

SUPPORTING INFORMATION

Combining enzyme and photoredox catalysis for aminoalkylation of indoles via a relay catalysis strategy in one pot

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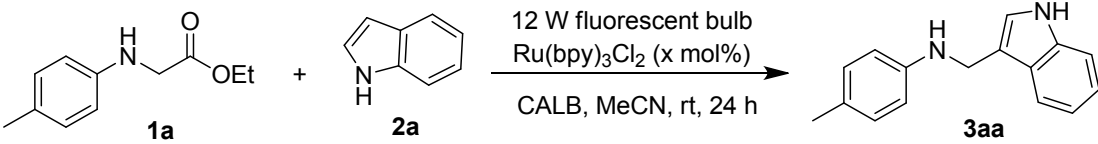
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1. Analytical methods

Reactions were monitored by thin-layer chromatography (TLC) with Haiyang GF254 silica gel plates (Qingdao Haiyang chemical industry Co Ltd. Qingdao, China) using UV light and vanillic aldehyde as visualizing agents. Flash column chromatography was performed using 200-300 mesh silica gel at increased pressure. ^1H NMR and ^{13}C NMR spectra were recorded on Bruker-AM 600 (600 MHz) (Bruker BioSpin AG Ltd., Beijing, China). Chemical shifts were reported in ppm from TMS with the solvent resonance as the internal standard. Data were reported as follows: chemical shifts (δ) in ppm, coupling constants (J) in Hz, and solvent (CDCl_3). High-resolution mass spectra were obtained by using ESI ionization sources (Varian 7.0T FTICR-MS).

2. Extra information for optimization of the reaction conditions

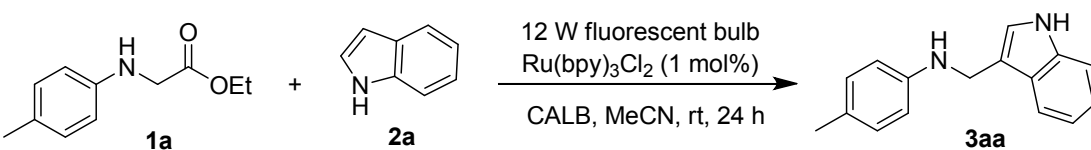
Supplementary Table S1 Effect of photocatalyst loading on the model reaction ^a

		
Entry	$\text{Ru}(\text{bpy})_3\text{Cl}_2$ (x mol%)	Yield (%) ^b
1	1	66
2	2	70
3	3	71
4	4	73
5	5	75
6	7	79
7	10	76

^a Reaction conditions: **1a** (0.2 mmol), **2a** (0.3 mmol), MeCN (1 mL, containing 1.59% water), $\text{Ru}(\text{bpy})_3\text{Cl}_2$ (1-10 mol%) and CALB (114 U) under irradiation of 12 W fluorescent bulb at rt for 24 h.

^b Determined by HPLC.

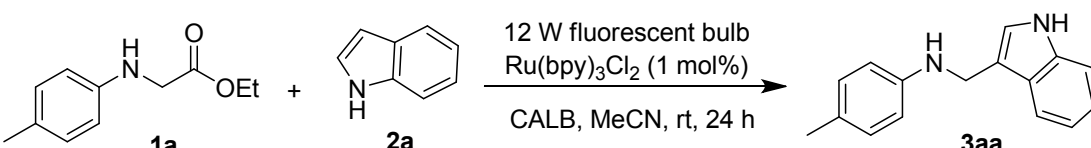
Supplementary Table S2 Effect of molar ratio of substrates on the model reaction ^a

		
Entry	Molar ratio (1a : 2a)	Yield (%) ^b
1	1 : 1	48
2	1 : 1.5	66
3	1 : 2	80
4	1 : 2.5	74
5	1 : 3	72
6	1.5 : 1	88
7	2 : 1	75
8	2.5 : 1	72
9	3 : 1	73

^a Reaction conditions: **1a** (0.2-0.6 mmol), **2a** (0.2-0.6 mmol), MeCN (1 mL, containing 1.59% water), Ru(bpy)₃Cl₂ (1 mol%) and CALB (114 U) under irradiation of 12 W fluorescent bulb at rt for 24 h.

^b Determined by HPLC.

Supplementary Table S3 Effect of enzyme loading on the model reaction ^a

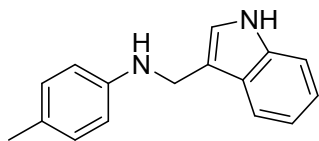
		
Entry	Enzyme loading (U)	Yield (%) ^b
1	38	29
2	76	59
3	114	88
4	152	90
5	190	95
6	228	79

^a Reaction conditions: **1a** (0.3 mmol), **2a** (0.2 mmol), MeCN (1 mL, containing 1.59% water), Ru(bpy)₃Cl₂ (1 mol%) and CALB (38-228 U) under irradiation of 12 W fluorescent bulb at rt for 24 h.

^b Determined by HPLC.

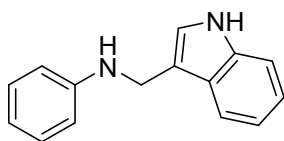
3. Characterization data of the products

N-((1*H*-indol-3-yl)methyl)-4-methylaniline (3aa)



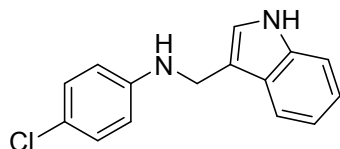
Brown liquid; ^1H NMR (600 MHz, CDCl_3) δ 8.02 (br, 1H), 7.66 (d, J = 7.9 Hz, 1H), 7.36 (d, J = 8.2 Hz, 1H), 7.23 – 7.18 (m, 1H), 7.17 – 7.10 (m, 2H), 7.01 (d, J = 8.2 Hz, 2H), 6.64 (d, J = 8.4 Hz, 2H), 4.72 (br, 1H), 4.45 (s, 2H), 2.25 (s, 3H); ^{13}C NMR (150 MHz, CDCl_3) δ 146.4, 136.5, 129.8, 126.9, 126.6, 122.7, 122.4, 119.8, 119.0, 114.4, 113.2, 111.3, 40.5, 20.4. HRMS (ESI) m/z 235.1240 ($\text{M} - \text{H}^+$), Cal. $\text{C}_{16}\text{H}_{15}\text{N}_2$, 235.1240.

N-((1*H*-indol-3-yl)methyl)aniline (3ba)



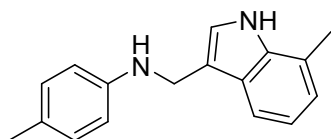
Brown liquid; ^1H NMR (600 MHz, CDCl_3) δ 7.93 (br, 1H), 7.75 (d, J = 7.9 Hz, 1H), 7.43 (d, J = 8.0 Hz, 1H), 7.30 - 7.27 (m, 3H), 7.24 - 7.22 (m, 2H), 6.81 (t, J = 7.3 Hz, 1H), 6.78 (d, J = 8.1 Hz, 2H), 4.83 (br, 1H), 4.55 (s, 2H); ^{13}C NMR (150 MHz, CDCl_3) δ 148.6, 136.6, 129.3, 126.9, 122.9, 122.5, 119.8, 119.1, 117.5, 114.2, 113.0, 111.4, 40.2. HRMS (ESI) m/z 221.1080 ($\text{M} - \text{H}^+$), Cal. $\text{C}_{15}\text{H}_{13}\text{N}_2$, 221.1084.

N-((1*H*-indol-3-yl)methyl)-4-chloroaniline (3ca)



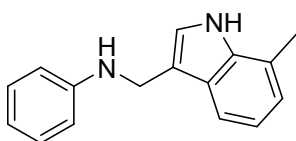
Brown liquid; ^1H NMR (600 MHz, CDCl_3) δ 7.74 (d, $J = 7.8$ Hz, 1H), 7.46 (d, $J = 7.0$ Hz, 1H), 7.31 (s, 1H), 7.24 – 7.20 (m, 3H), 7.18 – 7.16 (m, 1H), 6.68 (d, $J = 7.8$ Hz, 2H), 4.82 (br, 1H), 4.52 (s, 2H); ^{13}C NMR (150 MHz, CDCl_3) δ 147.1, 136.7, 129.1, 126.9, 123.0, 122.6, 122.0, 119.9, 119.0, 116.4, 114.0, 111.6, 40.3. HRMS (ESI) m/z 255.0698 ($\text{M} - \text{H}^+$), Cal. $\text{C}_{15}\text{H}_{12}\text{ClN}_2$, 255.0694.

4-methyl-*N*-((7-methyl-1*H*-indol-3-yl)methyl)aniline (3ab)



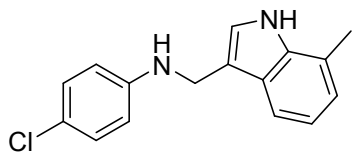
Brown liquid; ^1H NMR (600 MHz, CDCl_3) δ 8.00 (br, 1H), 7.56 (d, $J = 7.7$ Hz, 1H), 7.21 (s, 1H), 7.10 (d, $J = 7.7$ Hz, 1H), 7.08 – 7.04 (m, 3H), 6.67 (d, $J = 8.3$ Hz, 2H), 4.49 (s, 2H), 2.53 (s, 3H), 2.29 (s, 3H); ^{13}C NMR (150 MHz, CDCl_3) δ 146.4, 136.1, 129.8, 129.7, 126.6, 126.3, 122.9, 122.3, 120.0, 116.7, 115.3, 113.1, 40.6, 20.4, 16.5. HRMS (ESI) m/z 249.1399 ($\text{M} - \text{H}^+$), Cal. $\text{C}_{17}\text{H}_{17}\text{N}_2$, 249.1397.

***N*-((7-methyl-1*H*-indol-3-yl)methyl)aniline (3bb)**



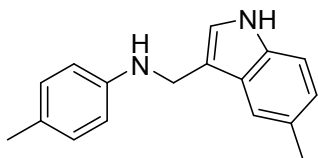
Brown liquid; ^1H NMR (600 MHz, CDCl_3) δ 8.00 (br, 1H), 7.58 (d, $J = 7.8$ Hz, 1H), 7.25 – 7.23 (m, 2H), 7.21 (s, 1H), 7.12 (d, $J = 7.4$ Hz, 1H), 7.09 – 7.07 (m, 1H), 6.78 (t, $J = 7.3$ Hz, 1H), 6.75 (d, $J = 8.3$ Hz, 2H), 4.52 (s, 2H), 2.54 (s, 3H); ^{13}C NMR (150 MHz, CDCl_3) δ 148.6, 136.1, 129.3, 126.3, 122.9, 122.4, 120.5, 120.0, 117.4, 116.7, 114.6, 112.9, 40.2, 16.5. HRMS (ESI) m/z 235.1241 ($\text{M} - \text{H}^+$), Cal. $\text{C}_{16}\text{H}_{15}\text{N}_2$, 235.1240.

4-chloro-*N*-((7-methyl-1*H*-indol-3-yl)methyl)aniline (3cb)



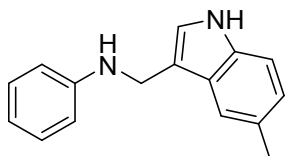
Brown liquid; ^1H NMR (600 MHz, CDCl_3) δ 8.03 (br, 1H), 7.54 (d, $J = 7.8$ Hz, 1H), 7.20 (s, 1H), 7.17 (d, $J = 8.7$ Hz, 1H), 7.15 (d, $J = 8.4$ Hz, 2H), 7.09 – 7.07 (m, 1H), 6.64 (d, $J = 4.8$ Hz, 2H), 4.48 (s, 2H), 2.54 (s, 3H); ^{13}C NMR (150 MHz, CDCl_3) δ 147.1, 136.1, 129.0, 126.2, 123.0, 122.4, 122.4, 121.9, 120.1, 116.6, 116.3, 114.0, 40.3, 16.5. HRMS (ESI) m/z 269.0853 ($\text{M} - \text{H}^+$), Cal. $\text{C}_{16}\text{H}_{14}\text{ClN}_2$, 269.0851.

4-methyl-*N*-((5-methyl-1*H*-indol-3-yl)methyl)aniline (3ac)



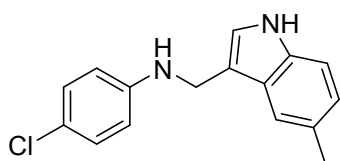
Brown liquid; ^1H NMR (600 MHz, CDCl_3) δ 7.98 (br, 1H), 7.50 (s, 1H), 7.29 (d, $J = 4.0$ Hz, 1H), 7.16 (s, 1H), 7.09 (d, $J = 7.9$ Hz, 1H), 7.06 (d, $J = 8.2$ Hz, 2H), 6.68 (d, $J = 8.2$ Hz, 2H), 4.46 (s, 2H), 2.49 (s, 3H), 2.30 (s, 3H); ^{13}C NMR (150 MHz, CDCl_3) δ 146.4, 134.8, 129.7, 129.0, 127.0, 126.6, 124.0, 122.8, 118.6, 113.3, 113.1, 110.9, 40.5, 21.4, 20.4. HRMS (ESI) m/z 249.1395 ($\text{M} - \text{H}^+$), Cal. $\text{C}_{17}\text{H}_{17}\text{N}_2$, 249.1397.

***N*-((5-methyl-1*H*-indol-3-yl)methyl)aniline (3bc)**



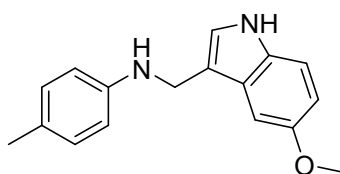
Brown liquid; ^1H NMR (600 MHz, CDCl_3) δ 7.99 (br, 1H), 7.50 (s, 1H), 7.30 (d, $J = 8.2$ Hz, 1H), 7.25 – 7.24 (m, 2H), 7.16 (s, 1H), 7.10 (d, $J = 8.1$ Hz, 1H), 6.75 (d, $J = 8.1$ Hz, 2H), 6.66 (d, $J = 8.1$ Hz, 1H), 4.48 (s, 2H), 2.50 (s, 3H); ^{13}C NMR (150 MHz, CDCl_3) δ 148.6, 134.8, 129.3, 129.1, 127.0, 124.0, 122.9, 118.6, 117.4, 113.1, 112.9, 110.9, 40.1, 21.4. HRMS (ESI) m/z 237.1395 ($M + \text{H}^+$), Cal. $\text{C}_{16}\text{H}_{17}\text{N}_2$, 237.1386.

4-chloro-*N*-((5-methyl-1H-indol-3-yl)methyl)aniline (3cc)



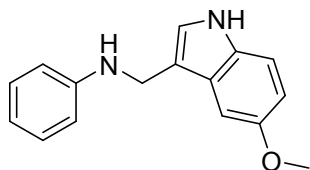
Brown liquid; ^1H NMR (600 MHz, CDCl_3) δ 8.00 (br, 1H), 7.47 (s, 1H), 7.31 (d, $J = 8.3$ Hz, 1H), 7.18 – 7.16 (m, 3H), 7.09 (d, $J = 7.9$ Hz, 1H), 6.64 (d, $J = 8.6$ Hz, 2H), 4.44 (s, 2H), 2.49 (s, 3H); ^{13}C NMR (150 MHz, CDCl_3) δ 147.1, 134.8, 129.2, 129.0, 126.9, 124.1, 122.8, 118.5, 116.2, 114.1, 113.9, 111.0, 40.2, 21.4. HRMS (ESI) m/z 269.0844 ($M - \text{H}^+$), Cal. $\text{C}_{16}\text{H}_{14}\text{ClN}_2$, 269.0851.

***N*-((5-methoxy-1H-indol-3-yl)methyl)-4-methylaniline (3ad)**



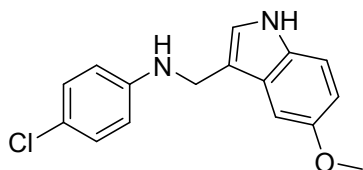
Brown liquid; ^1H NMR (600 MHz, CDCl_3) δ 8.00 (br, 1H), 7.28 (d, $J = 2.8$ Hz, 1H), 7.17 (s, 1H), 7.12 (s, 1H), 7.06 (d, $J = 8.2$ Hz, 2H), 6.92 (d, $J = 8.8$ Hz, 1H), 6.69 (d, $J = 8.3$ Hz, 2H), 4.45 (s, 2H), 3.87 (s, 3H), 2.30 (s, 3H); ^{13}C NMR (150 MHz, CDCl_3) δ 154.3, 146.5, 131.6, 129.7, 127.2, 126.6, 123.4, 113.2, 112.8, 112.0, 100.8, 55.9, 40.5, 20.4. HRMS (ESI) m/z 265.1343 ($M - \text{H}^+$), Cal. $\text{C}_{17}\text{H}_{17}\text{N}_2\text{O}$, 265.1346.

***N*-((5-methoxy-1H-indol-3-yl)methyl)aniline (3bd)**



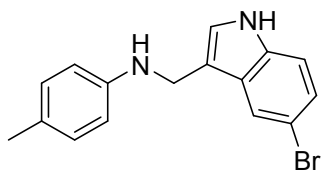
Brown liquid; ^1H NMR (600 MHz, CDCl_3) δ 8.01 (br, 1H), 7.29 (s, 1H), 7.24 (m, 2H), 7.17 (s, J = 2.1 Hz, 1H), 7.13 (s, J = 2.2 Hz, 1H), 6.93 (d, J = 8.8 Hz, 1H), 6.76 (d, J = 7.8 Hz, 2H), 6.65 (d, J = 7.8 Hz, 1H), 4.47 (s, 2H), 3.87 (s, 3H); ^{13}C NMR (150 MHz, CDCl_3) δ 154.3, 148.7, 129.3, 127.2, 123.5, 117.4, 115.1, 113.0, 112.8, 112.0, 100.8, 56.0, 40.2. HRMS (ESI) m/z 253.1337 ($\text{M} - \text{H}^+$), Cal. $\text{C}_{16}\text{H}_{17}\text{N}_2\text{O}$, 253.1336.

4-chloro-*N*-((5-methoxy-1H-indol-3-yl)methyl)aniline (3cd)



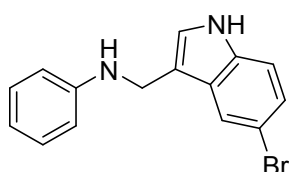
Brown liquid; ^1H NMR (600 MHz, CDCl_3) δ 8.01 (br, 1H), 7.17 – 7.16 (m, 3H), 7.09 (d, J = 1.8 Hz, 2H), 6.92 (d, J = 8.7 Hz, 1H), 6.65 (d, J = 8.8 Hz, 2H), 4.43 (s, 2H), 3.86 (s, 3H); ^{13}C NMR (150 MHz, CDCl_3) δ 154.4, 147.1, 129.1, 127.1, 123.5, 123.1, 116.2, 114.1, 112.8, 112.1, 100.7, 56.0, 40.3. HRMS (ESI) m/z 285.0798 ($\text{M} - \text{H}^+$), Cal. $\text{C}_{16}\text{H}_{14}\text{ClN}_2\text{O}$, 285.0800.

***N*-((5-bromo-1H-indol-3-yl)methyl)-4-methylaniline (3ae)**



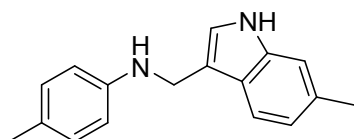
Brown liquid; ^1H NMR (600 MHz, CDCl_3) δ 8.17 (br, 1H), 7.83 (s, 1H), 7.32 (d, $J = 8.6$ Hz, 1H), 7.28 (d, $J = 8.1$ Hz, 1H), 7.19 (s, 1H), 7.05 (d, $J = 8.1$ Hz, 2H), 6.66 (d, $J = 8.2$ Hz, 2H), 4.44 (s, 2H), 2.29 (s, 3H); ^{13}C NMR (150 MHz, CDCl_3) δ 146.1, 135.1, 129.8, 128.6, 127.0, 125.2, 123.8, 121.7, 114.1, 113.2, 113.0, 112.7, 40.3, 20.4. HRMS (ESI) m/z 315.0498 ($\text{M} + \text{H}^+$), Cal. $\text{C}_{16}\text{H}_{16}\text{BrN}_2$, 315.0492.

***N*-((5-bromo-1H-indol-3-yl)methyl)aniline (3be)**



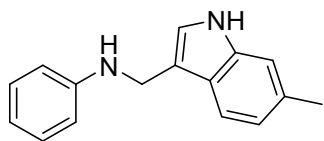
Brown liquid; ^1H NMR (600 MHz, CDCl_3) δ 8.16 (br, 1H), 7.84 (s, 1H), 7.33 (d, $J = 8.0$ Hz, 1H), 7.29 - 7.25 (m, 3H), 7.20 (s, 1H), 6.79 (t, $J = 7.3$ Hz, 1H), 6.74 (d, $J = 7.8$ Hz, 2H), 4.46 (s, 2H); ^{13}C NMR (150 MHz, CDCl_3) δ 148.3, 135.1, 129.3, 128.5, 125.3, 123.9, 121.7, 117.7, 113.8, 113.1, 112.7, 40.0. HRMS (ESI) m/z 301.0341 ($\text{M} + \text{H}^+$), Cal. $\text{C}_{15}\text{H}_{14}\text{BrN}_2$, 301.0335.

4-methyl-*N*-((6-methyl-1H-indol-3-yl)methyl)aniline (3af)



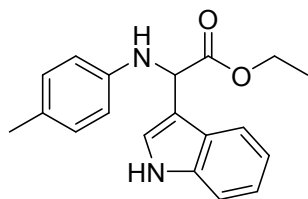
Brown liquid; ^1H NMR (600 MHz, CDCl_3) δ 7.95 (br, 1H), 7.59 (d, $J = 8.0$ Hz, 1H), 7.20 (s, 1H), 7.12 (s, 1H), 7.06 (d, $J = 8.2$ Hz, 2H), 7.02 (d, $J = 8.0$ Hz, 1H), 6.67 (d, $J = 8.3$ Hz, 2H), 4.47 (s, 2H), 2.51 (s, 3H), 2.30 (s, 3H); ^{13}C NMR (150 MHz, CDCl_3) δ 146.4, 136.9, 132.2, 129.7, 126.6, 122.0, 121.5, 118.7, 114.1, 113.1, 111.2, 40.5, 21.7, 20.4. HRMS (ESI) m/z 251.1549 ($\text{M} + \text{H}^+$), Cal. $\text{C}_{17}\text{H}_{19}\text{N}_2$, 251.1543.

***N*-((6-methyl-1H-indol-3-yl)methyl)aniline (3bf)**



Brown liquid; ^1H NMR (600 MHz, CDCl_3) δ 7.94 (br, 1H), 7.59 (d, $J = 8.0$ Hz, 1H), 7.26 – 7.23 (m, 2H), 7.20 (s, 1H), 7.12 (s, 1H), 7.02 (d, $J = 8.1$ Hz, 1H), 6.74 (d, $J = 7.7$ Hz, 2H), 6.66 (t, $J = 7.2$ Hz, 1H), 4.49 (s, 2H), 2.51 (s, 3H); ^{13}C NMR (150 MHz, CDCl_3) δ 148.6, 136.9, 132.3, 129.3, 122.0, 121.6, 118.7, 117.4, 115.2, 112.9, 111.2, 111.0, 40.2, 21.7. HRMS (ESI) m/z 237.1389 ($M + \text{H}^+$), Cal. $\text{C}_{16}\text{H}_{17}\text{N}_2$, 237.1386.

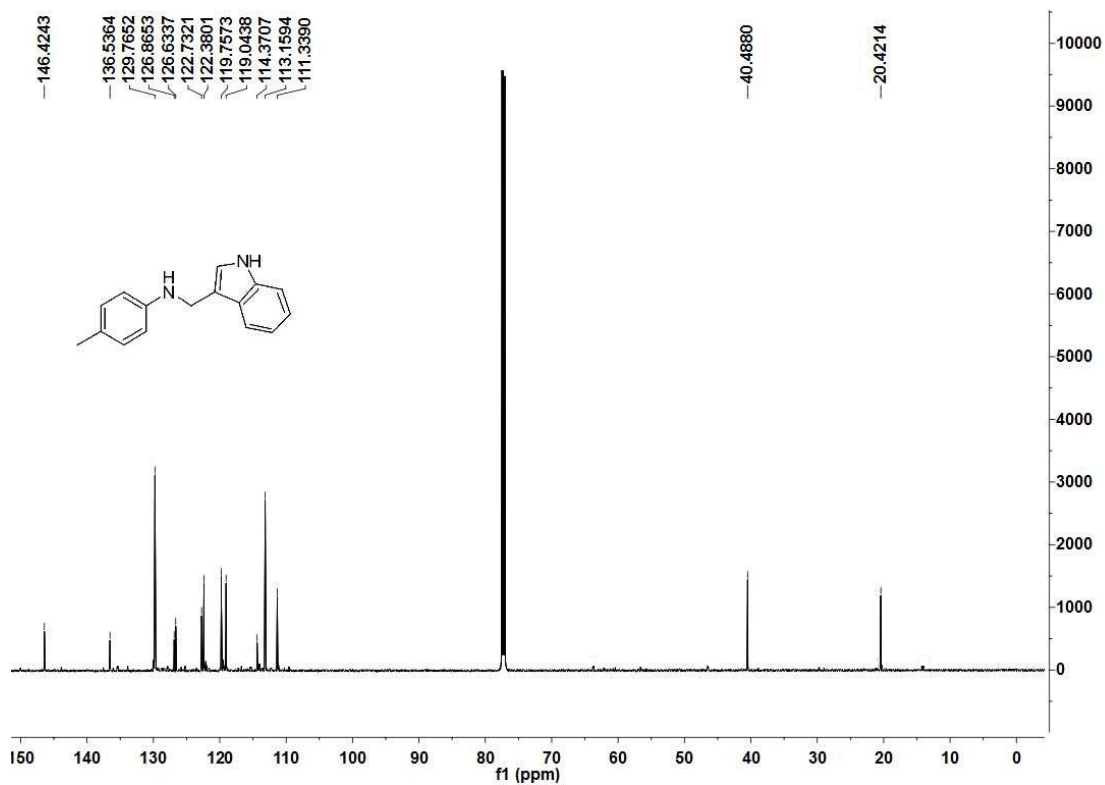
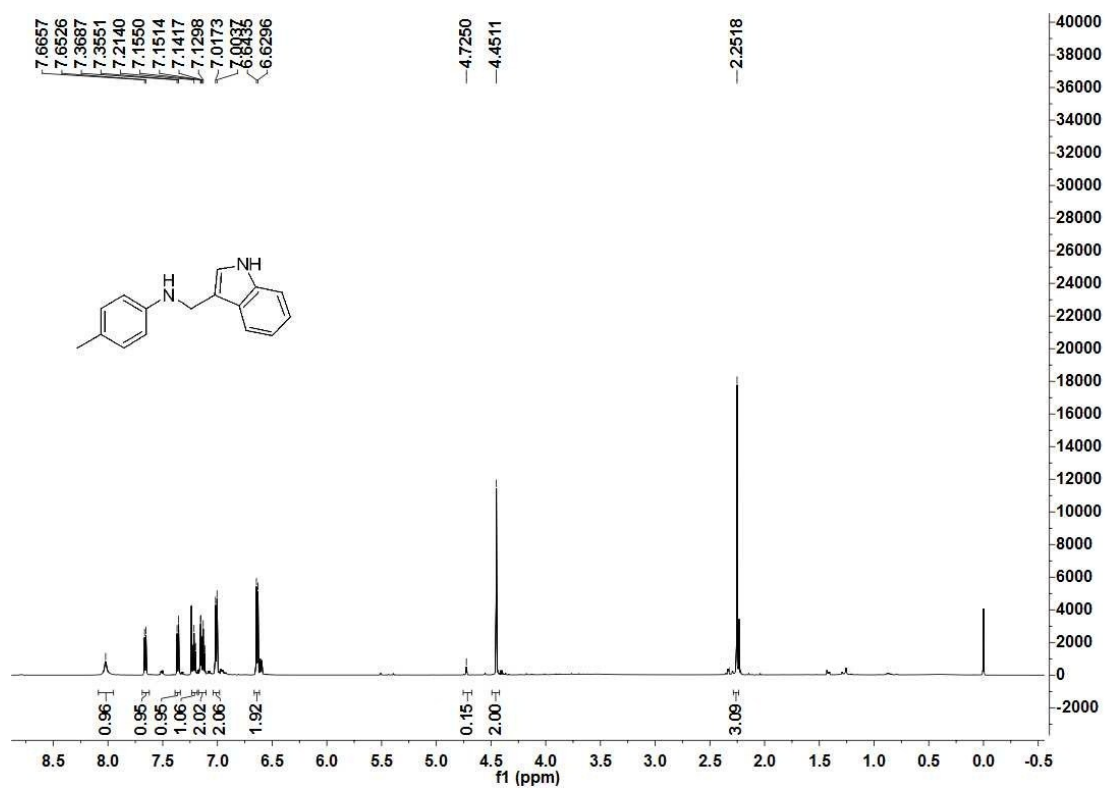
ethyl 2-(1*H*-indol-3-yl)-2-(*p*-tolylamino)acetate (4)¹



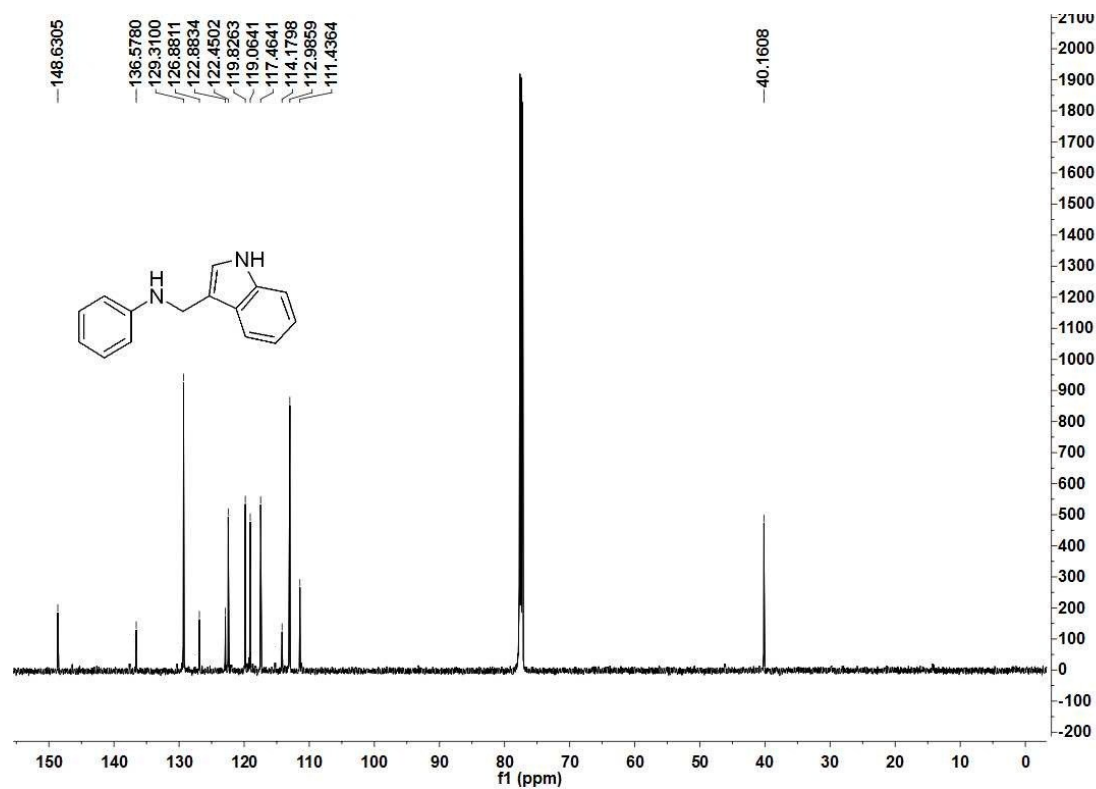
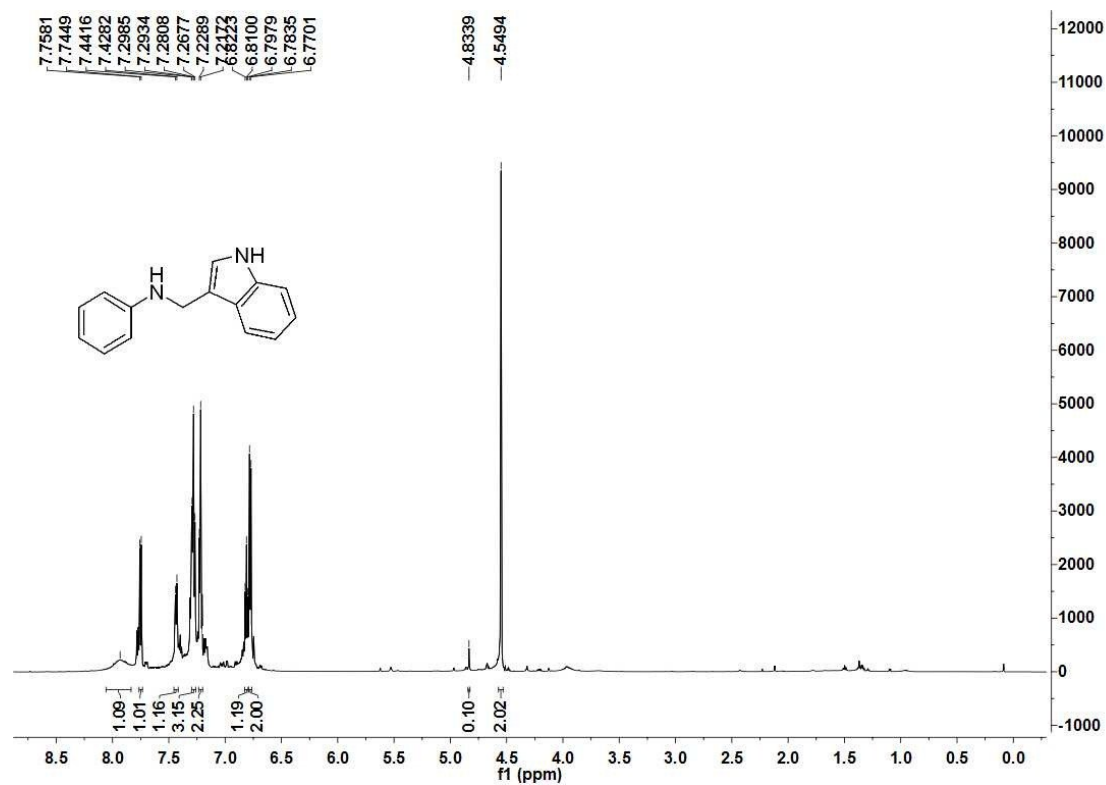
Yellow liquid; ^1H NMR (600 MHz, CDCl_3) δ 8.15 (s, 1H), 7.82 (d, $J = 7.9$ Hz, 1H), 7.33 (d, $J = 8.1$ Hz, 1H), 7.23 – 7.12 (m, 3H), 6.95 (d, $J = 8.1$ Hz, 2H), 6.56 (d, $J = 8.2$ Hz, 2H), 5.36 (s, 1H), 4.18 (ddd, $J = 75.3, 10.8, 7.1$ Hz, 2H), 2.21 (s, 3H), 1.20 (t, $J = 7.1$ Hz, 3H); ^{13}C NMR (150 MHz, CDCl_3) δ 172.7, 144.3, 136.5, 129.8, 127.4, 125.9, 123.1, 122.5, 120.0, 119.6, 113.7, 112.8, 111.4, 61.5, 54.7, 20.4, 14.2.

4. ^1H NMR and ^{13}C NMR spectra of the products

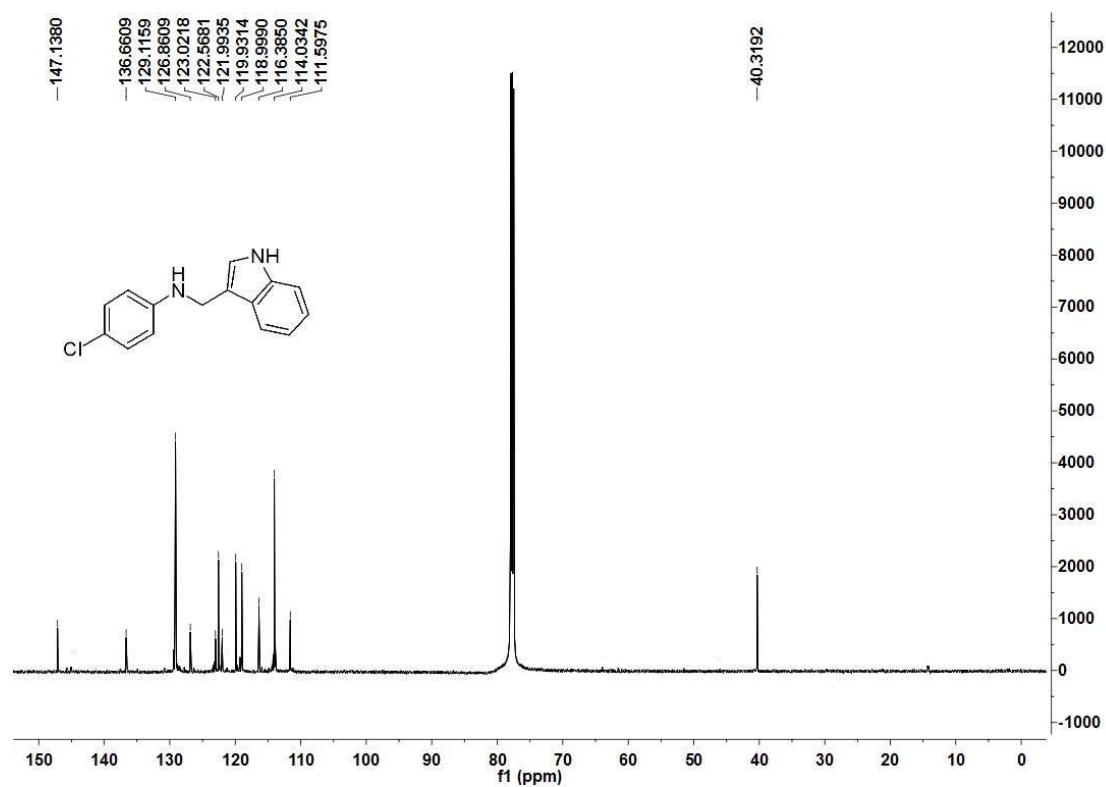
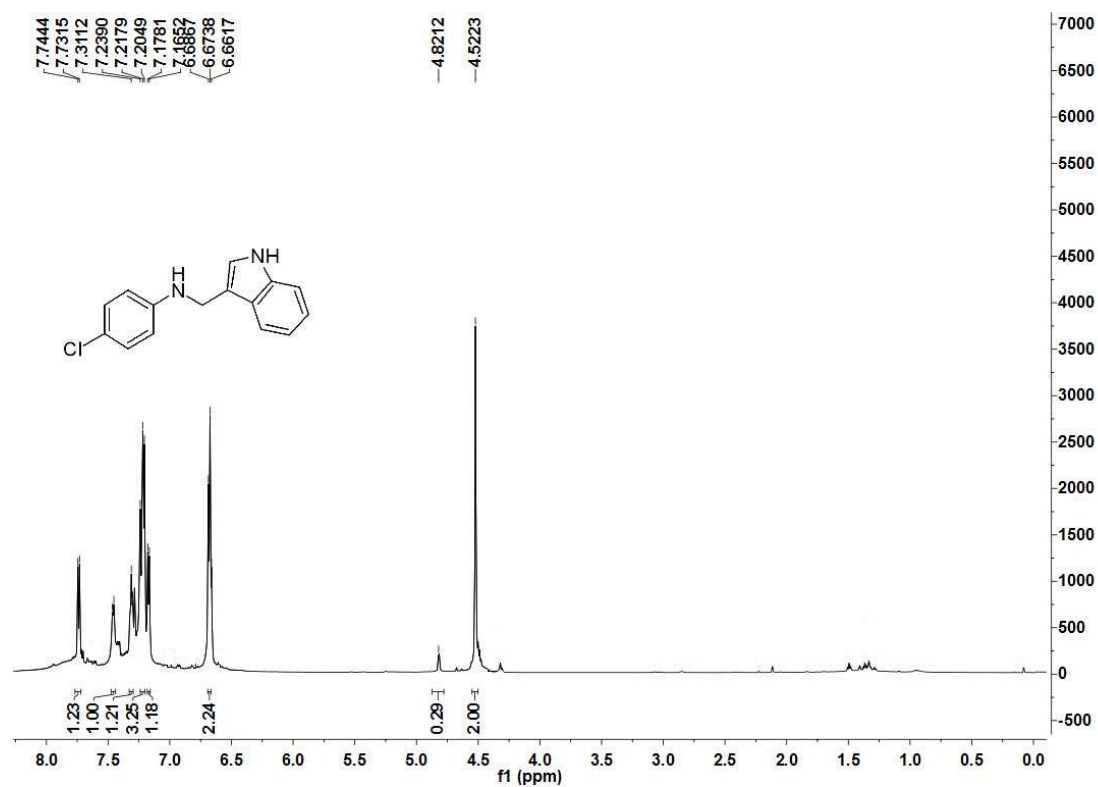
3aa



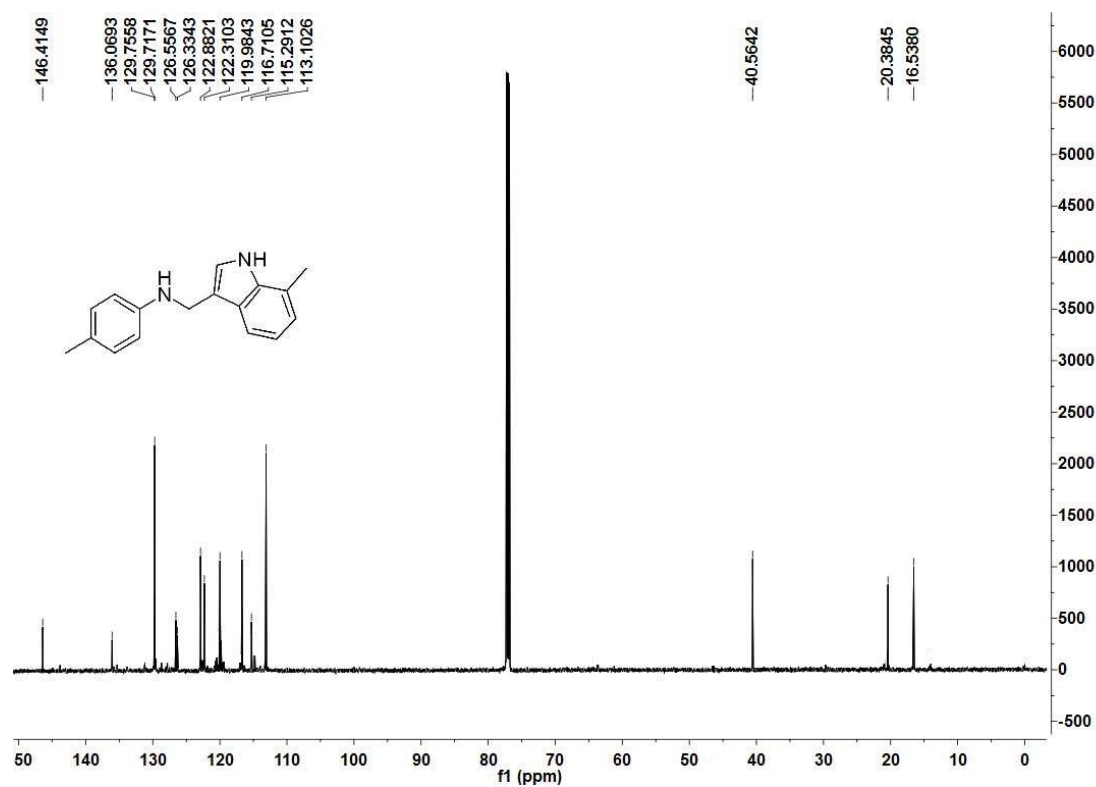
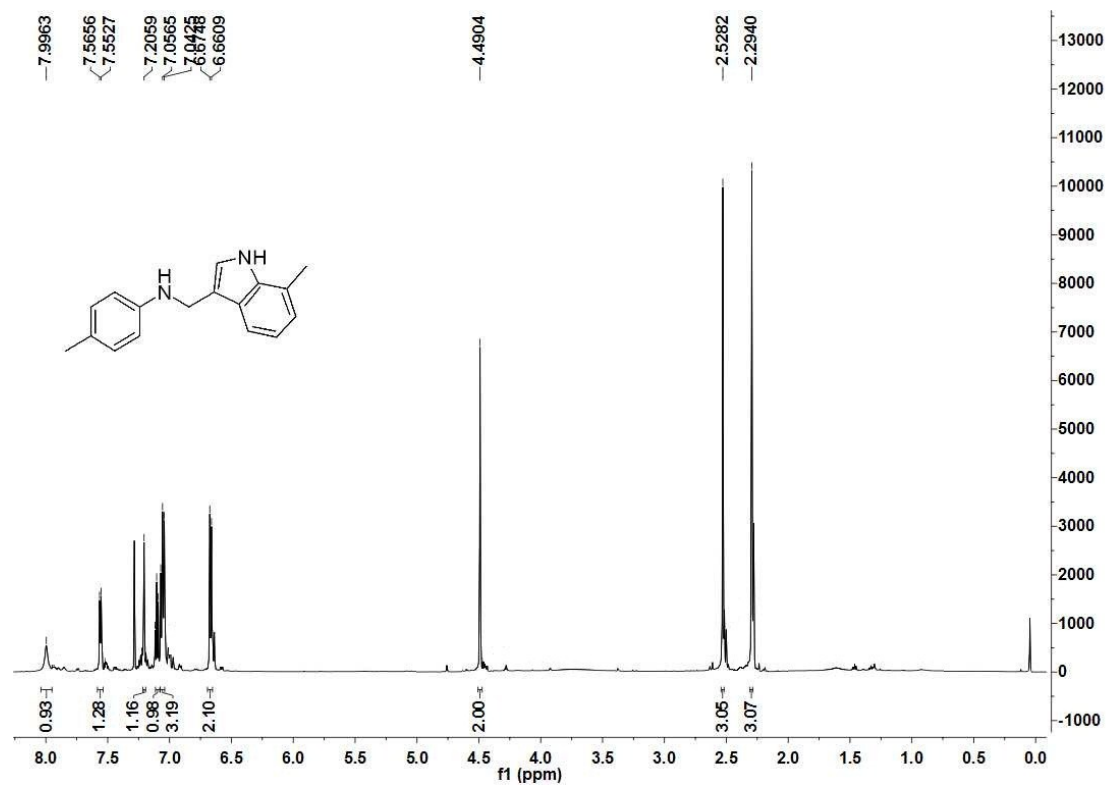
3ba



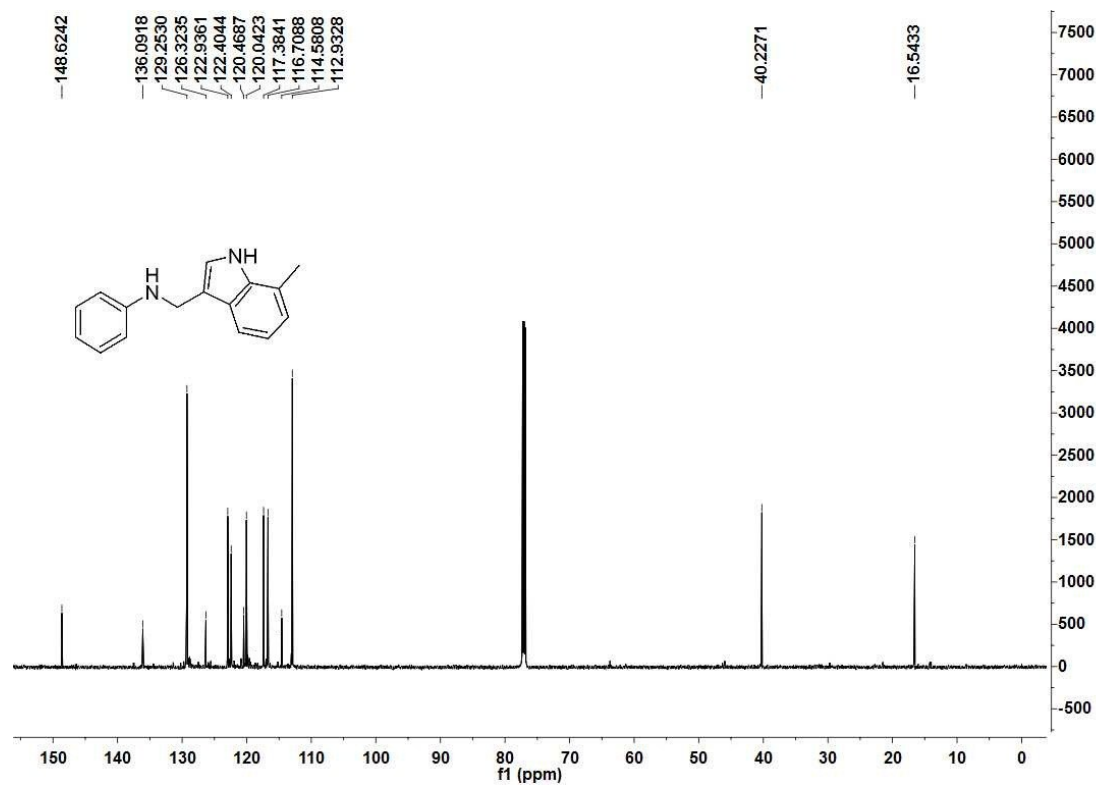
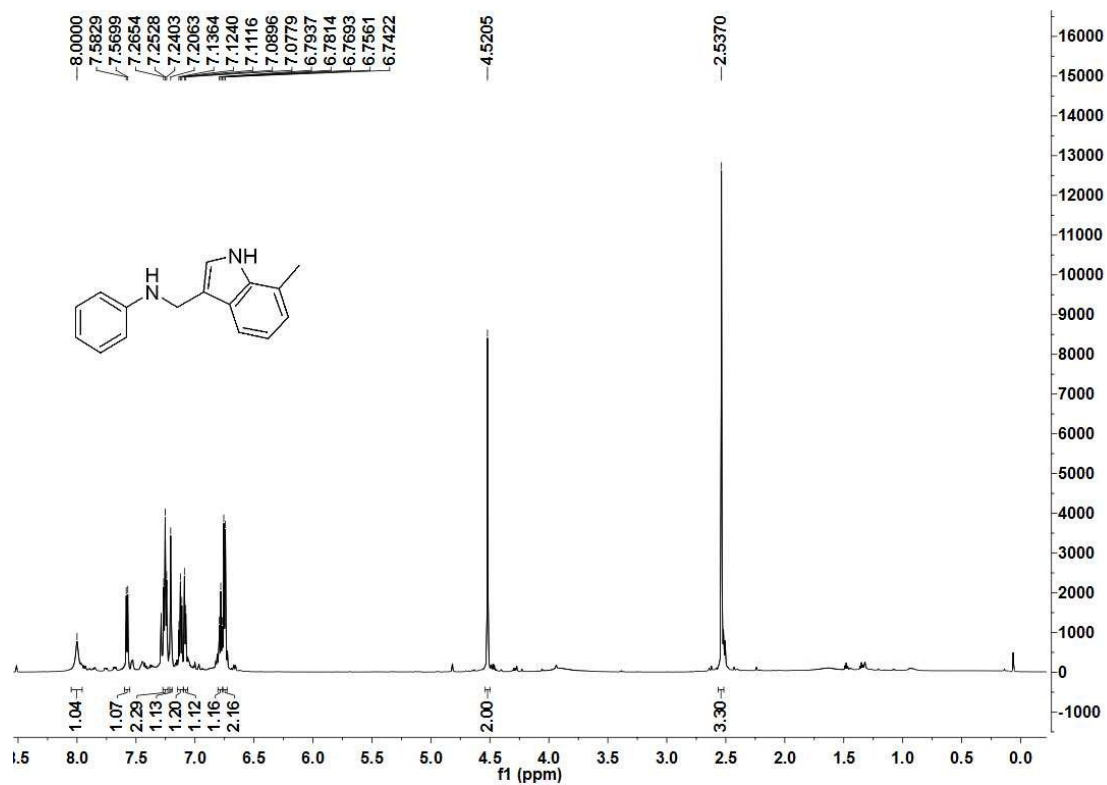
3ca



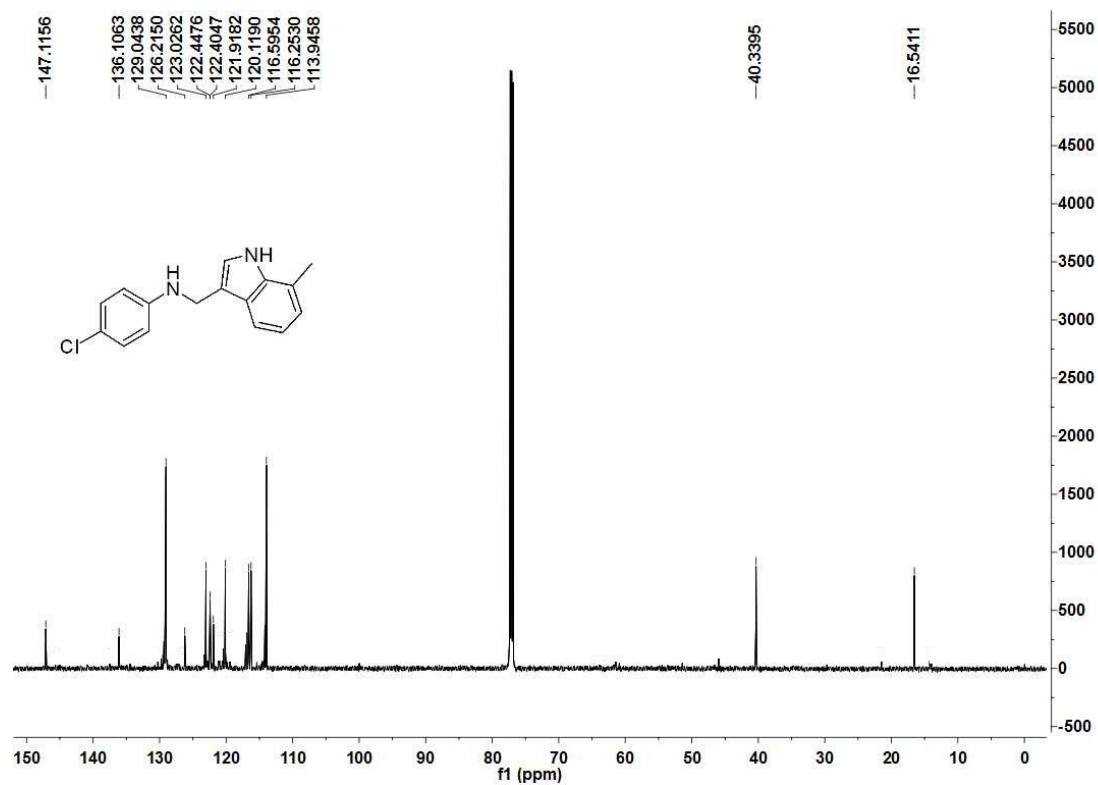
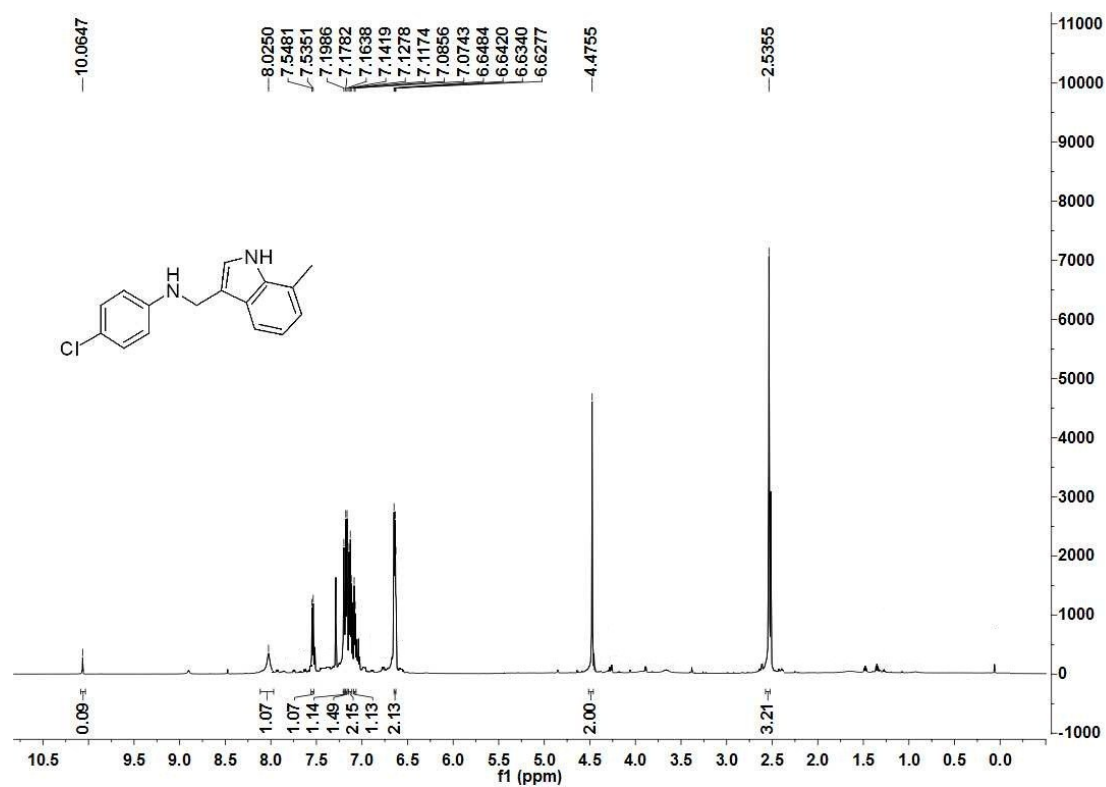
3ab



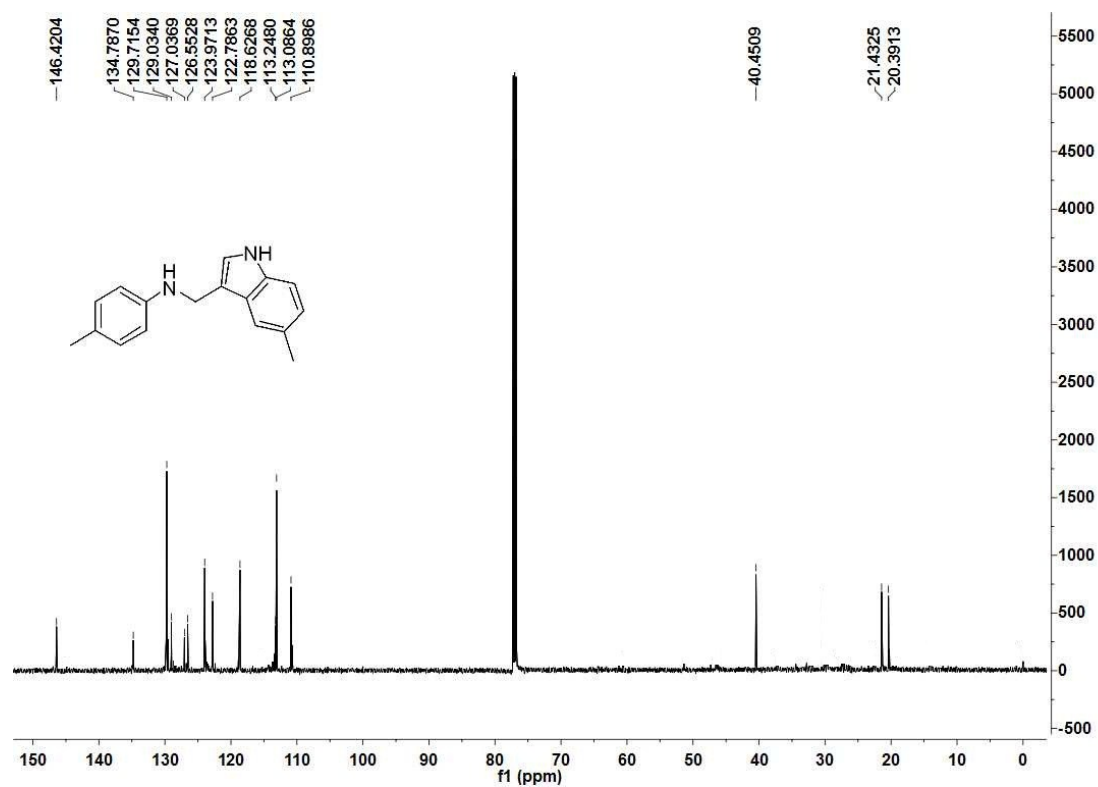
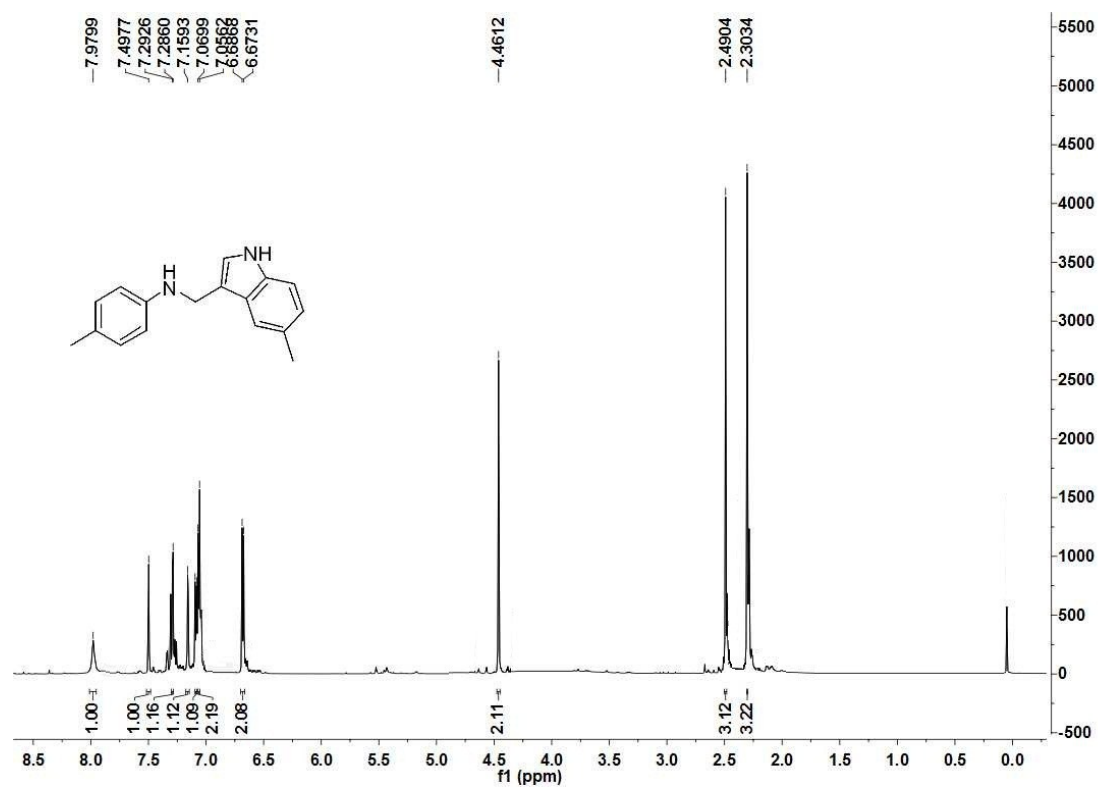
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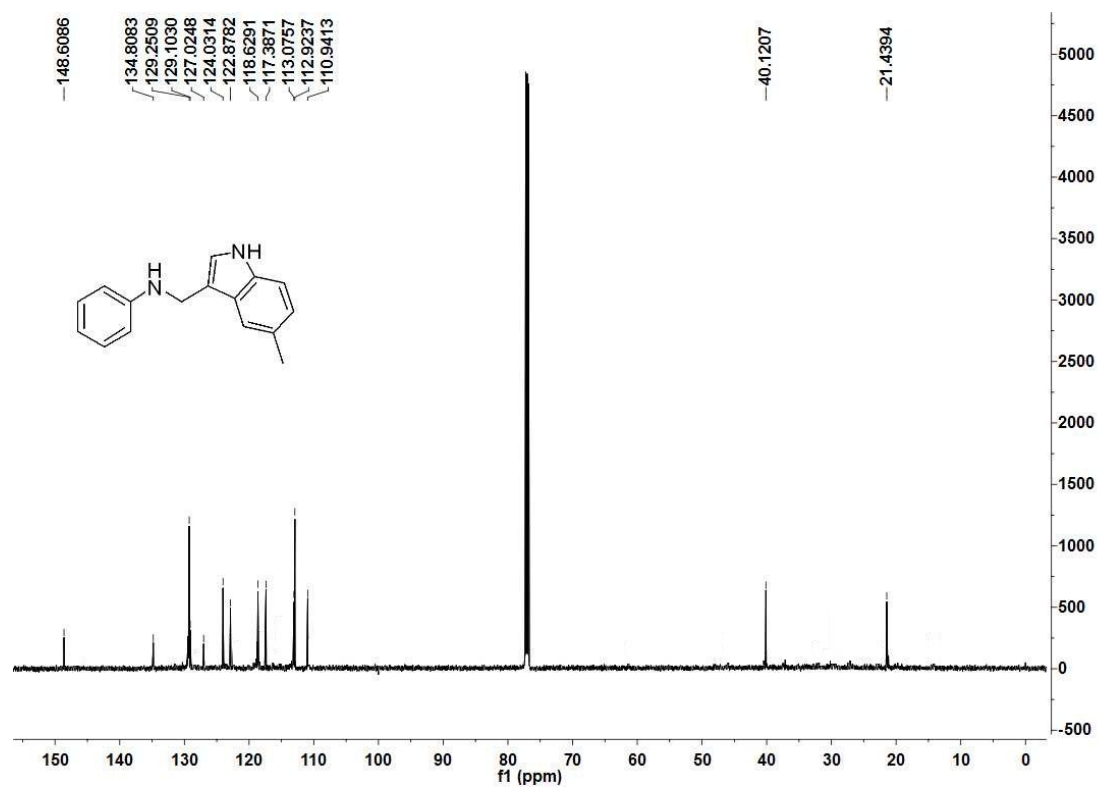
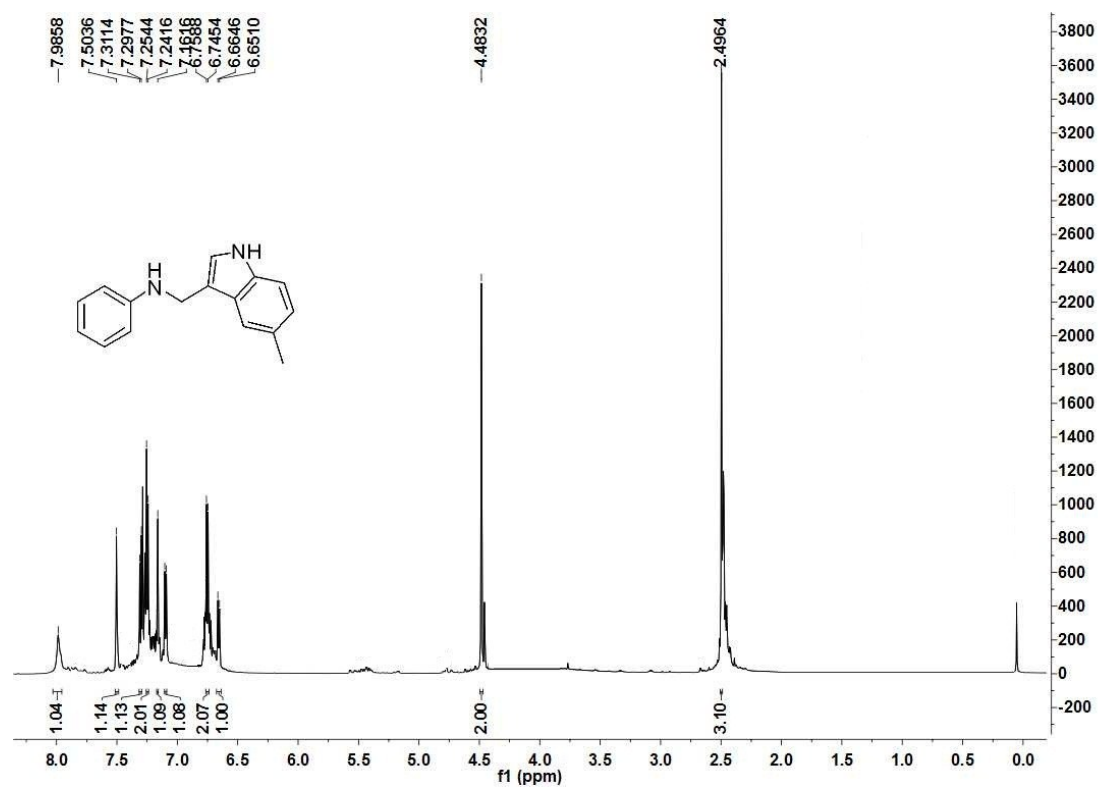
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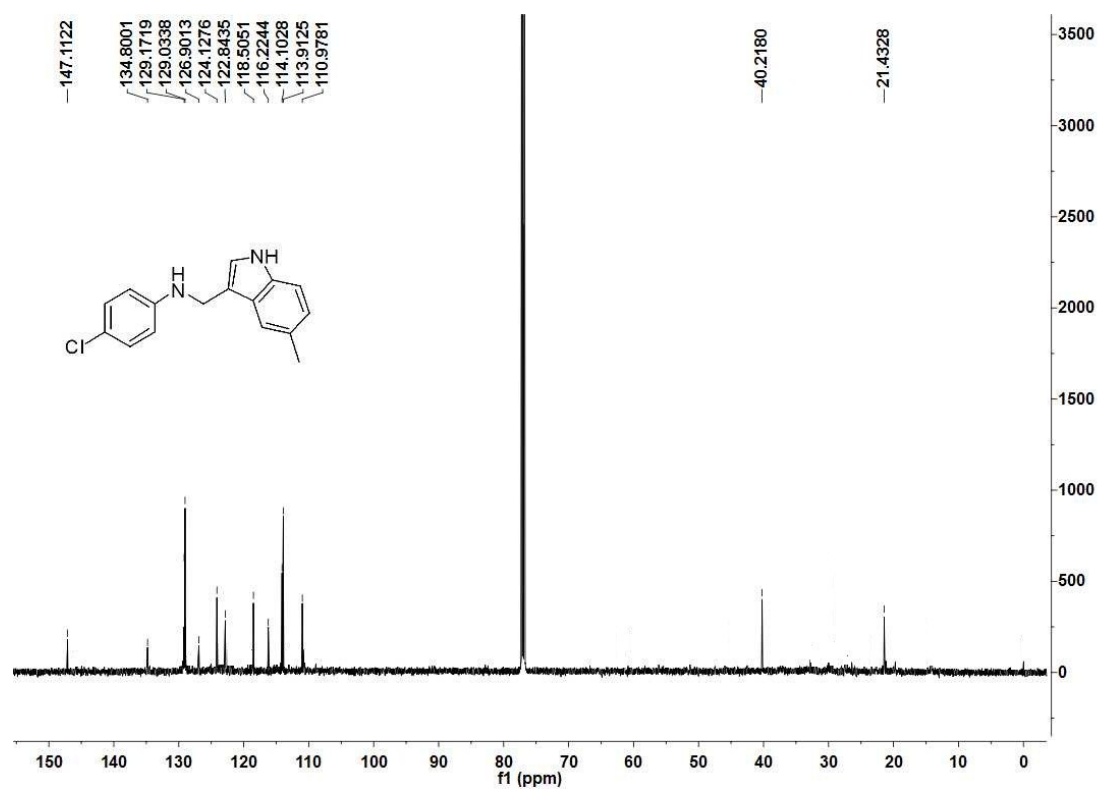
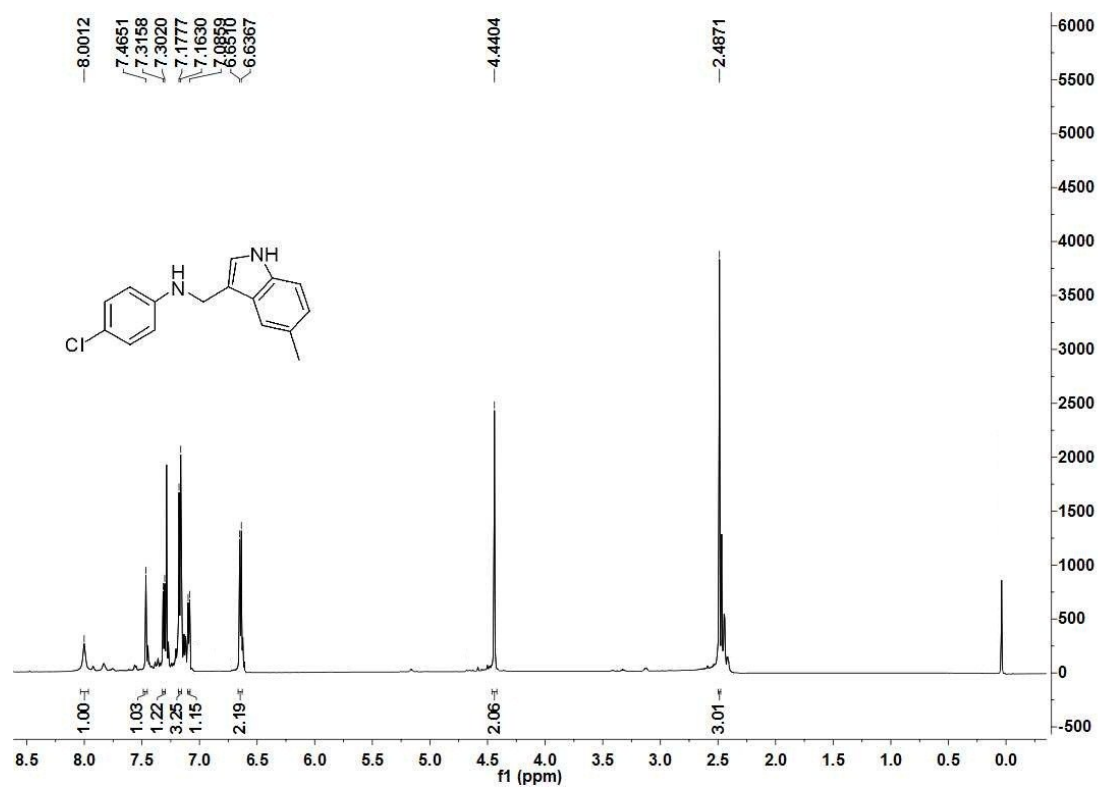
3ac



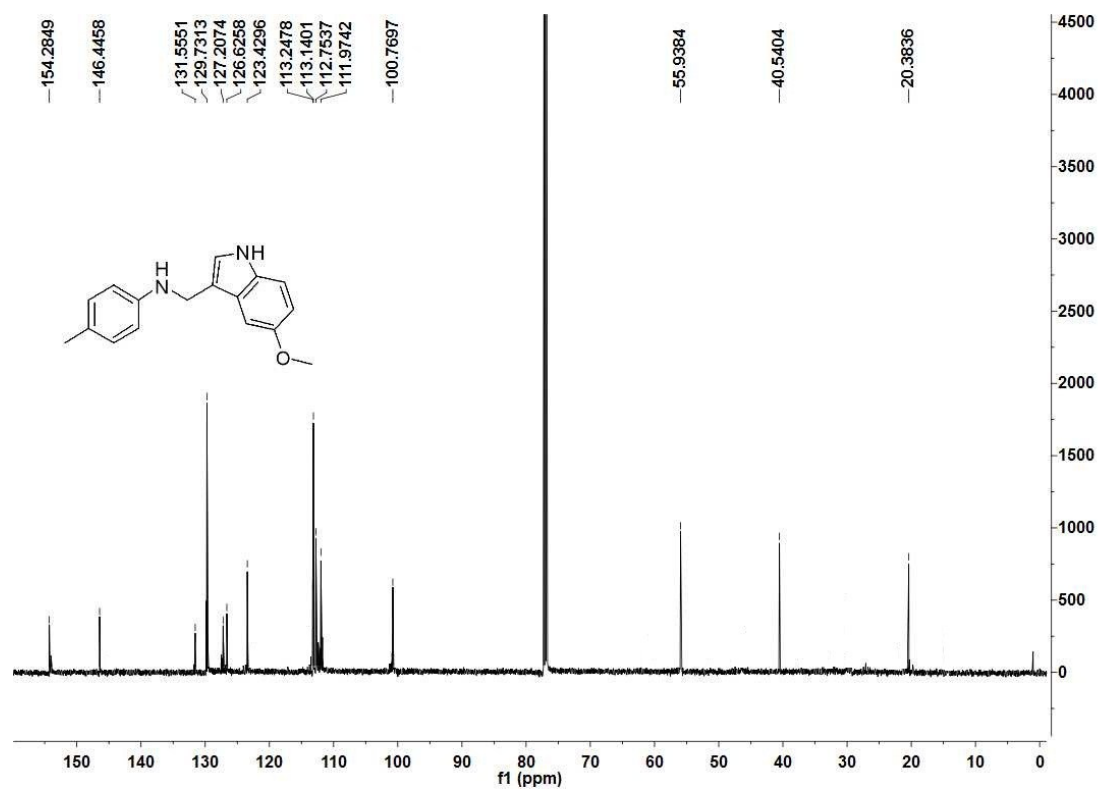
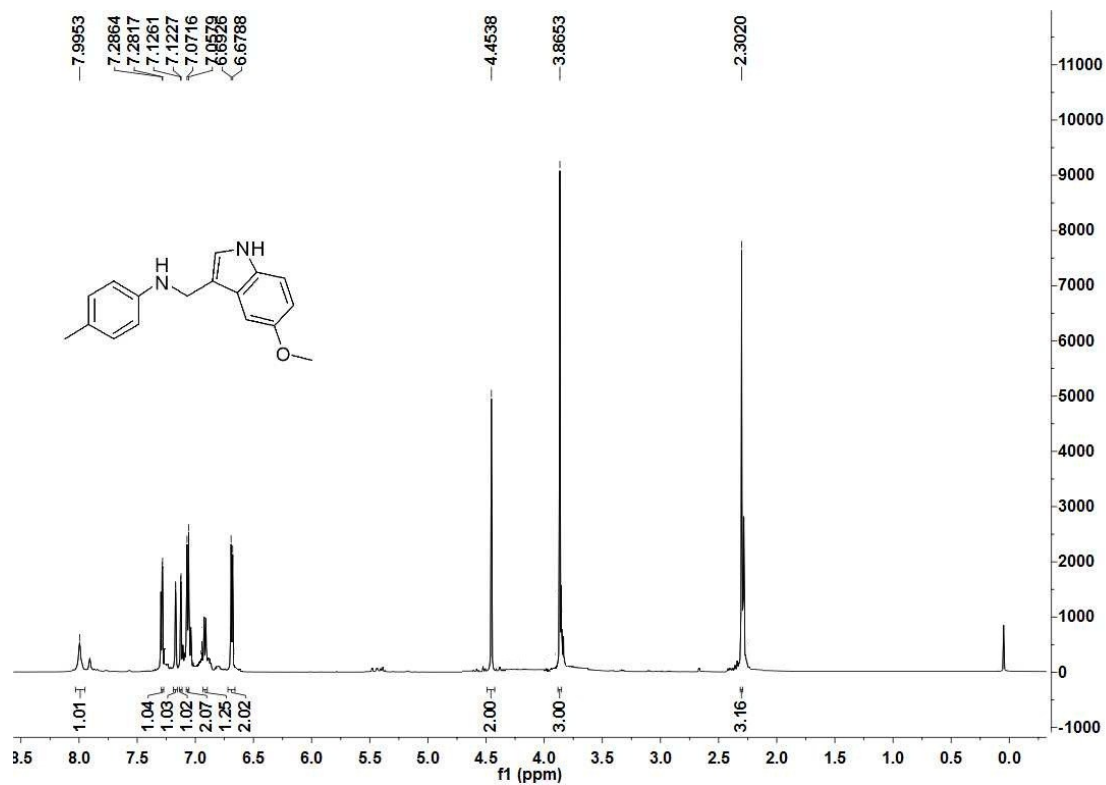
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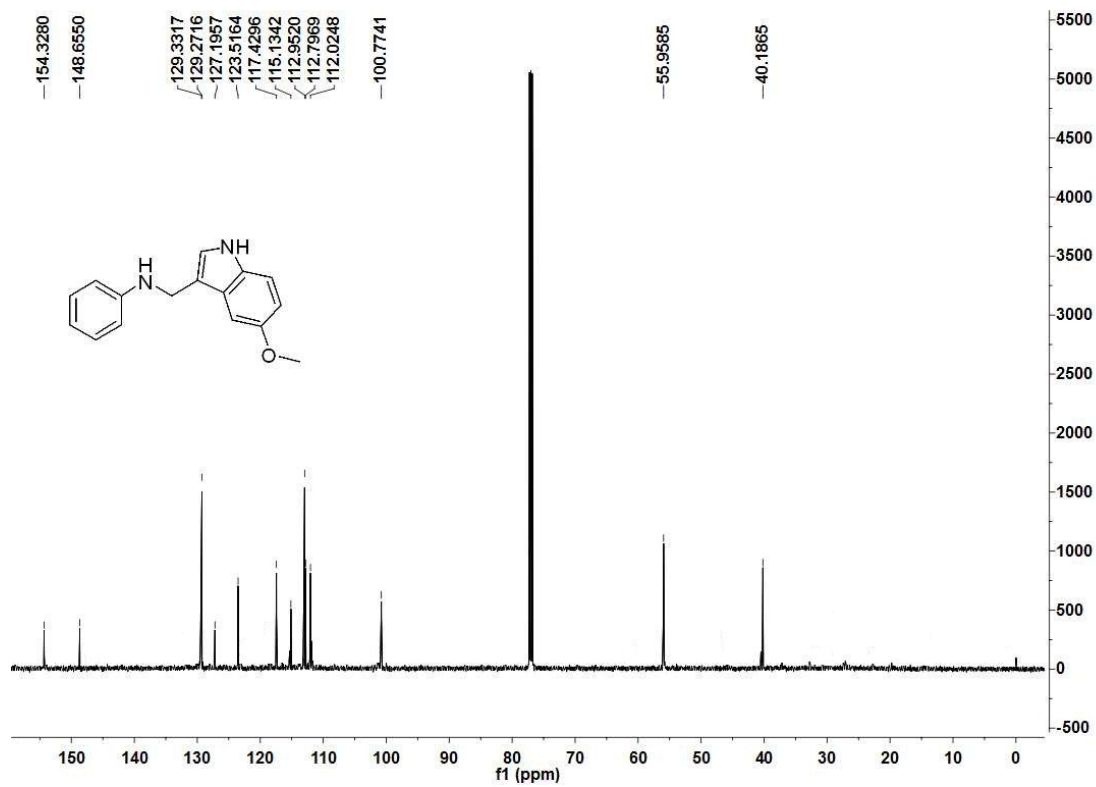
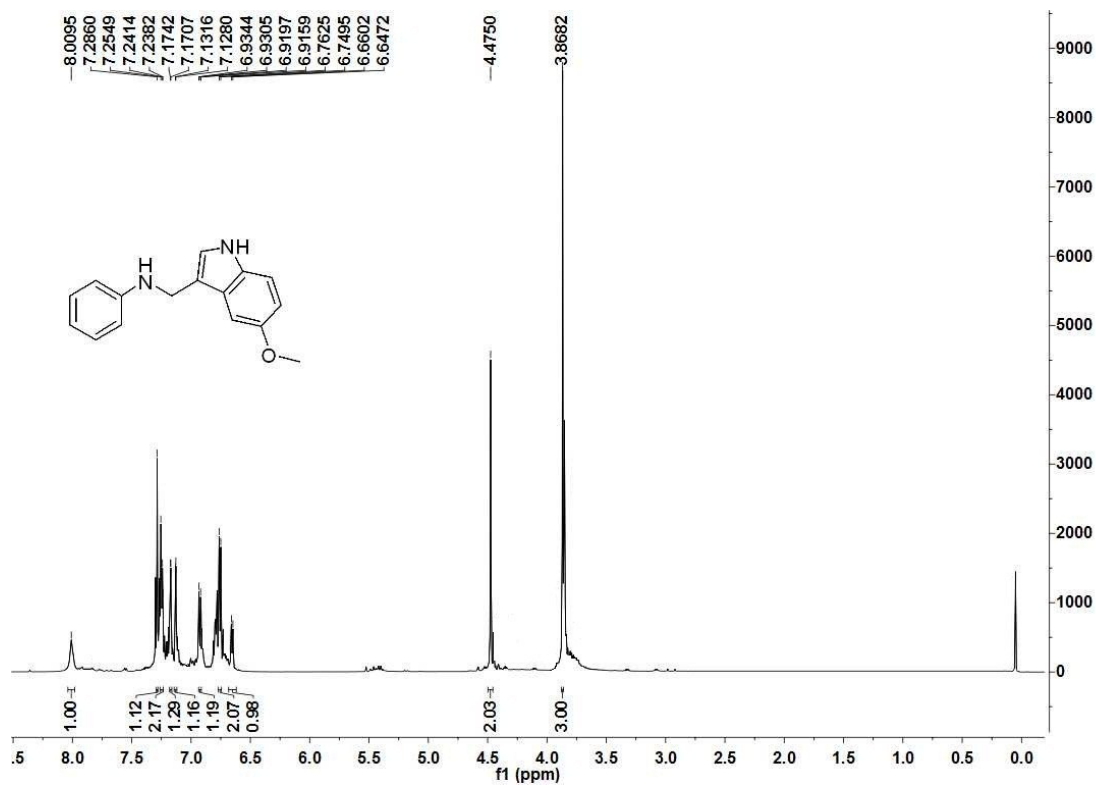
3cc



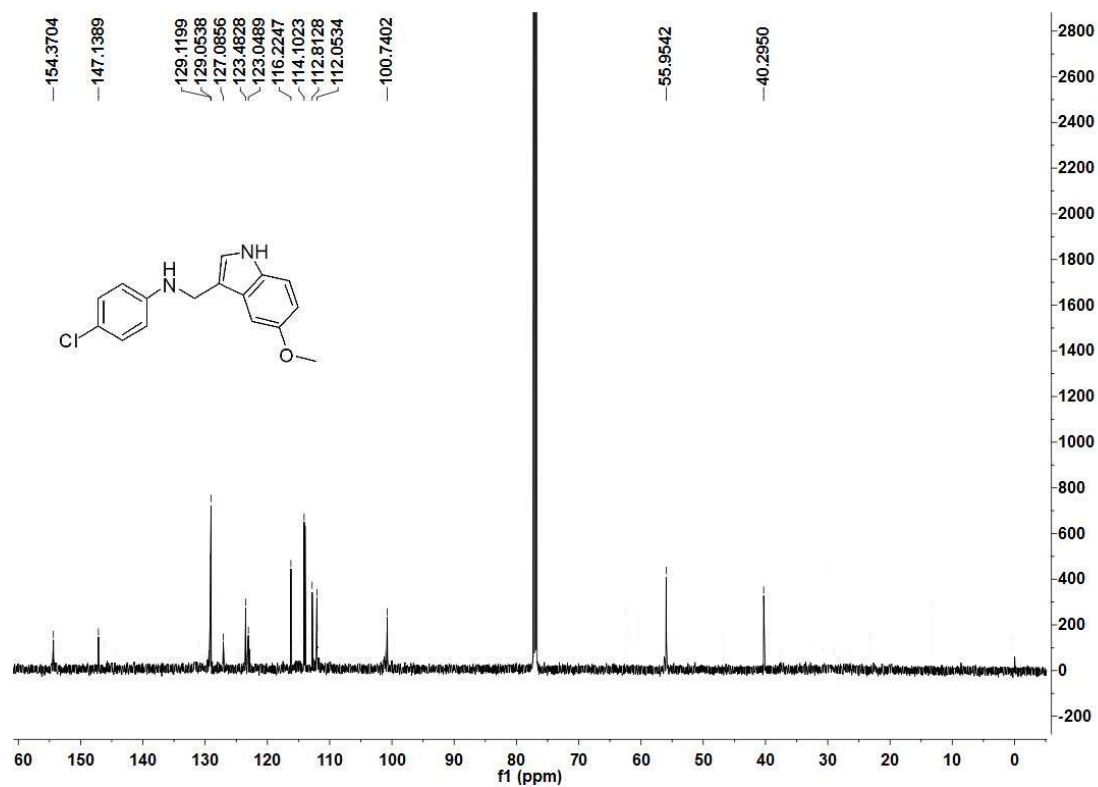
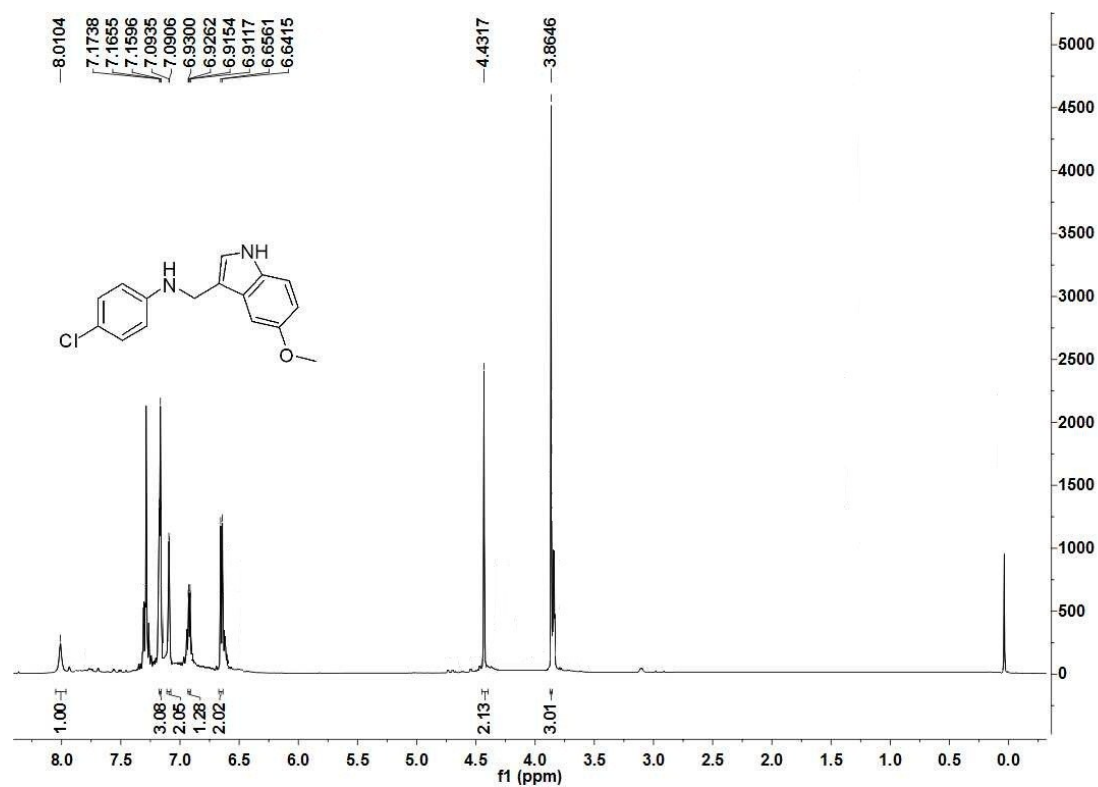
3ad



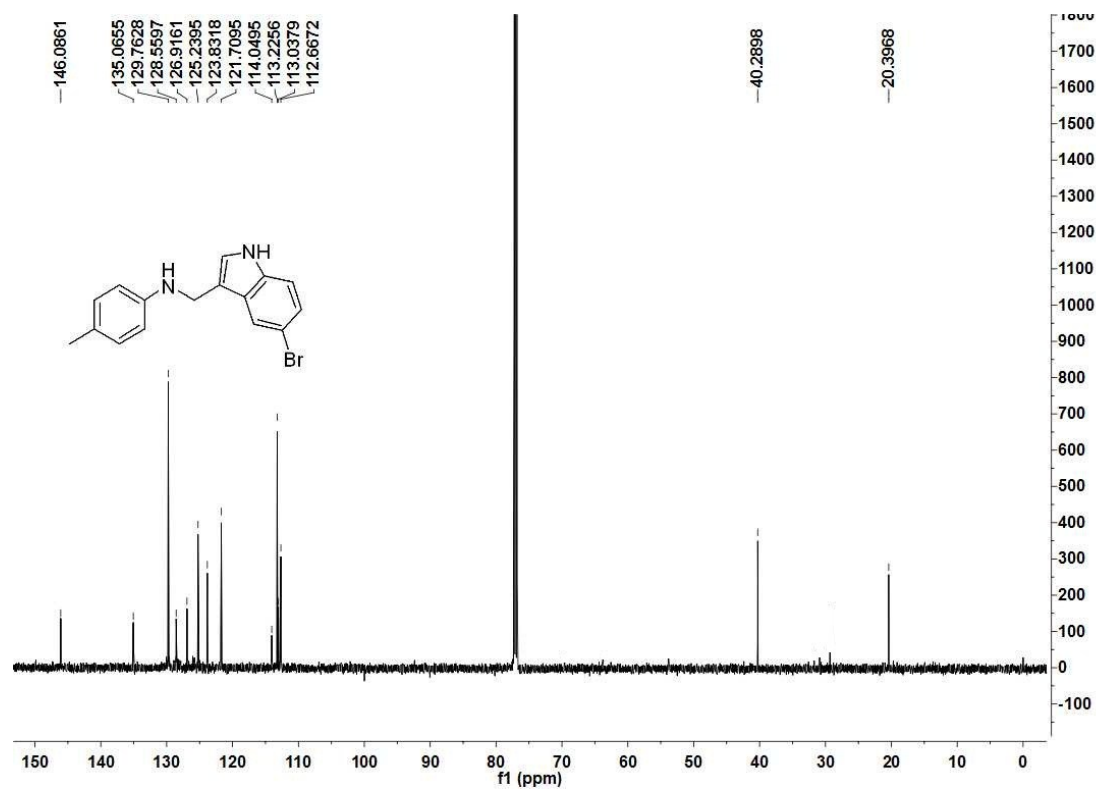
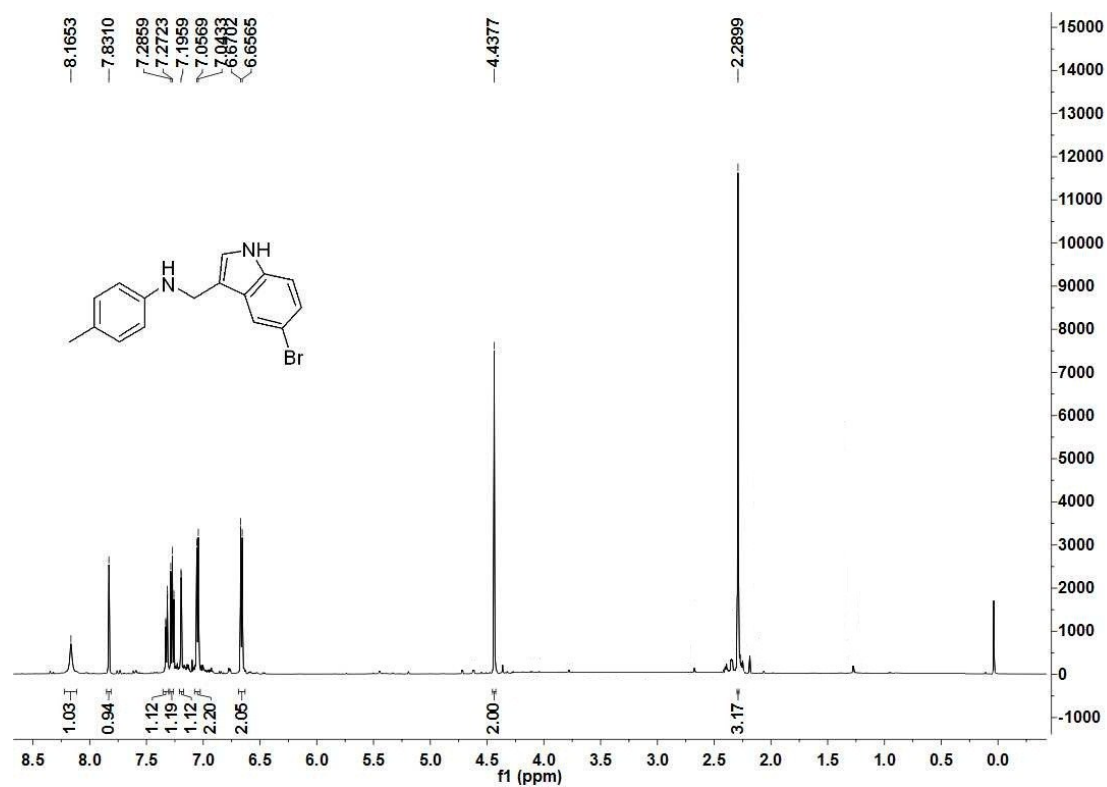
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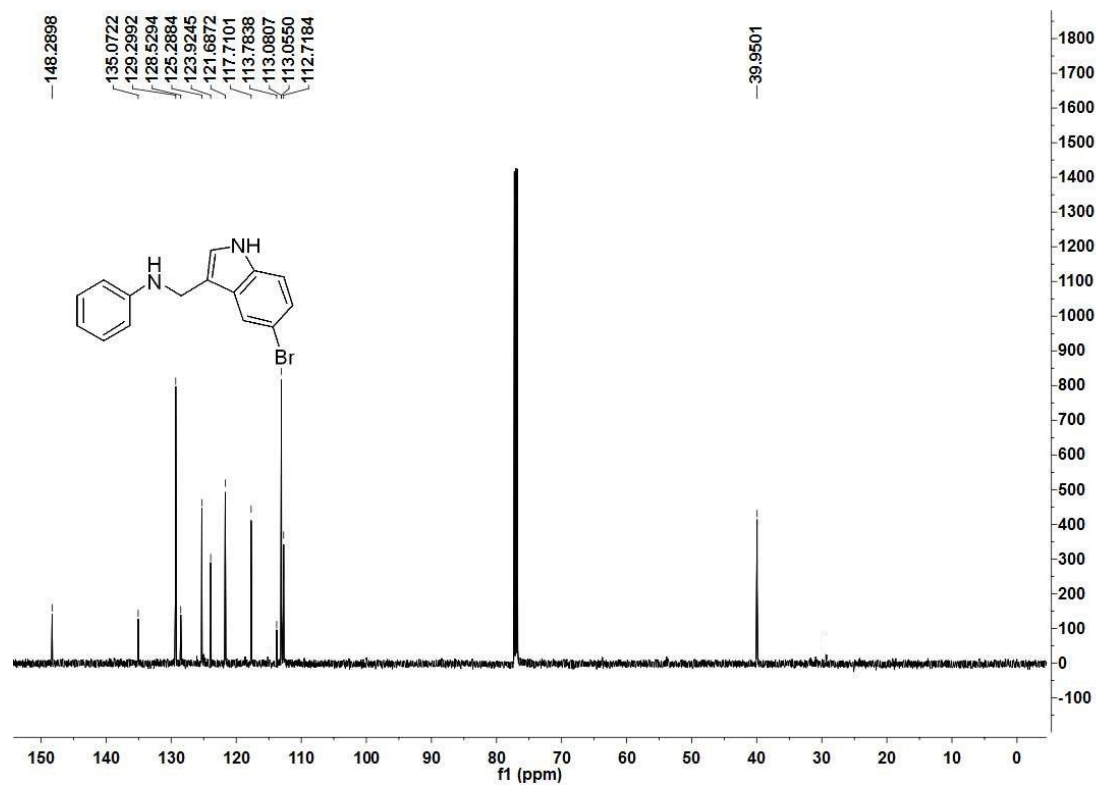
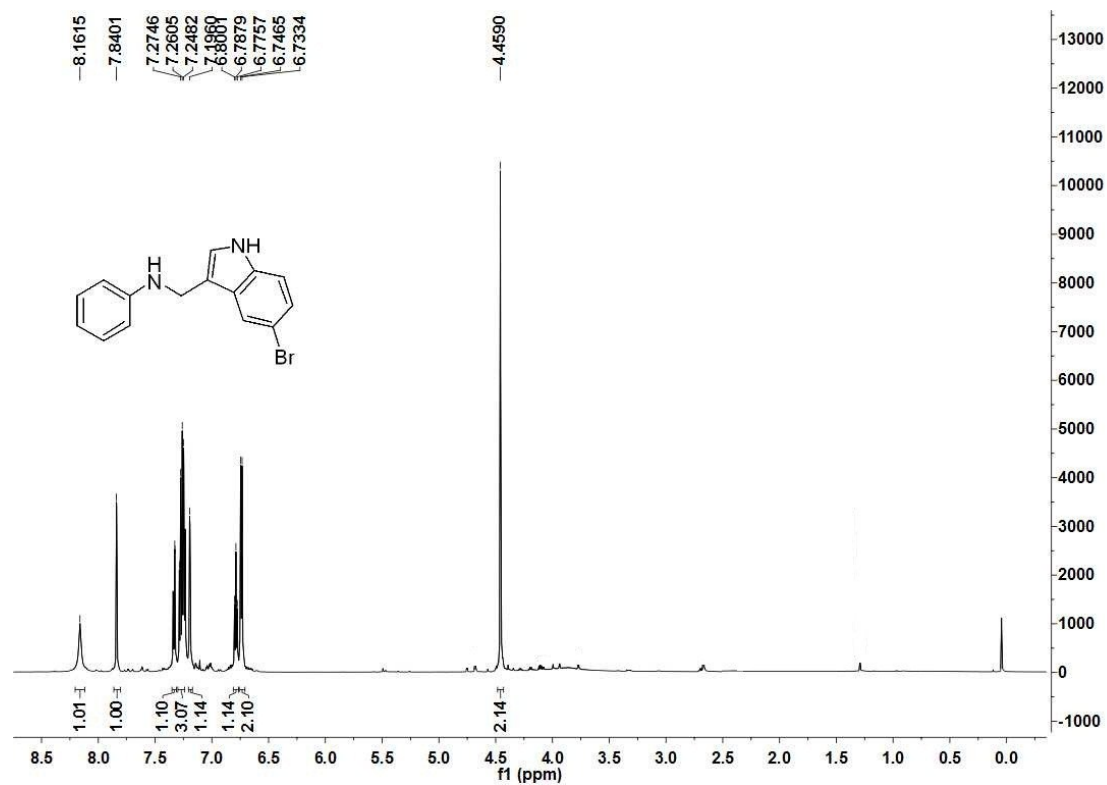
3cd



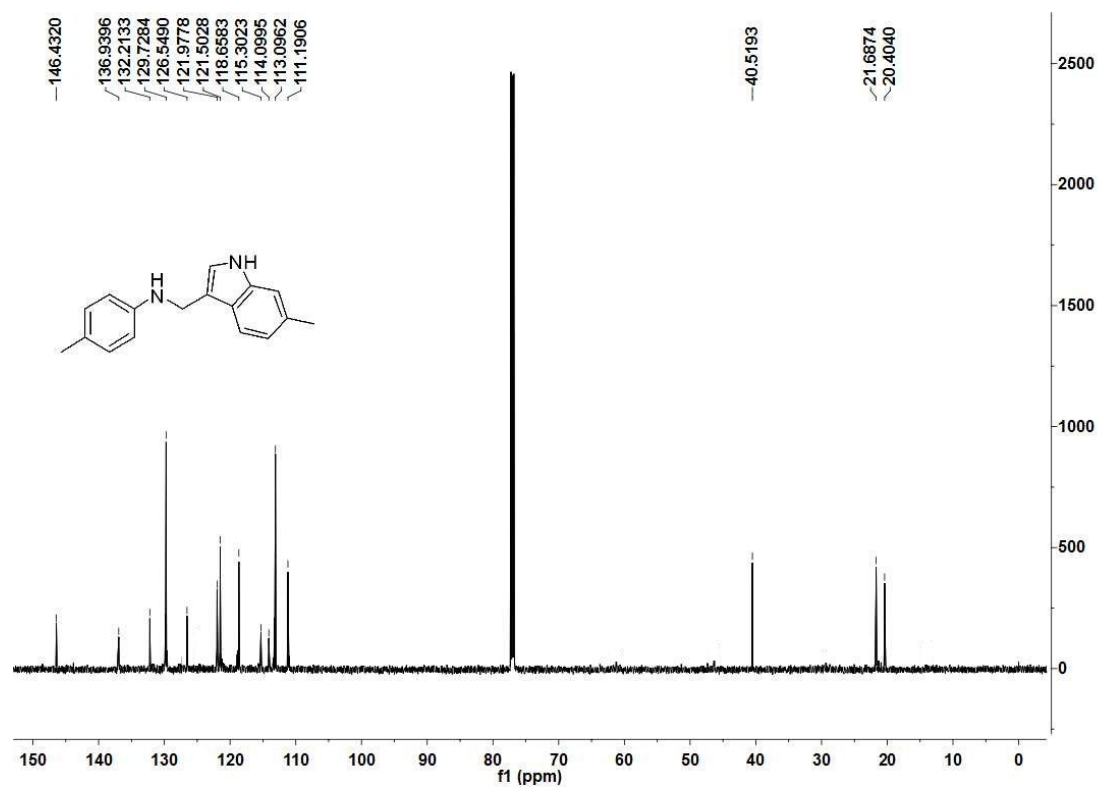
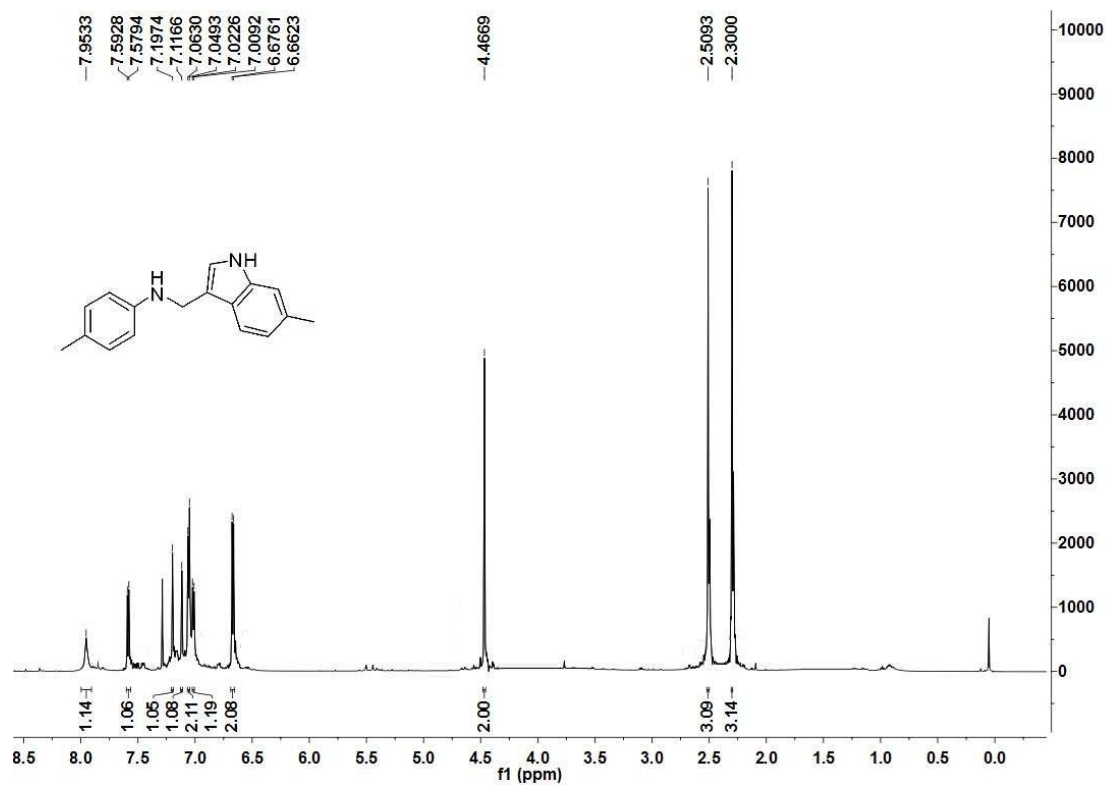
3ae



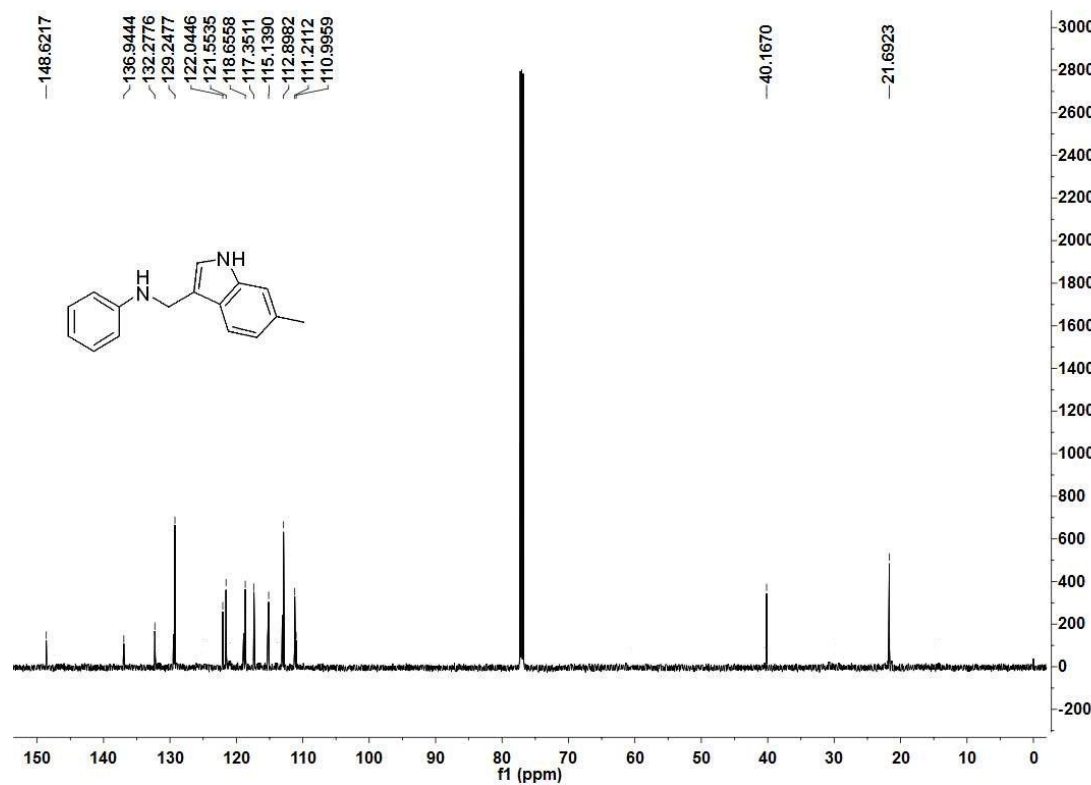
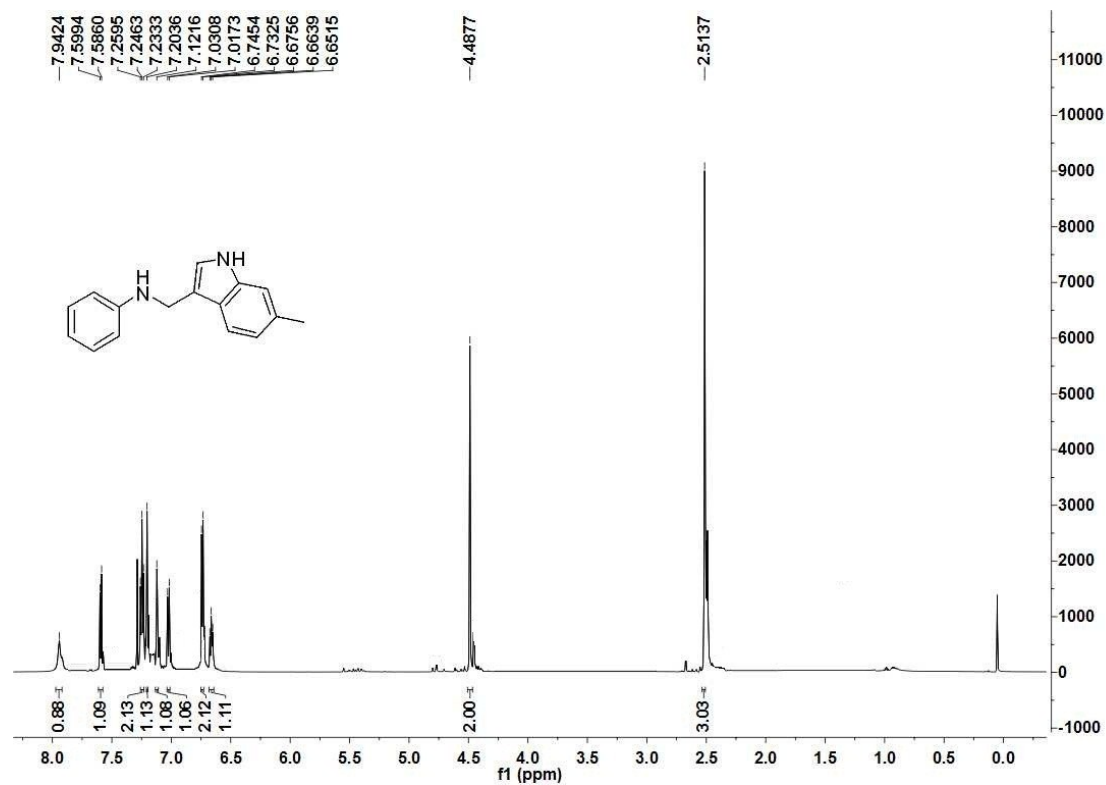
3be



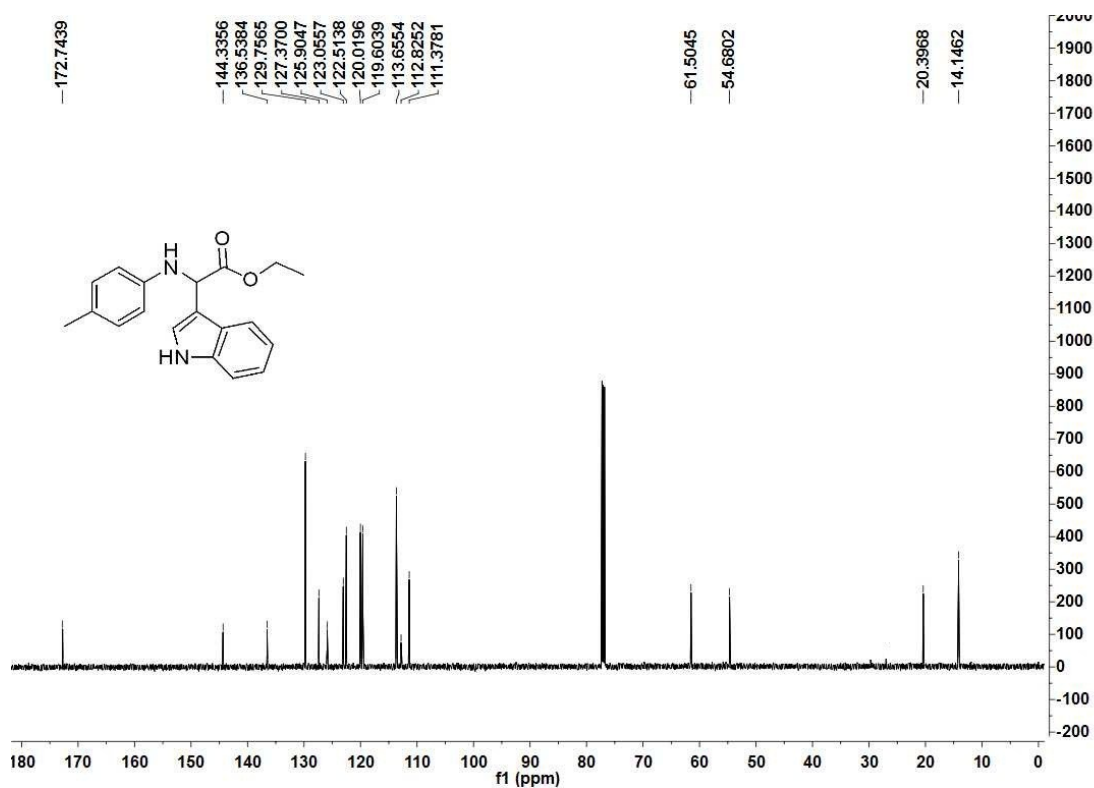
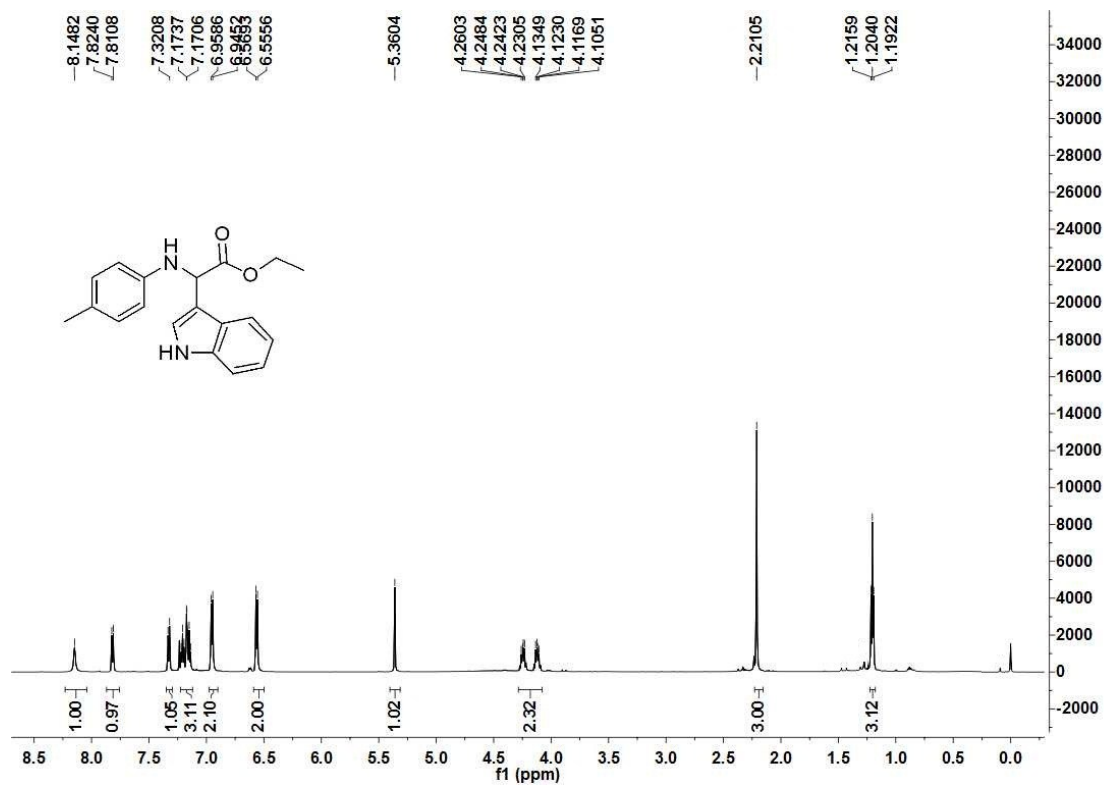
3af



3bf



4

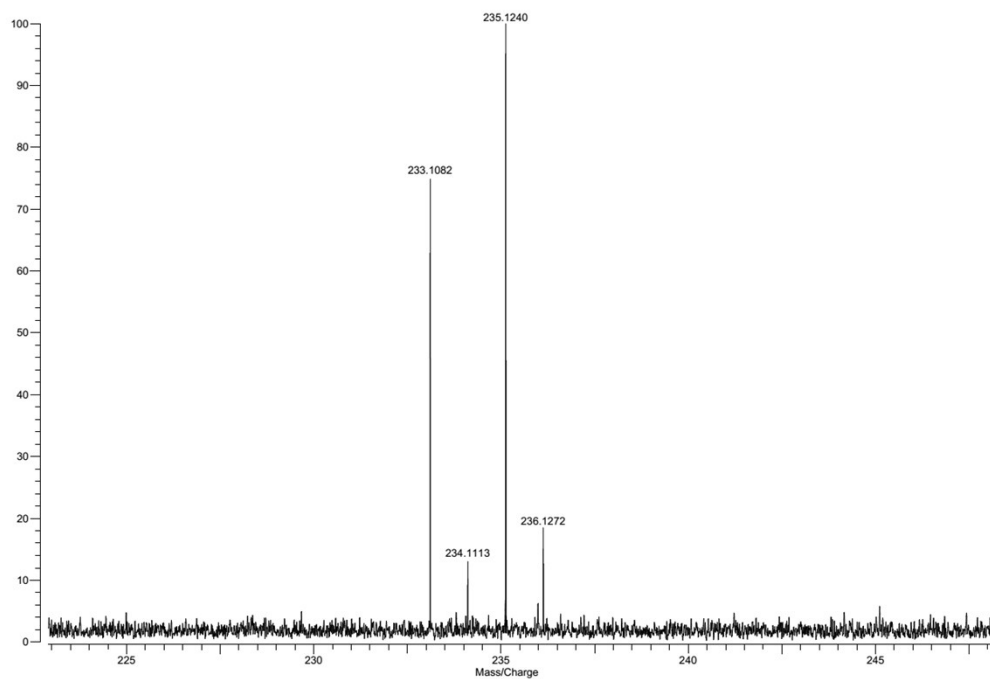


5. HRMS spectra of new compounds

3aa

Varian QFT-ESI
File: XY-7(2)_ESI.trans

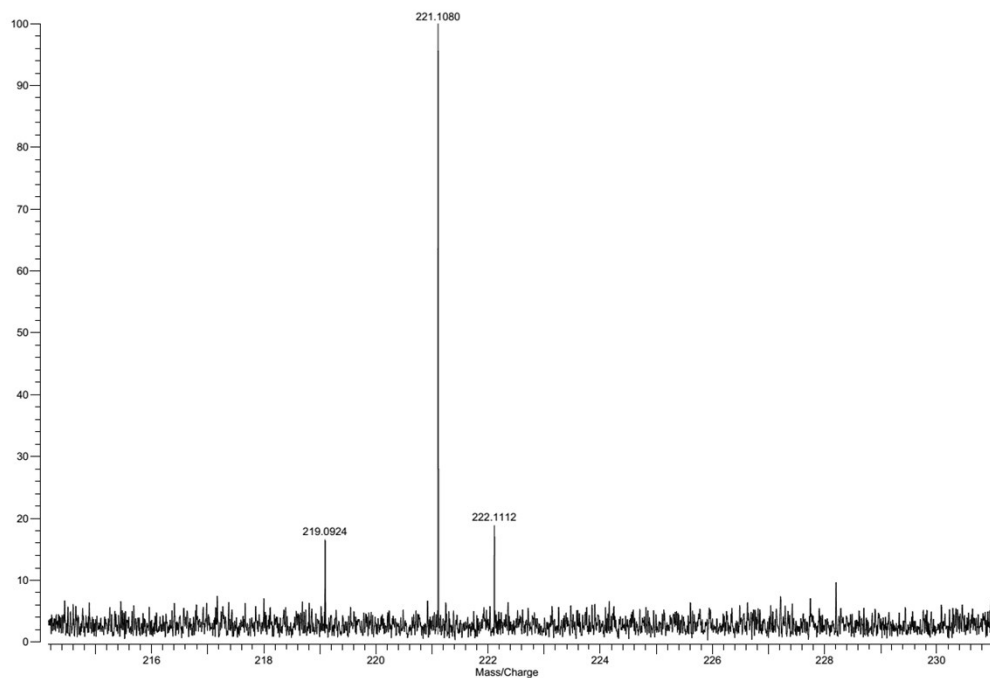
Mode: Negative
Scans: 2
Date: 02-JUN-2015
Time: 17:09:32
Scale: 41.1820



3ba

Varian QFT-ESI
File: XY-8_ESI.trans

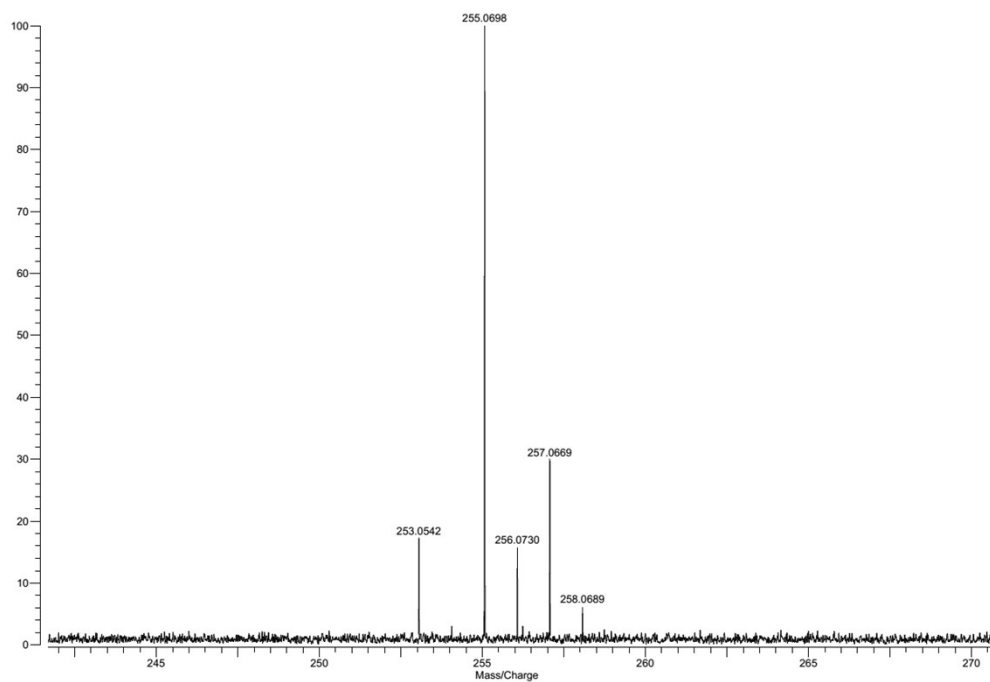
Mode: Negative Date: 02-JUN-2015
Scans: 5 Time: 16:53:29
Scale: 40.7318



3ca

Varian QFT-ESI
File: XY-8_ESI.trans

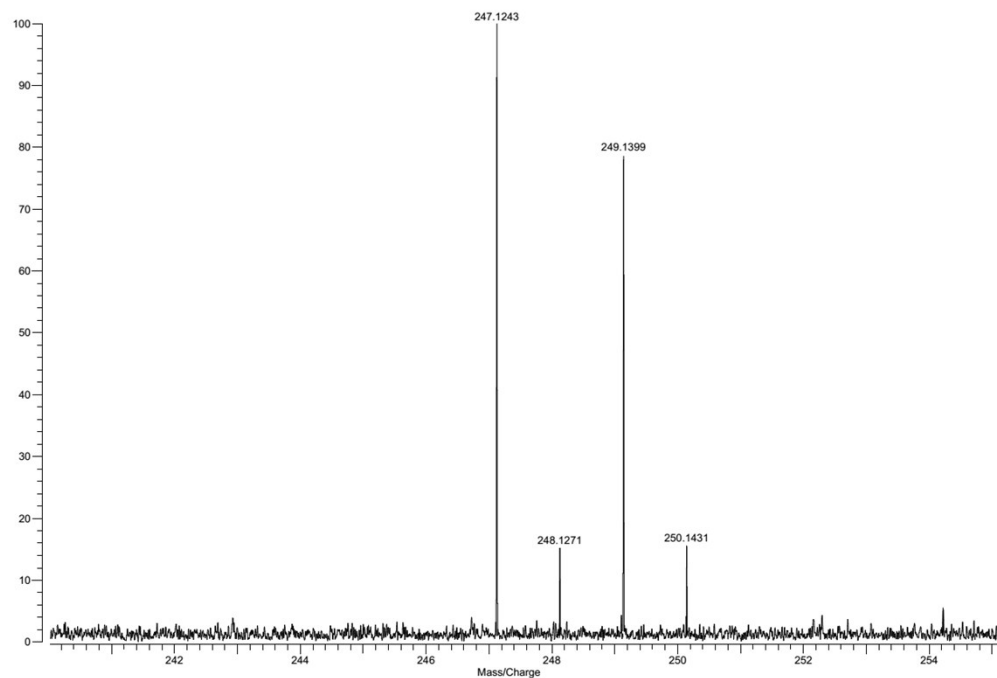
Mode: Negative Date: 02-JUN-2015
Scans: 1 Time: 20:35:09
Scale: 28.5574



3ab

Varian QFT-ESI
File: XY-10_ESI.trans

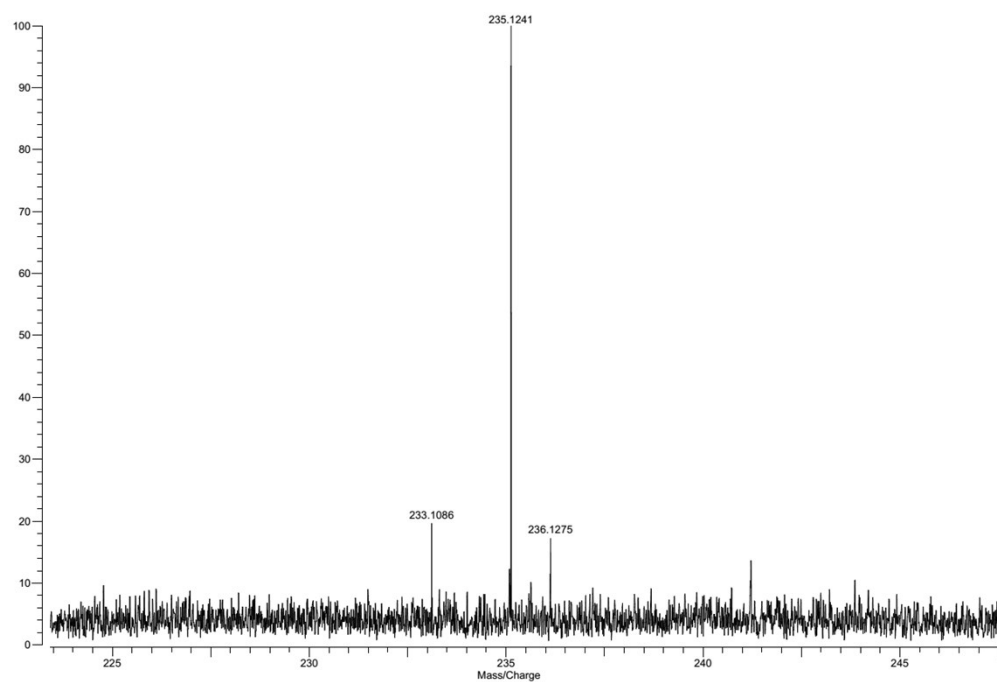
Mode: Negative Date: 02-JUN-2015
Scans: 1 Time: 17:27:45
Scale: 44.5356



3bb

Varian QFT-ESI
File: XY-9_ESI.trans

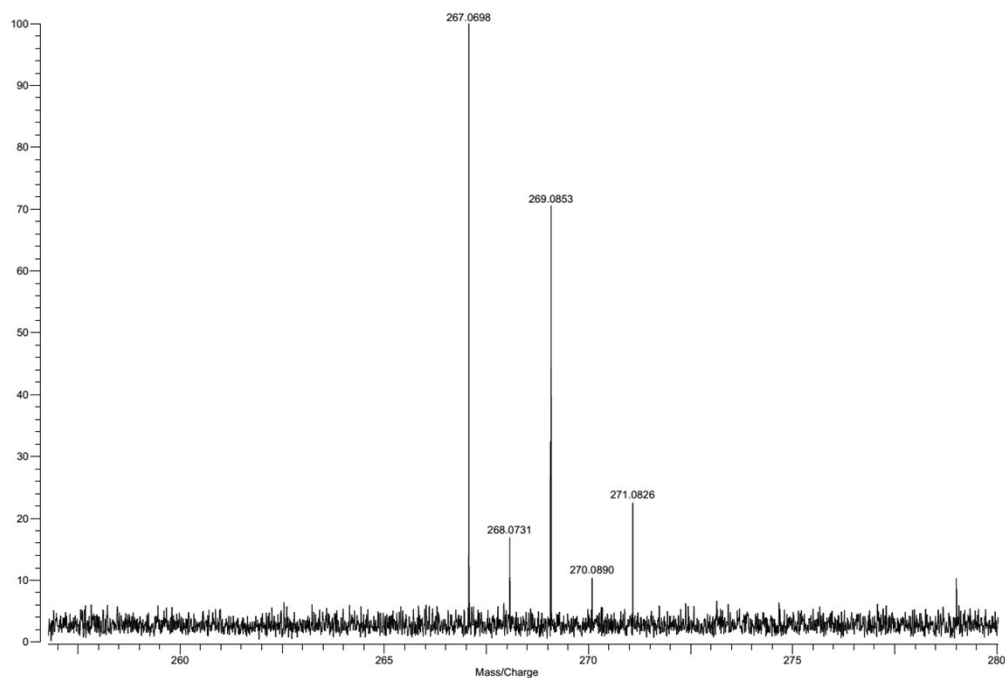
Mode: Negative Date: 02-JUN-2015
Scans: 5 Time: 17:21:52
Scale: 55.0144



3cb

Varian QFT-ESI
File: XY-11_ESI.trans

Mode: Negative Date: 02-JUN-2015
Scans: 1 Time: 17:41:23
Scale: 111.4810



3ac

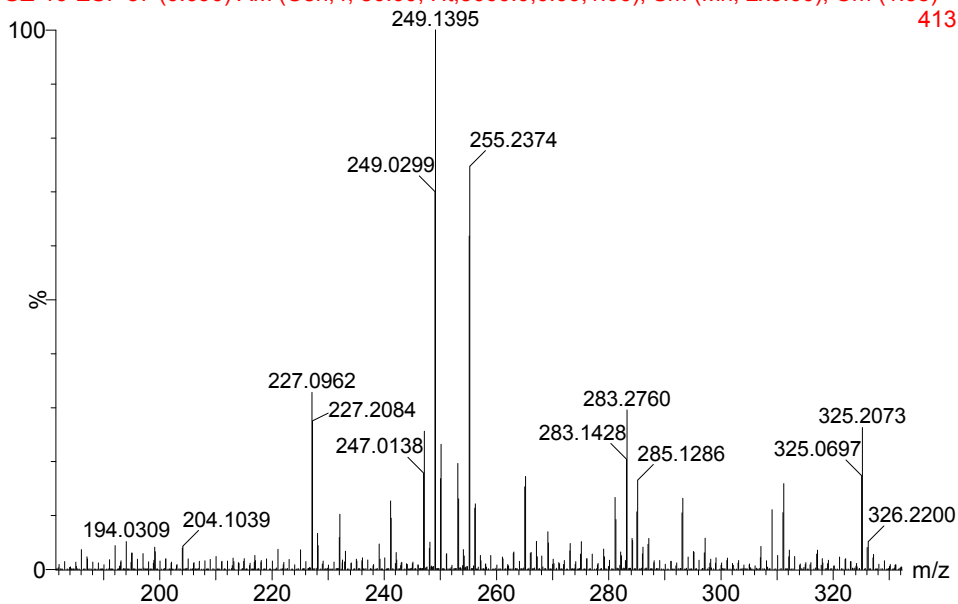
XY-2

15:17:33

25-Jan-2016

GZ-16-ESI- 37 (0.690) AM (Cen,4, 80.00, Ht,5000.0,0.00,1.00); Sm (Mn, 2x3.00); Cm (1:58)

413



3bc

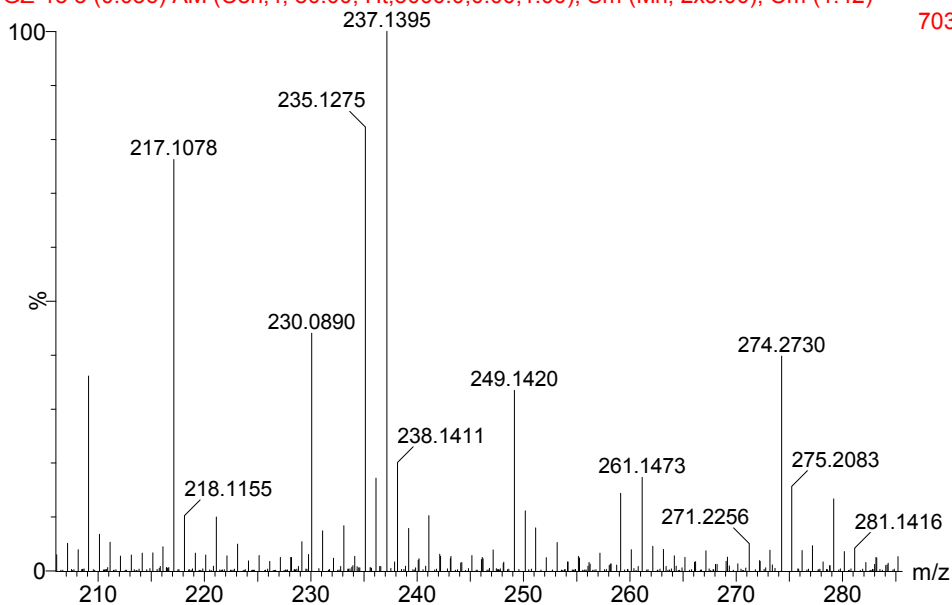
XY-1

09:27:16

25-Jan-2016

GZ-15 3 (0.056) AM (Cen,4, 80.00, Ht,5000.0,0.00,1.00); Sm (Mn, 2x3.00); Cm (1:42)

703



3cc

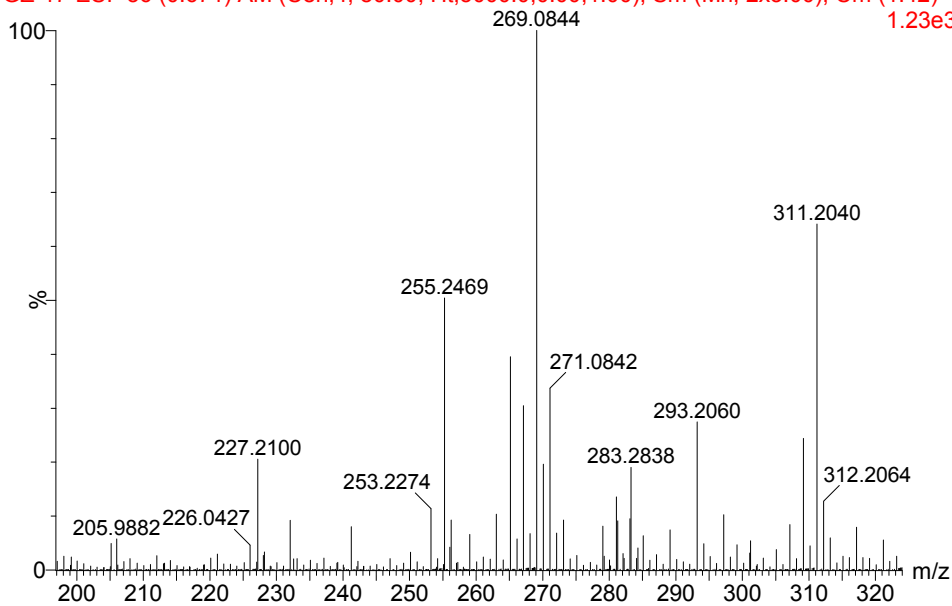
XY-3

15:23:19

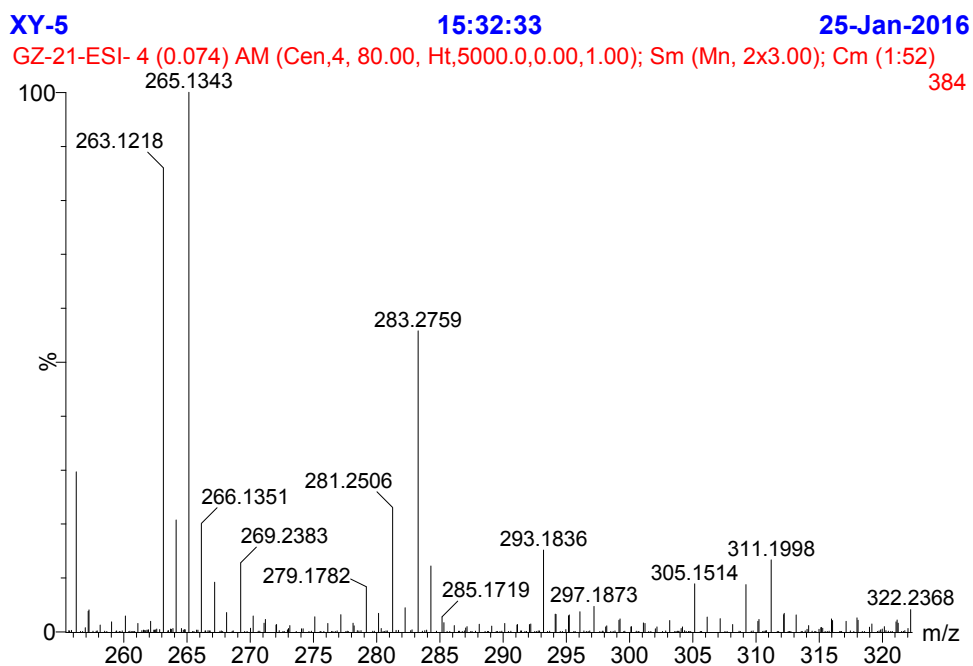
25-Jan-2016

GZ-17-ESI- 36 (0.671) AM (Cen,4, 80.00, Ht,5000.0,0.00,1.00); Sm (Mn, 2x3.00); Cm (1:42)

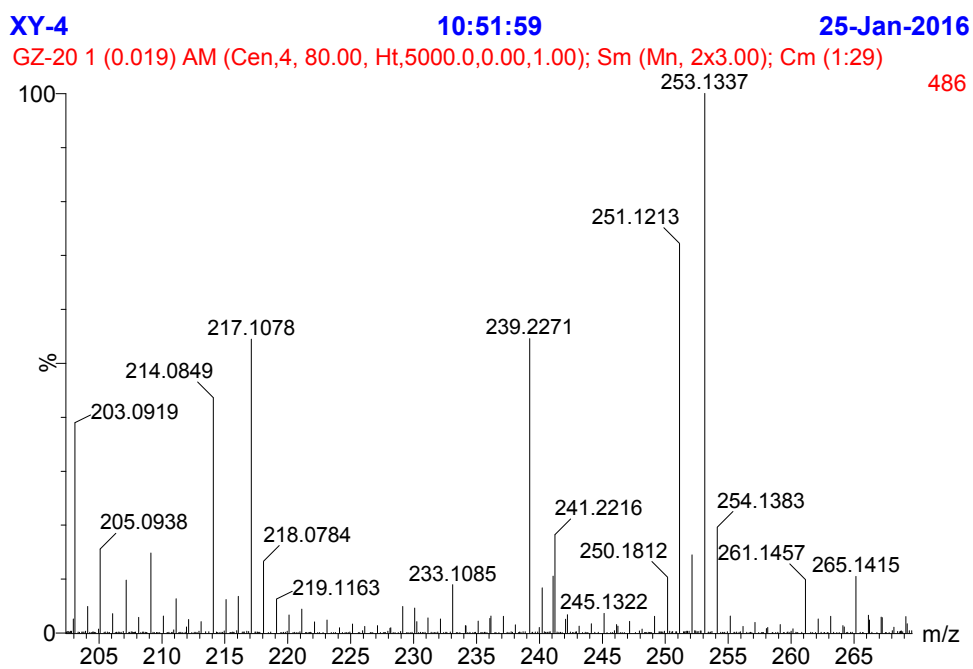
1.23e3



3ad



3bd



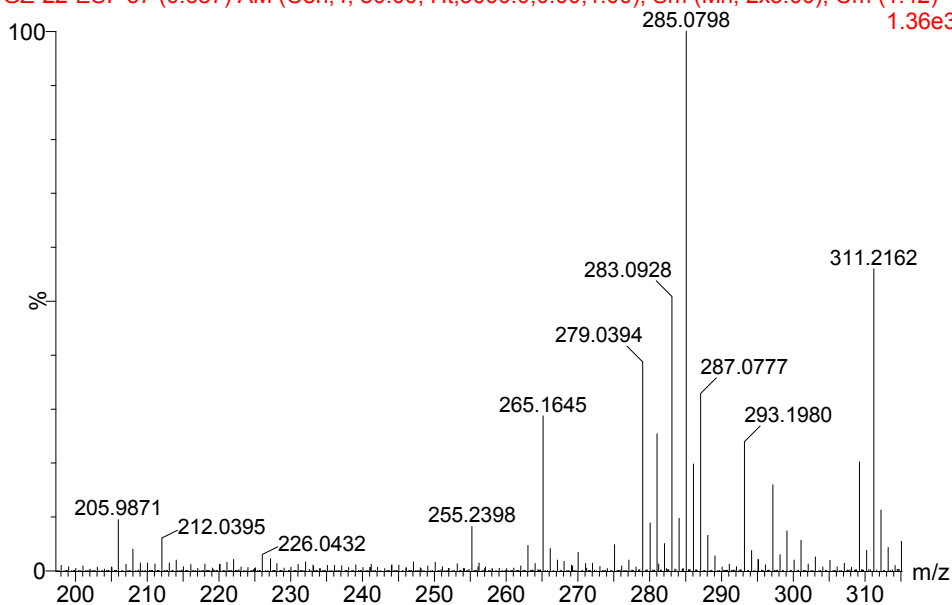
3cd

XY-6

16:18:29

25-Jan-2016

GZ-22-ESI- 37 (0.687) AM (Cen,4, 80.00, Ht,5000.0,0.00,1.00); Sm (Mn, 2x3.00); Cm (1:42)
1.36e3



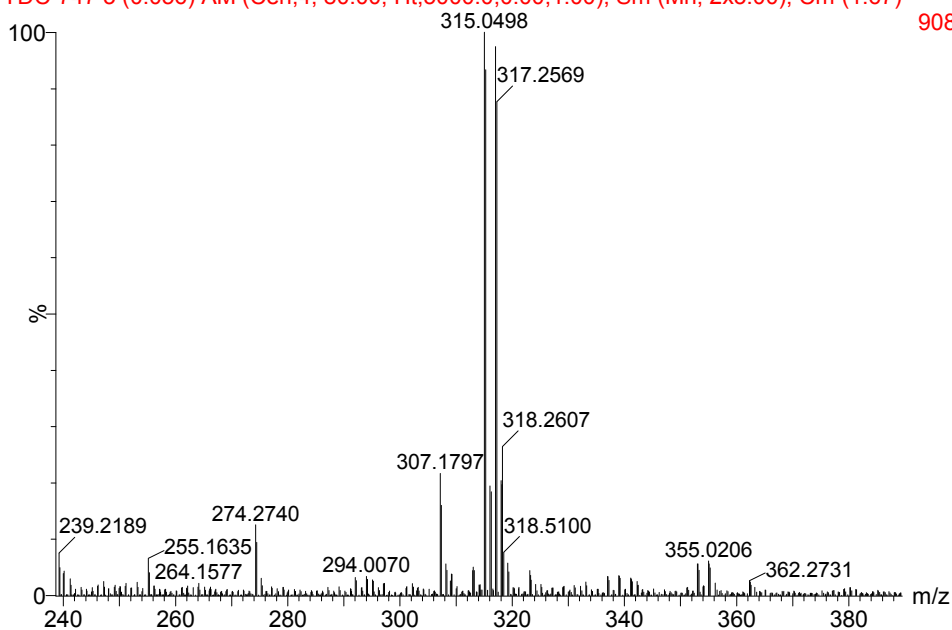
3ae

XY-2

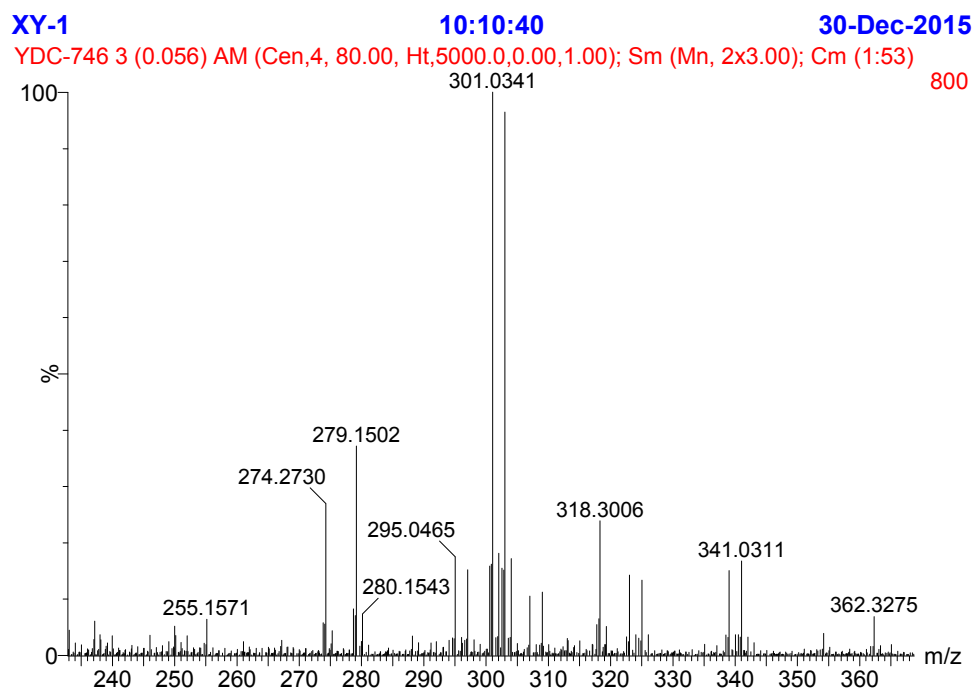
10:15:44

30-Dec-2015

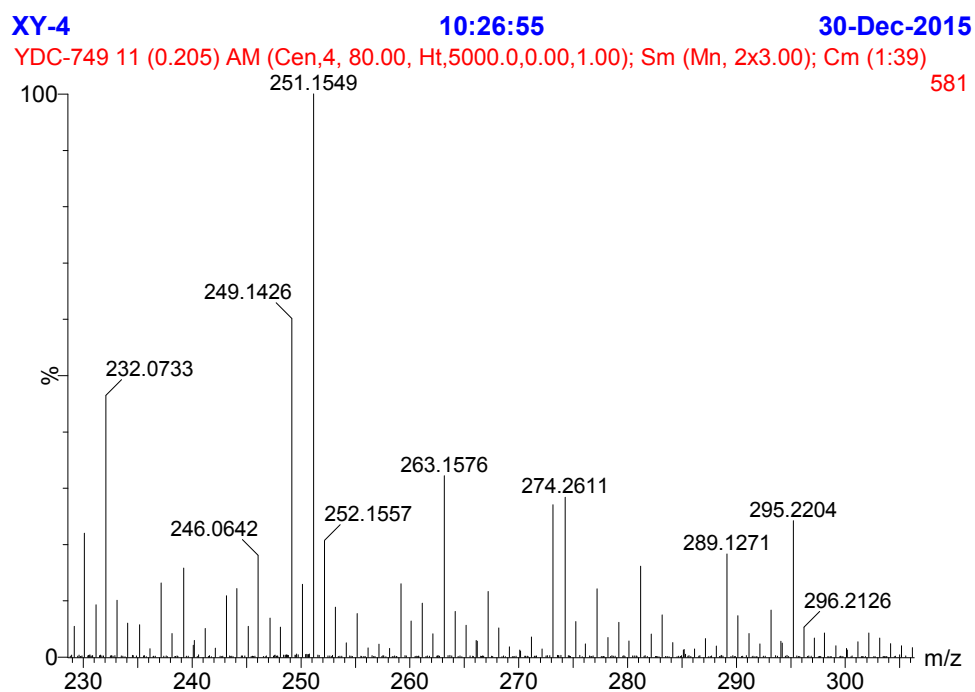
YDC-747 3 (0.056) AM (Cen,4, 80.00, Ht,5000.0,0.00,1.00); Sm (Mn, 2x3.00); Cm (1:67)
908



3be



3af



3bf

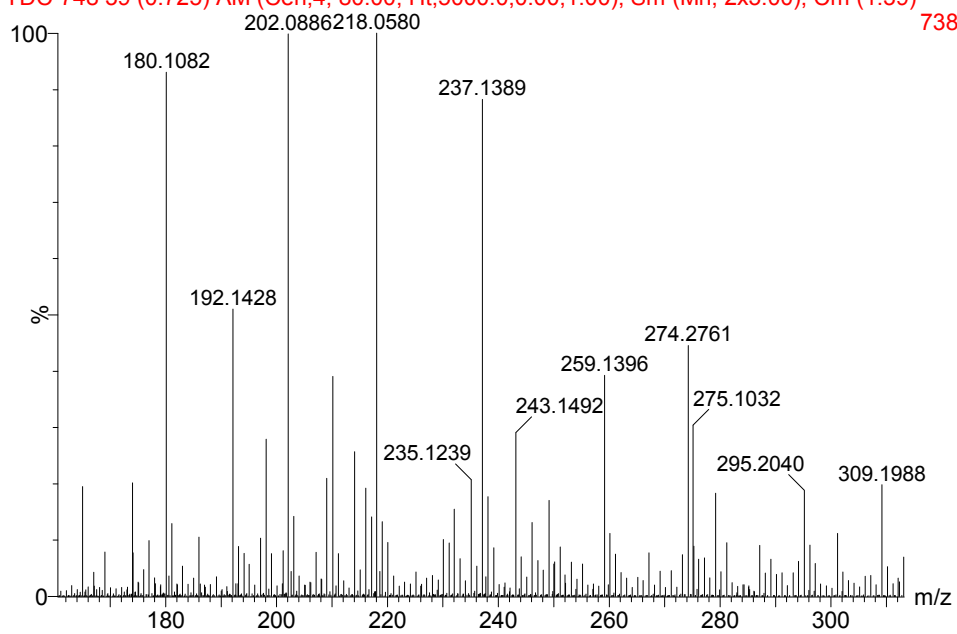
XY-3

10:30:06

30-Dec-2015

YDC-748 39 (0.725) AM (Cen,4, 80.00, Ht,5000.0,0.00,1.00); Sm (Mn, 2x3.00); Cm (1:39)

738



6. References

- 1 S.-Q. Zhu and M. Rueping, *Chem. Commun.*, 2012, **48**, 11960.