

# Repurposing designed mutants: a valuable strategy for computer-aided laccases engineering. The case of POXA1b.

Valerio Guido Giacobelli<sup>a</sup>#, Emanuele Monza<sup>b</sup>#, M. Fatima Lucas<sup>b</sup>, Cinzia Pezzella<sup>a</sup>, Alessandra Piscitelli<sup>a</sup>, Victor Guallar<sup>b,c</sup>\*, Giovanni Sannia<sup>a</sup>

<sup>a</sup>Department of Chemical Sciences, University of Naples Federico II, Via Cintia 4, 80126 Naples, Italy

<sup>b</sup>Joint BSC-CRG-IRB Research Program in Computational Biology, Barcelona Supercomputing Center, c/Jordi Girona 29, 08034 Barcelona, Spain

<sup>c</sup>Institució Catalana de Recerca i Estudis Avançats (ICREA), Passeig Lluís Companys 23, 08010 Barcelona, Spain

#Equally contributed to this work

\*Corresponding authors: [victor.guallar@bsc.es](mailto:victor.guallar@bsc.es), [sannia@unina.it](mailto:sannia@unina.it).

## Information presented here includes:

**Table S1.** Variants from round 1 showing improved descriptor values.

**Table S2.** Variants from round 2 of the V162H branch showing improved descriptor values.

**Table S3.** Variants from round 2 of the V162S branch showing improved descriptor values.

**Table S4.** Variants from round 3 of the V162H branch showing improved descriptor values.

**Table S5.** Variants from round 3 of the V162S branch showing improved descriptor values.

**Table S6.** Computational ionization energy of 2,4- and 2,5- dabsa.

**Figure S1.** PELE sampling result for the V162H branch triple mutant.

**Figure S2.** Sequence alignment of POXA1b with its homology model template.

**Figure S3.** PELE binding energy vs. 2,4-dabsa-Cu distance.

**Figure S4.** PELE binding energy vs. 2,5-dabsa-Cu distance.

## Examples of PELE input:

- Sampling
- Design

**Table S1.** Variants from round 1 showing improved descriptor values.

Protein variant	Binding energy (kcal/mol)	SASA
Wild type	-8	0.19
A336R	-22	0.10
A336Q	-13	0.17
A336K	-15	0.14
G161R	-16	0.12
G161Q	-20	0.11
G161K	-16	0.11
G161M	-15	0.09
G161S	-19	0.13
G161Y	-14	0.10
G392R	-18	0.10
G392F	-14	0.04
G392W	-15	0.02
G392Y	-13	0.04
L511R	-18	0.13
L511N	-18	0.18
L511Q	-17	0.17
L511H	-14	0.16
L511K	-14	0.15
L511T	-17	0.17
F331R	-16	0.12
F331Q	-15	0.17
F331K	-15	0.17
F331Y	-19	0.15
P163R	-17	0.18
P163N	-20	0.19
P163Q	-16	0.14
P163K	-14	0.13
V162N	-20	0.10
V162C	-18	0.09

<b>V162H</b>	<b>-20</b>	<b>0.09</b>
<b>V162S</b>	<b>-16</b>	<b>0.17</b>
V162T	-18	0.21

**Table S2.** Variants from round 2 of the V162H branch showing improved descriptor values.

Protein variant	Binding energy (kcal/mol)	SASA
V162H_A336N	-28	0.09
V162H_A336K	-26	0.10
V162H_L511R	-28	0.08
V162H_L511Q	-27	0.10
<b>V162H_F331Y</b>	<b>-29</b>	<b>0.09</b>

**Table S3.** Variants from round 2 of the V162S branch showing improved descriptor values.

Protein variant	Binding energy (kcal/mol)	SASA
V162S_A336R	-22	0.16
V162S_A336Q	-21	0.16
V162S_A336K	-22	0.17
V162S_G161R	-22	0.10
V162S_G392R	-22	0.05
V162S_G392Q	-22	0.12
V162S_G392F	-23	0.03
V162S_G392W	-23	0.03
V162S_G392Y	-23	0.03
V162S_L511R	-25	0.14
V162S_L511N	-25	0.20
V162S_L511Q	-24	0.17
V162S_L511T	-24	0.18
V162S_F331R	-23	0.13
V162S_F331K	-23	0.16
<b>V162S_F331Y</b>	<b>-26</b>	<b>0.14</b>
V162S_P163R	-24	0.14
V162S_P163N	-28	0.18
V162S_P163K	-23	0.14
V162S_P163M	-21	0.17

**Table S4.** Variants from round 3 of the V162H branch showing improved descriptor values.

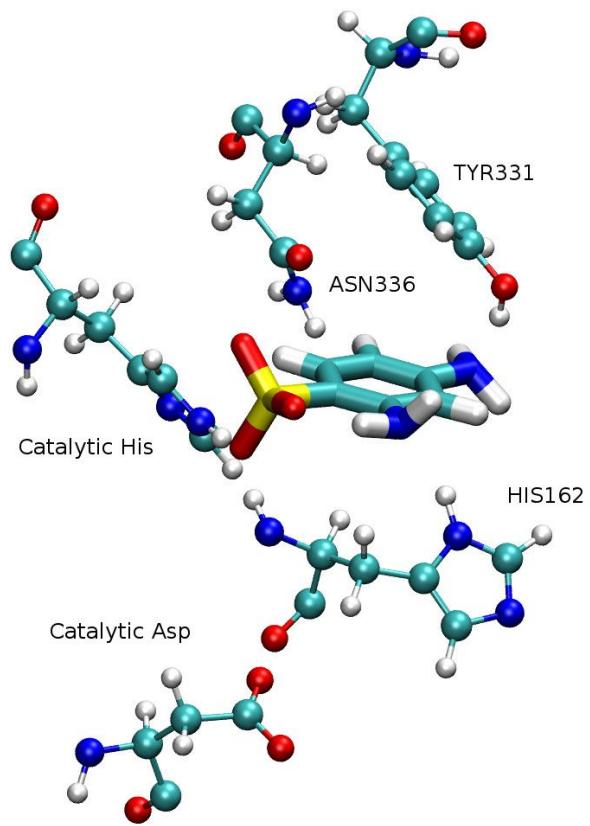
Protein variant	Binding energy (kcal/mol)	SASA
V162H_F331Y_A336R	-36	0.08
<b>V162H_F331Y_A336N</b>	<b>-35</b>	<b>0.07</b>
V162H_F331Y_G392R	-35	0.10
V162H_F331Y_L511R	-35	0.08
V162H_F331Y_L511N	-36	0.10
V162H_F331Y_P163K	-35	0.03
V162H_F331Y_P510R	-35	0.08

**Table S5.** Variants from round 3 of the V162S branch showing improved descriptor values.

Protein variant	Binding energy (kcal/mol)	SASA
<b>V162S_F331Y_A336N</b>	<b>-31</b>	<b>0.15</b>
V162S_F331Y_G161K	-34	0.15
V162S_F331Y_P163R	-33	0.10
V162S_F331Y_P163N	-35	0.13
V162S_F331Y_P163Q	-32	0.13
V162S_F331Y_P163H	-32	0.10
V162S_F331Y_P163K	-33	0.09

**Table S6.** Computational ionization energy of 2,4- and 2,5- dabsa.

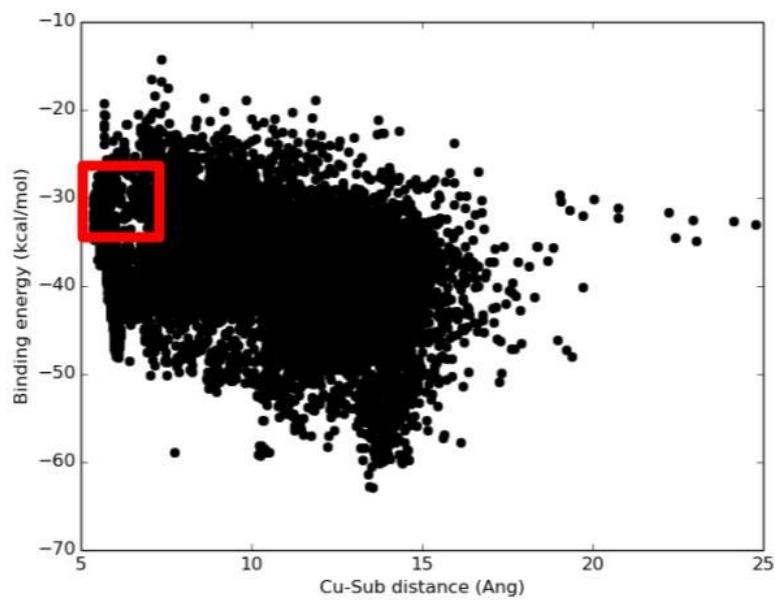
Chemical species	Ionization energy (eV)
2,4-dabsa	4.44008397
2,5-dabsa	4.26511468



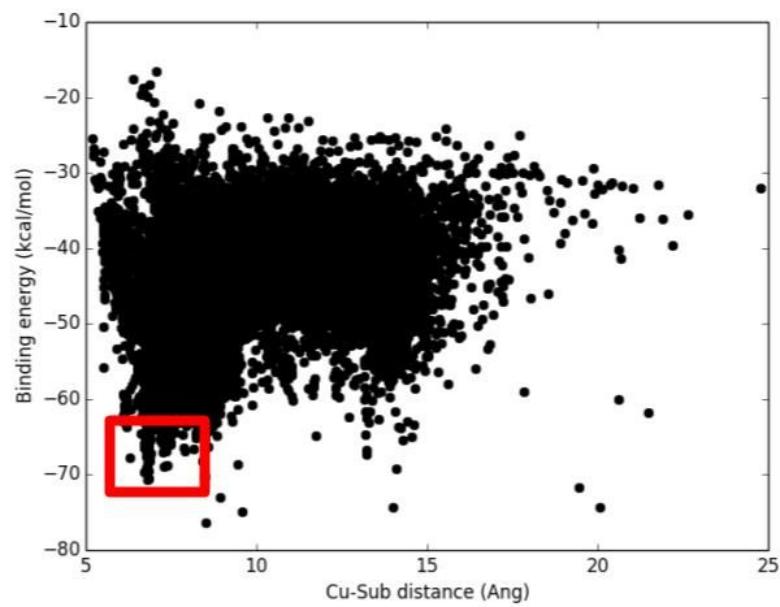
**Figure S1.** PELE sampling result for the V162H branch triple mutant.

Target	SIGPGRGTLNIAHKVIQEDGFYRSTVLAGGSYPGPLIRGKTGDRFQIHVVNHKLADTSNPVD	60
1gyc.1.A	DGPESALVANAPCPDGFERDAIIVNGEPIPLITGEIGDRFQIHVVVIDTLTHTMILK	60
Target	TSIHNRGLFVKGHHWADGPAMVTQCPIVPGHSLFLYDFWVPDFQAGTFWYHSHLGTQYCDGL	120
1gyc.1.A	TSIHNRGPFQAGTHWADGPAPVHQCPIAAGHSLFLYDFWPDFQAGTFWYHSHLSTQYCDGL	120
Target	RGPLVVYSENDPHKRLYDWDDESTVLTVDWYHAPSLSLTGVPH-PDSTLFWHGLGRSLNG	179
1gyc.1.A	RGPEVVYSEKDPHESLYDWDDESTVLTIDQYHSTAPLGPFPPIGRD-TLNGLGRS-LT	180
Target	PASPLYVHNVVEGKRYTRIRLILHTSCDSHYQFSIDGHTETVIRADGEHTQLQVDOVQIF	239
1gyc.1.A	PTAAALAVIHNQHGERYTRERLISCDPHVYTESIDGHLITVIREOGIH-QPLVQDLOQIF	240
Target	GQRYSLVLHANQAVGHYGIEAHPHSGDPGFENQHMSAILEYEGARSIDPTTPEQHATHPL	299
1gyc.1.A	GQRYSLVLAHQIVGNYWIFAHPHSGT/GFACGCCISATELFYQAFIARPTTIGT-TVPL	300
Target	HRYHMLRPLIKKPAPGKFPGGADHNLLHLHEZEDPATLFTAHNHTFVPPTVPVVLLQILRG	359
1gyc.1.A	IETIHLHPKLRHPVPGSPIPGGIDALILQAFEPY-EIHFEDHCFPPPTVPPVLLQILRG	358
Target	TRDAHDLAPAGSIYDILKLGDVVRITMPA--LVEAGPHPIHLHGHTFAVVRSAAGSSTYHYE	417
1gyc.1.A	EQTAQDLIPEASLYEYIANGTLEITLPAIALAPGAPHEPHIHLGH-FAVVESAGSSTYHKH	418
Target	IPVERDVVSIGDDE---TDNVTIREVADHAGPWFELHCHIDWHLIDLGFIAVVFAEVGVNQTAA	474
1gyc.1.A	IPVERDVVSIGDDE---TPASGDUVTIRETDMPGPWFELHCHIDWHLIDLGFIAEVFAEVGVNQTAA	476
Target	AHPVPEAWHULCPIYHSSNPSKLLMGTNAIGRLPAPLKA	513
1gyc.1.A	AHPVPEAWHULCPIYDELSEALQ-----	499

**Figure S2.** Sequence alignment of POXA1b with its homology model template.



**Figure S3.** PELE binding energy vs. 2,4-dabsa-Cu distance. Reactive enzyme-substrate conformations are localized in the red rectangle.



**Figure S4.** PELE binding energy vs. 2,5-dabsa-Cu distance. Reactive enzyme-substrate conformations are localized in the red rectangle.

# PELE sampling input file

```
## POXA1b-2,4-dabsa sampling

file datadir /home/bsc72/bsc72328/plop/data
file log dyn_24dabsa.log
energy params solvent vdgbnp
energy params ionic 0.15
load pdb poxa1b-24dabsa_fixed.pdb waters no ions no het yes

constraint atom A:597:CU__ current 200 0.0 &
atom A:451:_SG_ current 200 0.0 &
atom A:451:_CB_ current 200 0.0 &
atom A:456:_ND1 current 200 0.0 &
atom A:456:_CB_ current 200 0.0 &
atom A:456:_CG_ current 200 0.0 &
atom A:394:_ND1 current 200 0.0 &
atom A:394:_CB_ current 200 0.0 &
atom A:394:_CG_ current 200 0.0 &
atom A:453:_CD1 current 200 0.0 &
atom A:453:_CG1 current 200 0.0 &
atom A:461:_CE1 current 200 0.0 &
atom A:461:_CD1 current 200 0.0 &
atom A:461:_CG_ current 200 0.0 &
atom A:598:CU__ current 200 0.0 &
atom A:599:CU__ current 200 0.0 &
atom A:600:CU__ current 200 0.0 &
atom A:399:_NE2 current 200 0.0 &
atom A:111:_NE2 current 200 0.0 &
atom A:450:_NE2 current 200 0.0 &
atom A:452:_NE2 current 200 0.0 &
atom A:109:_NE2 current 200 0.0 &
atom A:66:_ND1 current 200 0.0 &
atom A:397:_NE2 current 200 0.0 &
atom A:64:_NE2 current 200 0.0 &
```

```
pele &
het L:1 &
pdbmodel yes &
top_side 10 &
init_min no &
task &
    show bind_ene 1 &
    show SASA 1 &
    if random 1 gt 0.5 then rot_r 0.02 else rot_r 0.25 endif &
    if random 2 gt 0.5 then tra_r 2.0 else tra_r 0.75 endif &
    spawn atom 1 A:597:CU__ within 15.0 &
    exit steps gt 2000 &
end_task &
temp 1000 &
tries 1 &
anmfreq 1 &
spfreq 1 &
mifreq 1 &
wrfreq 1 &
omit_sp A:394 A:394 &
omit_sp A:451 A:451 &
omit_sp A:456 A:456 &
omit_sp A:453 A:453 &
omit_sp A:461 A:461 &
side &
    randomize no &
    verbose no &
    failsafe no &
sideend &
path traj_24dabsa_ &
mirad 20 &
sprad 10 &
min &
    rmsg 0.02 &
    nbup yes &
```

```
gbup yes &
alphaup yes &
minimend &
caconst 0.05 &
lcom_con 0.1 &
rem_bulk_mov 3 &
anm_eig_freq 100000 &
anm_altm_freq 1 &
anm_altm_type 3 &
lanmanm neig 6 &
lanmanm ualig YES &
lanmanm move_ca 0.2 &
lanmanm mix_modes 0.75 &
lanmin &
mxitn 100 &
iter 1 &
rmsg 0.02 &
nbup yes &
alphaup no &
minimend
```

# PELE design input file

```
## POXA1b-2,4-dabsa design

file datadir /home/bsc72/bsc72328/plop/data
file log dyn.log
energy params solvent vdgbnp
energy params ionic 0.15
load pdb candidate_refined.pdb waters no ions no het yes

constraint atom A:597:CU__ current 200 0.0 &
atom A:451:_SG_ current 200 0.0 &
atom A:451:_CB_ current 100 0.0 &
atom A:456:_ND1 current 200 0.0 &
atom A:456:_CB_ current 100 0.0 &
atom A:456:_CG_ current 100 0.0 &
atom A:394:_ND1 current 200 0.0 &
atom A:394:_CB_ current 100 0.0 &
atom A:394:_CG_ current 100 0.0 &
atom A:453:_CD1 current 100 0.0 &
atom A:453:_CG1 current 100 0.0 &
atom A:461:_CE1 current 100 0.0 &
atom A:461:_CD1 current 100 0.0 &
atom A:461:_CG_ current 100 0.0 &
atom A:598:CU__ current 200 0.0 &
atom A:599:CU__ current 200 0.0 &
atom A:600:CU__ current 200 0.0 &
atom A:399:_NE2 current 100 0.0 &
atom A:111:_NE2 current 100 0.0 &
atom A:450:_NE2 current 100 0.0 &
atom A:452:_NE2 current 100 0.0 &
atom A:109:_NE2 current 100 0.0 &
atom A:66:_ND1 current 100 0.0 &
atom A:397:_NE2 current 100 0.0 &
```

```
atom A:64:_NE2 current 100 0.0 &

pele &
    het L:1 &
    pdbmodel yes &
    top_side 10 &
    init_min no &
    task &
        show bind_ene 1 &
        show SASA 1 &
        spawn atom 1 A:597:CU__ within 15.0 &
        exit steps gt 2000 &
    end_task &
    rot_r 0.02 &
    tra_r 0.25 &
    temp 1000 &
    tries 1 &
    anmfreq 1 &
    spfreq 1 &
    mifreq 1 &
    wrfreq 1 &
    omit_sp A:394 A:394 &
    omit_sp A:451 A:451 &
    omit_sp A:456 A:456 &
    omit_sp A:453 A:453 &
    omit_sp A:461 A:461 &
    side &
        randomize no &
        verbose no &
        failsafe no &
    sideend &
    path traj_24dabsa_ &
    mirad 20 &
    sprad 10 &
    min &
    rmsg 0.02 &
```

```
nbup yes &
gbup yes &
alphaup yes &
minimend &
caconst 0.05 &
lcom_con 0.1 &
rem_bulk_mov 3 &
anm_eig_freq 100000 &
anm_altm_freq 1 &
anm_altm_type 3 &
lanmanm neig 6 &
lanmanm ualig YES &
lanmanm move_ca 0.5 &
lanmanm mix_modes 0.75 &
lanmanm omit_sp A:597 A:600 &
lanmin &
mxitn 100 &
iter 1 &
rmsg 0.02 &
nbup yes &
alphaup no &
minimend
```