Lanthanum Complexes Containing a Bis(phenolate) Ligand with a Ferrocenediyl-1,1'-dithio Backbone: Synthesis, Characterization, and Ringopening Polymerization of *rac*-Lactide

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9. References

1. NMR Spectra of Complex [(L)LaN(SiMe₃)₂] (1)



Fig. S1 1 H NMR spectrum (C₆D₆, 400 MHz) of complex [(L)LaN(SiMe₃)₂] (1).



Fig. S2 $^{13}\text{C}\left\{^{1}\text{H}\right\}$ NMR spectrum (C₆D₆, 101 MHz) of complex 1.



Fig. S3 $^{29}\text{Si}\ \{^1\text{H}\}$ NMR spectrum (C6D6, 79.5 MHz) of complex 1.



Fig. S5 1 H- 13 C Heteronuclear single quantum correlation (HSQC) experiment of complex 1 (in C₆D₆).

2. NMR Spectra of Complex [(L)LaOⁱPr] (2)



Fig.S8 Homonuclear correlation spectroscopy (COSY) of complex 2 (in C_6D_{6r} , containing toluene residue).



Fig. S9 ¹H-¹³C Heteronuclear single quantum correlation (HSQC) experiment of complex 2 (in C₆D₆, containing toluene residue).

3. NMR Spectra of Complex [(L)LaBH₄] (3)



Fig. S10¹H NMR spectrum (C₆D₆, 400 MHz) of complex [(L)LaBH₄] (3).



Fig. S11 $^{13}C{^{1}H}$ NMR spectrum (C₆D₆, 101 MHz) of complex 3.



Fig. S12 ^{11}B { ^{1}H } NMR spectrum (C₆D₆, 128.4 MHz) of complex 3.



Fig. S13 Homonuclear correlation spectroscopy (COSY) of complex 3 (in C₆D₆, containing toluene residue).



Fig. S14 1 H- 13 C Heteronuclear single quantum correlation (HSQC) experiment of complex 3 (in C₆D₆, containing toluene residue).

4. Polymerization Kinetics



Fig. S15 First-order kinetic plot for the polymerization of *rac*-LA in THF at 25 °C with complex **1** as initiator (THF = 6 mL, $[M]_0$ = 0.5 M, [I] = 0.833 mM, $k_{app} = 0.032 \text{ min}^{-1}$).



Fig. S16 First-order kinetic plot for the polymerization of *rac*-LA in THF at 25 °C with complex **2** as initiator (THF = 6 mL, $[M]_0$ = 0.5 M, [I] = 0.833 mM, k_{app} = 0.036 min⁻¹).



Fig. S17 First-order kinetic plots for the polymerization of *rac*-LA in THF at 60 °C with complex **3** as initiator (triangle: THF = 2 mL, $[M]_0 = 1 M$, [I] = 5 mM, $k_{app} = 0.022 min^{-1}$; square: THF = 4 mL, $[M]_0 = 0.5 M$, [I] = 2.5 mM, $k_{app} = 0.011 min^{-1}$).

5. Polymerization Kinetics



Fig. S18 ^1H { $^1\text{H}\}$ NMR spectrum of PLA (sample Table 2, Entry 6) in CDCl_3.



Molar mass [Da]

Probe :	Vial 2: WR/518/4 - 1		
Integration von :	Dienstag 19.05.15 10:16:04		21.236 ml
Integration bis :	Dienstag 19.05.15 10:23:27		28.821 ml
Kalibration :	thf20132406.CAL	Eluent :	THE
MHK - A (Kal.):	1.000E+0	MHK - K (Kal.):	0.000E+0 ml/g
Int.StandK :	37.250 ml	Int.StandM :	36.300 ml
Pumpe :	PSS SECcurity	Flußrate :	1.000 ml/min
Konzentration :	0.000 g/l	Injektvolumen :	20.000 ul
Säule 1 :	PSS SDV 5µm	Temperatur :	25.000 °C
Säule 2 :	PSS SDV 5µm	Temperatur :	25.000 °C
Säule 3 :	PSS SDV 5µm	Temperatur :	25.000 °C
Detektor 1 :	I1: RID 1, RI Signal	Versatz :	0.217 ml
Detektor 2 :	I1: VWD 1, Signal A	Versatz :	0.000 ml
Operateur :	mpaul	Messintervall :	1.000 sec

	11: RID 1	, RI Signal	11: VWD	1, Signal A	
		Unsicherheit [%]	Unsicherheit [%]	
Mn :	2.5153e4	11.92	1.3036e4	n/v	g/mol
Mw :	3.4830e4	11.92	2.9148e4	n/v	g/mol
Mz:	4.4841e4	11.92	5.5166e4	n/v	g/mol
Mv:	3.4830e4	11.92	2.9148e4	n/v	g/mol
D :	1.3848e0	16.85	2.2359e0	n/v	
[n]:	0.000000	11.92	0.000000	n/v	ml/g
Vp:	2.4166e1	11.92	2.5141e1	n/v	ml
Mp:	3.5493e4	11.92	2.1440e4	n/v	g/mol
FI:	7.2417e3	11.92	1.0091e0	n/v	ml*V
< 3340	0.00	11.92	0.00	n/v	
w%:	100.00	11.92	100.00	n/v	
> 16602	0.00	11.92	0.00	n/v	

Projekt : Datum : C:\GPC-Daten\Installation.LDX Montag 29.02.16 14:48:33 Kostenstelle : Zeichen :

Fig. S19 GPC chromatogram of PLA (sample Table 2, Entry 10).



Molar mass [Da]

Probe :	Vial 1: WR/513/1 - 1		
Integration von :	Montag 18.05.15 09:45:4	0	21.712 ml
Integration bis :	Montag 18.05.15 09:53:1	5	29.484 ml
Kalibration :	thf20132406.CAL	Eluent :	THE
MHK - A (Kal.):	1.000E+0	MHK - K (Kal.):	0.000E+0 ml/g
Int.StandK :	37.250 ml	Int.StandM :	36.284 ml
Pumpe :	PSS SECcurity	Flußrate :	1.000 ml/min
Konzentration :	0.000 g/l	Injektvolumen :	20.000 ul
Säule 1 :	PSS SDV 5µm	Temperatur :	25.000 °C
Säule 2 :	PSS SDV 5µm	Temperatur :	25.000 °C
Säule 3 :	PSS SDV 5um	Temperatur :	25.000 °C
Detektor 1 :	11: RID 1, RI Signal	Versatz :	0.217 ml
Detektor 2 :	11: VWD 1, Signal A	Versatz :	0.000 ml
Operateur :	mpaul	Messintervall :	1.000 sec

	11: RID 1	, RI Signal	I1: VWD	1, Signal A	
		Unsicherheit	[%]	Unsicherheit	[%]
Mn :	2.1822e4	4.04	1.5553e4	n/v	g/mol
Mw :	2.7027e4	4.04	2.3501e4	n/v	g/mol
Mz:	3.2042e4	4.04	3.3252e4	n/v	g/mol
Mv:	2.7027e4	4.04	2.3501e4	n/v	g/mol
D :	1.2385e0	5.71	1.5110e0	n/v	-
[n]:	0.000000	4.04	0.000000	n/v	ml/g
Vp:	2.4588e1	4.04	2.4743e1	n/v	ml
Mp:	2.8519e4	4.04	2.6332e4	n/v	g/mol
FI :	1.2958e4	4.04	6.003e-1	n/v	mI*V
< 2406	0.00	4.04	0.00	n/v	
w%:	100.00	4.04	100.00	n/v	
> 12888	84 0.00	4.04	0.00	n/v	

Projekt : Datum : C:\GPC-Daten\Installation.LDX Montag 29.02.16 14:46:38 Kostenstelle : Zeichen :

Fig. S20 GPC chromatogram of PCL (sample Table 2, Entry 17).

6. Redox Reactions



Fig. S21 ¹H NMR spectrum (THF-d₈, 400 MHz) of complex 2 after adding one equivalent of FcBAr^F.



Fig S 22 13 C { 1 H} NMR spectrum (THF- d_{8} , 400 MHz) of complex 2 after adding one equivalent of FcBAr^F.



Fig. S23 ^{11}B { ^{1}H } NMR spectrum (THF- d_{8} , 400 MHz) of complex 2 after adding one equivalent of FcBAr^F.



Fig. S24 ¹⁹F {¹H} NMR spectrum (THF-*d*₈, 400 MHz) of complex 2 after adding one equivalent of FcBAr^F.



Fig. S25 ¹H NMR spectrum (THF-*d*₈, 400 MHz) of complex **3** after adding one equivalent of FcBAr^F.



Fig. S26 13 C { 1 H} NMR spectrum (THF- d_{8r} 400 MHz) of complex 3 after adding one equivalent of FcBAr^F.



Fig. S27 ^{11}B { ^{1}H } NMR spectrum (THF- d_{8} , 400 MHz) of complex 3 after adding one equivalent of FcBAr^F.



Fig. S28 19 F { 1 H} NMR spectrum (THF- d_8 , 400 MHz) of complex 3 after adding one equivalent of FcBAr^F.

7. Crystallography

X-ray diffraction data were collected on a Bruker CCD area-detector diffractometer with Mo K α radiation (monolayer optics, λ = 0.71073 Å) using ω scans.^{S1} The SMART program package was used for the data collection and unit cell determination; processing of the raw frame data was performed using SAINT; absorption corrections were applied with SADABS.^{S2} The structures were solved by direct methods (SIR-92).^{S3} All non-hydrogen atoms were refined anisotropically using all reflections with the program SHELXL-2013 as implemented in the program system WinGX.^{S4} All hydrogen atoms were placed in calculated positions and treated as riding. The graphical representations were performed with the program DIAMOND.^{S5}

	$1 \cdot C_6H_{14}$
formula	$C_{46}H_{70}FeLaNO_2S_2Si_2 + C_6H_{14}$
M _w	1070.26
cryst. system and space group	monoclinic, $P 2_1/c$
a [Å]	10.0044(10)
b [Å]	29.326(3)
c [Å]	19.1690(19)
в [°]	92.678(2)
V [Å ³]	5617.8(10)
Ζ	4
d _{calcd} [g cm⁻³]	1.265
radiation (λ [Å])	ΜοΚα (0.71073)
2ϑ _{max} [°]	55.8
μ [mm ⁻¹]	1.16
F [000]	2248
T _{min} , T _{max}	0.533, 0.746
reflns	67369 (R _{int} = 0.0899)
indep. reflns	12368
obs. refins with $l > 2\sigma(l)$	9220
parameters refined	570
GOF	1.047
R1, wR2 for $l > 2\sigma(l)$	0.0423, 0.0905
R1, wR2 for all data	0.0697, 0.0957
$\Delta e_{\max}, \Delta e_{\min} [e \text{ Å}^{-3}]$	0.952, - 0.654

Table S1 Crystallographic data and structure refinement details for complex 1

8. Cyclic Voltammograms



Fig S29a Cyclic voltammogram of LH₂, background (black), complex (red) vs ferrocene/ferrocenium (2.0 mmol/L in 1,2difluorobenzene, 200 mV/s scan rate, 0.10 mol/L [NⁿBu₄][B(C₆F₅)₄]).



 $\label{eq:Fig S29b} \mbox{ Cyclic voltammogram of LH}_2 \mbox{ vs ferrocene/ferrocenium, } E_{1/2} = 0.11 \mbox{ V} (2.0 \mbox{ mmol/L in 1,2-difluorobenzene, 20 mV/s scan rate, 0.10 mol/L } [N'^Bu_4] [B(C_6F_5)_4]).$



Fig S29c Cyclic voltammogram of LH₂ vs ferrocene/ferrocenium, 20 mV/s scan rate (purple), 50 mV/s scan rate (blue), 100 mV/s scan rate (red), 200 mV/s scan rate (black), E_{1/2} = 0.11 V (2.0 mmol/L in 1,2-difluorobenzene, 0.10 mol/L [NⁿBu₄][B(C₆F₅)₄]).



Fig. S30a Cyclic voltammogram of LLaN(SiMe₃)₂ (1), background (red), complex (black) vs ferrocene/ferrocenium (2.0 mmol/L in 1,2-difluorobenzene, 200 mV/s scan rate, 0.10 mol/L [NⁿBu₄][B(C₆F₅)₄]).



Fig. S30b Cyclic voltammogram of 1 vs ferrocene/ferrocenium, $E_{1/2} = 0.093$ V (2.0 mmol/L in 1,2-difluorobenzene, 20 mV/s scan rate, 0.10 mol/L [NⁿBu₄][B(C₆F₅)₄]).



Fig. S30c Cyclic voltammogram of 1 vs ferrocene/ferrocenium, 20 mV/s scan rate (black), 50 mV/s scan rate (red), 100 mV/s scan rate (blue), 200 mV/s scan rate (purple), E_{1/2} = 0.093 V (2.0 mmol/L in 1,2-difluorobenzene, 0.10 mol/L [NⁿBu₄][B(C₆F₅)₄]).



Fig. S31a Cyclic voltammogram of 2, background (red), complex (black) vs ferrocene/ferrocenium (2.0 mmol/L in 1,2-difluorobenzene, 200 mV/s scan rate, 0.10 mol/L $[N^n Bu_4][B(C_6F_5)_4)$.



Fig. S31b Cyclic voltammogram of 2 vs ferrocene/ferrocenium, $E_{1/2} = 0.055$ V (2.0 mmol/L in 1,2-difluorobenzene, 20 mV/s scan rate, 0.10 mol/L [NⁿBu₄][B(C₆F₅)₄]).



Fig. S31c Cyclic voltammogram of 2 vs ferrocene/ferrocenium, 20 mV/s scan rate (black), 50 mV/s scan rate (red), 100 mV/s scan rate (blue), 200 mV/s scan rate (purple), E_{1/2} = 0.055 V (2.0 mmol/L in 1,2-difluorobenzene, 0.10 mol/L [NⁿBu₄][B(C₆F₅)₄]).



Fig. S32a Cyclic voltammogram of 3, background (black), complex (red) vs ferrocene/ferrocenium (2.0 mmol/L in 1,2-difluorobenzene, 200 mV/s scan rate, 0.10 mol/L [NⁿBu₄][B(C₆F₅)₄]).



Fig. S32b Cyclic voltammogram of 3 vs ferrocene/ferrocenium, $E_{1/2} = 0.084$ V (2.0 mmol/L in 1,2-difluorobenzene, 20 mV/s scan rate, 0.10 mol/L [NⁿBu₄][B(C₆F₅)₄]).



Fig. S32c Cyclic voltammogram of 3 vs ferrocene/ferrocenium, 20 mV/s scan rate (black), 50 mV/s scan rate (red), 100 mV/s scan rate (blue), 200 mV/s scan rate (purple), E_{1/2} = 0.084 V (2.0 mmol/L in 1,2-difluorobenzene, 0.10 mol/L [NⁿBu₄][B(C₆F₅)₄]).

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