Supplementary information

Structure-based Design of Potent Human Dihydroorotate

Dehydrogenase Inhibitors as Anticancer Agents

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Chemistry

All chemical reagents and solvents were obtained from commercial sources and used without further purification. Thin-layer chromatography (TLC) was performed to monitor the process of reactions. Purification of compounds was achieved by column

chromatography with silica gel (Haiyang, Qingdao), 200–300 mesh. ¹H NMR and ¹³C NMR spectra were recorded on a Bruker AM-400 and a Bruker AM-500 spectrometer

with chemical shifts expressed as ppm (in $CDCl_3$ or $DMSO-d_6$, Me_4Si as internal standard). Melting points were analyzed on a WRS-1B digital melting point apparatus. The mass spectra were measured at the Institute of Fine Chemistry of ECUST.

General procedure for 5a-9a. A mixture of thiosemicarbazide (0.01 mol) and acetone (0.01 mol) in ethanol (40 mL) was refluxed for about 7 h in the presence of five drops of glacial acetic acid as catalyst. The progress of reaction was monitored by TLC. After completion of the reaction, the solution was cooled. The resulting precipitate was filtered and washed with ethanol. White crystal was obtained in good yield without further purification.

General procedure for 5b-9b, 18b-23b. Substituted acetophenone (10 mmol) was solved in 20 mL anhydrous ether in ice bath. One drop of glacial acetic acid was added as catalyst, and then bromine (10 mmol) was added gradually through a separatory funnel. After the addition of bromine, the ice bath was removed and the solution was stirred at ambient temperature overnight. After completion of the reaction, the mixture was washed with saturated aqueous sodium bicarbonate and brine, dried over anhydrous sodium sulfate and concentrated in reduced pressure. The obtained crude product was further purified by column chromatography, eluting with PE/DCM (PE/DCM = 4:1, v/v) to give the colorless oil product.

General procedure for 5c-9c, 11c-17c. The corresponding intermediates from 5a-9a and 5b-9b or 18b-23b were dissolved in ethanol, and then the mixture was heated to reflux until the TLC indicated the reaction was complete. After being cooled down to room temperature, the resulting yellow solid was filtrated off, washed with ethanol and dried. The product can be used directly without further purification.

General procedure for 5d-9d. 80% Hydrazine hydrate (30 mmol) was added into the solution of corresponding compound from **5c-9c** (10 mmol) in ethanol, and the mixture was heated to reflux for about 24 h. Then the solution was cooled to ambient temprature, then to ice bath for a while, finally the resulting precipitate was filtrated off and washed with ethanol. The obtained crude product was further purified through recrystallization using ethanol, and the product with about 30% yield as needle crystal

was obtained.

General procedure for 5-9. To a solution of the corresponding thiazolyl hydrazines (1 mmol) in ethanol, 2-carboxybenzaldehyde (1 mmol) was added. The mixture was stired at room temperature and yellow precipitate formed very quickly. After about 20 min, the target product was filtered, and washed with ethanol without any further purification.

General procedure for 11e-17e. To a solution of compounds 11c (5 mmol) and Cs_2CO_3 (25 mmol) dissolved in DMF, the corresponding alkyl halides were added gradually through a separatory funnel in ice bath. After addition of alkyl halides, the ice bath was removed and the mixture was heated to 80 °C. TLC monitored the reaction until the starting material was consumed totally. Then cooled down the solution to room temperature, extracted with ethyl acetate, collected the organic layer, and concentrated in reduced pressure. The crude product can be directly used for the next step without further purification.

General procedure for 11d-17d. To a solution of compound 11e (1 mmol) in 10 mL THF and 5 mL H₂O, ten drops of concentrate hydrochloride acid were added to provide an acidic environment. Then the solution was heated to 50 °C for about 3 h until TLC indicated the reaction was complete. After cooling down the mixture to room temperature, the mixture was extracted with ethyl acetate, neutralized with saturated aqueous sodium bicarbonate. The organic layer was washed with brine, dried over anhydrous sodium sulphate, and concentrated with reduced pressure. The crude product was purified by column chromatography, eluting with DCM/MeOH (DCM/MeOH = 100:1, v/v) to give brown needle solid in good yield.

General procedure for 11-17. To a solution of the corresponding thiazolyl hydrazines (1 mmol) in ethanol, 2-carboxybenzaldehyde (1 mmol) was added and the mixture was heated to reflux until TLC indicated the reaction was complete. Cooling down the solution to room temperature, removed the solvent under reduced pressure. The crude product was purified by column chromatography with DCM/MeOH (DCM/MeOH = 50:1, v/v) to get yellow solid in good yield.

General procedure for 10f, 18f-23f. To a solution of n-butyl lithium (25 mmol, 2.5 mol/L in hexane) in 10 mL THF at -78 °C, was added a solution of substituted 2-bromobenzoic acid (10 mmol) in THF over 1 h. After stirring at -78°C for 1 h, anhydrous N, N-dimethylformamide (25 mmol) was added dropwise and the mixture was allowed to warm to room temperature slowly. After stirring for around 6 h, a solution of 2 N sodium hydroxide was added, and then the mixture was extracted with

ethyl eater. The aqueous layer was separated and concentrated hydrochloric acid was added until the pH was adjusted to 4-5. The mixture was extracted with ethyl acetate and the organic layer was combined, washed with brine, dried with sodium sulfate, filtered and concentrated. The residue was purified by column chromatography with PE/EA/MeOH (PE/EA/MeOH = 9:1:0.1, v/v/v) to achieve the target product in medium yield.

General procedure for 10g, 18g-23g. Methylhydrazine sulfate (0.1 mol) and ammonium thiocyanate (0.12 mol) were dissolved in 250 ml of ethanol, and the mixture was heated to reflux. After refluxing for 72 h, cooled down the mixture to room temperature, filtrated off the precipitate, concentrated the filtrate under reduced pressure and purified the residue by column chromatography with DCM/MeOH (DCM/MeOH = 100:1, v/v) to get the white power.

General procedure for 10h, 18h-23h. 2-Methyl thiosemicarbazide (1 mmol) and corresponding 2-carboxybenzaldehyde (1 mmol) were dissolved in ethanol and the mixture was heated to reflux until the starting material was consumed totally. After cooling down the mixture to room temperature, removed the solvent under reduced pressure and the residue was purified by column chromatography with DCM/MeOH (DCM/MeOH = 50:1, v/v) to obtain the white solid as the product in good yield.

General procedure for 10, 18-23. To a solution of the corresponding (E)-2-((2-carbamothioyl-2-methylhydrazono)methyl)benzoic acid (1 mmol) in ethanol, substituted 2-bromoacetophenone or 2-bromopropiophenone (1 mmol) was added. Then the mixture was heated to reflux and the process of reaction was monitored by TLC. After completion of the reaction, cooled down the solution to room temperature, removed the solvent under reduced pressure and purified the residue by column chromatography with DCM/MeOH (DCM/MeOH = 50:1, v/v) to obtain the yellow solid as the product in good yield.

(E)-4-phenyl-2-(((2-carboxyl)benzylidene)hydrazinyl)thiazole 5. Mp 196.8-197.5 °C. ¹H NMR (400 MHz, DMSO-d₆, ppm): δ 13.45 (br, 1H), 12.49 (br, 1H), 8.81 (s, 1H), 7.99 (d, J = 7.6 Hz, 1H), 7.87 (m, 3H), 7.63 (t, J = 7.4 Hz, 1H), 7.47 (t, J = 7.2 Hz, 1H), 7.41 (t, J = 7.6 Hz, 2H), 7.33 (s, 1H), 7.30 (t, J = 7.4 Hz, 1H). ¹³C NMR (125 MHz, DMSO-d₆, ppm): δ 168.53, 168.53, 150.97, 140.58, 135.02, 134.99, 132.31, 130.74, 130.15, 129.15, 128.96, 128.96, 127.90, 126.34, 125.90, 125.90, 104.20. HRMS (ESI) calcd for C₁₈H₁₆N₃O₂S [M-H]⁻ 322.0650, found 322.0656.

(E)-5-methyl-4-phenyl-2-(((2-carboxyl)benzylidene)hydrazinyl)thiazole 6. Mp 215.9-216.0 °C. ¹H NMR (400 MHz, DMSO-d₆, ppm): δ 12.7 (br, 2H), 8.77 (s, 1H),

7.99 (d, J = 7.6 Hz, 1H), 7.87 (d, J = 8.0Hz, 1H), 7.63-7.59 (m, 3H), 7.48-7.43(m, 3H), 7.33 (t, J = 7.2 Hz, 1H), 2.43 (s, 3H). ¹³C NMR (125 MHz, DMSO-d₆, ppm): δ 168.66, 164.72, 146.00, 140.05, 135.64, 135.24, 132.36, 130.83, 130.12, 129.08, 128.73, 128.73, 128.35, 128.35, 127.51, 126.33, 117.61, 12.74. HRMS (ESI) calcd for C₁₈H₁₆N₃O₂S [M+H]⁺ 338.0963, found 338.0954.

(E)-4-(2-chlorophenyl)-2-(((2-carboxyl)benzylidene)hydrazinyl)thiazole 7. Mp 195.2-195.2 °C. ¹H NMR (400 MHz, DMSO-d₆, ppm): δ 13.2 (br, 1H), 12.4 (br, 1H), 8.82 (s, 1H), 8.00 (d, J = 7.6 Hz, 1H), 7.88 (t, J = 7.0 Hz, 2H), 7.64 (t, J = 7.4 Hz, 1H), 7.6 (d, J = 7.6 Hz, 1H), 7.48 (t, J = 7.6 Hz, 1H), 7.42 (t, J = 7.8 Hz, 1H), 7.36 (m, 2H). ¹³C NMR (125 MHz, DMSO-d₆, ppm): δ 168.63, 167.77, 147.63, 140.76, 135.08, 133.71, 132.43, 131.54, 131.23, 130.83, 130.83, 130.26, 129.47, 129.28, 127.69, 126.47, 109.26. HRMS (ESI) calcd for C₁₇H₁₃N₃O₂SCl [M+H]⁺ 358.0417, found 358.0417.

(E)-5-methyl-4-(2-chlorophenyl)-2-(((2-carboxyl)benzylidene)hydrazinyl)thiazole 8. Mp 220.0-220.3 °C. ¹H NMR (400 MHz, DMSO-d₆, ppm): δ 12.7 (br, 2H), 8.77 (s,1H), 7.99 (d, J = 7.6 Hz, 1H), 7.87 (d, J = 7.2 Hz, 1H), 7.62 (t, J = 7.4 Hz, 1H), 7.562-7.543 (m, 1H), 7.481-7.397 (m, 4H), 2.14 (s, 3H). ¹³C NMR (125 MHz, DMSO-d₆, ppm): δ 168.65, 165.18, 144.49, 140.16, 135.25, 134.56, 133.21, 132.46, 130.83, 130.14, 130.03, 129.11, 127.45, 126.37, 119.49, 12.15. HRMS (ESI) calcd for C₁₈H₁₅N₃O₂SCl [M+H]⁺ 372.0574, found 372.0569.

(E)-5-ethyl-4-(2-chlorophenyl)-2-(((2-carboxyl)benzylidene)hydrazinyl)thiazole 9. Mp 223.9-223.9 °C. ¹H NMR (500 MHz, DMSO-d₆, ppm): δ 12.57 (br, 2H), 8.79 (s, 1H), 8.01 (d, J = 8.0 Hz, 1H), 7.88 (d, J = 8.0 Hz, 1H), 7.62 (t, J = 7.5 Hz, 1H), 7.56-7.51 (m, 1H), 7.46 (t, J = 7.5 Hz, 1H), 7.44-7.36 (m, 3H), 2.49 (q, J = 7.5 Hz, 1H), 1.13 (t, J = 7.5 Hz, 3H). ¹³C NMR (125 MHz, DMSO-d₆, ppm): δ 169.28, 165.92, 144.14, 140.84, 135.89, 135.46, 133.99, 133.03, 133.02, 131.46, 130.81, 130.73, 130.60, 129.73, 128.09, 127.73, 127.01, 20.99, 17.25. HRMS (ESI) calcd for C₁₉H₁₇N₃O₂SC1 [M+H]⁺ 386.0730, found 386.0729.

(E)-4-(2-chlorophenyl)-2-(1-methyl-2-((2-

carboxyl)benzylidene)hydrazinyl)thiazole 10. Mp 210.4-212.0 °C. ¹H NMR (400 MHz, DMSO-d6, ppm): δ 13.30 (s, 1H), 8.62 (s, 1H), 8.02 (d, J = 7.6 Hz, 1H), 7.98-7.90 (m, 2H), 7.66 (t, J = 7.6 Hz, 1H), 7.56-7.49 (m, 2H), 7.47 (s, 1H), 7.43 (td, J_1 = 7.4 Hz, J_2 = 1.2 Hz, 1H), 7.36 (td, J_1 = 7.6 Hz, J_2 =1.6 Hz, 1H), 3.68 (s, 3H). ¹³C NMR (125 MHz, DMSO-d₆, ppm): δ 168.98, 168.57, 147.11, 136.94, 135.17, 133.42, 132.48, 131.51, 131.09, 130.98, 130.75, 130.16, 129.45, 129.19, 127.63, 126.51,

111.41, 32.88. HRMS (ESI) calcd for $C_{18}H_{15}N_3O_2SC1$ [M+H]⁺ 372.0574, found 372.0575.

(E)-4-(2-chlorophenyl)-2-(1-ethyl-2-((2-carboxyl)benzylidene)hydrazinyl)thiazole 11. Mp 203.3-204.1 °C. ¹H NMR (500 MHz, DMSO-d₆, ppm): δ 13.32 (br, 1H), 8.64 (s, 1H), 7.98-7.91 (m, 3H), 7.66 (t, *J* = 7.5 Hz, 1H), 7.56-7.46 (m, 2H), 7.45-7.39 (m, 2H), 7.34 (t, *J* = 6.8 Hz,1H), 4.32 (q, *J* = 6.8 Hz, 2H), 1.27 (t, *J* = 7.0 Hz, 3H). ¹³C NMR (125 MHz, DMSO-d₆, ppm): δ 168.56, 168.36, 147.34, 137.00, 135.42, 133.53, 132.50, 131.53, 131.12, 130.92, 130.72, 130.22, 129.42, 129.19, 127.62, 126.62, 111.22, 40.38, 10.33. HRMS (ESI) calcd for C₁₉H₁₇N₃O₂SCI [M+H]⁺ 386.0730, found 386.0728.

(E)-4-(2-chlorophenyl)-2-(1-propyl-2-((2-

carboxyl)benzylidene)hydrazinyl)thiazole 12. Mp 172.7-173.4 °C. ¹H NMR (400 MHz, DMSO-d₆, ppm): δ 13.35 (br, 1H), 8.65 (s, 1H), 7.98 (d, J = 8.0 Hz, 1H), 7.95-7.92 (m, 2H), 7.67 (t, J = 7.2 Hz, 1H), 7.54 (d, J = 8.0 Hz, 1H), 7.5 (t, J = 7.6 Hz, 1H), 7.45-7.42 (m, 2H), 7.36 (t, J_1 = 7.6 Hz, J_2 = 1.6 Hz, 1H), 4.25 (t, J = 7.2 Hz, 2H), 1.82-1.73 (m, 2H), 0.97 (t, J = 7.4 Hz, 1H). ¹³C NMR (100 MHz, DMSO-d₆, ppm): δ 168.54, 168.22 146.95, 136.63, 134.96, 133.17, 132.11, 131.10, 130.74, 130.55, 130.34, 129.82, 129.04, 128.80, 127.26, 126.09, 110.77, 46.31, 17.98,11.17. HRMS (ESI) calcd for C₂₀H₁₉N₃O₂SCI [M+H]⁺ 400.0887, found 400.0879.

(E)-4-(2-chlorophenyl)-2-(1-isopropyl-2-((2-

carboxyl)benzylidene)hydrazinyl)thiazole 13. Mp 185.4-187.0 °C. ¹H NMR (400 MHz, DMSO-d₆, ppm): δ 13.35 (br, 1H), 8.91 (s, 1H), 7.99 (d, J = 8.0 Hz, 1H), 7.95-7.92 (m, 2H), 7.67 (t, J = 7.6 Hz, 1H), 7.54 (d, J = 8.0 Hz, 1H), 7.52 (t, J = 7.6 Hz, 1H), 7.46-7.42 (m, 2H), 7.36 (td, J_1 = 7.6 Hz, J_2 = 1.6 Hz, 1H), 5.22-5.11 (m, 1H), 1.57 (d, J = 6.8 Hz, 6H). ¹³C NMR (100 MHz, DMSO-d₆, ppm): δ 168.22, 168.21, 146.90, 137.32, 135.23, 133.14, 132.07, 131.06, 130.64, 130.58, 130.39, 129.88, 128.97, 128.78, 127.28, 125.91, 111.12, 49.61, 18.07, 18.07. HRMS (ESI) calcd for C₂₀H₁₉N₃O₂SC1 [M+H]⁺400.0887, found 400.0885.

(E)-4-(2-chlorophenyl)-2-(1-hydroxyethyl-2-((2-

carboxyl)benzylidene)hydrazinyl)thiazole 14. Mp 187.9-188.9 °C. ¹H NMR (400 MHz, DMSO-d₆, ppm): δ 13.24 (br, 1H), 8.75 (s, 1H), 7.98-7.01 (m, 3H), 7.66 (t, J = 7.6 Hz, 1H), 7.54 (d, J = 8.0 Hz, 1H), 7.49 (t, J = 7.6 Hz, 1H), 7.45-7.41 (m, 2H), 7.36 (td, J_1 = 7.6 Hz, J_2 = 1.6 Hz, 1H), 4.35 (t, J = 6.4 Hz, 2H), 3.76 (t, J = 6.4 Hz, 2H). ¹³C NMR (100 MHz, DMSO-d₆, ppm): δ 168.64, 168.23, 146.87, 136.89, 134.91, 133.15, 131.99, 131.19, 130.71, 130.46, 130.32, 130.16, 129.04, 128.79, 127.22,

126.28, 110.78, 56.24, 47.46. HRMS (ESI) calcd for $C_{19}H_{17}N_3O_3SC1$ [M+H]⁺ 402.0679, found 402.0678.

(E)-4-(2-chlorophenyl)-2-(1-(2-butyl)-2-((2-

carboxyl)benzylidene)hydrazinyl)thiazole 15. Mp 172.7-173.4 °C. ¹H NMR (400 MHz, DMSO-d₆, ppm): δ 13.32 (br, 1H), 8.91 (s, 1H), 7.99 (d, J = 8.0 Hz, 1H), 7.95-7.90 (m, 2H), 7.67 (t, J = 7.6 Hz, 1H), 7.55-7.48 (m, 2H), 7.46-7.42 (m, 2H), 7.36 (td, J_1 = 7.6 Hz, J_2 = 1.6 Hz, 1H), 5.00-4.93 (m, 1H), 2.35-2.24 (m, 1H), 1.92-1.81 (m, 1H), 1.54 (d, J = 6.8 Hz, 3H), 0.88 (t, J = 7.2 Hz, 3H). ¹³C NMR (100 MHz, DMSO-d₆, ppm): δ 168.66, 168.19, 146.97, 137.03, 135.24, 133.19, 132.13, 131.04, 130.67, 130.58, 130.39, 129.70, 128.98, 128.77, 127.29, 125.91, 111.04, 55.67, 25.26, 16.33, 11.14. HRMS (ESI) calcd for C₂₁H₂₁N₃O₂SCl [M+H]⁺ 414.1043, found 414.1029.

(E)-4-(2-chlorophenyl)-2-(1-(pentan-2-yl)-2-((2-

carboxyl)benzylidene)hydrazinyl)thiazole 16. Mp 146.7-148.0 °C. ¹H NMR (400 MHz, DMSO-d₆, ppm): δ 13.35 (br, 1H), 8.91 (s, 1H), 7.98 (d, *J* = 7.6 Hz, 1H), 7.94 (d, *J* = 8.0 Hz, 1H), 7.91 (dd, *J*₁ = 7.6 Hz, *J*₂ = 1.6 Hz, 1H), 7.67 (t, *J* = 7.6 Hz, 1H), 7.53 (d, *J* = 8.0 Hz, 1H), 7.49 (t, *J* = 7.6 Hz, 1H), 7.46-7.42 (m, 2H), 7.36 (td, *J*₁ = 7.6 Hz, *J*₂ = 1.6 Hz, 1H), 5.14-5.06 (m, 1H), 2.34-2.25 (m, 1H), 1.82-1.73 (m, 1H), 1.53 (d, *J* = 6.8 Hz, 3H), 1.34-1.24 (m, 2H), 0.89 (t, *J* = 7.4 Hz, 1H). ¹³C NMR (100 MHz, DMSO-d₆, ppm): δ 168.68, 168.20, 146.98, 137.04, 135.30, 133.20, 132.14, 131.00, 130.68, 130.59, 130.40, 129.68, 128.99, 127.30, 125.90, 111.09, 53.81, 34.20, 19.46, 16.46, 13.60. HRMS (ESI) calcd for $C_{22}H_{23}N_3O_2SC1$ [M+H]⁺ 428.1200, found 428.1193.

(E)-5-methyl-4-(2-chlorophenyl)-2-(1-(pentan-2-yl)-2-((2-

carboxyl)benzylidene)hydrazinyl)thiazole 17. Mp. 89.9-90.1 °C. ¹H NMR (400 MHz, DMSO-d₆, ppm): δ 13.29 (br, 1H), 8.85 (s, 1H), 7.98 (d, J = 8.0 Hz, 1H), 7.92 (d, J = 7.6 Hz, 1H), 7.65 (t, J = 7.6 Hz, 1H), 7.56-7.54 (m, 1H), 7.51-7.41 (m, 4H), 5.05-4.96 (m, 1H), 2.29-2.22 (m, 1H), 2.16 (s, 3H), 1.75-1.67 (m, 1H), 1.46 (d, J = 7.2 Hz, 3H), 1.28-1.23 (m, 2H), 0.87 (t, J = 7.6 Hz, 3H). ¹³C NMR (100 MHz, DMSO-d₆, ppm): δ 168.23, 166.06, 144.08, 136.14, 135.46, 134.09, 132.79, 132.09, 131.93, 130.58, 129.64, 129.60, 129.48, 128.56, 126.95, 125.73, 121.62, 53.39, 34.01, 19.41, 16.35, 13.58, 11.52. HRMS (ESI) calcd for C₂₃H₂₅N₃O₂SCI [M+H]⁺ 442.1356, found 442.1354.

(E)-4-(2,5-dichlorophenyl)-2-(1-methyl-2-((2-

carboxyl)benzylidene)hydrazinyl)thiazole 18. Mp. 268.4-269.5 °C. ¹H NMR (400 MHz, DMSO-d₆, ppm): δ 13.31 (br, 1H), 8.63 (s, 1H), 8.02-8.00 (m, 2H), 7.93 (d, J =

7.2 Hz, 1H), 7.67 (t, J = 7.6 Hz, 1H), 7.63 (s, 1H), 7.58 (d, J = 7.6 Hz, 1H), 7.50 (t, J = 7.2 Hz, 1H), 7.43 (dd, $J_1 = 7.2$ Hz, $J_2 = 2.8$ Hz, 1H), 3.68 (s, 3H). ¹³C NMR (100 MHz, DMSO-d₆, ppm): δ 168.68, 168.15, 145.13, 136.79, 134.70, 134.21, 132.14, 132.06, 131.85, 130.59, 130.17, 129.76, 129.20, 128.84, 128.58, 126.13, 112.31, 32.49. HRMS (ESI) calcd for C₁₈H₁₄N₃O₂SCl2 [M+H]⁺ 406.0184, found 406.0187.

(E)-4-(3-chlorophenyl)-2-(1-methyl-2-((2-

carboxyl)benzylidene)hydrazinyl)thiazole 19. Mp. 244.7-245.4 °C. ¹H NMR (400 MHz, DMSO-d6, ppm): δ 13.32 (br, 1H), 8.63 (s, 1H), 8.01 (d, J = 7.6 Hz, 1H), 7.97 (s, 1H), 7.93 (d, J = 7.6 Hz, 1H), 7.88 (d, J = 8.0 Hz, 1H), 7.66 (t, J = 7.6 Hz, 1H), 7.61 (s, 1H), 7.50 (t, J = 7.6 Hz, 1H), 7.45 (t, J = 7.6 Hz, 1H), 7.37 (d, J = 8.0 Hz, 1H), 3.71 (s, 3H). ¹³C NMR (100 MHz, DMSO-d₆, ppm): δ 169.57, 168.18, 148.56, 136.64, 136.50, 134.72, 133.48, 132.06, 130.59, 130.44, 129.76, 128.81, 127.28, 126.12, 125.16, 124.05, 107.56, 32.52. HRMS (ESI) calcd for C₁₈H₁₅N₃O₂SC1 [M+H]⁺ 372.0574, found 372.0574.

(E)-4-(2-chlorophenyl)-2-(1-methyl-2-((2-carboxyl)-(4-

trifluoromethyl)benzylidene)hydrazin-yl)thiazole 20. Mp 209.3-210.7 °C. ¹H NMR (400 MHz, DMSO-d₆, ppm): δ 14.02 (br, 1H), 8.66 (s, 1H), 8.21 (d, J = 8.0 Hz, 1H), 8.17 (s, 1H), 8.02 (d, J = 8.0 Hz, 1H), 7.95 (d, J = 7.6 Hz, 1H), 7.56-7.53 (m, 2H), 7.43 (t, J = 7.6 Hz, 1H), 7.37 (t, J = 7.6 Hz, 1H), 3.70 (s, 3H). ¹³C NMR (100 MHz, DMSO-d₆, ppm): δ 168.33, 167.19, 146.79, 138.35, 135.04, 132.91, 131.12, 130.70, 130.37, 129.14, 128.57, 128.24, 128.18, 127.26, 127.02, 125.10, 122.40, 111.51, 32.68. HRMS (ESI) calcd for C₁₉H₁₄N₃O₂SClF₃ [M+H]⁺ 440.0447, found 440.0433.

(E)-4-(2-chlorophenyl)-2-(1-methyl-2-((2-carboxyl)-(4-

methyl)benzylidene)hydrazinyl)thiazole 21. Mp 231.3-232.5 °C. ¹H NMR (400 MHz, DMSO-d₆, ppm): δ 13.30 (br, 1H), 8.59 (s, 1H), 7.96 (d, J = 7.6 Hz, 1H), 7.92 (d, J = 8.0 Hz, 1H), 7.75 (s, 1H), 7.54 (d, J = 7.6 Hz, 1H), 7.49-7.46 (m, 2H), 7.43 (t, J = 7.6 Hz, 1H), 7.36 (t, J = 7.6 Hz, 1H), 3.66 (s, 3H), 2.38 (s, 3H). ¹³C NMR (100 MHz, DMSO-d₆, ppm): δ 168.60, 168.35, 146.68, 138.54, 136.64, 133.03, 132.70, 132.07, 131.11, 130.86, 130.67, 130.34, 129.83, 129.00, 127.21, 126.04, 110.84, 32.39, 20.69. HRMS (ESI) calcd for C₁₉H₁₇N₃O₂SC1 [M+H]⁺ 386.0730, found 386.0738.

(E)-4-(2-chlorophenyl)-2-(1-methyl-2-((2-carboxyl)-(4-

fluoro)benzylidene)hydrazinyl)thiazole 22. Mp 236.2-237.1 °C. ¹H NMR (400 MHz, DMSO-d₆, ppm): δ 8.61 (s, 1H), 8.06-8.02 (dd, J_1 = 5.6 Hz, J_2 = 8.4 Hz, 1H), 7.96 (d, J = 7.2 Hz, 1H), 7.67-7.64 (dd, J_1 = 9.6 Hz, J_2 = 2.4 Hz, 1H), 7.55-7.51 (m, 2H), 7.47

(s, 1H), 7.43 (t, J = 7.2 Hz, 1H), 7.36 (t, J = 7.2 Hz, 1H), 3.68 (s, 3H). ¹³C NMR (100 MHz, DMSO-d₆, ppm): δ 168.55, 168.01, 159.74 (d, ¹J = 249 Hz), 146.67, 133.68, 133.10, 131.50, 131.12, 130.72, 130.32, 130.10 (d, ³J = 8.8 Hz), 129.06, 127.23, 125.53 (d, ⁴J = 3.1 Hz), 121.75 (d, ²J = 11.8 Hz), 118.76 (d, ²J = 22 Hz), 111.13, 32.28. HRMS (ESI) calcd for C₁₈H₁₄N₃O₂FSC1 [M+H]⁺ 390.0479, found 390.0475.

(E)-5-methyl-4-phenyl-2-(1-methyl-2-((4-methyl)-(2-

carboxyl)benzylidene)hydrazinyl)thiazole 23. Mp 243.2-245.2 °C. ¹H NMR (400 MHz, DMSO-d6, ppm): δ 13.24 (br, 1H), 8.54 (s, 1H), 7.91(d, J = 8.0 Hz, 1H), 7.73 (s, 1H), 7.64-7.53 (m, 2H), 7.46-7.42 (m, 3H), 7.33 (t, J = 7.4 Hz, 1H), 3.60 (s, 3H), 2.44 (s, 3H), 2.38 (s, 3H). ¹³C NMR (100 MHz, DMSO-d₆, ppm): δ 168.35, 165.40, 145.43, 138.33, 135.76, 135.16, 132.68, 132.22, 130.86, 129.62, 128.24, 128.24, 127.85, 127.85, 127.06, 125.89, 119.27, 31.85, 20.68, 12.24. HRMS (ESI) calcd for C₂₀H₂₀N₃O₂S [M+H]⁺ 366.1276, found 366.1273.

In vitro enzyme assay

Recombined *h*DHODH was expressed and purified as described in our previous works.¹⁻³ The *h*DHODH inhibition assays were performed using the DCIP-based assay. For each inhibitor, eight different concentrations were used to determine the IC_{50} value. Brequinar was measured as the positive control. Each inhibitor concentration point was tested in triplicate and IC_{50} values were determined using origin 8.0 and summarized in Table 1.

Cell proliferation assay

Human colon cancer cell line HCT-116 and human pancreatic cancer cell line BxPC-3 were cultured in RPMI-1640 (Hyclone) and DMEM (Hyclone), respectively, and human lung fibroblast cell line Wi-38 was cultured in MEM (Hyclone), all containing 10% fetal bovine serum (FBS). Cell proliferation assay was evaluated by the MTT reduction assay.⁴ Firstly, 5000 cells were placed in each well of a 96-well plate. After 24 h incubation, the cells attached to the plate. 10 μ L compounds diluted to seven different concentrations (ranging from 0.016 μ M to 10 μ M for cancer cell lines and ranging from 0.082 μ M to 100 μ M for normal cell line) were added to each well and cell cultures were incubated for another 72 h. The concentration of DMSO in each well was 0.5% (ν/ν). Then 10 μ L MTT solutions (5 mg/mL) were added to each well. Subsequently, the plate was incubated for 4 h at 37 °C in a cell culture incubator. Afterwards, the culture medium was removed and aliquots (100 μ L) of DMSO were added to each well to dissolve the formazan crystals. Finally, the absorbance was measured at 570 nm. The final results were recorded by averaging at least three

independent determinations. The IC_{50} values were analyzed using Origin 8.0.

Analysis of apoptosis

HCT-116 and BxPC-3 cell lines were also used for the cell apoptosis assay by flow cytometry. The cells were incubated with 200 nM compound **22** for 24 h, 36 h and 48 h, respectively. Then the cells were collected and washed twice with cold PBS and resuspended in the binding buffer. The suspension was stained with 5 μ L propidium iodide (PI) and 5 μ L Annexin V-FITC and then incubated in the dark for 15 min at RT.

Analysis of cell cycle

HCT-116 and BxPC-3 cell lines were used for the cell cycle analysis. The cells were harvested after incubation with compound **22** (5 nM, 10 nM, 20 nM) for 12 h and washed twice in ice-cold PBS buffer before being fixed with cold ethanol (70%) for 12 h at 4 °C. Fixed cells were rinsed twice with cold PBS and resuspended in PBS. The suspension was incubated with RNase and propidium iodides (PI) for 15 min at RT. The distribution of the cell cycle was analyzed by flow cytometry.

Molecular docking simulation

To thoroughly elucidate the binding modes of this series of derivatives, molecular docking method was employed. The X-ray crystal structure of *h*DHODH in complex with compound 7 (PDB ID: 4LS1) was used for protein preparation. Hydrogen atoms and charges were added to the receptor using the Protein Preparation workflow in Maestro (Schrödinger Inc, version 9.0). The grid-enclosing box was centered on the centroid of ligand (compound 7) and defined so as to enclose residues located within 20 Å around the ubiquinone binding site, and a scaling factor of 1.0 was set to van der Waals (VDW) radii of those receptor atoms with the partial atomic charge less than 0.25. Then compounds 17 and 22 were prepared by the LigPrep module in Maestro, and Epik method was selected to predict the ionization states of the compounds within the default pH range of 7.0 ± 2.0 . Finally, Extra-Precision (XP) mode in Glide was used to perform the molecular docking simulations, and the top 10 docked poses of each compound ranked by GlideScore were remained for further analysis.

References

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Compound 6

Single Mass Analysis Tolerance = 50.0 PPM / DBE: min = -1.5, max = 100.0 Element prediction: Off Number of isotope peaks used for i-FIT = 2 Monoisotopic Mass, Even Electron Ions 104 formula(e) evaluated with 4 results within limits (up to 1 best isotopic matches for each mass) Elements Used: C: 0-30 H: 0-50 N: 0-3 O: 0-2 S: 0-1 WP-ZHU ECUST institute of Fine Chem 30-Mar-2015 21:39:45 1: TOF MS ES+ 1.55e+003 ZWP-SWL-084 4 (0.200) Cm (1:6) 338.0954 100-%-331.3409 342.0686 339,0980 334.2927 340.0988 332.3448 332.0 333.0 331.0 334.0 336.0 340.0 341.0 ---- m/z 337.0 342.0 335.0 338.0 339.0 Minimum: Maximum: -1.5100.0 300.0 50.0 mDa PPM DBE i-FIT i-FIT (Norm) Formula Mass Calc. Mass 338.0954 338.0963 C18 H16 N3 O2 S -0.9 -2.7 12.5 6.0 0.0

Compound 7



S13

Page 1



Compound 8





Compound 10



Page 1



Compound 11



Single Mass Analysis Tolerance = 50.0 PPM / DBE: min = -1.5, max = 100.0 Element prediction: Off Number of isotope peaks used for i-FIT = 2 Monoisotopic Mass, Even Electron Ions 41 formula(e) evaluated with 1 results within limits (up to 1 best isotopic matches for each mass) Elements Used:



Compound 14





Compound 16



Single Mass Analysis Tolerance = 50.0 PPM / DBE: min = -1.5, max = 100.0 Element prediction: Off Number of isotope peaks used for i-FIT = 2 Monoisotopic Mass, Even Electron Ions 41 formula(e) evaluated with 1 results within limits (up to 1 closest results for each mass) Elements Used: C: 0-22 H: 0-23 N: 0-3 O: 0-2 S: 0-1 CI: 0-1 ECUST institute of Fine Chem XH-QIAN 20-Jul-2015 14:36:40 1: TOF MS ES+ ZWP-SWL-192 160 (1.079) Cm (159:160) 2.44e+003 428.1193 100-430.1209 %-431.1262 346.3308 362.3244 374.3760 512,4966 -1.5 Minimum: 300.0 50.0 Maximum: i-FIT (Norm) Formula Calc. Mass Mass mDa PPM DBE i-FIT 428.1193 428.1200 -0.7 12.5 32.0 0.0 C22 H23 N3 O2 S C1 -1.6

Page 1

Compound 18







Single Mass Analysis Tolerance = 50.0 PPM / DBE: min = -1.5, max = 100.0 Element prediction: Off Number of isotope peaks used for i-FIT = 2

Monoisotopic Mass, Even Electron Ions 41 formula(e) evaluated with 1 results within limits (up to 1 closest results for each mass) Elements Used: C: 0-19 H: 0-17 N: 0-3 O: 0-2 S: 0-1 CI: 0-1 WP-ZHU ECUST institute of Fine Chem

ZWP-SWL-179 83 (0.608) Cm (80:83)



Compound 23



S25

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03-Jun-2015 20:45:37 1: TOF MS ES+

