

Supporting Information

Design, Synthesis and Biological Profiling of Aryl Piperazine Based Scaffold for the Management of Androgen Sensitive Prostatic Disorders[#]

Sonal Gupta,^{a,e} Deepti Pandey,^b Dhanaraju Mandalapu,^a Veenu Bala,^{a,e} Vikas Sharma,^b Mahendra Shukla,^{c,e} Santosh K.Yadav,^b Nidhi Singh,^d Swati Jaiswal,^{c,e} Jagdamba P. Maikhuri,^b Jawahar Lal,^{c,e} Mohammad I. Siddiqi,^d Gopal Gupta^b and Vishnu L. Sharma^{a,e,*}

^a*Medicinal & Process Chemistry Division, CSIR-Central Drug Research Institute, Lucknow-226031 (India)*

^b*Endocrinology Division, CSIR-Central Drug Research Institute, Lucknow-226031 (India)*

^c*Pharmacokinetics and Metabolism Division, CSIR-Central Drug Research Institute, Lucknow-226031 (India)*

^d*Molecular & Structural Biology Division, CSIR-Central Drug Research Institute, Lucknow-226031 (India)*

^e*Academy of Scientific and Innovative Research (AcSIR), New Delhi-110001 (India)*

*Corresponding author. Dr. V. L. Sharma

Medicinal and Process Chemistry Division,

CSIR-Central Drug Research Institute, Sector 10, Jankipuram ext.,

Lucknow, Uttar Pradesh 226031, India.

Tel.: 91-522-2772450; Ext. 4671; Fax: 91-522-2771941

E-mail address: vl_sharma@cdri.res.in; vlscdri@gmail.com

[#]CDRI Manuscript No. 276/2015/VLS

Communication No. 9294

1. Experimental details of chemistry and biology.....	S3-S17
2. ¹ H NMR and ¹³ C NMR spectra of compounds.....	S18-S45
3. HRMS of compounds.....	S46-S58
4. Representative Chromatogram of compound 9a	S59
5. References.....	S59

1. EXPERIMENTAL

1.1. Chemistry

General information

All solvents and reagents were commercial available and used without further purification. Analytical thin layer chromatography (TLC) was performed on pre-coated silica gel plates 60 GF₂₅₄ (Aldrich) to monitor the progress of the reaction. Electrospray ionization mass spectra (ESI-MS) were recorded on Ion Trap LCQ Advantage Max-IT (Thermo Electron Corporation). High-resolution mass spectra (HRMS) were recorded on a 6520 Agilent Q ToF LC MS/MS (accurate mass). ¹H and ¹³C NMR spectra were done on Bruker Supercon Magnet Avance spectrometers at 300/400 and 75.4/100 MHz respectively in deuterated solvents taking TMS as internal reference (chemical shifts δ in ppm, J in Hz.). IR spectra (ν_{\max} in cm⁻¹) were taken on Perkin Elmer's FT-IR RX1 PC spectrophotometer. Melting points were determined in open capillary tubes on the melting point apparatus. Elemental analyses were done on Carlo Erba EA-1108 micro analyzer/Vario ELIII C H N S analyzer. All compounds were analyzed for C, H, N and the results were within $\pm 0.3\%$ of calculated values. All compounds were characterized by TLC, ¹H and ¹³C NMR, MS, and HRMS. Elemental analyses data meet the criteria of >95% purity.

2-Chloro-1-(4-(4-nitrophenyl)piperazin-1-yl)ethanone (8a)

The mixture of 1-(4-nitrophenyl)piperazine (**7a**, 1 g, 4.83 mmol), Et₃N (1.34 mL, 9.66 mmol) in dry DCM (15 mL) was stirred at 0 to 5 °C for 15 min under dry condition using CaCl₂ guard tube and then chloroacetyl chloride (0.76 mL, 9.66 mmol) in dry DCM (5 mL), was added dropwise within 1 h duration. Reaction mixture was further stirred at room temperature for 2 h. The resultant reaction mixture was concentrated under reduced pressure, dissolved in EtOAc (15 mL) and washed with distilled water (10 mL \times 3) and organic layer was separated. The combined organic layer dried (anhyd. Na₂SO₄) and concentrated under reduced pressure. The obtained solid residue was recrystallized using EtOAc/Hexane to give the title compound as yellow solid (yield 92%); mp: 100-101 °C; IR (KBr) ν (cm⁻¹): 3012, 2853, 1653, 1596, 1500, 1442, 1389, 1327; ¹H NMR (400 MHz, CDCl₃): δ 8.14-8.19 (2H, m), 6.84-6.82 (2H, m), 4.11 (2H, s), 3.81-3.79 (2H, m), 3.74-3.72 (2H, m), 3.53-3.50 (2H, m), 3.47-3.45 (2H, m); ¹³C NMR (100 MHz, CDCl₃): δ 165.3, 154.3, 139.3, 125.9, 113.2, 47.1, 46.7, 45.4, 41.5, 40.6; HRMS (ESI positive)

m/z calcd. for $C_{12}H_{14}ClN_3O_3$ $[M+H]^+$: 284.0802, found: 284.0799; Anal calcd. for $C_{12}H_{14}ClN_3O_3$: C, 50.80; H, 4.97; N, 14.81, found: C, 50.66; H, 4.73; N, 14.61.

Further the compounds **8b-e** was synthesized using the procedure similar to compound **8a** from the corresponding substituted piperazines.

2-Chloro-1-(4-(pyrimidin-2-yl)piperazin-1-yl)ethanone (8b)

The title compound was synthesized from 2-(piperazine-1-yl)pyrimidine (**7b**) as off-white solid (yield 81.5%); mp: 75-76 °C; IR (KBr) ν (cm^{-1}): 3018, 2399, 1647, 1586, 1552, 1497, 1442, 1358; 1H NMR (300 MHz, $CDCl_3$): δ 8.35 (2H, d, $J = 4.4$ Hz), 6.58 (1H, t, $J = 4.4$ Hz), 4.14-4.12 (2H, m), 3.92-3.85 (4H, m), 3.73-3.71 (2H, m), 3.62-3.61 (2H, m); HRMS (ESI positive) m/z calcd. for $C_{10}H_{13}ClN_4O$ $[M+H]^+$ 241.0856, found: 241.0855; Anal calcd. for $C_{10}H_{13}ClN_4O$: C, 49.90; H, 5.44; N, 23.28, found: C, 49.66; H, 5.28; N, 23.02.

2-Chloro-1-(4-(2-methoxyphenyl)piperazin-1-yl)ethanone (8c)

The title compound was synthesized from 1-(2-methoxyphenyl)piperazine (**7c**) as off-white solid (yield 54%); mp: 78-79 °C; IR (KBr) ν (cm^{-1}): 3018, 2832, 2401, 1648, 1595, 1500, 1450; 1H NMR (300 MHz, $CDCl_3$): δ 7.07-7.02 (1H, m), 6.96-6.87 (3H, m), 4.11 (2H, bs), 3.88 (3H, s), 3.81 (2H, t, $J = 4.2$ Hz), 3.71-3.68 (2H, m) 3.12-3.03 (4H, m); ^{13}C NMR (100 MHz, $CDCl_3$): δ 165.1, 152.2, 140.3, 123.7, 121.0, 118.5, 111.4, 55.4, 50.8, 50.3, 46.5, 42.3, 40.9; HRMS (ESI positive) m/z calcd. for $C_{13}H_{17}ClN_2O_2$ $[M+H]^+$: 269.1057, found: 269.1052; Anal calcd. for $C_{13}H_{17}ClN_2O_2$: C, 58.10; H, 6.38; N, 10.42, found: C, 58.29; H, 6.58; N, 10.69.

2-Chloro-1-(4-(4-nitro-2-(trifluoromethyl)phenyl)piperazin-1-yl)ethanone (8d)

The title compound was synthesized from 1-(4-nitro-2-(trifluoromethyl)phenyl)piperazine (**7d**) to give pure yellow oily compound (yield 80%); IR (neat): ν (cm^{-1}): 3019, 2925, 1648, 1613, 1526, 1425, 1384, 1347; 1H NMR (400 MHz, $CDCl_3$): δ 8.51 (1H, d, $J = 2.6$ Hz), 8.36 (1H, dd, $J = 2.6, 8.9$ Hz), 7.34 (1H, d, $J = 8.9$ Hz), 4.10 (2H, bs), 3.79 (2H, t, $J = 4.8$ Hz), 3.71-3.69 (2H, m), 3.14 (2H, t, $J = 4.7$ Hz), 3.09 (2H, t, $J = 4.8$ Hz); ^{13}C NMR (100 MHz, $CDCl_3$): δ 165.2, 156.5, 143.3, 128.0, 126.0, 125.7, 124.2, 123.4, 52.9, 52.3, 46.3, 42.1, 40.8; HRMS (ESI positive) m/z calcd. for $C_{13}H_{13}ClF_3N_3O_3$ $[M+H]^+$: 352.0676, found: 352.0675; Anal calcd. for $C_{13}H_{13}ClF_3N_3O_3$: C, 44.39; H, 3.73; N, 11.95, found: C, 44.56; H, 3.91; N, 12.19.

2-Chloro-1-(4-(2-nitro-4-(trifluoromethyl)phenyl)piperazin-1-yl)ethanone (8e)

The title compound was prepared from 1-(2-nitro-4-(trifluoromethyl)phenyl)piperazine (**7e**) as pure yellow oily compound (yield 89%); IR (neat) ν (cm^{-1}): 3429, 3016, 2967, 1654, 1535, 1436,

1328; ¹H NMR (300 MHz, CDCl₃): δ 8.11 (1H, bs), 7.73 (1H, d, *J* = 8.8 Hz, Ar-H), 7.21 (1H, d, *J* = 8.7 Hz), 4.11 (2H, bs), 3.83 (2H, bs),), 3.73 (2H, bs), 3.21-3.20 (4H, m); HRMS (ESI positive) *m/z* calcd for C₁₃H₁₃ClF₃N₃O₃ [M+H]⁺: 352.0676, found: 352.0657; Anal calcd for C₁₃H₁₃ClF₃N₃O₃: C, 44.39; H, 3.73; N, 11.95, found: C, 44.15; H, 3.51; N, 11.70.

1, 2-Bis(4-(4-nitrophenyl)piperazin-1-yl)ethanone (9a)

To the mixture of **8a** (0.3 g, 1.06 mmol) and Et₃N (0.3 mL, 2.12 mmol) in CHCl₃ (5 mL) was added 1-(4-nitrophenyl)piperazine (**7a**, 0.320 g, 1.59 mmol) in 5 mL CHCl₃ dropwise within 1 h. After complete addition reaction mixture was further stirred in an oil bath at 80-85 °C for 15 h. The reaction mixture was cooled, washed with water (5 mL × 3) and the organic layer was separated. Combined organic layer was dried (anhyd. Na₂SO₄) and concentrated under reduced pressure in rotavapor. The solid obtained was purified by recrystallization using EtOAc/Hexane which furnished yellow crystals (yield 81%); mp: 156-157 °C; IR (KBr) ν (cm⁻¹): 3019, 2399, 1640, 1597, 1506, 1423, 1330; ¹H NMR (400 MHz, CDCl₃): δ 8.14-8.09 (4H, m), 6.84-6.81 (4H, m), 3.84-3.83 (4H, m), 3.49-3.44 (8H, m), 3.33 (2H, s), 2.72 (4H, t, *J* = 5.0 Hz); ¹³C NMR (75.4 MHz, CDCl₃): δ 167.7, 154.7, 154.3, 138.8, 138.4, 125.9, 125.8, 112.9, 112.7, 60.8, 52.5, 46.9, 46.7, 44.6; HRMS (ESI positive) *m/z* calcd. for C₂₂H₂₆N₆O₅ [M+H]⁺: 455.2043, found: 455.2034; Anal calcd. for C₂₂H₂₆N₆O₅: C, 58.14; H, 5.77; N, 18.49, found: C, 58.31; H, 5.92; N, 18.66.

The compounds **9b-n** were prepared by adopting the procedure similar to **9a** using their respective precursors.

2-(4-(2-Methoxyphenyl)piperazin-1-yl)-1-(4-(4-nitrophenyl)piperazin-1-yl)ethanone (9b)

The title compound was synthesized from **8a** and 1-(2-methoxyphenyl)piperazine (**7c**) as yellow solid (yield 78%); mp: 123-124 °C; IR (KBr) ν (cm⁻¹): 3020, 2833, 2401, 1641, 1596, 1503, 1445, 132; ¹H NMR (300 MHz, CDCl₃): δ 8.14 (2H, d, *J* = 8.5 Hz), 7.01-6.98 (1H, m), 6.93 (2H, bs), 6.88-6.82 (3H, m), 3.86-3.80 (7H, m), 3.48-3.44 (4H, m), 3.31 (2H, s), 3.10 (4H, bs), 2.73 (4H, bs); ¹³C NMR (100 MHz, CDCl₃): δ 168.3, 154.5, 152.2, 141.0, 138.9, 125.9, 123.0, 120.9, 118.1, 112.9, 111.3, 61.6, 55.4, 53.4, 50.6, 47.4, 46.8, 44.8, 41.2; HRMS (ESI positive) *m/z* calcd. for C₂₃H₂₉N₅O₄ [M+H]⁺: 440.2298, found: 440.2283; Anal calcd. for C₂₃H₂₉N₅O₄: C, 62.85; H, 6.65; N, 15.93, found: C, 62.63; H, 6.20; N, 15.82.

1-(4-(4-Nitrophenyl)piperazin-1-yl)-2-(4-(pyrimidin-2-yl)piperazin-1-yl)ethanone (9c)

The title compound was synthesized from **8a** and 2-(piperazine-1-yl)pyrimidine (**7b**) as yellow solid (yield 68%); mp: 151-152 °C; IR (KBr) ν (cm⁻¹): 3020, 2401, 1642, 1592, 1506, 1443, 1329; ¹H NMR (300 MHz, CDCl₃): δ 8.31 (2H, d, J = 4.5 Hz), 8.13 (2H, d, J = 8.6 Hz), 6.84 (2H, d, J = 8.7 Hz), 6.51 (1H, t, J = 4.5 Hz), 3.85 (8H, bs), 3.49-3.46 (4H, m), 3.29 (2H, s), 2.60 (4H, bs); ¹³C NMR (100 MHz, CDCl₃): δ 168.0, 161.6, 157.6, 154.4, 138.9, 125.9, 112.9, 110.0, 61.4, 53.0, 47.3, 46.8, 44.8, 43.6, 41.2; HRMS (ESI positive) m/z calcd. for C₂₀H₂₅N₇O₃ [M+H]⁺: 412.2097, found: 412.2096; Anal calcd. for C₂₀H₂₅N₇O₃: C, 58.38; H, 6.12; N, 23.83, found: C, 58.60; H, 6.31; N, 23.99.

1-(4-(4-Nitrophenyl)piperazin-1-yl)-2-(4-(pyridin-2-yl)piperazin-1-yl)ethanone (9d)

The title compound was synthesized from **8a** and 1-(pyridine-2-yl)piperazine (**10a**) as yellow solid (yield 65%); mp: 135-136 °C; IR (KBr) ν (cm⁻¹): 3018, 2402, 1699, 1641, 1596, 1485, 1439, 1384, 1325; ¹H NMR (300 MHz, CDCl₃): δ 8.19-8.13 (3H, m), 7.49 (1H, t, J = 7.4 Hz), 6.83 (2H, d, J = 8.8 Hz), 6.67-6.62 (2H, m), 3.84 (4H, d, J = 16.2 Hz), 3.56-3.45 (8H, m), 3.30 (2H, bs), 2.66-2.65 (4H, m); ¹³C NMR (75.4 MHz, CDCl₃): δ 168.0, 161.6, 157.6, 154.4, 138.9, 125.9, 110.0, 61.4, 53.0, 47.3, 46.8, 44.8, 43.6, 41.2; HRMS (ESI positive) m/z calcd. for C₂₁H₂₆N₆O₃ [M+H]⁺: 411.2145, found: 411.2160; Anal calcd. for C₂₁H₂₆N₆O₃: C, 61.45; H, 6.38; N, 20.47, found: C, 61.62; H, 6.55; N, 20.69.

2-(4-(4-Nitrophenyl)piperazin-1-yl)-1-(4-(pyrimidin-2-yl)piperazin-1-yl)ethanone (9e)

The title compound was synthesized from **8b** and 1-(4-nitrophenyl)piperazine (**7a**) as yellow solid (yield 65%); mp: 174-175 °C; IR (KBr) ν (cm⁻¹): 3020, 2401, 1639, 1592, 1502, 1441, 1329; ¹H NMR (300 MHz, CDCl₃): δ 8.33 (2H, d, J = 4.6 Hz), 8.11 (2H, d, J = 9.1 Hz), 6.82 (2H, d, J = 8.6 Hz), 6.57-6.54 (1H, m), 3.85 (4H, bs), 3.70 (4H, bs), 3.45 (4H, bs), 3.32 (2H, s), 2.72-2.71 (4H, m); ESI-MS m/z calcd for C₂₀H₂₅N₇O₃ [M+H]⁺: 412; Anal calcd. for C₂₀H₂₅N₇O₃: C, 58.38; H, 6.12; N, 23.83, found: C, 58.63; H, 6.28; N, 23.99.

1, 2-Bis(4-(pyrimidin-2-yl)piperazin-1-yl)ethanone (9f)

The title compound was synthesized from **8b** and 2-(piperazine-1-yl) pyrimidine (**7b**) as off-white solid (yield 63%); mp: 150-151 °C; IR (KBr) ν (cm⁻¹): 3011, 2853, 1637, 1585, 1549, 1495, 1446, 1306; ¹H NMR (300 MHz, CDCl₃): δ 8.33-8.29 (4H, m), 6.56-6.48 (2H, m), 3.86-3.84 (8H, m), 3.72 (4H, bs), 3.28 (2H, s) 2.60 (4H, bs); ¹³C NMR (100 MHz, CDCl₃): δ 168.0, 161.6, 161.5, 157.7, 157.6, 110.4, 109.9, 61.5, 53.0, 45.5, 44.1, 43.6, 41.7; HRMS (ESI positive)

m/z calcd. for $C_{18}H_{24}N_8O$ $[M+H]^+$: 369.2151, found: 369.2142; Anal calcd. for $C_{18}H_{24}N_8O$: C, 58.68; H, 6.57; N, 30.41, found C, 58.86; H, 6.66; N, 30.58.

2-(4-(2-Methoxyphenyl)piperazin-1-yl)-1-(4-(pyrimidin-2-yl)piperazin-1-yl)ethanone (9g)

The title compound was synthesized from **8b** and 1-(2-methoxyphenyl)piperazine (**7c**) as off-white solid (yield 78%); mp: 96-97 °C; IR (KBr) ν (cm^{-1}): 3430, 3017, 1637, 1586, 1551, 1498, 1447, 1305; 1H NMR (300 MHz, $CDCl_3$): δ 8.32 (2H, d, $J = 4.5$ Hz), 6.99-6.84 (4H, m), 6.53 (1H, t, $J = 4.6$ Hz), 3.85 (7H, bs), 3.72 (4H, bs) 3.30 (2H, s), 3.10 (4H, bs), 2.73 (4H, bs); ^{13}C NMR (75.4 MHz, $CDCl_3$): δ 168.4, 161.6, 157.8, 152.3, 141.2, 123.0, 121.0, 118.3, 111.4, 110.5, 61.6, 55.4, 53.4, 50.6, 45.6, 44.2, 43.7, 41.8; HRMS (ESI positive) m/z calcd. for $C_{21}H_{28}N_6O_2$ $[M+H]^+$: 397.2352, found: 397.2351; Anal calcd. for $C_{21}H_{28}N_6O_2$: C, 63.62; H, 7.12; N, 21.20, found: C, 63.76; H, 7.22; N, 21.38.

2-(4-(Pyridine-2-yl)piperazin-1-yl)-1-(4-(pyrimidin-2-yl)piperazin-1-yl)ethanone (9h)

The title compound was synthesized from **8b** and 1-(pyridine-2-yl)piperazine (**10a**) as white solid (yield 80%); mp: 123-124 °C; IR (KBr) ν (cm^{-1}): 3020, 2401, 1593, 1519, 1478, 1431; 1H NMR (300 MHz, $CDCl_3$): δ 8.32 (2H, d, $J = 4.6$ Hz), 8.18 (1H, d, $J = 4.1$ Hz), 7.50-7.45 (1H, m), 6.66-6.61 (2H, m), 6.55-6.52 (1H, m), 3.85-3.84 (4H, m), 3.73-3.71 (4H, m), 3.57-3.56 (4H, m), 3.29 (2H, s), 2.67-2.65 (4H, m); ^{13}C NMR (75.4 MHz, $CDCl_3$): δ 168.2, 161.7, 159.6, 157.9, 148.1, 137.6, 113.6, 110.6, 107.3, 61.7, 53.1, 45.7, 45.4, 44.3, 43.8, 41.9; HRMS (ESI positive) m/z calcd. for $C_{19}H_{25}N_7O$ $[M+H]^+$: 368.2199, found: 368.2185; Anal calcd. for $C_{19}H_{25}N_7O$: C, 62.10; H, 6.86; N, 26.68, found: C, 62.28; H, 6.95; N, 26.86.

1-(4-(2-Methoxyphenyl)piperazin-1-yl)-2-(4-(4-nitrophenyl)piperazin-1-yl)ethanone (9i)

The title compound was synthesized from **8c** and 1-(4-nitrophenyl)piperazine (**7a**) as yellow solid (yield 84%); mp: 85-86 °C; IR (KBr) ν (cm^{-1}): 3018, 1633, 1597, 1500, 1329; 1H NMR (300 MHz, $CDCl_3$): δ 8.13 (2H, d, $J = 8.1$ Hz), 7.06-7.03 (1H, m), 6.94-6.89 (3H, m), 6.83 (2H, d, $J = 8.6$ Hz), 3.89 (3H, s), 3.82-3.80 (4H, m), 3.46 (4H, bs), 3.32 (2H, s), 3.08-3.05 (4H, m), 2.72 (4H, bs); ^{13}C NMR (75.4 MHz, $CDCl_3$): 167.4, 154.8, 152.2, 140.5, 138.5, 125.9, 123.6, 121.0, 118.3, 112.7, 111.3, 60.8, 55.4, 52.6, 51.2, 50.7, 47.0; HRMS (ESI positive) m/z calcd. for $C_{23}H_{29}N_5O_4$ $[M+H]^+$: 440.2298, found: 440.2274; Anal calcd. for $C_{23}H_{29}N_5O_4$: C, 62.85; H, 6.65; N, 15.93, found: C, 62.99; H, 6.76; N, 16.09.

1, 2-Bis(4-(2-methoxyphenyl)piperazin-1-yl)ethanone (9j)

The title compound was synthesized from **8c** and 1-(2-methoxyphenyl)piperazine (**7c**) as brown crystals (yield 63%); mp: 144-145 °C; IR (KBr) ν (cm⁻¹): 3019, 2854, 1632, 1500, 1465, 1384; ¹H NMR (400 MHz, CDCl₃): δ 7.03-6.97 (2H, m), 6.94-6.84 (6H, m), 3.88-3.80 (10H, m), 3.30 (2H, s), 3.09-3.01 (8H, m) 2.73 (4H, bs); ¹³C NMR (75.4 MHz, CDCl₃): δ 167.9, 152.2, 141.2, 140.7, 123.5, 122.9, 121.0, 120.9, 118.4, 118.2, 111.4, 111.3, 61.4, 55.5, 55.4, 53.3, 51.3, 50.7, 50.5, 45.9, 42.0; HRMS (ESI positive) m/z calcd. for C₂₄H₃₂N₄O₃ [M+H]⁺: 425.2553, found: 425.2551; Anal calcd. for C₂₄H₃₂N₄O₃: C, 67.90; H, 7.60; N, 13.20, found: C, 67.79; H, 7.48; N, 13.03.

1-(4-(2-Methoxyphenyl)piperazin-1-yl)-2-(4-(pyridin-2-yl)piperazin-1-yl)ethanone (9k)

The title compound was synthesized from **8c** and 1-(pyridine-2-yl)piperazine (**10a**) as brown crystals (yield 55%); mp: 124-125 °C; IR (KBr) ν (cm⁻¹): 3018, 2833, 2402, 1634, 1478, 1439, 1385; ¹H NMR (400 MHz, CDCl₃): δ 8.18-8.16 (1H, m), 7.49-7.44 (1H, m), 7.04-7.00 (1H, m), 6.94-6.86 (3H, m), 6.65-6.60 (2H, m) 3.87 (3H, s), 3.83-3.79 (4H, m), 3.55 (4H, t, *J* = 4.5 Hz), 3.28 (2H, s), 3.06 (2H, t, *J* = 4.8 Hz), 3.02 (2H, t, *J* = 5.0 Hz), 2.65 (4H, t, *J* = 5.0 Hz); ¹³C NMR (75.4 MHz, CDCl₃): δ 167.8, 159.4, 152.2, 147.8, 140.7, 137.4, 123.5, 121.0, 118.4, 113.4, 111.3, 107.1, 61.3, 55.4, 52.9, 51.2, 50.7, 45.9, 45.2, 42.0; HRMS (ESI positive) m/z calcd. for C₂₂H₂₉N₅O₂ [M+H]⁺: 396.2400, found: 396.2387; Anal calcd. for C₂₂H₂₉N₅O₂: C, 66.81; H, 7.39; N, 17.71, found: C, 66.99; H, 7.52; N, 17.96.

1-(4-(2-Methoxyphenyl)piperazin-1-yl)-2-(4-(pyrimidin-2-yl)piperazin-1-yl)ethanone (9l)

The title compound was synthesized from **8c** and 2-(piperazine-1-yl)pyrimidine (**7b**) as brown crystals (yield 67%); mp: 129-130 °C; IR (KBr) ν (cm⁻¹): 3022, 1634, 1587, 1549, 1500, 1447, 1384, 1307; ¹H NMR (400 MHz, CDCl₃): δ 8.30 (2H, d, *J* = 4.7 Hz), 7.05-7.01 (1H, m), 6.95-6.87 (3H, m), 6.50-6.47 (1H, m), 3.88 (3H, s), 3.86-3.82 (8H, m), 3.28 (2H, s), 3.09-3.02 (4H, m), 2.60 (4H, t, *J* = 4.9 Hz); ¹³C NMR (100 MHz, CDCl₃): δ 167.8, 161.6, 157.6, 152.2, 140.7, 123.5, 121.0, 118.4, 111.3, 109.9, 61.3, 55.4, 52.9, 51.2, 50.7, 46.0, 43.6, 42.0; HRMS (ESI positive) m/z calcd. for C₂₁H₂₈N₆O₂ [M+H]⁺: 397.2352, found: 397.2351; Anal calcd. for C₂₁H₂₈N₆O₂: C, 63.62; H, 7.12; N, 21.20, found: C, 63.48; H, 7.00; N, 21.06.

1, 2-Bis(4-(4-nitro-2-(trifluoromethyl)phenyl)piperazin-1-yl)ethanone (9m)

The title compound was synthesized from **8d** and 1-(4-nitro-2-(trifluoromethyl)phenyl)piperazine (**7d**) as yellow oily compound (yield 86%); IR (neat) ν (cm⁻¹

¹): 3020, 2834, 2402, 1710, 1643, 1611, 1523, 1428, 1344; ¹H NMR (400 MHz, CDCl₃): δ 8.56-8.51 (2H, m), 8.39-8.32 (2H, m), 7.34-7.30 (2H, m), 3.82 (4H, bs), 3.34 (2H, s), 3.21-3.19 (4H, m), 3.13-3.10 (4H, m), 2.76-2.75 (4H, m); ¹³C NMR (75.4 MHz, CDCl₃): δ 167.8, 156.8, 156.5, 143.0, 142.2, 128.0, 127.8, 124.6, 124.5, 124.4, 124.3, 123.0, 122.3, 60.8, 53.0, 52.6, 45.6, 41.7; HRMS (ESI positive) *m/z* calcd. for C₂₄H₂₄F₆N₆O₅ [M+H]⁺: 591.1791, found: 591.1790; Anal calcd. for C₂₄H₂₄F₆N₆O₅: C, 48.82; H, 4.10; N, 14.23, found: C, 48.66; H, 4.05; N, 14.06.

1, 2-Bis(4-(2-nitro-4-(trifluoromethyl)phenyl)piperazin-1-yl)ethanone (9n)

The title compound was synthesized from **8e** and 1-(2-nitro-4-(trifluoromethyl)phenyl)piperazine (**7e**) as pure yellow oily compound (yield 83%); IR (neat) ν (cm⁻¹): 3020, 2928, 2401, 1672, 1532, 1428, 1326; ¹H NMR (500 MHz, CDCl₃) δ 8.03-7.99 (2H, m), 7.63-7.62 (2H, m), 7.20-7.13 (2H, m), 3.71-3.57 (6H, m), 3.42-3.33 (4H, m) 3.11 (8H, bs); ¹³C NMR (100 MHz, CDCl₃): δ 164.9, 147.6, 147.4, 141.5, 141.2, 130.4, 124.1, 121.6, 121.1, 58.5, 52.7, 51.3, 50.7, 50.4, 49.6, 49.4, 45.0, 41.6; HRMS (ESI positive) *m/z* calcd. for C₂₄H₂₄F₆N₆O₅ [M+H]⁺: 591.1791, found: 591.1776; Anal calcd. for C₂₄H₂₄F₆N₆O₅: C, 48.82; H, 4.10; N, 14.23, found: C, 48.96; H, 4.25; N, 14.36.

4-Nitrophenyl 4-(pyridin-2-yl)piperazine-1-carboxylate (11a)

To the mixture of 1-(pyridin-2-yl)piperazine (**10a**, 0.4 mL, 2.74 mmol) and Et₃N (0.6 mL, 4.11 mmol) in DCM (10 mL) was added 4-nitrophenyl chloroformate (718 mg, 3.56 mmol) in 5 mL DCM) dropwise in 0.5 h duration in ice-bath (0-5 °C). After complete addition reaction mixture was further stirred at room temperature for 1 h. It was concentrated under reduced pressure and extracted with EtOAc (10 mL x 3). EtOAc layer was washed with water (5 mL x 3), dried (anhyd. Na₂SO₄) and concentrated on rotavapor. The solid obtained was further purified by recrystallization using EtOAc/Hexane as yellow solid (yield 87%); mp: 138-139 °C; IR (KBr) ν (cm⁻¹): 3021, 2483, 1723, 1657, 1594, 1340; ¹H NMR (400 MHz, CDCl₃) δ 8.23-8.20 (3H, m), 7.56-7.52 (1H, m), 7.34-7.30 (2H, m), 6.72-6.69 (2H, m), 3.71-3.63 (8H, m); ¹³C NMR (75.4 MHz, CDCl₃): δ 163.5, 158.9, 156.0, 152.3, 147.8, 144.8, 137.9, 125.0, 122.3, 115.5, 114.1, 107.6, 58.0, 45.0, 44.3; ESI-MS *m/z* calcd for C₁₆H₁₆N₄O₄ [M+H]⁺: 329; Anal calcd for C₁₆H₁₆N₄O₄: C, 58.53; H, 4.91; N, 17.06; Found C, 58.61; H, 4.85; N, 17.18.

4-Nitrophenyl 4-(4-nitrophenyl)piperazine-1-carboxylate (11b)

The title compound was synthesized using the procedure similar to **11a** from 1-(4-nitrophenyl)piperazine (**7a**) as yellow solid (yield 89%); mp: 125-126 °C; IR (neat) ν (cm⁻¹):

3020, 2836, 1611, 1567, 1434, 1346; ¹H NMR (400 MHz, CDCl₃): δ 8.18 (2H, d, *J* = 8.2 Hz), 8.07 (2H, d, *J* = 8.6 Hz), 7.25 (2H, d, *J* = 8.2 Hz), 6.78 (2H, d, *J* = 8.5 Hz), 3.76 (4H, d, *J* = 41.3 Hz), 3.48 (4H, bs); ¹³C NMR (75.4 MHz, CDCl₃): δ 162.2, 155.8, 154.4, 152.3, 145.0, 139.0, 126.1, 126.0, 125.1, 122.3, 115.6, 113.1, 46.7, 43.9, 43.3; ESI-MS *m/z* calcd for C₁₇H₁₆N₄O₆ [M+H]⁺: 373; Anal calcd. for C₁₇H₁₆N₄O₆: C, 54.84; H, 4.83; N, 15.05, found: C, 54.99; H, 4.66; N, 15.26.

(4-(4-Nitro-2-(trifluoromethyl)phenyl)piperazin-1-yl)(4-(pyridin-2-yl)piperazin-1-yl)methanone (12a)

The mixture of compound **11a** (411 mg, 1.25 mmol) and 1-(4-nitro-2-(trifluoromethyl)phenyl)piperazine (**7d**, 0.23 mL, 1.62 mmol) in DMF (10 mL) was stirred at 100-110 °C for 60 h. The reaction mixture was cooled and water (25 mL) was added. The crude product was extracted with EtOAc (10 mL x 3). EtOAc layer was washed with water (5 mL x 3), dried (anhyd. Na₂SO₄) and concentrated under reduced pressure. The product obtained was purified over column chromatography (230-400 mesh) using EtOAc/Hexane as eluent as yellow solid (yield 65%); mp: 162-163 °C; IR (neat) ν (cm⁻¹): 2924, 2854, 1596, 1482, 1426, 1338; ¹H NMR (400 MHz, CDCl₃): δ 8.54 (1H, d, *J* = 2.7 Hz), 8.37 (1H, dd, *J* = 2.6, 8.9 Hz), 8.21 (1H, dd, *J* = 1.9, 5.3 Hz), 7.52-7.50 (1H, m), 7.34-7.32 (1H, m), 6.69-6.66 (2H, m), 3.60-3.57 (4H, m), 3.52 (4H, t, *J* = 4.6 Hz), 3.46-3.43 (4H, m), 3.13 (4H, t, *J* = 4.7 Hz); ¹³C NMR (100 MHz, CDCl₃): δ 163.6, 159.3, 156.8, 147.9, 142.8, 137.6, 127.9, 125.5, 125.2, 124.5, 124.4, 124.3, 122.9, 113.8, 107.2, 52.6, 46.7, 46.5, 45.1; HRMS (ESI positive) *m/z* calcd. for C₂₁H₂₃F₃N₆O₃ [M+H]⁺: 465.1862, found: 465.1859; Anal calcd. for C₂₁H₂₃F₃N₆O₃: C, 54.31; H, 4.99; N, 18.09, found: C, 54.52; H, 4.89; N, 18.16.

The following compounds (**12b-g**) were prepared using a procedure similar to that described for compound **12a** from corresponding arylpiperazine and compound **11a-b**.

(4-(4-Nitro-3-(trifluoromethyl)phenyl)piperazin-1-yl)(4-(pyridin-2-yl)piperazin-1-yl)methanone (12b)

The title compound was synthesized from **11a** and 1-(4-nitro-3-(trifluoromethyl)phenyl)piperazine as yellow solid (yield 76%); mp: 133-134 °C; IR (neat) ν (cm⁻¹): 3020, 2927, 2401, 1598, 1522, 1478, 1427, 1338; ¹H NMR (400 MHz, CDCl₃): δ 8.21-8.19 (1H, m), 8.02 (1H, d, *J* = 9.2 Hz), 7.54-7.50 (1H, m), 7.16 (1H, d, *J* = 2.4 Hz), 6.95 (1H, dd, *J* = 2.7, 9.2 Hz), 6.69-6.66 (2H, m), 3.60-3.45 (16H, m); ¹³C NMR (100 MHz, CDCl₃): δ 163.5,

159.2, 153.1, 147.9, 137.6, 128.5, 126.3, 126.0, 115.0, 113.9, 112.3, 112.2, 107.3, 46.7, 46.4, 46.1, 45.0; HRMS (ESI positive) m/z calcd. for $C_{21}H_{23}F_3N_6O_3$ $[M+H]^+$: 465.1862, found: 465.1858; Anal calcd. for $C_{21}H_{23}F_3N_6O_3$: C, 54.31; H, 4.99; N, 18.09, found: C, 54.42; H, 4.81; N, 18.20.

Bis(4-(pyridin-2-yl)piperazin-1-yl)methanone (12c)

The title compound was synthesized from **11a** and 1-(pyridin-2-yl)piperazine (**10a**) as pure yellow solid compound (yield 73%); mp: 142-143 °C; IR (neat) ν (cm^{-1}): 3010, 2925, 2854, 2462, 1664, 1597, 1314; 1H NMR (400 MHz, $CDCl_3$): δ 8.19-8.11 (2H, m), 7.51-7.47 (2H, m), 6.66-6.63 (4H, m), 3.65-3.41 (16H, m); ^{13}C NMR (100 MHz, $CDCl_3$): δ 163.8, 160.8, 159.3, 147.9, 137.5, 113.7, 107.2, 46.5, 45.9, 45.2, 45.0; HRMS (ESI positive) m/z calcd. for $C_{19}H_{24}N_6O$ $[M+H]^+$: 353.2090, found: 353.2090; Anal calcd. for $C_{19}H_{24}N_6O$: C, 64.75; H, 6.86; N, 23.85, found: C, 64.60; H, 6.77; N, 23.94.

(4-(4-Nitrophenyl)piperazin-1-yl)(4-(pyridin-2-yl)piperazin-1-yl)methanone (12d)

The title compound was synthesized from **11a** and 1-(4-nitrophenyl)piperazine (**7a**) as solid compound of yellow colour (yield 72%); mp: 130-131 °C; IR (neat) ν (cm^{-1}): 3018, 2926, 2402, 1673, 1522, 1437; 1H NMR (400 MHz, $CDCl_3$): δ 8.21-8.19 (1H, m), 8.15-8.12 (2H, m), 7.54-7.49 (1H, m), 6.85-6.82 (2H, m), 6.69-6.66 (2H, m), 3.60-3.57 (4H, m), 3.50-3.44 (12H, m); ^{13}C NMR (100 MHz, $CDCl_3$): δ 163.6, 159.2, 154.6, 147.9, 138.8, 137.6, 125.9, 113.8, 112.8, 107.3, 46.7, 46.4, 46.2, 45.0; HRMS (ESI positive) m/z calcd. for $C_{20}H_{24}N_6O_3$ $[M+H]^+$: 397.1988, found: 397.1985; Anal calcd. for $C_{20}H_{24}N_6O_3$: C, 60.59; H, 6.10; N, 21.20, found: C, 60.44; H, 6.29; N, 21.36.

Bis(4-(4-nitrophenyl)piperazin-1-yl)methanone (12e)

The title compound was synthesized from **11b** and 1-(4-nitrophenyl)piperazine (**7a**) as yellow solid (yield 76%); mp: 145-146 °C; IR (neat) ν (cm^{-1}): 3020, 2926, 2401, 1638, 1596, 1505; 1H NMR (400 MHz, $CDCl_3$): δ 8.15-8.12 (4H, m), 6.86-6.82 (4H, m), 3.53-3.48 (16H, m); ^{13}C NMR (100 MHz, $CDCl_3$): δ 163.3, 154.6, 139.0, 125.9, 112.9, 46.8, 46.1; HRMS (ESI positive) m/z calcd. for $C_{21}H_{24}N_6O_5$ $[M+H]^+$: 441.1886, found: 441.1876; Anal calcd. for $C_{21}H_{24}N_6O_5$: C, 57.26; H, 5.49; N, 19.08, found: C, 57.44; H, 5.31; N, 19.19.

(4-(4-Nitro-2-(trifluoromethyl)phenyl)piperazin-1-yl)(4-(4-nitrophenyl)piperazin-1-yl)methanone (12f)

The title compound was synthesized from **11b** and 1-(4-nitro-2-(trifluoromethyl)phenyl)piperazine (**7d**) as yellow solid compound (yield 70%); mp: 137-138 °C; IR (neat) ν (cm⁻¹): 3021, 1596, 1407, 1328; ¹H NMR (400 MHz, CDCl₃): δ 8.53 (1H, d, *J* = 2.6 Hz), 8.36 (1H, dd, *J* = 2.6, 8.9 Hz), 8.14-8.11 (2H, m), 7.33 (1H, m, *J* = 8.9 Hz), 6.85-6.83 (2H, m), 3.55-3.47 (12H, m), 3.14 (4H, t, *J* = 4.8 Hz); ¹³C NMR (100 MHz, CDCl₃): δ 163.3, 156.7, 154.6, 142.9, 138.9, 128.0, 125.9, 124.5, 124.4, 124.3, 123.0, 112.9, 52.6, 46.8, 46.6, 46.2; HRMS (ESI positive) *m/z* calcd. for C₂₂H₂₃F₃N₆O₅ [M+H]⁺: 509.1760, found: 509.1739; Anal calcd. for C₂₂H₂₃F₃N₆O₅: C, 51.97; H, 4.56; N, 16.53, found: C, 51.77; H, 4.42; N, 16.62.

(4-(2-Nitro-4-(trifluoromethyl)phenyl)piperazin-1-yl)(4-(4-nitrophenyl)piperazin-1-yl)methanone (12g)

The title compound was synthesized from **11b** and 1-(2-nitro-4-(trifluoromethyl)phenyl)piperazine (**7e**) as yellow solid compound (yield 74%); mp: 141-142 °C; IR (neat) ν (cm⁻¹): 3016, 2433, 1627, 1404, 1326; ¹H NMR (400 MHz, CDCl₃): δ 8.15-8.09 (3H, m), 7.72 (1H, dd, *J* = 1.9, 8.7 Hz), 7.20 (1H, d, *J* = 8.6 Hz), 6.84 (2H, d, *J* = 9.4 Hz), 3.55-3.48 (12H, m), 3.20 (4H, t, *J* = 5 Hz); ¹³C NMR (100 MHz, CDCl₃): δ 163.2, 154.6, 147.9, 141.0, 138.9, 130.4, 130.3, 125.9, 124.2, 124.1, 123.3, 120.8, 112.9, 50.7, 46.8, 46.3, 46.2; HRMS (ESI positive) *m/z* calcd. for C₂₂H₂₃F₃N₆O₅ [M+H]⁺: 509.1760, found: 509.1743; Anal calcd. for C₂₂H₂₃F₃N₆O₅: C, 51.97; H, 4.56; N, 16.53, found: C, 51.81; H, 4.62; N, 16.44.

1.2. BIOLOGY

1.2.1. Cell cultures

The prostate cancer PC3 and DU145, monkey kidney Cos-7, and LNCaP cells were procured from National Centre for Cell sciences (NCCS, Pune, India). Human prostate cancer cells, PC3 and DU145, were maintained in DMEM/ HAM'S F-12 medium (Sigma-Aldrich, St. Louis, MO), and Cos-7 cells were grown in DMEM medium (Sigma-Aldrich, St. Louis, MO) supplemented with 10% FBS and 0.01% antibiotic/antimycotic solutions. The LNCaP cells were grown in Roswell Park Memorial Institute (RPMI) supplemented with 12.5% fetal bovine serum (charcoal stripped, Life Technologies Inc), and 0.01% antibiotic/antimycotic solutions in an atmosphere of 5% CO₂/95% air at 37 °C.

Ready-to-Assay α_{1a} Adrenergic Receptor GPCR frozen cells were used for rapid calcium assays and procured from Millipore (cat no. **HTS087RTA**).

1.2.2. Cell proliferation assay

The prostate cancer and normal cells were seeded in 96-well plates at a density of 2×10^4 cells/well and allowed 24 h for attachment. The treatment were proceeded in triplicate with flutamide (Stock 10mM; Sigma-Aldrich, USA) and with compounds (Stock 10 mM) at concentrations ranging from 2.5 μ M to 80 μ M in 2% charcoal stripped serum for 24 h at 37 °C in 5% CO₂ atmosphere. The stock solution of flutamide (10 mM) was dissolved in molecular grade dimethyl sulfoxide (DMSO) and diluted with culture medium to different concentrations. Final concentration of DMSO was not more than 0.05%. Controls were treated with DMSO (0.05% in culture medium). After 24 h of incubation in CO₂ incubator, 5 μ l of 5mg/ml MTT [3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetra-zolium bromide)] was added to the cells. After incubation of 3 h, the formazan crystals formed in viable cells were dissolved in DMSO with gentle shaking on a plate shaker for 5 min. The absorbance was measured at 540 nm using a microplate reader (Microquant, Bio-Tek, USA). The percent viability was calculated according to formula given below:

$$\% \text{ viability of cells} = \{(\text{OD of treatment})/\text{OD of Vehicle control}\} \times 100$$

1.2.3. Western blot analysis of PSA

LNCaP cells were seeded in culture flasks and incubated for overnight. After 60% confluency cells were treated with agonists, antagonists and respective compounds (testosterone, flutamide compounds **8a**, **8c** and **9a**). Cells were washed after 24 h of incubation, with ice cold PBS and whole cell lysates of LNCaP prepared in lysis buffer [25 mM tris-HCl, pH 7.6, 150 mM NaCl, 1% sodium deoxycholate, 0.1% SDS, 1% NP-40, 1% protease inhibitor cocktail] were incubated at room temperature for 30 min followed by centrifugation at 14,000 g (4 °C, 25 min). The protein concentration of the supernatant was determined by Bradford protein assay. Samples were boiled for 10 min in denaturing sample buffer (10% glycerol, 1% SDS, 1% β -mercaptoethanol, 10 mM tris-HCl, 0.01% bromophenol blue, pH 6.8), and then 20 μ g (protein) of each sample was separated on 10% acrylamide gel and transferred to immobilon-P PVDF membranes (Millipore). After blocking non-specific sites with 5% skimmed milk (in 150 mM NaCl, 10 mM Tris-HCl, pH-7.6) the membranes were probed overnight with antibodies for PSA (1:1000, Sigma-Aldrich), and re-probed with β -actin antibody (1:10,000, Sigma-Aldrich) for loading correction. Subsequently, the blots were washed three times with 0.1% tween 20 in TBS

and incubated with 1:3000 and 1:30,000 dilution of secondary antibody (anti-mouse-IgG-HRP conjugate). After washing in 0.1% tween-20/TBS, substrate solution was added to the membrane, which was incubated for 5 min and exposed at room temperature. The membranes were developed with enhanced chemiluminescence (ECL) kit by following the manufacturer's protocol.

1.2.4. Reporter Gene Assay by Luciferase Expression

AR-negative Cos7 cells were seeded at a density of 5×10^4 cells/well into 48-well plates on the day prior to transfection with 500ng of luciferase reporter gene construct pARE-eEIB-LUC along with the expression vector for AR, pCR 3.1.AR, using DharmaFECT transfection reagent according to manufacturer's protocol. At the end of 8 h transfection period, cells were switched to complete medium and thereafter, treated with test compounds and Bicalutamide (AR-antagonist, $3 \mu\text{M}$) was used as positive control for 24 h. Luciferase activity was determined with luciferase assay systems (Promega, Madison, WI) following the manufacturer's protocol to detect the AR mediated transcriptional activity. Luciferase activity was normalized for transfection efficiency using pRL-SV40-luc as an internal control.

1.2.5. Calcium mobilization assay

All the analyses was performed with α_{1a} -adrenergic receptor GPCR cells (Millipore) which was cultured immediately by immersing in a 37°C water bath in a vial pre-sterilized with 70% ethanol. 1 mL of pre-warmed media component was added to the vial of cells and the volume was raised to 10 mL with media component. Then cell suspension was centrifuged at $190 \times g$ for 4 min, supernatant removed and 5 mL of pre-warmed media was added to re-suspend the cell pellet. 4×10^4 cells per well were seeded overnight in 100 μL using 96-well plates and growth medium. After incubation overnight, the growth medium was removed from the cells and 100 μL /well fluoforte™ dye-loading solution (Enzo life sciences FluoForte Calcium Assay Kit cat. No. ENZ-51017) was added to each well. Thereafter, the culture plates were incubated for 45 min at 37°C and 15 min at room temperature. α_1 -agonist (epinephrine, $20 \mu\text{M}$), and antagonists (tamsulosin $1 \mu\text{M}$) and the test compounds **8a**, **8c** and **9a** ($10 \mu\text{M}$) were prepared and the calcium flux assay was run by monitoring the fluorescence at excitation wavelength at 490 nm and emission wavelength of 525 nm, using a fluorescence plate reader (Biotek Flx 800), for 180

sec with agonist addition at 10 sec. Calcium assay was optimized for agonist at one fifth of the final volume and relative fluorescence unit (RFU) values were determined for Ca²⁺ released in the cells.

1.2.6. *In vivo* experiment

Adult mature Sprague–Dawley rats (weighing 250–280 g), for *in vivo* study, were procured from the Animal Division of the CSIR-Central Drug Research Institute, Lucknow, India. BPH was induced in rats by a known disease causing agent citral.¹ Animals were erratically divided into five groups, viz. Group I (vehicle control); Group II (Citral 100 mg/kg); Group III (Citral 100 mg/kg + Flutamide 10 mg/kg); Group IV (Citral 100 mg/kg + **8a**) and Group V (Citral 100 mg/kg + **9a**). Groups II, III, IV, and V were given citral at a dose of 100 mg/kg once daily for 28 days while animals in Group III, IV and V were coadministered with flutamide, **8a** and **9a** respectively, at 10 mg/kg, treatment beginning from the 8th day of citral treatment and ongoing once daily for remaining 21 days. All compounds were administered orally. At the completion of experiment, animals were sacrificed according to the guidelines of Institutional Animal Ethics Committee, and the weights of prostate, epididymis, seminal vesicles and testis were taken. Prostatic tissues were fixed in 10% formalin solution for histology.

1.2.7. Histology

Prostate tissues kept overnight in 10% formalin solution were processed by following the standard protocol. 5.0 μ sections were taken and stained with hematoxyline & eosin and examined under a light microscope (Eclipse 80i, Nikon Corporation, Japan).

1.2.8. Pharmacokinetic Studies

The pharmacokinetic studies of **9a** was performed in young and healthy male *sprague dawley* rats weighing 250 ± 25 g obtained from Laboratory Animal Division, CDRI, Lucknow. The animals were housed in plastic cages in standard laboratory conditions with a regular 12 hour day-night cycle. Standard pelleted laboratory chow (Goldmohar Laboratory Animal Feed, Lipton India Ltd, Chandigarh, India) and water were allowed *ad libitum*. The rats were acclimatized to this environment for at least five days before conducting the experiment. The study was conducted on overnight fasted (12-16 hour) rats (n = 3 per time point). All experiments,

euthanasia and disposal of carcasses were done as per the guidelines of Local Ethics Committee for animal experimentation.

Suspension formulation containing 2.5 mg/mL of **9a** was prepared by triturating **9a**, gum acacia (1%, w/v) and water (drop wise addition) in a mortar and pestle. A single 10 mg/kg oral dose was given to conscious rats using rat feeding needle. The animals were sacrificed at various predefined times up to 72 h post dose and blood, prostate and hypothalamus were collected. Serum samples were harvested. The hypothalamus of three rats were pooled and homogenized in order to ensure a measurable quantity. All the samples were stored at -80 °C until analysis.

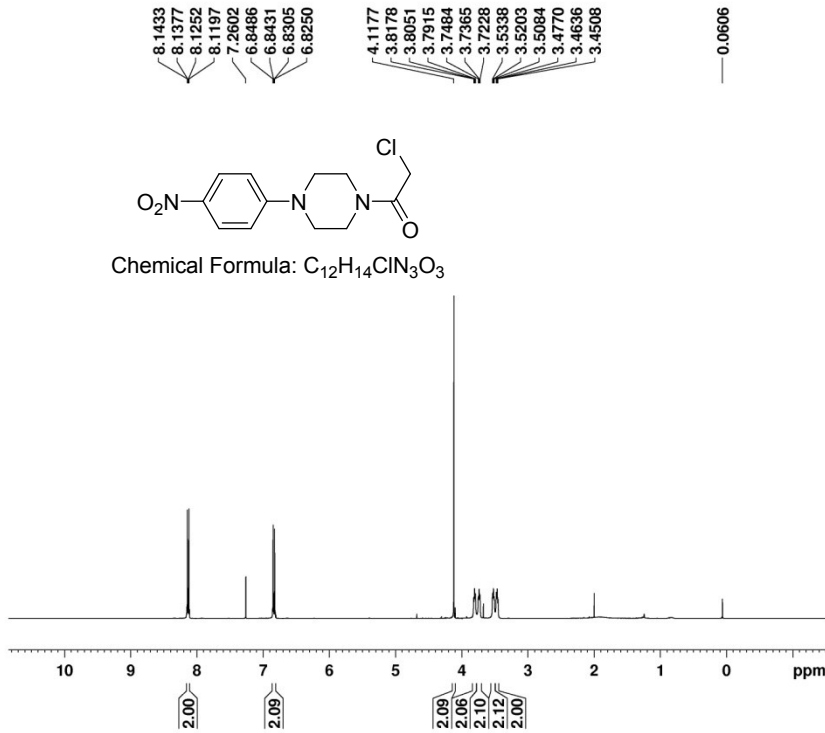
A Shimadzu UFLC pump (LC-20AD) with online degasser (DGU-20A3), an auto-sampler (SIL-HTc) with a temperature-controlled peltier-tray and a triple quadrupole API 4000 Q trap mass spectrometer (Applied Biosystems, Toronto, Canada) was used for analysis. Chromatographic separation was made on a Discovery HS C-18 column (5 µm, 100 x 4.6 mm id) preceded with a guard column (5 µm, 50 x 4.0 mm, id) packed with the same material with mobile phase [85% acetonitrile in aqueous ammonium acetate buffer (0.01 M)] pumped at a flow rate of 0.7 mL/min under isocratic condition. The mobile phase was degassed by ultrasonication for 15 min before use. LC-MS/MS system was equilibrated for approximately 20 min before commencement of analysis. The column oven temperature was 40 °C. Total analysis time was 3 min per sample. The mass spectral analysis was performed in positive ionization mode at 5500 V using multiple reaction monitoring technique to monitor the transitions m/z 455.3 \rightarrow m/z 220.1 for **9a** and m/z 180.1 \rightarrow m/z 138.2 for phenacetin (internal standard). Data acquisition and quantitation were performed using analyst software (version 1.4.2; AB Sciex, Toronto, Canada). The method was linear over the range of 2 – 200 ng/mL with recovery of >60% and acceptable accuracy and precision.²

1.2.9. Docking studies

The compounds were drawn by using the sketch module of the molecular modeling suite Sybyl 2.1.³ Tripos force field and Gasteiger–Huckel charges was applied to optimize the geometry of compounds. The energy minimization was done using the Powell method with an energy convergence gradient of 0.001 kcalmol⁻¹. To obtain the probable binding pose the active compounds were subjected to docking program Surflex-Dock.⁴ Surflex uses an empirical scoring function and a patented search engine to dock ligands into a protein's binding site and is based on

the Hammerhead scoring function and a consensus score that is the linear combination of non-linear functions of protein-ligand atomic surface distances. The interactions include steric, polar, entropic, and solvation terms. In addition, a total score is also generated. The crystal structure of wild type Androgen receptor was retrieved from the protein databank (PDBID: 2AXA), this is co-crystallised with bound ligand designated as S-1. For surflex program the protein receptor was prepared by removing the substructures and extracting ligand. The side chains of the protein structure then were fixed using default settings, water atoms removed, hydrogen added, unknown atom types were assigned and bumps were relaxed. The Kollman-all atom charges were assigned to protein atoms. Protocol was generated using the ligand based mode, residues Leu 704, Asn 705 and Arg 752 were kept as flexible. Docking poses were generated using Geom mode of Surflex-Dock program.

¹H NMR of compound 8a



VLS-SG-4/3

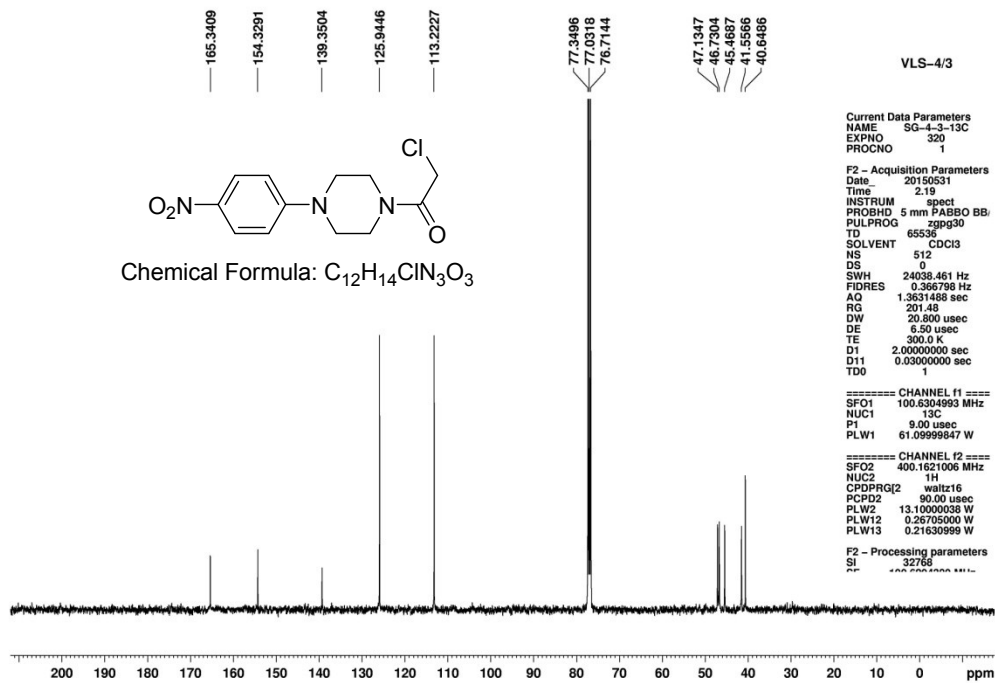
Current Data Parameters
 NAME SG-4-3-1H-NEW
 EXPNO 350
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20151005
 Time 23.42
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 8
 DS 0
 SWH 8012.820 Hz
 FIDRES 0.122266 Hz
 AQ 4.0894465 sec
 RG 100.41
 DW 62.400 usec
 DE 6.50 usec
 TE 300.0 K
 D1 1.00000000 sec
 TDO 1

==== CHANNEL f1 =====
 SFO1 400.1629712 MHz
 NUC1 1H
 P1 13.20 usec
 PLW1 13.00000000 W

F2 - Processing parameters
 SI 65536
 SF 400.1605091 MHz
 WDW no
 SSB 0
 LB 0 Hz
 GB 0
 PC 1.00

¹³C NMR of compound 8a



VLS-4/3

Current Data Parameters
 NAME SG-4-3-13C
 EXPNO 320
 PROCNO 1

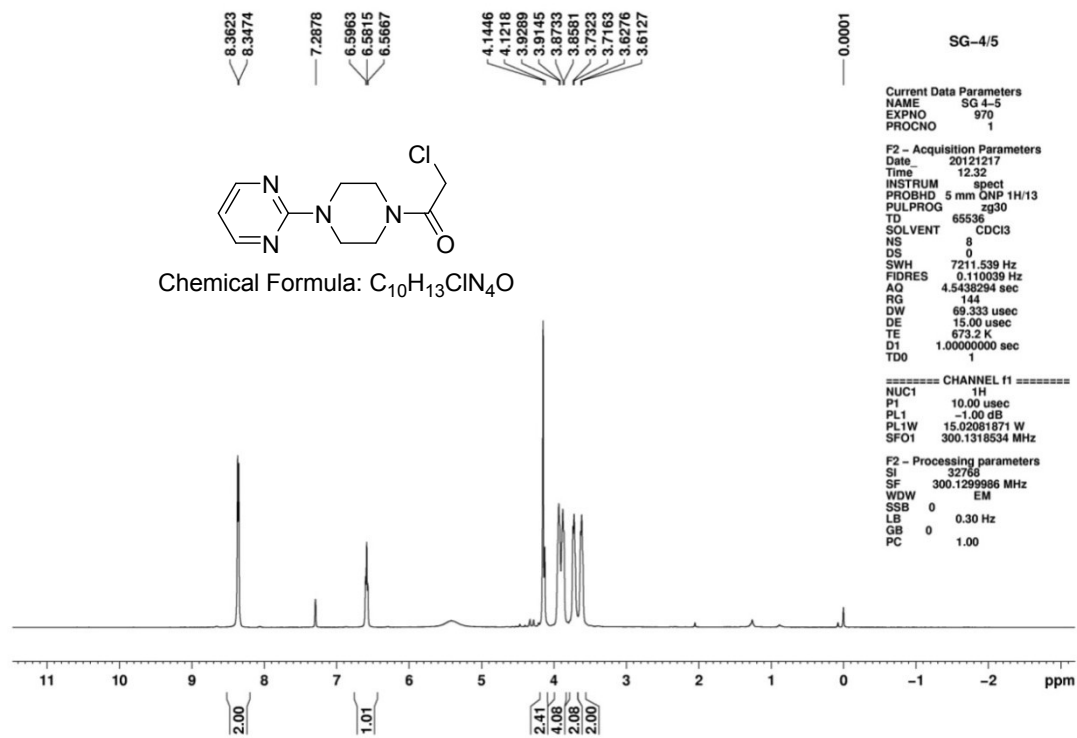
F2 - Acquisition Parameters
 Date_ 20150531
 Time 2.19
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 512
 DS 0
 SWH 24038.461 Hz
 FIDRES 0.366798 Hz
 AQ 1.3631468 sec
 RG 201.48
 DW 20.800 usec
 DE 6.50 usec
 TE 300.0 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TDO 1

==== CHANNEL f1 =====
 SFO1 100.6304993 MHz
 NUC1 13C
 P1 9.00 usec
 PLW1 61.09999847 W

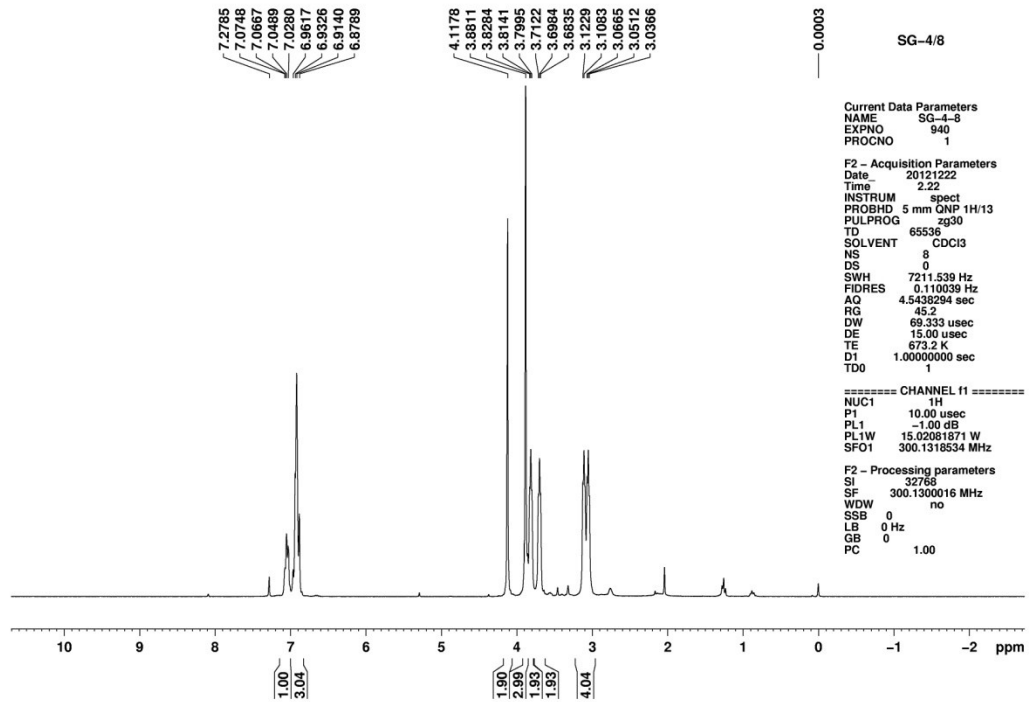
==== CHANNEL f2 =====
 SFO2 400.1621006 MHz
 NUC2 1H
 CPDPRG2 waltz16
 PCPD2 80.00 usec
 PLW2 13.10000000 W
 PLW12 0.26705000 W
 PLW13 0.21630999 W

F2 - Processing parameters
 SI 32768

¹H NMR of compound 8b



¹H NMR of compound 8c



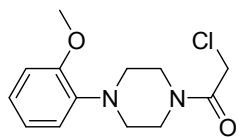
SG-4/8

Current Data Parameters
 NAME SG-4-8
 EXPNO 940
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20121222
 Time 2.22
 INSTRUM spect
 PROBHD 5 mm QNP 1H/13
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 8
 DS 0
 SWH 7211.539 Hz
 FIDRES 0.110039 Hz
 AQ 4.5438294 sec
 RG 45.2
 DW 69.333 usec
 DE 15.00 usec
 TE 673.2 K
 D1 1.00000000 sec
 TDO 1

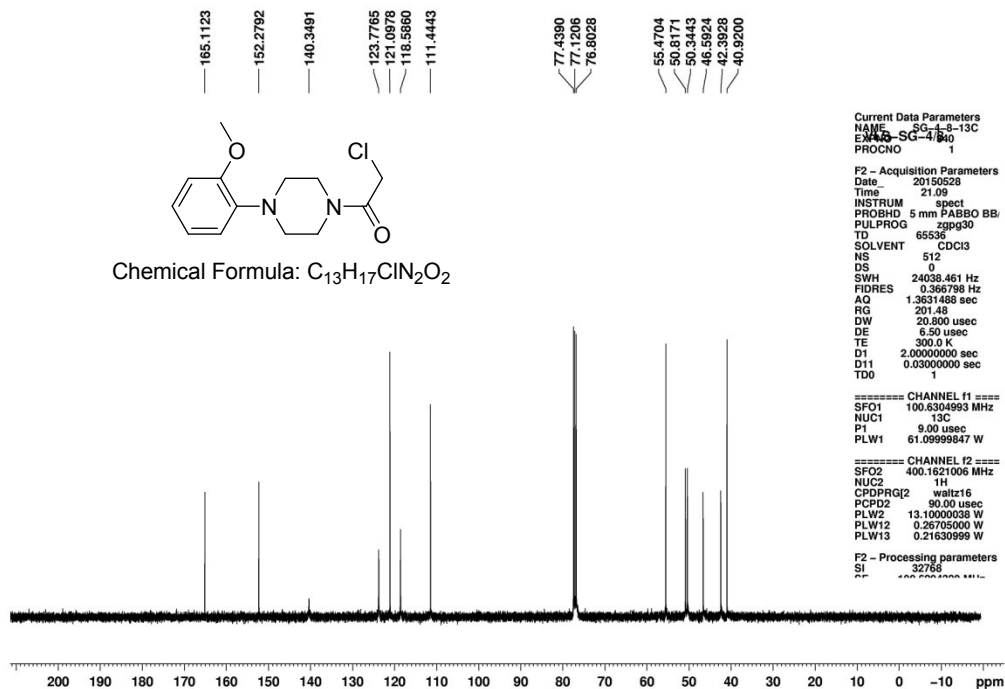
===== CHANNEL f1 =====
 NUC1 1H
 P1 10.00 usec
 PL1 -1.00 dB
 PL1W 15.02081871 W
 SFO1 300.1318534 MHz

F2 - Processing parameters
 SI 32768
 SF 300.1300016 MHz
 WDW no
 SSB 0
 LB 0 Hz
 GB 0
 PC 1.00

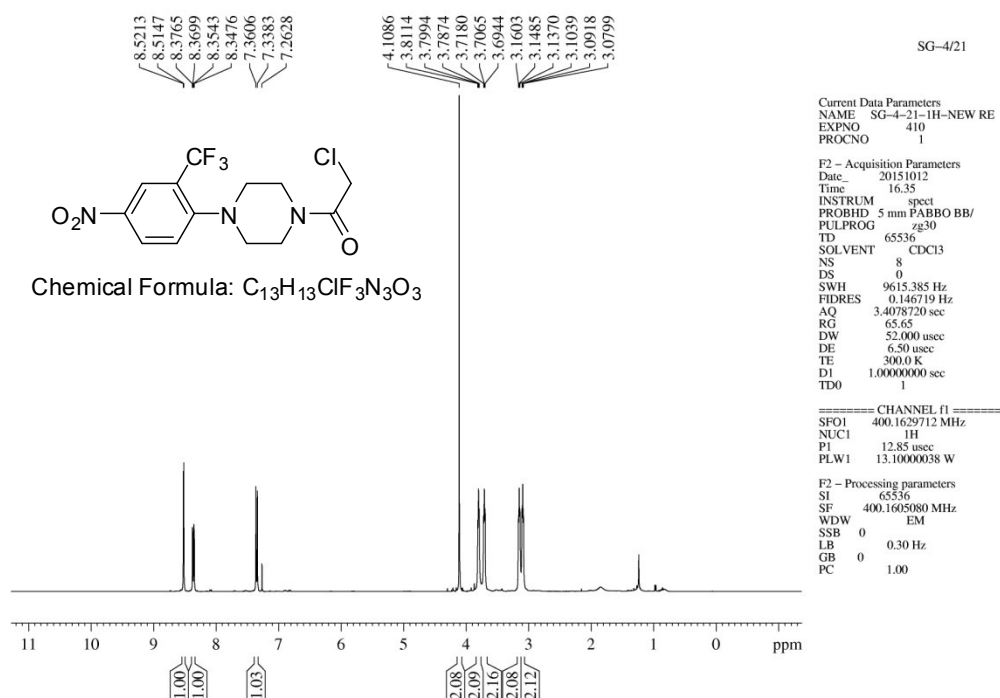


Chemical Formula: C₁₃H₁₇ClN₂O₂

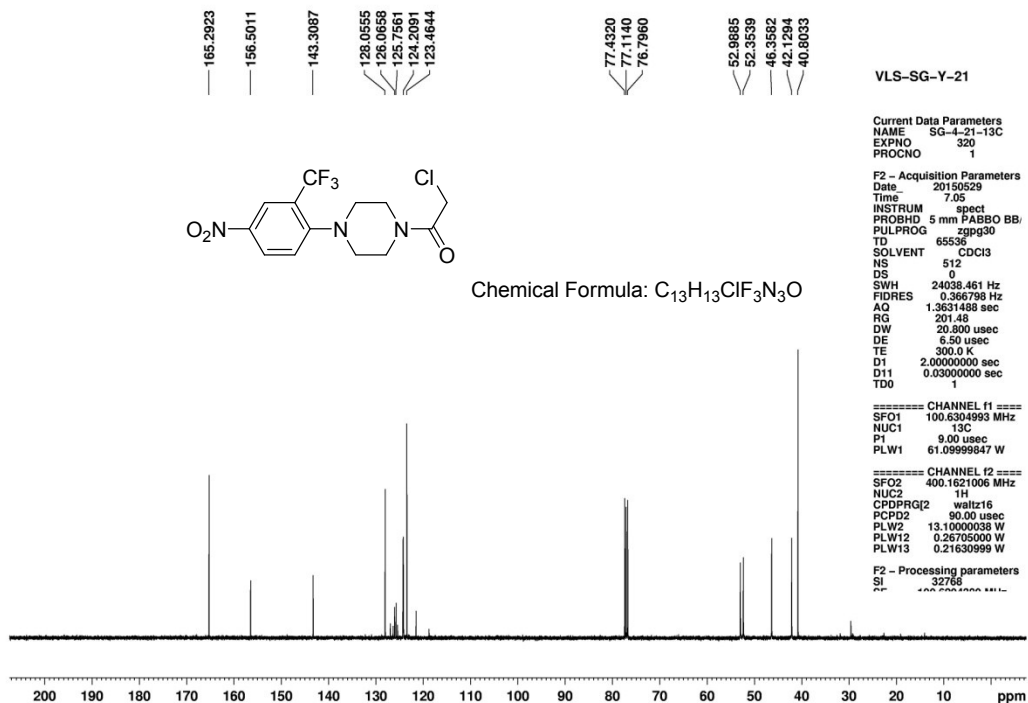
¹³C NMR of compound 8c



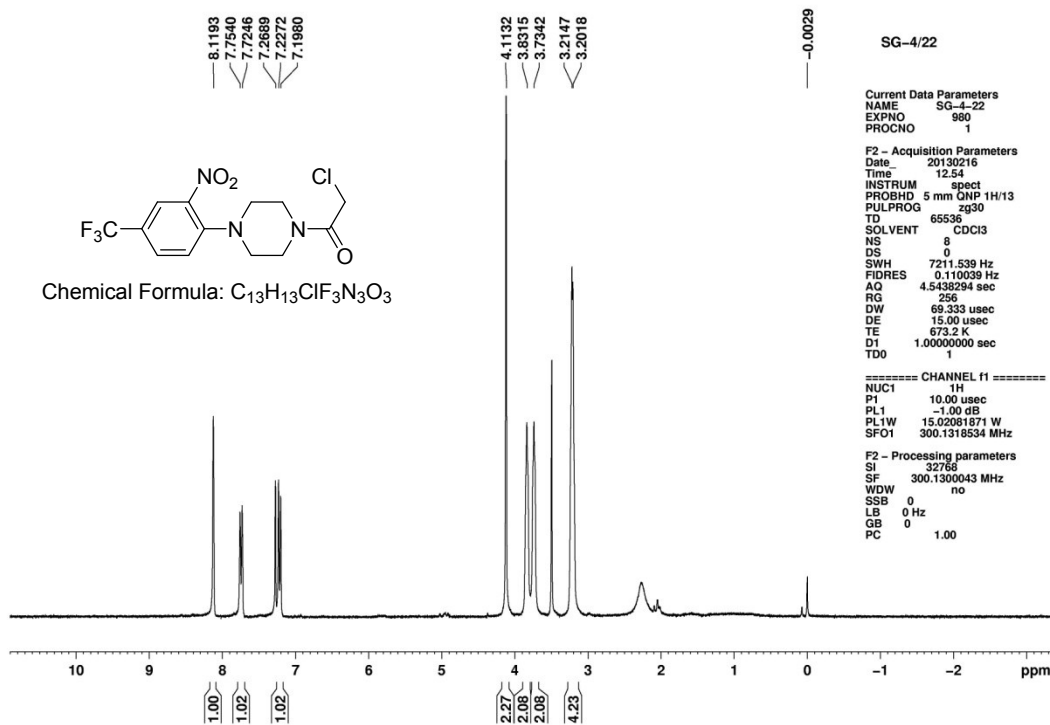
¹H NMR of compound 8d



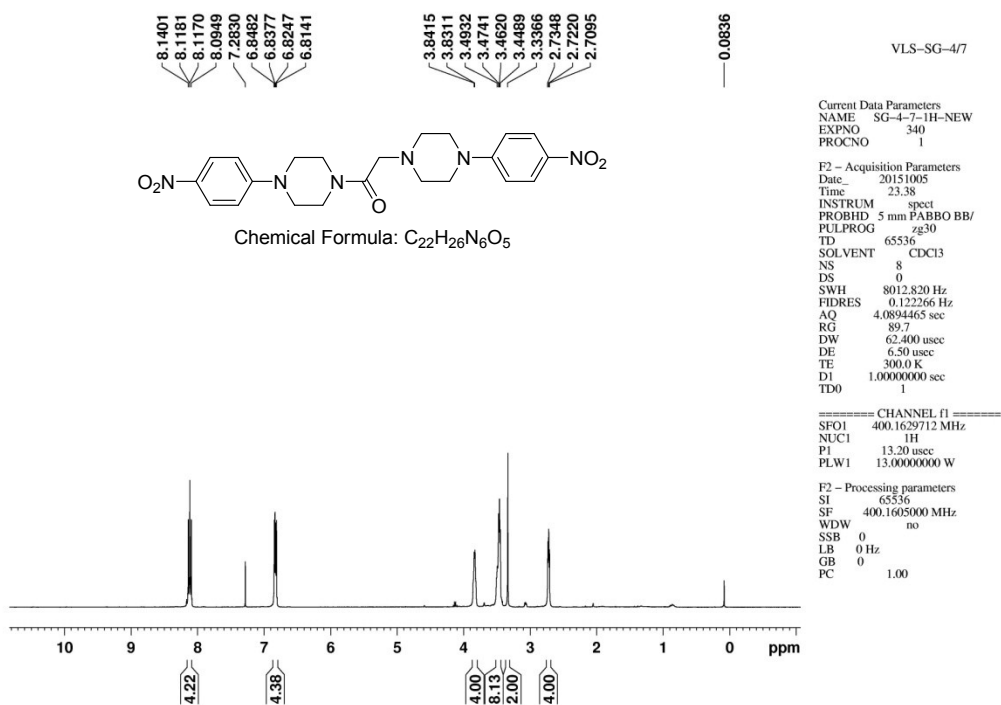
¹³C NMR of compound 8d



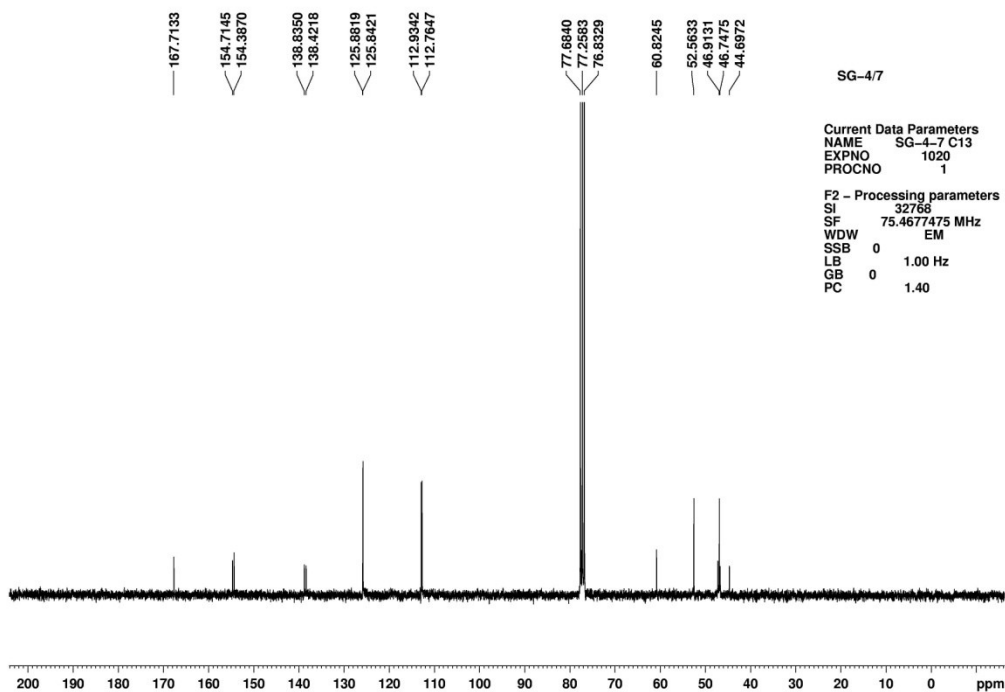
¹H NMR of compound 8e



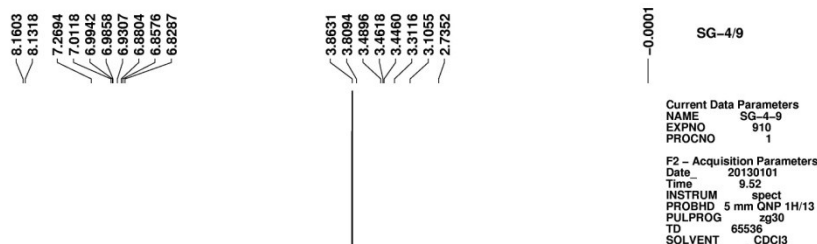
¹H NMR of compound 9a

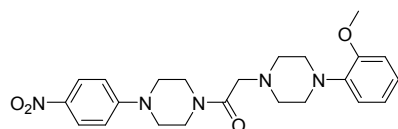


¹³C NMR of compound 9a



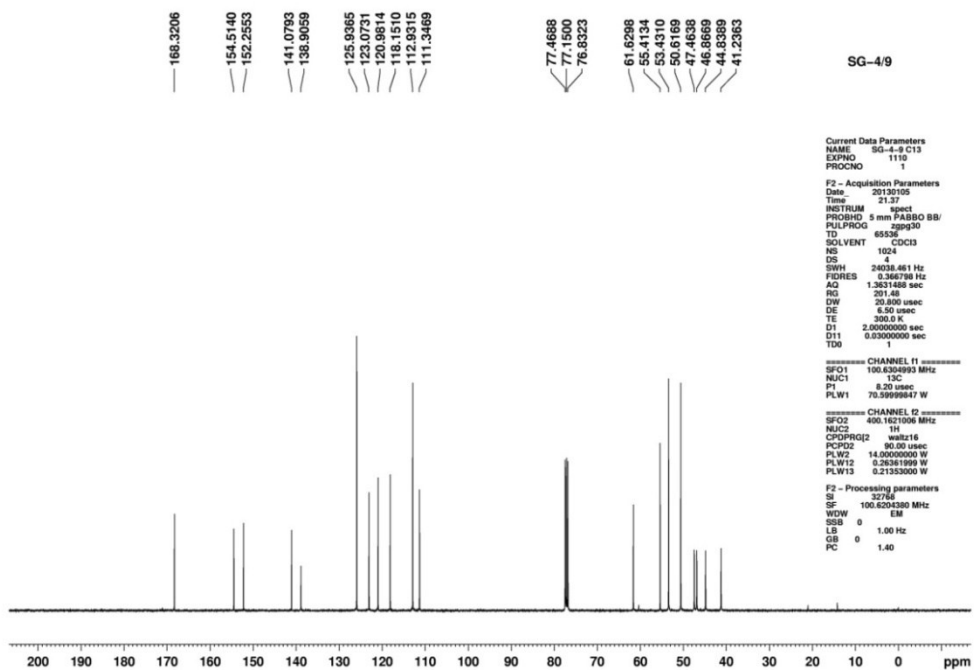
¹H NMR of compound 9b

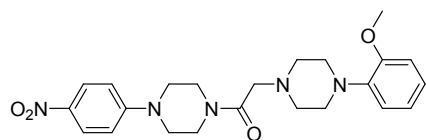




Chemical Formula: C₂₃H₂₉N₅O₄

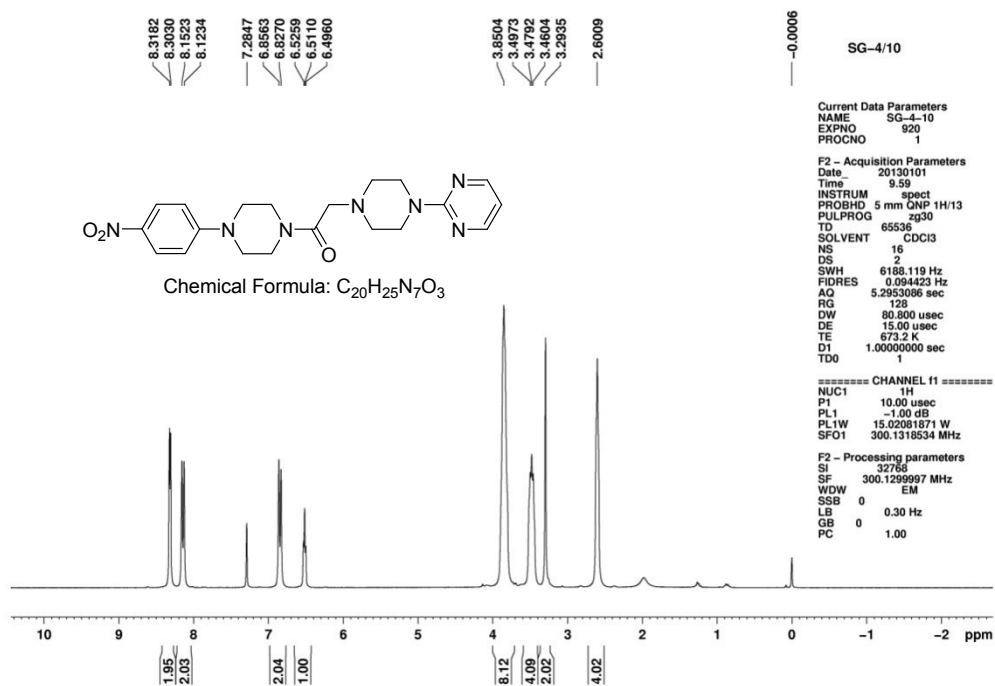
¹³C NMR of compound 9b



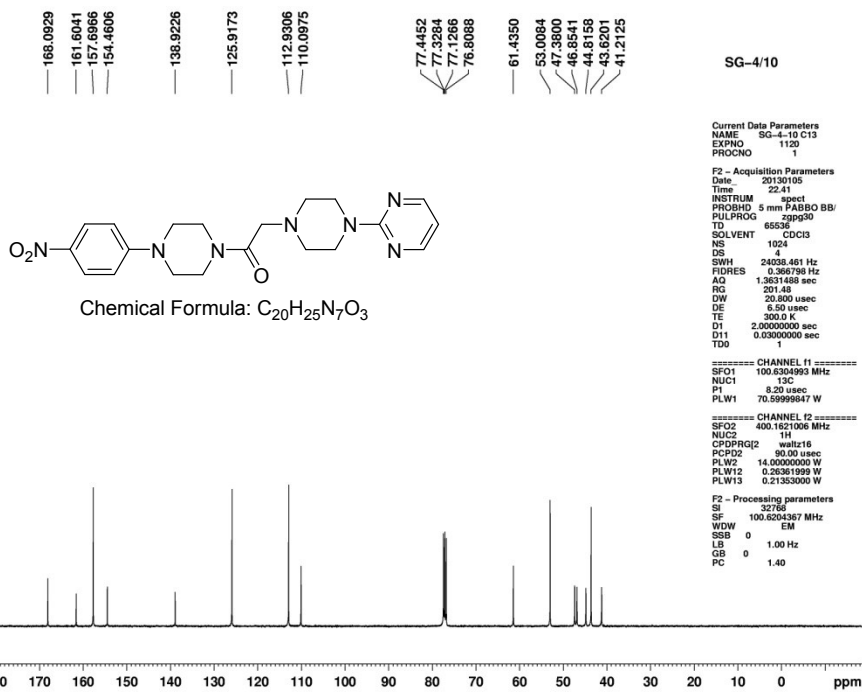


Chemical Formula: $C_{23}H_{29}N_5O_4$

1H NMR of compound 9c

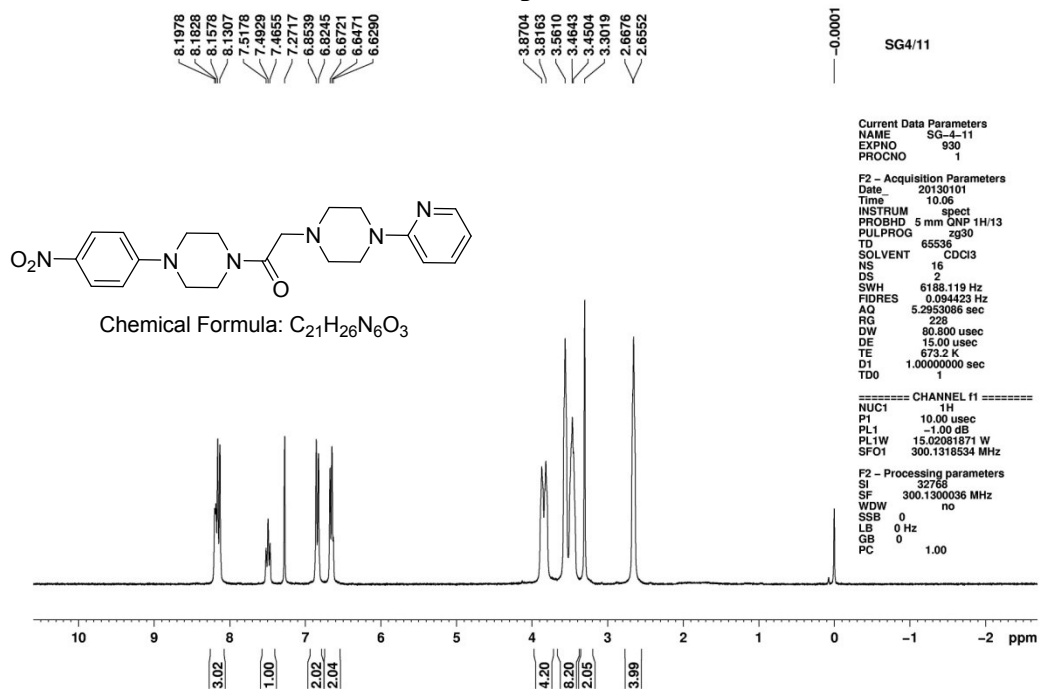


¹³C NMR of compound 9c

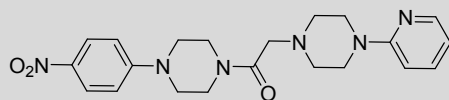


¹H

NMR of compound 9d



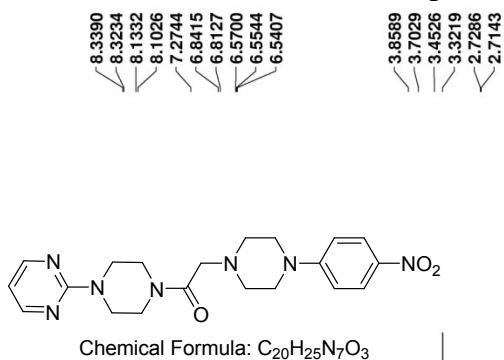
¹³C NMR of compound 9d



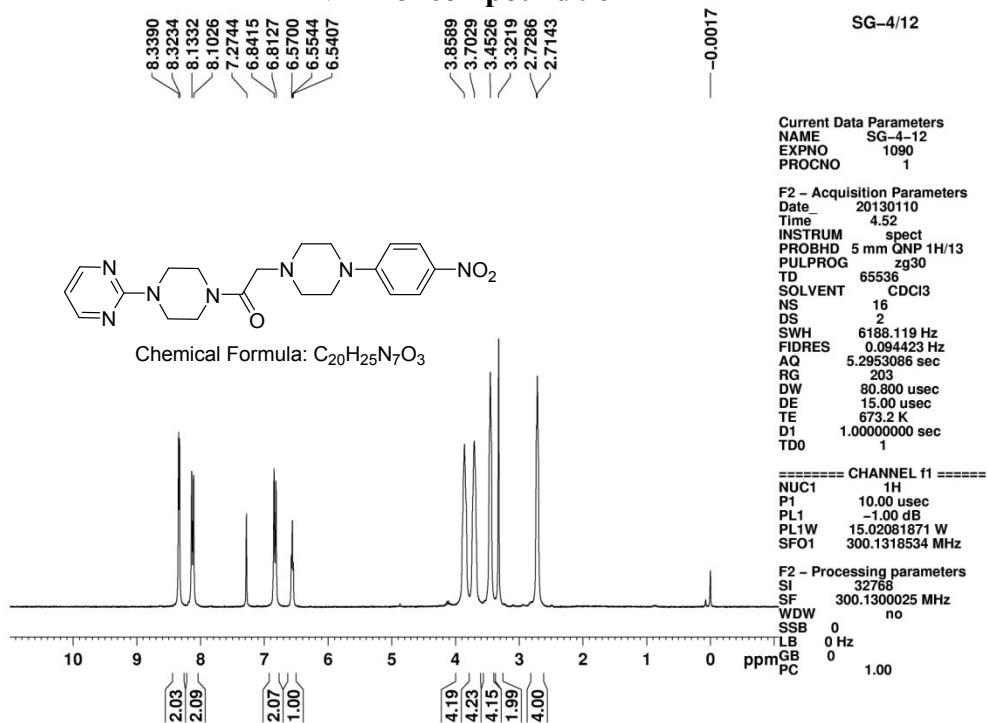
Chemical Formula: C₂₁H₂₆N₆O₃

¹H

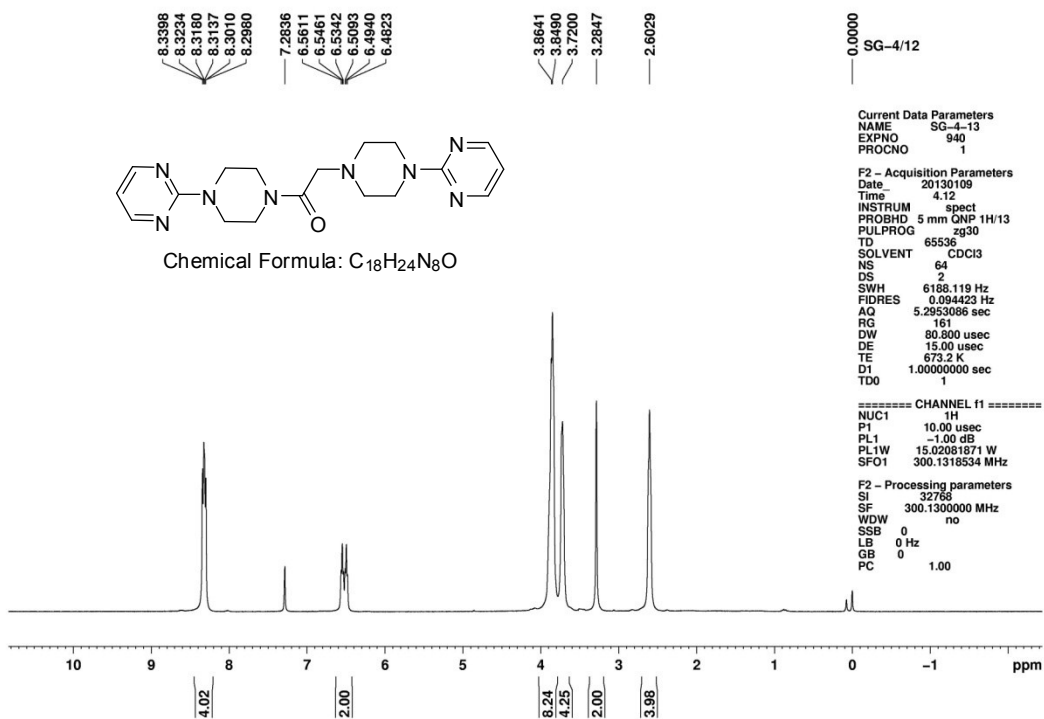
NMR of compound 9e



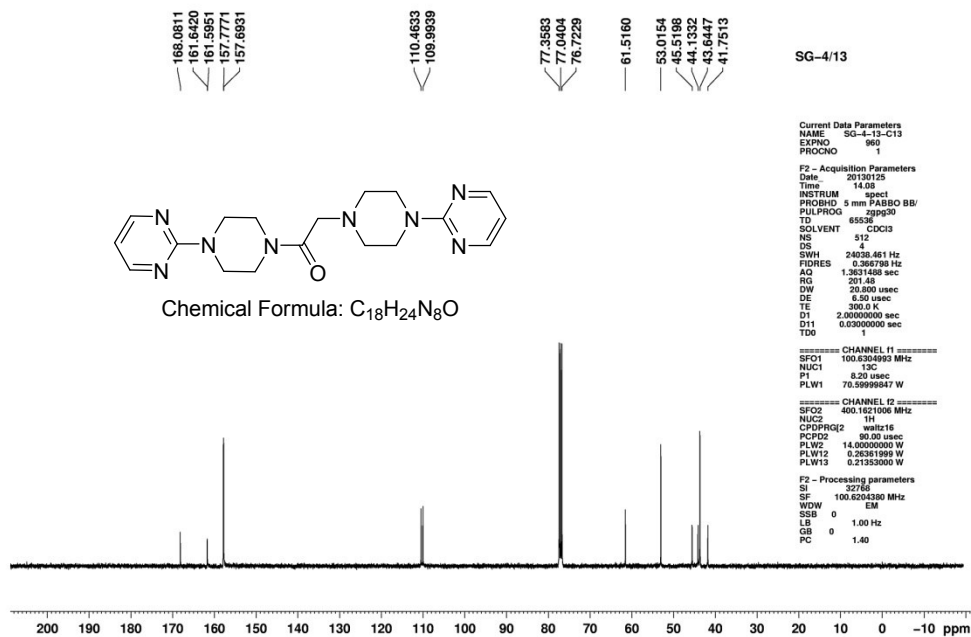
Chemical Formula: C₂₀H₂₅N₇O₃



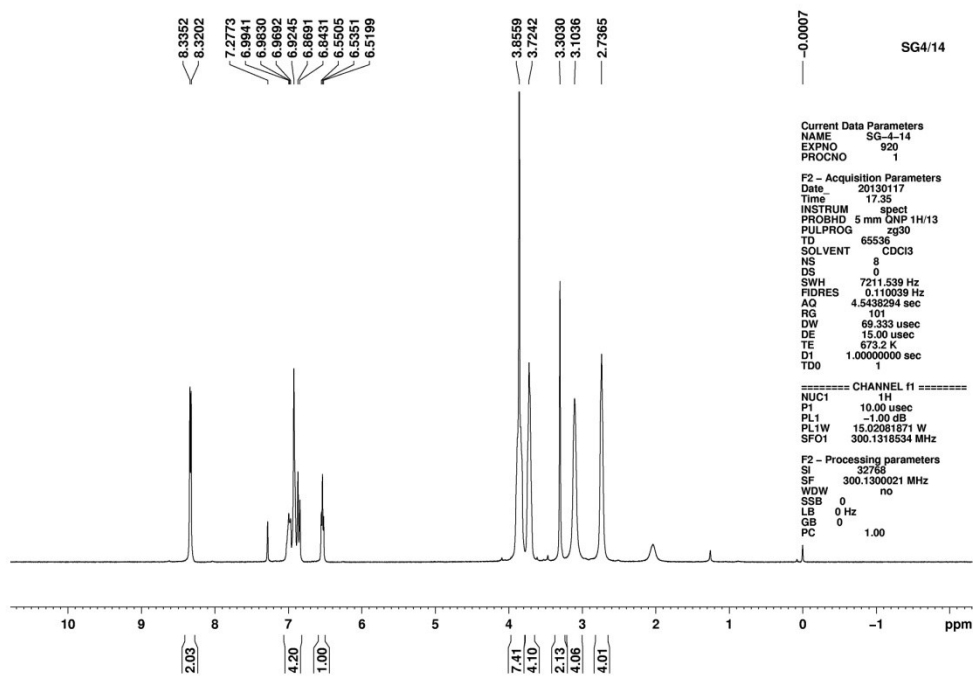
¹H NMR of compound 9f



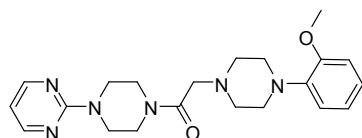
¹³C NMR of compound 9f



¹H NMR of

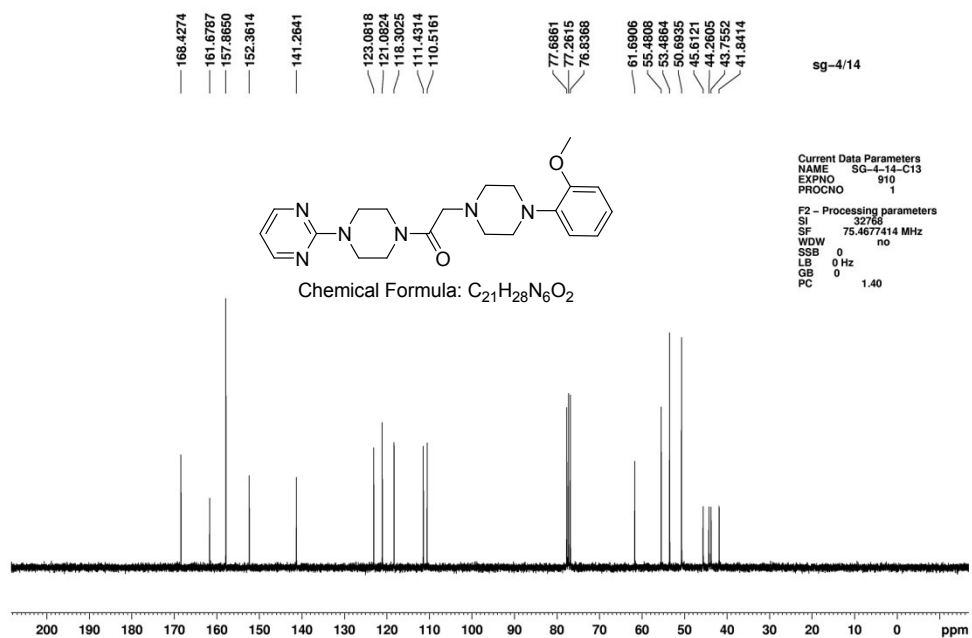


compound 9g

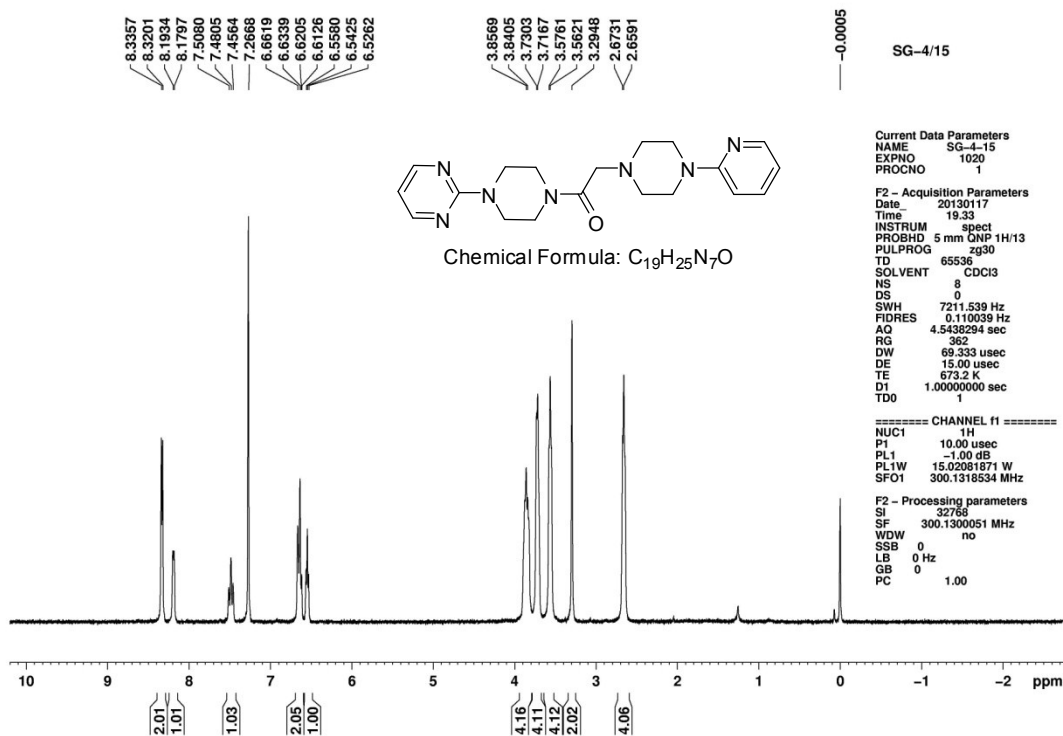


Chemical Formula: C₂₁H₂₈N₆O₂

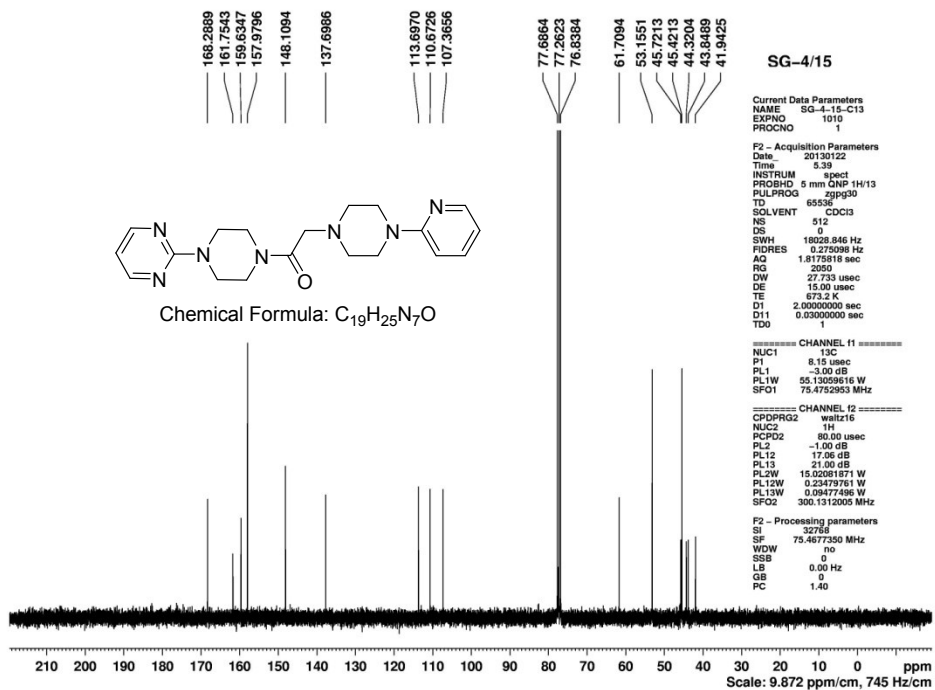
¹³C NMR of compound 9g



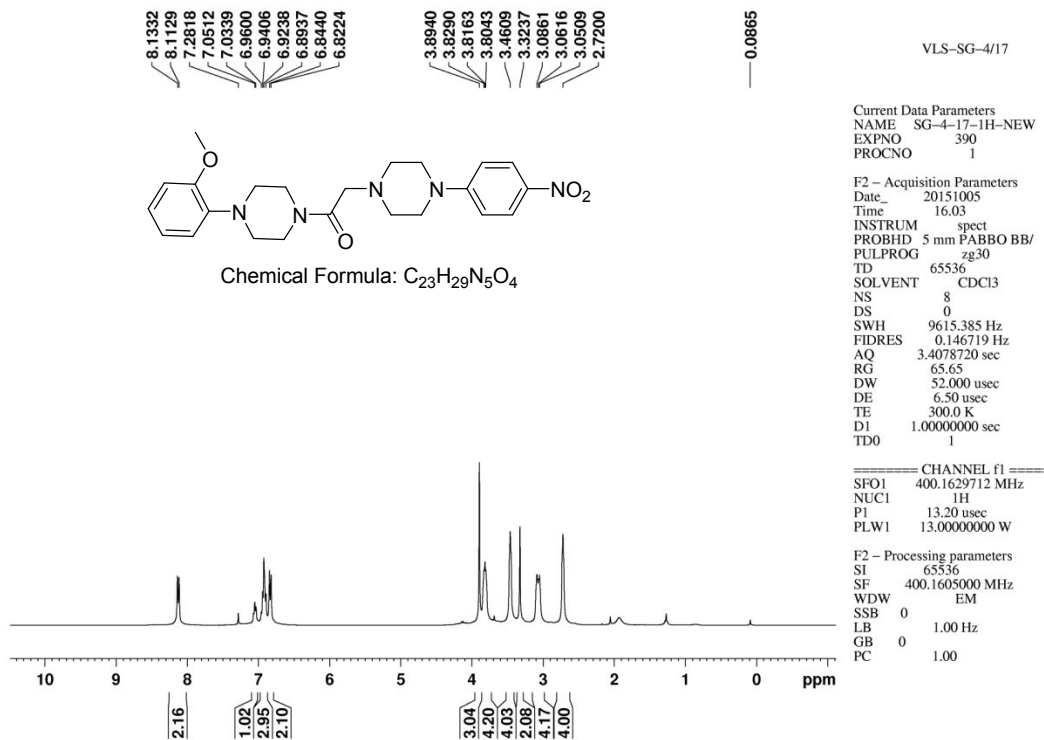
¹H NMR of compound 9h



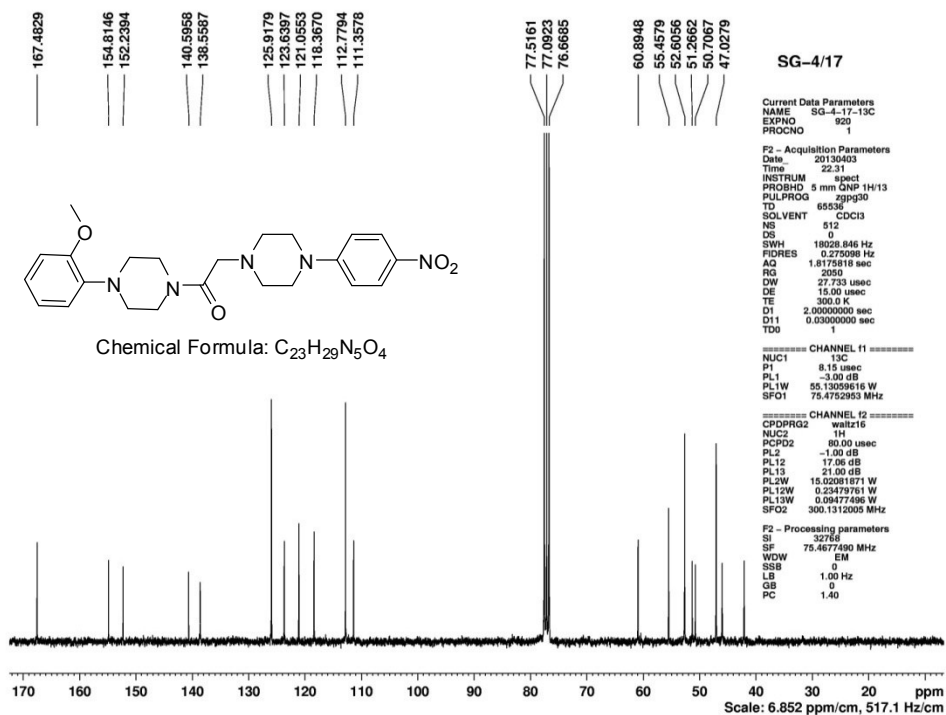
¹³C NMR of compound 9h



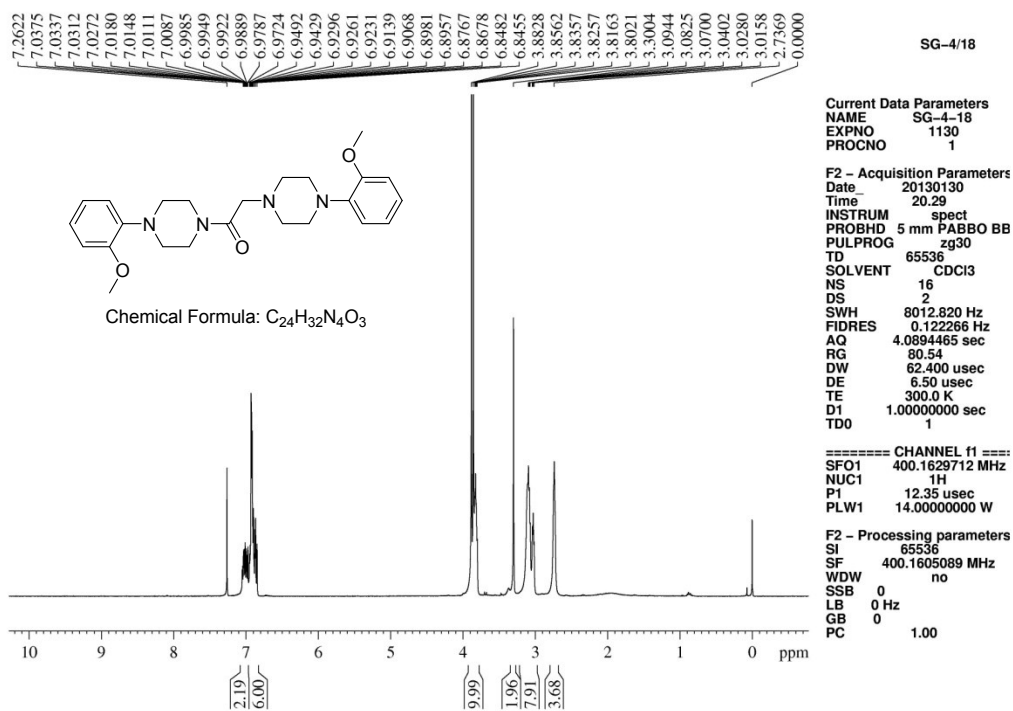
¹H NMR of compound 9i



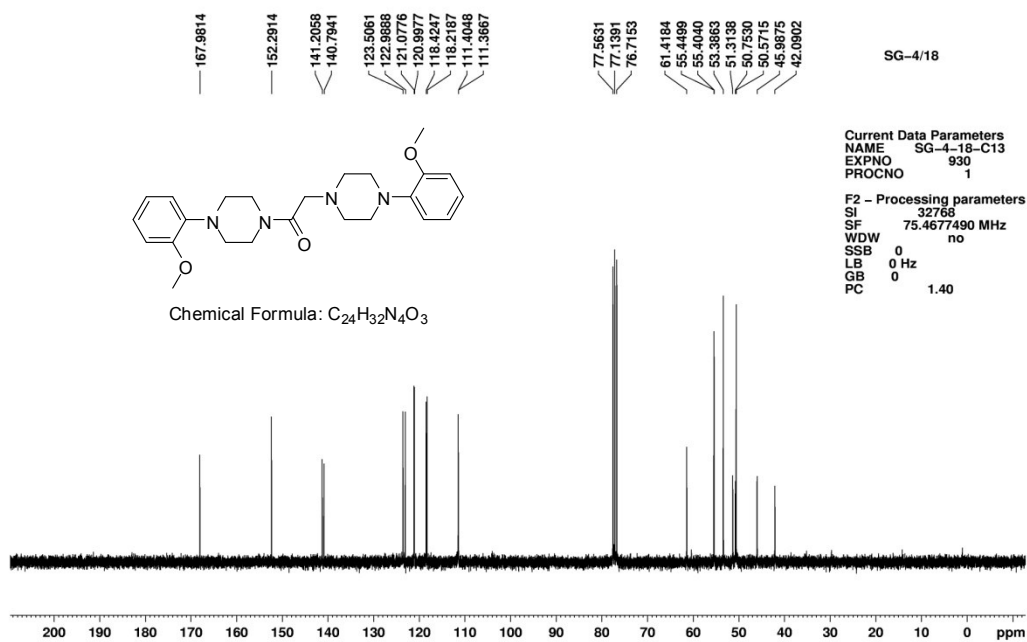
¹³C NMR of compound 9i



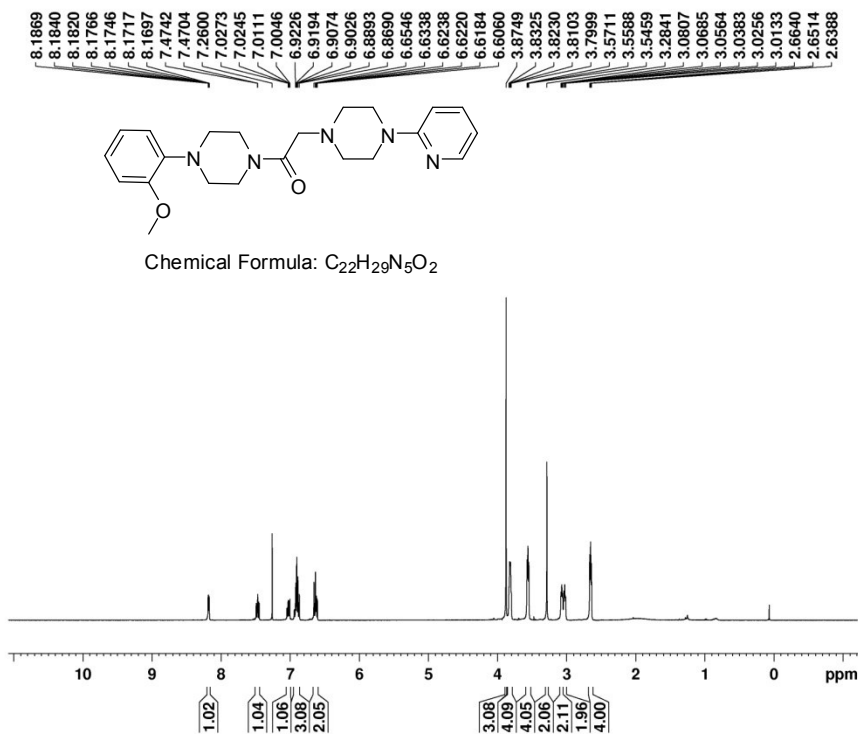
¹H NMR of compound 9j



¹³C NMR of compound 9j



¹H NMR of compound 9k



VLS-SG-4/19

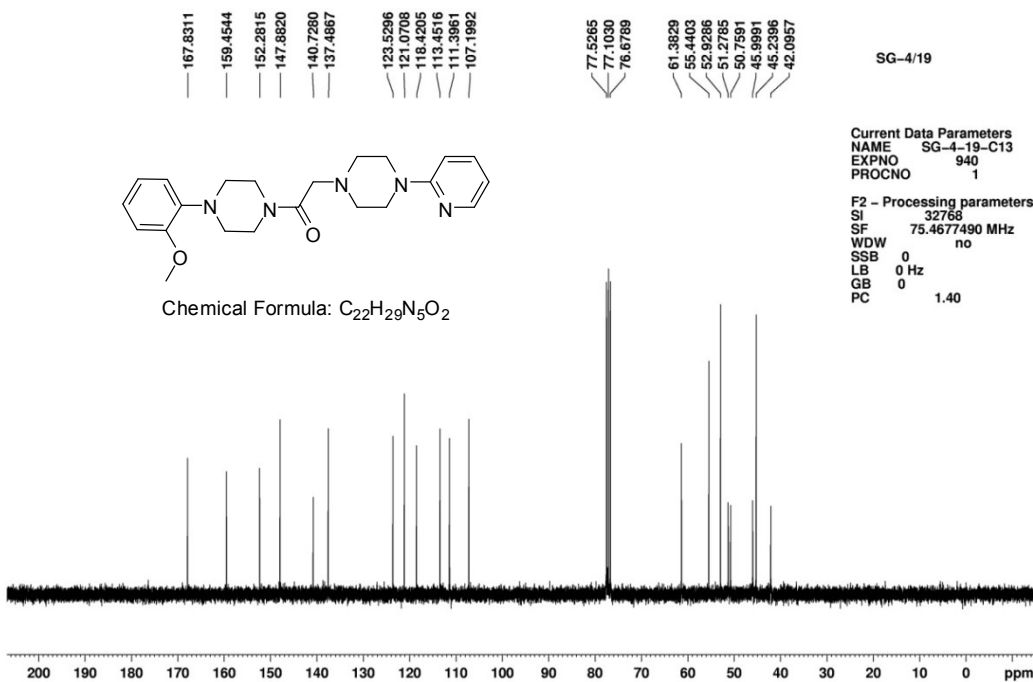
Current Data Parameters
 NAME SG-4-19-1H-NEW
 EXPNO 360
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20151005
 Time 23:46
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 8
 DS 0
 SWH 8012.820 Hz
 FIDRES 0.122266 Hz
 AQ 4.089465 sec
 RG 89.7
 DW 62.400 usec
 DE 6.50 usec
 TE 300.0 K
 D1 1.0000000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 400.1629712 MHz
 NUC1 1H
 P1 13.20 usec
 PLW1 13.0000000 W

F2 - Processing parameters
 SI 65536
 SF 400.1605093 MHz
 WDW no
 SSB 0
 LB 0 Hz
 GB 0
 PC 1.00

¹³C NMR of compound 9k

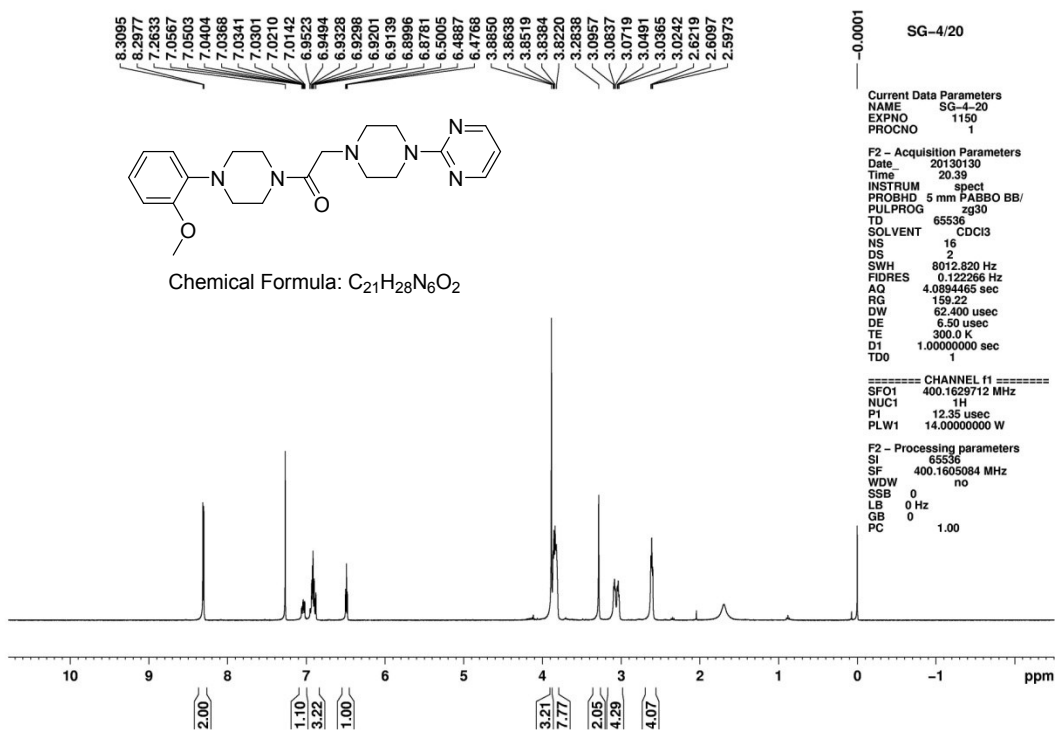


SG-4/19

Current Data Parameters
 NAME SG-4-19-C13
 EXPNO 940
 PROCNO 1

F2 - Processing parameters
 SI 32768
 SF 75.4677490 MHz
 WDW no
 SSB 0
 LB 0 Hz
 GB 0
 PC 1.40

¹H NMR of compound 9l



SG-4/20

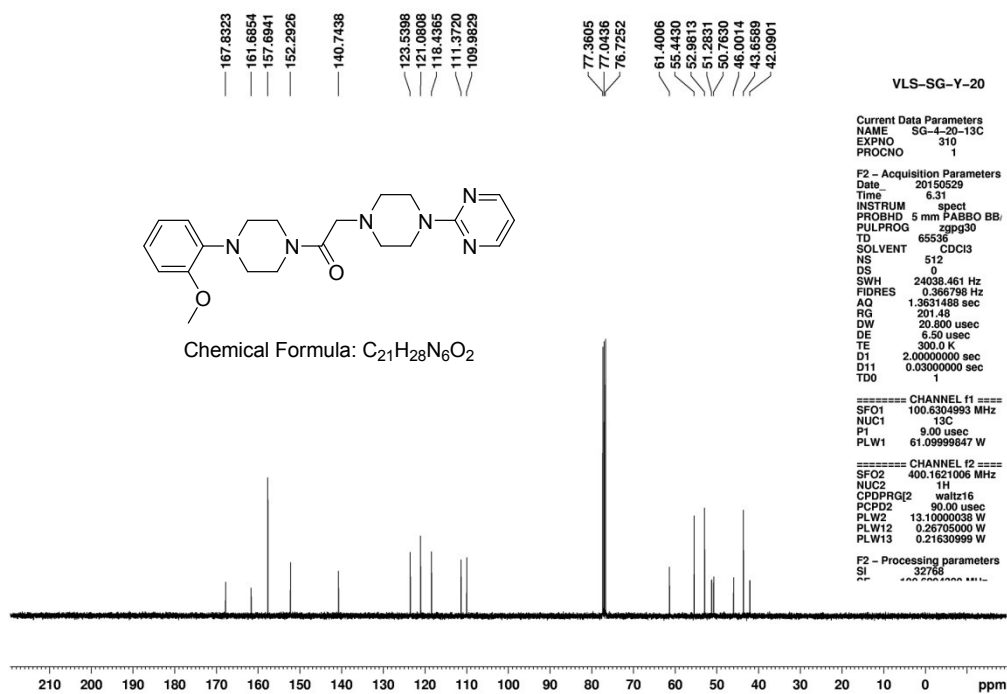
Current Data Parameters
 NAME SG-4-20
 EXPNO 1150
 PROCNO 1

F2 - Acquisition Parameters
 Date 20130130
 Time 20.39
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.122266 Hz
 AQ 4.0894465 sec
 RG 156.22
 DW 82.400 usec
 DE 6.50 usec
 TE 300.0 K
 D1 1.0000000 sec
 TDO 1

==== CHANNEL f1 =====
 SFO1 400.1629712 MHz
 NUC1 1H
 P1 12.35 usec
 PLW1 14.0000000 W

F2 - Processing parameters
 SI 65536
 SF 400.1605084 MHz
 WDW no
 SSB 0
 LB 0 Hz
 GB 0
 PC 1.00

¹³C NMR of compound 9l



VLS-SG-Y-20

Current Data Parameters
 NAME SG-4-20-13C
 EXPNO 310
 PROCNO 1

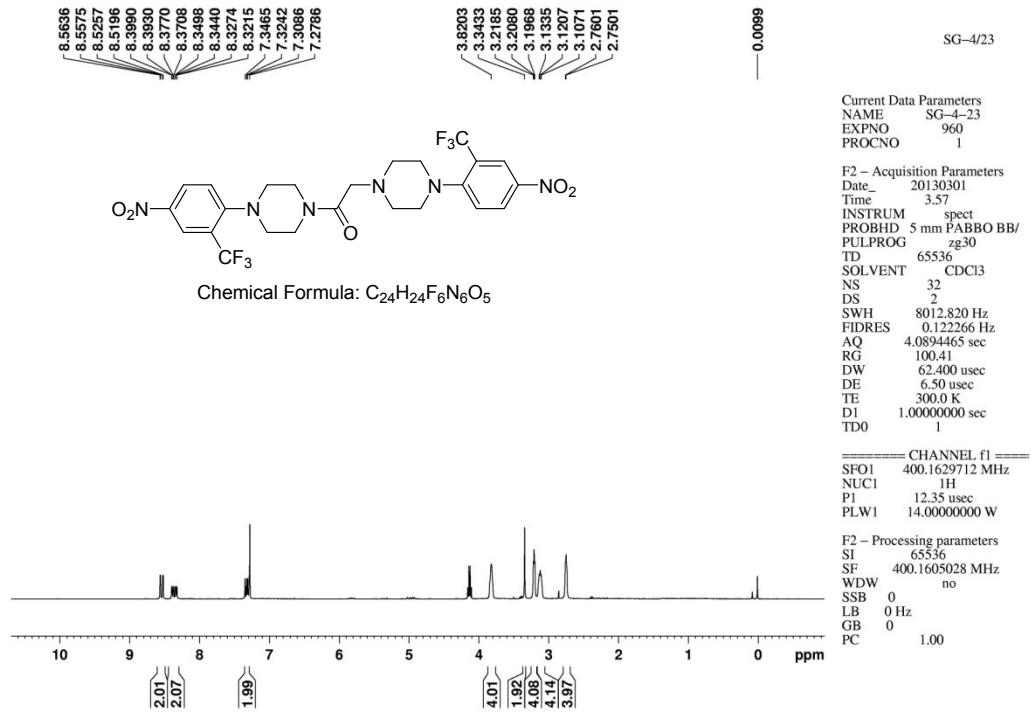
F2 - Acquisition Parameters
 Date 20150529
 Time 6.31
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 512
 DS 0
 SWH 24038.461 Hz
 FIDRES 0.366798 Hz
 AQ 1.3631488 sec
 RG 201.48
 DW 20.800 usec
 DE 6.50 usec
 TE 300.0 K
 D1 2.0000000 sec
 D11 0.0300000 sec
 TDO 1

==== CHANNEL f1 =====
 SFO1 100.6304993 MHz
 NUC1 13C
 P1 9.00 usec
 PLW1 61.09999847 W

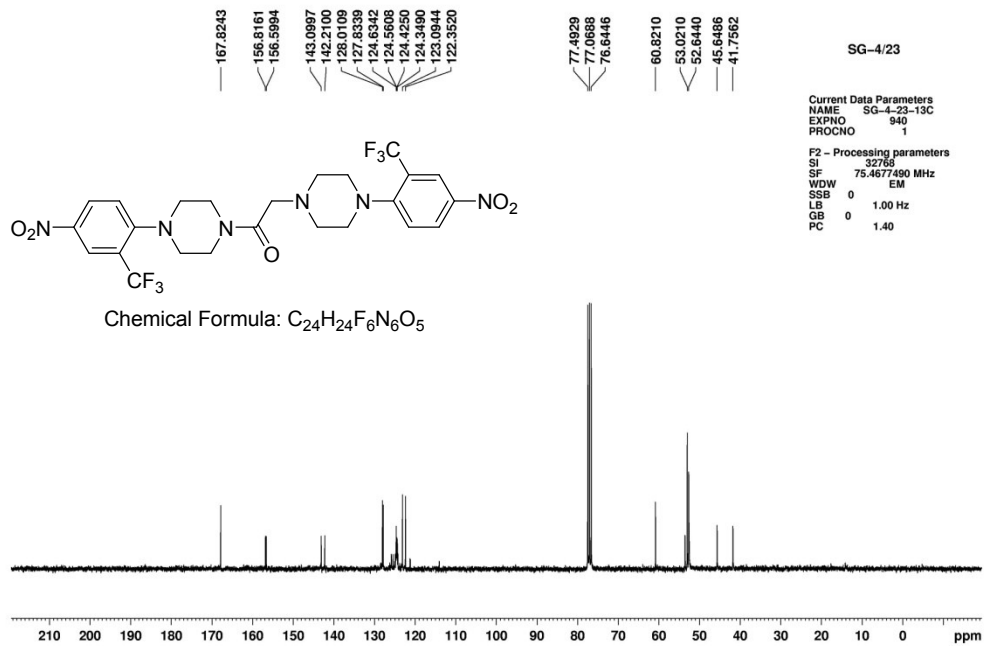
==== CHANNEL f2 =====
 SFO2 400.1621006 MHz
 NUC2 1H
 CPDPRG2 waltz16
 PCPD2 90.00 usec
 PLW2 13.10000038 W
 PLW12 0.26705000 W
 PLW13 0.21630999 W

F2 - Processing parameters
 SI 32768

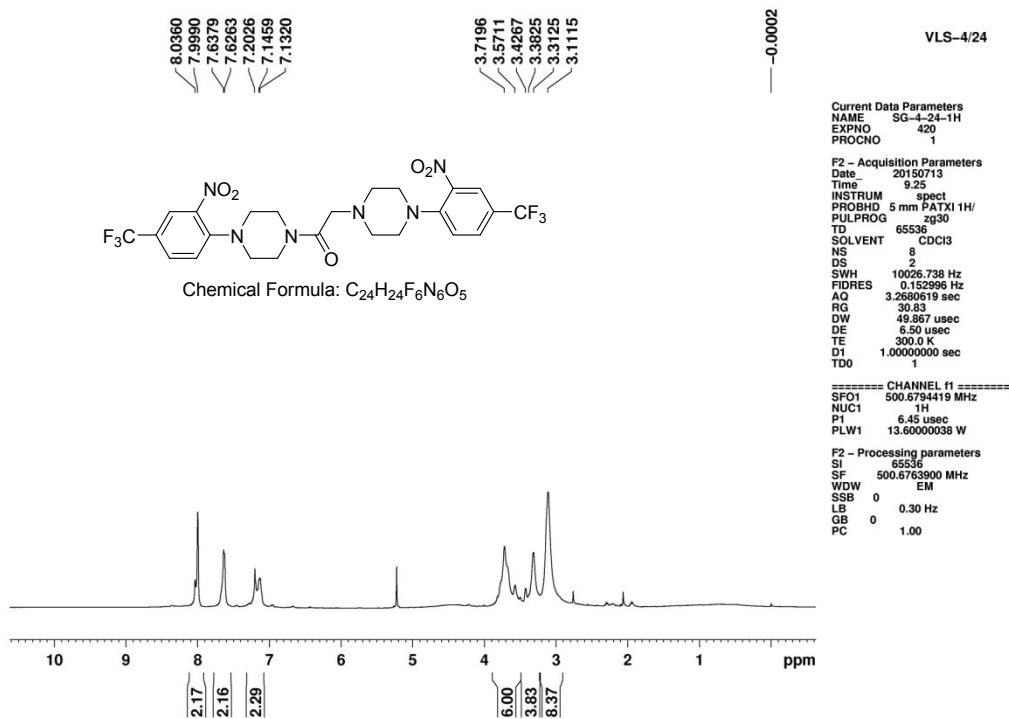
¹H NMR of compound 9m



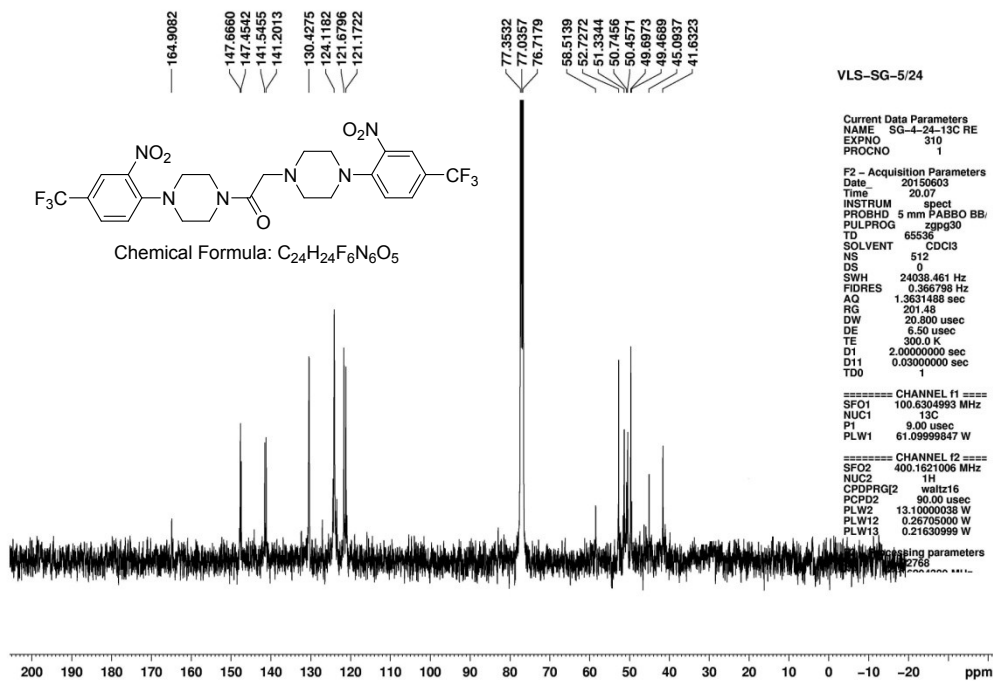
¹³C NMR of compound 9m



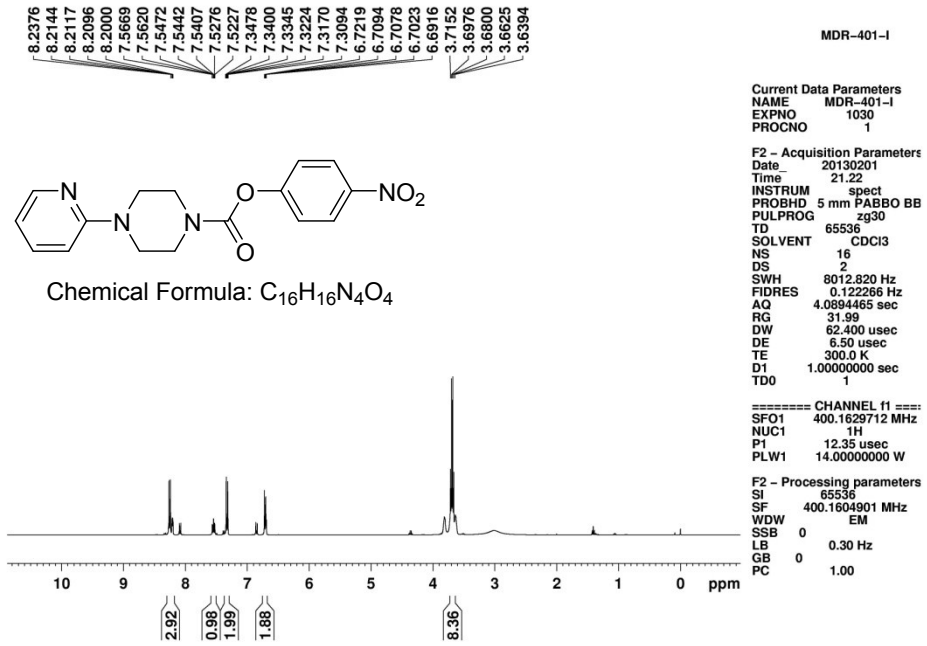
¹H NMR of compound 9n



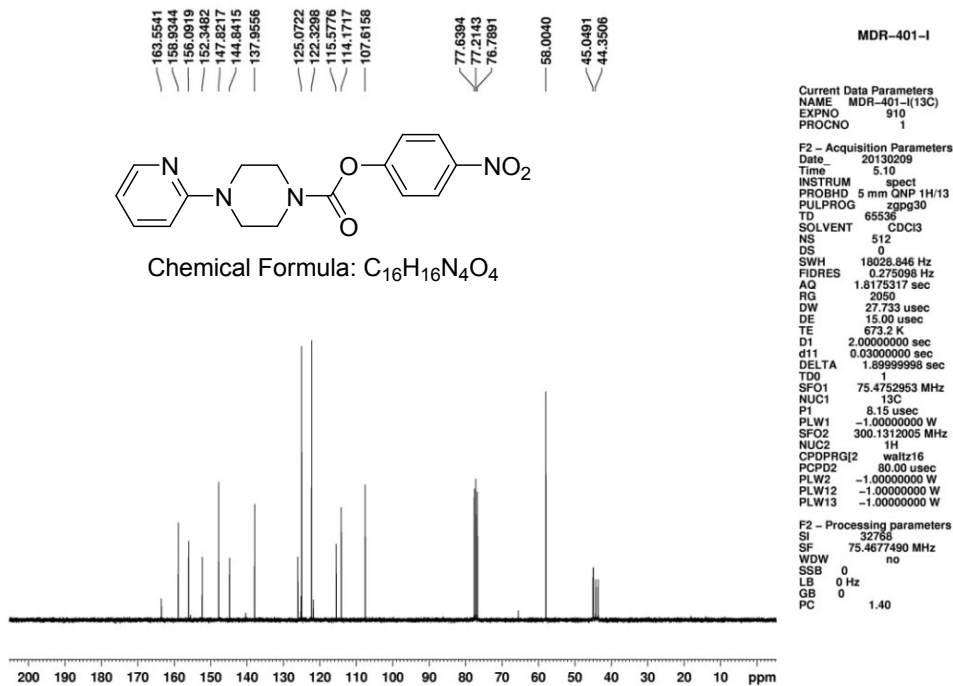
¹³C NMR of compound 9n



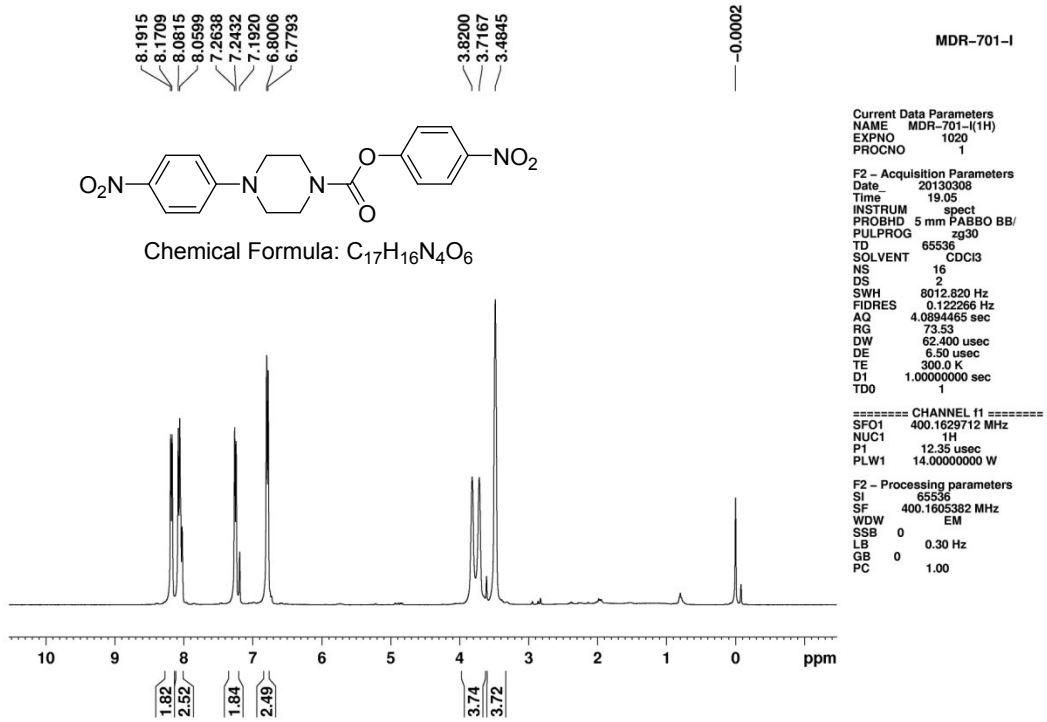
¹H NMR of compound 11a



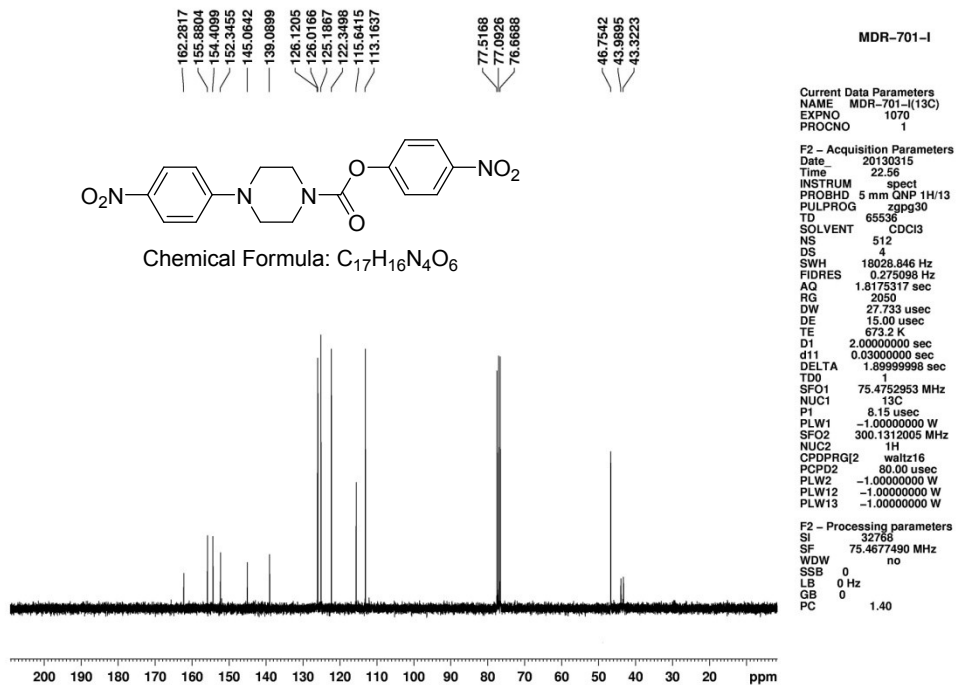
¹³C NMR of compound 11a



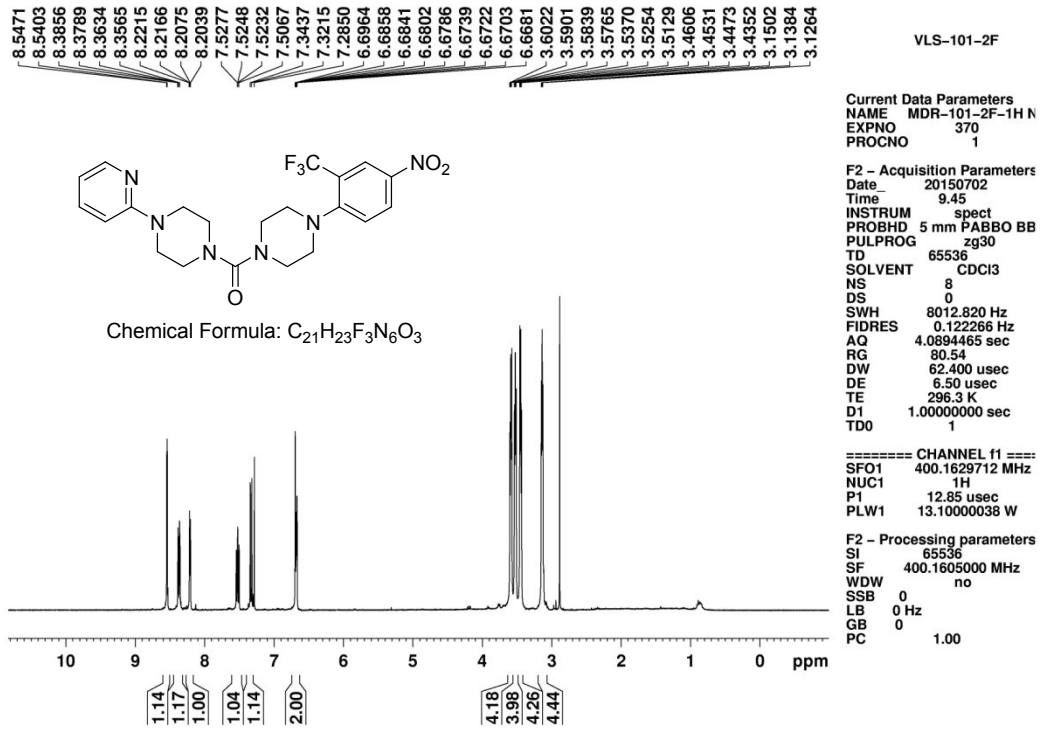
¹H NMR of compound 11b



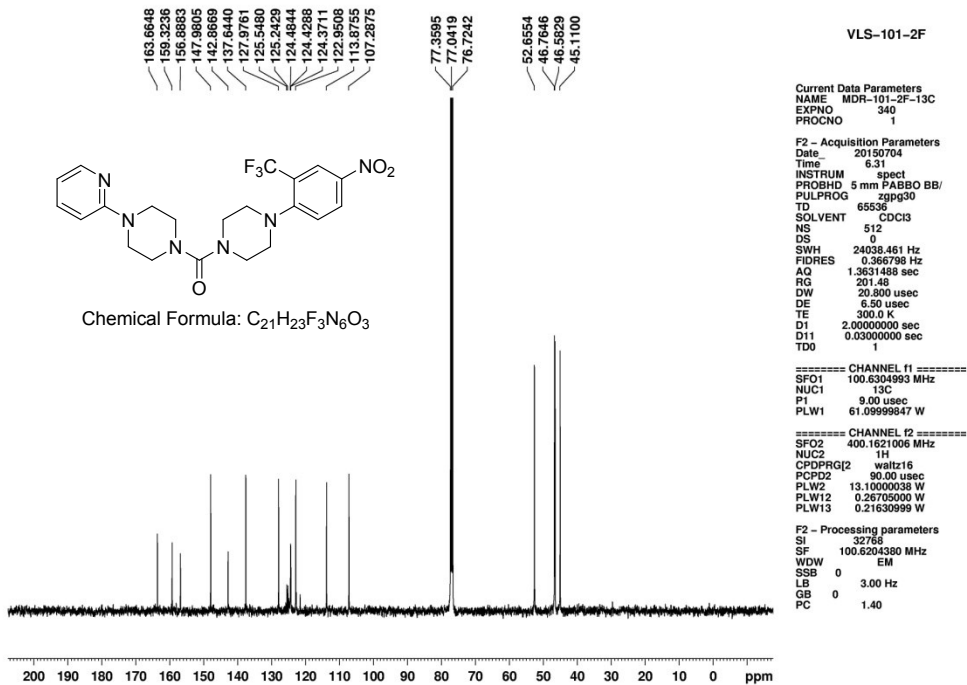
¹³C NMR of compound 11b



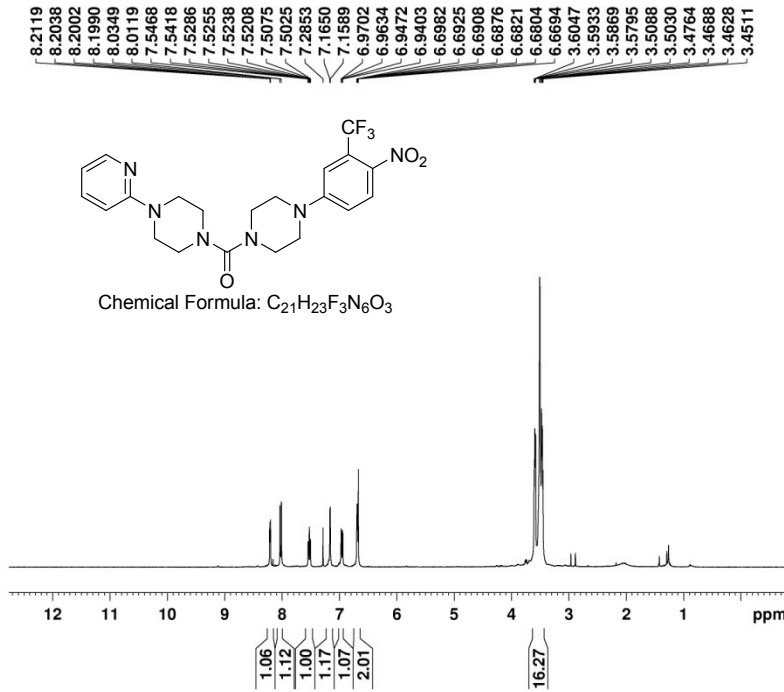
¹H NMR of compound 12a



¹³C NMR of compound 12a



¹H NMR of compound 12b



VLS-401-2F

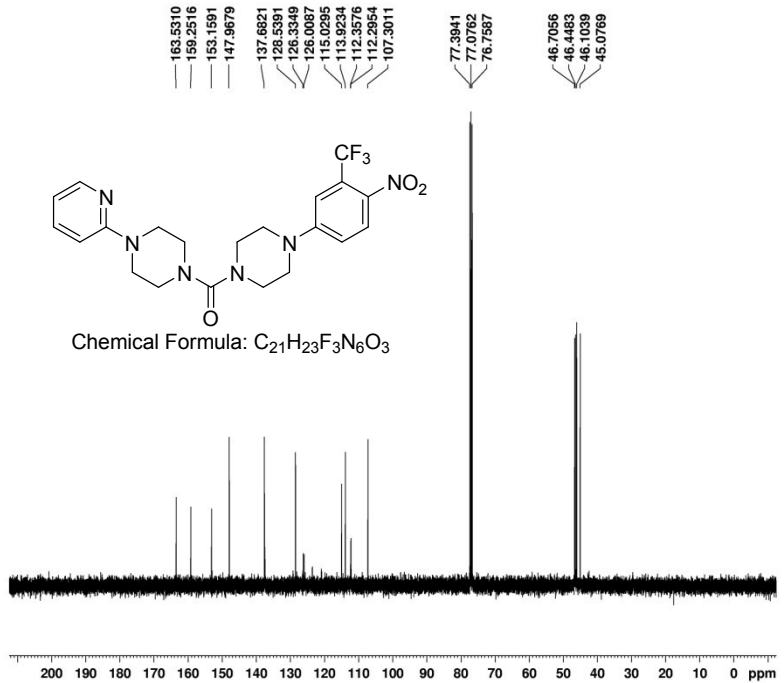
Current Data Parameters
 NAME MDR-401-2F-1H-I
 EXPNO 380
 PROCNO 1

F2 - Acquisition Parameters
 Date 20150702
 Time 9.50
 INSTRUM spect
 PROBHD 5 mm PABBO BB
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 8
 DS 0
 SWH 8012.820 Hz
 FIDRES 0.122266 Hz
 AQ 4.0894465 sec
 RG 65.65
 DW 62.400 usec
 DE 6.50 usec
 TE 296.3 K
 D1 1.00000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 400.1629712 MHz
 NUC1 1H
 P1 12.85 usec
 PLW1 13.1000038 W

F2 - Processing parameters
 SI 65536
 SF 400.1605000 MHz
 WDW no
 SSB 0
 LB 0 Hz
 GB 0
 PC 1.00

¹³C NMR of compound 12b



VLS-2F

Current Data Parameters
 NAME MDR-401-2F-13C
 EXPNO 310
 PROCNO 1

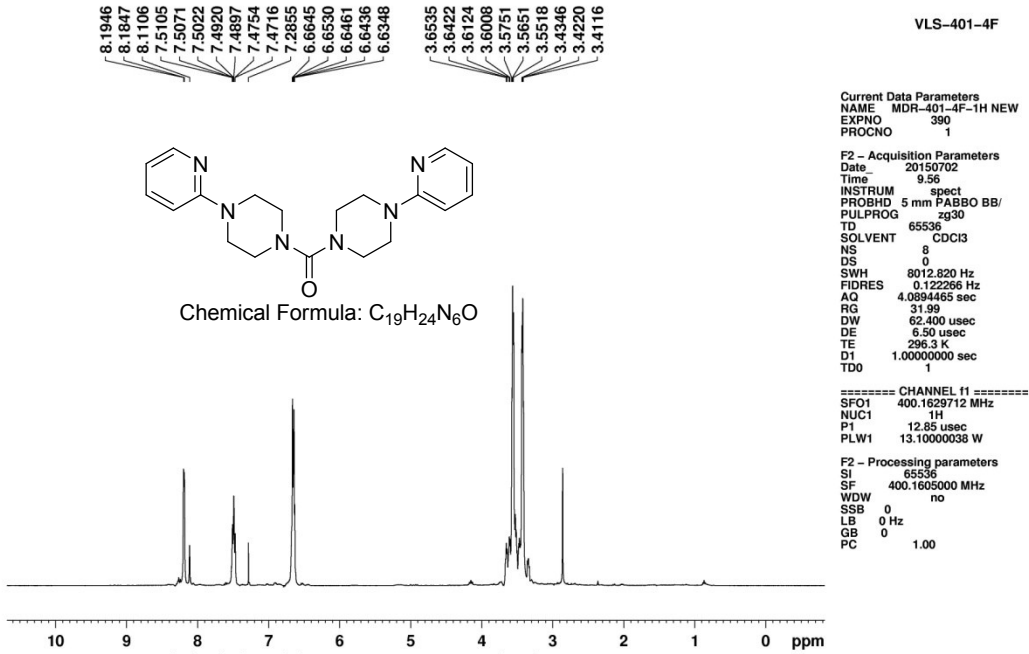
F2 - Acquisition Parameters
 Date 20150707
 Time 19.18
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 256
 DS 0
 SWH 24038.461 Hz
 FIDRES 0.366798 Hz
 AQ 1.3631498 sec
 RG 201.48
 DW 20.800 usec
 DE 6.50 usec
 TE 300.0 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TDO

===== CHANNEL f1 =====
 SFO1 100.6304993 MHz
 NUC1 13C
 P1 9.00 usec
 PLW1 61.09999847 W

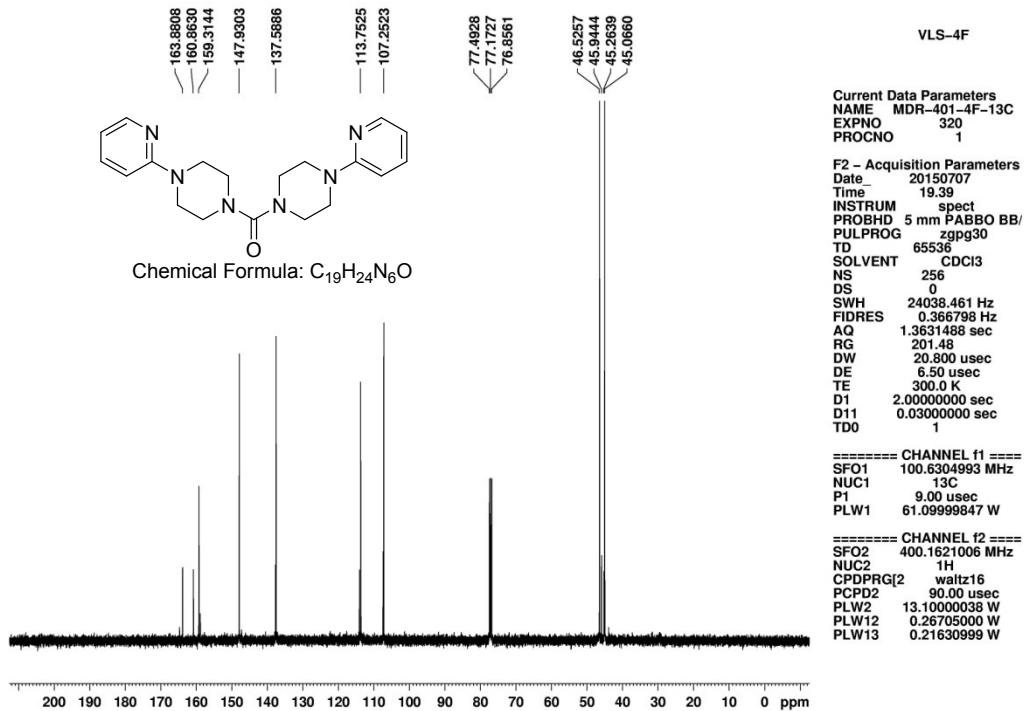
===== CHANNEL f2 =====
 SFO2 400.1621006 MHz
 NUC2 1H
 CPDPRG2 waltz16
 PCPDZ 90.00 usec
 PLW2 13.1000038 W
 PLW12 0.28705000 W
 PLW13 0.21630998 W

F2 - Processing parameters
 SI 32768
 SF 100.6204380 MHz
 WDW no
 SSB 0
 LB 0 Hz
 GB 0
 PC 1.40

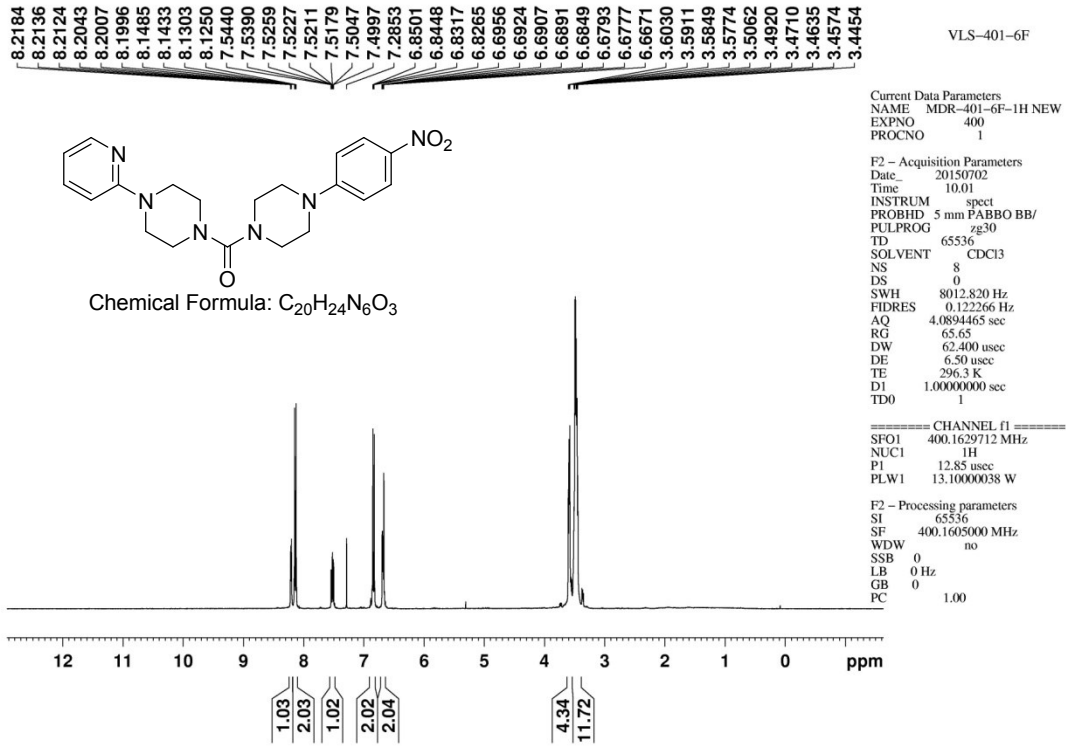
¹H NMR of compound 12c



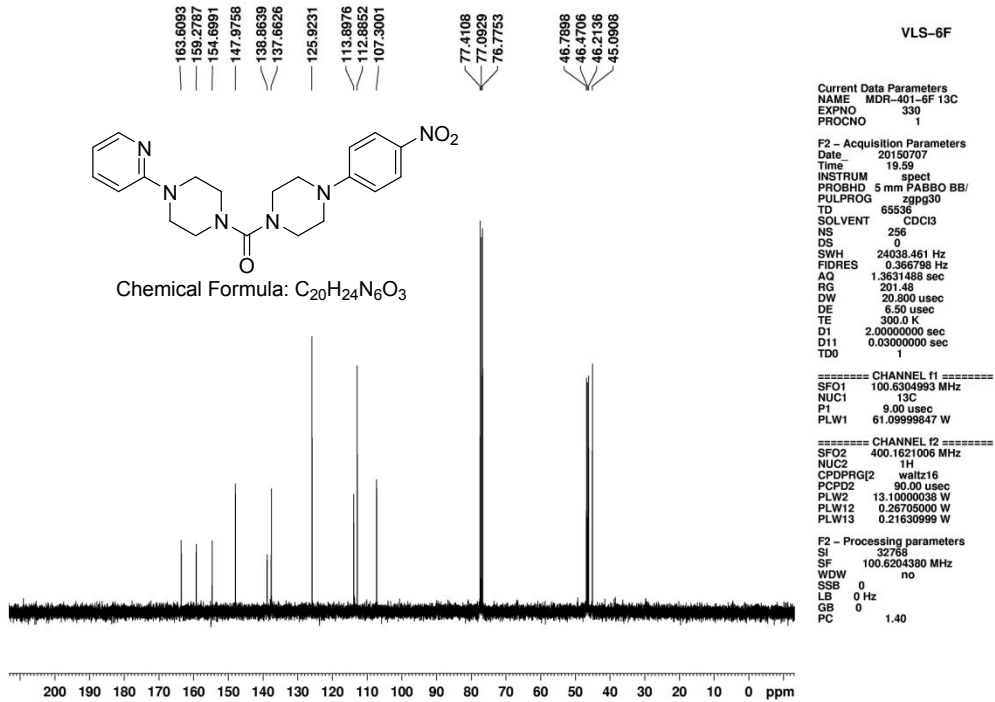
¹³C NMR of compound 12c



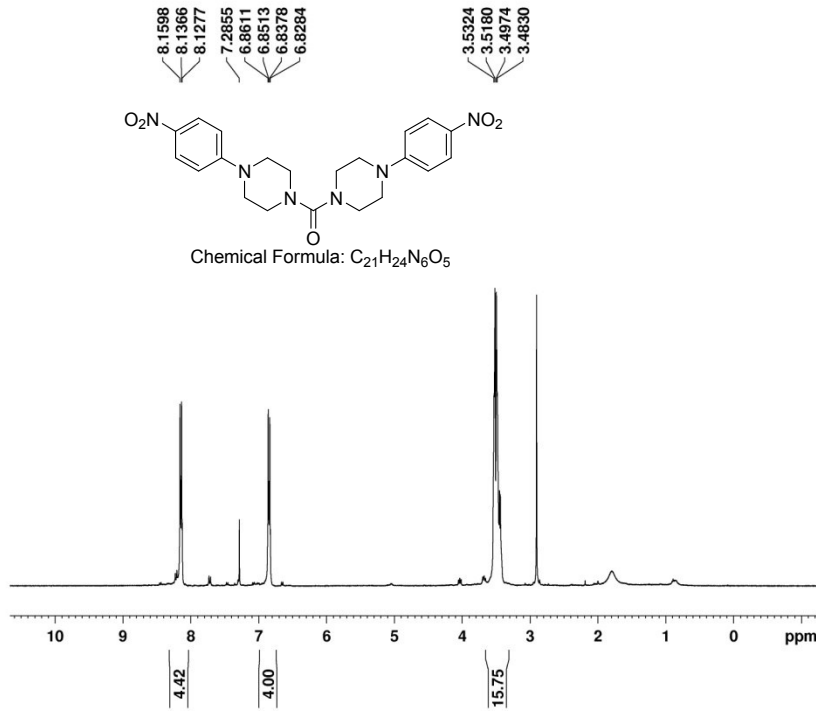
¹H NMR of compound 12d



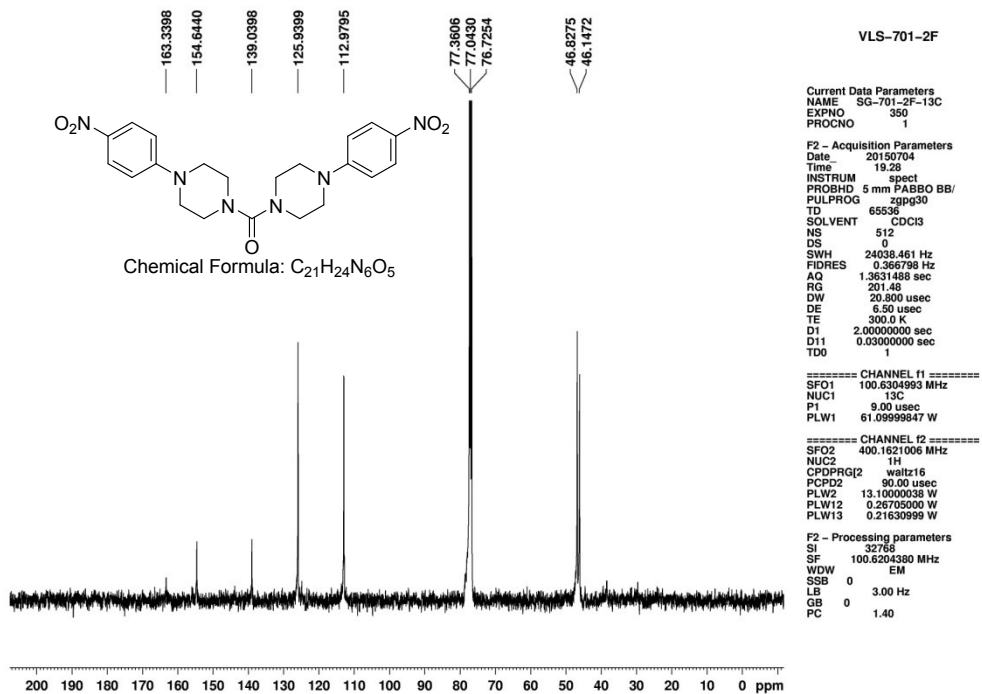
¹³C NMR of compound 12d



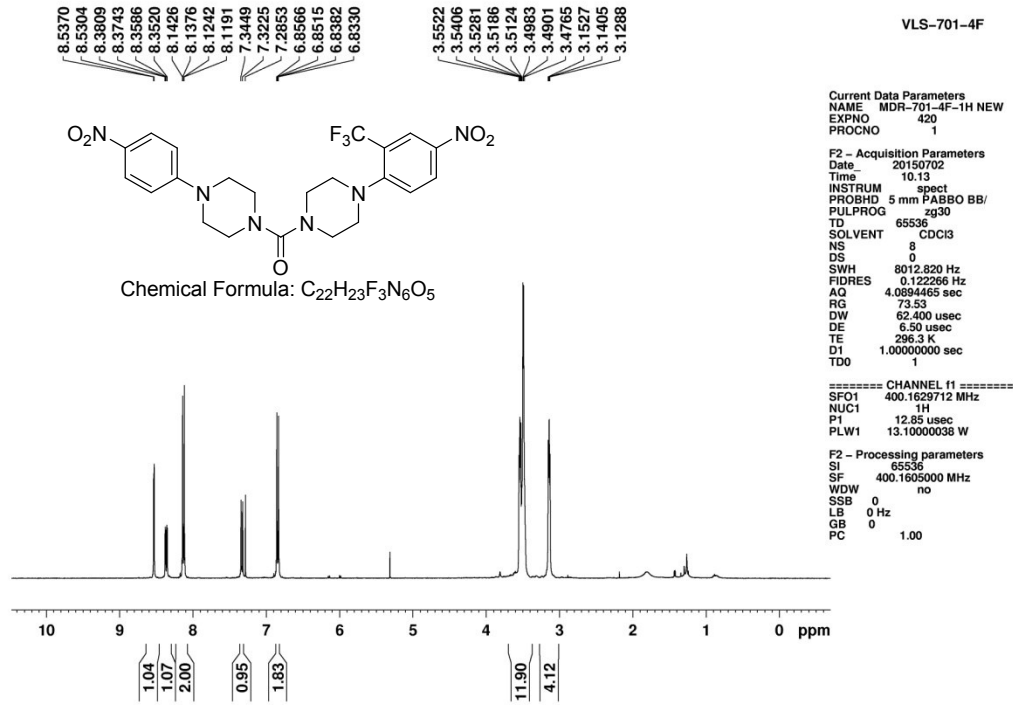
¹H NMR of compound 12e



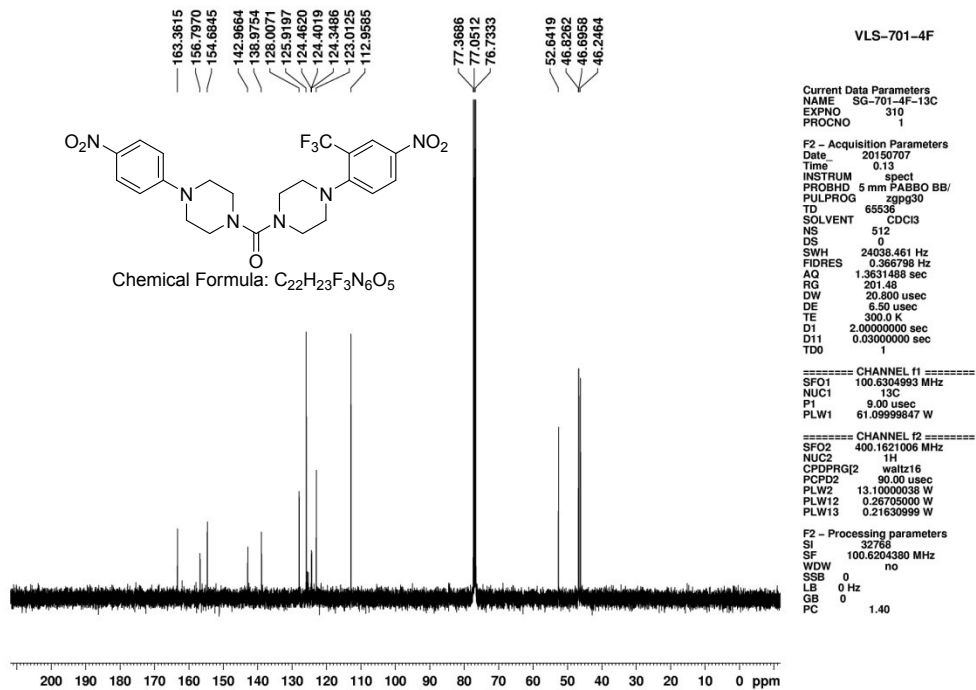
¹³C NMR of compound 12e



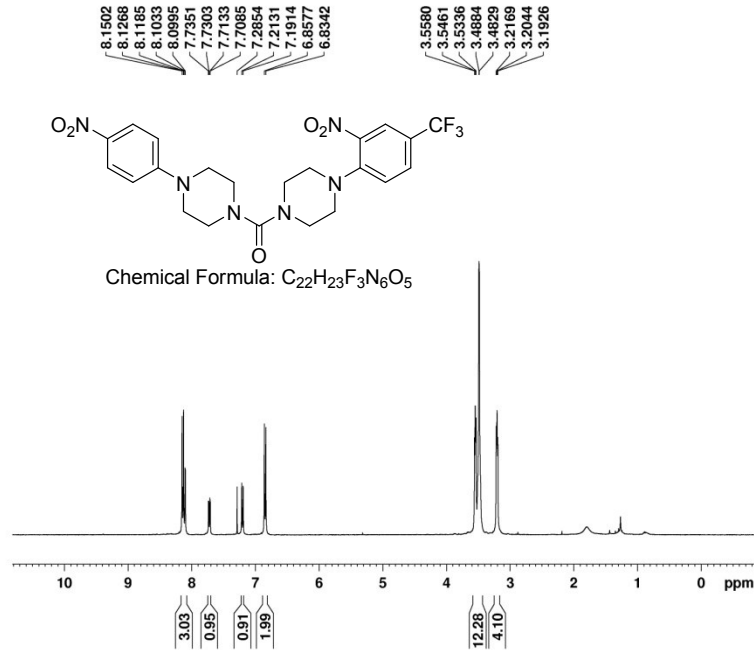
¹H NMR of compound 12f



¹³C NMR of compound 12f



¹H NMR of compound 12g



VLS-701-5F

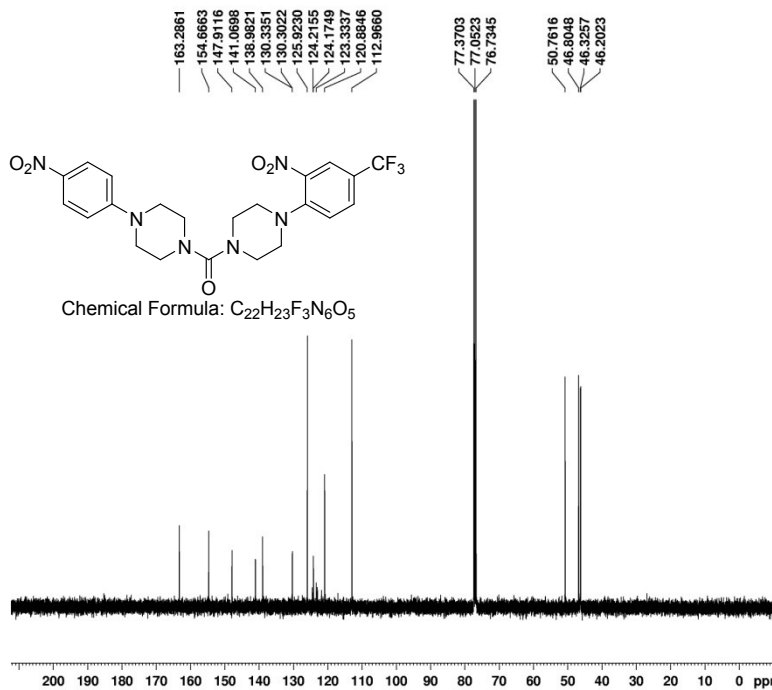
Current Data Parameters
 NAME MDR-701-5F-1H NEW
 EXPNO 430
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20150702
 Time 10.18
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 8
 DS 0
 SWH 8012.820 Hz
 FIDRES 0.122266 Hz
 AQ 4.0894465 sec
 RG 80.54
 DW 62.400 usec
 DE 6.50 usec
 TE 296.3 K
 D1 1.0000000 sec
 TDO 1

==== CHANNEL f1 =====
 SFO1 400.1629712 MHz
 NUC1 1H
 P1 12.85 usec
 PLW1 13.10000038 W

F2 - Processing parameters
 SI 65536
 SF 400.1605000 MHz
 WDW no
 SSB 0
 LB 0 Hz
 GB 0
 PC 1.00

¹³C NMR of compound 12g



VLS-701-5F

Current Data Parameters
 NAME SG-701-5F-13C
 EXPNO 320
 PROCNO 1

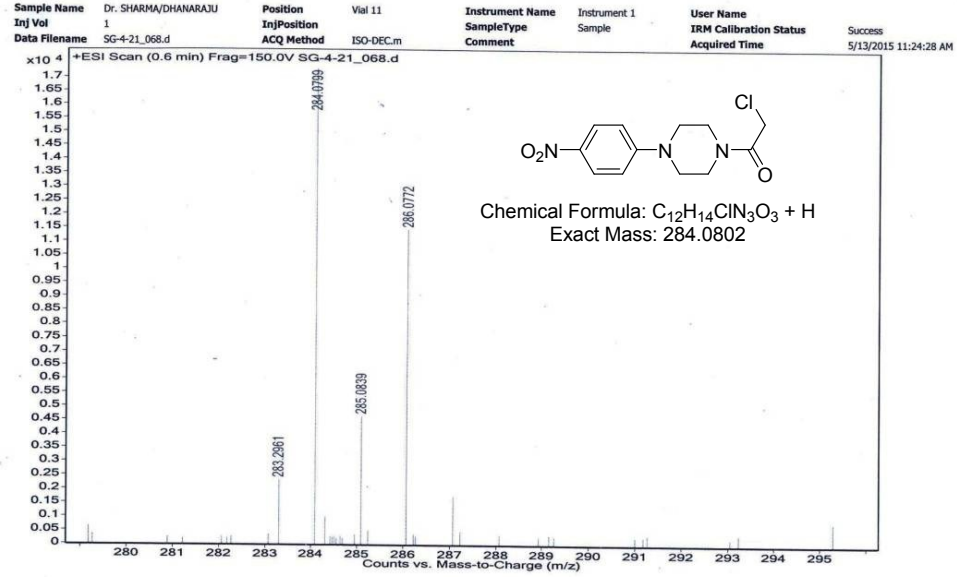
F2 - Acquisition Parameters
 Date_ 20150707
 Time 0.47
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 512
 DS 0
 SWH 24038.461 Hz
 FIDRES 0.366798 Hz
 AQ 1.3631488 sec
 RG 201.48
 DW 20.800 usec
 DE 6.50 usec
 TE 300.0 K
 D1 2.0000000 sec
 D11 0.03000000 sec
 TDO 1

==== CHANNEL f1 =====
 SFO1 100.6304993 MHz
 NUC1 13C
 P1 9.00 usec
 PLW1 61.09999847 W

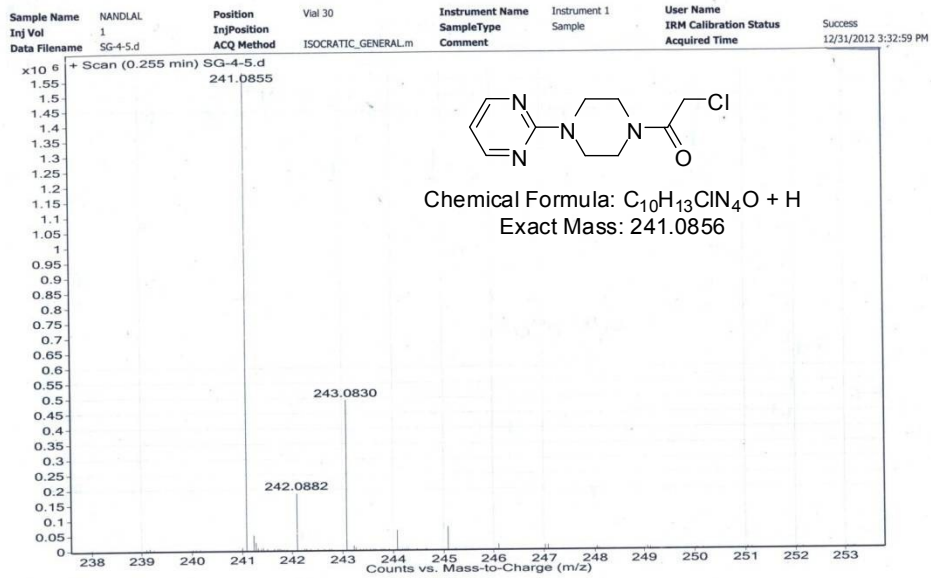
==== CHANNEL f2 =====
 SFO2 400.1621006 MHz
 NUC2 1H
 CPDPRG2 waltz16
 PCPD2 90.00 usec
 PLW2 13.10000038 W
 PLW12 0.26705000 W
 PLW13 0.21630999 W

F2 - Processing parameters
 SI 32768
 SF 100.6204380 MHz
 WDW no
 SSB 0
 LB 0 Hz
 GB 0
 PC 1.40

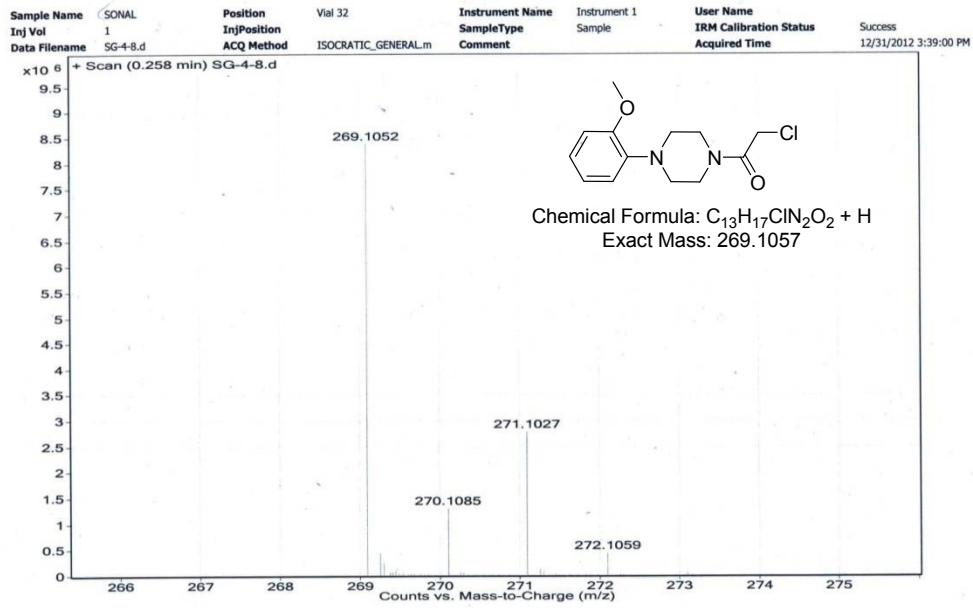
HRMS of compound 8a



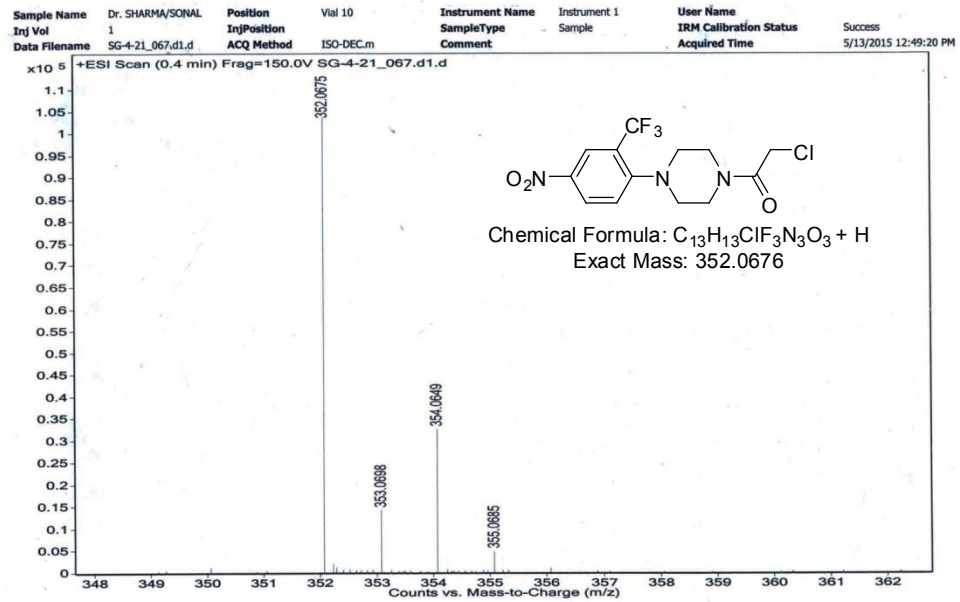
HRMS of compound 8b



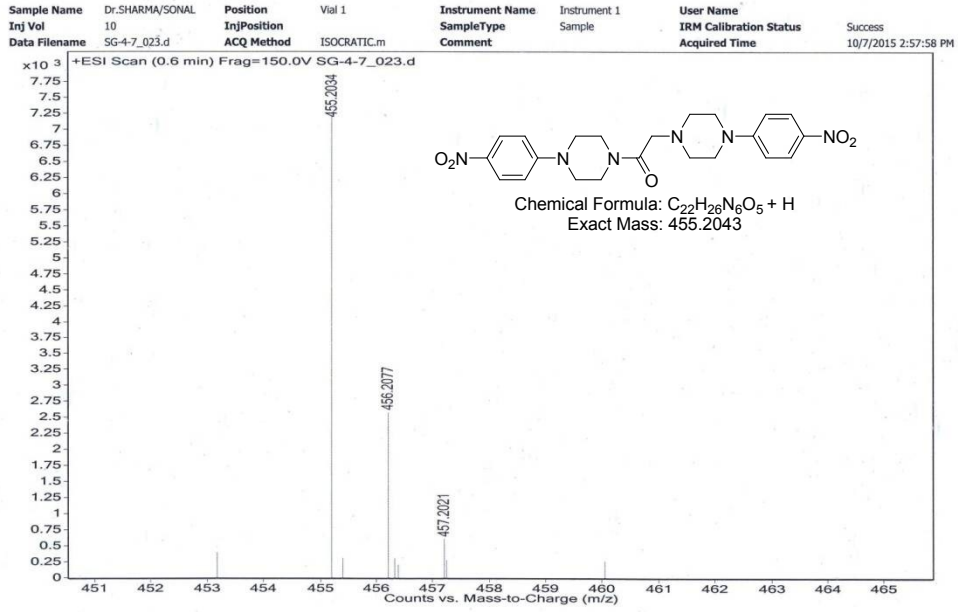
HRMS of compound 8c



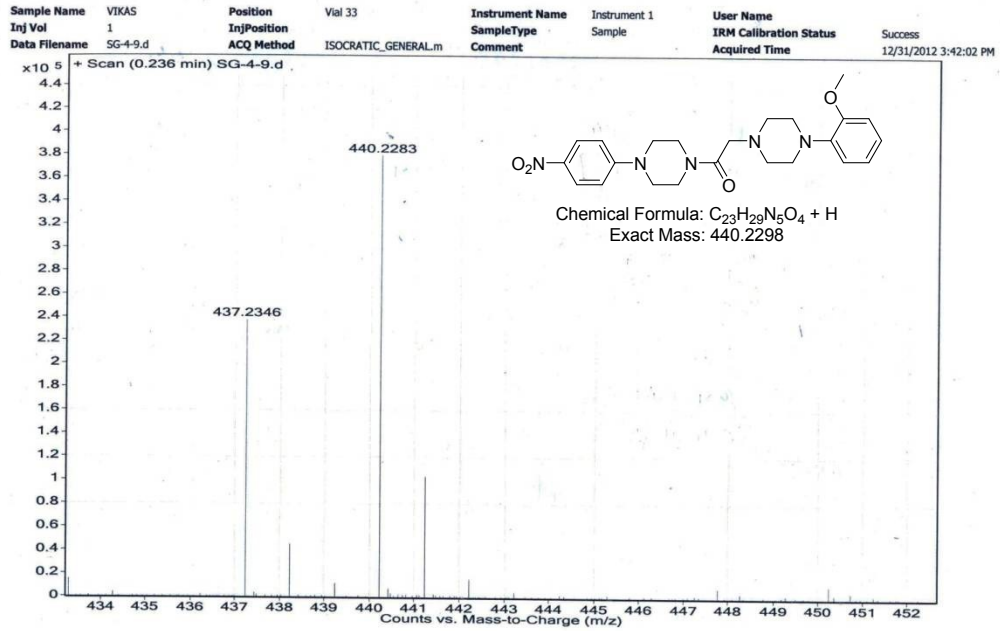
HRMS of compound 8d



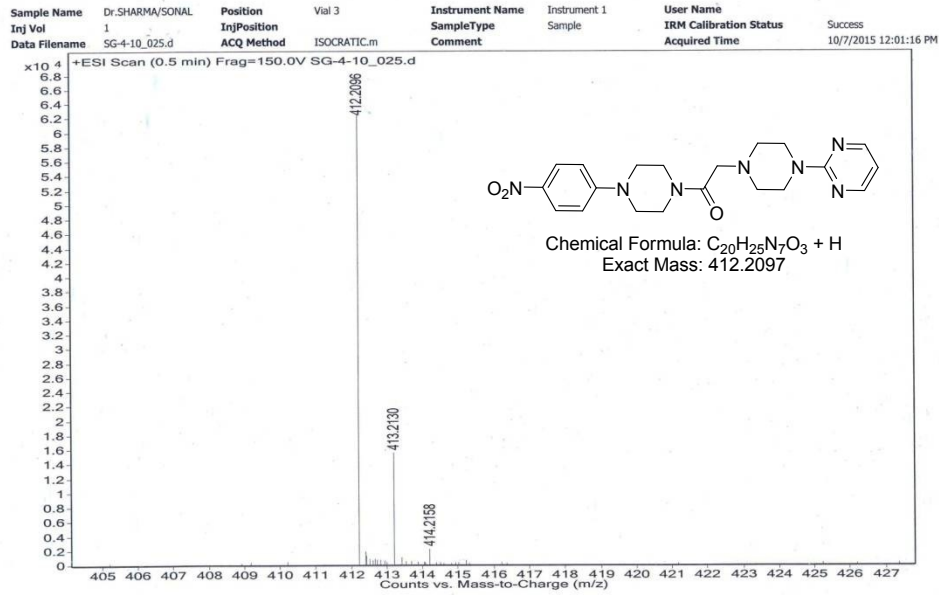
HRMS of compound 9a



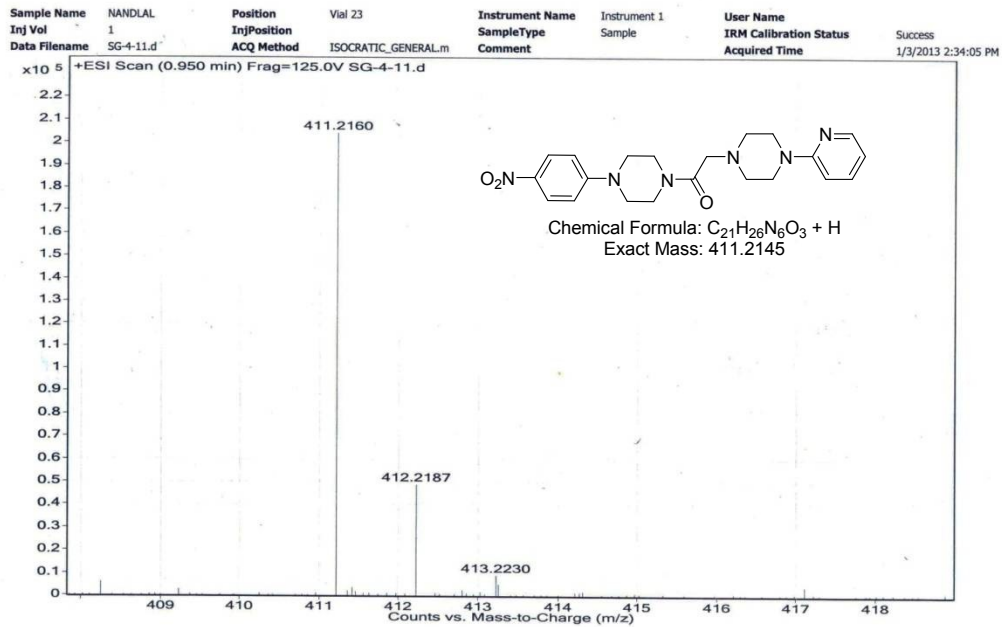
HRMS of compound 9b



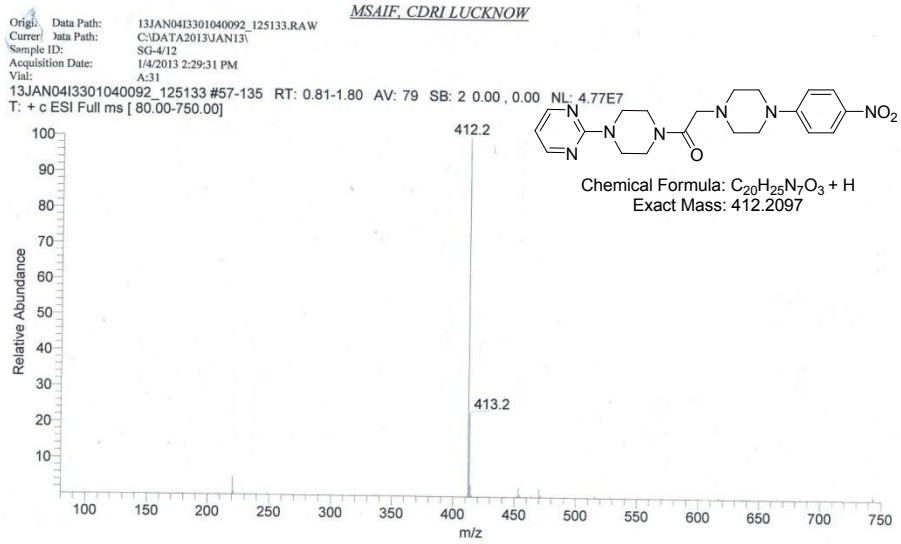
HRMS of compound 9c



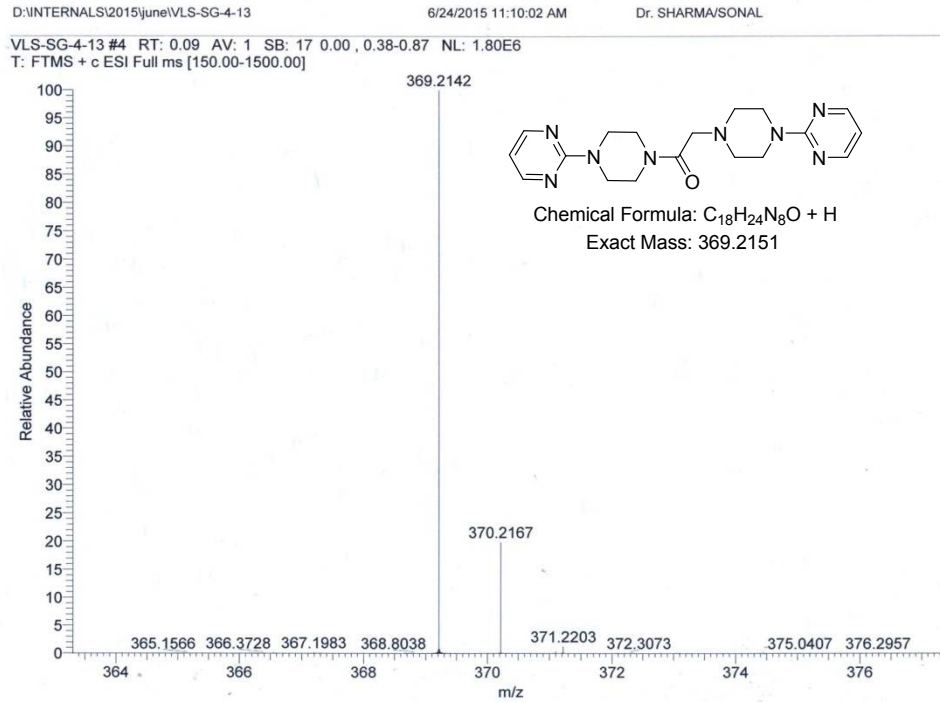
HRMS of compound 9d



ESI-MS of compound 9e



HRMS of compound 9f



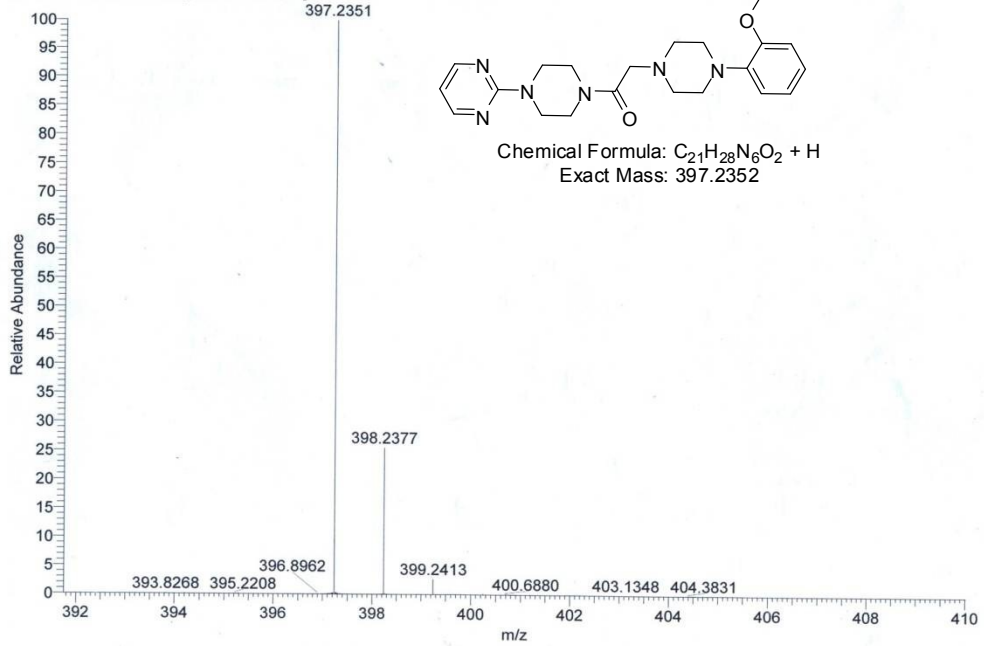
HRMS of compound 9g

D:\INTERNALS\2015\may\SG-4-14

5/12/2015 11:09:04 AM

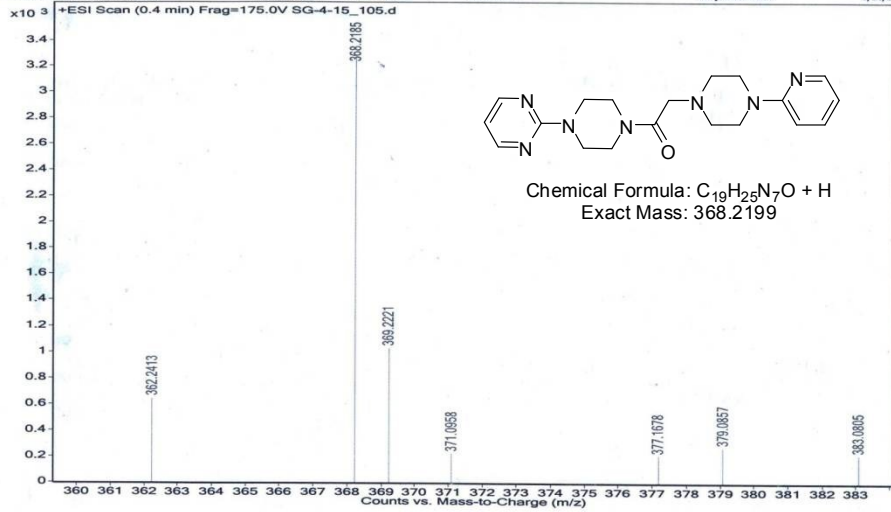
Dr. V.L.S/SONAL

SG-4-14 #9 RT: 0.13 AV: 1 NL: 1.42E7
T: FTMS + c ESI Full ms [150.00-2000.00]



HRMS of compound 9h

Sample Name	Dr. SHARMA/DHANARAJU	Position	Vial 7	Instrument Name	Instrument 1	User Name	Success
Inj Vol	1	InjPosition		SampleType	Sample	IRM Calibration Status	5/20/2015 12:56:46 PM
Data Filename	SG-4-15_105.d	ACQ Method	ISO-DEC.m	Comment		Acquired Time	



HRMS of compound 9i

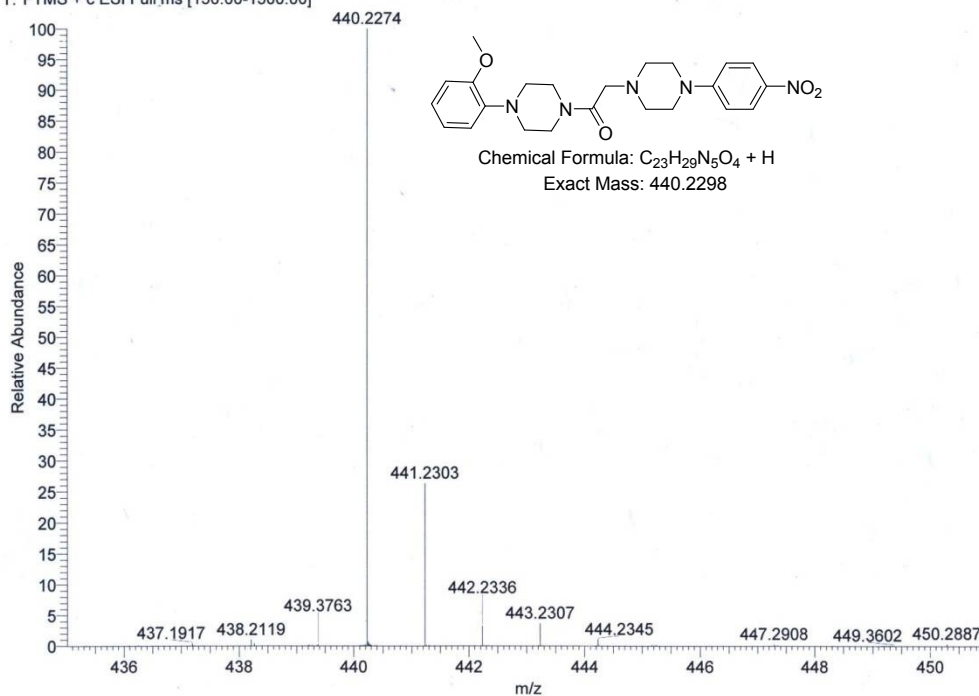
D:\INTERNALS\2015\june\VLS-SG-4-17

6/23/2015 11:33:29 AM

Dr. SHARMA/SONAL

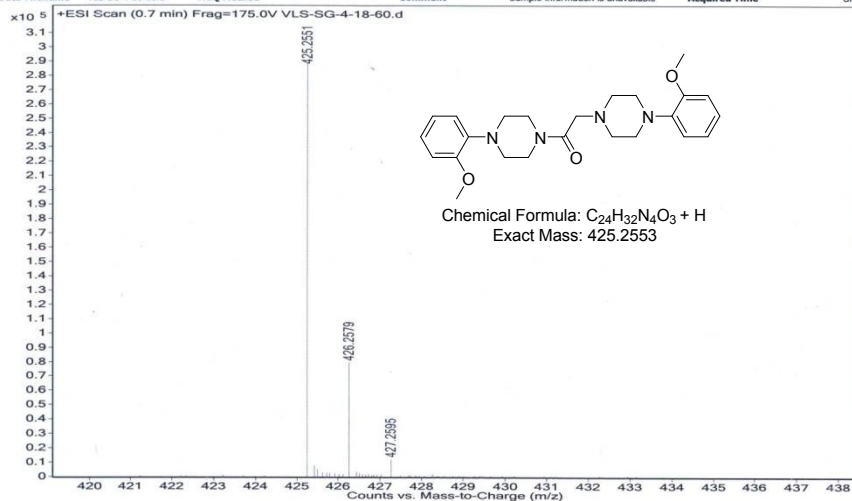
VLS-SG-4-17 #7 RT: 0.11 AV: 1 SB: 23 0.00, 0.29-0.71 NL: 8.13E5

T: FTMS + c ESI Full ms [150.00-1500.00]



HRMS of compound 9j

Sample Name	Unavailable	Position	Unavailable	Instrument Name	Unavailable	User Name	Unavailable
Inj Vol	Unavailable	InjPosition	Unavailable	SampleType	Unavailable	IRM Calibration Status	Success
Data Filename	VLS-SG-4-18-60.d	ACQ Method	Unavailable	Comment	Sample information is unavailable	Acquired Time	Unavailable



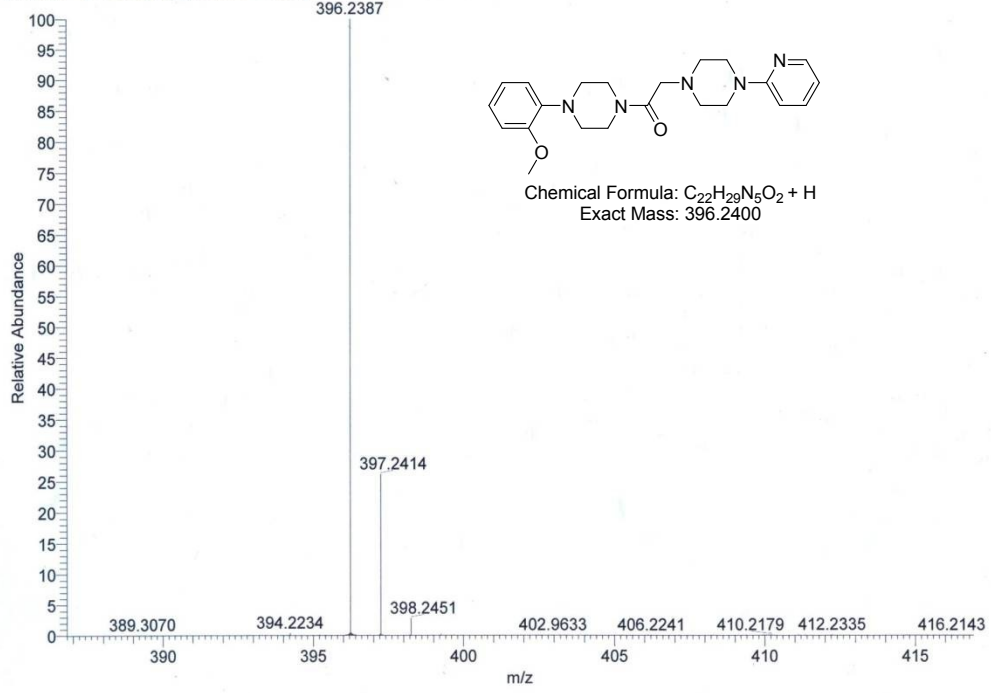
HRMS of compound 9k

D:\INTERNALS\2015\june\VLS-SG-9-14

6/19/2015 2:32:56 PM

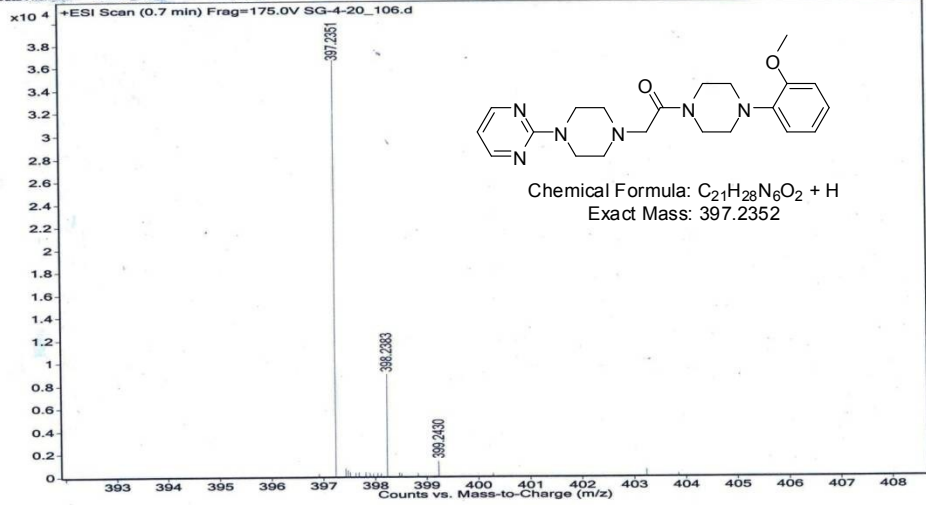
Dr. SHARMA/SONAL

VLS-SG-9-14 #11 RT: 0.17 AV: 1 SB: 27 0.00-0.02, 0.39-0.86 NL: 1.29E6
T: FTMS + c ESI Full ms [150.00-1500.00]

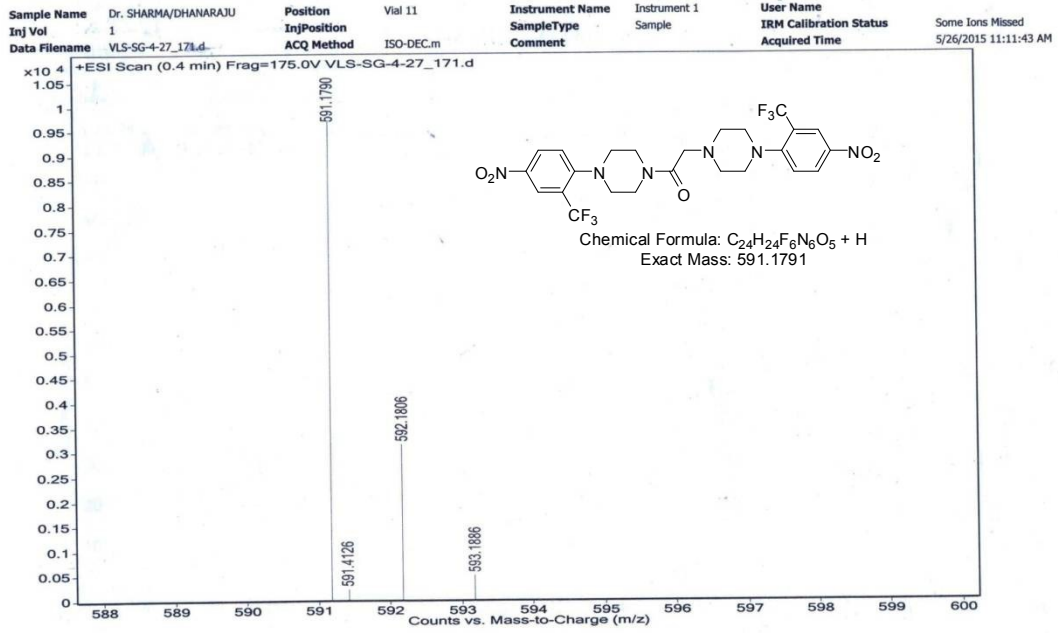


HRMS of compound 9l

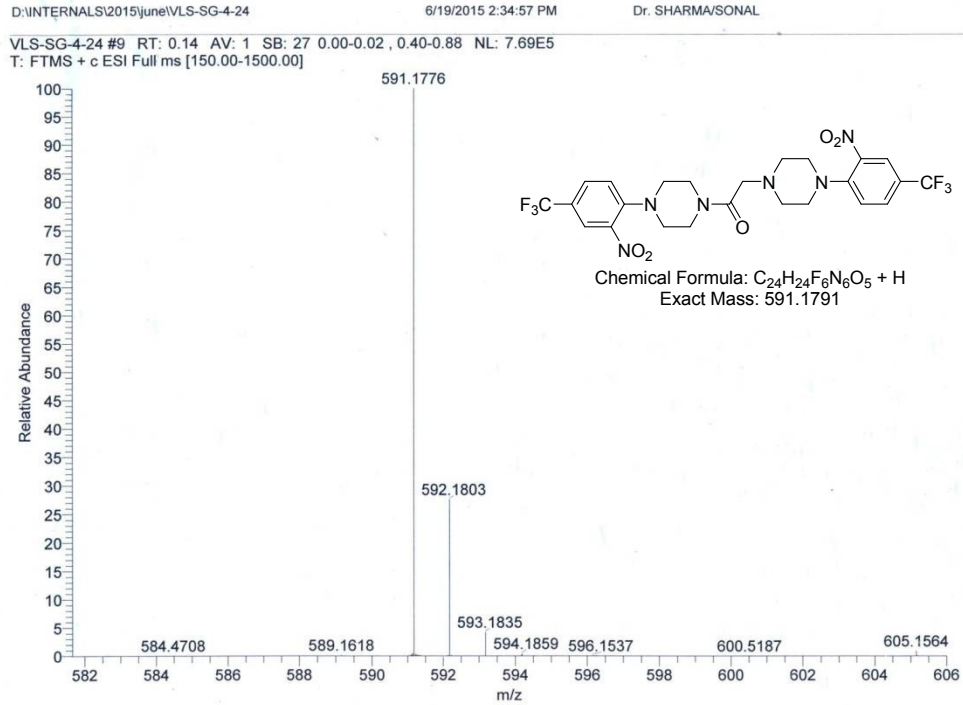
Sample Name	Dr. SHARMA/SONAL	Position	Vial 8	Instrument Name	Instrument 1	User Name	
Inj Vol	1	InjPosition		SampleType	Sample	IRM Calibration Status	Success
Data Filename	SG-4-20_106.d	ACQ Method	ISO-DEC.m	Comment		Acquired Time	5/20/2015 1:00:26 PM



HRMS of compound 9m

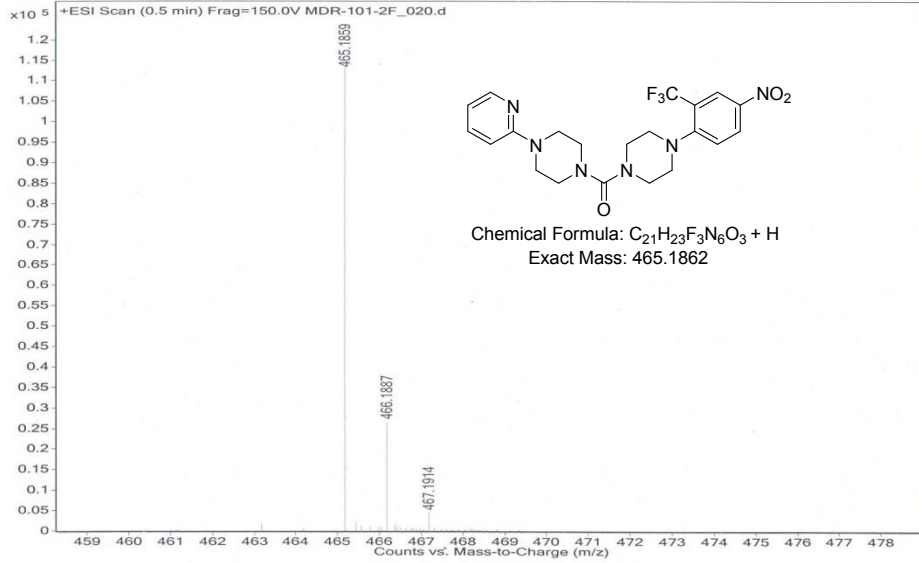


HRMS of compound 9n



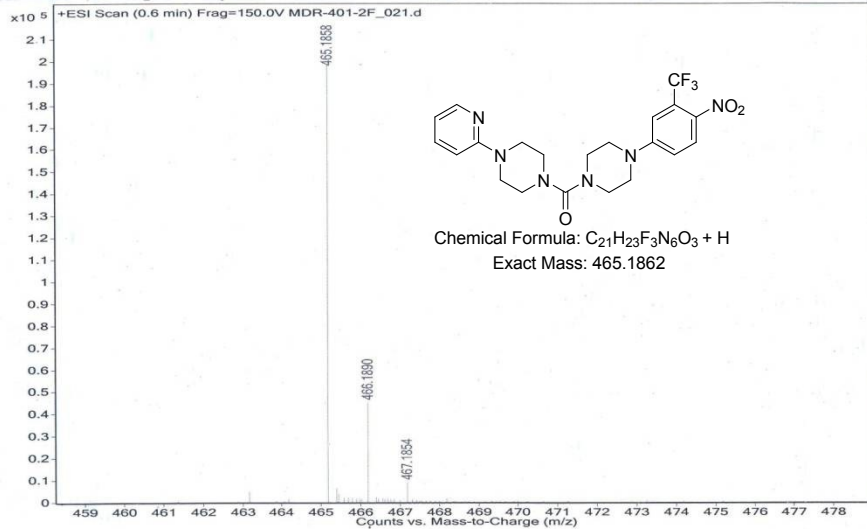
HRMS of compound 12a

Sample Name	Dr. SHARMA/MALA	Position	Vial 11	Instrument Name	Instrument 1	User Name	
Inj Vol	1	InjPosition		SampleType	Sample	IRM Calibration Status	Success
Data Filename	MDR-101-2F_020.d	ACQ Method	ISOCRATIC.m	Comment		Acquired Time	10/6/2015 12:48:34 PM

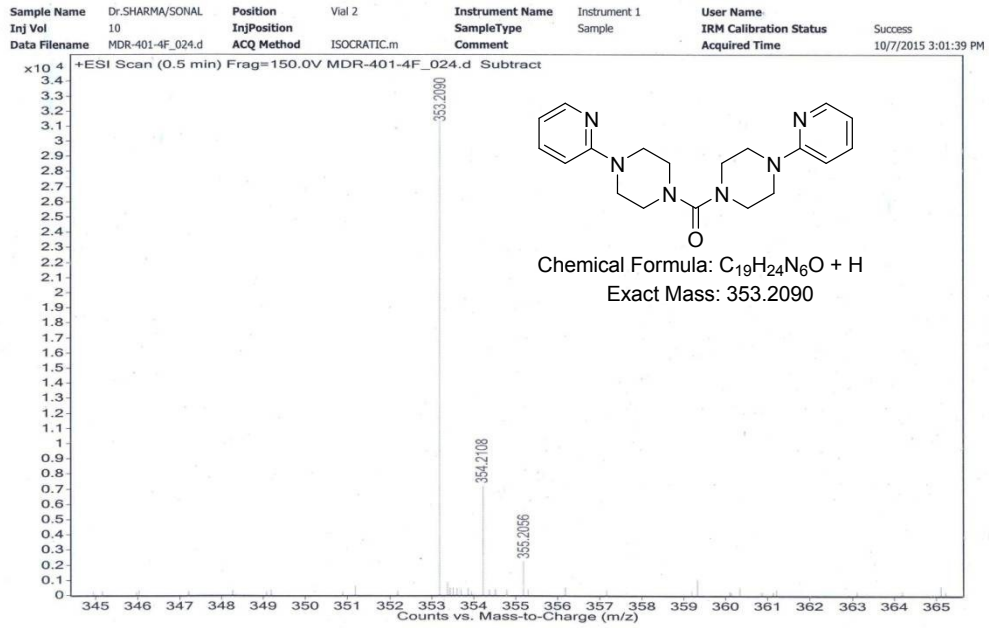


HRMS of compound 12b

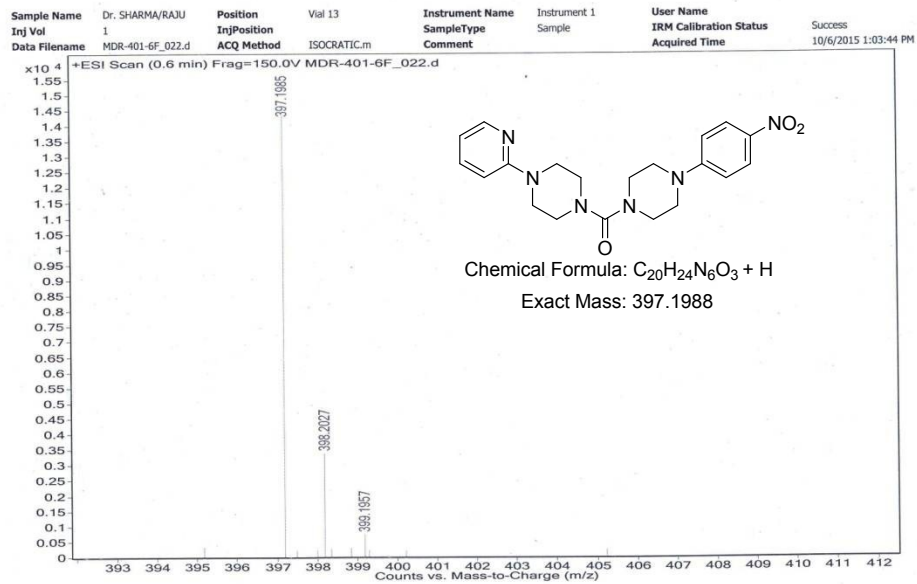
Sample Name	Dr. SHARMA/SONAL	Position	Vial 12	Instrument Name	Instrument 1	User Name	
Inj Vol	1	InjPosition		SampleType	Sample	IRM Calibration Status	Success
Data Filename	MDR-401-2F_021.d	ACQ Method	ISOCRATIC.m	Comment		Acquired Time	10/6/2015 1:00:07 PM



HRMS of compound 12c



HRMS of compound 12d



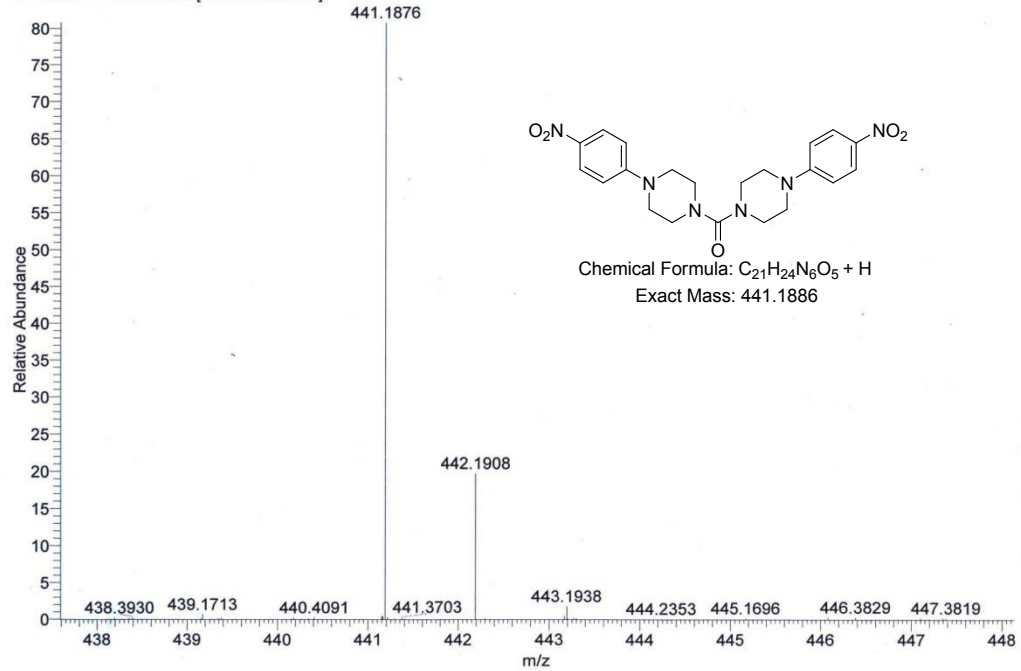
HRMS of compound 12e

D:\INTERNALS\2015\july\VLS-701-2F

7/8/2015 2:50:38 PM

Dr. SHARMA/SONAL

VLS-701-2F #5 RT: 0.12 AV: 1 SB: 11 0.29-0.63 NL: 8.91E4
T: FTMS + c ESI Full ms [150.00-1500.00]



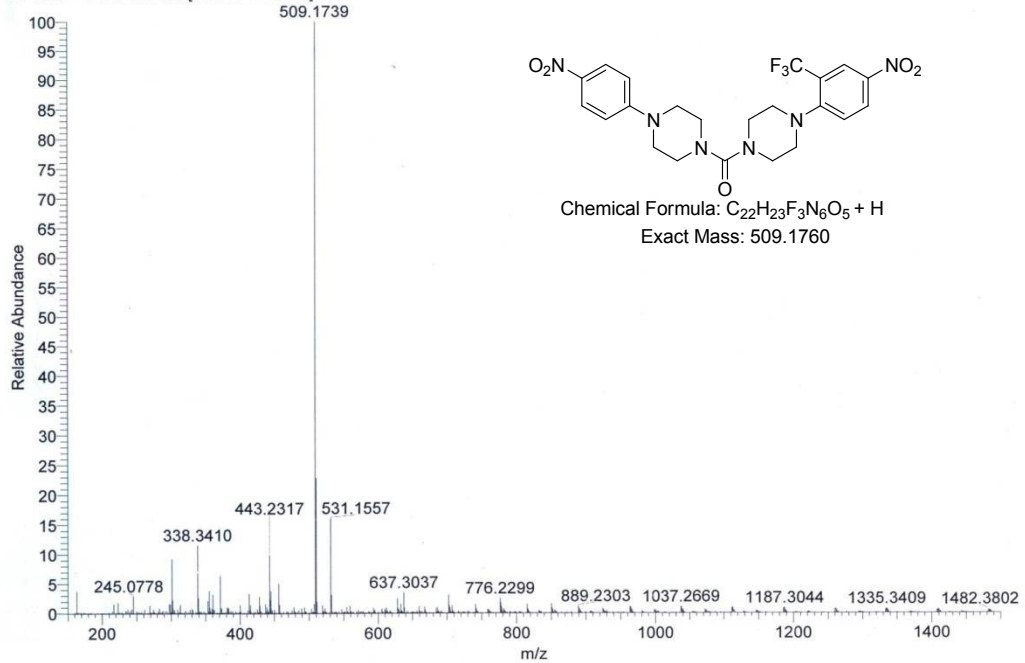
HRMS of compound 12f

D:\INTERNALS\2015\july\VLS-701-4F

7/8/2015 2:48:38 PM

Dr. SHARMA/SONAL

VLS-701-4F #3 RT: 0.06 AV: 1 SB: 11 0.29-0.63 NL: 4.84E5
T: FTMS + c ESI Full ms [150.00-1500.00]



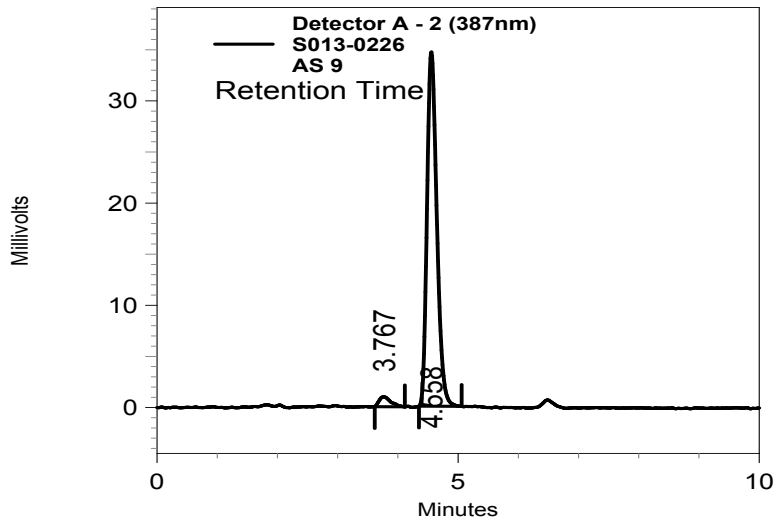
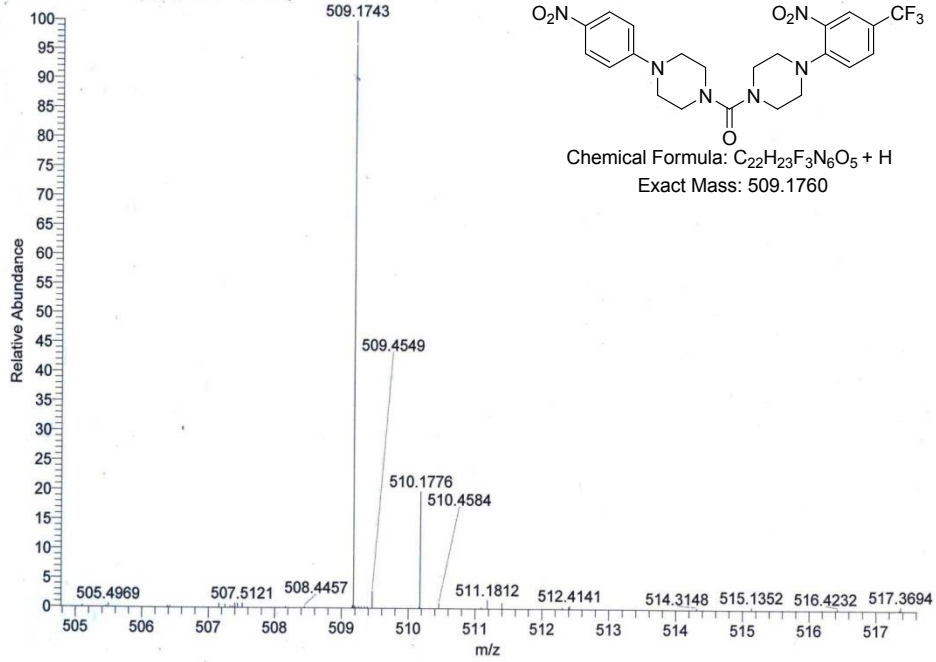
HRMS of compound 12g

D:\INTERNALS\2015\july\VLS-701-5F

7/10/2015 11:48:38 AM

Dr. SHARMA/SONAL

VLS-701-5F #9 RT: 0.26 AV: 1 NL: 6.54E4
T: FTMS + c ESI Full ms [150.00-1500.00]



Sample Conc. 1000 ng/mL
Mobile Phase: ACN: MeOH: AAB (10 mM) :: 50:10:40
Run Time: 10 min
 λ_{max} : 387 nm
Flow rate: 1 mL/min
Column: Discovery HS C-18 10 cm (Reverse phase)
Retention time: 4.5
Purity: 97%

Figure 1: Representative chromatogram of **9a**

REFERENCES

1. D. Engelstein, J. Shmueli, S. Bruhis, C. Servadio, A. Abramovici, *Comp. Biochem. Physiol. C. Pharmacol. Toxicol. Endocrinol.*, 1996, **115**, 169-77.
2. <http://www.fda.gov/downloads/Drugs/Guidances/ucm070107.pdf>, FDA, Guidance for Industry: Bioanalytical Method Validation.
3. SYBYL-X, version 2.1, Tripos, **1991-2012**, Certara.
4. A. N. Jain, *J. Med. Chem.*, 2003, **46**, 499-511.