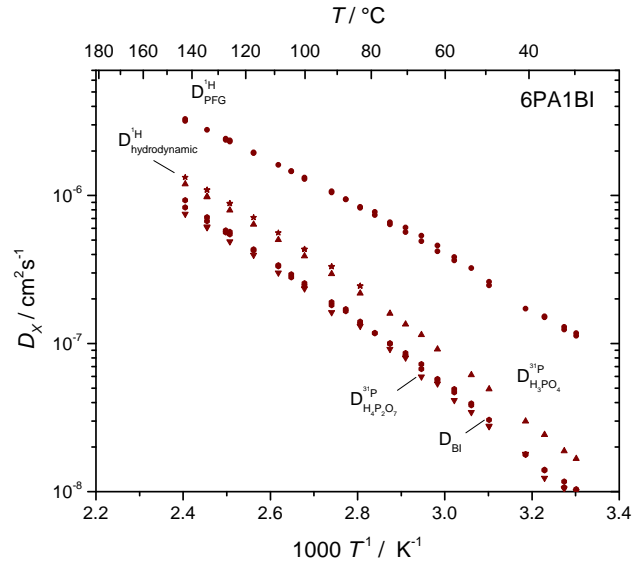


Figure 13: Diffusion coefficients for the different phosphoric acid species, and protons for 6PA1Imi.

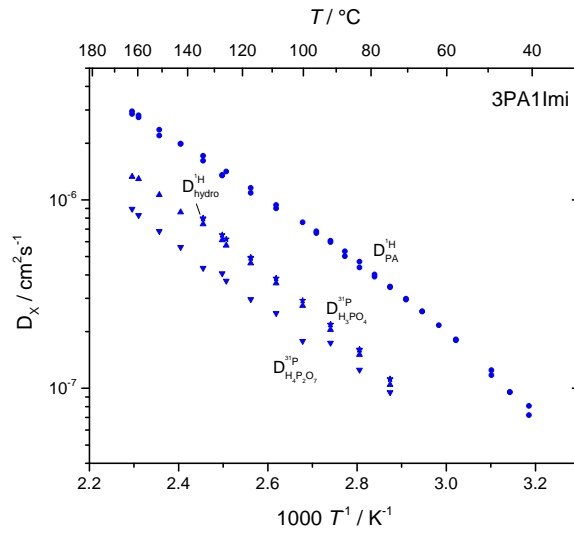


Figure 14: Diffusion coefficients for the different phosphoric acid species, and protons for 3PA1Imi.

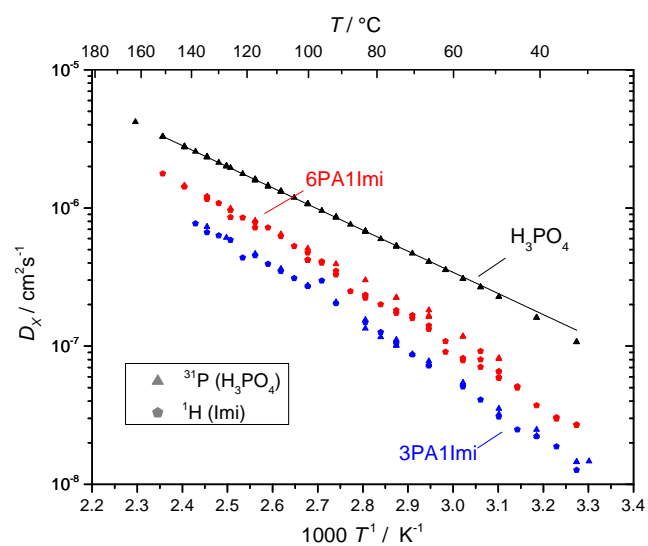


Figure 15: Diffusion coefficients for imidazole and orthophosphoric acid for 3PA1Imi and 6PA1Imi.

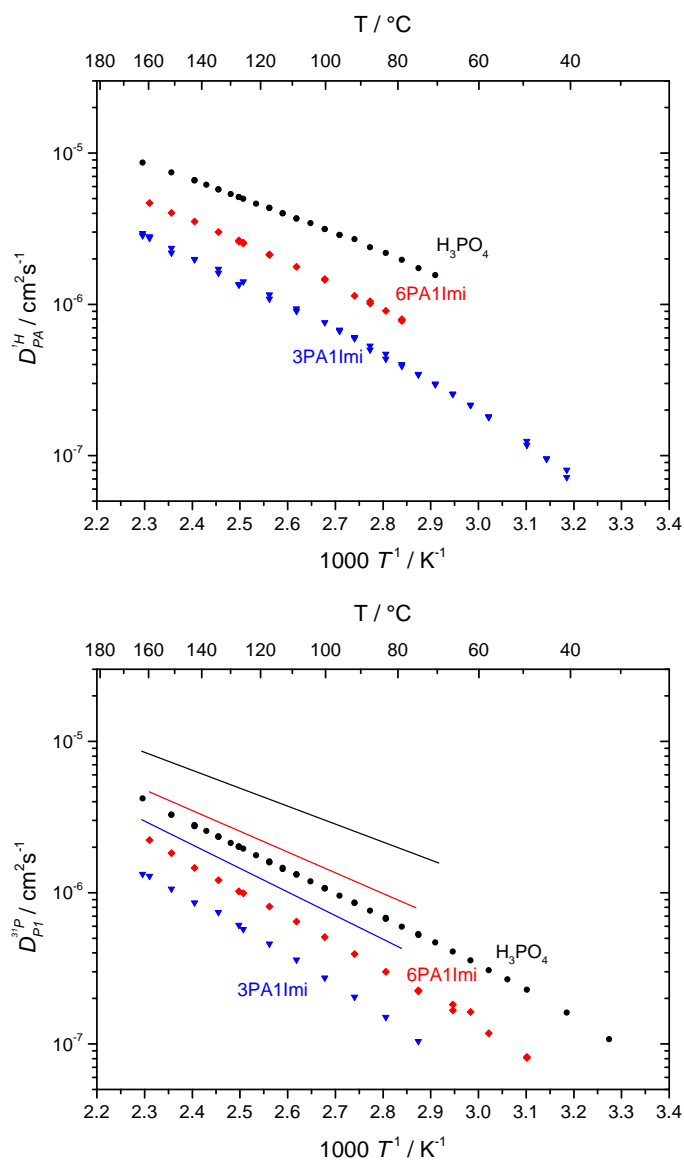


Figure 16: a) Diffusion coefficients for protons in the PA part of PA/Imi mixture D_{PA}^H as obtained from ^1H -PFG NMR and b) diffusion coefficient of orthophosphate species D_{P1}^{31P} for the same mixtures.

3 A Perspective on Exchange Effects in PFG-NMR Diffusion Measurements of PBI-PA membranes

By analysis of coalescing ^1H NMR spectra, the lifetimes for proton exchange between H_3PO_4 and (benz)imidazole have been found to be on the same *millisecond* scale as the diffusion times Δ in ^1H PFG-NMR. Accordingly the intermediate exchange model by Kärger [6, 7] was used to evaluate the decay of the echo intensity (see main text). Such exchange rates have not been reported for PBI-PA membranes before and in previous studies by PFG-NMR [5, 15] the Stejskal-Tanner equation was used to analyze the decay of the echo intensity. The relevant differences between evaluation of PFG-NMR echo attenuation with the Stejskal-Tanner or the Kärger model are briefly discussed.

3.1 Influence of Exchange on the Apparent Diffusion Coefficient in PA-BI Mixtures

On the intermediate exchange timescale the decay of the echo intensity f depends on the diffusion coefficients of both exchange sites D_a and D_b , their populations p_a and p_b , and the exchange life times τ_a and τ_b :

$$\begin{aligned} f &= p'_a e^{-b_i D'_a} + p'_b e^{-b_i D'_b} \\ D'_{a,b} &= C_{1D} \mp C_{2D} \\ C_{1D} &= \frac{1}{2} \cdot \left(D_a + D_b + \frac{1}{\gamma^2 \delta^2 G^2} \left(\frac{1}{\tau_a} + \frac{1}{\tau_b} \right) \right) \\ C_{2D} &= \frac{1}{2} \cdot \sqrt{\left(D_b - D_a + \frac{1}{\gamma^2 \delta^2 G^2} \left(\frac{1}{\tau_b} - \frac{1}{\tau_a} \right) \right)^2 + \frac{1}{\gamma^4 \delta^4 G^4} \frac{4}{\tau_a \tau_b}} \\ p'_b &= \frac{1}{D'_b - D'_a} (p_1 D_a + p_b D_b - D'_a) ; p'_a = 1 - p'_b \end{aligned} \quad (5)$$

The deviations of D_a from the apparent diffusion coefficients D_{app} obtained through the Stejskal-Tanner equation are estimated for the monomeric systems investigated in this work and for PBI-PA membranes in a short numerical simulation. Echo decay data were generated according to equation 5 by MATHEMATICA 10.0 and the Stejskal-Tanner equation was fitted by the least square fitting routines provided by the same program.

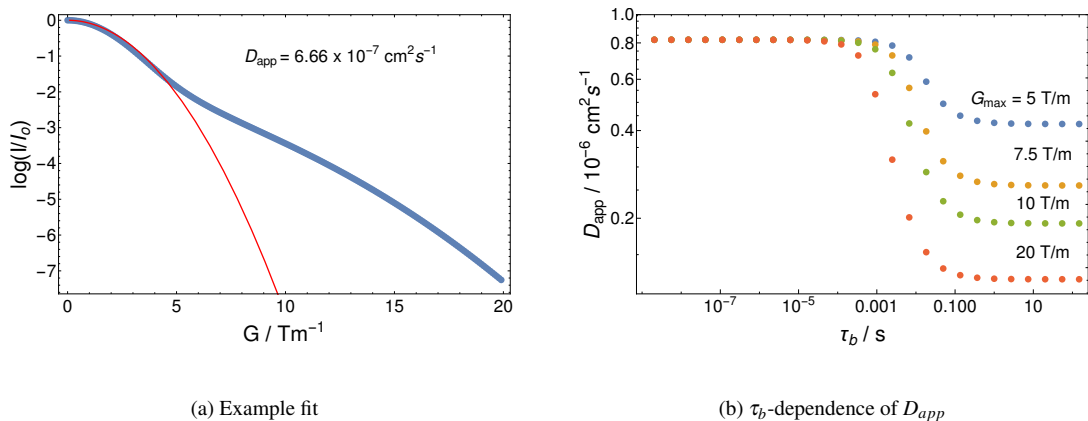


Figure 17: a) The blue line shows data generated according to equation 5 for the model system described in the text with $\tau_b = 0.01 \text{ s}$, $\Delta = 0.04 \text{ s}$. The red line shows a fit of the Stejskal-Tanner equation to the generated data and corresponds to an apparent diffusion coefficient $D_{app} = 6.66 \cdot 10^{-7} \text{ cm}^2/\text{s}$. b) Apparent diffusion coefficients D_{app} obtained as in a) as a function of τ_b with fixed $\Delta = 0.04 \text{ s}$ and different maximum gradient strength G_{max} .

A model system ($D_a = 1 \cdot 10^{-6} \text{ cm}^2/\text{s}$, $D_b = 1 \cdot 10^{-7} \text{ cm}^2/\text{s}$, $p_a = 0.8$, $p_b = 0.2$, $\gamma = 267.513 \cdot 10^6 \text{ s}^{-1}\text{T}^{-1}$ and $\delta = 0.001 \text{ s}$) roughly corresponding to high temperature measurements of 3PA1BI is discussed for constant $\Delta = 0.04 \text{ s}$ and variable τ_b (see figure 17), and for constant $\tau_b = 0.01 \text{ s}$ and variable Δ (see Figure 18). Population differences directly affect the prefactors (p'_a, p'_b) of the exponential terms in equation 5 and the corresponding exponents through detailed balance ($\frac{p'_a}{p'_b} = \frac{\tau_a}{\tau_b}$). These effects are, however, comparably small for $p_a > 0.8$, i.e., for all samples of the present work.

D_{app} decreases with the diffusion time Δ towards a plateau at $\Delta \geq 0.02 \text{ s}$ (see figure 18) with a plateau value depending on the choice of maximum gradients strength. For liquid samples the maximum gradient strength G_{max} is often limited, as above a certain G value the echo intensity is smaller than the signal-to-noise ratio. If, however, the signal-to-noise ratio limits the evaluated gradient range, higher diffusion coefficients (“complete” echo decay at low G) are less affected by the deviations than low diffusion coefficients (“complete” echo decay at high G). This and the direct dependence of D_{app} on τ_b (see figure 17) result in a systematic error of the temperature dependence of D_{app} . For the model system the total deviation of D_{app} can therefore be between 10% (for low exchange rates, low temperatures) and up to an order of magnitude (depending on the maximum gradient strength).

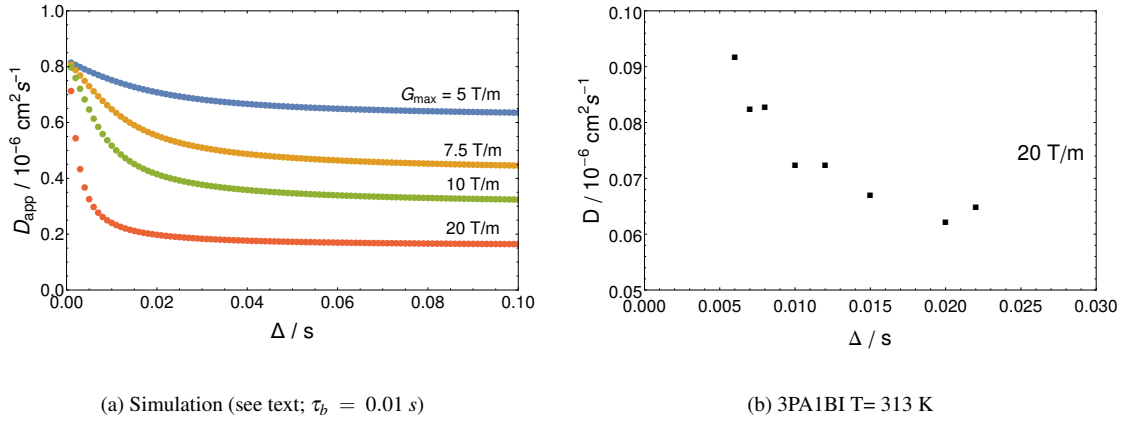


Figure 18: Dependence of D_{app} on Δ in simulation a) and experiment b).

This strong dependence of D_{app} on the experimental settings, exchange rates, population differences and differences of the diffusion coefficients D_a and D_b , results in different deviations for each investigated sample. A comparison of different samples and temperatures with the afforded accuracy of this study on structure diffusion was only possible by applying the intermediate exchange model with exchange rates obtained through lineshape analysis.

3.2 Influence of Exchange on the Apparent Diffusion Coefficient in PBI-PA Membranes

The numerical simulation was repeated for a model membrane with $D_b = 0 \text{ cm}^2/\text{s}$ and $D_a = 10^{-6} \text{ cm}^2/\text{s}$, $p_a = 0.8$, $p_b = 0.2$. Stejskal-Tanner diffusion coefficients in figure 19 exhibit the same qualitative dependence on Δ and τ_b as in the case of the liquid system. The minimum plateau values do, however, not reach D_b as in the liquid system; i.e., in all cases a diffusion coefficient other than zero can be fitted.

Recent literature diffusion data on PBI-PA membranes [5, 15], which include the first attempts to quantify proton diffusion in PBI-PA membranes, suffer from temperature dependent scattering and exhibit plateaus for certain temperature ranges and samples. As discussed in the main text, effects of proton exchange between polybenzimidazole and phosphoric acid, which will occur in the membranes, do not become evident in many cases through visual inspection of the echo decay in a PFG-NMR measurement. This statement is especially true for the membranes for which also the linewidth of protons associated to the polymer are wide. As argued above, the gradient and diffusion time Δ setting can result in artifacts in the temperature dependence of D_{app} which are

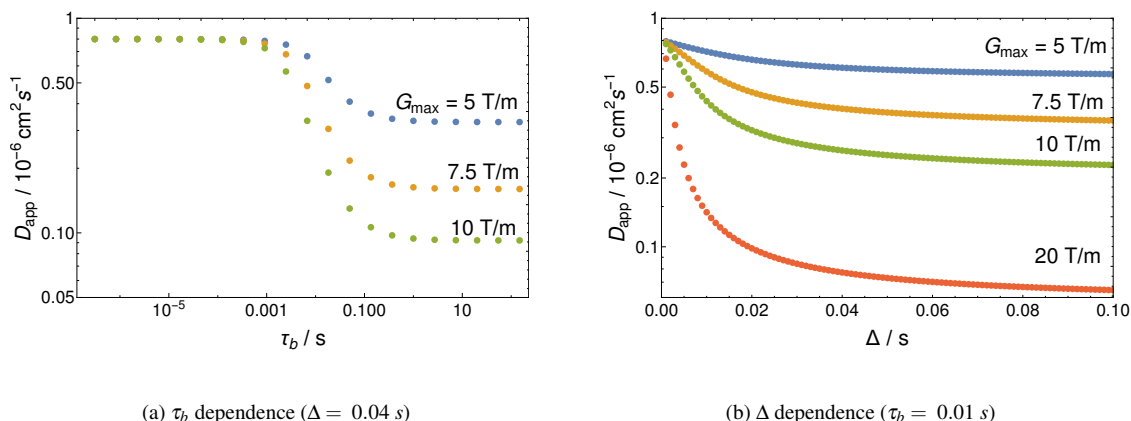


Figure 19: Dependence of D_{app} on τ_b and Δ for a membrane ($D_b = 0 \text{ m}^2/\text{s}$).

also τ_b dependent and can exceed the differences between diffusion coefficients assumed for different membrane polymer structures and phosphoric acid contents. Experimental settings for maximum gradient strength G_{max} , gradient duration δ , or diffusion time Δ are, however, customarily not indicated for individual measurements.

The discussed exchange model holds well for two state exchange, as it will also be found in simple phosphoric acid impregnated polybenzimidazole polymers like AB-PBI (see figure 2). Most of the PBI variations used in fuel cell membranes, however, have a more complex chemical structure than simple AB-PBI and contain benzene, pyridine or other modifications along their polymer repeat unit. Such groups may also act as proton acceptor with different basicity, thereby exchanging protons with phosphoric acid, but not directly with different proton acceptors along the polymer. Proton exchange rates are specific for each exchange partner along the polymer. A detailed description of the echo decay in PFG-NMR measurements of complex PBI-PA membranes will have to include the possibility of more than one exchange rate on the millisecond scale for exchange of protons of phosphoric acid and the polymer.

We feel future NMR studies need to address exchange of protons between phosphoric acid and the polymer, when diffusion coefficients are measured by PFG-NMR. If it is not possible to analyze the proton exchange rates between membrane and phosphoric acid, it might be required to confine diffusion studies to high concentrations of phosphoric acid and to a certain temperature range.

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