Electronic Supplementary Information

Antagonist Effects Leading to Turn-on Electrochemiluminescence in Thermoresponsive Hydrogel Films

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A. Additional figures

0.0 -5.0x10⁻⁶ Current / A -1.0x10⁻⁵ -1.5x10⁻⁵ -2.0x10⁻⁵ -2.5x10⁻⁵ -3.0x10⁻⁵ -0.8 -0.7 -0.6 -0.9 -0.5 -0.4 -0.3 Potential / V vs Ag/AgCl

Figure S1. Electrodeposition of a pNIPAM-Ru hydrogel film by cyclic voltammetry. Electrochemically assisted free radical polymerization was performed in a degassed solution containing 70 mM pNIPAM, 0.14 mM Ru(bpy)₃ monomer, 1.5 mM K₂S₂O₈, 1.75 mM BIS and

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 0.2 M KNO_3 . The potential was scanned 60 times at a scan rate of 100 mV s⁻¹ between -0.35 and -0.9 V *vs*. Ag/AgCl. The arrow indicates the increasing scan number.



Figure S2. Scanning electron microscopy image of lyophilized hydrogel films on the GC electrode.



Figure S3. Voltammetric and ECL signals of a pNIPAM-modified GC electrodes recorded in a 100 mM PBS (pH7.4) solution containing 0.5 mM $Ru(bpy)_3^{2+}$ and 100 mM TPA at 25°C (red curve) and at 40°C (blue curve). The film were prepared as in Figure S1 but <u>without</u> $Ru(bpy)_3$ monomer in the electrodeposition solution.



Figure S4. ECL spectra of the pNIPAM-Ru hydrogel film in the swollen state at 25°C (blue curve) and in the collapsed state at 40°C (red curve). Comparison with the photoluminescent spectrum (black curve) of the 10 μ M Ru(bpy)₃²⁺ complex in aqueous solution at 25°C. ECL spectra were acquired in 100 mM PBS solution (pH 7.4) containing 100 mM TPA during cyclic voltammetry experiments. The potential was scanned from 0.2 V to 1.45 V at a scan rate of 50 mV s¹.



Figure S5. Voltammetric (a) and ECL signal (b) of a GC electrode modified with a pNIPAM-Ru film in a 100 mM PBS solution containing 20 mM DBAE at 25°C (blue curve) and at 40°C (red curve). Scan rate: 100 mV s⁻¹.

| Coreactant ^(a) | Faradaic charge at 25°C <i>Q</i> ^{25°C} (μC) | Faradaic charge at 40°C <i>Q^{4θ°C}</i> (μC) | ECL signal at 25°C ^{I^{25°C} (10³ a.u.)} | ECL signal at 40°C ^{I^{40°C}_{ECL} (10³ a.u.)} | Faradaic charge ratio Q ^{40°C} / Q ^{25°C} | ECL signal ratio I ^{40°C} / I ^{25°C} |
|---------------------------|---|--|---|---|---|--|
| ТРА | 46±3 | 14.3±13 | 5±0.4 | 188±53 | 0.31±0.28 | 38±12 |
| DBAE | 39±2 | 13.7±3 | 1.7 ± 0.07 | 67±3 | 0.31±0.1 | 40±1 |
| Oxalate | 13.7±0.4 | 17.5±0.7 | 2.68 ± 0.07 | 155±4 | 1.28±0.06 | 58±3 |

Table S1. Variations of the faradaic charge ratio and of the ECL signal ratio for different coreactants between 25°C (swollen state) and 40°C (collapsed state).

B. Experimental section

Chemicals.

All the reagents were purchased from Sigma-Aldrich unless otherwise noted. *N*-isopropylacrylamide (NIPAM) was recrystallized from hexane (ICS) and dried under vacuum prior to use. The cross-linker *N*,*N*'-methylenebis(acrylamide) (BIS) and the initiator potassium persulfate were used as received. Ruthenium(II) (4-vinyl-4'-methyl-2,2'-bipyridine)bis(2,2'-bipyridine)bis(hexafluorophosphate) [Ru(bpy) monomer)] was synthesized according to the procedure described by Spiro et al.¹ The coreactants, *n*-tripropylamine (TPA), dibutylaminoethanol (DBAE) and sodium oxalate, were purchased from Sigma Aldrich. De-ionized water, obtained with a Milli-Q system, was used for all synthesis reactions, purification and solution preparation.

Preparation of electrodes modified with pNIPAM-Ru films

Glassy carbon (GC) electrodes were polished with wet alumina powder on a polishing cloth, and then washed ultrasonically with ethanol and water, respectively. In a typical process, the electrode was immersed in an aqueous solution containing 70 mM NIPAM, 1.75 mM BIS (2.5 mol % against NIPAM), 0.14 mM of Ru(bpy) monomer (0.2 mol % of Ru monomer against NIPAM), and 1.5 mM K₂S₂O₈ in 0.2 M KNO₃ solution. Prior to electrochemically assisted polymerization, the reaction solution was deoxygenated by bubbling nitrogen gas for 30 min at room temperature. The potential was applied by cycling between -0.35 and -1.35 V at 100 mV s⁻¹ for 60 cycles. After the polymerization, the electrodes modified with pNIPAM-Ru films were thoroughly washed with double-distilled water to remove the monomers and stored in ultrapure water.

Characterization of the hydrogel film

Scanning electron microscopy (SEM)

A flat glassy carbon plate was used as working electrode and modified using the same procedure as described on the macroscopic electrode. After rinsing with distilled water, it was immersed in a water bath and frozen rapidly in liquid nitrogen. After freeze drying, the electrode was sputtered with gold using a sputter-coater (Emitech K550X) and observed in SEM (Hitachi TM3000).

Electrochemical measurements

Differential pulse voltammetry (DPV) experiments were performed with a μ -Autolab Type III and PGSTAT30 electrochemical stations. The three-electrode system consisted in a glassy carbon (GC) electrode as a working electrode, Ag wire as a pseudo-reference and a platinum wire as a counter-electrode. The temperature of the sample was controlled using a thermostatically controlled cell compartment. The measurements were taken under equilibrium conditions after holding the sample for 15 min at the required temperature.

Electrochemiluminescence. ECL experiments were performed with a μ -Autolab Type III electrochemical station. ECL intensity was measured by using a Hamamatsu photomultiplier tube R4632. The signal was amplified by a Keithley Picoammeter before acquisition *via* the second input channel of the μ -Autolab. ECL spectra were recorded using a Princeton Instrument spectrograph Spectra pro 2300i under potential scanning between 0.2 and 1.45 V at a scan rate of 50 mV/s, in an aqueous solution containing the coreactant and the supporting electrolyte. The temperature of the sample was controlled using a thermostatically controlled cell compartment. The measurements were taken under equilibrium conditions after holding the sample for 15 min at each temperature.

C. References

1 P. K. Ghosh and T. G. Spiro, J. Am. Chem. Soc., 1980, **102**, 5543-5549.