

Electronic Supplementary Information (ESI)

Biocatalytic Access to Nonracemic γ -Oxo Esters *via* Stereoselective Reduction Using Ene-Reductases

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1. General information

Unless stated otherwise, all solvents and commercially available reagents were used as purchased. Cyclohexane was distilled prior to use. Anhydrous toluene, dichloromethane and tetrahydrofuran were distilled over sodium, calcium hydride and sodium/potassium alloy, respectively, and stored under nitrogen on 4 Å molecular sieves.

Melting points were recorded on a Büchi M-565 melting point apparatus and are uncorrected. Nuclear magnetic resonance (NMR) spectra were recorded on a Bruker Avance III 300 MHz NMR (75.47 MHz for ^{13}C) with chemical shifts reported relative to TMS ($\delta = 0.00$ ppm) or a Bruker Avance 500 (125.78 MHz for ^{13}C) or Bruker Avance 400 (100.62 MHz for ^{13}C) using the residual solvent as internal standard (^1H : δ 7.26 ppm, $^{13}\text{C}\{^1\text{H}\}$: δ 77.16 ppm for CDCl_3 , ^1H : δ 2.50 ppm, $^{13}\text{C}\{^1\text{H}\}$: δ 39.52 ppm for DMSO- d_6). Chemical shifts (δ) are given in ppm and coupling constants (J) are quoted in Hertz (Hz). Multiplicities are described as s (singlet), d (doublet), t (triplet), q (quartet), quint (quintet), sex (sextet), sep (septet), br (broad singlet) and m (multiplet) or combinations thereof. Infrared (IR) spectra were recorded neat using a Shimadzu FTIR-8400s spectrophotometer and wavelengths are reported in cm^{-1} . Electrospray Ionization (ESI) high-resolution mass spectrometry (HRMS) was carried out using a Bruker microTOF-Q instrument in positive ion mode (capillary potential of 4500 V). Flash chromatography was performed on Silicycle Silia-P Flash Silica Gel (particle size 40-63 μm , pore diameter 60 Å) using the indicated eluent. Thin Layer Chromatography (TLC) was performed using TLC plates from Merck (Kieselgel 60 F₂₅₄ neutral, on aluminium sheets with fluorescence indicator).

2. Access to enzymes

2.1. List of protein accession numbers (Uniprot)

Enzyme	Protein accession number
OYE1	Q02899
OYE2	Q03558
OYE3	P41816
NCR	Q5NLA1
YqjM	P54550
OPR1	Q9XG54
OPR3	Q9FEW9
XenA	Q88NF7
EBP1	P43084

2.2. Preparation of biocatalysts: typical procedure

Synthetic genes and *E. coli* BL21 DE3 cells were obtained from Invitrogen. NEB5 α cells were obtained from New England Biolabs. Restriction enzymes, buffers and ligases were obtained from Thermo Fisher Scientific. For DNA extraction, the QIAGEN QIAquick gel extraction kit was used. Plasmid preparation was

performed with the QIAGEN QIAprep Spin Miniprep Kit. For enzyme purification, a HisTrap FF column (GE Healthcare, 5 mL column volume) was employed. Desalination was achieved by passing the purified enzyme through a PD-10 column (GE Healthcare).

2.2.1 Cloning

The synthetic genes (plasmid) were cloned into a pET28a(+)-vector, followed by transformation into NEB5 α cells. Six overnight cultures (ONCs) were prepared and the plasmid was isolated the following day (QIAGEN). Sequencing showed successful incorporation of the proper gene into the vector. Agarose-gel confirmed the insertion of the desired DNA-fragment.

2.2.2 Transformation of plasmids

To 100 μ L of cell suspension (*E. coli* BL21 DE3) in 1.5 mL Eppendorf tubes, 10 μ L of plasmid were added and incubated on ice for 30 min. The samples were then placed in a thermoshaker at 42 °C for 10 s. Afterwards, 250 μ L of LB-medium were added and the cells were incubated in the thermomixer for 1 h at 37 °C and 300 rpm. 100 μ L of the resulting product were plated on kanamycin-containing agar plates.

2.2.3 Overexpression

Colonies obtained from the transformation step were used to prepare overnight cultures (10 mL LB-medium, 50 μ g/mL kanamycin). They were placed in the shaker overnight (37 °C, 120 rpm) and used on the next day to inoculate the main cultures (330 mL LB-medium per shaking flask; 3.3 mL of the ONCs, 50 μ g/mL kanamycin). The main cultures were placed in the shaker at 37 °C and 140 rpm until an OD₆₀₀ of 0.6 was reached. Then, IPTG (0.2 mM) was used for induction. The cultures were again placed in the shaker overnight at 37 °C and 140 rpm. On the next day, the cells were harvested by centrifugation (4000 rpm, 20 min, 4 °C) and the supernatant was discarded. The pellet was resuspended with cold 0.9% NaCl solution and centrifuged again (4000 rpm, 20 min, 4°C). The supernatant was discarded and the pellets were stored (-20 °C freezer) until purification was performed.

2.2.4 Enzyme purification

The pellets obtained from the overexpression were all treated as follows. Lysis buffer (Tris-HCl buffer, 50 mM, pH 7.5) was added to resuspend the pellet. The cells were then ultrasonicated (40% amplitude, 1 s pulse on, 4 s pulse off, total pulse time: 5 min), centrifuged (14000 rpm, 30 min, 4 °C). The supernatant was filtered through a 0.45 μ m syringe filter and transferred into a 50 mL Sarstedt tube. Purification was performed by Ni-affinity chromatography. Wash buffer: TrisHCl (50 mM, 25 mM imidazole and 300 mM

NaCl), elution buffer: TrisHCl (50 mM, 250 mM imidazole, 300 mM NaCl). Following buffer exchange (lysis buffer), the enzyme concentration was determined by Bradford-assay. The enzymes were divided into aliquots of 800 μ L and stored at -20 °C.

3. General synthetic procedures

3.1. Synthesis of substrates

Note: some substrates were found to be volatile and/or unstable over time (polymerization).

Procedure A. Wittig-Horner reaction with diketones^[1]

To a solution of (alkoxycarbonylmethyl)triphenylphosphonium bromide (20 mmol, 1 equiv) in water (200 mL) was added NaOH (22 mmol, 1.1 equiv) precipitating the phosphorane as white solid. The suspension was extracted twice with CH_2Cl_2 , dried over Na_2SO_4 and concentrated *in vacuo*. The resulting white powder was redissolved in CH_2Cl_2 (100 mL) and the diketone (20 mmol, 1 equiv) was added. The mixture was stirred at room temperature for 1-5 days, concentrated *in vacuo* and subjected to purification by column chromatography on silica gel.

Procedure B. Isomerization of *exo*- to *endocyclic* alkenes^[2]

To a solution of substrate (3 mmol, 1 equiv) in cyclohexane (30 mL) was added DBU (0.1-0.4 equiv). The solution was refluxed for 3-24 hours (until TLC indicated full consumption of starting material) and then cooled to room temperature. The product was washed with saturated NH_4Cl solution, dried over Na_2SO_4 and concentrated *in vacuo* yielding a yellow oil after purification by column chromatography on silica gel.

Procedure C. γ -Oxidation of α,β -unsaturated esters^[3]

To a solution of α,β -unsaturated ester (4 mmol, 1 equiv) in CH_2Cl_2 (7 mL) cooled on an ice bath was added dropwise over 30 minutes a CrO_3 solution (11 mmol, 2.75 equiv) in acetic acid (2.6 mL) and acetic anhydride (1.25 mL). The solution stirred at room temperature for 90 minutes and then cautiously neutralized with concentrated KOH solution. The product was extracted with CH_2Cl_2 , washed with water and concentrated *in vacuo* yielding a colorless oil after purification by column chromatography on silica gel.

3.2. Synthesis of product standards

Procedure D. Hydrogenation of activated alkenes

To a solution of alkene (1 mmol) in ethyl acetate (10 mL) was added 10 w% Pd/C (25% w/w to substrate). The mixture was stirred under atmospheric pressure of H_2 (balloon) at room temperature for 1-2 days. The mixture was then filtered over Celite and concentrated *in vacuo* yielding the product. If necessary, the product was purified by column chromatography on silica gel.

Procedure E. Basic hydrolysis of esters

To a solution of ester (5 mmol, 1 equiv) in a water/methanol mixture (10:1, 5 mL) was added NaOH (10 mmol, 2 equiv). The solution was stirred at room temperature for 2-24 hours before acidifying to pH = 1 with 6 M HCl. The mixture was extracted with ethyl acetate twice, dried over Na₂SO₄ and concentrated *in vacuo* to give the pure product.

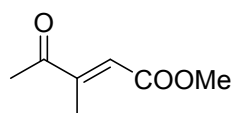
Procedure F. Esterification of carboxylic acids

To a solution of carboxylic acid (1 mmol) in ethanol or methanol (2 mL) was added a solution of thionyl chloride (1 mmol) in toluene (0.1 mL) dropwise over 30 minutes at 0-5 °C. The solution was then stirred at room temperature for 5 hours and concentrated *in vacuo*. The crude product was taken up in CH₂Cl₂, washed twice with saturated NaHCO₃ solution, dried over Na₂SO₄ and concentrated *in vacuo* to give the ethyl or methyl ester, respectively.

Ethyl esters were generally prepared by hydrogenation of corresponding alkenes on Pd/C (Procedure D). Methyl esters were generally prepared by esterification from corresponding carboxylic acids (Procedure F). Carboxylic acids were generally prepared by saponification of the ethyl esters (Procedure E). Racemic **9b** was purchased from Sigma Aldrich. All product standards are known compounds: **1b**,^[4] **2b**,^[5] **3b**,^[8] **4b**,^[9] **5b**,^[6] **6b**,^[7] **7b**,^[10] **8b**,^[11] **9b**,^[12] **10b**,^[13] **11b**,^[8] **12b**,^[11] **13b**.^[14]

4. Characterization of substrates

Methyl 3-methyl-4-oxopent-2-enoate (*E*)- and (*Z*)-**1a**^[15]



Prepared according to Procedure A from diacetyl (878 μ L, 10 mmol) and methoxycarbonylmethyltriphenylphosphonium bromide (4.17 g, 10 mmol). Purification: column chromatography on silica gel (cyclohexane/ethyl acetate, 19/1); diastereoisomers could be separated [R_f = 0.22 (*E*) and R_f = 0.14 (*Z*)] and

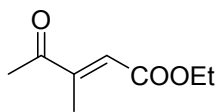
were isolated as colorless oils. Yield: 0.457 g, 0.36 mmol, 36% of combined (*E*) and (*Z*) isomers, 2.5:1 ratio. Note: The *Z* isomer isomerizes slowly to the *E*.

(*E*)-**1a**: ¹H-NMR (CDCl₃, 500 MHz) δ 2.12 (s, 3H), 2.31 (s, 3H), 3.70 (s, 3H), 6.51 (s, 1H) ppm; ¹³C{¹H}-NMR (CDCl₃, 125 MHz) δ 13.1 (CH₃), 26.2 (CH₃), 51.8 (CH₃), 126.0 (CH), 150.7 (C), 166.6 (C), 199.8 (C) ppm; IR (neat): ν_{max} (cm⁻¹) = 1955 (w), 1724 (s), 1682 (s), 1437 (m), 1364 (m), 1246 (s), 1196 (s), 1180 (s), 1105 (m), 1040 (m), 876 (m);

(*Z*)-**1a**: ¹H-NMR (CDCl₃, 500 MHz) δ 1.96 (s, 3H), 2.32 (s, 3H), 3.66 (s, 3H), 5.66 (s, 1H) ppm; ¹³C{¹H}-NMR (CDCl₃, 125 MHz) δ 20.3 (CH₃), 28.6 (CH₃), 51.8 (CH₃), 116.6 (CH), 157.7 (C), 165.7 (C), 206.3 (C) ppm; IR (neat): ν_{max} (cm⁻¹) = 2953 (w), 1705 (s), 1643 (m), 1437 (m), 1362 (m), 1348 (m), 1248 (s), 1161 (s), 1011 (m), 858 (m);

HRMS (ESI): m/z calculated for C₇H₁₀NaO₃ [M+Na]⁺ 165.0523, found 165.0522.

Ethyl 3-methyl-4-oxopent-2-enoate (*E*)- and (*Z*)-2a^[16]



Prepared according to Procedure A from diacetyl (1.78 mL, 20 mmol) and ethoxycarbonylmethyltriphenylphosphonium bromide (8.59 g, 20 mmol). Purification: column chromatography on silica gel (cyclohexane/ethyl acetate, 15/1); diastereoisomers could be separated [$R_f = 0.22$ (*E*) and $R_f = 0.14$ (*Z*)] and

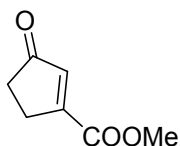
were isolated as colorless oils. Yield: 2.41 g, 15.6 mmol, 78% of combined (*E*) and (*Z*) isomers, 2.5:1 ratio. Note: The *Z* isomer isomerizes slowly to the *E*.

(*E*)-2a: $^1\text{H-NMR}$ (CDCl_3 , 500 MHz) δ 1.31 (t, $J = 7.0$ Hz, 3H), 2.19 (s, 3H), 2.37 (s, 3H), 4.23 (q, $J = 7.0$ Hz, 2H), 6.56 (s, 1H); $^{13}\text{C}\{^1\text{H}\}\text{-NMR}$ (CDCl_3 , 125 MHz) δ 13.2 (CH_3), 14.3 (CH_3), 26.3 (CH_3), 60.9 (CH_2), 126.7 (CH), 150.5 (C), 166.3 (C), 200.0 (C) ppm; IR (neat): ν_{max} (cm^{-1}) = 2981 (w), 1720 (s), 1686 (s), 1366 (m), 1246 (s), 1188 (s), 1103 (s), 1045 (m), 633 (m), 536 (s), 498 (s);

(*Z*)-2a: $^1\text{H-NMR}$ (CDCl_3 , 500 MHz) δ 1.25 (t, $J = 7.0$ Hz, 3H), 1.97 (s, 3H), 2.34 (s, 3H), 4.14 (q, $J = 7.0$ Hz, 2H), 5.67 (s, 1H) ppm; $^{13}\text{C}\{^1\text{H}\}\text{-NMR}$ (CDCl_3 , 125 MHz) δ 14.3 (CH_3), 20.4 (CH_3), 28.8 (CH_3), 60.8 (CH_2), 117.1 (CH), 157.4 (C), 166.3 (C), 200.0 (C) ppm; IR (neat): ν_{max} (cm^{-1}) = 2982 (w), 1720 (s), 1686 (s), 1366 (m), 1246 (s), 1188 (s), 1045 (m), 633 (m), 536 (s), 498 (s);

HRMS (ESI): m/z calculated for $\text{C}_8\text{H}_{12}\text{NaO}_3$ [$\text{M}+\text{Na}$] $^+$ 179.0679, found 179.0684.

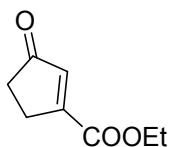
Methyl 3-oxocyclopent-1-ene-1-carboxylate 3a^[20]



Prepared according to procedure C from methyl cyclopent-1-ene-1-carboxylate (0.49 mL, 4 mmol). Purification by column chromatography on silica gel (pentane/diethyl ether, 2/1). Isolated as a pale yellow solid. Yield: 230 mg, 1.6 mmol, 41% (contains traces of AcOH and Ac_2O).

m.p.: 35.0-38.5 °C. $^1\text{H-NMR}$ (CDCl_3 , 500 MHz) δ 2.42-2.47 (m, 2H), 2.74-2.79 (m, 2H), 3.78 (s, 3H), 6.65 (s, 1H) ppm; $^{13}\text{C}\{^1\text{H}\}\text{-NMR}$ (CDCl_3 , 125 MHz) δ 27.4 (CH_2), 35.5 (CH_2), 52.5 (CH_3), 138.0 (CH), 163.9 (C), 164.7 (C), 209.2 (C) ppm; IR (neat): ν_{max} (cm^{-1}) = 2959 (w), 1709 (s), 1431 (m), 1219 (s), 1161 (s), 991 (m), 737 (m), 435 (m); HRMS (ESI): m/z calculated for $\text{C}_7\text{H}_8\text{NaO}_3$ [$\text{M}+\text{Na}$] $^+$ 163.0371, found 163.0364.

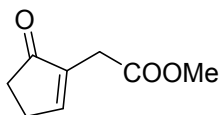
Ethyl 3-oxocyclopent-1-ene-1-carboxylate 4a^[21]



Prepared according to procedure C from ethyl cyclopent-1-ene-1-carboxylate (530 mg, 3.8 mmol). Purification by column chromatography on silica gel (pentane/diethyl ether, 1/2). Isolated as a pale yellow oil. Yield: 153 mg, 1 mmol, 26%.

$^1\text{H-NMR}$ (CDCl_3 , 500 MHz) δ 1.33 (t, $J = 7.0$ Hz, 3H), 2.50-2.55 (m, 2H), 2.82-2.87 (m, 2H), 4.30 (q, $J = 7.0$ Hz, 2H), 6.73 (t, $J = 2.0$ Hz, 1H) ppm; $^{13}\text{C}\{^1\text{H}\}\text{-NMR}$ (CDCl_3 , 125 MHz) δ 14.3 (CH_3), 27.5 (CH_2), 35.7 (CH_2), 61.8 (CH_2), 138.2 (CH), 164.5 (C), 209.4 (C) ppm; IR (neat): ν_{max} (cm^{-1}) = 2982 (w), 1713 (s), 1613 (w), 1213 (s), 1161 (s), 1070 (m), 1009 (m), 741 (m), 631 (m); HRMS (ESI): m/z calculated for $\text{C}_8\text{H}_{12}\text{O}_3$ [$\text{M}+\text{H}$] $^+$ 155.0704, found 155.0703.

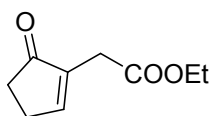
Methyl 2-(5-oxocyclopent-1-en-1-yl)acetate **5a**^[17]



To a solution of *exo*-**6a** (1.30 g, 7.75 mmol, 1 equiv) in methanol (16 mL) was added 0.5 mL 37% HCl. The mixture was refluxed for 18 hours and then cooled to room temperature. The mixture was neutralized by adding solid NaHCO₃, the salts were filtered off and the solution was concentrated *in vacuo* to give the product as a yellow oil (696 mg, 4.5 mmol, 58%).

¹H-NMR (CDCl₃, 500 MHz) δ 2.40-2.46 (m, 2H), 2.62-2.67 (m, 2H), 3.24 (s, 2H), 3.70 (s, 3H), 7.60 (quint, J = 1.0 Hz, 1H) ppm; ¹³C{¹H}-NMR (CDCl₃, 125 MHz) δ 27.0 (CH₂), 30.2 (CH₂), 34.1 (CH₂), 52.2 (CH₃), 139.0 (C), 160.9 (CH), 171.1 (C), 208.6 (C) ppm; IR (neat): ν_{max} (cm⁻¹) = 2953 (w), 1734 (s), 1697 (s), 1437 (m), 1258 (m), 1163 (m), 1001 (m), 791 (w), 633 (w), 492 (m); HRMS (ESI): *m/z* calculated for C₈H₁₀NaO₃ [M+Na]⁺ 177.0523, found 177.0522.

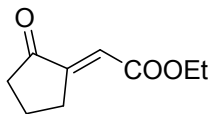
Ethyl 2-(5-oxocyclopent-1-en-1-yl)acetate *endo*-**6a**^[18]



To a solution of *exo*-**6a** (317 mg, 1.88 mmol, 1 equiv) in ethanol (4 mL) was added 0.125 mL of 37% HCl. The mixture was refluxed for 17 hours and then cooled to room temperature. The mixture was neutralized by adding solid NaHCO₃, the salts were filtered off and the solution was concentrated *in vacuo* to give the product as a dark brown oil (298 mg, 1.76 mmol, 94%).

¹H-NMR (CDCl₃, 500 MHz) δ 1.23 (t, J = 7.0 Hz, 3H), 2.40-2.48 (m, 2H), 2.62-2.68 (m, 2H), 3.22 (s, 2H), 4.16 (q, J = 7.0 Hz, 2H), 7.60 (quint, J = 1.5 Hz, 1H) ppm; ¹³C{¹H}-NMR (CDCl₃, 125 MHz) δ 14.2 (CH₃), 26.9 (CH₂), 30.4 (CH₂), 34.0 (CH₂), 61.0 (CH₂), 139.1 (C), 160.8 (CH), 170.6 (C), 208.6 (C) ppm; IR (neat): ν_{max} (cm⁻¹) = 2982 (w), 2928 (w), 1732 (s), 1697 (s), 1258 (m), 1159 (m), 1026 (m), 1026 (m), 789 (w), 498 (w); HRMS (ESI): *m/z* calculated for C₉H₁₃O₃ [M+H]⁺ 169.0860, found 169.0859.

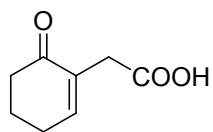
Ethyl (*E*)-2-(2-oxocyclopentylidene)acetate *exo*-**6a**^[19]



To a solution of 1-morpholinocyclopentene (1.7 mL, 10 mmol, 1 equiv) in cyclohexane (10 mL) was added ethyl glyoxylate (50% in toluene, 2 mL, 10 mmol, 1 equiv). The mixture was refluxed using a Dean-Stark apparatus for 23 hours. After cooling to room temperature 6 M HCl (6 mL) was added and the mixture was stirred for 5 hours at room temperature. The product was extracted with toluene (two times), dried over Na₂SO₄ and concentrated *in vacuo*. The product was isolated by column chromatography on silica gel (cyclohexane/ethyl acetate, 13/1, R_f = 0.24) as a brown oil (574 mg, 3.4 mmol, 34%).

¹H-NMR (CDCl₃, 500 MHz) δ 1.28 (t, J = 7.5 Hz, 3H), 2.00 (quint, J = 7.5 Hz, 2H), 2.39 (t, J = 7.5 Hz, 2H), 3.08 (td, J = 7.5, 3.0 Hz, 2H), 4.21 (q, J = 7.5 Hz, 2H), 6.48 (t, J = 3.0 Hz, 1H) ppm; ¹³C{¹H}-NMR (CDCl₃, 125 MHz) δ 14.3 (CH₃), 19.7 (CH₂), 29.5 (CH₂), 38.0 (CH₂), 60.9 (CH₂), 119.8 (CH), 150.9 (C), 166.5 (C), 207.5 (C) ppm; IR (neat): ν_{max} (cm⁻¹) = 2380 (w), 1709 (s), 1369 (m), 1346 (m), 1223 (m), 1202 (s), 1173 (m), 1022 (m), 525 (m); HRMS (ESI): *m/z* calculated for C₉H₁₃O₃ [M+H]⁺ 169.0860, found 169.0832.

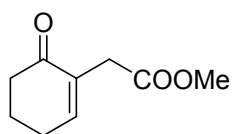
2-(6-Oxocyclohex-1-en-1-yl)acetic acid **7a**^[22]



Prepared according to Procedure E from *endo*-**9a** (60 mg, 0.33 mmol). Isolated as an off-white amorphous solid used without further purification. Yield: 45 mg, 0.3 mmol, 90%.

¹H-NMR (CDCl₃-DMSO-d₆, 500 MHz) δ 1.64 (quint, *J* = 6.5 Hz, 2H), 1.95-2.10 (m, 4H), 2.73 (s, 2H), 6.51 (t, *J* = 4.5 Hz, 1H) ppm; ¹³C{¹H}-NMR (CDCl₃/DMSO-d₆, 125 MHz) δ 22.1 (CH₂), 25.2 (CH₂), 34.7 (CH₂), 37.2 (CH₂), 132.9 (C), 147.6 (CH), 172.3 (C), 197.5 (C) ppm; IR (neat): ν_{max} (cm⁻¹) = 2933 (w), 1715 (s), 1695 (s), 1231 (m), 1180 (s); HRMS (ESI): *m/z* calculated for C₈H₁₀NaO₃ [M+Na]⁺ 177.0522, found 177.0529.

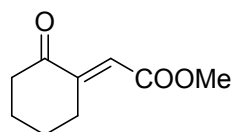
Methyl 2-(6-oxocyclohex-1-en-1-yl)acetate *endo*-**8a**^[22]



Prepared according to Procedure B from *exo*-**8a** (587 mg, 3 mmol) and DBU (179 μ L, 1.2 mmol). Purification: column chromatography on silica gel (cyclohexane/ethyl acetate, 9/1, *R_f* = 0.15). Isolated as a yellow oil. Yield: 297 mg, 1.77 mmol, 59%.

¹H-NMR (CDCl₃, 500 MHz) δ 2.03 (quint, *J* = 7.0 Hz, 2H), 2.41 (q, *J* = 4.5 Hz, 2H), 2.47 (t, *J* = 7.0 Hz, 2H), 3.20 (s, 2H), 3.67 (s, 3H), 6.88 (t, *J* = 4.0 Hz, 1H) ppm; ¹³C{¹H}-NMR (CDCl₃, 125 MHz) δ 23.0 (CH₂), 26.2 (CH₂), 35.3 (CH₂), 38.1 (CH₂), 52.1 (CH₃), 133.6 (C), 148.6 (CH), 172.0 (C), 198.5 (C) ppm; IR (neat): ν_{max} (cm⁻¹) = 2951 (w), 1736 (s), 1670 (s), 1435 (m), 1339 (m), 1258 (m), 1169 (s), 987 (w), 601 (m), 498 (m); HRMS (ESI): *m/z* calculated for C₉H₁₂NaO₃ [M+Na]⁺ 191.0679, found 191.0679.

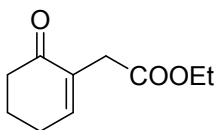
Methyl (*E*)-2-(2-oxocyclohexylidene)acetate *exo*-**8a**^[23]



Prepared according to Procedure A from 1,2-cyclohexanedione (1.12 g, 10 mmol) and methoxycarbonylmethyltriphenylphosphonium bromide (4.16 g, 10 mmol). Purification: column chromatography on silica gel (cyclohexane/ethyl acetate, 19/1, *R_f* = 0.17). Isolated as a yellow oil. Yield: 0.91 g, 5.4 mmol, 54%.

¹H-NMR (CDCl₃, 500 MHz) δ 1.68-1.77 (m, 2H), 1.80-1.91 (m, 2H), 2.47 (t, *J* = 6.5 Hz, 2H), 2.98-3.06 (m, 2H), 3.68 (s, 3H), 6.39 (t, *J* = 2.0 Hz, 1H) ppm; ¹³C{¹H}-NMR (CDCl₃, 125 MHz) δ 23.4 (two CH₂s), 28.8 (CH₂), 41.0 (CH₂), 51.6 (CH₃), 121.6 (CH), 151.7 (C), 166.5 (C), 201.1 (C) ppm; IR (neat): ν_{max} (cm⁻¹) = 2949 (w), 1718 (s), 1693 (s), 1435 (m), 1362 (m), 1256 (m), 1196 (s), 1175 (s), 1138 (s), 1015 (m), 816 (m), 631 (m), 527 (m); HRMS (ESI): *m/z* calculated for C₉H₁₃O₃ [M+H]⁺ 169.0860, found 169.0859.

Ethyl 2-(6-oxocyclohex-1-en-1-yl)acetate *endo*-**9a**^[2]

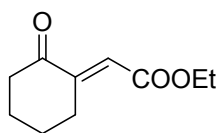


Prepared according to Procedure B from *exo*-**9a** (400 mg, 2.2 mmol) and DBU (132 μ L, 0.4 mmol) and used without further purification. Isolated as a brown oil. Yield: 311 mg, 1.71 mmol, 78%.

¹H-NMR (CDCl₃, 500 MHz) δ 1.24 (t, *J* = 7.0 Hz, 3H), 2.03 (quint, *J* = 6.5 Hz, 2H), 2.38-2.44 (m, 2H), 2.46 (t, *J* = 7.0 Hz, 2H), 3.18 (d, *J* = 1 Hz, 2H), 4.13 (q, *J* = 7.0 Hz, 2H), 6.87 (t, *J* = 3.5 Hz, 1H) ppm; ¹³C{¹H}-NMR (CDCl₃, 125 MHz) δ 14.3 (CH₃), 23.1 (CH₂), 26.2 (CH₂), 35.5 (CH₂), 38.1 (CH₂), 60.9 (CH₂), 133.7 (C), 148.4

(CH), 171.6 (C), 198.4 (C) ppm; IR (neat): ν_{max} (cm^{-1}) = 2935 (w), 1732 (s), 1670 (s), 1369 (m), 1331 (m), 1258 (m), 1173 (s), 1030 (m), 625 (m), 463 (m); HRMS (ESI): m/z calculated for $\text{C}_{10}\text{H}_{14}\text{NaO}_3$ $[\text{M}+\text{Na}]^+$ 205.0835, found 205.0846.

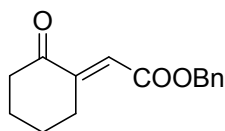
Ethyl (*E*)-2-(2-oxocyclohexylidene)acetate *exo*-9a^[1]



Prepared according to Procedure A from 1,2-cyclohexanedione (113 mg, 1 mmol) and ethoxycarbonylmethyltriphenylphosphonium bromide (430 mg, 1 mmol). Purification: column chromatography on silica gel (cyclohexane/ethyl acetate, 19/1, R_f = 0.17). Isolated as a colorless oil. Yield: 90 mg, 0.55 mmol, 55%.

^1H -NMR (CDCl_3 , 500 MHz) δ 1.28 (t, J = 7.0 Hz, 3H), 1.7-1.82 (m, 2H), 1.87-1.95 (m, 2H), 2.52 (t, J = 7.0 Hz, 2H), 3.08 (td, J = 7.0 Hz, J = 2.0 Hz, 2H), 4.19 (q, J = 7.0 Hz, 2H), 6.46 (t, J = 2.0 Hz, 1H) ppm; $^{13}\text{C}\{^1\text{H}\}$ -NMR (CDCl_3 , 125 MHz) δ 14.3 (CH_3), 23.5 (CH_2), 23.6 (CH_2), 28.9 (CH_2), 41.1 (CH_2), 60.6 (CH_2), 122.3 (CH), 151.4 (C), 166.2 (C), 201.4 (C) ppm; IR (neat): ν_{max} (cm^{-1}) = 2939 (w), 1717 (s), 1369 (m), 1184 (s), 633 (m), 532 (s), 498 (s); HRMS (ESI): m/z calculated for $\text{C}_{10}\text{H}_{14}\text{NaO}_3$ $[\text{M}+\text{Na}]^+$ 205.0835, found 205.0846.

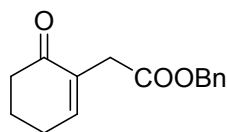
Benzyl (*E*)-2-(2-oxocyclohexylidene)acetate *exo*-10a



Prepared according to Procedure A from 1,2-cyclohexanedione (560 mg, 5 mmol) and benzyloxycarbonylmethyltriphenylphosphonium bromide (2.95 g, 6 mmol). Purification: column chromatography on silica gel (cyclohexane/ethyl acetate, 20/1, R_f = 0.12). Isolated as a yellow oil. Yield: 950 mg, 3.9 mmol, 78%.

^1H -NMR (CDCl_3 , 500 MHz) δ 1.78 (quint, J = 5.0 Hz, 2H), 1.90 (quint, J = 6.0 Hz, 2H), 2.52 (t, J = 7.0 Hz, 2H), 3.10 (td, J = 7.0 Hz, J = 2.0 Hz, 2H), 5.18 (s, 2H), 6.52 (t, J = 2.0 Hz, 1H), 7.28-7.38 (m, 5H) ppm; $^{13}\text{C}\{^1\text{H}\}$ -NMR (CDCl_3 , 125 MHz) δ 23.4 (two CH_2 s), 28.9 (CH_2), 41.0 (CH_2), 66.3 (CH_2), 121.7 (CH), 128.2 (CH), 128.3 (CH), 128.6 (CH), 135.7 (C), 152.0 (C), 165.8 (C), 201.1 (C) ppm; IR (neat): ν_{max} (cm^{-1}) = 2900 (w), 1716 (s), 1696 (s), 1685 (s), 1231 (m), 1168 (s), 737 (m), 696 (s); HRMS (ESI): m/z calculated for $\text{C}_{15}\text{H}_{16}\text{NaO}_3$ $[\text{M}+\text{Na}]^+$ 267.0992, found 267.0985.

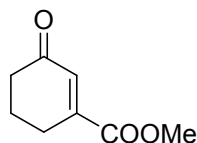
Benzyl 2-(6-oxocyclohex-1-en-1-yl)acetate *endo*-10a



Prepared according to Procedure B from *exo*-10a (500 mg, 2 mmol) and DBU (60 μL , 0.4 mmol). Purification by column chromatography on silica gel (cyclohexane/ethyl acetate, 8/1, R_f = 0.25). Isolated as a yellow oil. Yield: 400 mg, 1.6 mmol, 80%.

^1H -NMR (CDCl_3 , 500 MHz) δ 2.02 (quint, J = 7.0 Hz, 2H), 2.39 (q, J = 5.0 Hz, 2H), 2.46 (t, J = 5.0 Hz, 2H), 3.24 (s, 2H), 5.11 (s, 2H), 6.86 (t, J = 4.0 Hz, 1H), 7.28-7.38 (m, 5H) ppm; $^{13}\text{C}\{^1\text{H}\}$ -NMR (CDCl_3 , 125 MHz) δ 23.0 (CH_2), 26.1 (CH_2), 35.5 (CH_2), 38.0 (CH_2), 66.6 (CH_2), 128.2 (CH), 128.3 (CH), 128.6 (CH), 133.5 (C), 136.0 (C), 148.6 (CH), 171.3 (C), 198.3 (C) ppm; IR (neat): ν_{max} (cm^{-1}) = 2940 (w), 1743 (s), 1730 (s), 1684 (s), 1231 (m), 1163 (s), 745 (m), 698 (m); HRMS (ESI): m/z calculated for $\text{C}_{15}\text{H}_{16}\text{NaO}_3$ $[\text{M}+\text{Na}]^+$ 267.0992, found 267.0982.

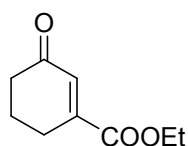
Methyl 3-oxocyclohex-1-ene-1-carboxylate **11a**^[3]



Prepared according to Procedure C from methyl cyclohex-1-ene-1-carboxylate (1 g, 7.1 mmol). Purification by column chromatography on silica gel (cyclohexane/ethyl acetate, 4/1, $R_f = 0.26$). Isolated as a yellow oil. Yield: 600 mg, 3.9 mmol, 55%.

$^1\text{H-NMR}$ (CDCl_3 , 500 MHz) δ 1.97 (quint, $J = 7.0$ Hz, 2H), 2.34 (t, $J = 7.0$ Hz, 2H), 2.45-2.53 (m, 2H), 3.73 (s, 3H), 6.61 (s, 1H) ppm; $^{13}\text{C}\{^1\text{H}\}\text{-NMR}$ (CDCl_3 , 125 MHz) δ 22.0 (CH_2), 24.7 (CH_2), 37.6 (CH_2), 52.5 (CH_3), 132.9 (CH), 148.7 (C), 166.8 (C), 199.9 (C) ppm; IR (neat): ν_{max} (cm^{-1}) = 2953 (w), 1721 (s), 1682 (s), 1435 (m), 1250 (s), 1225 (s), 1076 (m), 1043 (m), 743 (m), 727 (m); HRMS (ESI): m/z calculated for $\text{C}_{18}\text{H}_{10}\text{NaO}_3$ $[\text{M}+\text{Na}]^+$ 177.0521, found 177.0522.

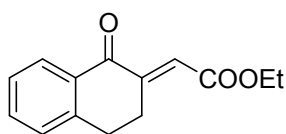
Ethyl 3-oxocyclohex-1-ene-1-carboxylate **12a**^[24]



Prepared according to Procedure C from ethyl cyclohex-1-ene-1-carboxylate (325 mg, 2.1 mmol). Purification by column chromatography on silica gel (cyclohexane/ethyl acetate, 9/1, $R_f = 0.19$). Isolated as a pale yellow oil. Yield: 184 mg, 1.1 mmol, 52%.

$^1\text{H-NMR}$ (CDCl_3 , 500 MHz) δ 1.26 (t, $J = 7.0$ Hz, 3H), 2.00 (quint, $J = 7.0$ Hz, 2H), 2.37 (t, $J = 7.0$ Hz, 2H), 2.51 (td, $J = 7.0$ Hz, $J = 1.0$ Hz, 2H), 4.20 (q, $J = 7.0$ Hz, 2H), 6.65 (t, $J = 1.0$ Hz, 1H) ppm; $^{13}\text{C}\{^1\text{H}\}\text{-NMR}$ (CDCl_3 , 125 MHz) δ 14.1 (CH_3), 22.1 (CH_2), 24.8 (CH_2), 37.7 (CH_2), 61.6 (CH_2), 132.8 (CH), 149.2 (C), 166.4 (C), 200.1 (C) ppm; IR (neat): ν_{max} (cm^{-1}) = 2955 (w), 1717 (s), 1682 (s), 1250 (s), 1223 (s), 1076 (m), 1042 (m), 964 (m), 741 (m), 633 (m), 498 (s); HRMS (ESI): m/z calculated for $\text{C}_9\text{H}_{13}\text{O}_3$ $[\text{M}+\text{H}]^+$ 169.0865, found 169.0845.

Ethyl (*E*)-2-(1-oxo-3,4-dihydronaphthalen-2(1*H*)-ylidene)acetate **14a**

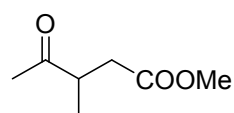


To a solution of α -tetralone (1.35 mL, 10 mmol) in toluene (50 mL) were added MgSO_4 (8 g), ethyl glyoxylate (50% solution in toluene, 2 mL, 10 mmol) and *p*-toluenesulfonic acid monohydrate (190 mg, 1 mmol). The mixture was refluxed for 15 hours. After cooling, MgSO_4 was filtered off. The solution was diluted with ethyl acetate and washed with water and brine, dried over MgSO_4 and concentrated *in vacuo*. The product was isolated by column chromatography on silica gel (cyclohexane/ethyl acetate, 13/1) as a purple solid (1.77g, 7.7 mmol, 77%). m.p.: 104-107.5 °C.

$^1\text{H-NMR}$ (CDCl_3 , 500 MHz) δ 1.34 (t, $J = 7.5$ Hz, 3H), 3.03 (t, $J = 6.5$ Hz, 2H), 3.44 (td, $J = 6.5$ Hz, $J = 1.5$ Hz, 2H), 4.26 (q, $J = 7.5$ Hz, 2H), 6.90 (t, $J = 1.5$ Hz, 1H), 7.28 (d, $J = 7.5$ Hz, 1H), 7.37 (t, $J = 7.5$ Hz, 1H), 7.52 (t, $J = 7.5$ Hz, 1H), 8.10 (t, $J = 7.5$ Hz, 1H) ppm; $^{13}\text{C}\{^1\text{H}\}\text{-NMR}$ (CDCl_3 , 125 MHz) δ 14.4 (CH_3), 27.2 (CH_2), 28.7 (CH_2), 60.8 (CH_2), 123.7 (CH), 127.4 (CH), 128.6 (CH), 128.6 (CH), 132.7 (C), 134.1 (CH), 144.3 (C), 149.7 (C), 166.4 (C), 187.1 (C) ppm; IR (neat): ν_{max} (cm^{-1}) = 2974 (w), 1705 (s), 1666 (m), 1593 (m), 1292 (s), 1242 (s), 1161(s), 903 (m), 744 (s); HRMS (ESI): m/z calculated for $\text{C}_{14}\text{H}_{14}\text{NaO}_3$ $[\text{M}+\text{Na}]^+$ 239.0679, found 239.0682.

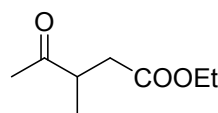
5. Characterization of products

Methyl 3-methyl-4-oxopentanoate *rac*-1b^[4]



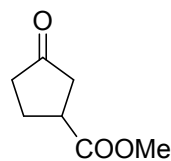
¹H-NMR (CDCl₃, 500 MHz) δ 1.12 (d, J = 7.0 Hz, 3H), 1.21 (m, 3H), 2.20 (s, 3H), 2.27 (dd, J = 17.0 Hz, J = 5.0 Hz, 1H), 2.74 (dd, J = 17.0 Hz, J = 8.5 Hz, 1H), 2.93-3.03 (m, 1H), 3.63 (s, 3H) ppm; ¹³C{¹H}-NMR (CDCl₃, 125 MHz) δ 16.6 (CH₃), 28.5 (CH₃), 36.7 (CH₂), 42.8 (CH), 51.8 (CH₃), 172.9 (C), 210.9 (C) ppm.

Ethyl 3-methyl-4-oxopentanoate *rac*-2b^[5]



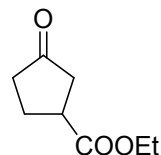
¹H-NMR (CDCl₃, 500 MHz) δ 1.11 (d, J = 7.0 Hz, 3H), 1.21 (t, J = 7.0 Hz, 3H), 2.18 (s, 3H), 2.25 (dd, J = 17.0 Hz, J = 5.0 Hz, 1H), 2.71 (dd, J = 17.0 Hz, J = 8.5 Hz, 1H), 2.92-3.02 (m, 1H), 4.07 (q, J = 7.0 Hz, 2H) ppm; ¹³C{¹H}-NMR (CDCl₃, 125 MHz) δ 14.2 (CH₃), 16.6 (CH₃), 28.5 (CH₃), 37.0 (CH₂), 42.8 (CH), 60.5 (CH₂), 172.4 (C), 210.9 (C) ppm.

Methyl 3-oxocyclopentane-1-carboxylate *rac*-3b^[8]



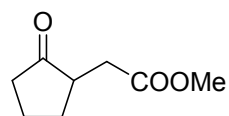
¹H-NMR (CDCl₃, 500 MHz) δ 2.13-2.51 (m, 6H), 3.13 (quint, J = 8.0 Hz, 1H), 3.73 (s, 3H) ppm; ¹³C{¹H}-NMR (CDCl₃, 125 MHz) δ 26.7 (CH₂), 37.6 (CH₂), 40.9 (CH₂), 41.3 (CH), 52.3 (CH₃), 174.9 (C), 216.7 (C) ppm.

Ethyl 3-oxocyclopentane-1-carboxylate *rac*-4b^[9]



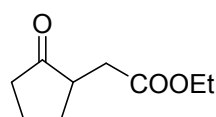
¹H-NMR (CDCl₃, 500 MHz) δ 1.27 (t, J = 7.0 Hz, 3H), 2.12-2.50 (m, 6H), 3.11 (quint, J = 8.0 Hz, 1H), 4.17 (q, J = 7.0 Hz, 2H) ppm; ¹³C{¹H}-NMR (CDCl₃, 125 MHz) δ 14.3 (CH₃), 26.7 (CH₂), 37.6 (CH₂), 41.1 (CH), 41.3 (CH₂), 61.2 (CH₂), 174.4 (C), 216.9 (C) ppm.

Methyl 2-(2-oxocyclopentyl)acetate *rac*-5b^[6]



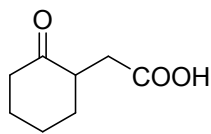
¹H-NMR (CDCl₃, 500 MHz) δ 1.55-1.65 (m, 1H), 1.75-1.87 (m, 1H), 2.02-2.10 (m, 1H), 2.11-2.23 (m, 1H), 2.29-2.44 (m, 4H), 2.68-2.80 (m, 1H), 3.68 (s, 3H) ppm; ¹³C{¹H}-NMR (CDCl₃, 125 MHz) δ 20.7 (CH₂), 29.5 (CH₂), 33.9 (CH₂), 37.6 (CH₂), 45.8 (CH), 51.9 (CH₃), 172.8 (C) ppm. Note: the cyclopentanone C=O appears at δ >217 ppm.

Ethyl 2-(2-oxocyclopentyl)acetate *rac*-6b^[7]



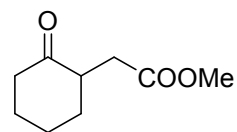
¹H-NMR (CDCl₃, 500 MHz) δ 1.21 (t, J = 7.0 Hz, 3H), 1.53-1.64 (m, 1H), 1.70-1.83 (m, 1H), 1.96-2.06 (m, 1H), 2.08-2.18 (m, 1H), 2.20-2.44 (m, 4H), 2.63-2.71 (m, 1H), 4.08 (q, J = 7.0 Hz, 2H) ppm; ¹³C{¹H}-NMR (CDCl₃, 125 MHz) δ 14.2 (CH₃), 20.7 (CH₂), 29.3 (CH₂), 34.0 (CH₂), 37.5 (CH₂), 45.6 (CH), 60.7 (CH₂), 172.1 (C) ppm. Note: the cyclopentanone C=O appears at δ >217 ppm.

2-(2-Oxocyclohexyl)acetic acid *rac*-7b^[10]



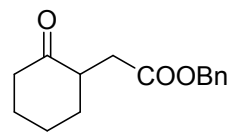
¹H-NMR (CDCl₃, 500 MHz) δ 1.41 (qd, J = 13.5 Hz, 2.0 Hz, 1H), 1.55-1.80 (m, 2H), 1.87 (d, J = 13.5 Hz, 1H), 2.03-2.25 (m, 3H), 2.30-2.48 (m, 2H), 2.74-2.88 (m, 2H), 11.31 (br, 1H) ppm; ¹³C{¹H}-NMR (CDCl₃, 125 MHz) δ 25.3 (CH₂), 27.9 (CH₂), 33.9 (CH₂), 34.4 (CH₂), 41.9 (CH₂), 47.0 (CH), 178.7 (C), 211.3 (C) ppm.

Methyl 2-(2-oxocyclohexyl)acetate *rac*-8b^[11]



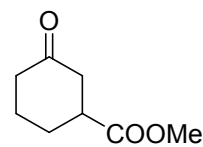
¹H-NMR (CDCl₃, 500 MHz) δ 1.36 (qd, J = 13.5 Hz, 2.0 Hz, 1H), 1.51-1.73 (m, 2H), 1.82 (d, J = 13.5 Hz, 1H), 2.03-2.14 (m, 3H), 2.25-2.38 (m, 2H), 2.71 (dd, J = 16.5 Hz, J = 7.5 Hz, 1H), 2.81 (sex, J = 7.0, 1H), 3.60 (s, 3H) ppm; ¹³C{¹H}-NMR (CDCl₃, 125 MHz) δ 25.1 (CH₂), 27.8 (CH₂), 33.9 (CH₂), 34.2 (CH₂), 41.8 (CH₂), 47.0 (CH), 51.6 (CH₃), 173.0 (C), 211.0 (C) ppm.

Benzyl 2-(2-oxocyclohexyl)acetate *rac*-10b^[13]



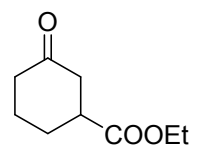
¹H-NMR (CDCl₃, 500 MHz) δ 1.40 (qd, J = 13.5, 2.0 Hz, 1H), 1.55-1.80 (m, 2H), 1.80-1.93 (m, 1H), 2.03-2.26 (m, 3H), 2.28-2.48 (m, 2H), 2.74-2.95 (m, 2H), 5.11 (s, 2H), 7.26-7.40 (m, 5H) ppm; ¹³C{¹H}-NMR (CDCl₃, 125 MHz) δ 25.3 (CH₂), 27.9 (CH₂), 34.0 (CH₂), 34.4 (CH₂), 41.9 (CH₂), 47.1 (CH), 66.3 (CH₂), 128.1 (CH), 128.2 (CH), 128.5 (CH), 136.0 (C), 172.6 (C), 211.0 (C) ppm.

Methyl 3-oxocyclohexane-1-carboxylate *rac*-11b^[8]



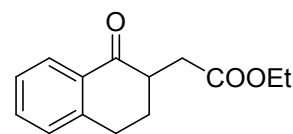
¹H-NMR (CDCl₃, 500 MHz) δ 1.67-1.78 (m, 1H), 1.85 (qd, J = 12.0 Hz, 3.5 Hz, 1H), 2.01-2.16 (m, 2H), 2.27-2.42 (m, 2H), 2.55 (d, J = 8.0 Hz, 2H), 2.75-2.86 (m, 1H), 3.71 (s, 3H) ppm; ¹³C{¹H}-NMR (CDCl₃, 125 MHz) δ 24.6 (CH₂), 27.9 (CH₂), 41.0 (CH₂), 43.1 (CH), 43.2 (CH₂), 52.3 (CH₃), 174.3 (C), 209.4 (C) ppm.

Ethyl 3-oxocyclohexane-1-carboxylate *rac*-12b^[11]



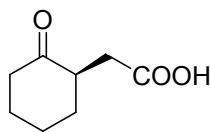
¹H-NMR (CDCl₃, 500 MHz) δ 1.26 (t, J = 7.0 Hz, 3H), 1.67-1.75 (m, 1H), 1.76-1.88 (m, 1H), 2.02-2.15 (m, 2H), 2.25-2.40 (m, 2H), 2.54 (d, J = 8.0 Hz, 2H), 2.74-2.82 (m, 1H), 4.15 (q, J = 7.0 Hz, 2H) ppm; ¹³C{¹H}-NMR (CDCl₃, 125 MHz) δ 14.3 (CH₃), 24.6 (CH₂), 27.9 (CH₂), 41.1 (CH₂), 43.3 (CH₂), 43.4 (CH), 61.0 (CH₂), 173.9 (C), 209.5 (C) ppm.

Ethyl 2-(1-oxo-1,2,3,4-tetrahydronaphthalen-2-yl)acetate *rac*-14b^[14]



¹H-NMR (CDCl₃, 500 MHz) δ 1.28 (t, J = 7.5 Hz, 3H), 1.97 (dq, J = 13.0 Hz, J = 4.0 Hz, 1H), 2.25 (dd, J = 9.0 Hz, J = 4.0 Hz, 1H), 2.42 (dd, J = 6.5 Hz, J = 15.5 Hz, 1H), 2.96-3.13 (m, 4H), 4.19 (q, J = 7.5 Hz, 2H), 7.24 (d, J = 7.5 Hz, 1H), 7.31 (t, J = 7.5 Hz, 1H), 7.47 (t, J = 7.5 Hz, 1H), 8.03 (d, J = 7.5 Hz, 1H) ppm; ¹³C{¹H}-NMR (CDCl₃, 125 MHz) δ 14.4 (CH₃), 29.4 (CH₂), 35.3 (CH₂), 44.9 (CH), 60.7 (CH₂), 126.8 (CH), 127.6 (CH), 128.9 (CH), 132.3 (C), 133.5 (CH), 144.1 (C), 172.7 (C), 198.6 (C) ppm.

(*R*)-2-(2-Oxocyclohexyl)acetic acid (*R*)-7b



To a solution of **10b** obtained from scale up of the bioreduction of *endo*-**10a** with NCR (90 mg, 0.36 mmol) in ethyl acetate (1.5 mL) was added 10 w% Pd/C (45 mg, 50% w/w). The mixture was stirred under atmospheric pressure of H₂ (balloon) at room temperature for 21 h. The mixture was then filtered over Celite (the filter thoroughly washed with ethyl acetate) and concentrated *in vacuo* yielding the product of sufficient purity.

Isolated as an off-white solid. Yield: 49 mg, 0.31 mmol, 70%. $[\alpha]_D^{20} = +36.7$ ($c = 1.8$, CHCl₃). NMR as described above.

The *ee* (85% for (*R*)-**7b**) was determined by chiral GC upon derivatization to **8b** with TMSCHN₂ in MeOH-CHCl₃. The compound most likely epimerizes slowly.

6. Interpretation of isotopic labeling experiments

Reduction of 2a

In the ¹H-NMR spectrum of product **2b** obtained in non-deuterated medium, the proton at C α of the ketone carbonyl gives a typical signal at 2.92-3.02 ppm (multiplet), while the methyl group attached to that C α gives an expected doublet at 1.11 ppm. The two protons on C β (i.e. C α of the ester moiety) give two distinguishable signals (doublet of doublets) at 2.25 and 2.71 ppm, respectively.

Upon reduction of (*Z*)-**2a** by NCR in D₂O yielding (*R*)-**2b**, the multiplet at 2.92-3.02 ppm disappears while the doublet at 1.11 ppm is replaced by a singlet, which is a strong evidence that the proton at C α has been replaced by deuterium. At the same time, the dd signals of both protons on C β are now doublets, confirming insertion of D at C α of the ketone carbonyl, which is the binding group in NCR.

Bioreduction of (*E*)-**2a** by NCR at the expense of NADD forming (*S*)-**2b** resulted in a complex ¹H-NMR spectrum, despite full conversion and purification of the product. Importantly, the doublet at 1.11 ppm corresponding to the methyl group on C α is still present, indicating the presence of a proton on that carbon. The corresponding signal of that proton clearly appears as a quintuplet, indicating that the neighboring carbon C β now bears a single proton. Additionally, the dd signal at 2.25 ppm corresponding to one of the protons on C β has disappeared. All this indicates that D has been incorporated on C β to the ketone carbonyl, which remains the binding group of (*E*)-**2a** in NCR. The ¹³C NMR spectrum supports this conclusion as it shows a splitting of the C β signal (37.0 ppm), a typical sign of a deuterium being present on a carbon atom.

Reduction of 3a

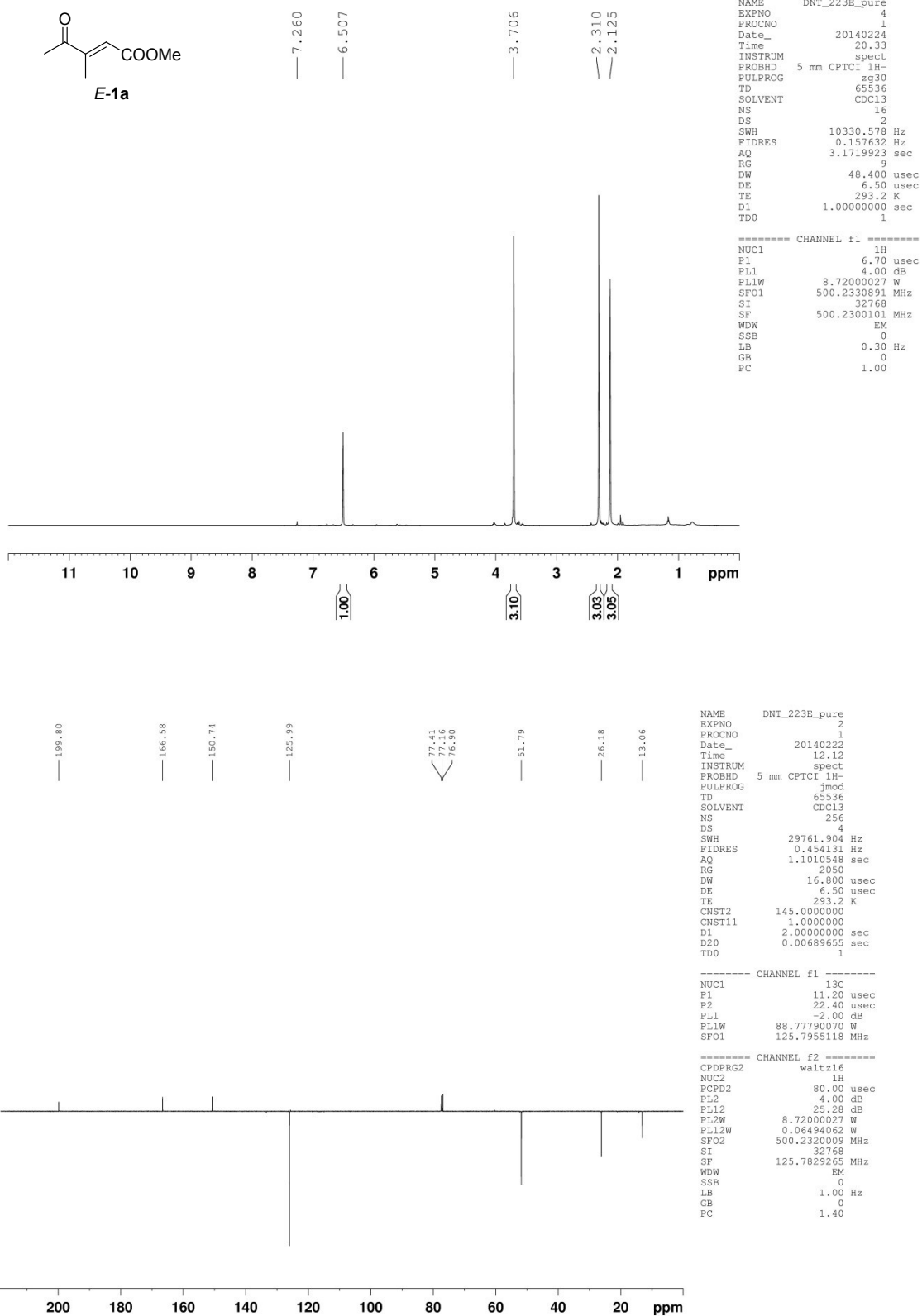
For product **3b**, the ¹H-NMR spectrum of a non-deuterated sample displays a typical signal for the proton on the most substituted carbon (C β to the ketone carbonyl) as a multiplet at 3.07-3.18 ppm. Upon reduction of **3a** at the expense of NADD by OYE2 to (*S*)-**3b** and by XenA to (*R*)-**3b**, a major decrease in the abundance of that signal is observed, without change of the shape, indicating that this position has been

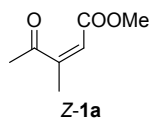
deuterated, and possibly that exchange with the solvent (H_2O) occurred (incorporation of D at $\text{C}\alpha$ would have modified the shape of that signal). Furthermore, in ^{13}C NMR, splitting of the $\text{C}\beta$ signal at position 40.9 ppm indicates incorporation of deuterium at that position. In both cases, the ketone carbonyl is activating. The HSQC-spectrum also clearly shows disappearance of the coupling between the $\text{C}\beta$ (40.9 ppm) and the corresponding H (replaced by D).

Reduction of 5a

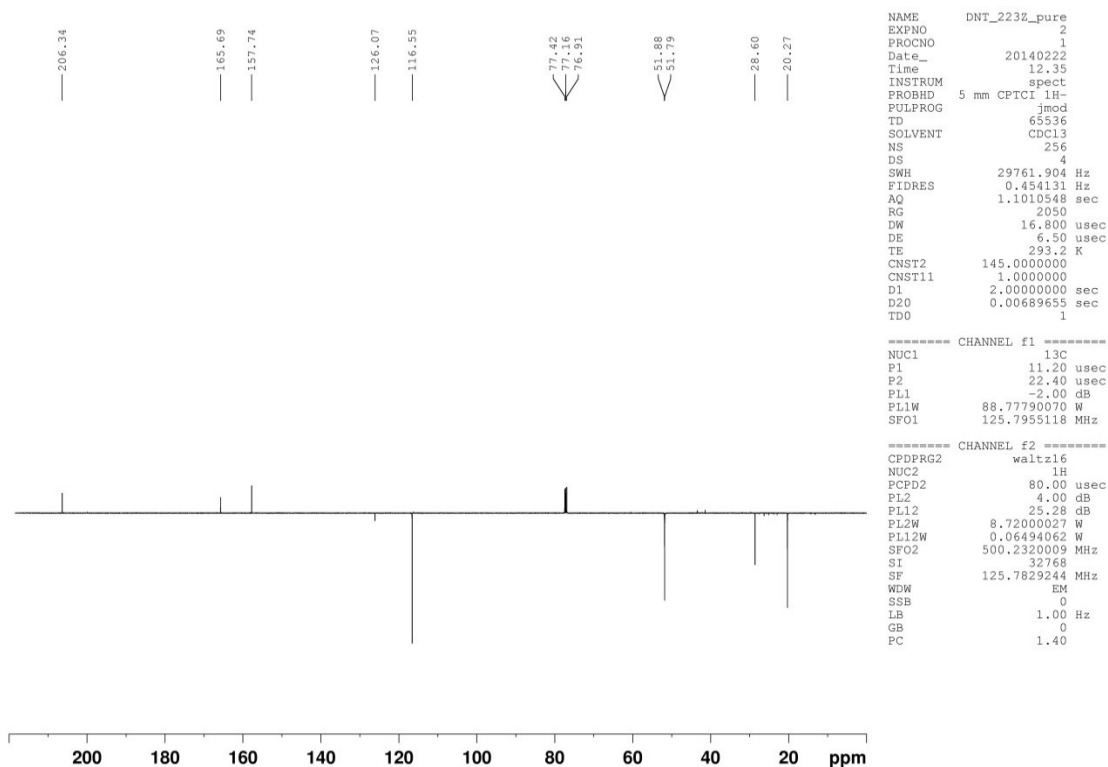
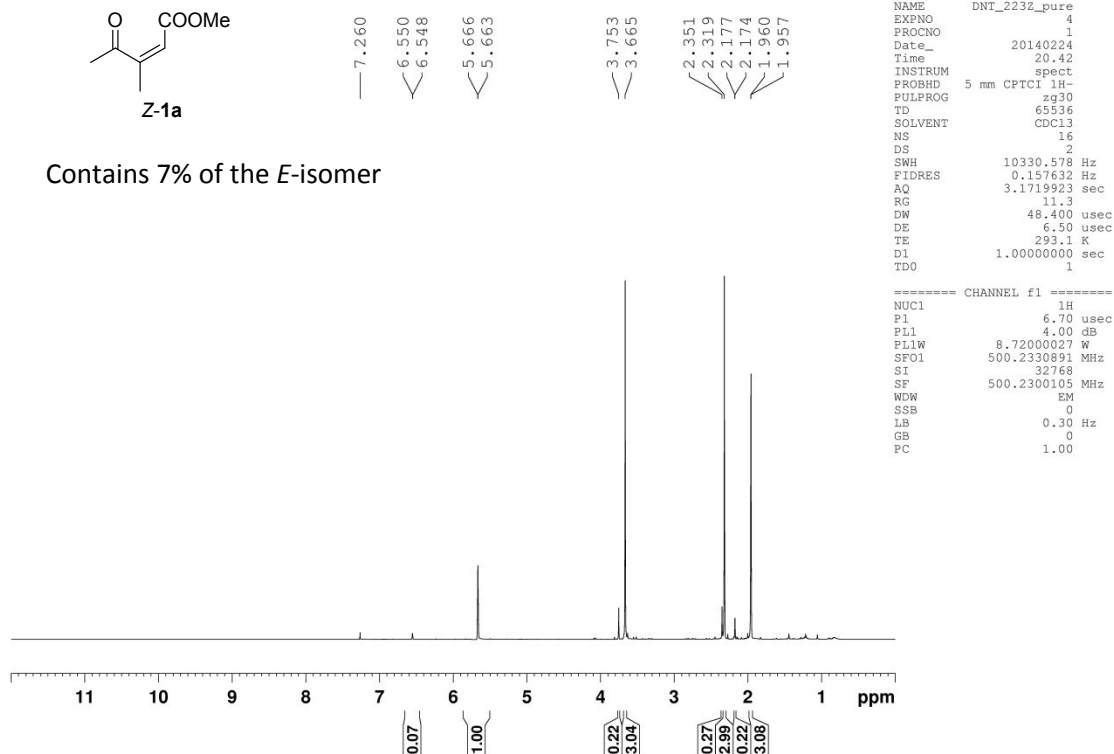
(*R*)-**5b** was obtained from reduction of **5a** by NCR in D_2O . Analysis of the resulting ^1H -NMR spectrum indicates a change in the shape and integral of signals at 2.71-2.76 ppm and 2.28-2.55 ppm. However, overlap with the signal of the proton on $\text{C}\alpha$ to the ketone carbonyl, which gives a multiplet under non-deuterating conditions, does not allow integration of the signals, which were significantly altered. This indicates (partial) incorporation of D at $\text{C}\alpha$ and thus suggests that the ketone carbonyl is the binding group. In addition, the carbon signal in ^{13}C NMR of $\text{C}\alpha$ at 45.7 ppm shows a splitting and reduced intensity, indicating incorporation of a deuterium atom. Finally, the HSQC-spectrum clearly shows no coupling of the $\text{C}\alpha$ (45 ppm) to any hydrogen (the signal at {2.4,45} ppm has disappeared). The ketone carbonyl is the sole binding group.

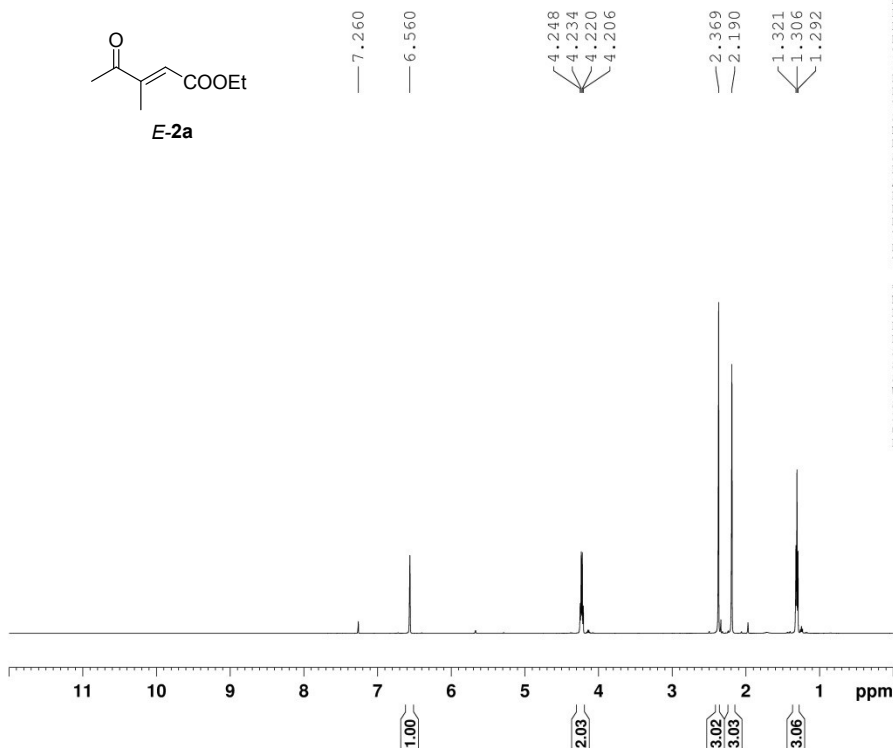
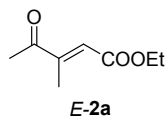
7. NMR spectra





Contains 7% of the *E*-isomer



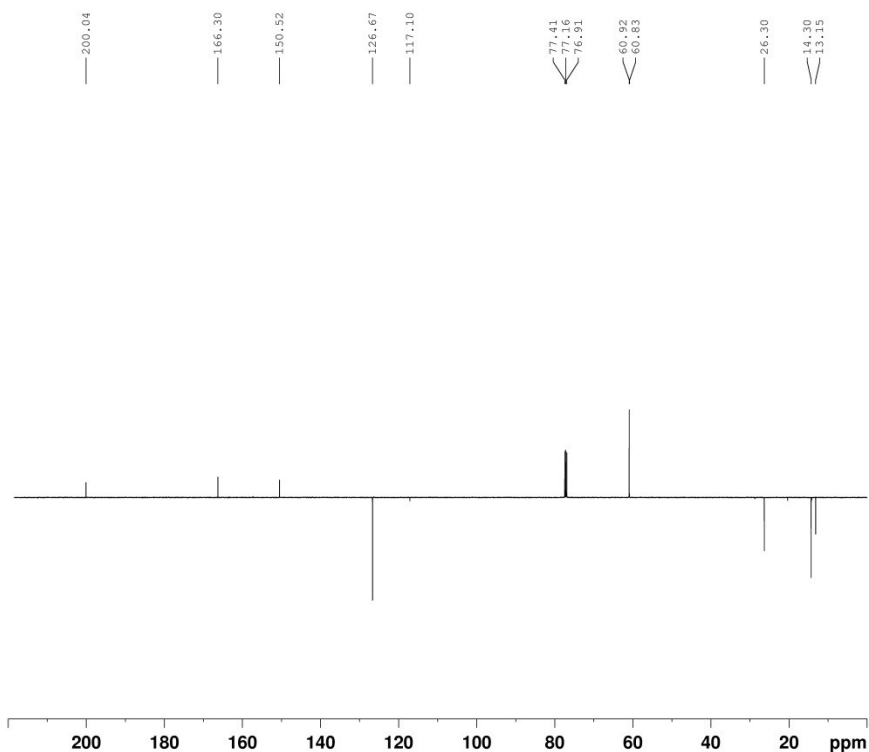


```

NAME      DNT_188_pure
EXPNO     1
PROCNO    1
Date_     20140203
Time      19.00
INSTRUM   spect
PROBHD    5 mm CPTCI 1H-
PULPROG   zg30
TD         65536
SOLVENT   CDCl3
NS         4
DS         2
SWH        10330.578 Hz
FIDRES     0.157632 Hz
AQ         3.1719923 sec
RG         14.2
DW         48.400 usec
DE         6.50 usec
TE         293.2 K
D1         1.00000000 sec
D10        1
  
```

```

===== CHANNEL f1 =====
NUC1      1H
P1        6.70 usec
PL1       4.00 dB
PL1W      8.72000027 W
SFO1      500.230891 MHz
SI        32768
SF        500.2300110 MHz
WDW       EM
SSB       0
LB        0.30 Hz
GB        0
PC        1.00
  
```



```

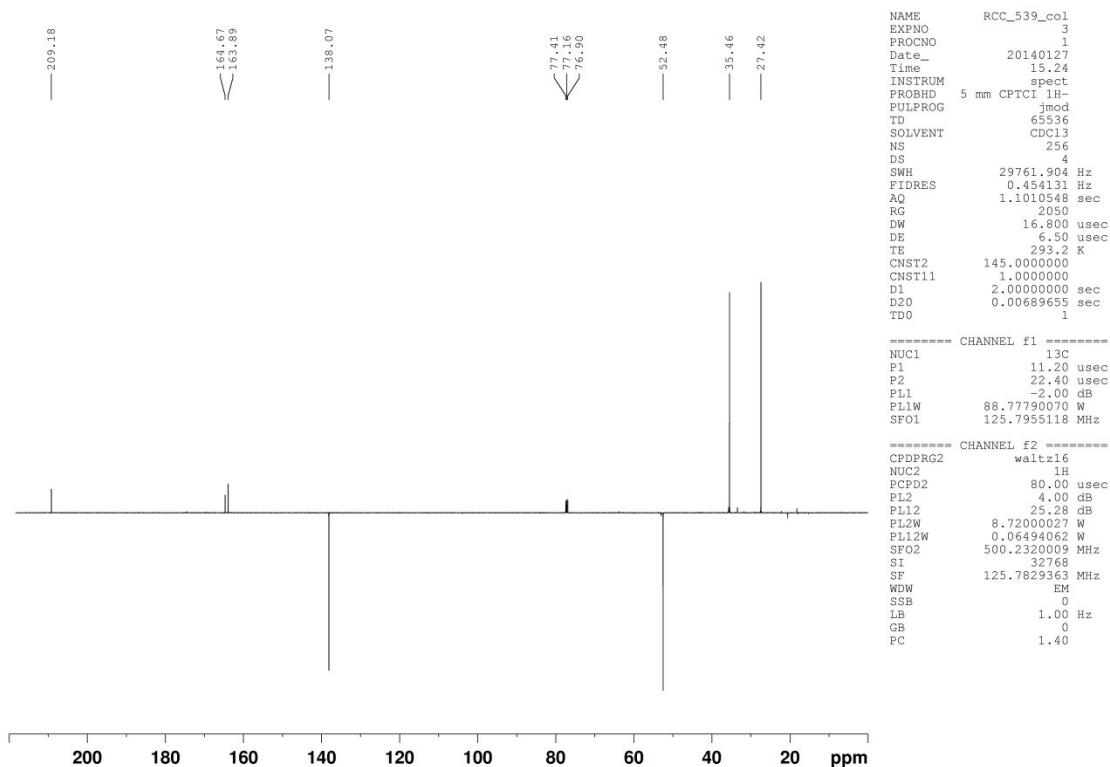
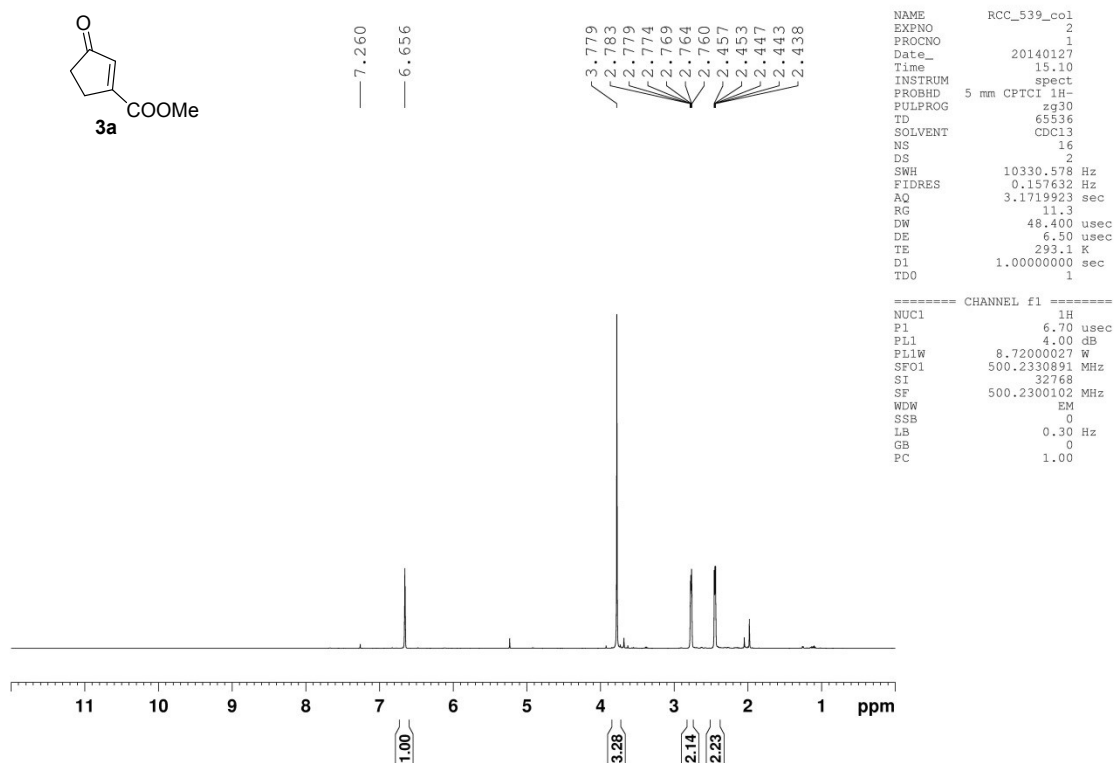
NAME      DNT_188_pure
EXPNO     2
PROCNO    1
Date_     20140203
Time      19.15
INSTRUM   spect
PROBHD    5 mm CPTCI 1H-
PULPROG   jmod
TD         65536
SOLVENT   CDCl3
NS         256
DS         4
SWH        29761.904 Hz
FIDRES     0.454131 Hz
AQ         1.1010548 sec
RG         2050
DW         16.800 usec
DE         6.50 usec
TE         293.2 K
CNST2     145.0000000
CNST11    1.0000000
D1         2.00000000 sec
D20        0.00689655 sec
D10        1
  
```

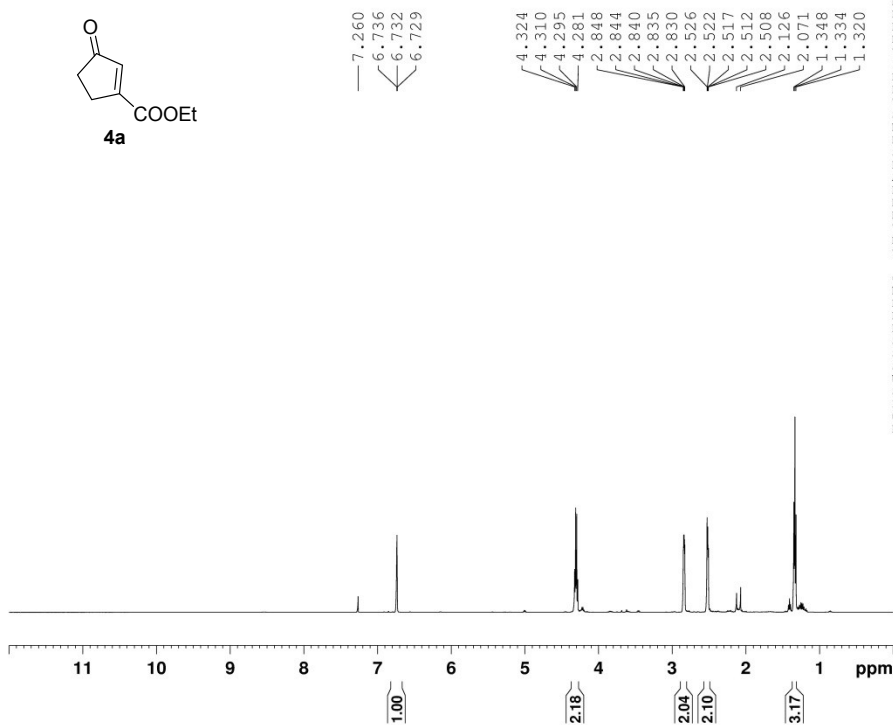
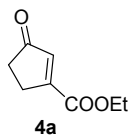
```

===== CHANNEL f1 =====
NUC1      13C
P1        11.20 usec
P2        22.40 usec
PL1       -2.00 dB
PL1W      88.77790070 W
SFO1      125.7955118 MHz
  
```

```

===== CHANNEL f2 =====
CPDPRG2   waltz16
NUC2      1H
PCPD2     80.00 usec
PL2       4.00 dB
PL12      25.28 dB
PL2W      8.72000027 W
PL12W     0.06494062 W
SFO2      500.2320009 MHz
SI        32768
SF        125.7829203 MHz
WDW       EM
SSB       0
LB        1.00 Hz
GB        0
PC        1.40
  
```

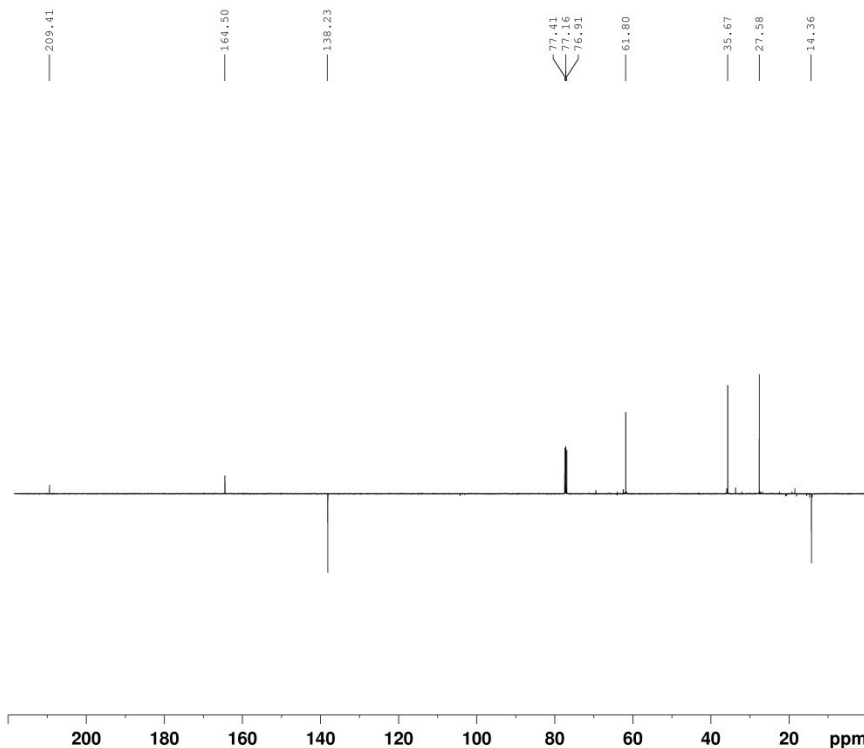




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NAME      RCC_577_pcol
EXPNO     1
PROCNO    1
Date_     20140303
Time      13.42
INSTRUM   spect
PROBHD    5 mm CPTCI 1H-
PULPROG   zg30
TD         65536
SOLVENT   CDCl3
NS         3
DS         2
SWH        10330.578 Hz
FIDRES     0.157632 Hz
AQ         3.1719923 sec
RG         14.2
DW         48.400 usec
DE         6.50 usec
TE         293.2 K
D1         1.0000000 sec
D10        1
TD0        1

===== CHANNEL f1 =====
NUC1       1H
P1         6.70 usec
PL1        4.00 dB
PL1W       8.72000027 W
SFO1       500.2330891 MHz
SI         32768
SF         500.2300107 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00
  
```

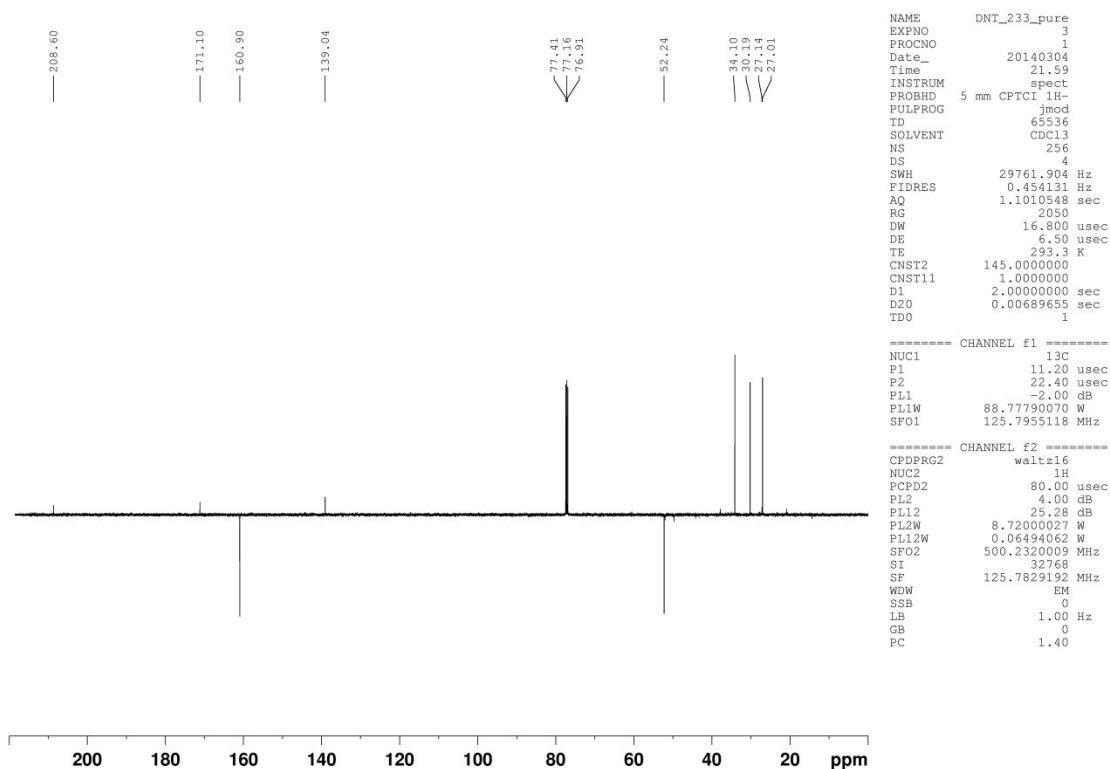
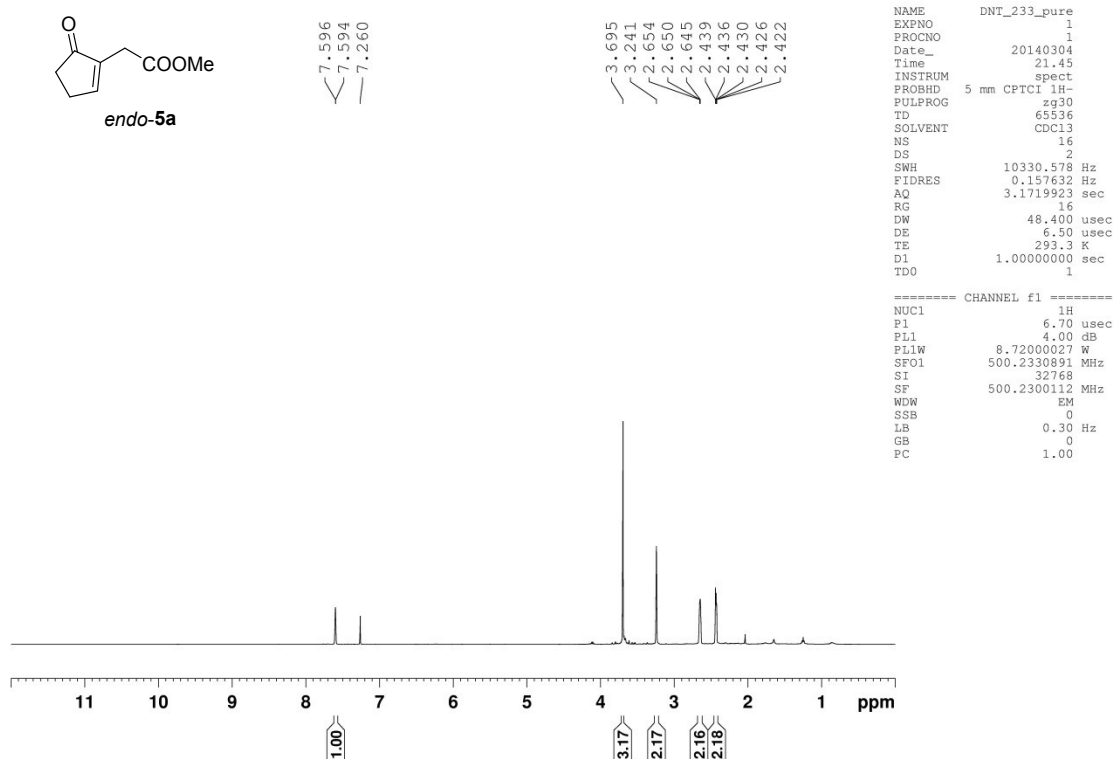


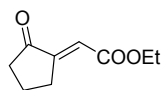
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NAME      RCC_577_pcol
EXPNO     2
PROCNO    1
Date_     20140303
Time      13.57
INSTRUM   spect
PROBHD    5 mm CPTCI 1H-
PULPROG   jmod
TD         65536
SOLVENT   CDCl3
NS         256
DS         4
SWH        29761.904 Hz
FIDRES     0.454131 Hz
AQ         1.1010548 sec
RG         2050
DW         16.800 usec
DE         6.50 usec
TE         293.2 K
CNST2     145.0000000
CNST11    1.0000000
D1         2.0000000 sec
D20        0.00689655 sec
TD0        1

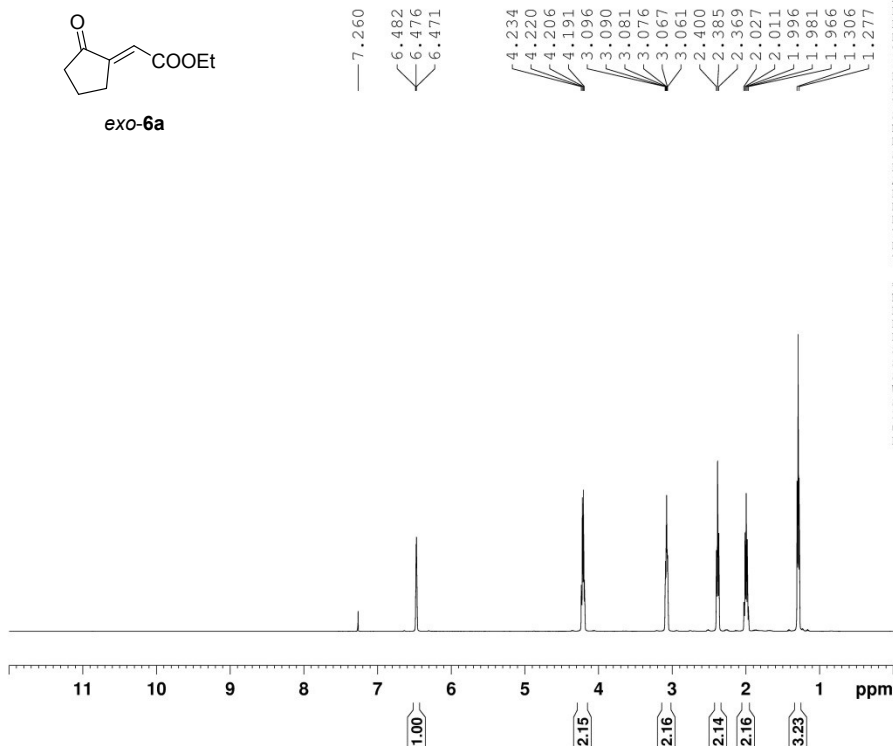
===== CHANNEL f1 =====
NUC1       13C
P1         11.20 usec
P2         22.40 usec
PL1        -2.00 dB
PL1W       88.77790070 W
SFO1       125.7955118 MHz

===== CHANNEL f2 =====
CPDPRG2   waltz16
NUC2       1H
PCPD2      80.00 usec
PL2        4.00 dB
PL12       25.28 dB
PL2W       8.72000027 W
PL12W      0.06494062 W
SFO2       500.2320009 MHz
SI         32768
SF         125.7829220 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         1.40
  
```





exo-6a



```

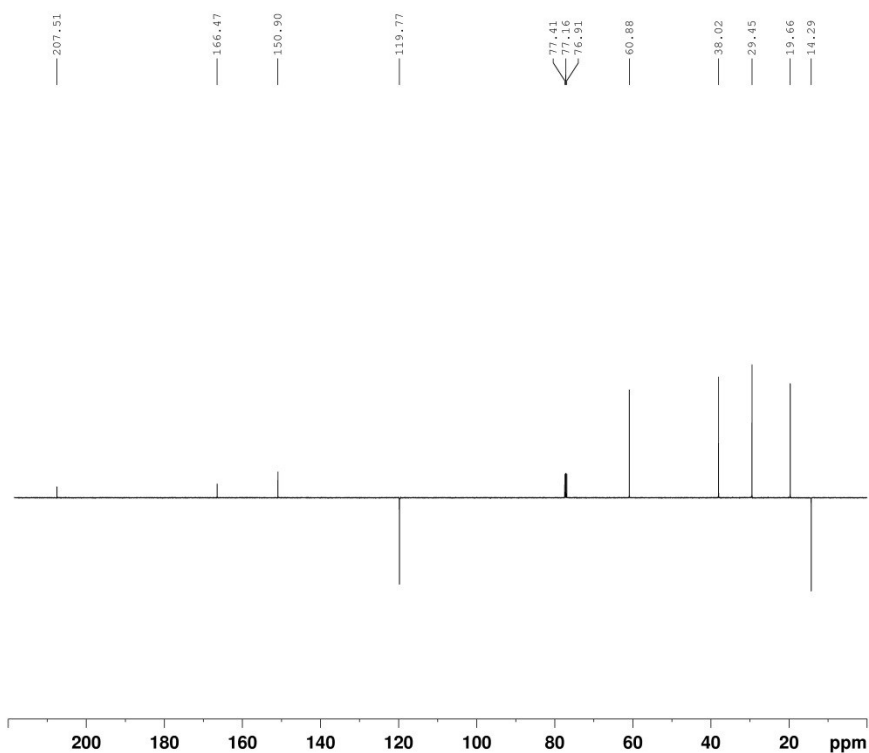
NAME      DNT_213_pure
EXPNO     2
PROCNO    1
Date_     20140211
Time      1.16
INSTRUM   spect
PROBHD    5 mm CPTCI 1H-
PULPROG   zg30
TD         65536
SOLVENT   CDCl3
NS         8
DS         2
SWH        10330.578 Hz
FIDRES     0.157632 Hz
AQ         3.1719923 sec
RG         18
DW         48.400 usec
DE         6.50 usec
TE         293.2 K
D1         1.00000000 sec
TD0        1

```

```

===== CHANNEL f1 =====
NUC1      1H
P1         6.70 usec
PL1        4.00 dB
PL1W      8.72000027 W
SFO1      500.2330891 MHz
SI         32768
SF         500.2300107 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00

```



```

NAME      DNT_213_pure
EXPNO     1
PROCNO    1
Date_     20140211
Time      1.16
INSTRUM   spect
PROBHD    5 mm CPTCI 1H-
PULPROG   jmod
TD         65536
SOLVENT   CDCl3
NS         256
DS         4
SWH        29761.904 Hz
FIDRES     0.454131 Hz
AQ         1.1010548 sec
RG         2050
DW         16.800 usec
DE         6.50 usec
TE         293.2 K
CNST2     145.0000000
CNST11    1.00000000
D1         2.00000000 sec
D20        0.00689655 sec
TD0        1

```

```

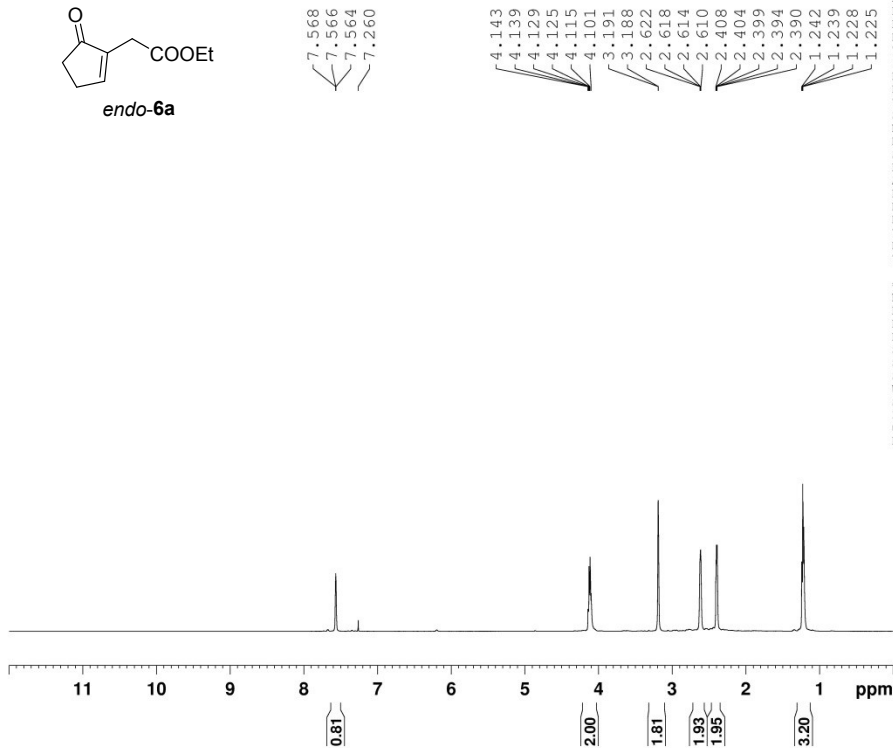
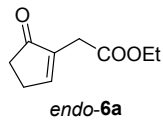
===== CHANNEL f1 =====
NUC1      13C
P1         11.20 usec
P2         22.40 usec
PL1        -2.00 dB
PL1W      88.77790070 W
SFO1      125.7955118 MHz

```

```

===== CHANNEL f2 =====
CPDPRG2   waltz16
NUC2      1H
PCPD2     80.00 usec
PL2        4.00 dB
PL12       25.28 dB
PL2W      8.72000027 W
PL12W     0.06494062 W
SFO2      500.2320009 MHz
SI         32768
SF         125.7829220 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         1.40

```

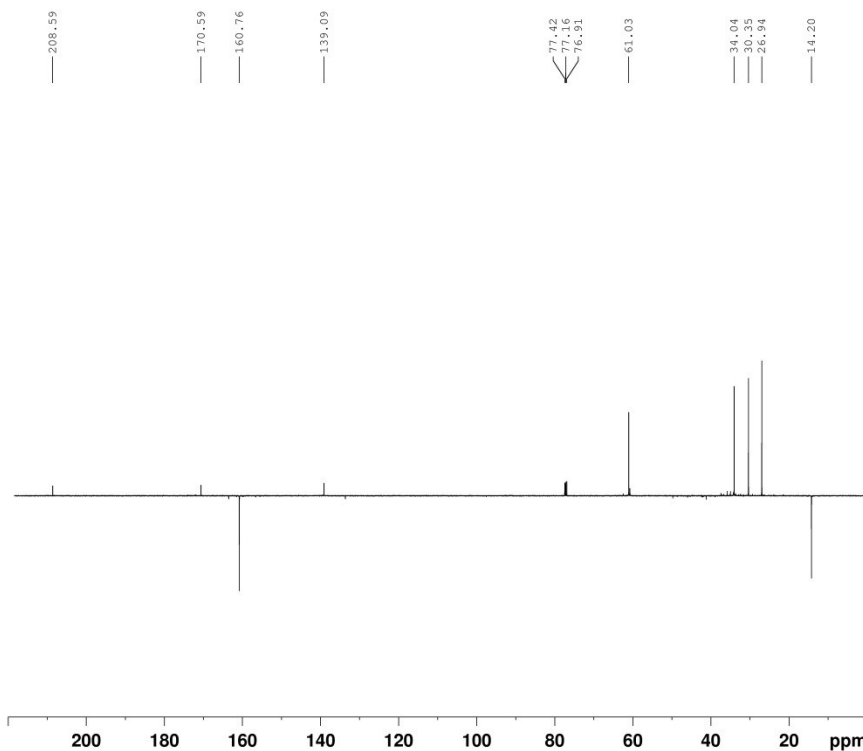


```

NAME      DNT_218_pure
EXPNO     1
PROCNO    1
Date_     20140212
Time      4.00
INSTRUM   spect
PROBHD    5 mm CPTCI 1H-
PULPROG   zg30
TD         65536
SOLVENT   CDC13
NS         3
DS         2
SWH        10330.578 Hz
FIDRES     0.157632 Hz
AQ         3.1719923 sec
RG         12.7
DW         48.400 usec
DE         6.50 usec
TE         293.3 K
D1         1.00000000 sec
TD0        1
  
```

```

===== CHANNEL f1 =====
NUC1      1H
P1         6.70 usec
PL1        4.00 dB
PL1W       8.72000027 W
SFO1      500.230891 MHz
SI         32768
SF         500.2300107 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00
  
```



```

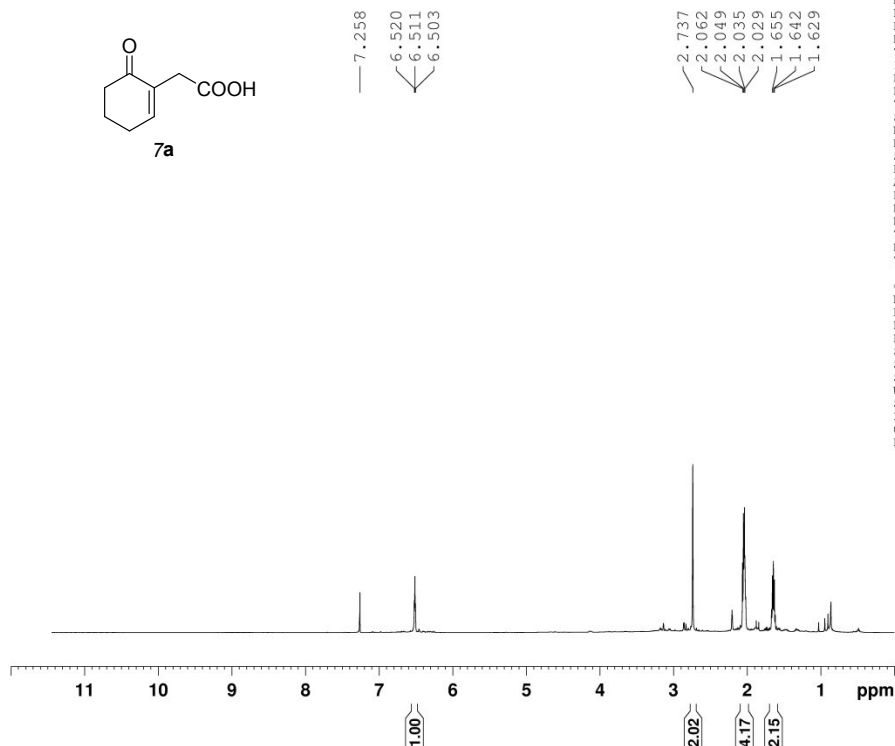
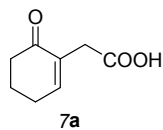
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EXPNO     2
PROCNO    1
Date_     20140212
Time      4.15
INSTRUM   spect
PROBHD    5 mm CPTCI 1H-
PULPROG   jmod
TD         65536
SOLVENT   CDC13
NS         256
DS         4
SWH        29761.904 Hz
FIDRES     0.454131 Hz
AQ         1.1010548 sec
RG         2050
DW         16.800 usec
DE         6.50 usec
TE         293.1 K
CNST2     145.0000000
CNST11    1.0000000
D1         2.00000000 sec
D20        0.00689655 sec
TD0        1
  
```

```

===== CHANNEL f1 =====
NUC1      13C
P1         11.20 usec
P2         22.40 usec
PL1        -2.00 dB
PL1W       88.77790070 W
SFO1      125.7955118 MHz
  
```

```

===== CHANNEL f2 =====
CPDPRG2   waltz16
NUC2       1H
PCPD2      80.00 usec
PL2        4.00 dB
PL12       25.28 dB
PL2W       8.72000027 W
PL12W      0.06494062 W
SFO2      500.2320009 MHz
SI         32768
SF         125.7829245 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         1.40
  
```



```

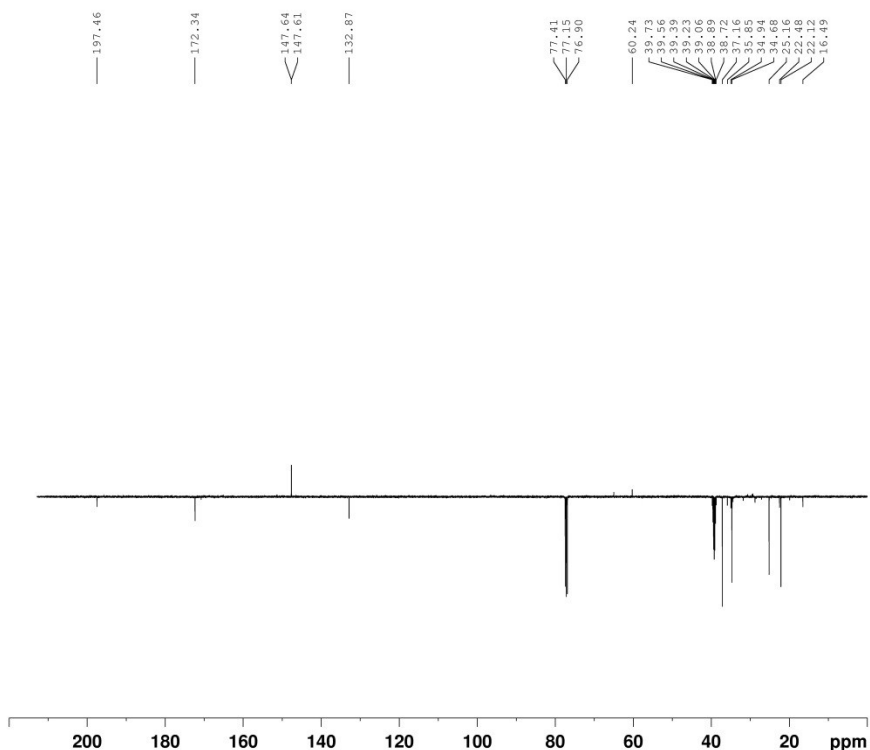
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EXPNO     4
PROCNO    1
Date_     20151002
Time      11.00
INSTRUM   spect
PROBHD    5 mm CPTCI 1H-
PULPROG   zg30
TD         65536
SOLVENT   CDCl3
NS         16
DS         2
SWH        10330.578 Hz
FIDRES     0.157632 Hz
AQ         3.1719923 sec
RG         14.2
DW         48.400 usec
DE         6.50 usec
TE         298.2 K
D1         1.00000000 sec
TD0        1

```

```

===== CHANNEL f1 =====
NUC1      1H
P1         6.70 usec
PL1        4.00 dB
PL1W      8.72000027 W
SFO1      500.2330891 MHz
SI         32768
SF         500.2325294 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00

```



```

NAME      RCC_977_pure
EXPNO     5
PROCNO    1
Date_     20151002
Time      11.14
INSTRUM   spect
PROBHD    5 mm CPTCI 1H-
PULPROG   jmod
TD         65536
SOLVENT   CDCl3
NS         256
DS         4
SWH        29761.904 Hz
FIDRES     0.454131 Hz
AQ         1.1010548 sec
RG         2050
DW         16.800 usec
DE         6.50 usec
TE         298.2 K
CNST2     145.0000000
CNST11    1.0000000
D1         2.00000000 sec
D20        0.00689655 sec
TD0        1

```

```

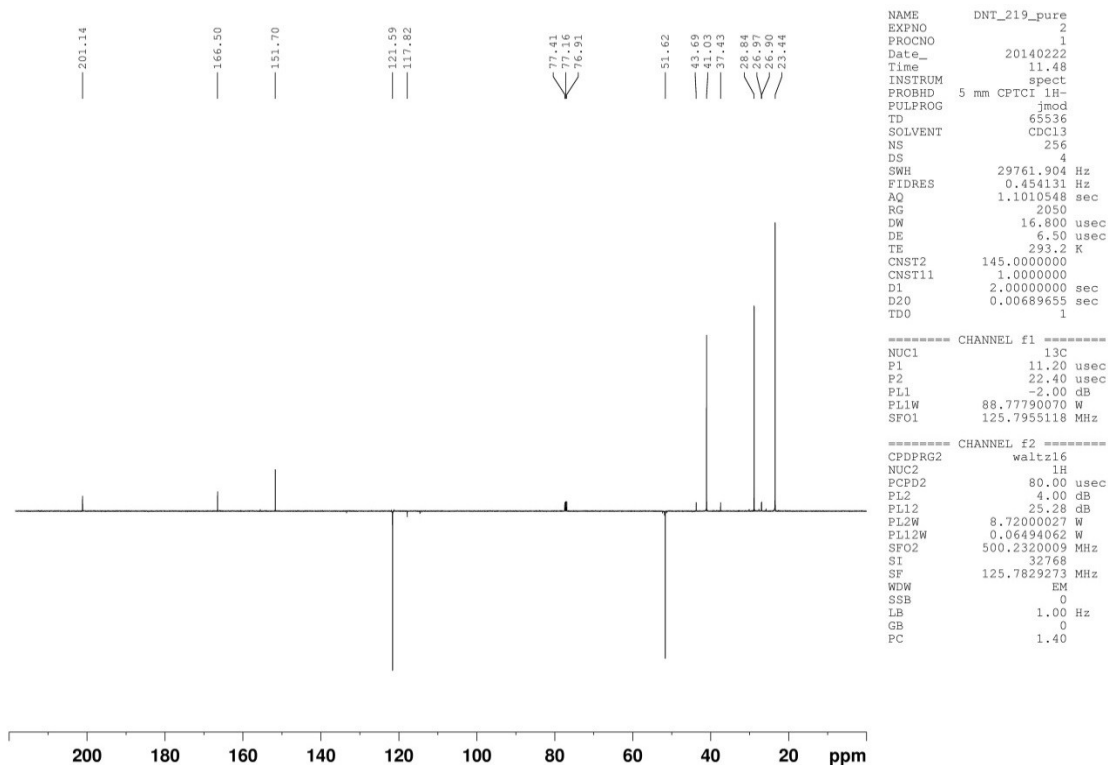
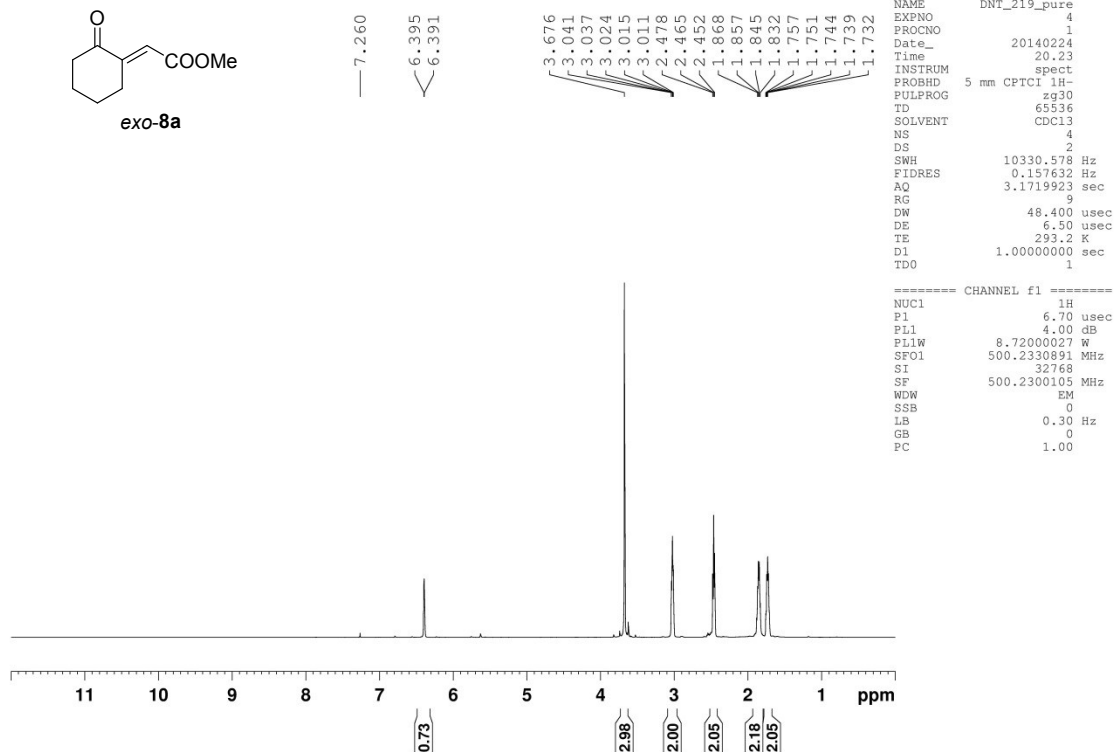
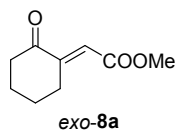
===== CHANNEL f1 =====
NUC1      13C
P1         11.20 usec
P2         22.40 usec
PL1        -2.00 dB
PL1W      88.77790070 W
SFO1      125.7955118 MHz

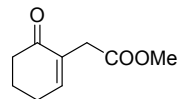
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```

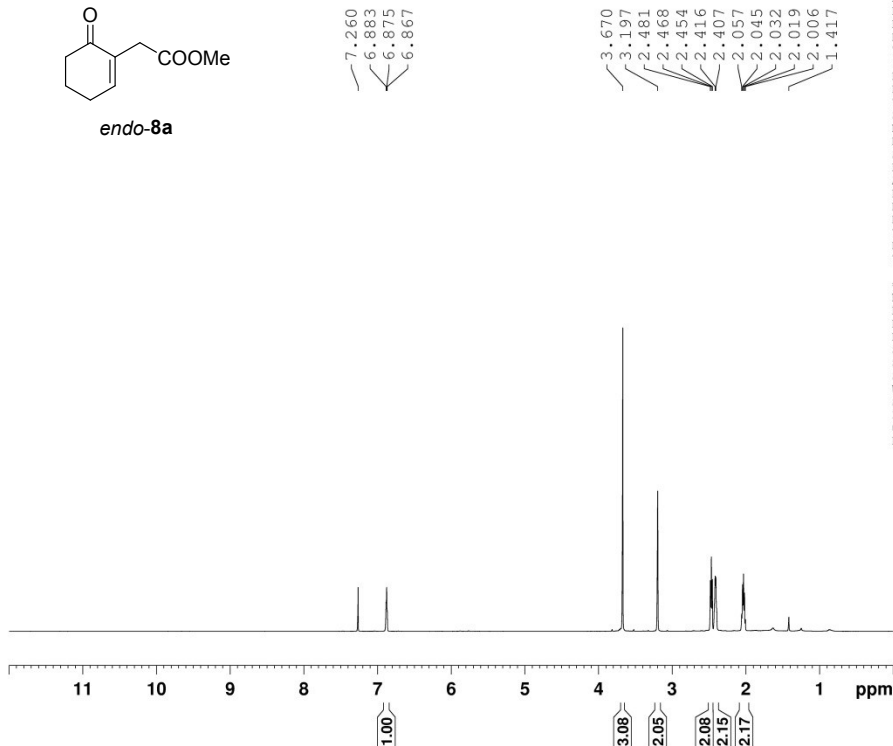
===== CHANNEL f2 =====
CPDPRG2   waltz16
NUC2      1H
PCPD2     80.00 usec
PL2        4.00 dB
PL12      25.28 dB
PL2W      8.72000027 W
PL12W     0.06494062 W
SFO2      500.2320009 MHz
SI         32768
SF         125.7836322 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         1.40

```





endo-8a



```

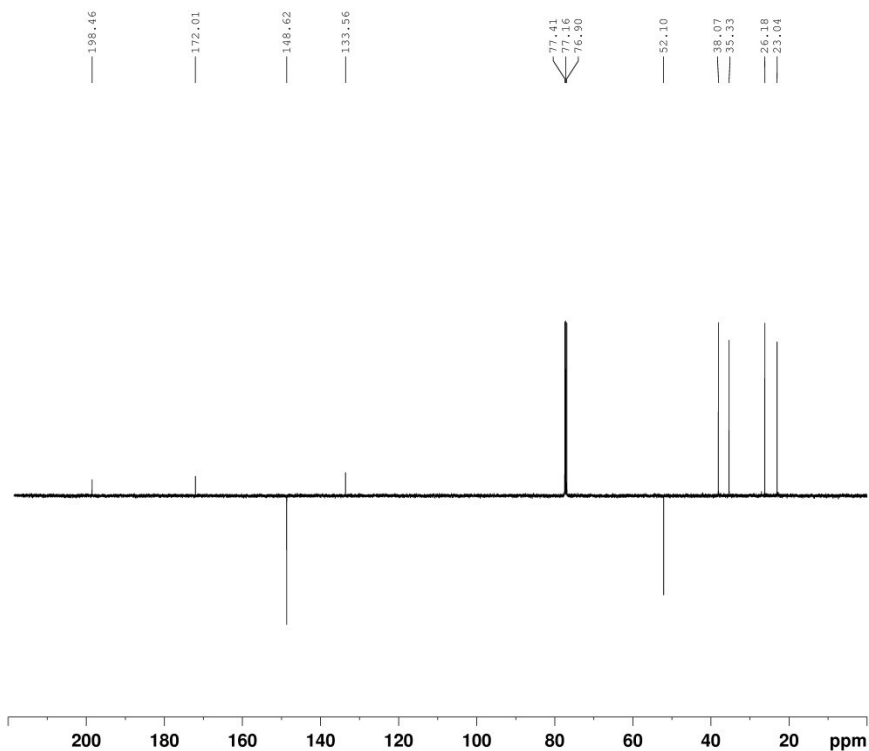
NAME      DNT_228_pure
EXPNO     1
PROCNO    1
Date_     20140304
Time      21.21
INSTRUM   spect
PROBHD    5 mm CPTCI 1H-
PULPROG   zg30
TD         65536
SOLVENT   CDCl3
NS         16
DS         2
SWH        10330.578 Hz
FIDRES     0.157632 Hz
AQ         3.1719923 sec
RG         16
DW         48.400 usec
DE         6.50 usec
TE         293.2 K
D1         1.00000000 sec
D10        1

```

```

===== CHANNEL f1 =====
NUC1      1H
P1         6.70 usec
PL1        4.00 dB
PL1W      8.72000027 W
SFO1      500.2330891 MHz
SI        32768
SF        500.2300112 MHz
WDW        EM
SSB         0
LB         0.30 Hz
GB         0
PC         1.00

```



```

NAME      DNT_228_pure
EXPNO     3
PROCNO    1
Date_     20140304
Time      21.35
INSTRUM   spect
PROBHD    5 mm CPTCI 1H-
PULPROG   jmod
TD         65536
SOLVENT   CDCl3
NS         256
DS         4
SWH        29761.904 Hz
FIDRES     0.454131 Hz
AQ         1.1010548 sec
RG         2050
DW         16.800 usec
DE         6.50 usec
TE         293.2 K
CNST2     145.0000000
CNST11    1.0000000
D1         2.00000000 sec
D20        0.00689655 sec
D10        1

```

```

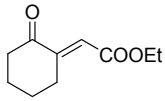
===== CHANNEL f1 =====
NUC1      13C
P1         11.20 usec
P2         22.40 usec
PL1        -2.00 dB
PL1W      88.77790070 W
SFO1      125.7955118 MHz

```

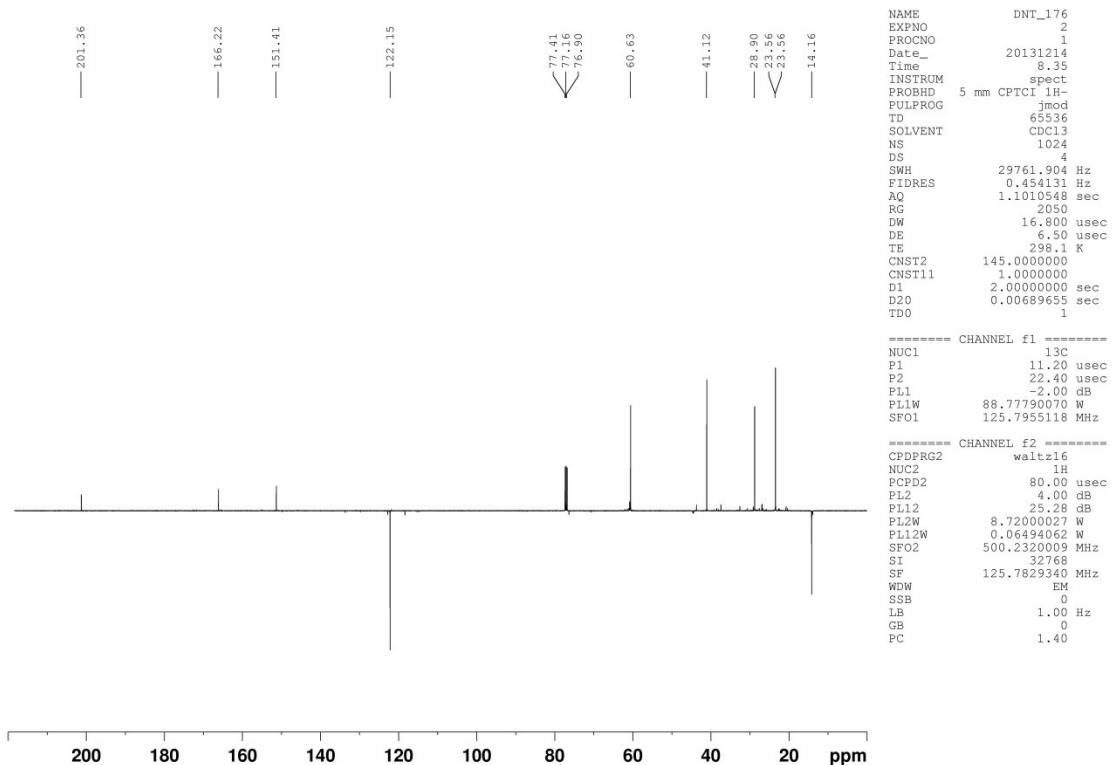
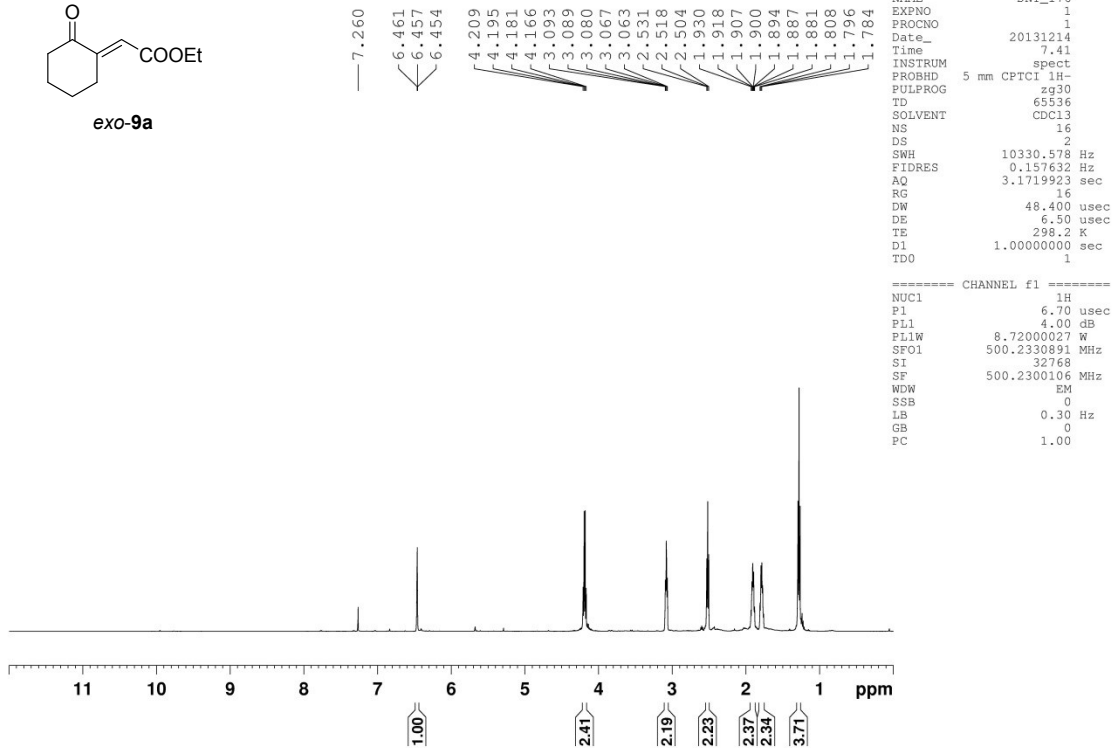
```

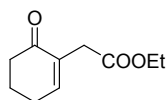
===== CHANNEL f2 =====
CPDPRG2   waltz16
NUC2      1H
PCPD2     80.00 usec
PL2        4.00 dB
PL12      25.28 dB
PL2W      8.72000027 W
PL12W     0.06494062 W
SFO2      500.2320009 MHz
SI        32768
SF        125.7829194 MHz
WDW        EM
SSB         0
LB         1.00 Hz
GB         0
PC         1.40

```

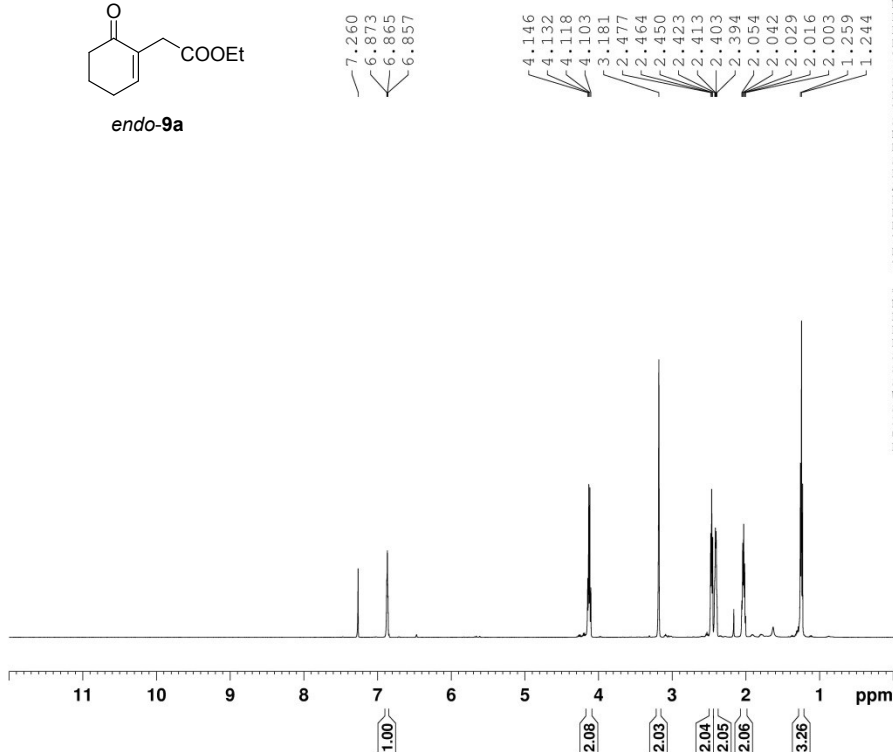


exo-9a





endo-9a



```

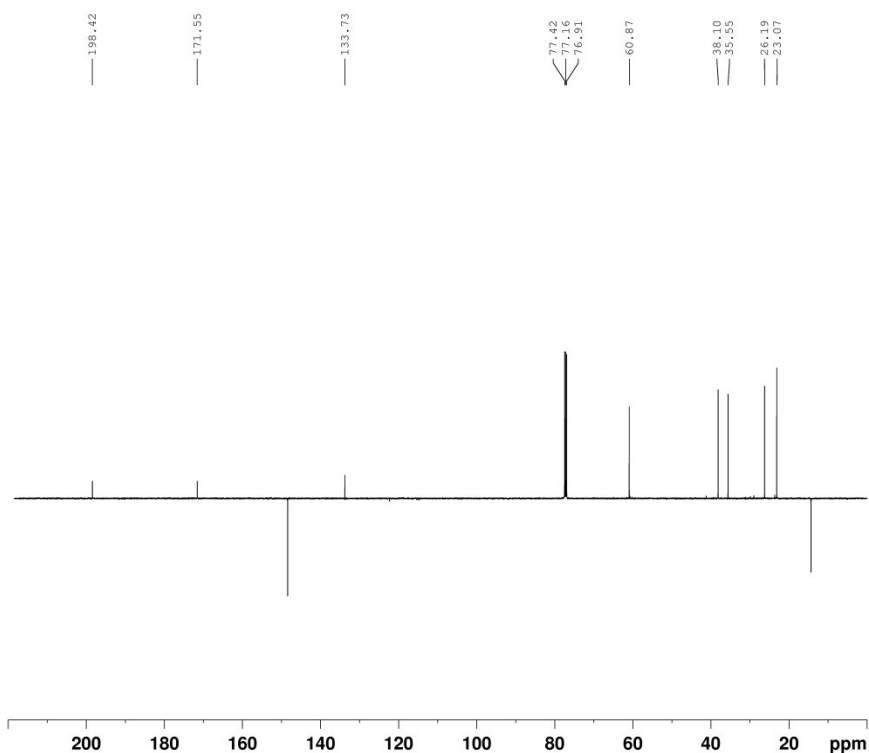
NAME          DNT_187
EXPNO         1
PROCNO        1
Date_         20131214
Time          8.44
INSTRUM       spect
PROBHD        5 mm CPTCI 1H-
PULPROG       zg30
TD            65536
SOLVENT       CDCl3
NS            16
DS            2
SWH           10330.578 Hz
FIDRES        0.157632 Hz
AQ            3.1719923 sec
RG            18
DW            48.400 usec
DE            6.50 usec
TE            298.2 K
D1            1.00000000 sec
D0            1

```

```

===== CHANNEL f1 =====
NUC1          1H
P1            6.70 usec
PL1           4.00 dB
PL1W          8.72000027 W
SFO1          500.2330891 MHz
SI            32768
SF            500.2300109 MHz
WDW           EM
SSB           0
LB            0.30 Hz
GB            0
PC            1.00

```



```

NAME          DNT_187
EXPNO         3
PROCNO        1
Date_         20131214
Time          9.39
INSTRUM       spect
PROBHD        5 mm CPTCI 1H-
PULPROG       jmod
TD            65536
SOLVENT       CDCl3
NS            1024
DS            4
SWH           29761.904 Hz
FIDRES        0.454131 Hz
AQ            1.1010548 sec
RG            2050
DW            16.800 usec
DE            6.50 usec
TE            298.2 K
CNST2         145.0000000
CNST11        1.0000000
D1            2.00000000 sec
D2            0.00689655 sec
D0            1

```

```

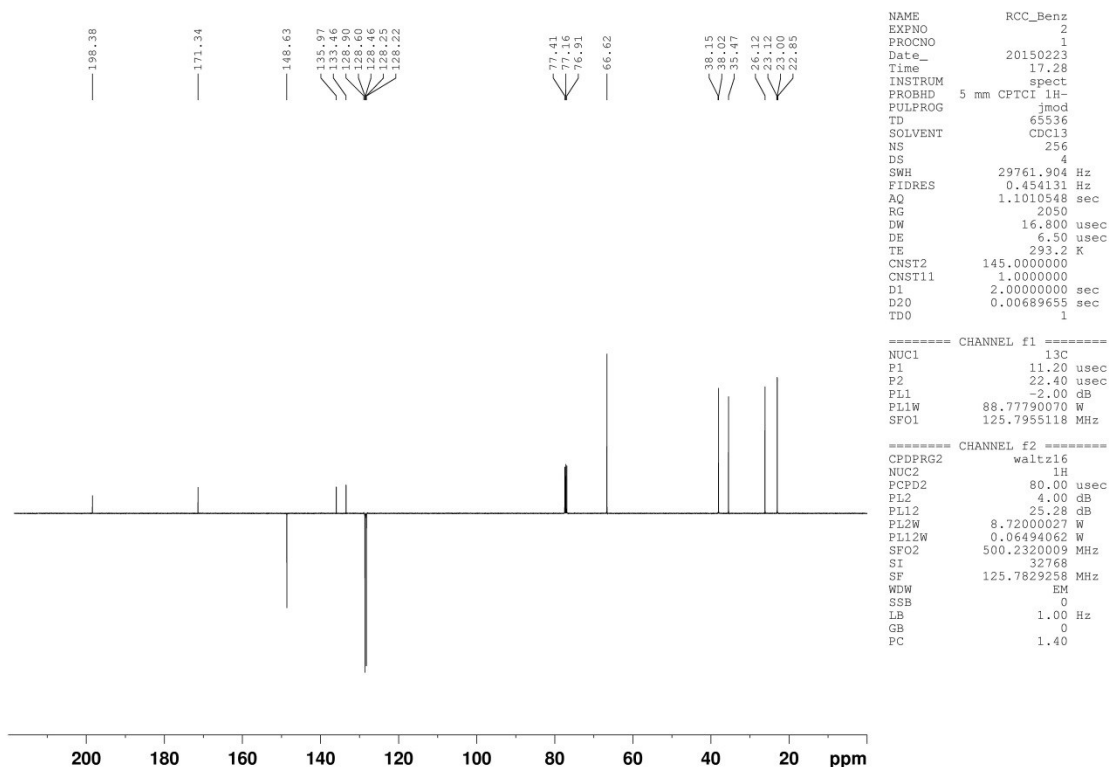
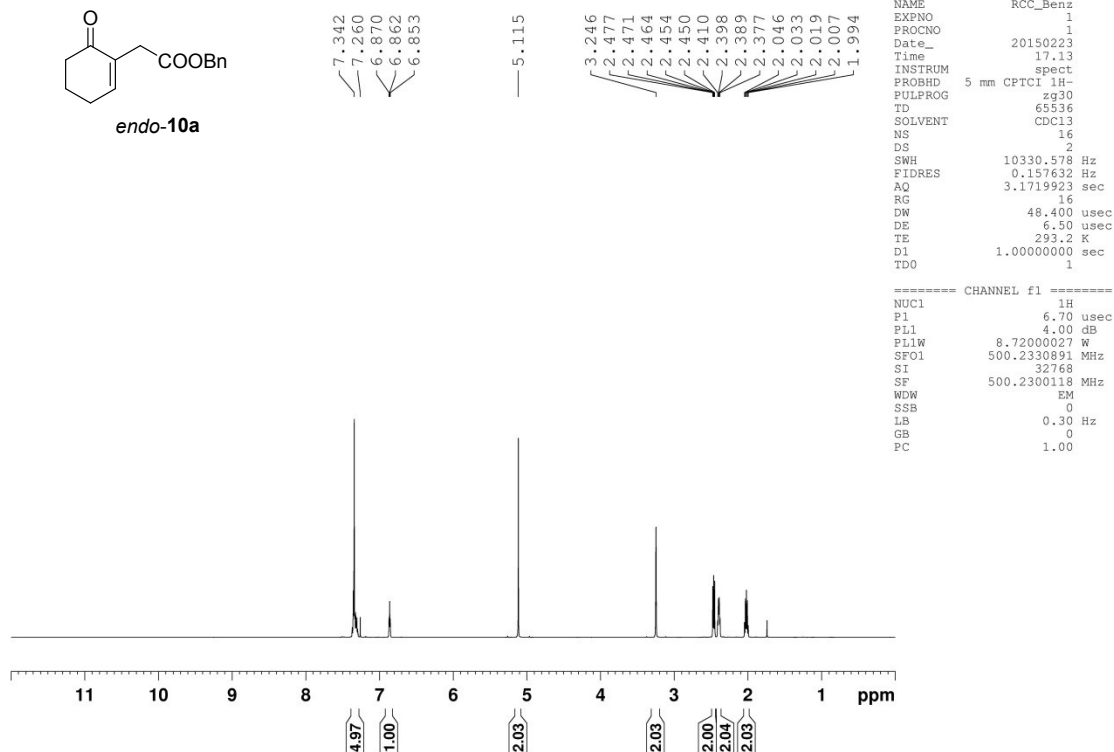
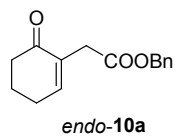
===== CHANNEL f1 =====
NUC1          13C
P1            11.20 usec
P2            22.40 usec
PL1           -2.00 dB
PL1W          88.77790070 W
SFO1          125.7955118 MHz

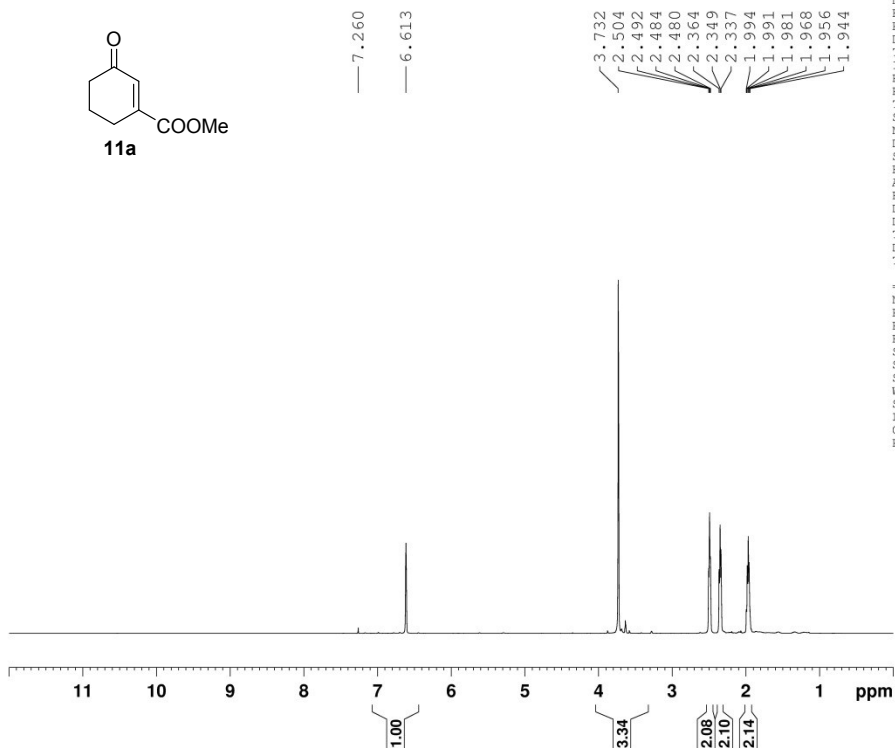
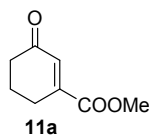
```

```

===== CHANNEL f2 =====
CPDPRG2       waltz16
NUC2          1H
PCPD2         80.00 usec
PL2           4.00 dB
PL12          25.28 dB
PL2W          8.72000027 W
PL12W         0.06494062 W
SFO2          500.2320009 MHz
SI            32768
SF            125.7829172 MHz
WDW           EM
SSB           0
LB            1.00 Hz
GB            0
PC            1.40

```



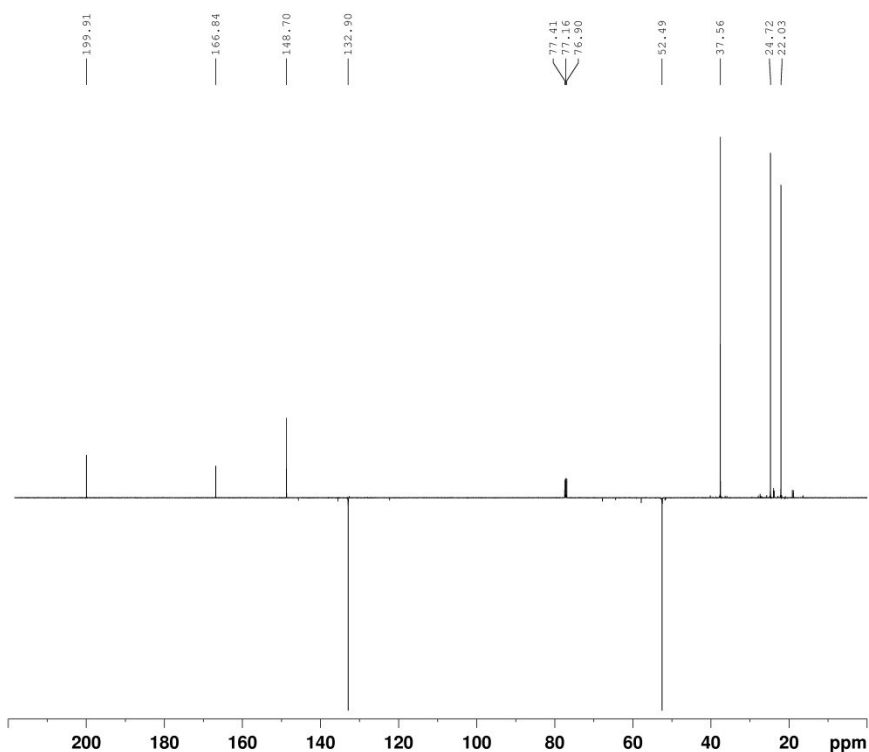


```

NAME      RCC_547_col
EXPNO     1
PROCNO    1
Date_     20140207
Time      18.40
INSTRUM   spect
PROBHD    5 mm CPTCI 1H-
PULPROG   zg30
TD         65536
SOLVENT   CDC13
NS         5
DS         2
SWH        10330.578 Hz
FIDRES     0.157632 Hz
AQ         3.1719923 sec
RG         9
DW         48.400 usec
DE         6.50 usec
TE         293.2 K
D1         1.00000000 sec
TD0        1
  
```

```

===== CHANNEL f1 =====
NUC1      1H
P1        6.70 usec
PL1       4.00 dB
PL1W      8.72000027 W
SFO1      500.2330891 MHz
SI        32768
SF        500.2300104 MHz
WDW       EM
SSB       0
LB        0.30 Hz
GB        0
PC        1.00
  
```



```

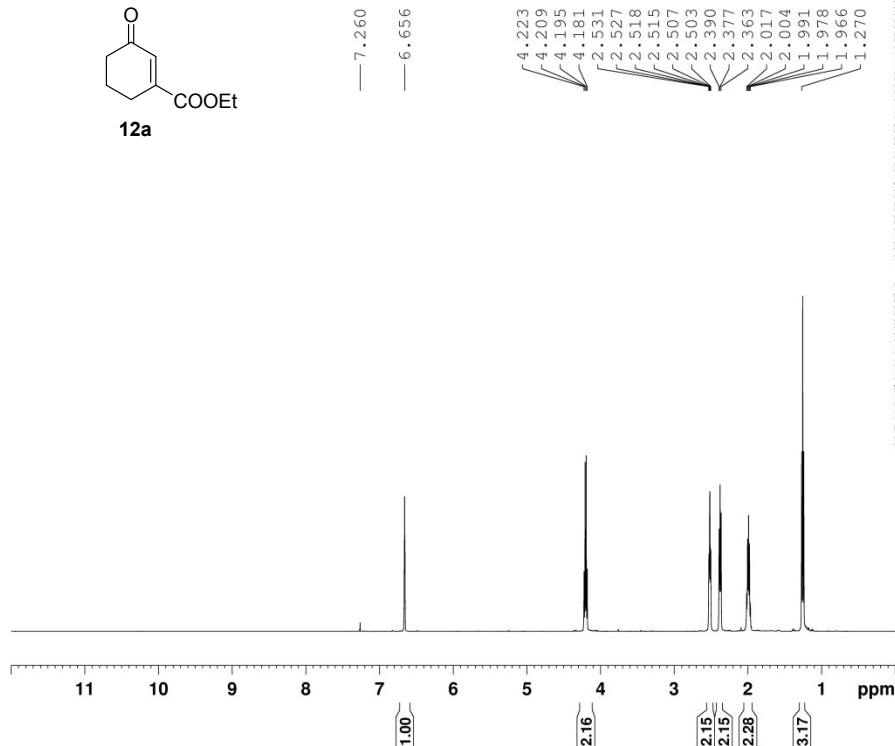
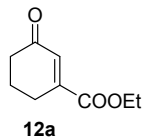
NAME      RCC_547_col
EXPNO     2
PROCNO    1
Date_     20140207
Time      18.55
INSTRUM   spect
PROBHD    5 mm CPTCI 1H-
PULPROG   jmod
TD         65536
SOLVENT   CDC13
NS         4
DS         4
SWH        29761.904 Hz
FIDRES     0.454131 Hz
AQ         1.1010548 sec
RG         2050
DW         16.800 usec
DE         6.50 usec
TE         293.2 K
CNST2     145.0000000
CNST11    1.0000000
D1         2.00000000 sec
D20        0.00689655 sec
TD0        1
  
```

```

===== CHANNEL f1 =====
NUC1      13C
P1        11.20 usec
P2        22.40 usec
PL1       -2.00 dB
PL1W      88.77790070 W
SFO1      125.7955118 MHz
  
```

```

===== CHANNEL f2 =====
CPDPRG2   waltz16
NUC2      1H
PCPD2     80.00 usec
PL2       4.00 dB
PL12      25.28 dB
PL2W      8.72000027 W
PL12W     0.06494062 W
SFO2      500.2320009 MHz
SI        32768
SF        125.7829377 MHz
WDW       EM
SSB       0
LB        1.00 Hz
GB        0
PC        1.40
  
```

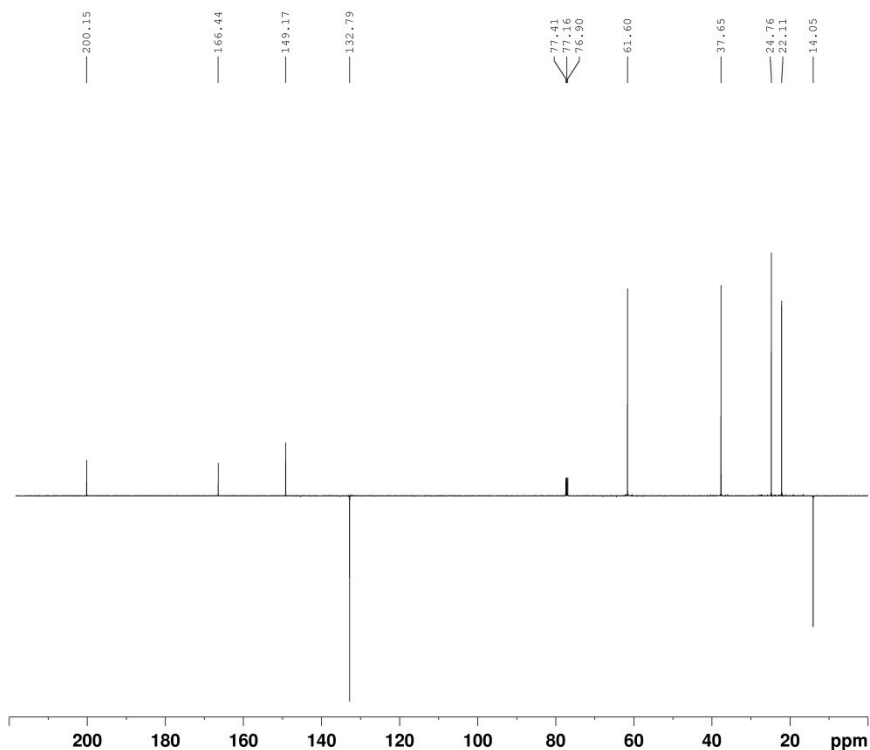


```

NAME      RCC_543_col
EXPNO     1
PROCNO    1
Date_     20140127
Time      15.32
INSTRUM   spect
PROBHD    5 mm CPTCI 1H-
PULPROG   zg30
TD         65536
SOLVENT   CDCl3
NS         3
DS         2
SWH        10330.578 Hz
FIDRES     0.157632 Hz
AQ         3.1719923 sec
RG         10
DW         48.400 usec
DE         6.50 usec
TE         293.2 K
D1         1.00000000 sec
TD0        1
  
```

```

===== CHANNEL f1 =====
NUC1      1H
P1         6.70 usec
PL1        4.00 dB
PL1W      8.72000027 W
SFO1      500.2330891 MHz
SI         32768
SF         500.2300103 MHz
WDW        EM
SSB         0
LB         0.30 Hz
GB         0
PC         1.00
  
```



```

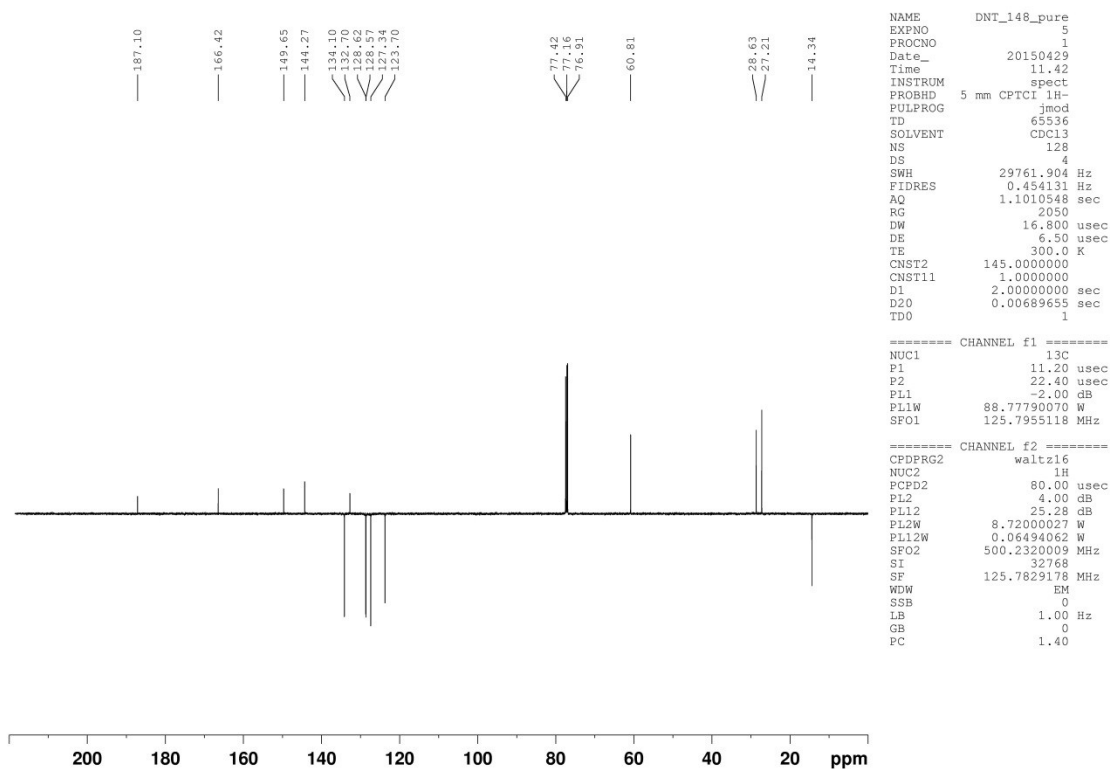
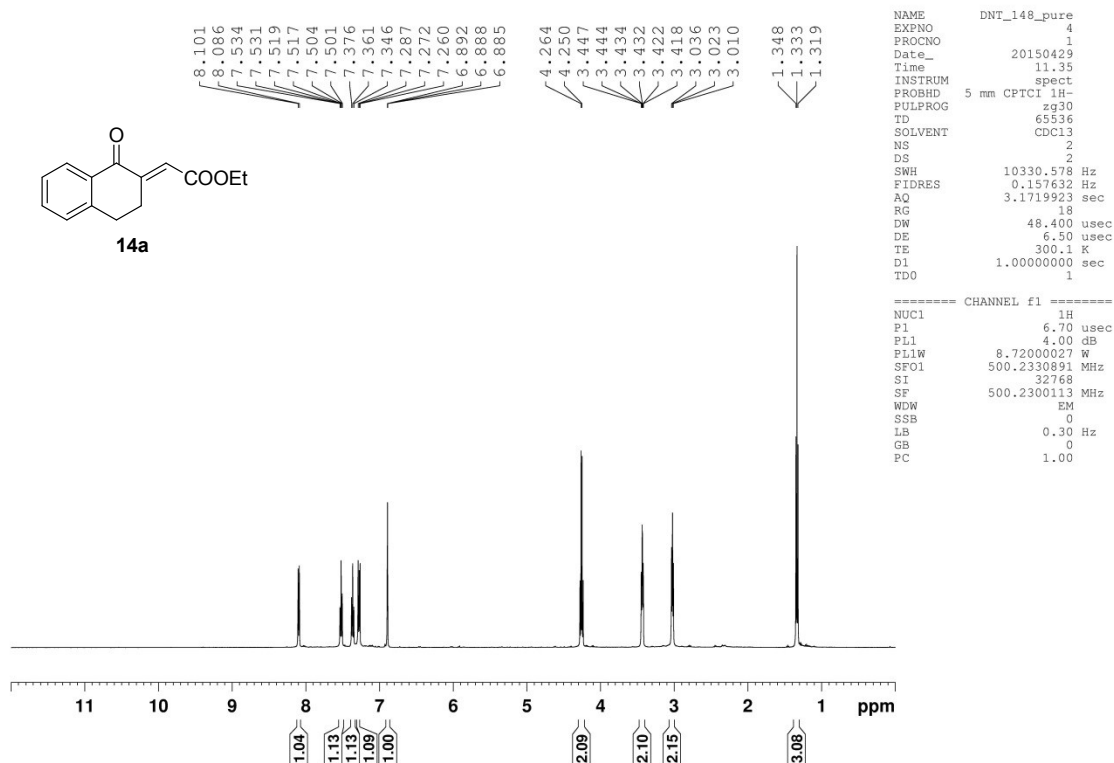
NAME      RCC_543_col
EXPNO     2
PROCNO    1
Date_     20140127
Time      15.47
INSTRUM   spect
PROBHD    5 mm CPTCI 1H-
PULPROG   jmod
TD         65536
SOLVENT   CDCl3
NS         256
DS         4
SWH        29761.904 Hz
FIDRES     0.454131 Hz
AQ         1.1010548 sec
RG         2050
DW         16.800 usec
DE         6.50 usec
TE         293.2 K
CNST2     145.0000000
CNST11    1.0000000
D1         2.00000000 sec
D20        0.00689655 sec
TD0        1
  
```

```

===== CHANNEL f1 =====
NUC1      13C
P1         11.20 usec
P2         22.40 usec
PL1        -2.00 dB
PL1W      88.77790070 W
SFO1      125.7955118 MHz
  
```

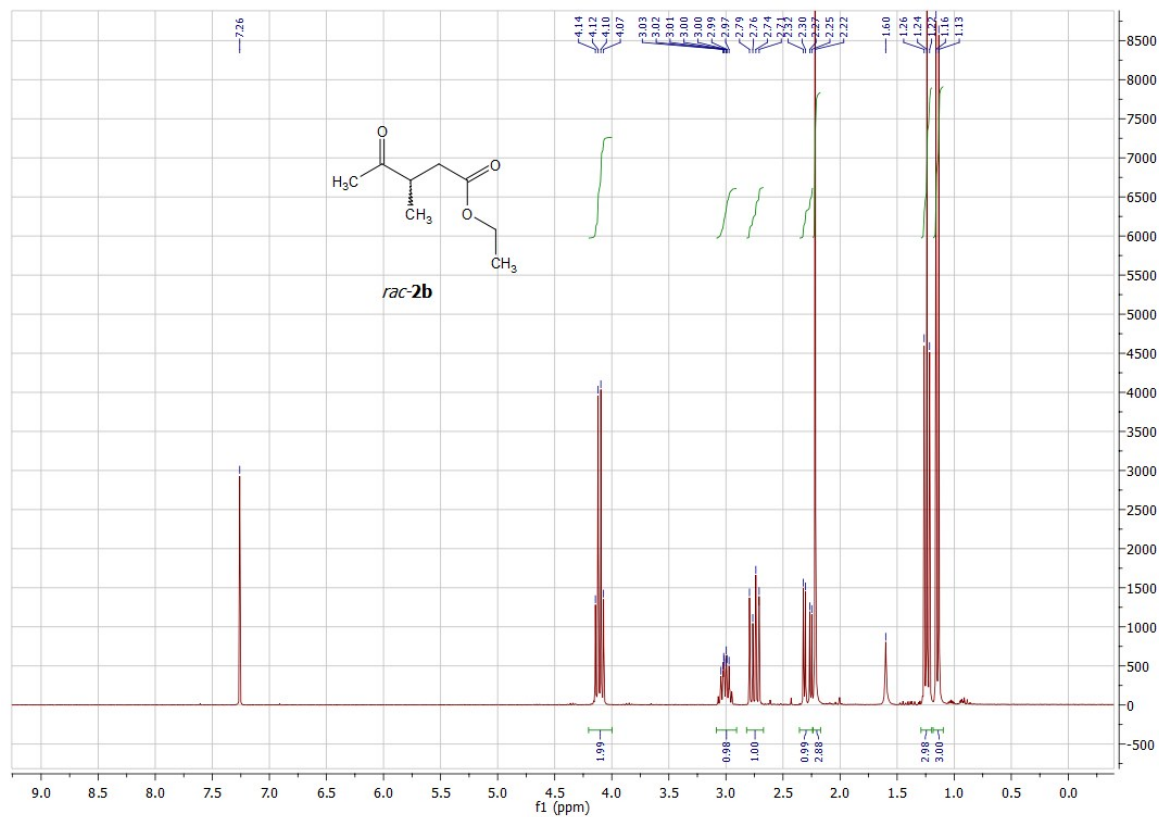
```

===== CHANNEL f2 =====
CPDPRG2   waitz16
NUC2      1H
PCPD2     80.00 usec
PL2        4.00 dB
PL12      25.28 dB
PL2W      8.72000027 W
PL12W     0.06494062 W
SFO2      500.2320009 MHz
SI         32768
SF         125.7829323 MHz
WDW        EM
SSB         0
LB         1.00 Hz
GB         0
PC         1.40
  
```

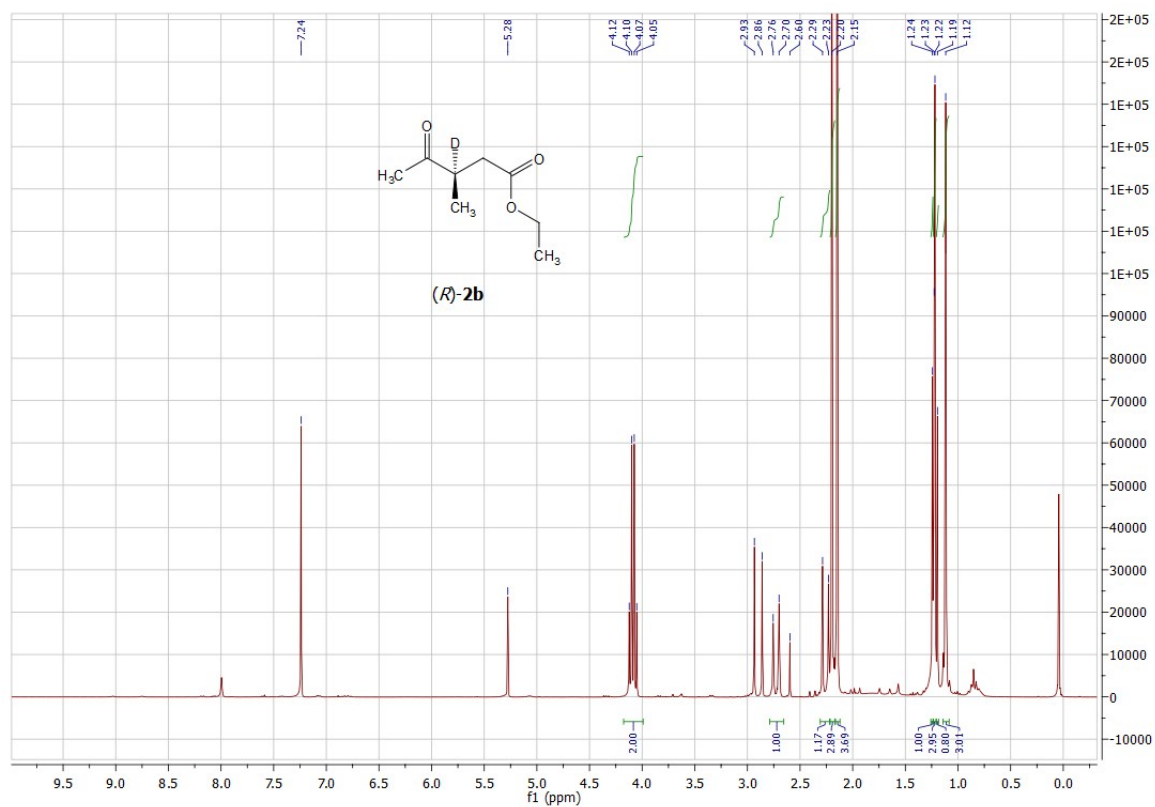


8. NMR spectra from isotopic labeling

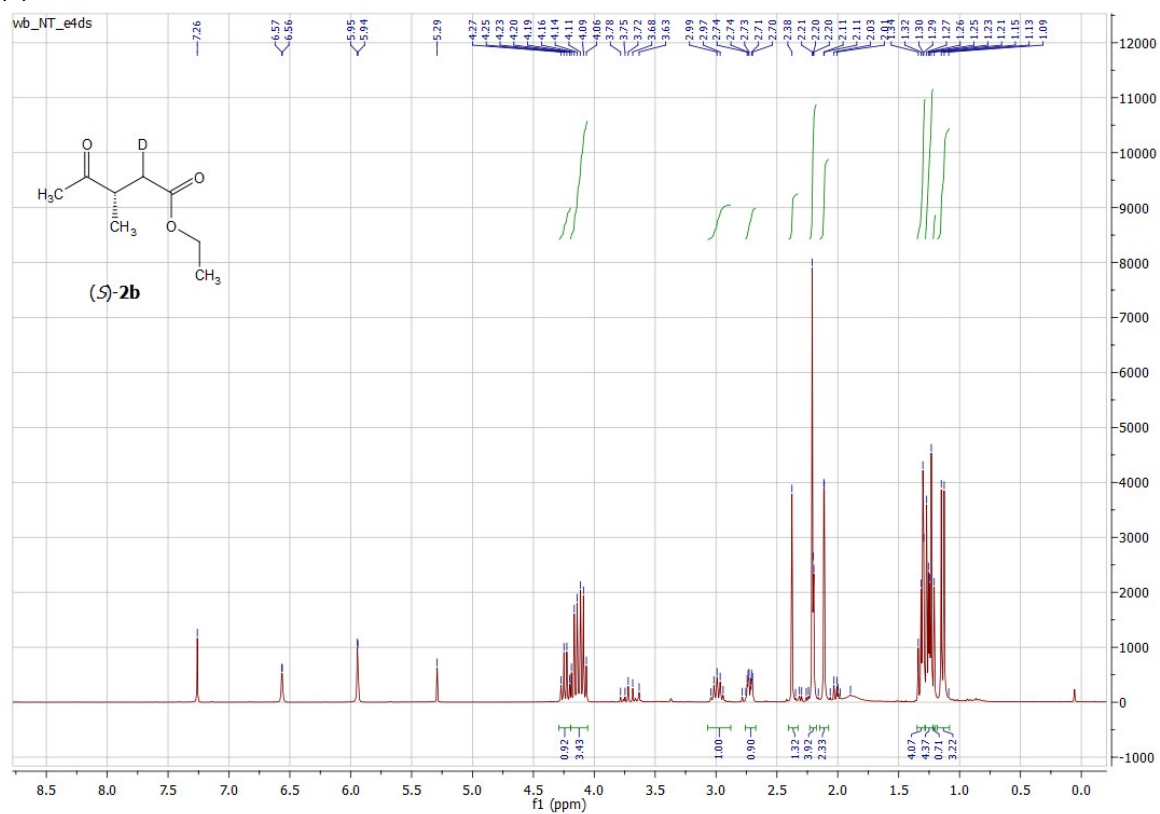
rac-2b



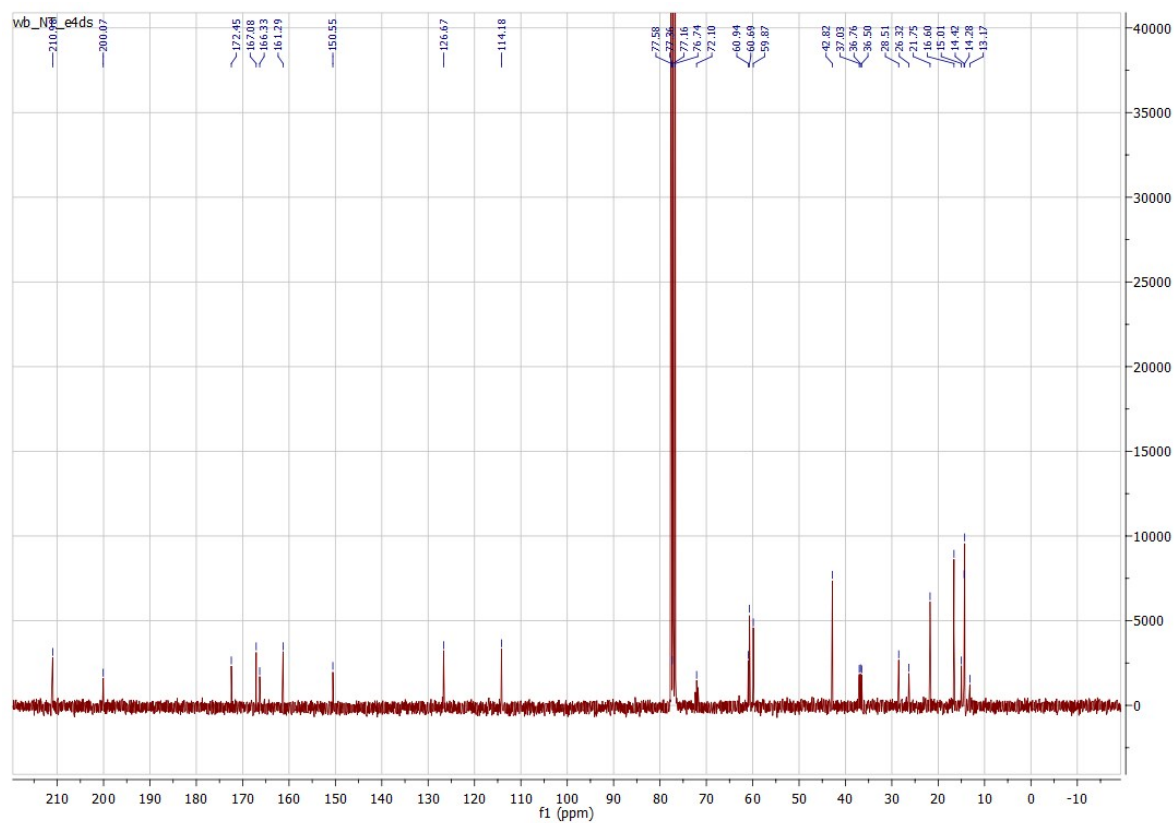
(*R*)-2b



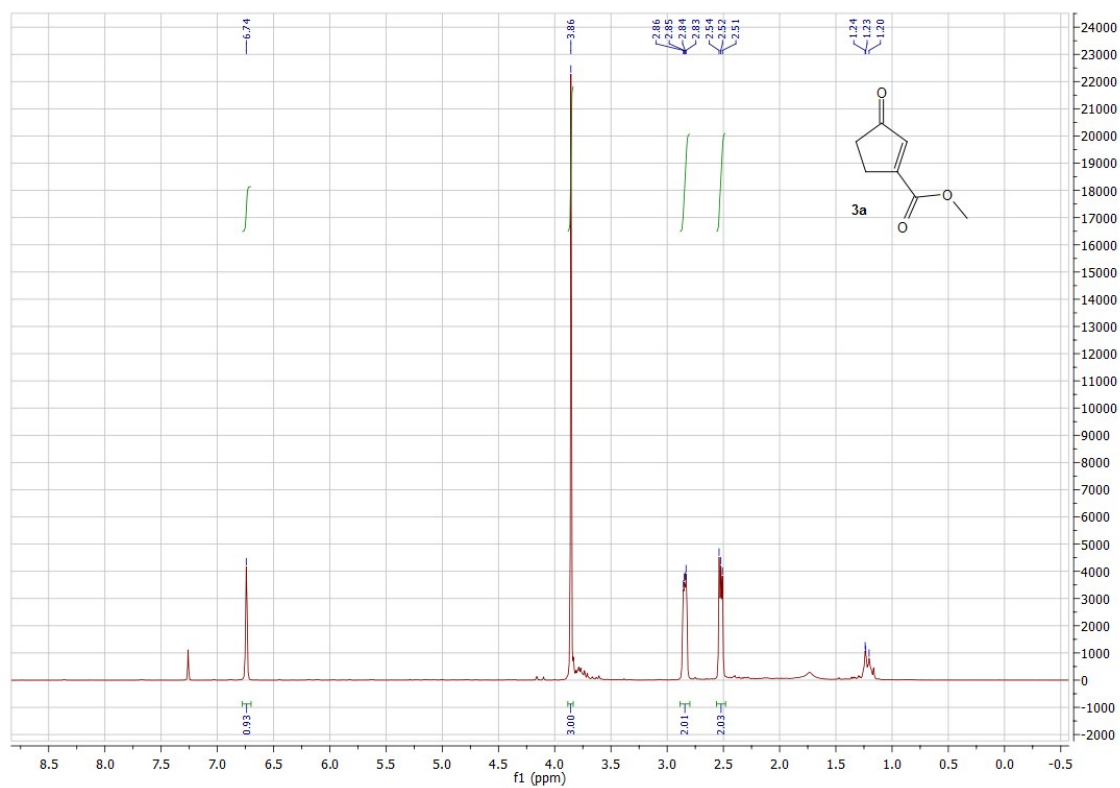
(S)-2b



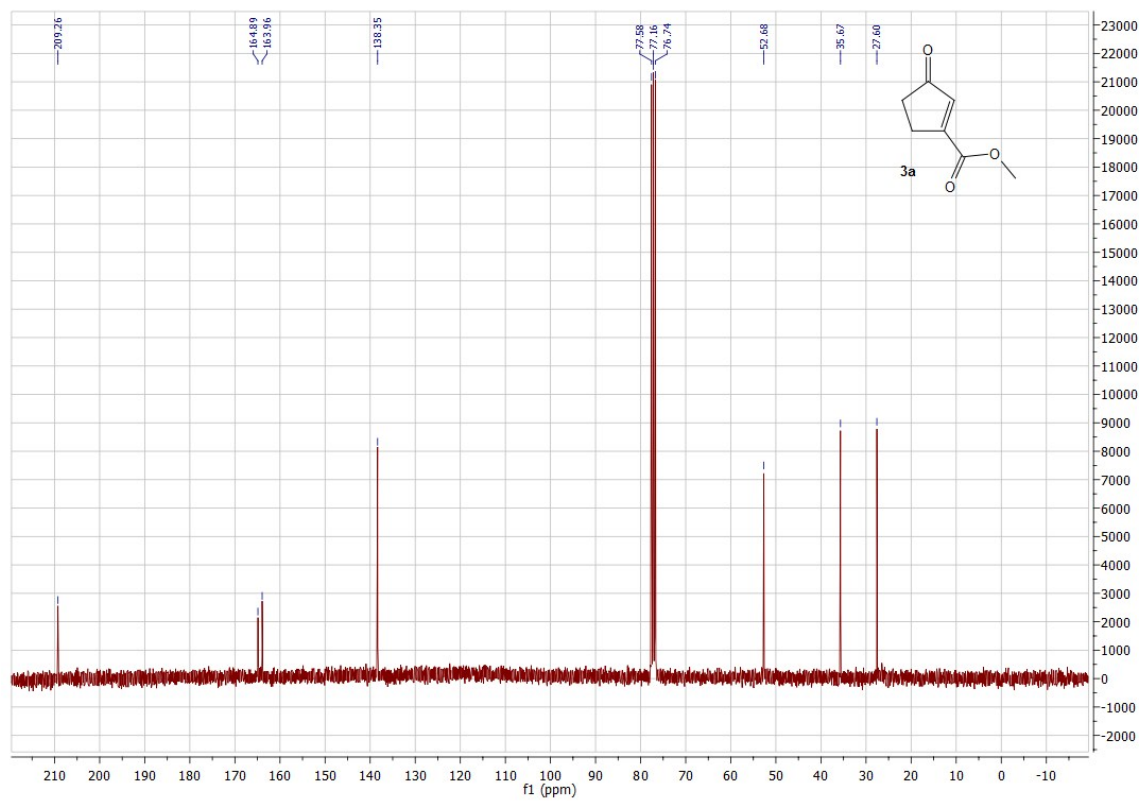
(S)-2b ¹³C



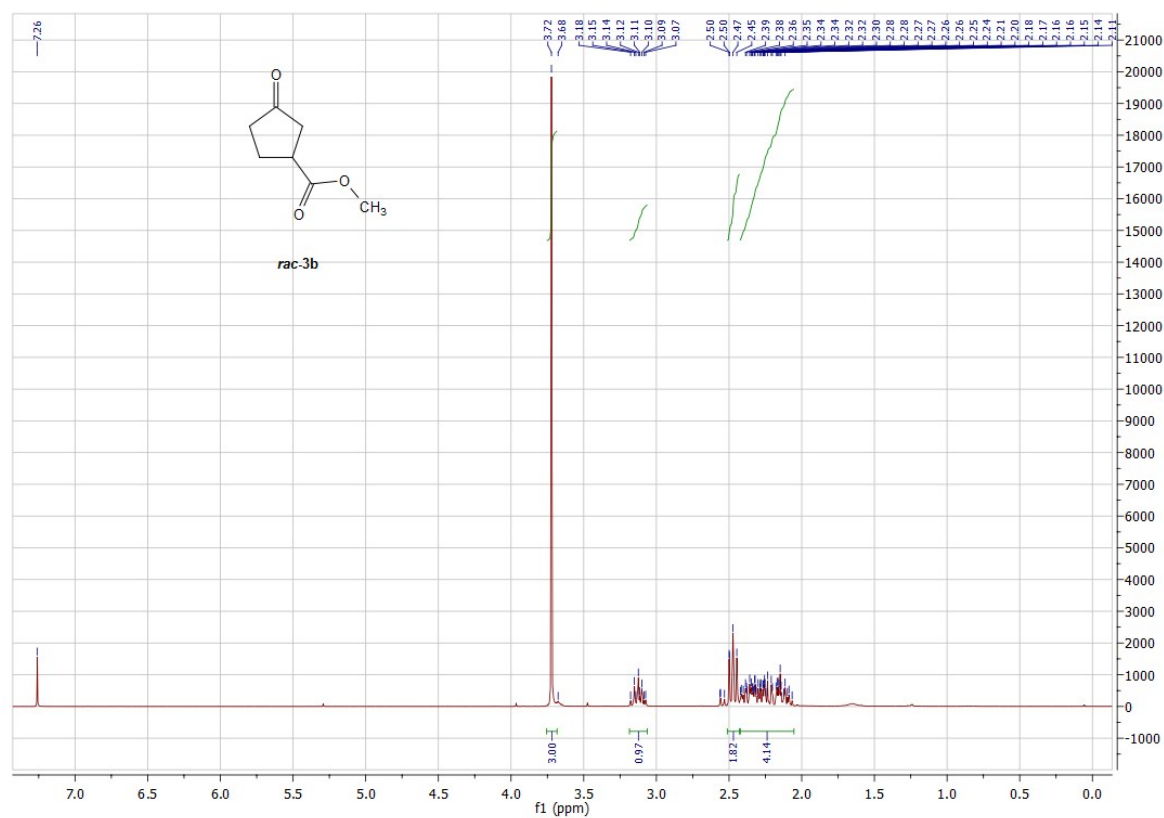
3a



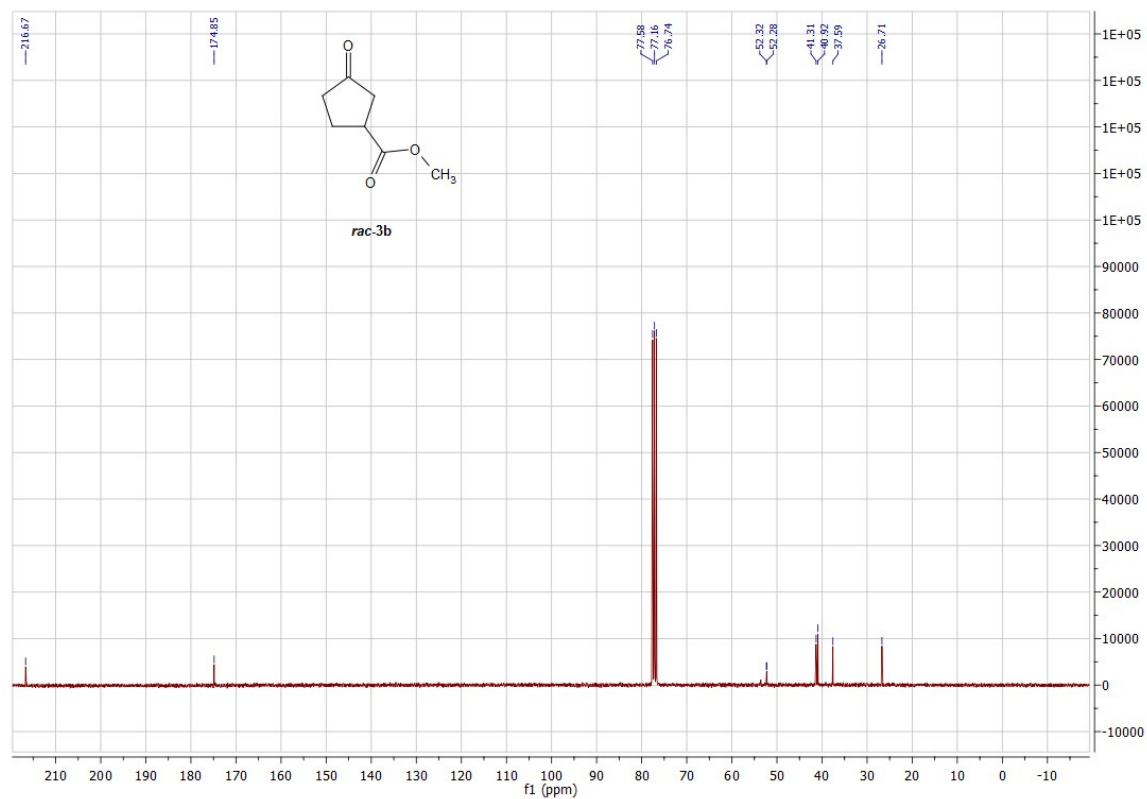
3a ¹³C



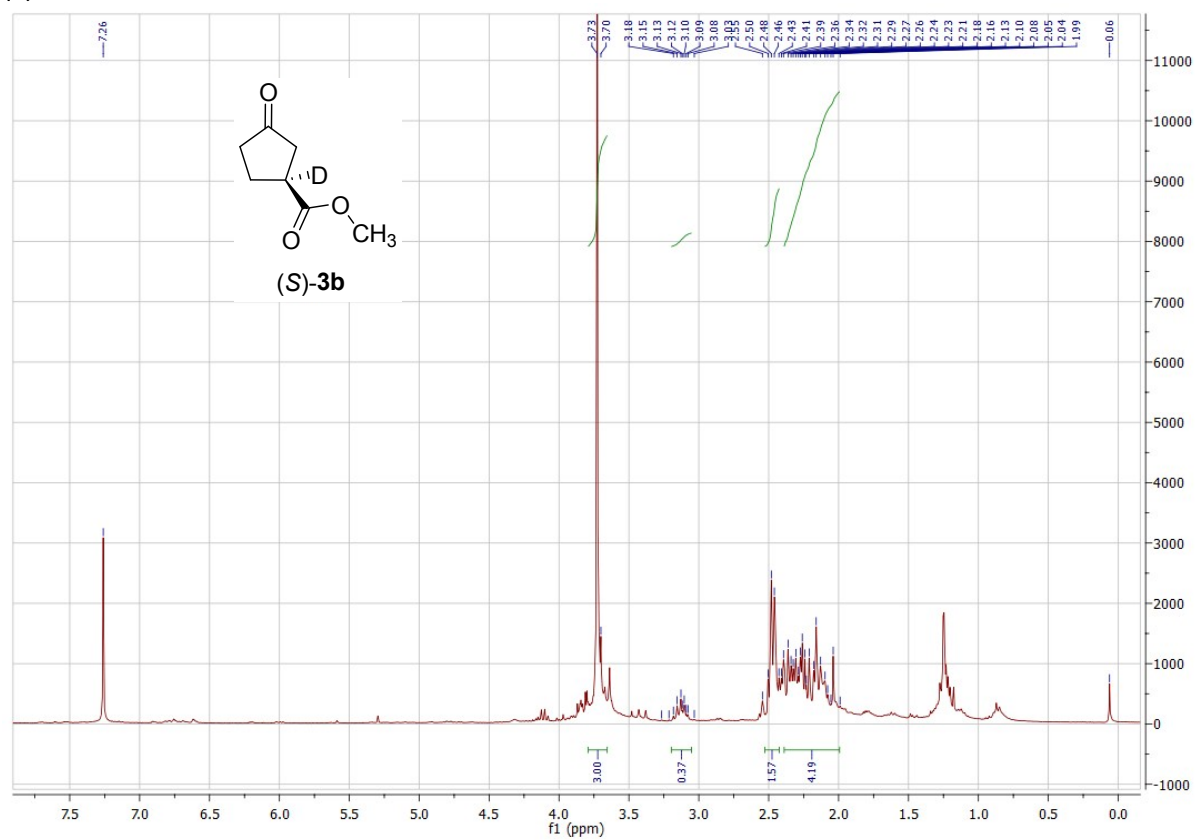
rac-3b



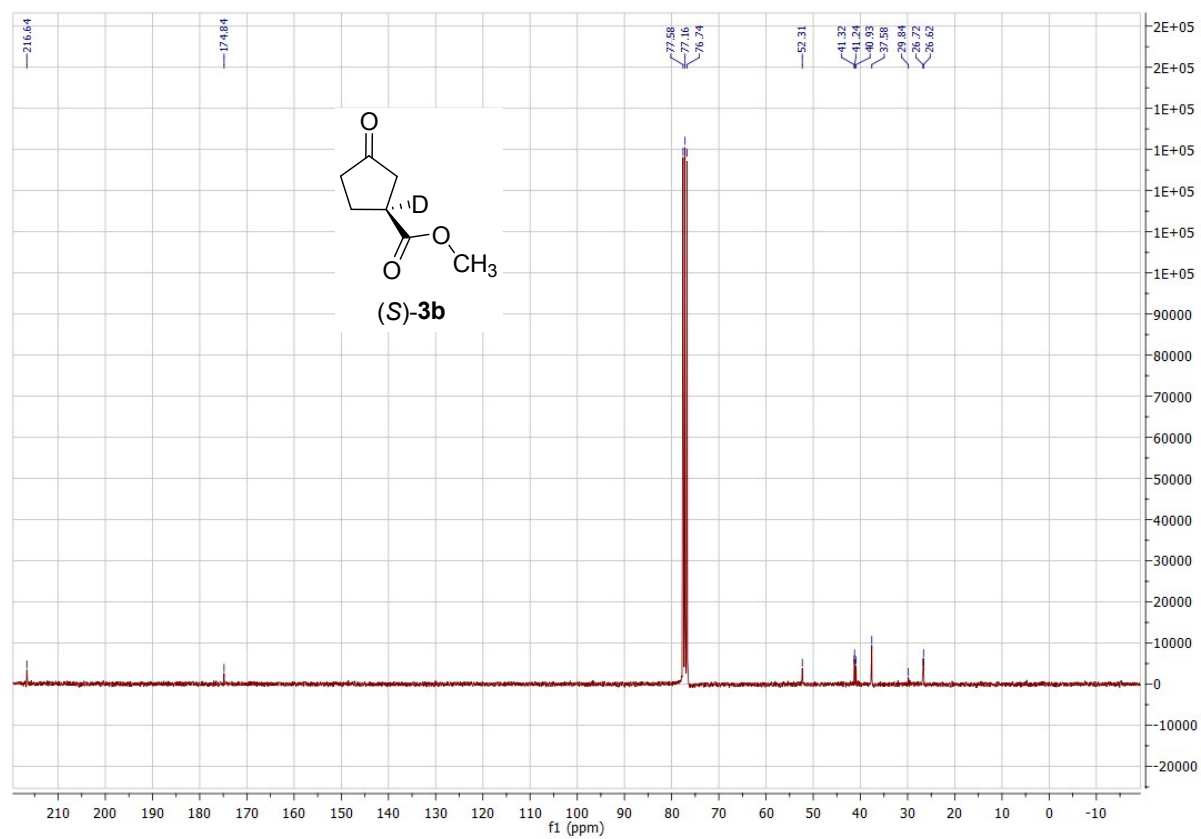
rac-3b ¹³C



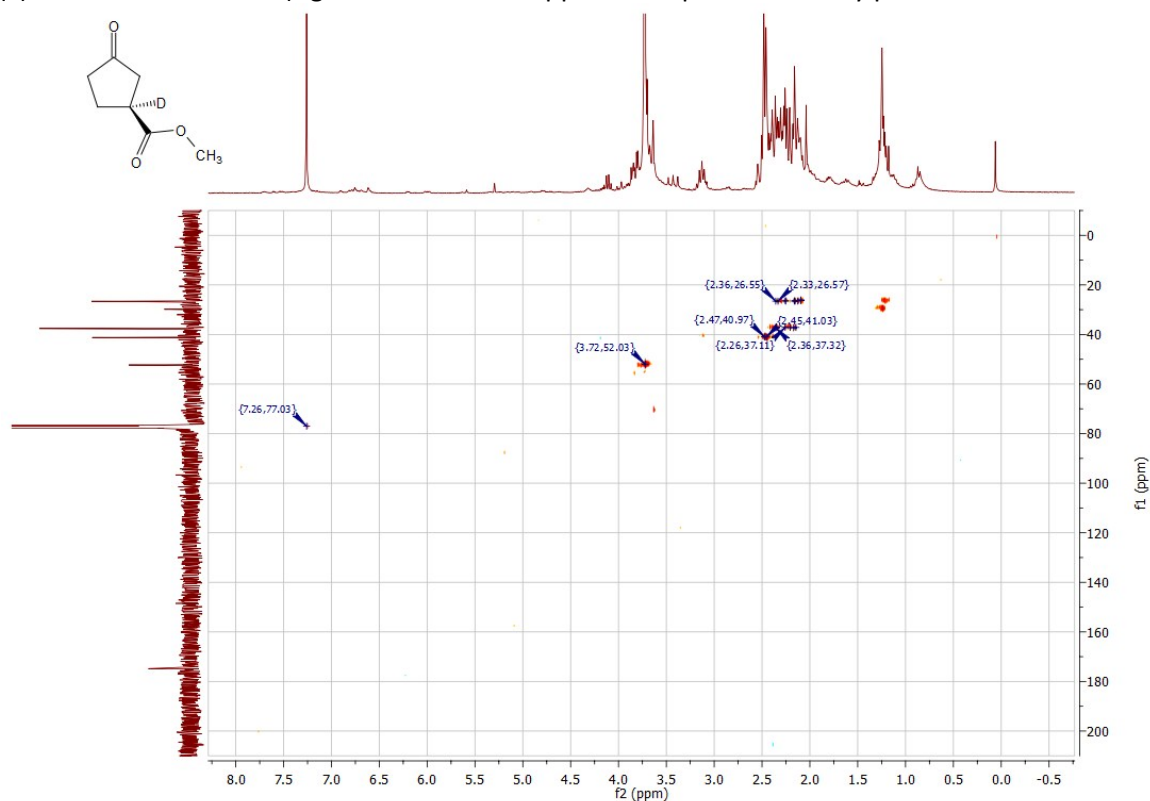
(S)-**3b** from OYE2



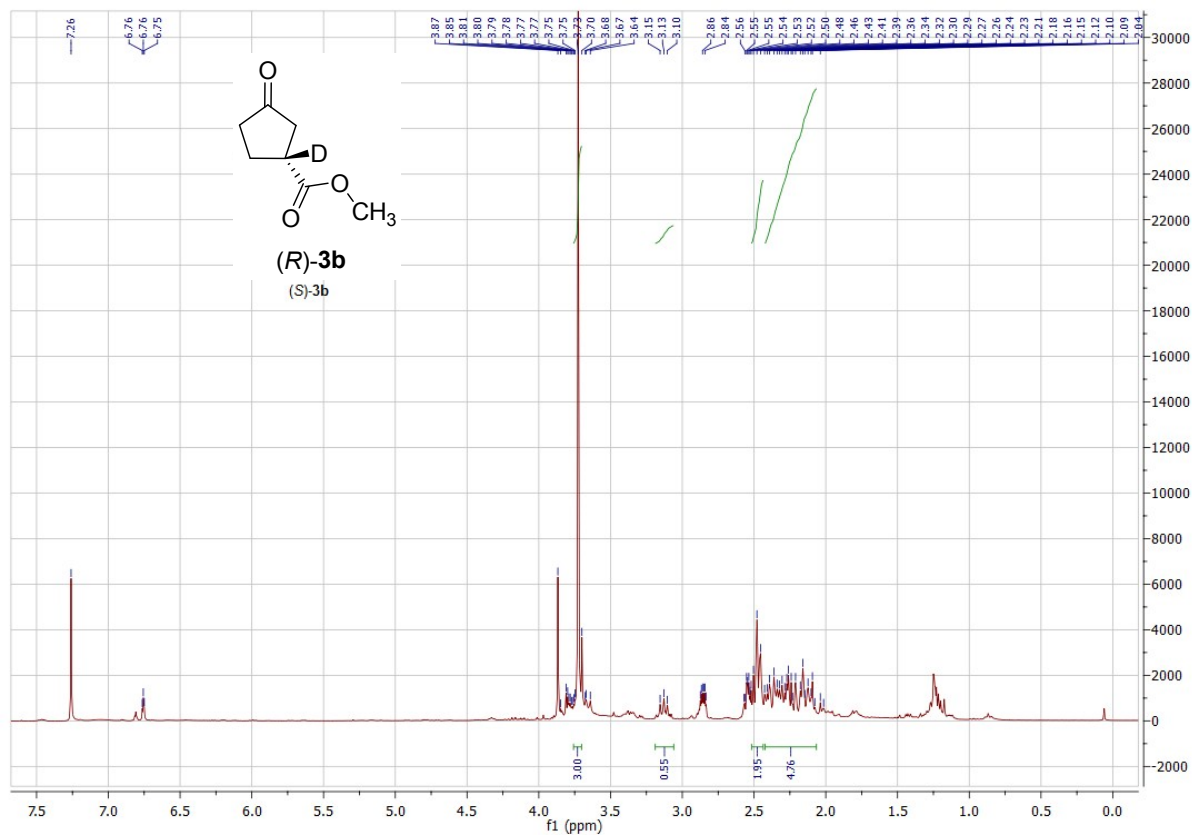
(S)-**3b** from OYE2 ¹³C



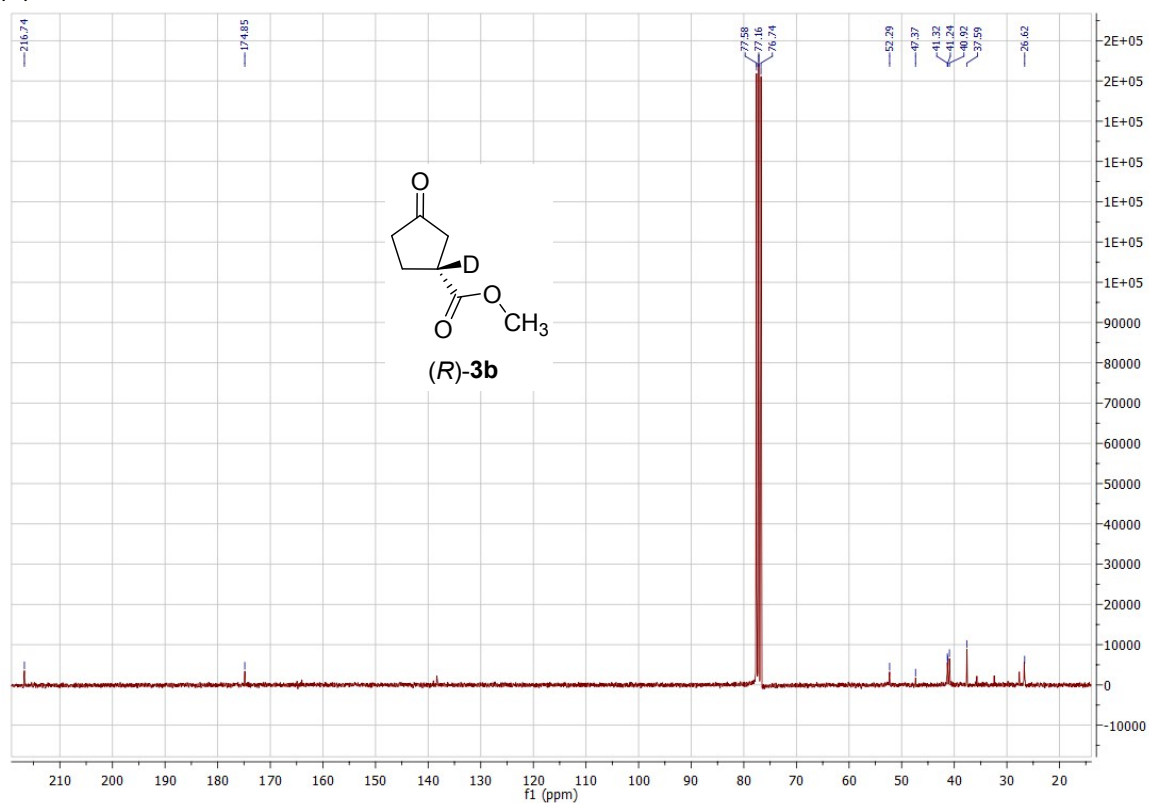
(S)-**3b** from OYE2 HSQC (signals at around 1.2 ppm are impurities already present in the substrate)



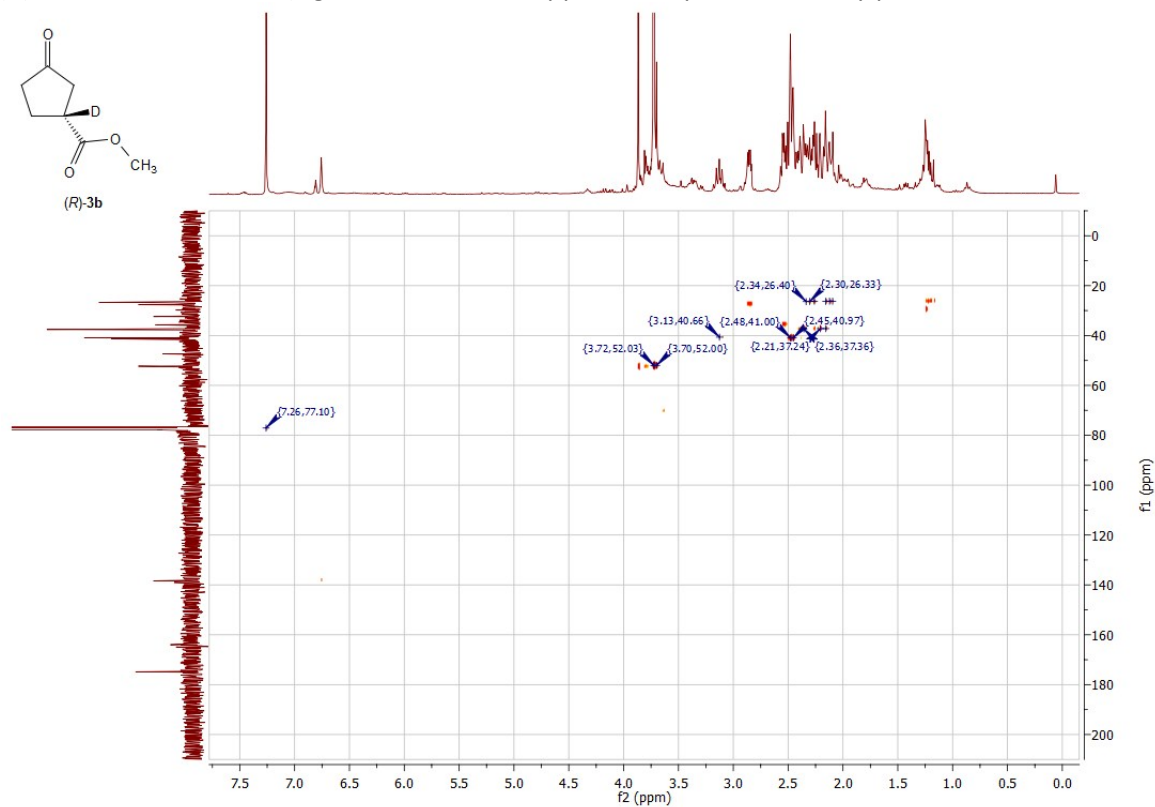
(R)-**3b** from Xena



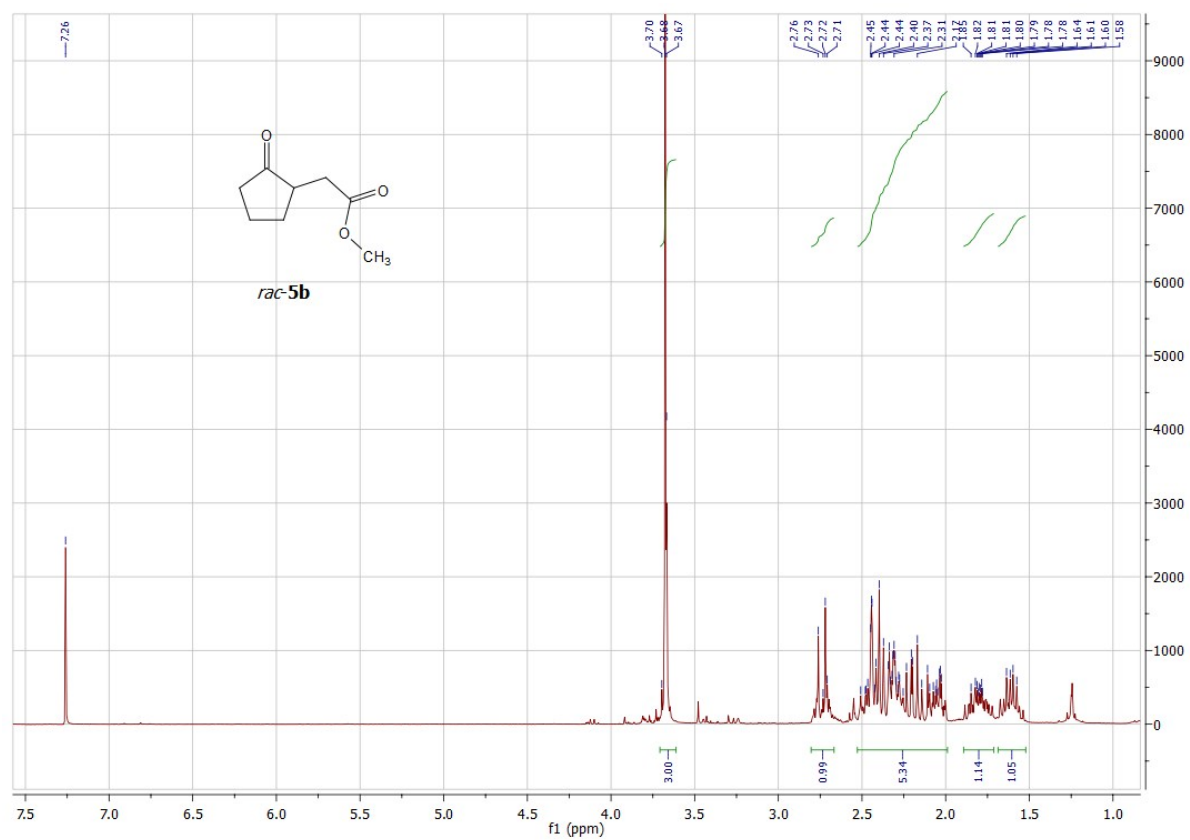
(*R*)-**3b** from XenA ^{13}C



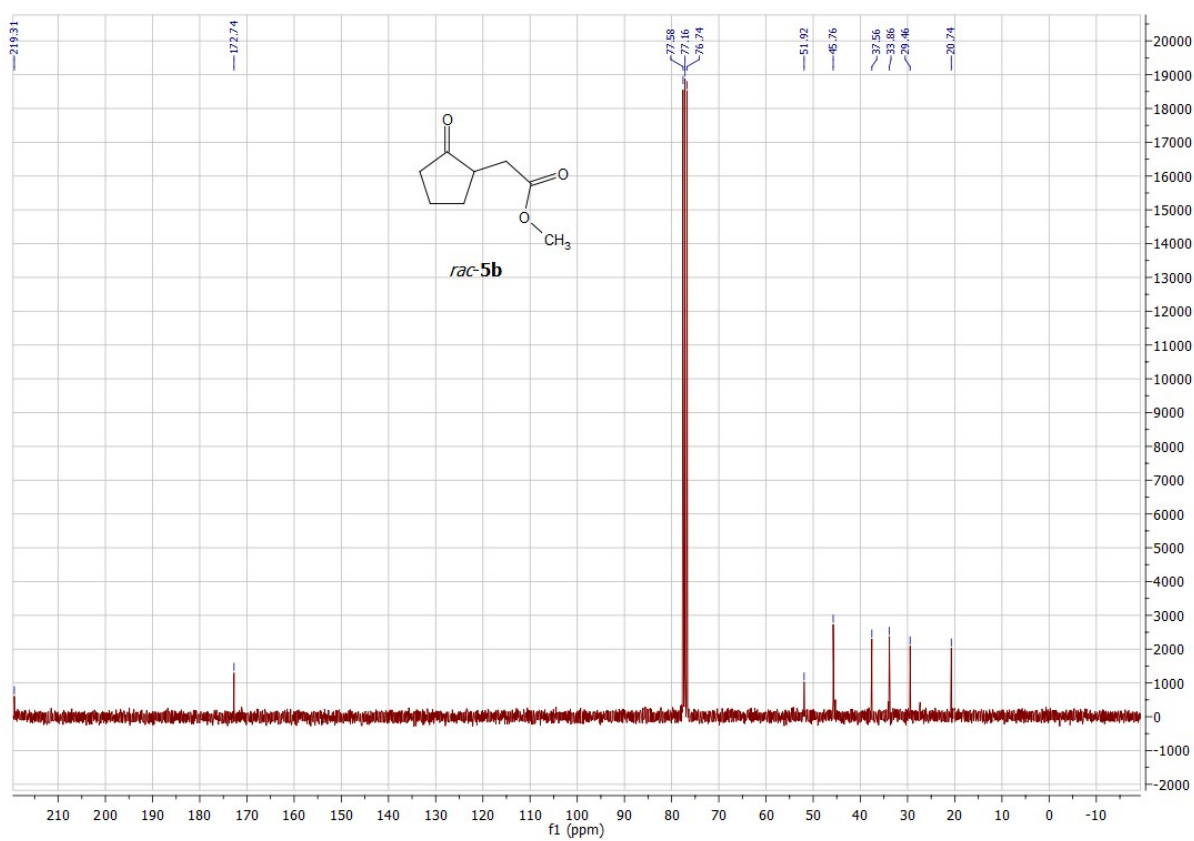
(*R*)-**3b** from XenA HSQC (signals at around 1.2 ppm are impurities already present in the substrate)



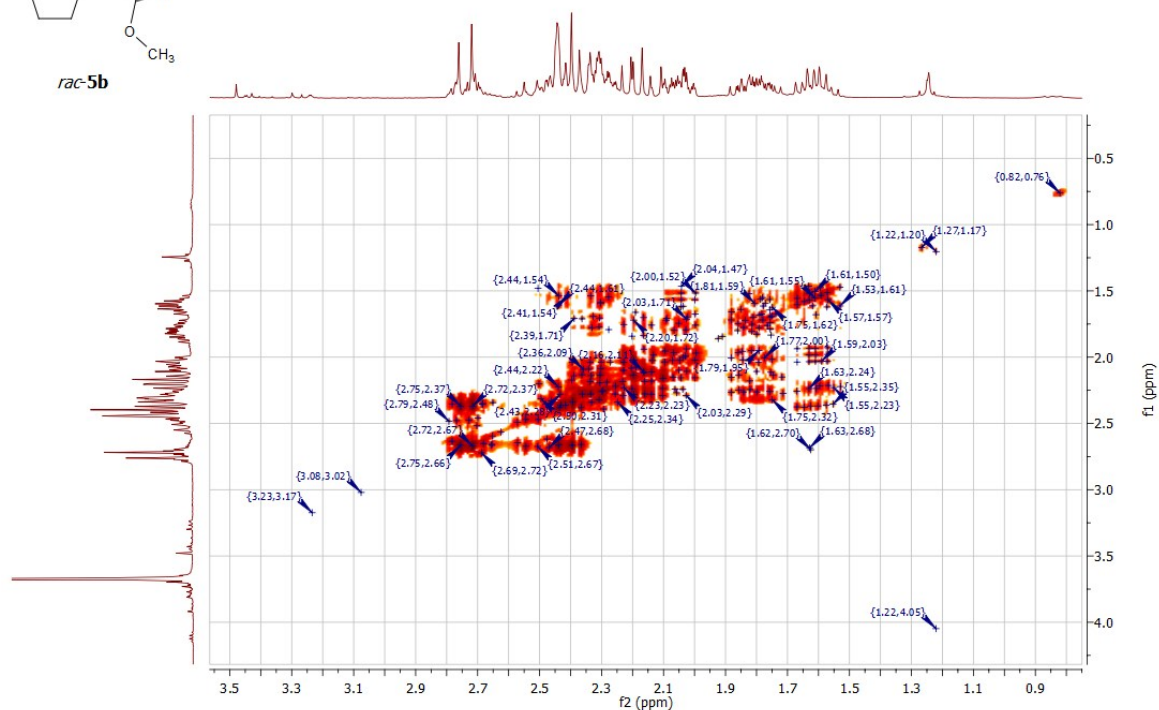
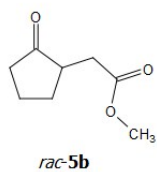
rac-5b



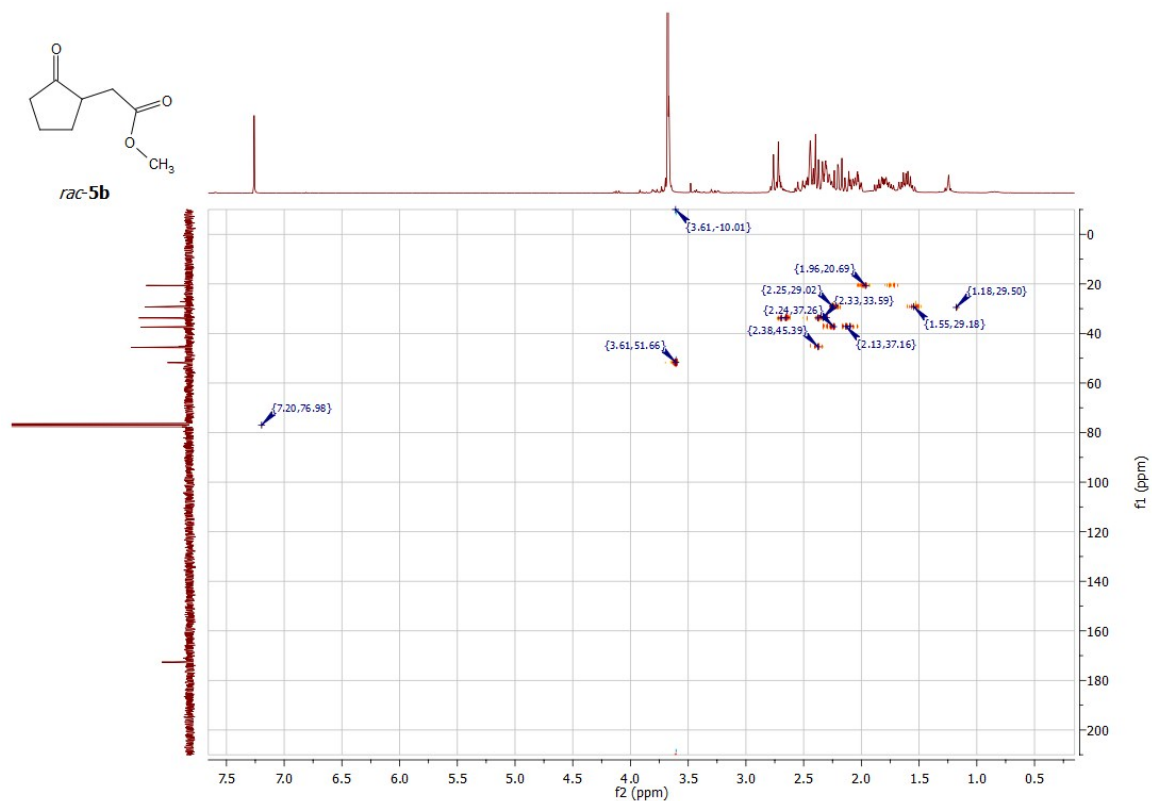
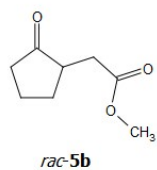
rac-5b ¹³C



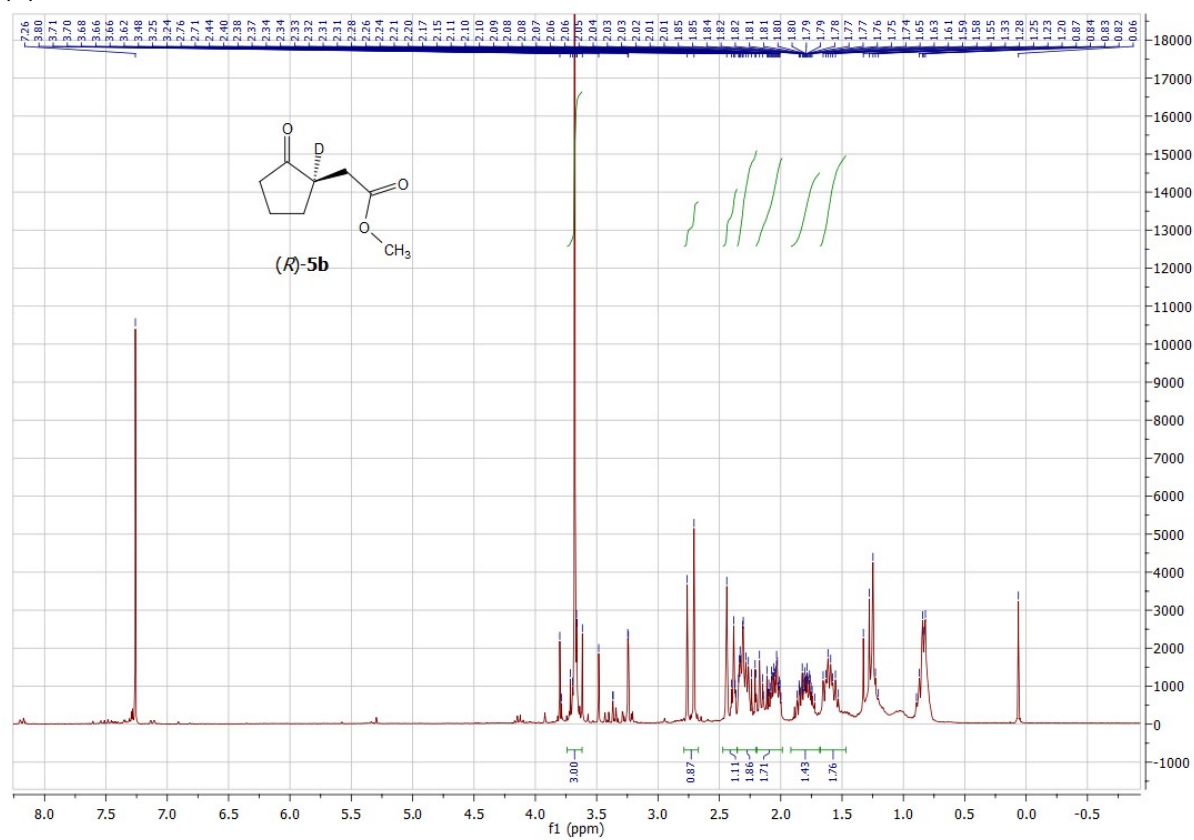
rac-5b COSY



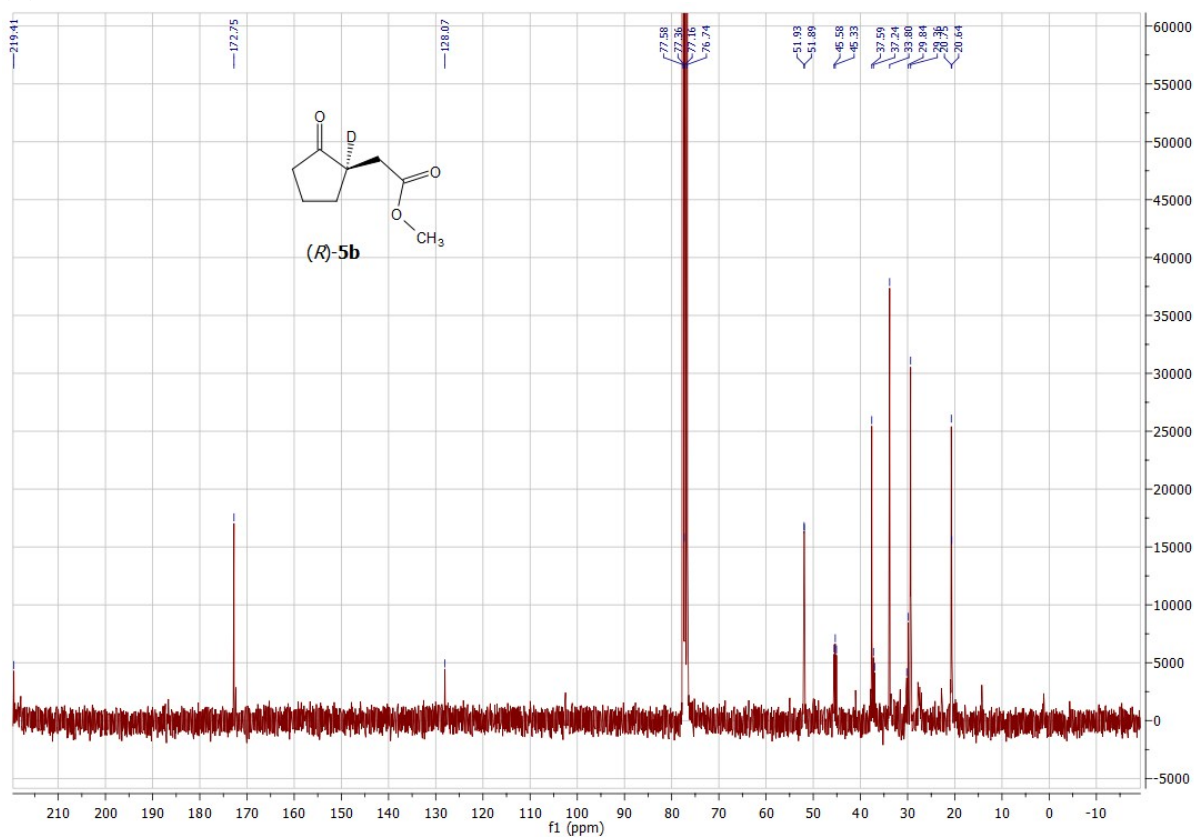
rac-5b HSQC



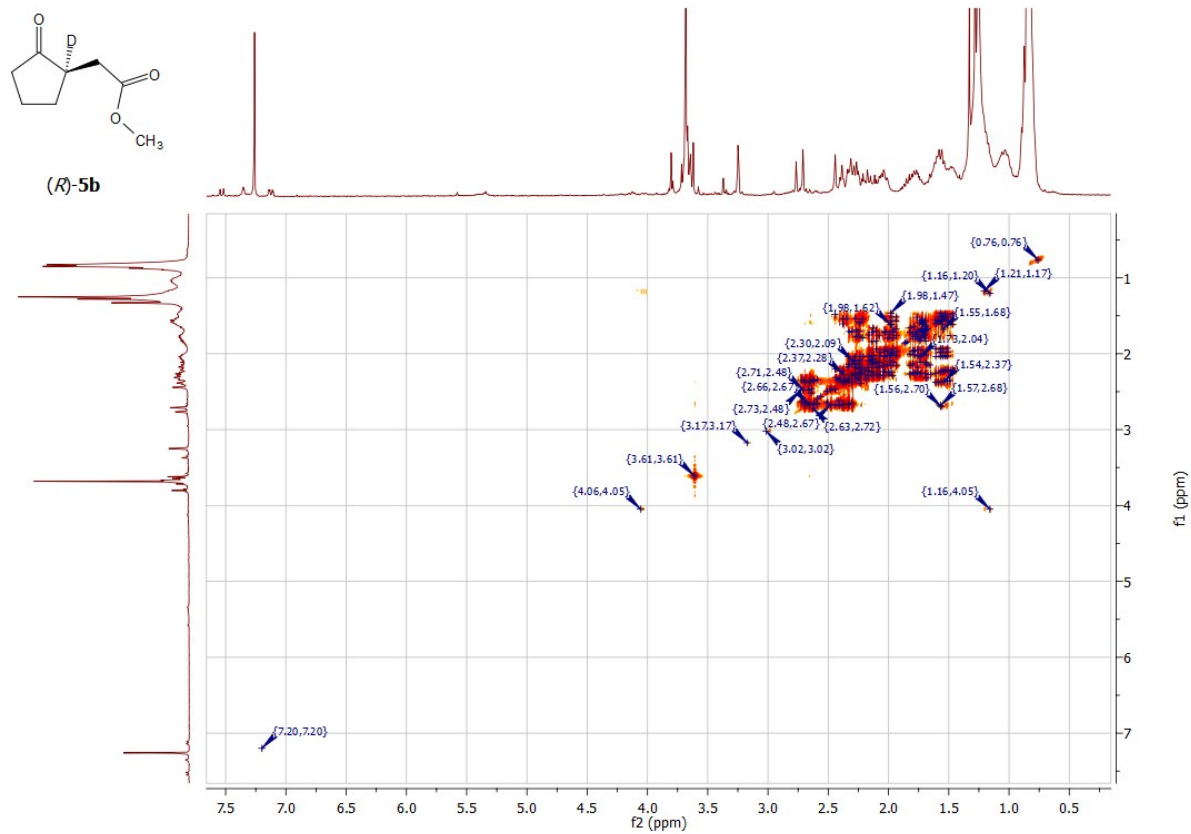
(R)-5b from NCR



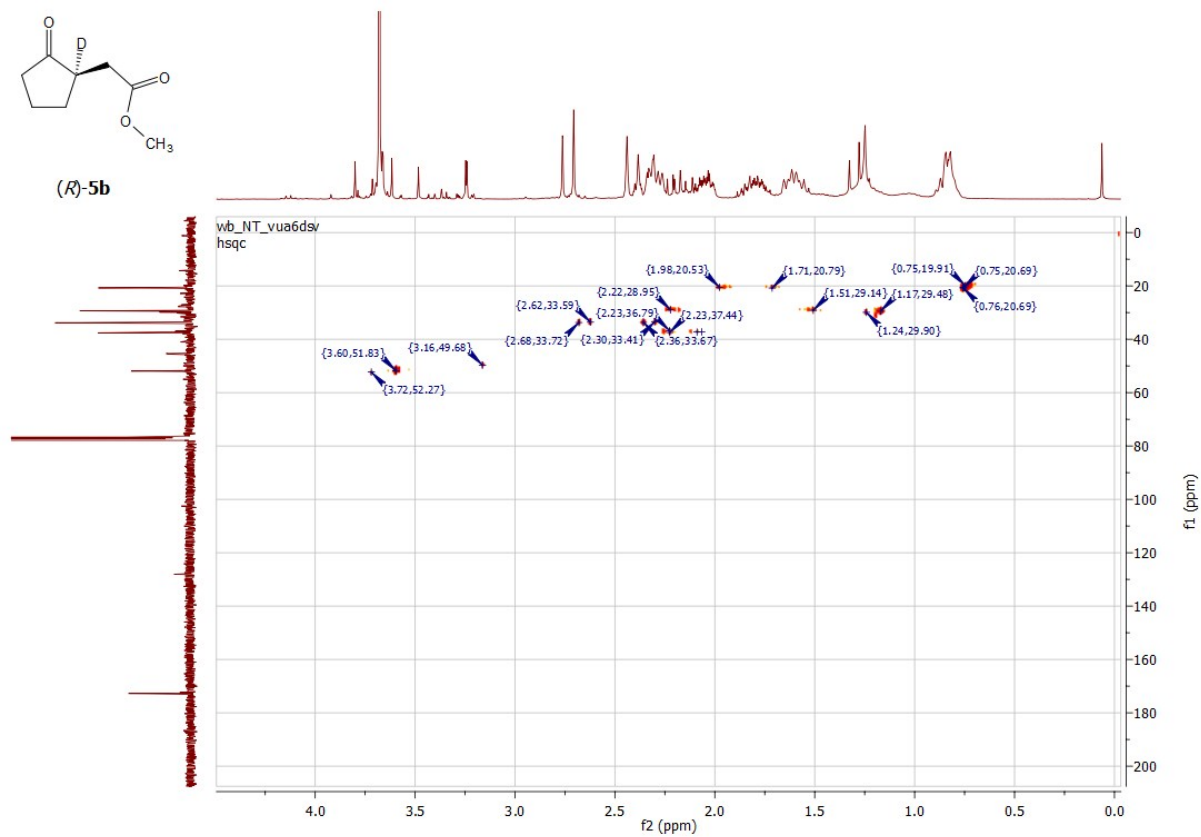
(R)-5b from NCR ¹³C



(R)-5b from NCR COSY



(R)-5b from NCR HSQC



9. References

- [1] S. Mawaziny, A. M. Lawny, *Phosphorus, Sulfur Silicon Relat. Elem.* **2000**, *163*, 99-120.
- [2] B. M. Trost, W. Tang, F. D. Toste, *J. Am. Chem. Soc.* **2005**, *127*, 14785-14803.
- [3] G. L. Lange, J. A. Otulakowski, *J. Org. Chem.* **1982**, *47*, 5093-5096.
- [4] P. Gottschalk, D. C. Neckers, *J. Org. Chem.* **1985**, *50*, 3498-3502.
- [5] Y. Yamamoto, H. Maekawa, S. Goda, I. Nishiguchi, *Org. Lett.* **2003**, *5*, 2755-2758.
- [6] K. Yoshikai, T. Hayama, K. Nishimura, K.-i. Yamada, K. Tomioka, *J. Org. Chem.* **2005**, *70*, 681-683.
- [7] M. J. Begley, K. Cooper, G. Pattenden, *Tetrahedron* **1981**, *37*, 4503-4508.
- [8] B. C. Ranu, J. Dutta, S. K. Guchhait, *Org. Lett.* **2001**, *3*, 2603-2605.
- [9] Y.-J. Chen, H.-L. Wang, N. R. Villarante, G. J. Chuang, C.-C. Liao, *Tetrahedron* **2013**, *69*, 9591-9599.
- [10] S. M. Allin, S. L. James, M. R. J. Elsegood, W. P. Martin, *J. Org. Chem.* **2002**, *67*, 9464-9467.
- [11] K. Nishikawa, T. Ando, K. Maeda, T. Morita, Y. Yoshimi, *Org. Lett.* **2013**, *15*, 636-638.
- [12] C. W. Jefford, A. W. Sledeski, L. Patrick, J. Boukouvalas, *Tetrahedron Lett.* **1992**, *33*, 1855-1858.
- [13] Z. D. Aron, T. Ito, T. L. May, L. E. Overman, J. Wang, *J. Org. Chem.* **2013**, *78*, 9929-9948.
- [14] T. Takeda, M. Terada, *J. Am. Chem. Soc.* **2013**, *135*, 15306-15309.
- [15] A. B. Smith, S. P. Walsh, M. Frohn, M. O. Duffey, *Org. Lett.* **2005**, *7*, 139-142.
- [16] G. H. Posner, J. C. Carry, R. D. Crouch, N. Johnson, *J. Org. Chem.* **1991**, *56*, 6987-6993.
- [17] U. Ravid, R. Ikan, R. M. Sachs, *J. Agric. Food Chem.* **1975**, *23*, 835-838.
- [18] Y. Chen, D. M. Ho, C. Lee, *J. Am. Chem. Soc.* **2005**, *127*, 12184-12185.
- [19] D. J. Coveney, V. F. Patel, G. Pattenden, D. M. Thompson, *J. Chem. Soc., Perkin Transactions 1* **1990**, 2721-2728.
- [20] Y. Aye, S. G. Davies, A. C. Garner, P. M. Roberts, A. D. Smith, J. E. Thomson, *Org. Biomol. Chem.* **2008**, *6*, 2195-2203.
- [21] K. Mori, *Tetrahedron* **1978**, *34*, 915-920.
- [22] A. Mondon, H.-U. Menz, J. Zander, *Chem. Ber.* **1963**, *96*, 826-839.
- [23] I. Fleming, J. Iqbal, *Tetrahedron Lett.* **1983**, *24*, 327-328.
- [24] W. C. Agosta, W. W. Lowrance, *J. Org. Chem.* **1970**, *35*, 3851-3856.