

Exploiting the Interplay of Quantum Interference and Backbone Rigidity on Electronic Transport in Peptides: A Step Towards Bio-Inspired Quantum Interferometers

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1. General Information

Chemicals

Fmoc-Aib-OH, Boc-Aib-OH, Fmoc-Lys(Boc)-OH, Boc-Lys(Cbz)-OH, Boc-Glu(OBzl)-OH, H-Leu-OMe, H-Ala-OMe, 1-(3-Dimethylaminopropyl)-3-ethylcarbodiimide HCl (EDC·HCl), Fmoc-OSu, 2-chlorotriyl chloride polystyrene resin, 1-hydroxy-7-azabenzotriazole (HOAt) and 2-(1H-7-azabenzotriazol-1-yl)-1,1,3,3-tetramethyl uranium hexafluorophosphate methanaminium (HATU) were purchased from GL Biochem (Shanghai) Ltd, China. Dichloromethane (DCM), diethyl ether (Et₂O), ethyl acetate (EtOAc), methanol and ethanol were purchased from Ajax Finechem Pty Ltd (Australia). Piperidine, acetonitrile, propan-2-ol, potassium carbonate, and *N,N*-dimethylformamide (DMF) were purchased from Merck, Australia. Anhydrous *N,N*-dimethylformamide (DMF), dimethyl sulfoxide (DMSO), tetrahydrofuran (THF), dioxane, 2,2,2-trifluoroethanol (TFE), trifluoroacetic acid (TFA), 4 M HCl/dioxane solution, cysteamine, methyl iodide, Pd/C and diisopropylethylamine (DIPEA) were purchased from Sigma-Aldrich, Australia. SOCl₂, CH₃COOH and NaOH were purchased from Chem Supply, Australia. Single-walled carbon nanotubes (P2-SWCNTs), were purchased from Carbon Solutions Inc., USA. Ferrocenylmethylamine^{1, 2} was prepared as published. All solvents and reagents were used without purification unless noted.

High-Performance Liquid Chromatography

The synthetic peptides were analyzed and purified by reverse phase HPLC, using an HP 1100 LC system equipped with a Phenomenex C18 column (250x4.6 mm) for analytical traces and a Phenomenex C18 column (250 x 21.2 mm) for purification, a photodiode array detector, and a Sedex evaporative light scattering detector. Water/TFA (100/0.1 by v/v) and ACN/TFA (100/0.08 by v/v) solutions were used as aqueous and organic buffers.

NMR Spectroscopy.

¹H NMR spectra were recorded in DMSO-d₆ or CDCl₃-d solutions using a Varian Gemini-300 NMR. ¹³C NMR and two-dimensional NMR experiments utilized COSY, ROESY, HSQC and HMBC were obtained on a Varian Inova 600 MHz spectrometer. Chemical shifts are reported in ppm (δ) using TMS (0.00 ppm) as the internal standard. Signals are reported as s (singlet), d (doublet), t (triplet) or m (multiplet).

Mass Spectroscopy

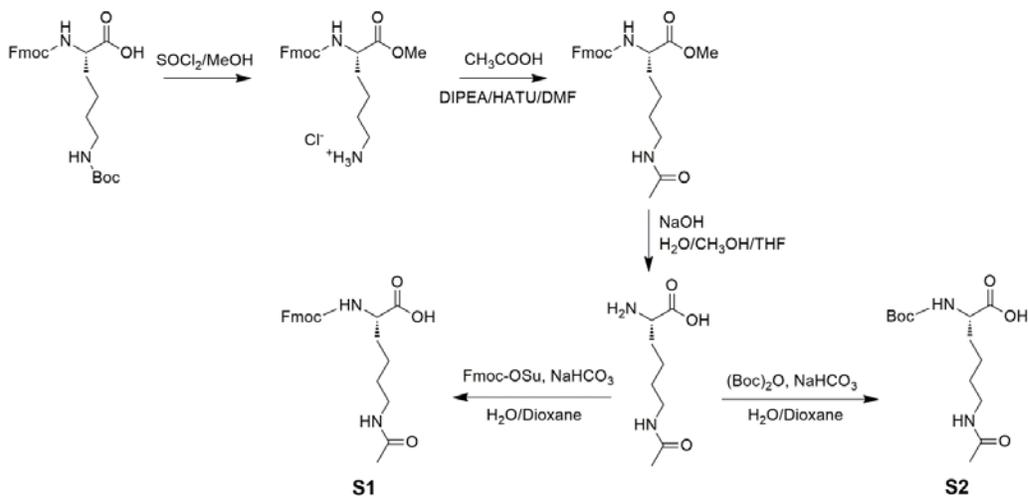
Low resolution mass spectral data were analyzed using a Finnigan MAT LCQ spectrometer with MS/MS and ESI probe, utilizing XCalibur software. High resolution mass spectral data were analyzed using an Ultimate 3000 RSL HPLC (Thermo Fisher Scientific Inc., MA) and an LTQ Orbitrap XL ETD using a flow injection method, with a flow rate of 5 μL/min. The HPLC flow is interfaced with the mass spectrometer using the Electrospray source (Thermo Fisher Scientific Inc., MA). Mass spectra were obtained over a range of 100 < m/z < 1000. Data was analyzed using XCalibur software (Version 2.0.7, Thermo Fisher Scientific).

FTIR Spectroscopy

Infrared spectra were collected on a Perkin Elmer Spectrum 100 FT-IR spectrometer, with attenuated total reflectance (ATR) imaging capabilities, fitted with a ZnSe crystal, with an average reading taken from 4 scans at 4 cm⁻¹ resolution.

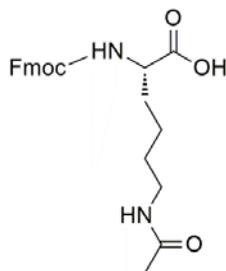
2. Synthesis of peptides

Scheme S1. The synthetic steps for building blocks S1 and S2.



Fmoc-Lys(Boc)-OH (4.19 g, 8.9 mmol) was dissolved in 40 mL dry methanol (over molecular sieves) and cooled to 0 °C. Thionyl chloride (2.5 mL) was added dropwise to the methanolic solution over 5 min. The mixture was stirred at 0 °C for 30 min, then warmed to room temperature and further stirred overnight. The volatiles were removed to yield the intermediate Fmoc-Lys-OMe, an off-white solid (3.80 g, quant). The resulting residue and CH₃COOH (2.0 mL) were dissolved in anhydrous DMF (15 mL) and stirred at rt under an N₂ atmosphere. HATU (6.50 g, 17.1mmol) and DIPEA (6.0 mL) were added, and the mixture stirred for 48 h. The solvent was removed and the residue taken up in EtOAc (200 mL) and H₂O (200 mL). The organic layer was separated and washed with NaHCO₃ (200 mL), brine (200 mL) and dried over Na₂SO₄. The volatiles were removed *in vacuo* to reveal a white solid (3.91 g, quant). The resulting residue was dissolved in the mixture of THF (23.0 mL) and methanol (16.0 mL). 9.0 mL of NaOH solution (1.6 M, aqueous) was added to the mixture, and the reaction stirred at rt for 18 h. The solvent was removed *in vacuo* and the residue redissolved in H₂O (200mL), washed with Et₂O (200 mL). The aqueous layer was separated and dried over MgSO₄. The solvent was removed *in vacuo* to yield the amphoteric intermediate, an off-white solid (1.85 g, quant).

Compound S1



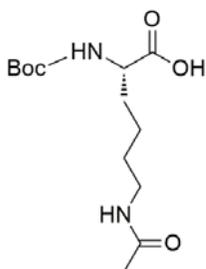
The amphoteric residue (1.85 g, 8.0 mmol) was dissolved in para-dioxane (20.0mL). A solution of NaHCO₃ (2.20 g in 20.0 mL H₂O, 16.0mmol, 2equiv) was added followed by Fmoc-OSu (2.90 g, 8.0mmol, 1 equiv). The reaction mixture was stirred overnight at rt after which the volatiles

were removed under reduced pressure. The residue was dissolved in 2.5% NaHCO₃ and washed with Et₂O (3x20 mL). The aqueous layer was then acidified to pH 4 by dropwise addition of 6 M aqueous HCl and extracted with EtOAc (3x50 mL). The combined organic layers were washed with brine (20 mL), dried over Na₂SO₄ and the solvent removed under reduced pressure to give the product as a white solid (2.51 g, 76%).

¹H NMR (300 MHz, DMSO-d₆): δ 7.88 (d, 2H, aromH), 7.81 (t, 1H, NHCH₂), 7.41 (t, 2H, aromH), 7.72 (d, 2H, aromH), 7.62 (d, 1H, NHCH), 7.46-7.28 (m, 4H, aromH), 4.32-4.16 (d, 3H, CH& CH₂ in Fmoc), 3.90 (m, 1H, NHCH), 3.06-2.96 (m, 2H, NHCH₂), 1.78 (s, 3H, CH₃), 1.64 (m, 2H, CH₂), 1.36 (m, 4H, 2xCH₂).

MS: [M+H]⁺_{calcd}=411.2, [M+H]⁺_{found}=411.2.

Compound S2

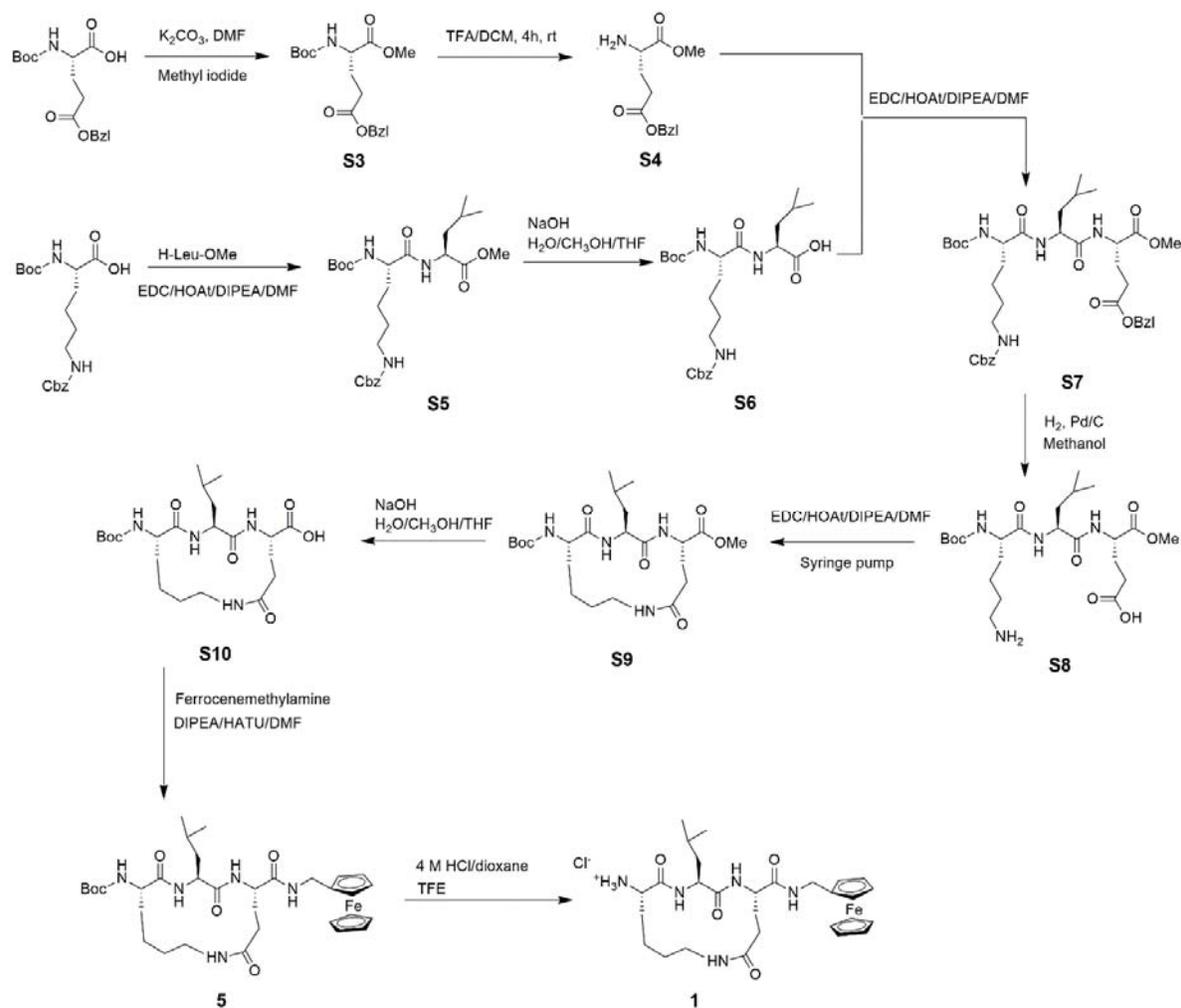


The amphoteric residue (3.93 g, 17.0mmol) and NaHCO₃ (2.86 g, 34.0 mmol) were dissolved in water (60.0 mL). A solution of (Boc)₂O (5.60 g in 60.0 mL para-dioxane) was added. The reaction mixture was stirred overnight at rt after which the volatiles were removed under reduced pressure. The residue was redissolved in 200 mL of water, washed with EtOAc (200 mL). The aqueous layer was then acidified to pH 3 by dropwise addition of 6 M aqueous HCl and extracted with EtOAc (3x100 mL). The combined organic layers were washed with brine (50 mL), dried over Na₂SO₄ and the solvent removed under reduced pressure to give the product as a white solid (4.40 g, 89%).

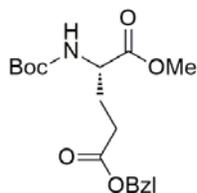
¹H NMR (300 MHz, DMSO-d₆): δ 7.78(t, 1H, NHCH₂), 7.05 (d, 1H, NHCH), 3.80 (m, 1H, NHCH), 3.06-2.92 (m, 2H, NHCH₂), 1.76 (s, 3H, CH₃), 1.58 (m, 2H, CH₂), 1.40-1.20 (m, 13H, Boc & 2xCH₂).

MS: [M+H]⁺_{calcd}=289.2, [M+H]⁺_{found}=289.2.

Scheme S2. The final synthetic steps for lactam-bridged β -strand peptides



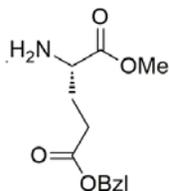
Compound S3



K_2CO_3 (982 mg, 7.11 mmol) was suspended in anhydrous DMF (20 mL). Boc-Glu(OBzl)-OH (2.0 g, 5.93 mmol) was added, followed by MeI (1.01 g, 7.11 mmol, 443 μ L). The reaction was stirred at rt under an N_2 atmosphere for 18 h. The solvent was removed and the residue taken up in EtOAc (200 mL) and H_2O (200 mL). The pH was adjusted to pH 3-4 and the organic layer separated and washed with brine (200 mL) and dried over $NaSO_4$. The solvent was removed *in vacuo* to reveal golden oil (1.80 g, 86%).

1H NMR (300 MHz, DMSO- d_6): δ 7.40-7.30 (m, 6H, benzene, NH), δ 5.09 (s, 2H, OCH_2), δ 4.06-3.98 (m, 1H, CaH), δ 3.61 (s, 3H, OCH_3), δ 2.45 (dd, 2H, CH_2 , $J=9.8, 5.8$ Hz), δ 2.01-1.74 (m, 2H, CH_2), δ 1.37 (s, 9H, Boc).

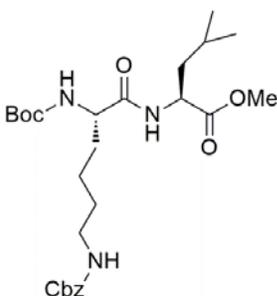
Compound S4



Compound S3 (877 mg, 2.50 mmol) was dissolved in DCM (5 mL). TFA (5 mL) was added dropwise and the reaction stirred at rt for 3 h. The solvent was removed *in vacuo* to reveal golden oil (1.21 g, quant).

^1H NMR (300 MHz, DMSO- d_6): δ 8.44 (br s, 3H, NH), 7.37 (m, 5H, benzene), 5.11 (s, 2H, OCH₂), 4.10 (br s, 1H, C α H), 3.72 (s, 3H, OCH₃), 2.56 (m, 2H, CH₂), 2.04 (m, 2H, CH₂).

Compound S5

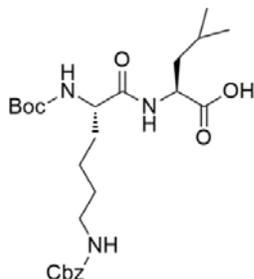


Boc-Lys(Z)-OH (1.00 g, 2.63 mmol) and HCl·H₂N-Leu-OMe (572 mg, 3.16 mmol) were dissolved in anhydrous DCM (20 mL) and stirred at rt under an N₂ atmosphere. Anhydrous DIPEA (1.6 mL), EDC·HCl (552 mg, 2.89 mmol) and HOAt (393 mg, 2.89 mmol) were added and the solution stirred for 36 h. DCM (30 mL) and H₂O (50 mL) were added and the pH adjusted to pH 2-3. The organic layer was separated and washed with brine (50 mL) and dried over MgSO₄. The solvent was removed *in vacuo* to yield clear oil (1.07 g, 81%).

^1H NMR (300 MHz, DMSO- d_6): δ 8.11 (d, 1H, NH, $J=7.6$ Hz), 7.40-7.30 (m, 5H, benzene), 7.23 (t, 1H, NH, $J=10.5$ Hz), 6.79 (d, 1H, NH, $J=8.1$ Hz), 5.00 (s, 2H, OCH₂), 4.33-4.26 (m, 1H, C α H), 3.90 (d, 1H, C α H, $J=5.4$ Hz), 3.60 (s, 3H, OCH₃), 2.97 (d, 2H, CH₂NH, $J=5.9$ Hz), 1.70-1.22 (m, 9H, 4xCH₂, CH), 1.37 (s, 9H, Boc), 0.90-0.82 (m, 6H, 2xCH₃ Leu).

MS: [M+Na]⁺_{calcd}=530.2, [M+Na]⁺_{found}=530.2.

Compound S6

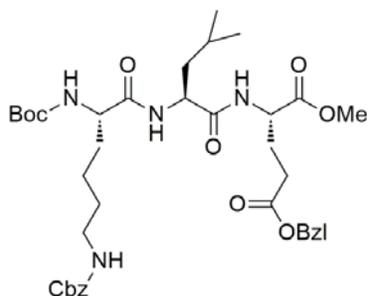


Compound **S5** (1.06 g, 2.10 mmol) was dissolved in THF (8.37 mL) and methanol (2.13 mL). NaOH (125 mg, 3.14 mmol) was dissolved in H₂O (2.13 mL) and added to the solution and stirred at rt for 24 h. The THF was removed *in vacuo* and EtOAc (50 mL) and H₂O (50 mL) added. The pH was adjusted to pH 2-3, the organic layer separated and washed with brine (50 mL) and dried over MgSO₄. The solvent was removed *in vacuo* to yield a clear solid (904 mg, 87%).

¹H NMR (300 MHz, DMSO-*d*₆): δ 7.93 (d, 1H, NH, *J*=7.7 Hz), 7.40-7.30 (m, 5H, benzene), 7.23 (t, 1H, NH, *J*=10.2 Hz), 6.79 (d, 1H, NH, *J*=8.3 Hz), 5.00 (s, 2H, OCH₂), 4.22 (dd, 1H, CaH, *J*=14.2, 8.5 Hz), 3.89 (d, 1H, CaH, *J*=4.8 Hz), 2.96 (d, 2H, CH₂NH, *J*=5.6 Hz), 1.70-1.22 (m, 9H, 4xCH₂, CH), 1.37 (s, 9H, Boc), 0.90-0.82 (m, 6H, 2xCH₃ Leu).

MS: [M+Na]⁺_{calcd}=516.2, [M+Na]⁺_{found}=516.2.

Compound S7

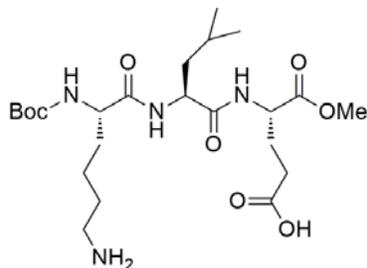


Compound **S4** (627 mg, 2.50 mmol) and compound **S6** (1.12 g, 2.27 mmol) were dissolved in anhydrous DCM (42 mL) and stirred at rt under an N₂ atmosphere. Anhydrous DIPEA (1.58 mL), HATU (949 mg, 2.50 mmol) and HOAt (308 mg, 2.27 mmol) were added and the solution stirred for 42 h. DCM (60 mL) and H₂O (100 mL) were added and the pH adjusted to pH 3. The organic layer was separated and washed with brine (100 mL) and dried over MgSO₄. The solvent was removed *in vacuo* to reveal a white solid, which was purified by column chromatography on silica gel (70/30 EtOAc/PE), yielding (785 mg, 48%).

¹H NMR (300 MHz, DMSO-*d*₆): δ 8.31 (d, 1H, NH, *J*=7.2 Hz), 7.77 (d, 1H, NH, *J*=7.9 Hz), 7.40-7.30 (m, 10H, 2x benzene), 7.23 (t, 1H, NH, *J*=9.0 Hz), 6.87 (d, 1H, NH, *J*=8.0 Hz), 5.09 (s, 2H, OCH₂), 4.99 (s, 2H, OCH₂), 4.30 (dd, 2H, 2x CaH, *J*=13.5, 6.2 Hz), 3.89-3.82 (m, 1H, CaH), 3.59 (s, 3H, OCH₃), 2.99-2.91 (m, 2H, CH₂NH), 2.43 (t, 2H, CH₂, *J*=7.5 Hz), 2.07-1.22 (m, 11H, 5x CH₂, CH), 1.36 (s, 9H, Boc), 0.89-0.82 (m, 6H, (CH₃)₂ Leu).

MS: [M+Na]⁺_{calcd}=749.3, [M+Na]⁺_{found}=749.3.

Compound S8

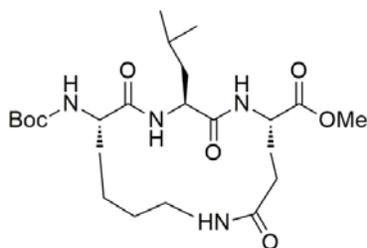


Compound S7 (464 mg, 0.639 mmol) was dissolved in (methanol over molecular sieves, 10 mL) and stirred at rt under an N₂ atmosphere for 10 min. Pd/C (15% w/w, 70 mg) was added. A H₂ balloon was fitted under vacuum and the solution stirred at rt for a further 24 h. The solution was filtered through celite and washed with methanol (3x 20 mL), and the solvent removed *in vacuo* to yield a white solid (232 mg, 72%).

¹H NMR (600 MHz, DMSO-d₆): δ 8.89 (d, 1H, NH, *J*=6.5 Hz), 8.02 (d, 1H, NH, *J*=8.5 Hz), 6.78 (d, 1H, NH, *J*=8.0 Hz), 4.33 (dd, 1H, CαH, *J*=14.4, 9.0 Hz), 4.23 (t, 1H, CαH, *J*=9.8 Hz), 3.95 (dd, 1H, CαH, *J*=14.0, 7.4 Hz), 3.58 (s, 3H, OCH₃), 2.03-1.26 (m, 15H, 7x CH₂, CH), 1.36 (s, 9H, Boc), 0.87-0.81 (m, 6H, 2xCH₃ Leu).

MS: [M+Na]⁺_{calcd}=525.3, [M+Na]⁺_{found}=525.3.

Compound S9



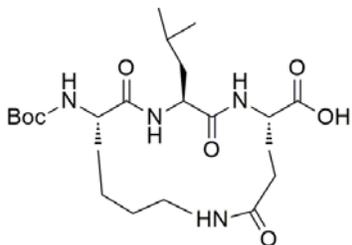
Compound S8 (127 mg, 0.253 mmol) was suspended in anhydrous DMF (26 mL) and anhydrous THF (20 mL) and sonicated. EDC·HCl (238 mg, 1.25 mmol), HOAt (170 mg, 1.25 mmol) and DIPEA (435 μL) were dissolved in anhydrous DMF (15 mL) and anhydrous THF (8 mL) and the solution stirred at rt under an N₂ atmosphere. The compound S8 mixture was placed into a syringe pump and added to the coupling reagents at the rate of 30 μL/min, and stirred for 48 h. The crude product was purified using reverse phase HPLC to yield a white solid (39 mg, 31%).

¹H NMR (600 MHz, DMSO-d₆): δ 8.25 (d, 1H, NH, *J*=7.4 Hz), 8.06 (d, 1H, NH, *J*=8.2 Hz), 7.42 (m, 1H, NH), 6.43 (d, 1H, NH, *J*=7.3 Hz), 4.43 (m, 1H, CαH), 4.34 (m, 1H, CαH), 4.05 (m, 1H, CαH), 3.60 (s, 3H, OCH₃), 3.32 (m, 1H, CHNH), 2.71 (m, 1H, CHNH), 2.28-1.04 (m, 13H, 6x CH₂, CH), 1.36 (s, 9H, Boc), 0.90-0.84 (m, 6H, 2xCH₃ Leu).

¹³C NMR (150 MHz, DMSO-d₆): δ 172.4, 172.1, 171.4, 170.7, 158.0, 154.5, 77.8, 53.2, 51.7, 50.8, 50.4, 48.5, 41.0, 36.9, 31.4, 28.4, 28.1, 27.9, 25.5, 24.6, 23.9, 23.8, 21.7.

MS: [M+Na]⁺_{calcd}=507.2, [M+Na]⁺_{found}=507.2.

Compound S10

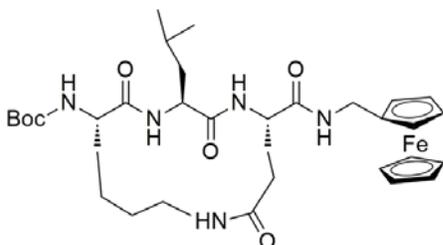


Compound **S9** (125 mg, 0.258 mmol) was dissolved in THF (2.6 mL) and methanol (750 μ L). NaOH (15 mg, 0.387 mmol) was dissolved in H₂O (250 μ L) and added to the acid, and the reaction stirred at rt for 17 h. The solvent was removed *in vacuo* and the residue redissolved in EtOAc (25 mL) and H₂O (25 mL). The pH was adjusted to pH 2 and the organic layer separated and washed with brine (25 mL), and dried over MgSO₄. The solvent was removed *in vacuo* to reveal a white solid (95 mg, 78%).

¹H NMR (500 MHz, DMSO-d₆): δ 8.15-8.01 (m, 2H, 2x NH), 7.37 (br s, 1H, NH), 6.43 (d, 1H, NH, $J=7.2$ Hz), 4.37-4.29 (m, 2H, 2x CaH), 4.06 (dd, 1H, CaH, $J=14.1, 7.0$ Hz), 3.30 (m, 1H, CHHNH), 2.73 (m, 1H, CHHNH), 2.28-1.06 (m, 13H, 6x CH₂, CH), 1.36 (s, 9H, Boc), 0.89-0.83 (m, 6H, 2xCH₃ Leu).

MS: [M+Na]⁺_{calcd}=493.2, [M+Na]⁺_{found}=493.2.

Peptide 5



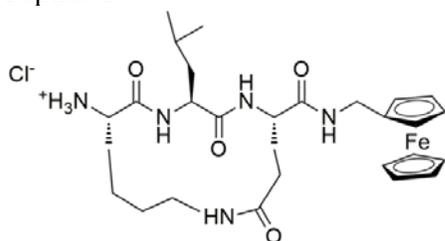
Compound **S10** (95 mg, 0.202 mmol) and ferrocenylmethylamine were dissolved in anhydrous DMF (4 mL) and stirred at rt under an N₂ atmosphere. HATU (84 mg, 0.222 mmol), HOBt (27 mg, 0.202 mmol) and DIPEA (140 μ L) were added, and the mixture stirred for 48 h. The solvent was removed and the residue taken up in EtOAc (25 mL) and H₂O (25 mL). The pH was adjusted to pH 3 and the organic layer separated and washed with NaHCO₃ (25 mL), brine (25 mL) and dried over Na₂SO₄. The solvent was removed *in vacuo* to yield a brown solid (98 mg, 73%).

¹H NMR (600 MHz, DMSO-d₆): δ 8.24-8.19 (m, 2H, 2x NH), 7.63 (t, 1H, NH, $J=5.8$ Hz), 7.49 (br s, 1H, NH), 6.37 (d, 1H, NH, $J=7.4$ Hz), 4.32 (m, 1H, CaH), 4.20 (m, 1H, CaH), 4.18-4.05 (m, 9H, Cp), 4.09 (m, 1H, CaH), 3.98 (d, 2H, CH₂Fc, $J=5.7$ Hz), 3.32 (m, 1H, CHHNH), 2.75 (m, 1H, CHHNH), 2.28-1.01 (m, 13H, 6xCH₂, CH), 1.36 (s, 9H, Boc), 0.89-0.83 (m, 6H, 2xCH₃ Leu).

¹³C NMR (150 MHz, DMSO-d₆): δ 172.8, 172.3, 172.0, 170.9, 155.0, 86.3, 78.3, 73.4, 70.7, 69.8, 69.3, 69.2, 68.8, 68.7, 67.98, 67.93, 67.6, 67.5, 60.1, 53.5, 52.4, 52.2, 46.3, 40.9, 38.0, 31.5, 30.7, 28.5, 26.3, 24.4, 23.3, 22.2.

MS: [M+Na]⁺_{calcd}=690.3, [M+Na]⁺_{found}=690.3.

Peptide 1



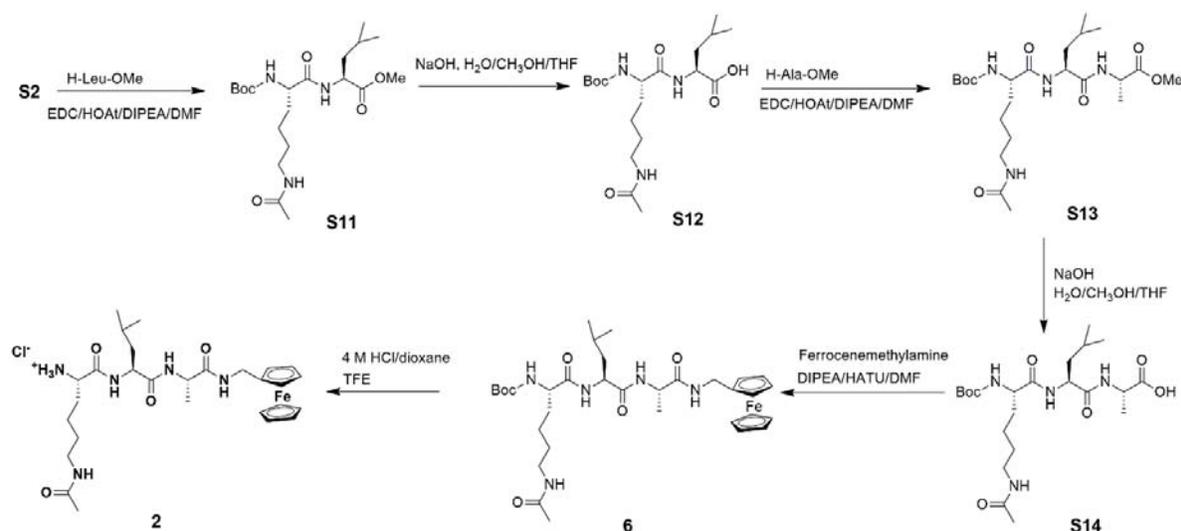
Peptide **5** (86 mg, 0.129 mmol) was dissolved in TFE (2 mL) and 4M HCl in dioxane (1 mL) added dropwise. The reaction was stirred at rt for 25 min. The solvent was removed *in vacuo* to reveal a brown solid. The crude product was purified using reverse phase HPLC to yield a sandy brown solid (15 mg, 21%).

¹H NMR (600 MHz, DMSO-d₆): δ 8.54 (d, 1H, NH, *J*=7.9 Hz), 8.36 (d, 1H, NH, *J*=7.6 Hz), 8.07 (br s, 3H, NH), 7.79 (br s, 1H, NH), 7.55 (m, 1H, NH), 4.41-4.34 (m, 2H, 2 x CaH), 4.19-3.93 (m, 9H, Cp), 4.07 (br s, 2H, CH₂Fc), 3.88 (br s, 1H, CaH), 3.28 (m, 1H, CHHNH), 2.79 (m, 1H, CHHNH), 2.32-1.05 (m, 13H, 6 x CH₂, CH), 0.90-0.87 (dd, 6H, (CH₃)₂Leu, *J*=9.3, 6.6 Hz).

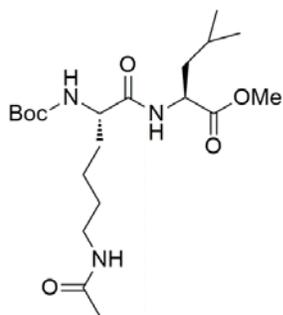
¹³C NMR (150 MHz, DMSO-d₆): δ 171.63, 171.29, 170.68, 168.36, 157.88, 70.92, 69.38, 68.92, 68.37, 67.33, 67.14, 51.81, 51.77, 51.58, 45.45, 41.49, 40.04, 37.45, 36.69, 30.19, 28.76, 25.20, 24.26, 23.98, 22.99, 22.63, 21.86, 19.50.

HRMS (*m/z*): [M+H]⁺ _{calcd}=568.2586, _{found}=568.2582.

Scheme S3. The final synthetic steps for linear β -strand peptides



Compound S11

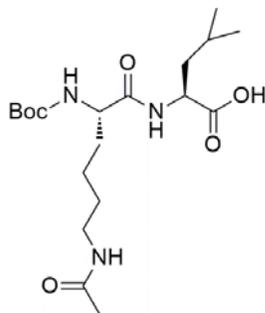


Compound **S2** (367 mg, 1.27 mmol) was dissolved in anhydrous DCM (10 mL). H-Leu-OMe (275 mg, 1.52 mmol) was added and stirred at rt under an N₂ atmosphere. Anhydrous DIPEA (883 μ L) was added, followed by EDC·HCl (267 mg, 1.40 mmol) and HOAt (173 mg, 1.27 mmol), and the reaction mixture stirred for 24 h. DCM (20 mL) and H₂O (30 mL) were added and the pH adjusted to pH 2-3. The organic layer was collected, washed with brine (30 mL), and dried over MgSO₄. The solvent was removed *in vacuo* to yield pale golden oil (373 mg, 71%).

¹H NMR (300 MHz, d-DMSO) δ 8.12 (d, 1H, NH, $J=7.6$ Hz), δ 7.79 (br s, 1H, NH), δ 6.80 (d, 1H, NH, $J=8.2$ Hz), δ 4.29 (m, 1H, C α H), δ 3.89 (m, 1H, C α H), δ 3.60 (s, 3H, OCH₃), δ 3.02-2.85 (m, 2H, CH₂NH), δ 1.78 (s, 3H, CH₃), δ 1.66-1.23 (m, 9H, 4xCH₂, CH), δ 1.37 (s, 9H, Boc), δ 0.90-0.81 (m, 6H, (CH₃)₂Leu).

MS: [M+Na]⁺_{calcd}=438.2, [M+Na]⁺_{found}=438.2.

Compound S12

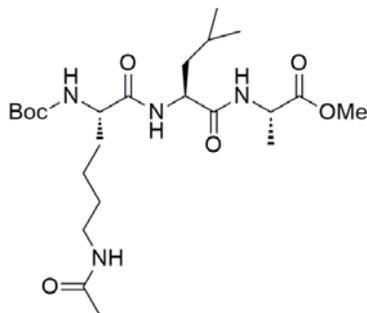


Compound **S11** (357 mg, 0.86 mmol) was dissolved in THF (2.5 mL) and methanol (714 μ L) and the mixture stirred. NaOH (52 mg, 1.29 mmol) was dissolved in H₂O (714 μ L) and this solution was added to the acid and stirred at rt for 31 h. The THF was removed *in vacuo* and the residue dissolved in EtOAc (25 mL) and H₂O (25 mL). The pH was adjusted to pH 2-3 and the organic layer collected and washed with brine (25 mL), and dried over MgSO₄. The solvent was removed *in vacuo* to reveal a clear solid (294 mg, 85%).

¹H NMR (300 MHz, DMSO-d₆): δ 7.94 (d, 1H, NH, $J=7.8$ Hz), 7.77 (t, 1H, NH, $J=9.0$ Hz), 6.79 (d, 1H, NH, $J=8.2$ Hz), 4.22 (dd, 1H, C α H, $J=13.9, 8.5$ Hz), 3.88 (d, 1H, C α H, $J=4.8$ Hz), 3.00-2.85 (m, 2H, CH₂NH), 1.77 (s, 3H, CH₃), 1.70-1.23 (m, 9H, 4xCH₂, CH), 1.36 (s, 9H, Boc), 0.89-0.81 (m, 6H, 2xCH₃Leu).

MS: [M+Na]⁺_{calcd}=400.2, [M+Na]⁺_{found}=400.2.

Compound S13

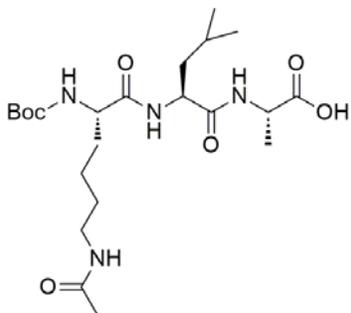


Compound **S12** (275 mg, 0.69 mmol) and HCl·H₂N-Ala-OMe (105 mg, 0.754 mmol) were dissolved in anhydrous DMF (11.5 mL) and stirred at rt under an N₂ atmosphere. Anhydrous DIPEA (477 μ L), HATU (287 mg, 0.754 mmol) and HOAt (93 mg, 0.686 mmol) were added and the reaction stirred for 25 h. The solvent was removed and the residue dissolved in EtOAc (50 mL) and H₂O (50 mL). The pH was adjusted to pH 3 and the organic layer washed with NaHCO₃ (50 mL) and brine (50 mL), before being dried over MgSO₄. The solvent was removed *in vacuo* to yield a pale golden oil (265 mg, 79%).

¹H NMR (300 MHz, DMSO-d₆): δ 8.36 (d, 1H, NH, $J=6.8$ Hz), 7.77 (t, 1H, NH, $J=9.0$ Hz), 7.72 (d, 1H, NH, $J=8.2$ Hz), 6.87 (d, 1H, NH, $J=8.1$ Hz), 4.35 (d, 1H, C α H, $J=7.6$ Hz), 4.23 (m, 1H, C α H), 3.87 (m, 1H, C α H), 3.60 (s, 3H, OCH₃), 3.03-2.90 (m, 2H, CH₂NH), 1.77 (s, 3H, CH₃), 1.70-1.34 (m, 9H, 4xCH₂, CH), 1.37 (s, 9H, Boc), 1.27 (d, 3H, CH₃, Ala, $J=7.3$ Hz), 0.89-0.81 (m, 6H, 2xCH₃Leu).

MS: [M+Na]⁺_{calcd}=509.3, [M+Na]⁺_{found}=509.3.

Compound S14

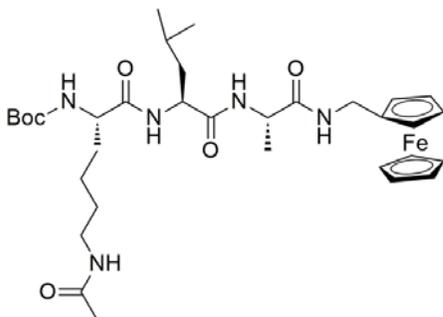


Compound **S13** (246 mg, 0.51 mmol) was dissolved in THF (1.72 mL) and methanol (492 μ L), and the mixture stirred. NaOH (30 mg, 0.75 mmol) was dissolved in H₂O (492 μ L) and this solution was added to the acid and stirred at rt for 27 h. The THF was removed *in vacuo* and the residue dissolved in EtOAc (35 mL) and H₂O (35 mL). The pH was adjusted to pH 2 and the organic layer collected and washed with brine (35 mL), and dried over MgSO₄. The solvent was removed *in vacuo* to reveal clear oil (189 mg, 79%).

¹H NMR (300 MHz, DMSO-*d*₆): δ 8.19 (d, 1H, NH, *J*=7.0 Hz), 7.78 (t, 1H, NH, *J*=9.9 Hz), 7.72 (d, 1H, NH, *J*=8.3 Hz), 6.88 (d, 1H, NH, *J*=8.1 Hz), 4.35 (dd, 1H, CaH, *J*=15.5, 7.7 Hz), 4.16 (m, 1H, CaH), 3.86 (d, 1H, CaH, *J*=4.7 Hz), 3.03-2.83 (m, 2H, CH₂NH), 1.77 (s, 3H, CH₃), 1.70-1.34 (m, 9H, 4xCH₂, CH), 1.37 (s, 9H, Boc), 1.25 (d, 3H, CH₃Ala, *J*=7.3 Hz), 0.89-0.82 (m, 6H, 2xCH₃ Leu).

MS: [M+Na]⁺_{calcd}=495.2, [M+Na]⁺_{found}=495.2.

Peptide 6



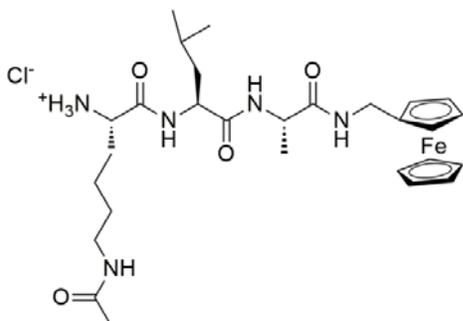
Compound **S14** (164 mg, 0.347 mmol) and ferrocenylmethylamine (82 mg, 0.382 mmol) were dissolved in anhydrous DMF (5.8 mL) and stirred at rt under an N₂ atmosphere. Anhydrous DIPEA (242 μ L), HATU (145 mg, 0.382 mmol) and HOAt (47 mg, 0.347 mmol) were added and the reaction stirred for 66 h. The solvent was removed and the residue dissolved in EtOAc (35 mL) and H₂O (35 mL). The pH was adjusted to pH 2 and the organic layer washed with NaHCO₃ (35 mL) and brine (35 mL), before being dried over MgSO₄. The solvent was removed *in vacuo* to yield a brown solid (192 mg, 83%).

¹H NMR (300 MHz, DMSO-*d*₆): δ 8.05-7.95 (m, 2H, 2xNH), 7.86-7.76 (m, 2H, 2xNH), 6.93 (d, 1H, NH, *J*=7.5 Hz), 4.35-3.82 (m, 14H, CH₂Fc, Cp, 3xCaH), 3.02-2.92 (m, 2H, CH₂NH), 1.77 (s, 3H, CH₃), 1.70-1.34 (m, 9H, 4xCH₂, CH), 1.37 (s, 9H, Boc), 1.22 (d, 3H, CH₃Ala, *J*= 8.1 Hz), 0.89-0.82 (m, 6H, (CH₃)₂ Leu).

^{13}C NMR (150 MHz, DMSO- d_6): 172.1, 171.5, 171.4, 168.5, 155.3, 123.1, 122.6, 115.5, 86.0, 78.0, 77.3, 68.9, 68.35, 68.30, 67.5, 67.3, 54.4, 50.7, 48.0, 40.8, 38.6, 37.4, 31.5, 31.3, 30.6, 28.8, 28.2, 28.1, 23.9, 23.1, 22.6, 21.4.

MS: $[\text{M}+\text{Na}]^+$ $_{\text{calcd}}=692.3$, $[\text{M}+\text{Na}]^+$ $_{\text{found}}=692.3$.

Peptide 2



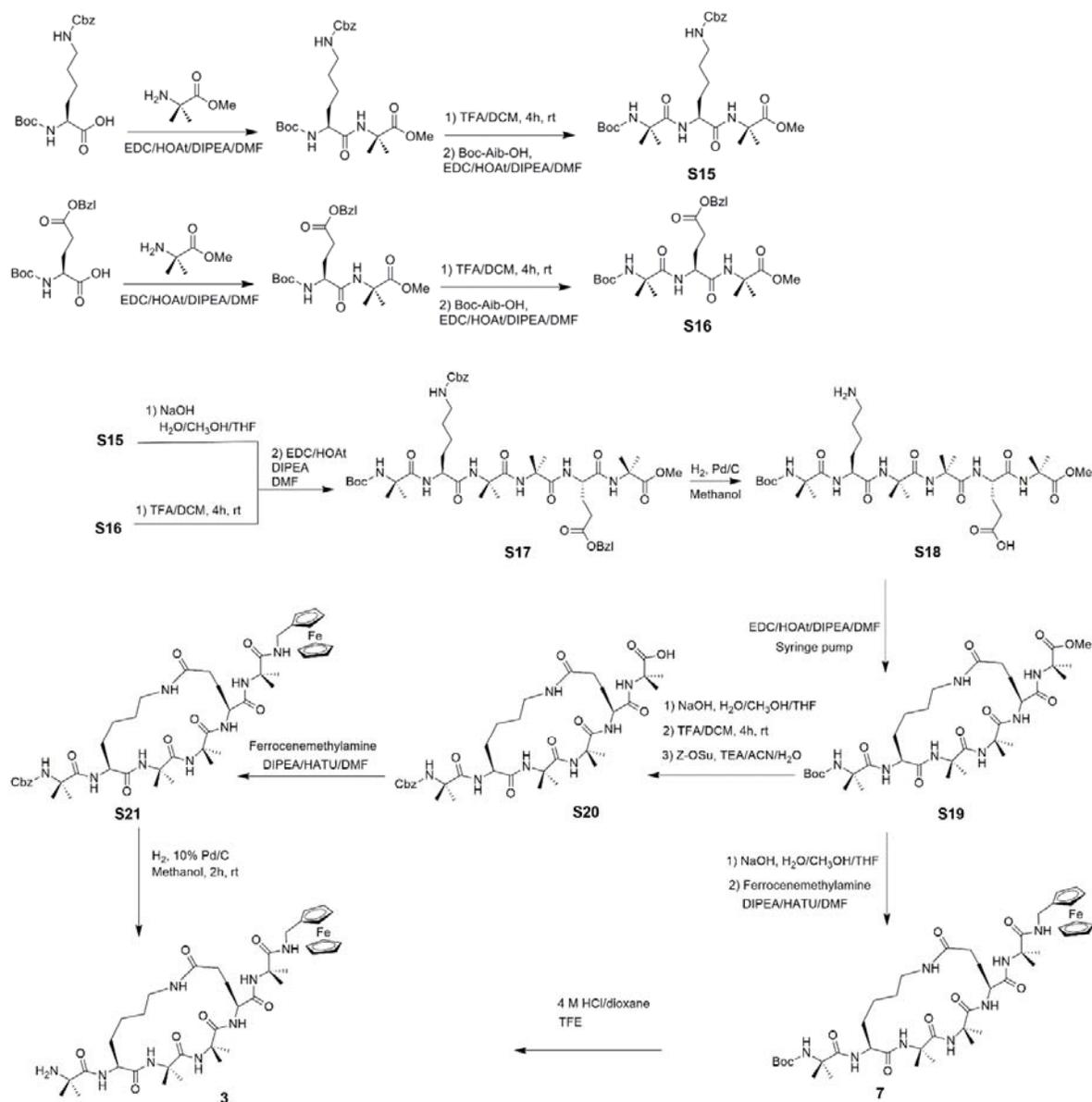
Peptide **6** (120 mg, 0.179 mmol) was dissolved in TFE (4 mL) and 4M HCl in dioxane (3 mL) added dropwise. The solution was stirred for 25 min., the solvent removed *in vacuo* and the residue washed with MeOH (2x10 mL). The crude product was purified using reverse phase HPLC to yield a sandy brown solid (25 mg, 25%).

^1H NMR (600 MHz, DMSO- d_6): δ 8.48 (d, 1H, NH, $J=8.1$ Hz), 8.16 (d, 1H, NH, $J=7.3$ Hz), 8.07 (d, 3H, NH, $J=4.0$ Hz), 8.01 (br s, 1H, NH), 7.77 (t, 1H, NH, $J=5.4$ Hz), 4.39 (dd, 1H, C α H, $J=14.1, 8.8$ Hz), 4.31 (m, 1H, C α H), δ 4.20-3.94 (m, 11H, Cp, CH₂Fc), 3.76 (dd, 1H, C α H, $J=11.2, 5.7$ Hz), 2.99 (dd, 2H, CH₂NH, $J=13.1, 6.8$ Hz), 1.78 (s, 3H, CH₃), 1.71-1.61 (m, 3H, CH₂, CH), 1.49-1.45 (m, 2H, CH₂), 1.39-1.35 (dd, 2H, CH₂, $J=14.3, 7.1$ Hz), 1.32-1.26 (dt, 2H, CH₂, $J=14.9, 7.2$ Hz), 1.22 (d, 3H, CH₃, $J=6.3$ Hz), 0.90-0.86 (dd, 6H, (CH₃)₂ Leu, $J=15.2, 6.6$ Hz).

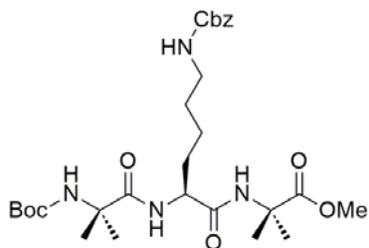
^{13}C NMR (150 MHz, DMSO- d_6): δ 171.8, 171.5, 169.3, 168.8, 158.3, 71.3, 69.8, 69.3, 68.8, 67.8, 67.5, 52.4, 51.5, 48.5, 41.2, 38.6, 37.8, 31.3, 24.4, 23.5, 23.0, 21.9, 18.8.

HRMS (m/z): $[\text{M}+\text{H}]^+$ $_{\text{calcd}}=570.2737$; $_{\text{found}}=570.2743$.

Scheme S4. The final synthetic steps for lactam-bridged 3₁₀-helical peptides.



Compound S15



Boc-Lys(Cbz)-OH (1.68 g, 4.4 mmol) and HCl·H₂N-Aib-OMe (880 mg, 4.4 mmol) were dissolved in anhydrous DMF (28 mL) and stirred at rt under an N₂ atmosphere. HOAt (780 mg, 5.7 mmol), EDC·HCl (1.09 g, 5.7 mmol) and DIPEA (3.50 mL, 20 mmol) were added, and the

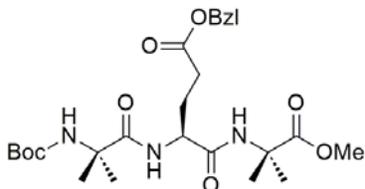
mixture stirred for 26 h. The solvent was removed and the residue taken up in EtOAc (200 mL) and H₂O (200 mL). The organic layer separated and washed with NaHCO₃ (200 mL), brine (200 mL) and dried over Na₂SO₄. The solvent was removed *in vacuo* and the crude product was purified by column chromatography (EtOAc: Petroleum ether 50:50 by v/v) to reveal the dipeptide Boc-Lys(Cbz)-Aib-OMe, a white solid (1.70 g, 81%).

The resulting was dissolved in DCM (10 mL) and TFA (10 mL) added dropwise. The reaction was stirred at rt for 3 h. The solvent was removed *in vacuo* to reveal brown oil. The oil and Boc-Aib-OH (880 mg, 4.4 mmol) were dissolved in anhydrous DMF (25 mL) and stirred at rt under an N₂ atmosphere. HOAt (884 mg, 6.5 mmol), EDC·HCl (1.24 g, 6.5 mmol) and DIPEA (3.5 mL, 20 mmol) were added, and the mixture stirred for 40 h. The solvent was removed and the residue taken up in EtOAc (200 mL) and H₂O (200 mL). The organic layer separated and washed with NaHCO₃ (200 mL), brine (200 mL) and dried over Na₂SO₄. The solvent was removed *in vacuo* and the crude product was purified by column chromatography (EtOAc: Petroleum ether 70:30 by v/v) to yield the product, a white solid (1.98 g, 80%).

¹H NMR (300 MHz, DMSO-d₆): δ 8.04 (s, 1H, NH), 7.35-7.25 (m, 6H, NH, benzene), 7.16 (t, 1H, NHCH₂), 7.08 (br s, 1H, NH), 4.96 (s, 2H, OCH₂), 4.09 (m, 1H, CaH), 3.51 (s, 3H, OCH₃), 2.92 (m, 2H, NHCH₂), 1.76 (s, 3H, COCH₃), 1.65-1.45 (m, 2H, CH₂), 1.40-1.10 (m, 25H, 2xCH₂, Boc, 4xCH₃).

MS: [M+H]⁺_{calcd}=565.3, [M+H]⁺_{found}=565.3.

Compound S16



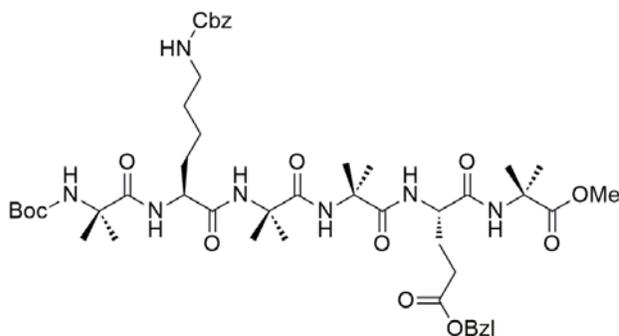
Boc-Glu(OBzl)-OH (1.68 g, 5 mmol) and HCl·H₂N-Aib-OMe (1.00 g, 5 mmol) were dissolved in anhydrous DMF (28 mL) and stirred at rt under an N₂ atmosphere. HOAt (884 mg, 6.50 mmol), EDC·HCl (1.24 g, 6.50 mmol) and DIPEA (3.50 mL, 20 mmol) were added, and the mixture stirred for 26 h. The solvent was removed and the residue taken up in EtOAc (200 mL) and H₂O (200 mL). The organic layer separated and washed with NaHCO₃ (200 mL), brine (200 mL) and dried over Na₂SO₄. The solvent was removed *in vacuo* and the crude product was purified by column chromatography (EtOAc: Petroleum ether 50:50 by v/v) to reveal the dipeptide Boc-Glu(OBzl)-Aib-OMe, a white solid (1.98 g, 92%).

Boc-Glu(OBzl)-Aib-OMe (453 mg, 1.04 mmol) was dissolved in DCM (5 mL) and TFA (5 mL) added dropwise. The reaction was stirred at rt for 3 h. The solvent was removed *in vacuo* to reveal brown oil. The resulting oil and Boc-Aib-OH (210 mg, 1.04 mmol) were dissolved in anhydrous DMF (2 mL) and stirred at rt under an N₂ atmosphere. HOBt (207 mg, 1.35 mmol), EDC·HCl (260 mg, 1.35 mmol) and DIPEA (0.75 mL, 4.16 mmol) were added, and the mixture stirred for 40 h. The solvent was removed and the residue taken up in EtOAc (50 mL) and H₂O (50 mL). The organic layer separated and washed with NaHCO₃ (50 mL), brine (50 mL) and dried over Na₂SO₄. The solvent was removed *in vacuo* and the crude product was purified by column chromatography (EtOAc: Petroleum ether 70:30 by v/v) to yield the product, a white solid (430 mg, 80%).

^1H NMR (300 MHz, DMSO- d_6): δ 8.15 (s, 1H, NH), 7.50 (d, 1H, NH, $J=6.0$ Hz), 7.42-7.25 (m, 5H, benzene), 7.16 (s, 1H, NH), 5.08 (s, 2H, OCH₂), 4.17 (m, 1H, C α H), 3.52 (s, 3H, OCH₃), 2.35 (t, 2H, CH₂), 1.98 (m, 1H, CHH), 1.83 (m, 1H, CHH), 1.45-1.20 (m, 12H, 4xCH₃).

MS: $[\text{M}+\text{H}]^+$ _{calcd}=522.3, $[\text{M}+\text{H}]^+$ _{found}=522.3.

Compound S17



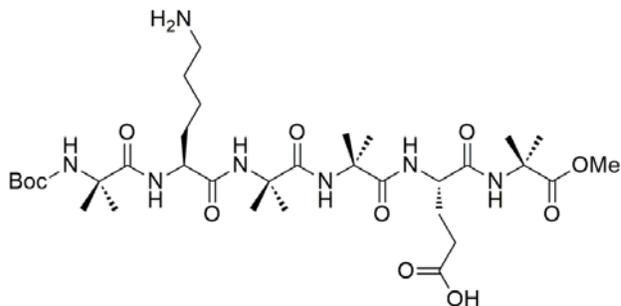
Compound **S15** (2.23 g, 4.0 mmol) was dissolved in THF (12.0 mL) and methanol (8.0 mL). 4.0 mL of NaOH solution (1.6 M, aqueous) was added to the mixture, and the reaction stirred at rt for 18 h. The solvent was removed *in vacuo* and the residue redissolved in EtOAc (200 mL) and H₂O (200 mL). The pH was adjusted to pH 3 and the organic layer separated and washed with brine (200 mL), and dried over MgSO₄. The solvent was removed *in vacuo* to reveal a white solid (2.29 g).

Compound **S16** (2.08 g, 4.0 mmol) was dissolved in DCM (5 mL) and TFA (5 mL) added dropwise. The reaction was stirred at rt for 3 h. The solvent was removed *in vacuo* to reveal brown oil. The resulting oil and the above hydrolysed compound **S15** (2.29 g) were dissolved in anhydrous DMF (28 mL) and stirred at rt under an N₂ atmosphere. HOAt (707 mg, 5.2 mmol), EDC·HCl (1.0 g, 5.2 mmol) and DIPEA (2.8 mL, 16 mmol) were added, and the mixture stirred for 40 h. The solvent was removed and the residue taken up in EtOAc (200 mL) and H₂O (200 mL). The organic layer separated and washed with NaHCO₃ (200 mL), brine (200 mL) and dried over Na₂SO₄. The solvent was removed *in vacuo* and the crude product was purified by column chromatography (EtOAc: Petroleum ether 90:10 by v/v) to yield the title product, a white solid (2.10 g, 55%).

^1H NMR (300 MHz, DMSO- d_6): δ 8.32-8.06 (m, 2H, 2xNH), 7.93-7.14 (m, 15H, 5xNH, 2x benzene), 5.06 (s, 2H, OCH₂), 4.98 (s, 2H, OCH₂), 4.05 (m, 1H, C α H), 3.94-3.67 (m, 1H, C α H), 3.53 (s, 3H, OCH₃), 2.95 (m, 2H, CH₂NH), 2.44 (m, 2H, CH₂), 2.14-1.22 (m, 41H, 4xCH₂, 8xCH₃, Boc).

MS: $[\text{M}+\text{H}]^+$ _{calcd}=954.5, $[\text{M}+\text{H}]^+$ _{found}=954.5.

Compound S18

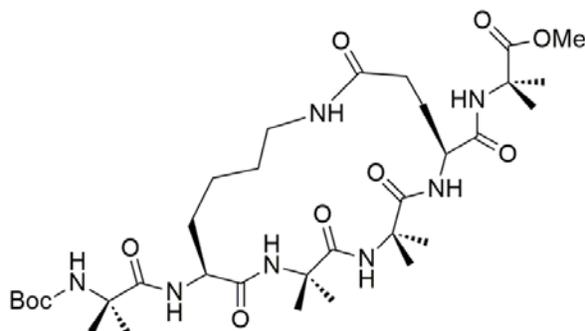


Compound **S17** (1.73 g, 1.81 mmol) was dissolved in (methanol over molecular sieves, 20 mL). Pd/C (15% w/w, 260 mg) was added and the mixture stirred. A H₂ balloon was fitted under vacuum and the solution stirred at rt for a further 18 h. The solution was filtered through celite and washed with methanol (3x 20 mL), and the solvent removed *in vacuo* to yield a white solid (1.26 g, 95%).

¹H NMR (300 MHz, DMSO-d₆): δ 8.60 (br s, 1H, NH), 8.29 (br s, 1H, NH), 8.19 (br s, 1H, NH), 7.65 (br s, 1H, NH), 7.56 (br s, 1H, NH), 7.47 (br s, 1H, NH), 3.90 (m, 1H, CαH), 3.74 (m, 1H, CαH), 3.55 (s, 3H, OCH₃), 2.75-1.52 (m, 12H, 6xCH₂), 1.41-1.30 (m, 24H, 8xCH₃), 1.34 (s, 9H, Boc).

MS: [M+H]⁺_{calcd}=730.4, [M+H]⁺_{found}=730.4.

Compound S19



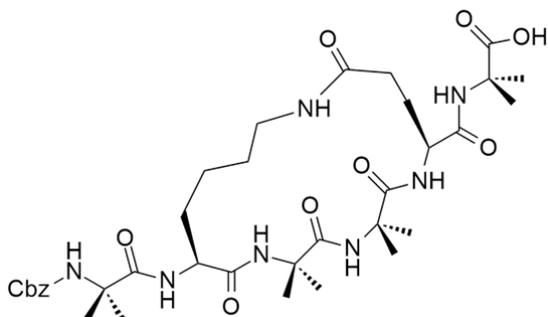
EDC·HCl (815 mg, 4.27 mmol), HOBt (576 mg, 4.27 mmol) and DIPEA (1.5 mL) were dissolved in anhydrous DMF (255 mL), and the solution stirred at rt under an N₂ atmosphere. Compound **S18** (622 mg, 0.853 mmol) was dissolved in anhydrous DMF (25 mL), placed into a syringe and pumped into the solution containing the coupling agents at the rate of 20 μL/min, and stirred for 68 h. The solvent was removed and the residue taken up in EtOAc (100 mL) and H₂O (100 mL). The pH was adjusted to pH 3, the organic layer separated and washed with NaHCO₃ (100 mL), brine (100 mL) and dried over Na₂SO₄. The solvent was removed *in vacuo* to give the crude product, which was purified using reverse phase HPLC, to yield a white solid (90 mg, 15%).

¹H NMR (600 MHz, DMSO-d₆): δ 8.33 (br s, 1H, NH), 8.04 (br s, 1H, NH), 7.79-7.70 (m, 2H, 2x NH), 7.63 (m, 1H, NH), 7.53 (s, 1H, NH), 7.45 (d, 1H, NH, J=8.0 Hz), 3.80-3.73 (m, 2H, 2x CαH), 3.54 (s, 3H, OCH₃), 3.07 (m, 1H, CHHNH), 2.90 (m, 1H, CHHNH), 2.15-1.22 (m, 10H, 5x CH₂), 1.41-1.27 (m, 24H, (CH₃)₈), 1.34 (s, 9H, Boc).

^{13}C NMR (150 MHz, DMSO-d_6): δ 175.6, 174.8, 174.7, 174.6, 172.9, 171.6, 171.5, 158.7, 158.5, 79.3, 79.2, 56.7, 56.6, 56.2, 55.3, 55.2, 52.1, 40.3, 28.6, 26.0, 25.2, 25.1.

MS: $[\text{M}+\text{H}]^+_{\text{calcd}}=712.4$, $[\text{M}+\text{H}]^+_{\text{found}}=712.4$.

Compound S20



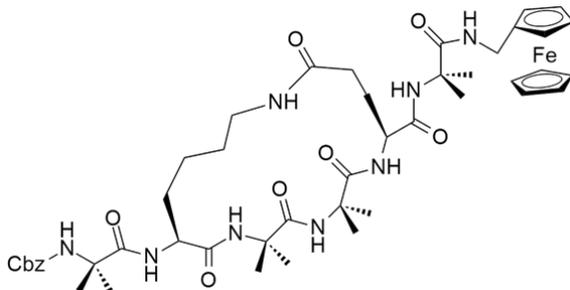
Compound **S19** (380 mg, 0.53mmol) was dissolved in THF (3.80 mL) and H_2O (3.80 mL). NaOH (85 mg) solid was added and the reaction stirred at rt for 2 h. The reaction was quenched by 1M HCl (2 mL) aqueous solution. The solvent was removed *in vacuo* to reveal a white solid (514 mg). The resulting intermediate (190 mg, 0.27 mmol) was dissolved in DCM (5 mL) and TFA (2 mL) added dropwise. The reaction was stirred at rt overnight. The solvent was removed *in vacuo* to reveal a brown oil (350 mg).

The resulting oil and triethylamine (150 μL) were dissolved in a solvent mixture of water (5 mL) and acetonitrile (3 mL). A solution of Z-OSu (82 mg in 2 mL acetonitrile) was added. The reaction mixture was stirred overnight at rt, after which the volatiles were removed under reduced pressure. The residue was redissolved in MeOH (5 mL), and purified using reverse phase HPLC to yield a white solid (120 mg).

^1H NMR (600 MHz, DMSO-d_6): δ 8.01 (d, 1H, NH), 7.79 (s, 1H, NH), 7.77 (m, 1H, NH), 7.71 (t, 1H, NH), 7.61 (d, 1H, NH), 7.58 (s, 1H, NH), 7.40 (s, 1H, NH). 7.38-7.26 (m, 5H, aromH), 5.05 (dd, 2H, Cbz CH_2), 4.12 (m, 1H, $\text{C}\alpha\text{H}$), 3.72 (m, 1H, $\text{C}\alpha\text{H}$), 3.06 (m, 2H, CH_2NH), 2.04 (m, 1H, CHHCO), 1.92 (m, 1H, CHHCO), 1.85 (m, 1H, CHHCO), 1.54 (m, 1H, CHHCO), 1.48-1.09 (m, 26H, CH_2 , 8x CH_3), 1.22 (m, 2H, CH_2).

MS: $[\text{M}+\text{H}]^+_{\text{calcd}}=732.4$, $[\text{M}+\text{H}]^+_{\text{found}}=732.4$.

Compound S21



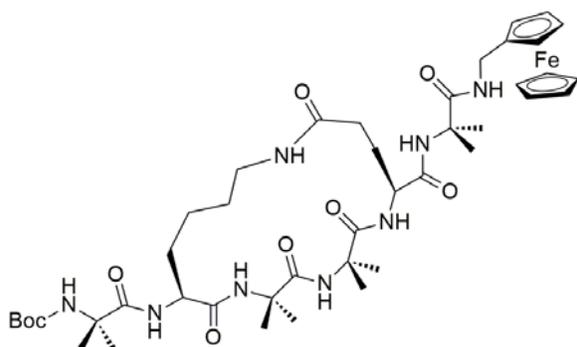
Compound **S20** (120 mg, 0.16mmol) and ferrocenylmethylamine (60 mg, 0.27mmol) were dissolved in anhydrous DMF (2 mL) and stirred at rt under an N_2 atmosphere. EDC·HCl (67 mg),

HOAt (50 mg) and DIPEA (200 μ L) were added, and the mixture stirred for 36 h. The solvent was removed and the residue redissolved in MeOH (15 mL), and purified using reverse phase HPLC to yield a sandy brown solid (90 mg).

^1H NMR (600 MHz, DMSO- d_6): δ 8.05 (d, 1H, NH), 7.84 (d, 1H, NH), 7.82 (s, 1H, NH), 7.78 (s, 1H, NH), 7.74 (s, 1H, NH), 7.72 (t, 1H, NH), 7.44 (s, 1H, NH), 7.39-7.28 (m, 5H, aromH), 7.26 (t, 1H, NH), 5.06(dd, 2H, CH_2), 4.25-3.90 (m, 12H, Cp, CH_2Fc , CaH), 3.70 (m, 1H, CaH), 3.18 (m, 2H, $\text{CH}_2\text{CH}_2\text{NH}$), 1.99 (m, 2H, CHHCO), 1.85 (m, 1H, CHHCO), 1.54 (m, 1H, CHHCO), 1.48-1.19 (m, 28H, 2x CH_2 , 8x CH_3).

MS: $[\text{M}+\text{H}]^+_{\text{calcd}}=929.9$, $[\text{M}+\text{H}]^+_{\text{found}}=929.9$.

Peptide 7



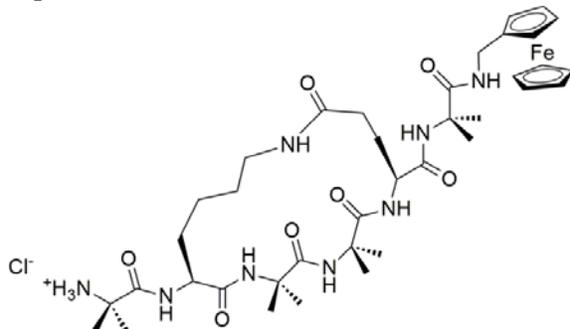
Compound **S19** (125 mg, 0.176 mmol) was dissolved in THF (1.25 mL) and H₂O (1.5 mL). NaOH aqueous solution (1.6 M, 352 μ L) was added and the reaction stirred at rt for 2 h. The solvent was removed *in vacuo* to reveal a white solid (157 mg, quant). The resulting intermediate (122 mg, 0.176 mmol) and ferrocenylmethylamine (75 mg, 0.352 mmol) were dissolved in anhydrous DMF (3.4 mL) and stirred at rt under an N₂ atmosphere. EDC·HCl (37 mg, 0.194 mmol), HOAt (24 mg, 0.176 mmol) and DIPEA (122 μ L) were added, and the mixture stirred for 36 h. The solvent was removed *in vacuo* and the residue redissolved in MeOH (15 mL), and purified using reverse phase HPLC to yield a sandy brown solid (40 mg, 29%).

^1H NMR (600 MHz, DMSO- d_6): δ 8.16 (br s, 1H, NH), 7.96 (s, 1H, NH), 7.86 (d, 1H, NH, $J=3.9$ Hz), 7.82 (s, 1H, NH), 7.70 (t, 1H, NH, $J=12.0$ Hz), 7.46 (s, 1H, NH), 7.35 (s, 1H, NH), 7.27 (br s, 1H, NH), 4.25-4.02 (m, 9H, Cp), 3.99-3.90 (m, 3H, CH_2Fc , CaH), 3.71 (m, 1H, CaH), 3.18 (m, 1H, CHHNH), 2.96 (m, 1H, CHHNH), 2.15-1.37 (m, 10H, 5x CH_2), 1.44-1.29 (m, 24H, 8 x CH_3), 1.34 (s, 9H, Boc).

^{13}C NMR (150 MHz, DMSO- d_6): 175.83, 175.37, 174.89, 173.61, 171.07, 170.75, 155.31, 86.55, 78.79, 68.66, 68.34, 66.99, 66.97, 66.92, 56.35, 56.29, 56.02, 55.70, 53.63, 38.12, 37.82, 33.63, 31.26, 31.02, 29.40, 28.97, 28.86, 28.66, 28.50, 28.19, 25.48, 24.55, 24.26, 22.96, 22.06.

HRMS: $[\text{M}]^+_{\text{calcd}}=894.4302$, $[\text{M}]^+_{\text{found}}=894.4303$.

Peptide 3



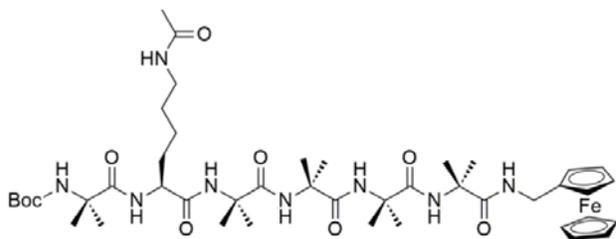
Peptide 7 (5 mg, 0.006 mmol) was dissolved in TFE (600 μ L) and 4M HCl in dioxane (200 μ L) added. The reaction was stirred at rt for 25 min. The solvent was removed *in vacuo* to reveal a light brown solid (quant). The crude product was purified using reverse phase HPLC to yield a sandy brown solid (2 mg, 45%).

¹H NMR (600 MHz, DMSO-d₆): δ 8.30 (s, 1H, NH), 8.08 (s, 1H, NH), 7.75 (d, 1H, NH), 7.72 (m, 1H, NH), 7.54 (s, 1H, NH), 7.49 (m, 1H, NH), 7.33 (s, 1H, NH), 7.18 (t, 1H, NH), 4.25-3.90 (m, 12H, Cp, CH₂Fc, C α H), 3.85 (m, 1H, C α H), 3.04-2.90 (m, 2H, CH₂NH), 2.30-1.80 (m, 4H, 4 x CHH), 1.80-1.10 (m, 28H, 8 x CH₃, 2 x CH₂).

¹³C NMR (150 MHz, DMSO-d₆): 173.66, 172.51, 172.26, 171.96, 169.20, 166.27, 149.91, 144.91, 105.60, 100.89, 85.08, 84.44, 78.30, 77.34, 68.80, 67.45, 68.34, 66.99, 66.97, 66.92, 56.38, 56.02, 55.70, 53.68, 44.99, 43.47, 38.72, 31.50, 29.46, 28.95, 28.53, 24.89.

HRMS (*m/z*): [M]⁺ _{calcd}=794.3778, _{found}=794.3778.

Peptide 8



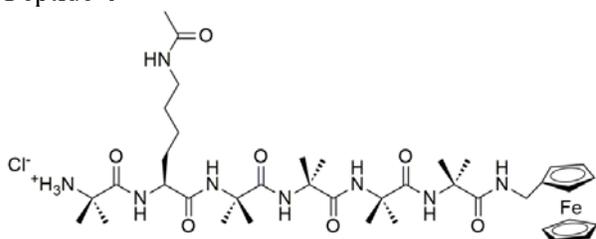
Peptide **S22** (410 mg, 0.57mmol) and ferrocenylmethylamine (140 mg, 0.65mmol) were dissolved in anhydrous DMF (8 mL). DIPEA (400 μ L, 4 equiv), HOAt (160 mg, 2 equiv) and HATU (430 mg, 2equiv) were added. Reaction mixture was stirred overnight under an N₂ atmosphere at rt. The solvent was removed *in vacuo* and the peptide purified using reverse phase HPLC.

¹H NMR (300 MHz, DMSO-d₆): δ 8.21 (s, 1H, NH), 7.99 (s, 1H, NH), 7.73 (t, 1H, NHCH₂), 7.61 (s, 1H, NH), 7.57 (s, 1H, NH), 7.47 (br s, 1H, NH), 7.28 (s, 1H, NH), 7.20 (s, 1H, NH), 4.31-3.40 (m, 12H, Cp, C α H, CH₂Fc), 2.95 (m, 2H, NHCH₂), 1.76 (s, 3H, COCH₃), 1.68 (m, 2H, CH₂), 1.45-1.20 (m, 43H, 2xCH₂, Boc, 10xCH₃).

¹³C NMR (150 MHz, DMSO-d₆): 175.59, 175.09, 175.08, 174.98, 174.82, 174.41, 173.73, 173.41, 172.80, 168.93, 158.29, 154.93, 78.59, 68.33, 66.94, 56.14, 56.03, 56.02, 55.63, 54.25, 42.69, 42.66, 42.62, 40.04, 38.24, 37.73, 35.12, 29.66, 28.91, 28.14, 25.39, 24.91, 24.70, 22.64, 22.57. .

MS: [M+H]⁺_{calcd}=911.5, [M+H]⁺_{found}=911.5.

Peptide 4



Peptide **8** (10 mg, 0.012 mmol) was dissolved in TFE (600 μ L) and 4M HCl in dioxane (200 μ L) added. The reaction was stirred at rt for 25 min. The solvent was removed *in vacuo* to reveal a light brown solid (quant). The crude product was purified using reverse phase HPLC to yield a sandy brown solid (4 mg, 45%).

¹H NMR (600 MHz, DMSO-d₆): δ 8.51 (s, 1H, NH), 8.13 (s, 1H, NH), 8.10 (s, 3H, NH₃), 7.84 (t, 1H, NHCH₂), 7.68 (s, 1H, NH), 7.63 (s, 1H, NH), 7.43 (m, 1H, NH), 7.34 (br s, 1H, NH), 4.55-3.50 (m, 12H, Cp, C α H, CH₂), 3.02 (m, 2H, NHCH₂), 1.76 (s, 3H, COCH₃), 1.68 (m, 2H, CH₂), 1.45-1.20 (m, 34H, 2 x CH₂, 10 x CH₃).

¹³C NMR (150 MHz, DMSO-d₆): δ 174.81, 174.35, 173.74, 173.47, 172.21, 171.81, 169.00, 128.89, 127.25, 121.35, 119.99, 109.72, 68.41, 66.90, 56.31, 56.10, 56.01, 55.93, 55.79, 53.26, 40.03, 38.30, 37.69, 30.45, 29.14, 28.87, 26.00, 25.38, 25.10, 24.97, 24.50, 24.20, 23.50, 23.37, 23.24, 22.60.

HRMS (*m/z*): [M]⁺_{calcd}=811.4164, _{found}=811.4162.

3. ROESY spectra for peptides 1-4 and 5-7

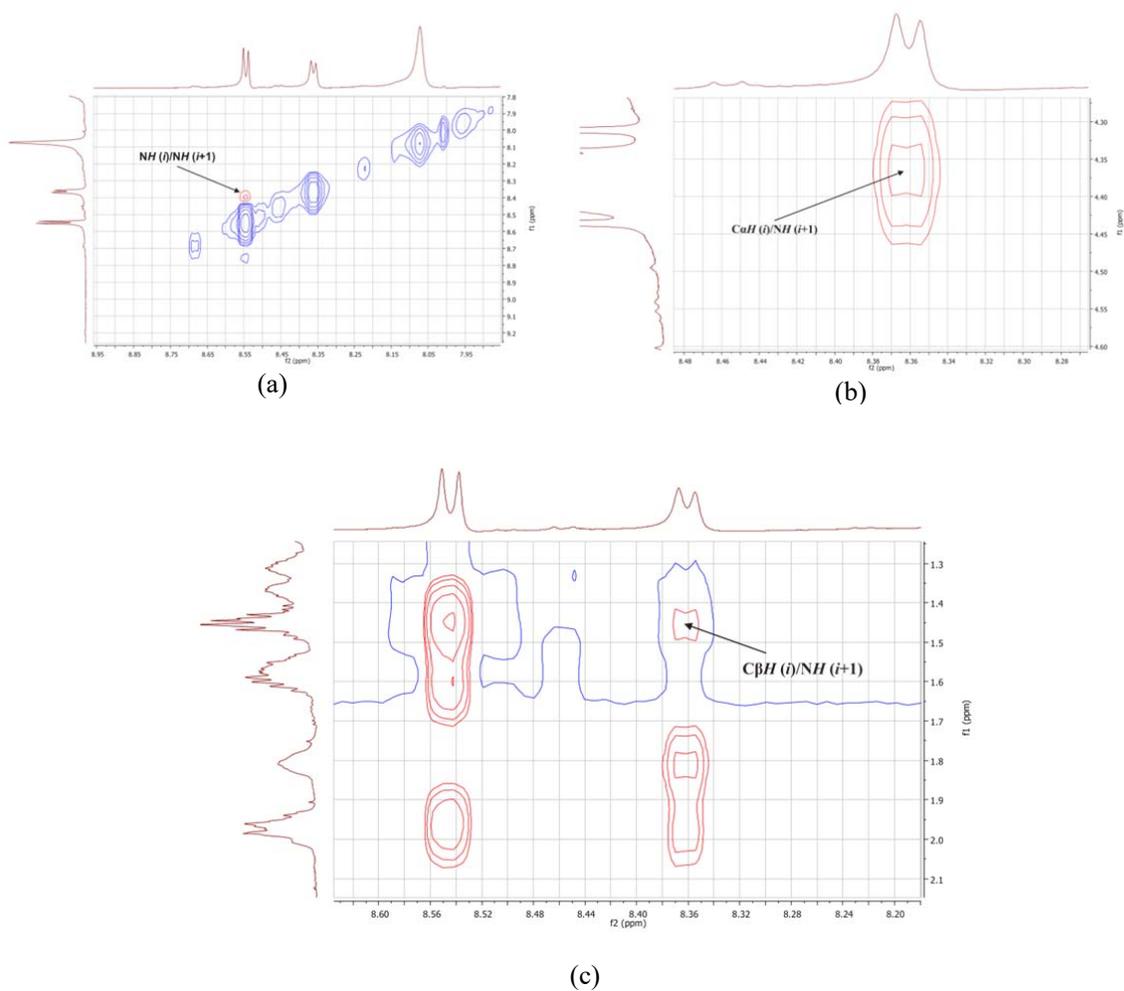


Figure S1. ROESY spectra of peptide 1, showing (a) $NH(i)$ to $NH(i+1)$, (b) $CaH(i)$ to $NH(i+1)$ and (c) $C\beta H(i)$ to $NH(i+1)$ correlations.

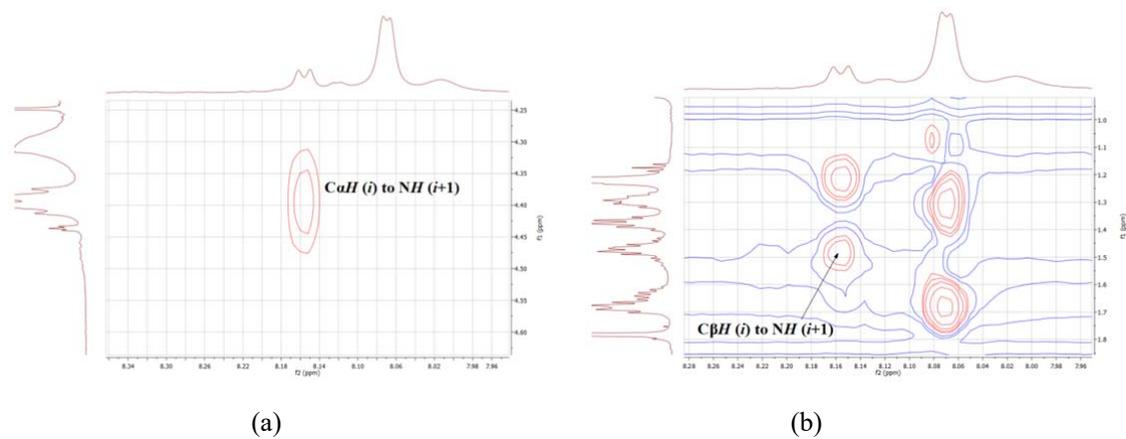


Figure S2. ROESY spectra of peptide 2, showing (a) $CaH(i)$ to $NH(i+1)$ and (b) $C\beta H(i)$ to $NH(i+1)$ correlations.

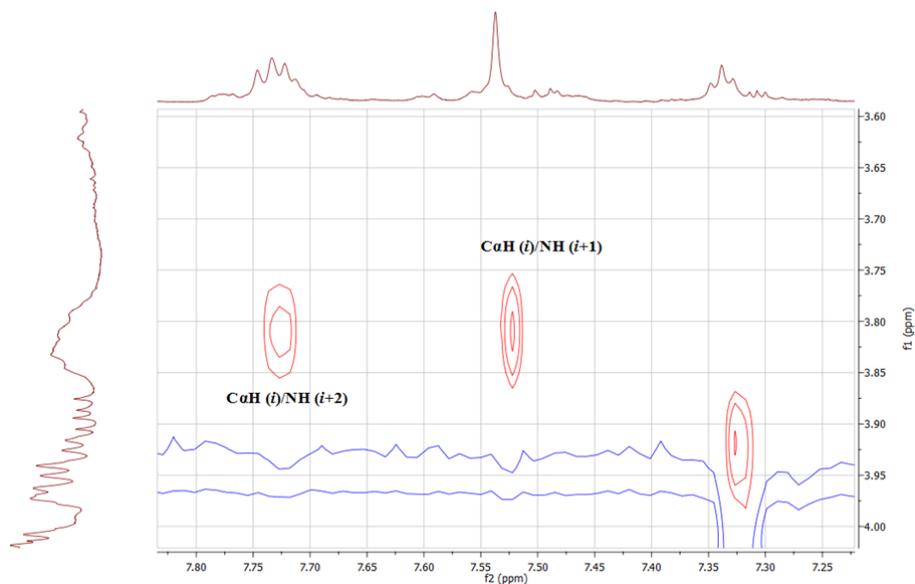


Figure S3. ROESY spectrum of peptide **3**, showing $\text{CaH}(i)$ to $\text{NH}(i+1)$, and $\text{CaH}(i)$ to $\text{NH}(i+2)$ correlations.

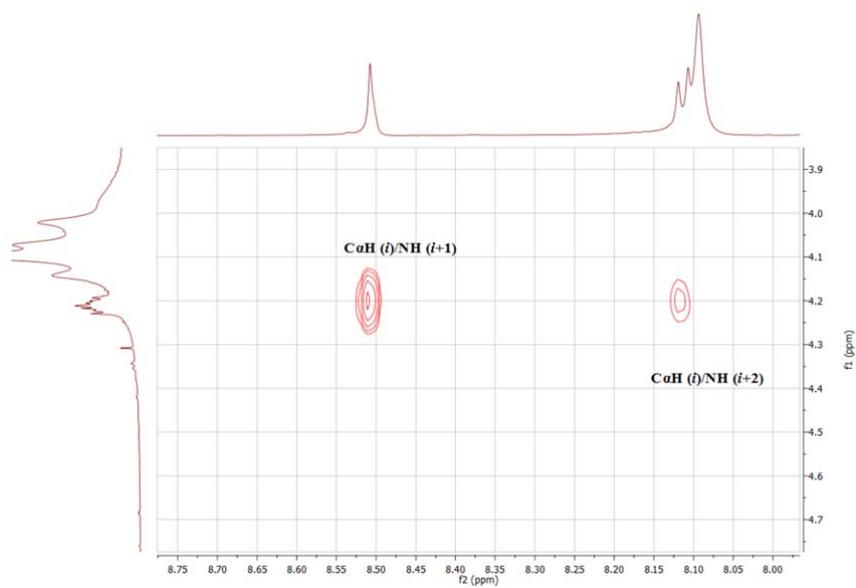
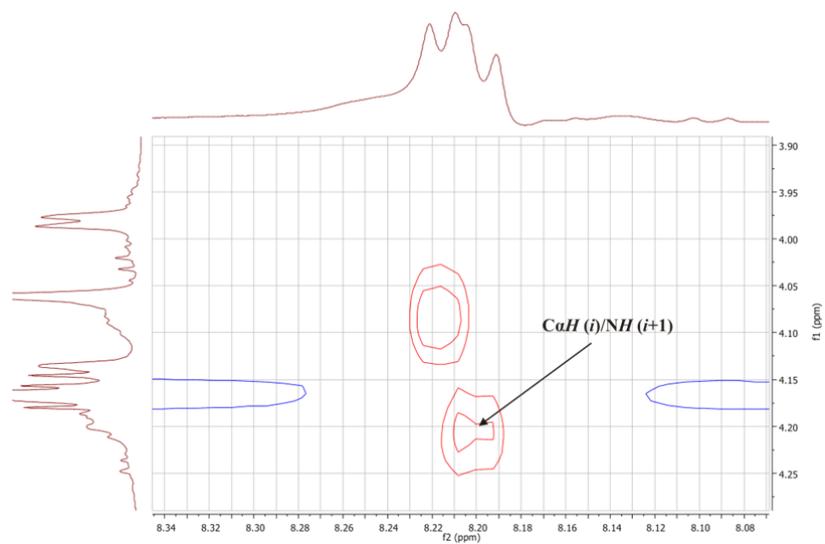
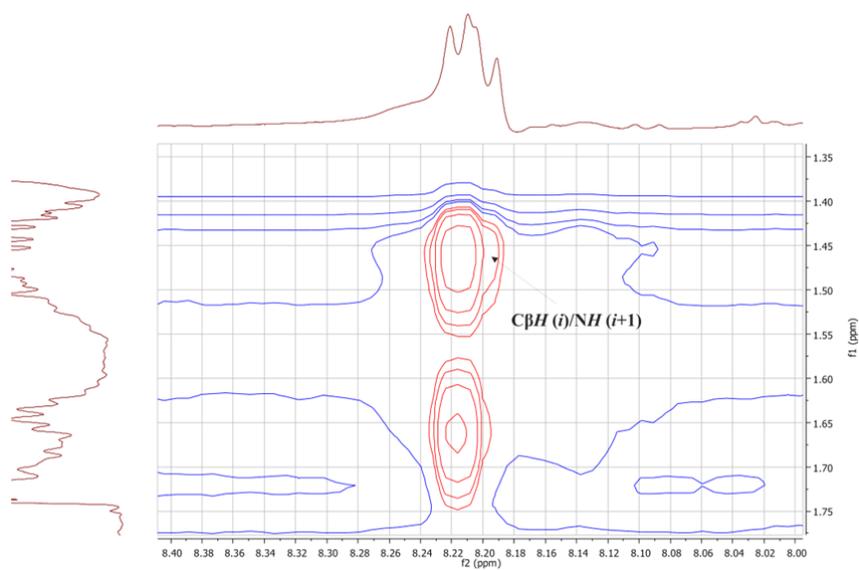


Figure S4. ROESY spectrum of peptide **4**, showing $\text{CaH}(i)$ to $\text{NH}(i+1)$, and $\text{CaH}(i)$ to $\text{NH}(i+2)$ correlations.



(a)



(b)

Figure S5. ROESY spectra of peptide **5**, showing (a) $C\alpha H(i)$ to $NH(i+1)$ and (b) $C\beta H(i)$ to $NH(i+1)$ correlations.

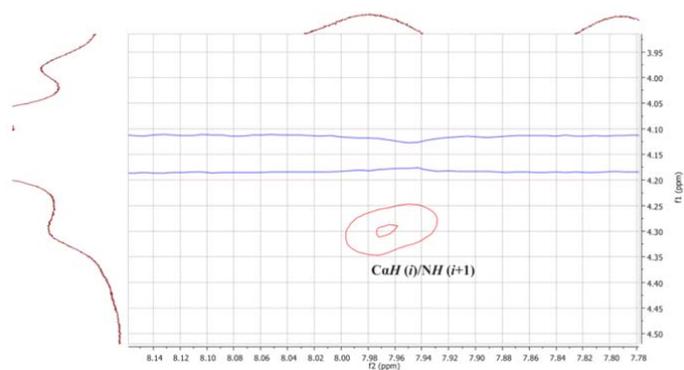


Figure S6. ROESY spectrum of peptide **6**, showing $CaH(i)$ to $NH(i+1)$ correlation.

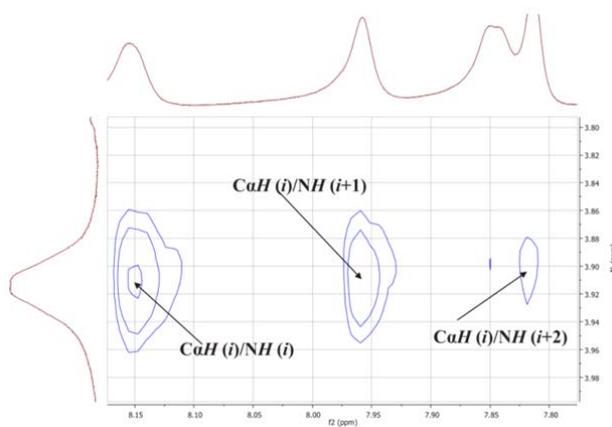


Figure S7. ROESY spectrum of peptide **7**, showing $CaH(i)$ to $NH(i+1)$ and medium range $CaH(i)$ to $NH(i+2)$ correlations.

4. IR spectrum for peptide 7

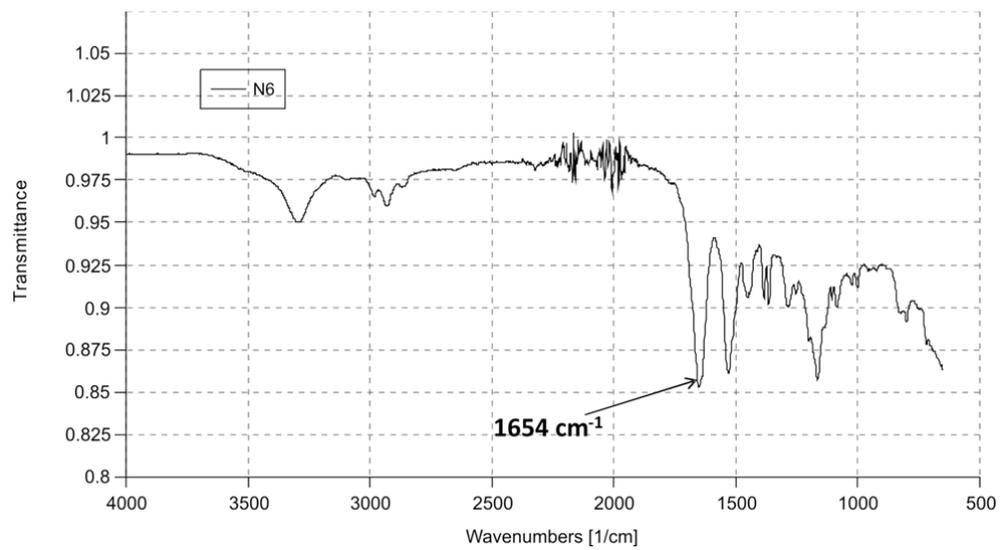


Figure S8. IR spectrum for peptide 7.

5. Analysis of computational models for peptides 5, 6 and 7, 8

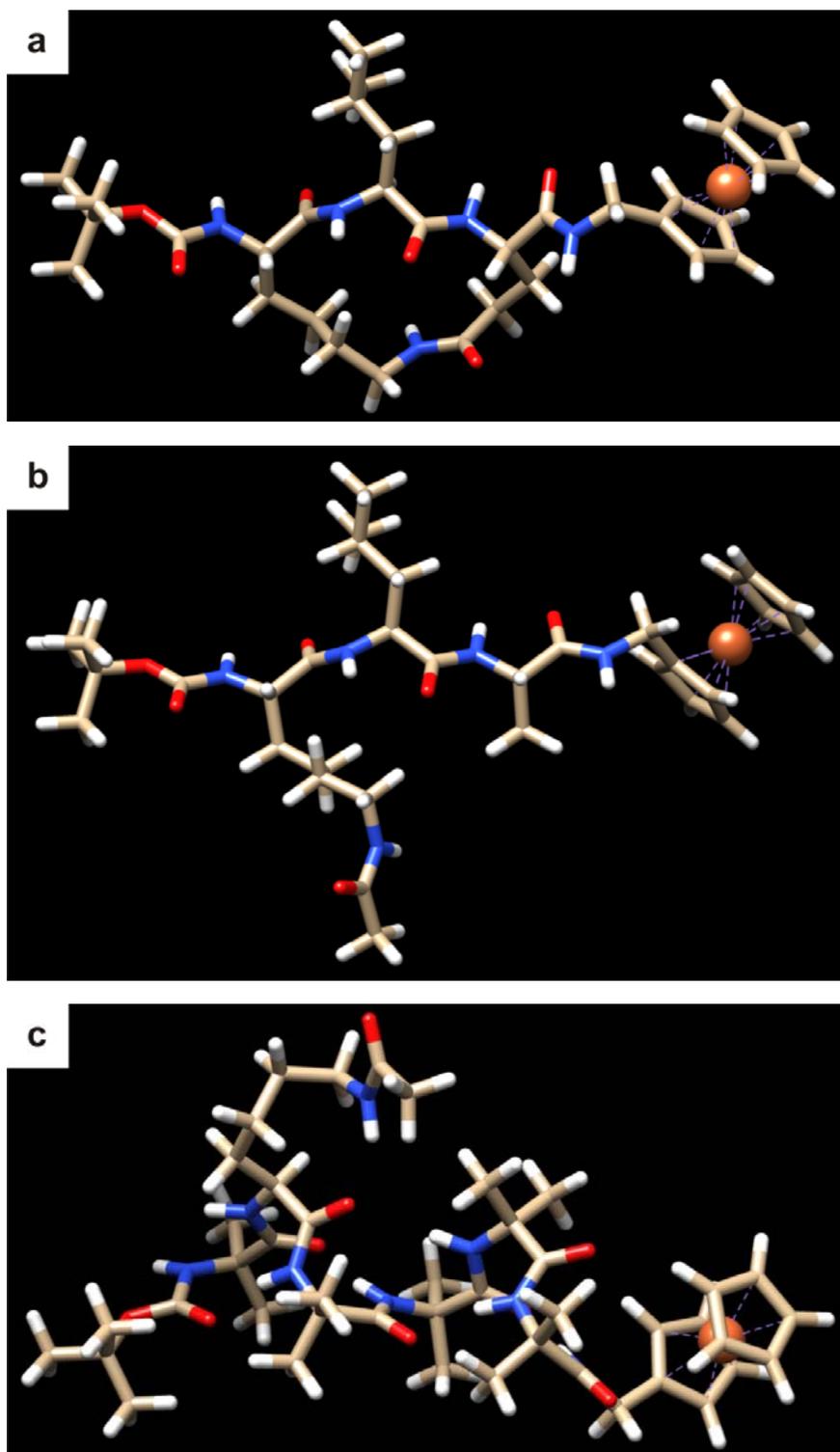


Figure S9. The lowest energy conformer for (a) peptide 5, (b) peptide 6, and (c) peptide 8, optimized by the hybrid B3LYP method with 6-31G** basis set for all C, H, N, O atoms, and Lanl2dz basis set for Fe atom.

5.1 Characteristics of *N*-protected β -strand peptides **5** and **6**.

Table S1. *NH* to *NH* distances and total lengths for peptides **5** and **6**.

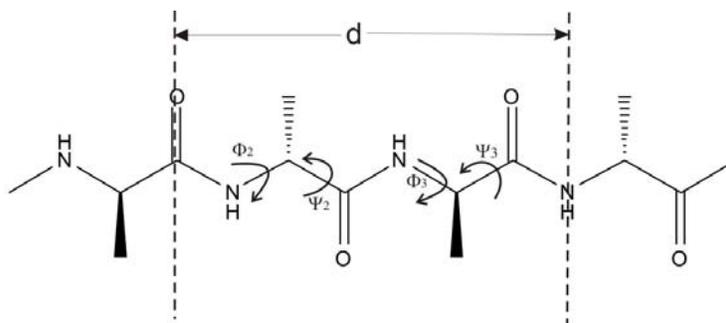
<i>NH</i> to <i>NH</i> distance (Å)	peptide 5	peptide 6
1-2	4.207	4.311
2-3	4.316	4.374
3-4	4.357	4.271
Distance from first to last carbonyl carbon (Å)	10.205	10.627

Table S2. Dihedral angles for all residues in the lowest energy conformers for peptides **5** and **6**.

	peptide 5		peptide 6	
	Φ	Ψ	Φ	Ψ
Residue 1	-150.366	122.799	-157.060	146.222
Residue 2	-126.024	158.039	-129.267	162.041
Residue 3	-118.162	151.364	-157.670	163.394

Table S3. Important characteristic correlations for peptides **5** and **6**, with comparison to optimal β -strand values. (all distances in Å)

	peptide 5	peptide 6	optimal β -strand conformation
Length (first to last carbonyl)	10.205	10.627	
Distance (d)*	8.0	8.2	8.0 ⁴
N-Leu to CO-Leu	2.4	2.4	2.5 ⁵
NH to NH (Average)	4.3	4.3	4.3 ⁵
α H to NH+1	2.5	2.5	2.2 ⁵
β H ₂ to NH+1	3.5	3.6	3.2 to 4.5 ⁵



Note: * This distance (d) is defined between the C atom (*i*) and N atom (*i*+3). This is indicative of an optimal extended β -strand, as shown above.

5.2 Characteristics of *N*-protected helical peptides **7** and **8**.

Table S4. Hydrogen bond lengths, *NH* to *NH* distances and total peptide length for peptide **7**.

Residue	Hydrogen bond lengths (Å)	Distance (<i>NH</i> to <i>NH</i>) (Å)
1	2.074	2.996
2	2.184	2.817
3	2.061	2.837
4	2.157	2.779
5	2.198	2.680
6		2.602
Average	2.13	2.78
• Distance from first to last carbonyl carbon 11.989 Å.		

Table S5. Hydrogen bond lengths, *NH* to *NH* distances and total peptide length for peptide **8**.

Residue	Hydrogen bond lengths (Å)	Distance (<i>NH</i> to <i>NH</i>) (Å)
1	2.123	2.987
2	2.073	2.790
3	2.535	2.743
4	2.140	2.832
5	2.066	2.804
6		2.680
Average	2.18	2.80
• Distance from first to last carbonyl carbon 11.816 Å.		

Table S6. Distances between $d\alpha\text{N}$, $d\beta\text{N}$ and $d\text{NN}$, characteristic of a 3_{10} -helix.

Distance	peptide 7	peptide 8	Ideal 3_{10} helix ⁵
$d\alpha\text{N}$ (Å)	3.5	3.5	3.4
$d\beta\text{N}$ (Å)	3.0-4.1	3.0-4.1	2.9-4.4
Averaged NN (Å)	2.7	2.8	2.6

Table S7. Dihedral angles for all residues in the lowest energy conformers for peptides **7** and **8**.

	peptide 7		peptide 8	
	Φ	ψ	Φ	ψ
Residue 1	-64.156	-28.640	-64.329	-28.300
Residue 2	-55.638	-27.008	-56.464	-28.097
Residue 3	-52.532	-31.510	-59.037	-23.420
Residue 4	-55.092	-29.286	-55.844	-28.940
Residue 5	-70.469	-10.385	-53.315	-31.545
Residue 6	-66.630	-24.062	-66.533	-20.283
Average	-60.75	-25.14	-59.25	-26.76
Differs from ideal 3_{10} helix	3.75°	4.86°	2.25°	3.24°

6. Electrochemical measurements

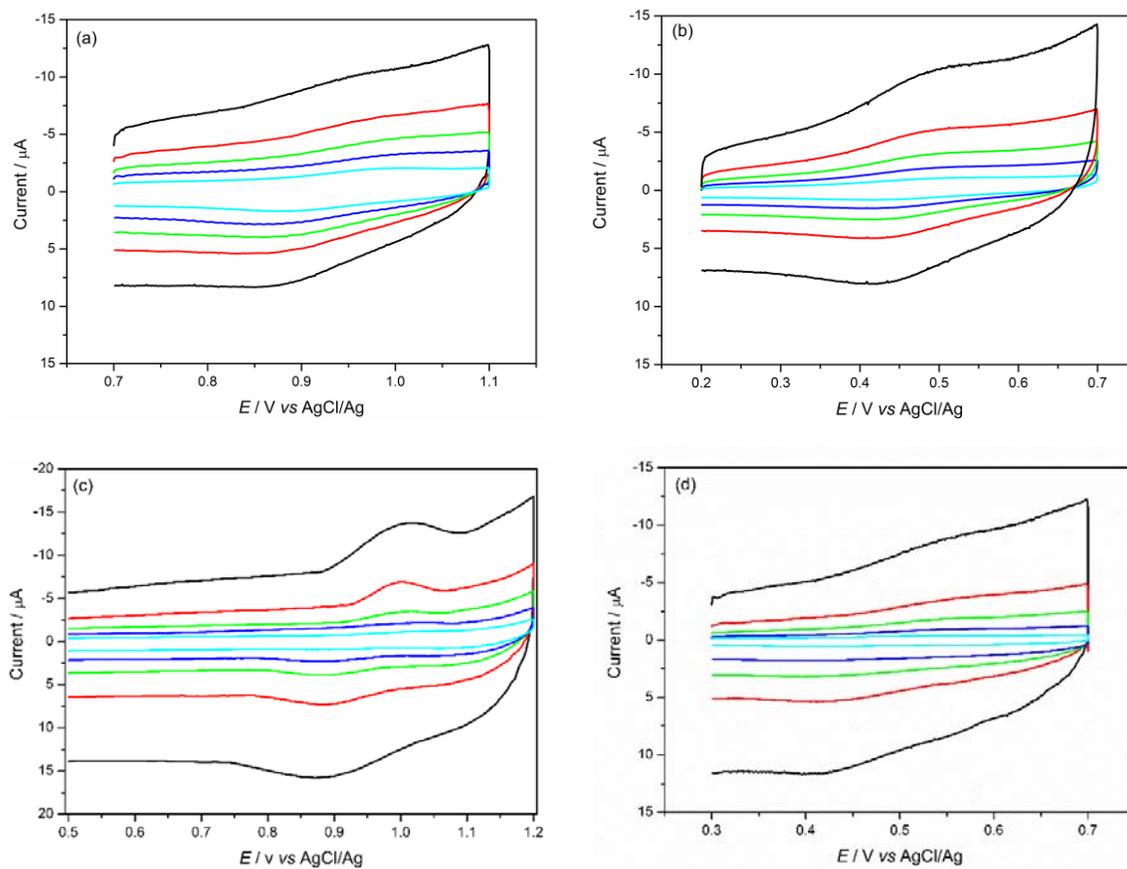


Figure S10. Cyclic voltammograms for (a) peptide **1** (b) peptide **2** (c) peptide **3** and (d) peptide **4** immobilized on SWCNTs/Au electrodes taken at 5, 2, 1, 0.5 and 0.2 V s^{-1} (from top to centre).

7. Electronic transport simulations

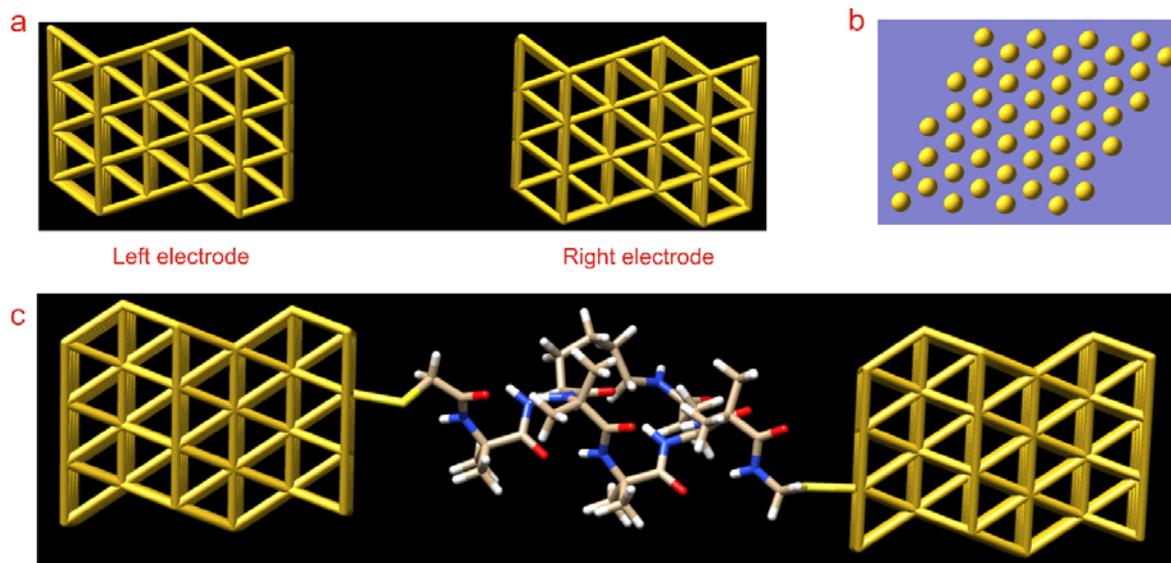


Figure S11. Scheme of the gold leads used for building the extended molecule, with (a) side view and (b) top view of 4 x 4 x 6 gold layers for each lead. The face of the leads corresponds to a (111) surface. (c) A typical Au-peptide-Au setup for electronic transport simulation, with peptide 11 incorporated here.

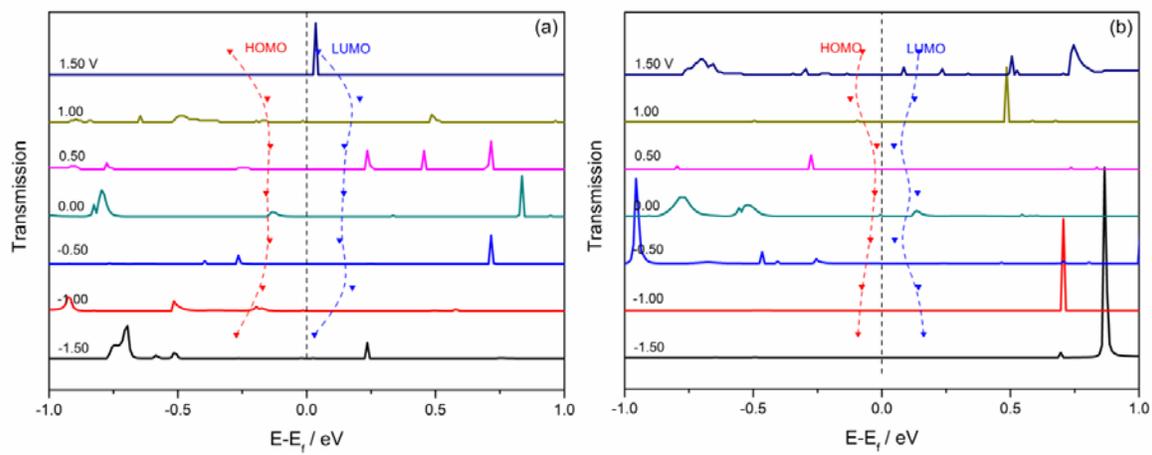


Figure S12. Transmission spectra of (a) the constrained **9** and (b) linear peptide **10** at different bias voltages, indicating the molecular orbital energy levels in proximity to the Au Fermi level (E_f).

**Sample input for TranSIESTA calculation:
0V calculation for Au-peptide 11-Au**

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SystemLabel cyclic_0d00V

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SPECIES AND BASIS

Chemical species

NumberOfSpecies 6

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2	16	S	# Species index, atomic number, species label
3	1	H	# Species index, atomic number, species label
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5	6	C	# Species index, atomic number, species label
6	7	N	# Species index, atomic number, species label

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K-points

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UNIT CELL AND ATOMIC POSITIONS

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xc.authors PBE
SpinPolarized .false. # Exchange-correlation version

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SaveHS                   .false.
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TS.ComplexContour.NLine  10
TS.biasContour.NumPoints  15

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# TBT OPTIONS
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TS.TBT.Emax              +5.00 eV

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TS.TBT.Eta 0.000001 Ry

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TS.SaveLead .false.

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TS.ReplicateA2Left 1
TS.NumUsedAtomsLeft 64
TS.BufferAtomsLeft 0

RIGHT ELECTRODE

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TS.ReplicateA1Right 1
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TS.NumUsedAtomsRight 64
TS.BufferAtomsRight 0

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AtomicCoordinatesFormat Ang

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-4.19062011	4.03507976	4.71004000	1	46	Au
-1.30634011	4.03508976	4.71004000	1	47	Au
1.57794989	4.03508976	4.71004000	1	48	Au
-1.30633011	-4.29114024	7.06505000	1	49	Au
1.57795989	-4.29114024	7.06505000	1	50	Au
4.46224989	-4.29114024	7.06505000	1	51	Au
7.34652989	-4.29114024	7.06505000	1	52	Au
-2.74847011	-1.79327024	7.06505000	1	53	Au
0.13580989	-1.79327024	7.06505000	1	54	Au
3.02009989	-1.79327024	7.06505000	1	55	Au
5.90438989	-1.79327024	7.06505000	1	56	Au
-4.19062011	0.70458976	7.06505000	1	57	Au
-1.30633011	0.70458976	7.06505000	1	58	Au
1.57794989	0.70458976	7.06505000	1	59	Au
4.46224989	0.70459976	7.06505000	1	60	Au
-5.63277011	3.20244976	7.06505000	1	61	Au
-2.74848011	3.20244976	7.06505000	1	62	Au
0.13580989	3.20244976	7.06505000	1	63	Au
3.02009989	3.20245976	7.06505000	1	64	Au
-1.30634011	-2.62590024	9.42006000	1	65	Au
1.57795989	-2.62590024	9.42006000	1	66	Au
4.46223989	-2.62590024	9.42006000	1	67	Au
7.34653989	-2.62589024	9.42006000	1	68	Au
-2.74848011	-0.12803024	9.42006000	1	69	Au
0.13580989	-0.12803024	9.42006000	1	70	Au
3.02009989	-0.12803024	9.42006000	1	71	Au
5.90438989	-0.12803024	9.42006000	1	72	Au
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1.57795989	2.36983976	9.42006000	1	75	Au
4.46223989	2.36983976	9.42006000	1	76	Au
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3.02009989	4.86770976	9.42006000	1	80	Au
-2.74847011	-3.45853024	11.77507000	1	81	Au
0.13580989	-3.45853024	11.77507000	1	82	Au
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5.90438989	-3.45851024	11.77507000	1	84	Au

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2.26117989	-1.31582024	17.46748000	5	101	C
2.54887989	-2.59059024	18.37520000	5	102	C
3.61701989	-0.61422024	17.05254000	5	103	C
0.81174989	0.79306976	17.78933000	5	104	C
-0.28596011	1.51981976	18.69994000	5	105	C
0.01368989	3.07117976	18.80443000	5	106	C
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-1.47192011	-0.78989024	21.73035000	5	108	C
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1.16321989	6.23156976	22.39127000	5	113	C
1.97191989	-1.26335024	24.82928000	5	114	C
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-0.01947011	3.12783976	25.89996000	6	129	N
1.46287989	1.15358976	24.39055000	6	130	N
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1.38072989	-0.37954024	18.28401000	6	132	N
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-3.07607011	-0.46713024	23.25751000	3	163	H
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-1.30633011	-4.29115024	33.47507000	1	196	Au

1.57795989	-4.29115024	33.47507000	1	197	Au
4.46224989	-4.29115024	33.47507000	1	198	Au
7.34653989	-4.29115024	33.47507000	1	199	Au
-2.74848011	-1.79329024	33.47507000	1	200	Au
0.13580989	-1.79329024	33.47507000	1	201	Au
3.02009989	-1.79329024	33.47507000	1	202	Au
5.90438989	-1.79329024	33.47507000	1	203	Au
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-1.30633011	4.03506976	38.18509000	1	241	Au
1.57794989	4.03506976	38.18509000	1	242	Au
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1.57795989	-4.29115024	40.54011000	1	244	Au
4.46224989	-4.29115024	40.54011000	1	245	Au
7.34653989	-4.29115024	40.54011000	1	246	Au
-2.74848011	-1.79329024	40.54011000	1	247	Au
0.13580989	-1.79329024	40.54011000	1	248	Au
3.02009989	-1.79329024	40.54011000	1	249	Au
5.90438989	-1.79329024	40.54011000	1	250	Au
-4.19062011	0.70457976	40.54011000	1	251	Au
-1.30633011	0.70457976	40.54011000	1	252	Au

1.57795989	0.70457976	40.54011000	1	253	Au
4.46224989	0.70457976	40.54011000	1	254	Au
-5.63277011	3.20244976	40.54011000	1	255	Au
-2.74848011	3.20244976	40.54011000	1	256	Au
0.13580989	3.20244976	40.54011000	1	257	Au
3.02009989	3.20244976	40.54011000	1	258	Au
-1.30633011	-2.62591024	42.89512000	1	259	Au
1.57795989	-2.62591024	42.89512000	1	260	Au
4.46223989	-2.62591024	42.89512000	1	261	Au
7.34652989	-2.62591024	42.89512000	1	262	Au
-2.74847011	-0.12804024	42.89512000	1	263	Au
0.13581989	-0.12804024	42.89512000	1	264	Au
3.02010989	-0.12804024	42.89512000	1	265	Au
5.90437989	-0.12804024	42.89512000	1	266	Au
-4.19062011	2.36981976	42.89512000	1	267	Au
-1.30633011	2.36981976	42.89512000	1	268	Au
1.57795989	2.36981976	42.89512000	1	269	Au
4.46224989	2.36981976	42.89512000	1	270	Au
-5.63277011	4.86768976	42.89512000	1	271	Au
-2.74848011	4.86768976	42.89512000	1	272	Au
0.13580989	4.86768976	42.89512000	1	273	Au
3.02009989	4.86768976	42.89512000	1	274	Au
-2.74848011	-3.45853024	45.25013000	1	275	Au
0.13580989	-3.45853024	45.25013000	1	276	Au
3.02009989	-3.45853024	45.25013000	1	277	Au
5.90438989	-3.45853024	45.25013000	1	278	Au
-4.19061011	-0.96067024	45.25013000	1	279	Au
-1.30634011	-0.96066024	45.25013000	1	280	Au
1.57794989	-0.96066024	45.25013000	1	281	Au
4.46223989	-0.96066024	45.25013000	1	282	Au
-5.63276011	1.53719976	45.25013000	1	283	Au
-2.74847011	1.53719976	45.25013000	1	284	Au
0.13580989	1.53720976	45.25013000	1	285	Au
3.02009989	1.53720976	45.25013000	1	286	Au
-7.07491011	4.03506976	45.25013000	1	287	Au
-4.19062011	4.03506976	45.25013000	1	288	Au
-1.30633011	4.03506976	45.25013000	1	289	Au
1.57794989	4.03506976	45.25013000	1	290	Au

%endblock AtomicCoordinatesAndAtomicSpecies

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8. Geometric similarity of peptide backbones in molecular junctions

8.1 Geometric similarity of β -strand peptide backbones (**9** and **10**).

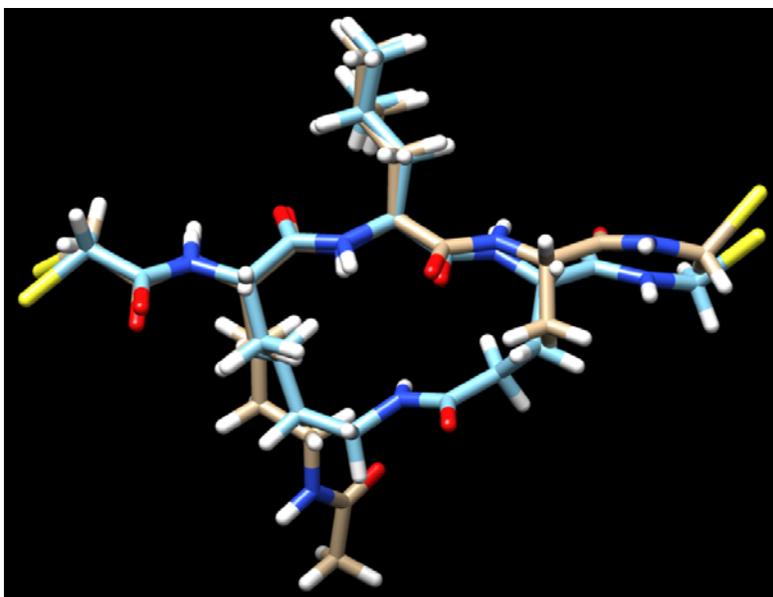


Figure S13. The superposition of the two β -strand peptides, **9** and **10**. (The light blue strand denotes the constrained peptide **9**, while the grey strand represents the linear peptide **10**).

8.2 Geometric similarity of helical peptide backbones (**11** and **12**).

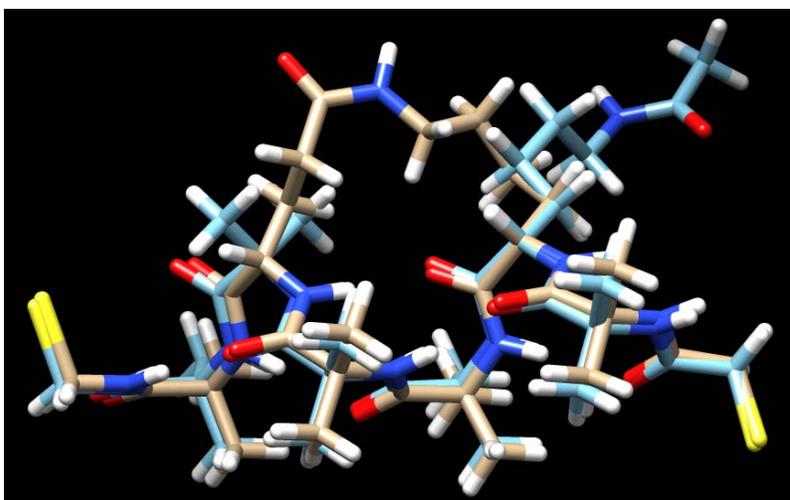
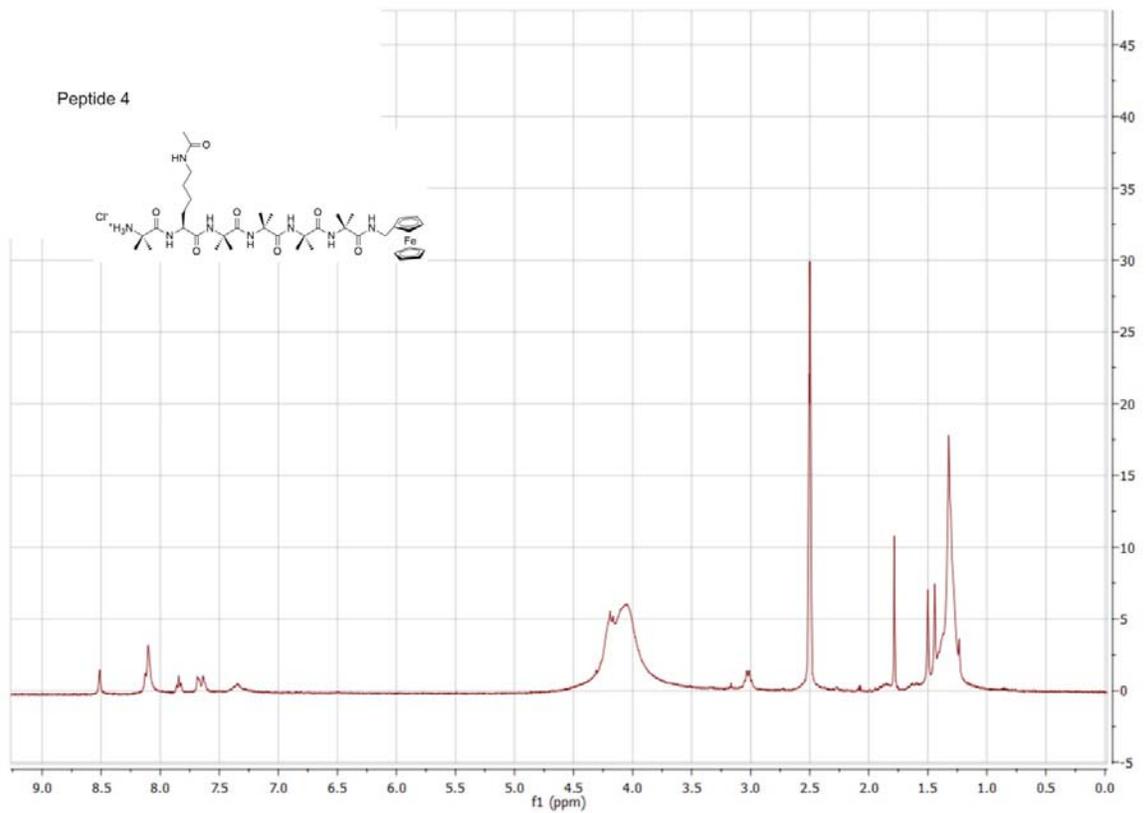
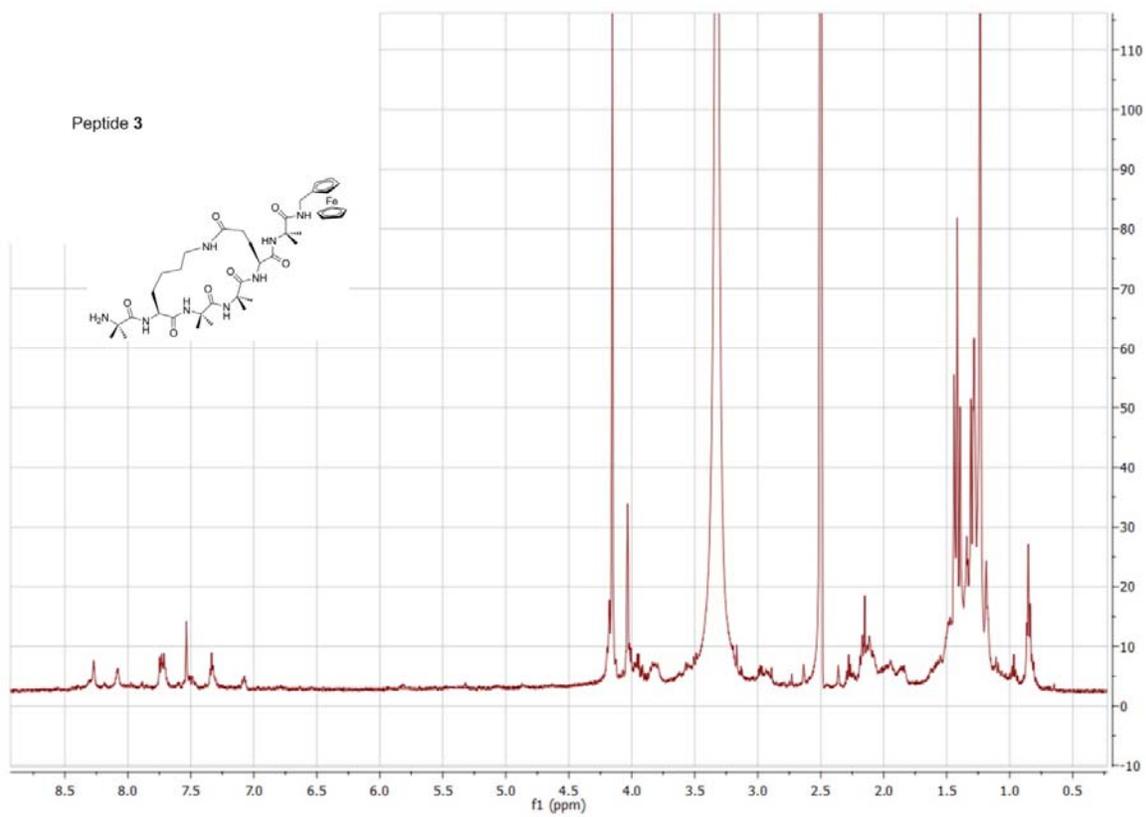


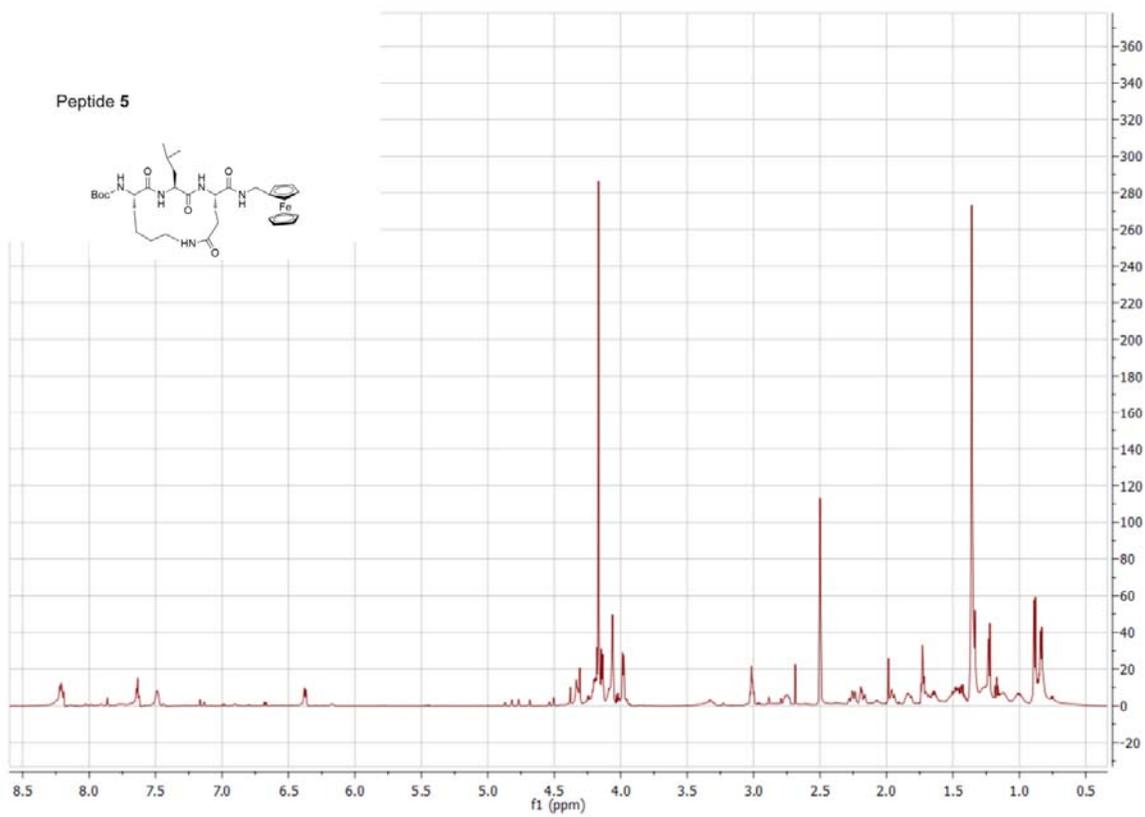
Figure S14. The superposition of the two helical peptides, **11** and **12**. (The grey strand denotes the constrained peptide **11**, while the light blue strand represents the linear peptide **12**).

Table S8. Distances between S-S atoms in molecular junctions containing peptides **9-12**.

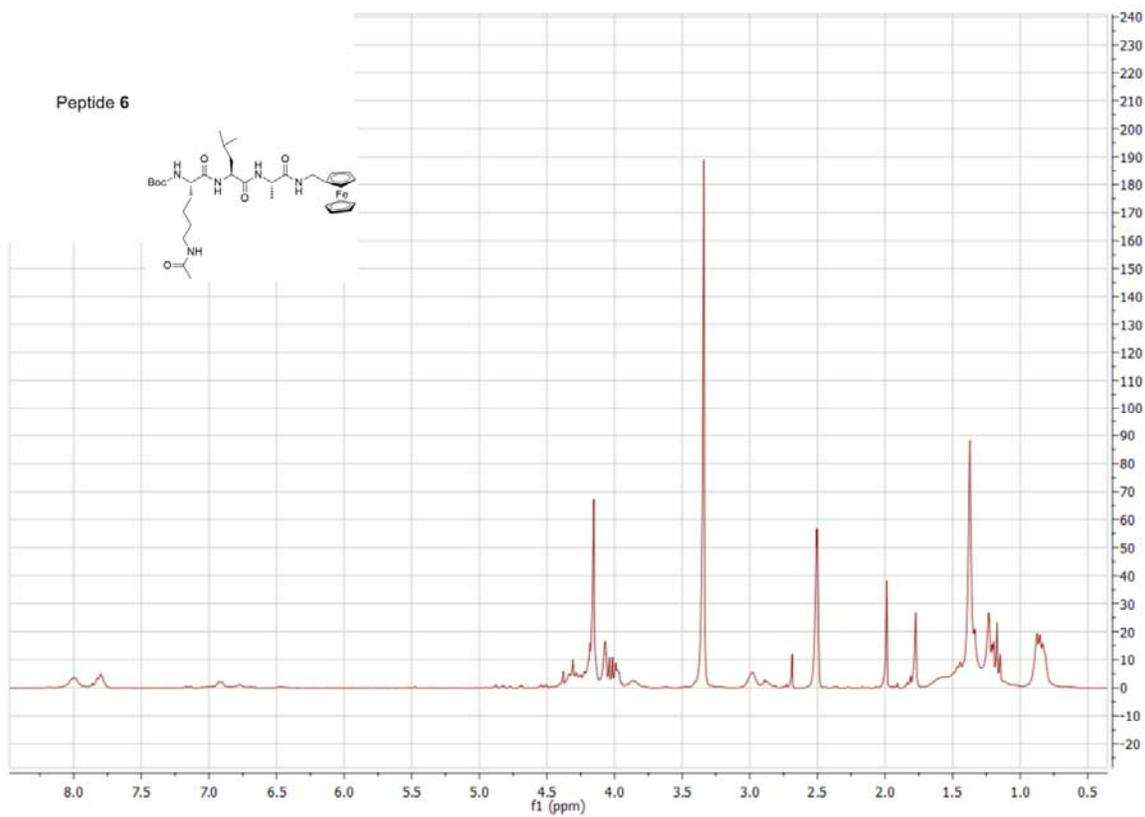
Peptide	Secondary structure	Linear or constrained	S-S distance (Å)
9	β -strand	Constrained	18.13
10	β -strand	Linear	18.02
11	3_{10} -helical	Constrained	17.36
12	3_{10} -helical	Linear	17.38

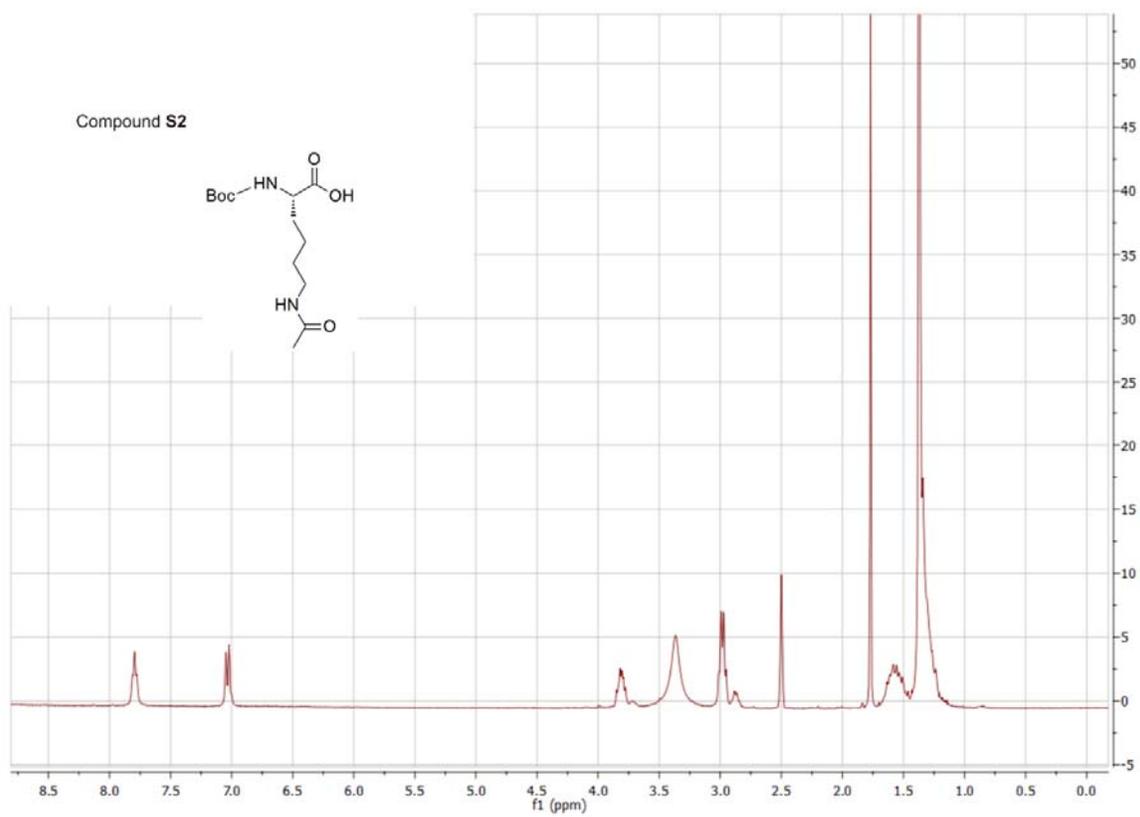
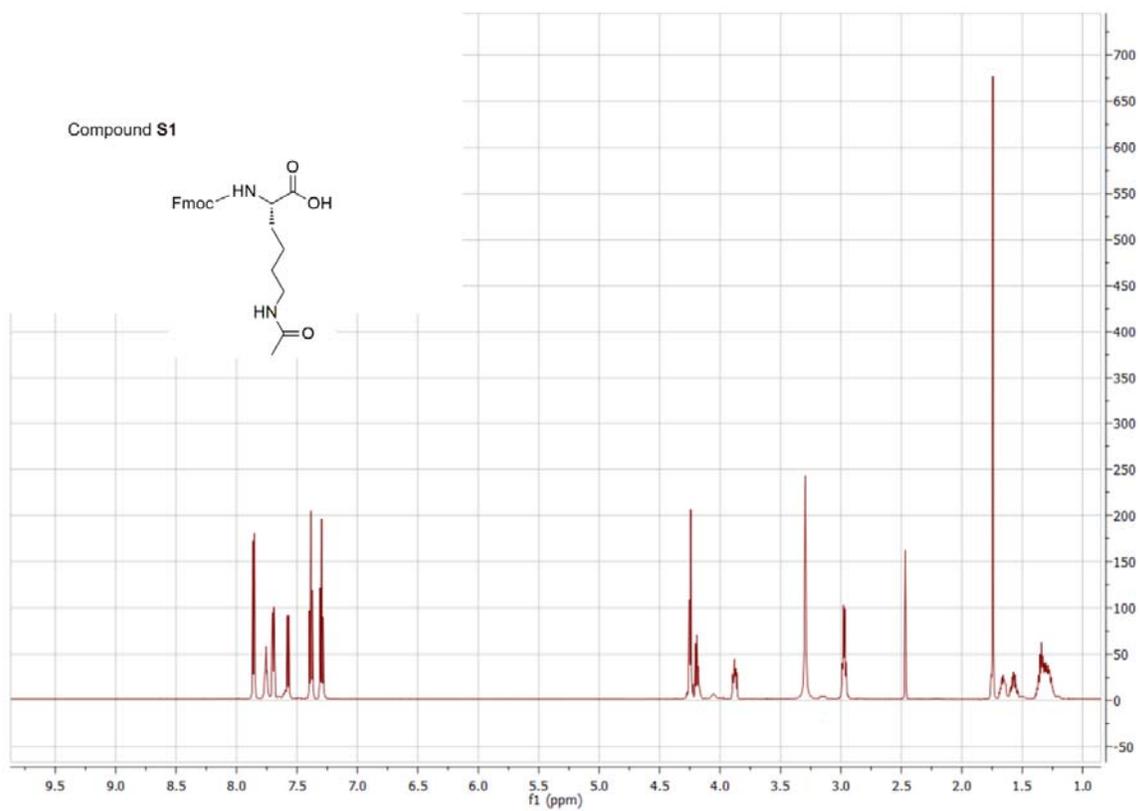


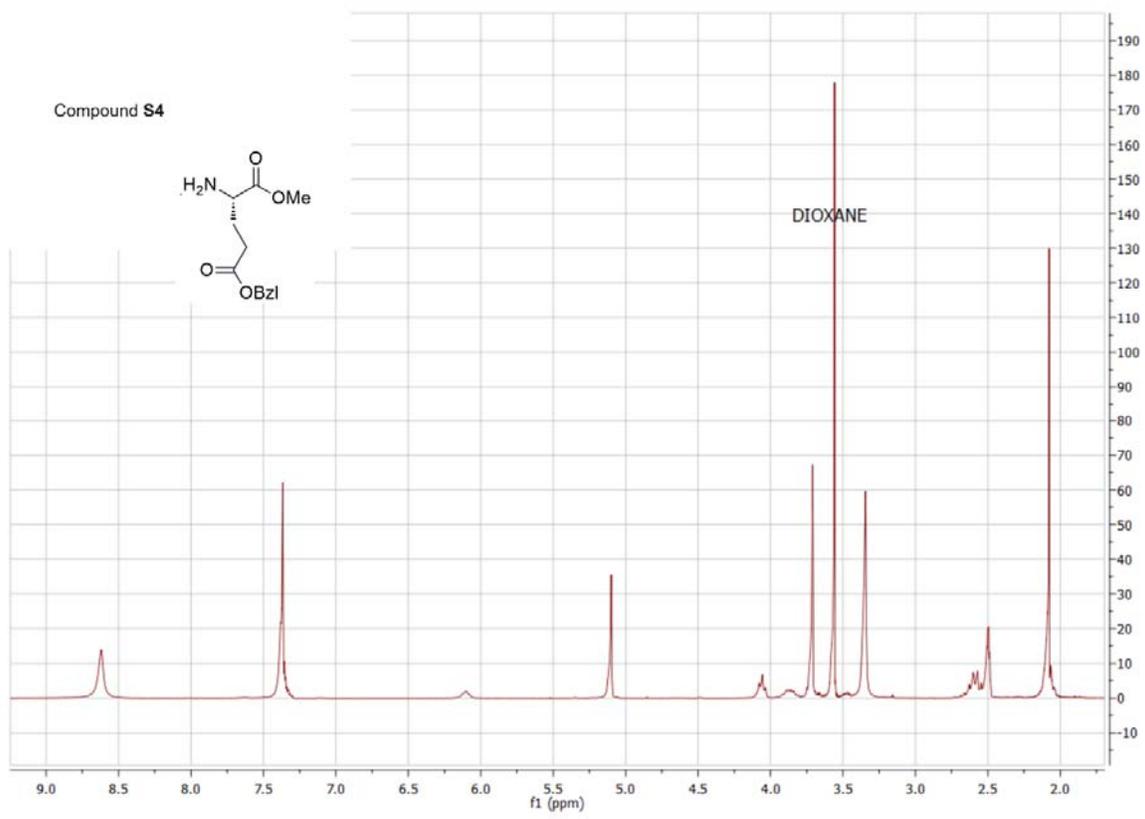
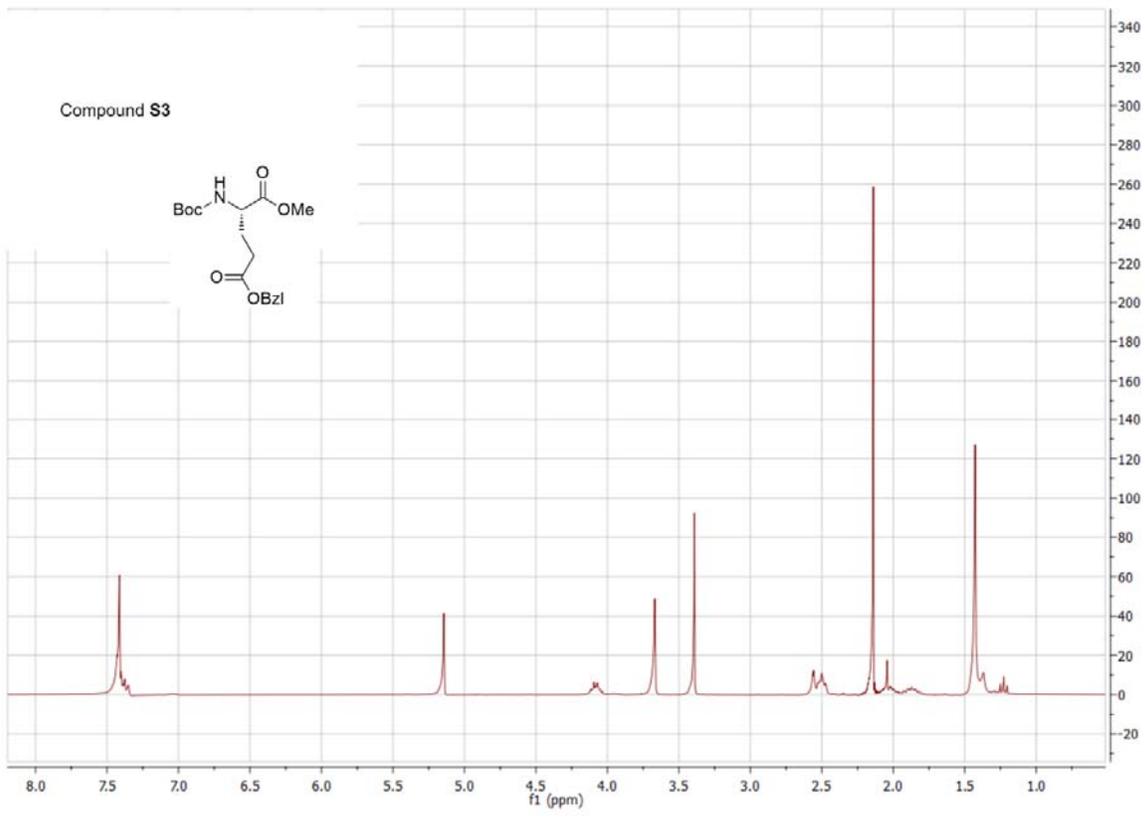
Peptide 5



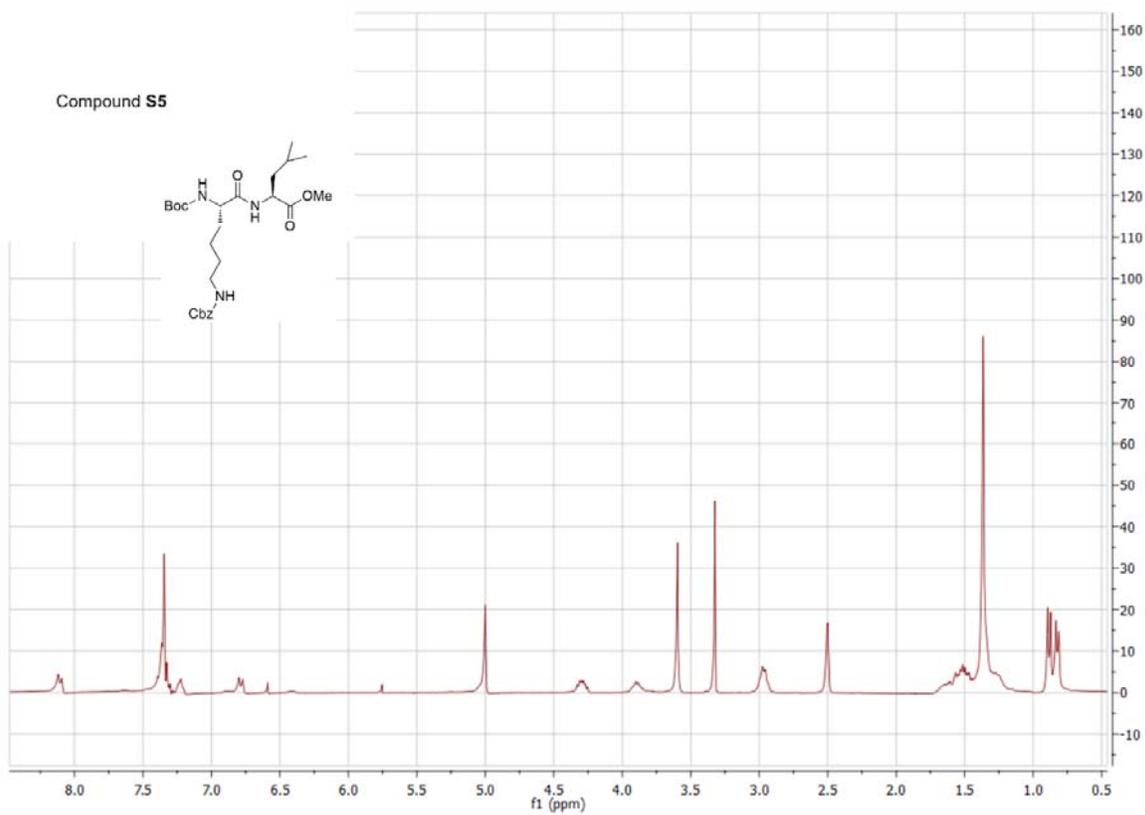
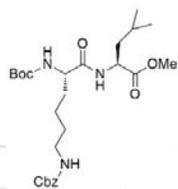
Peptide 6



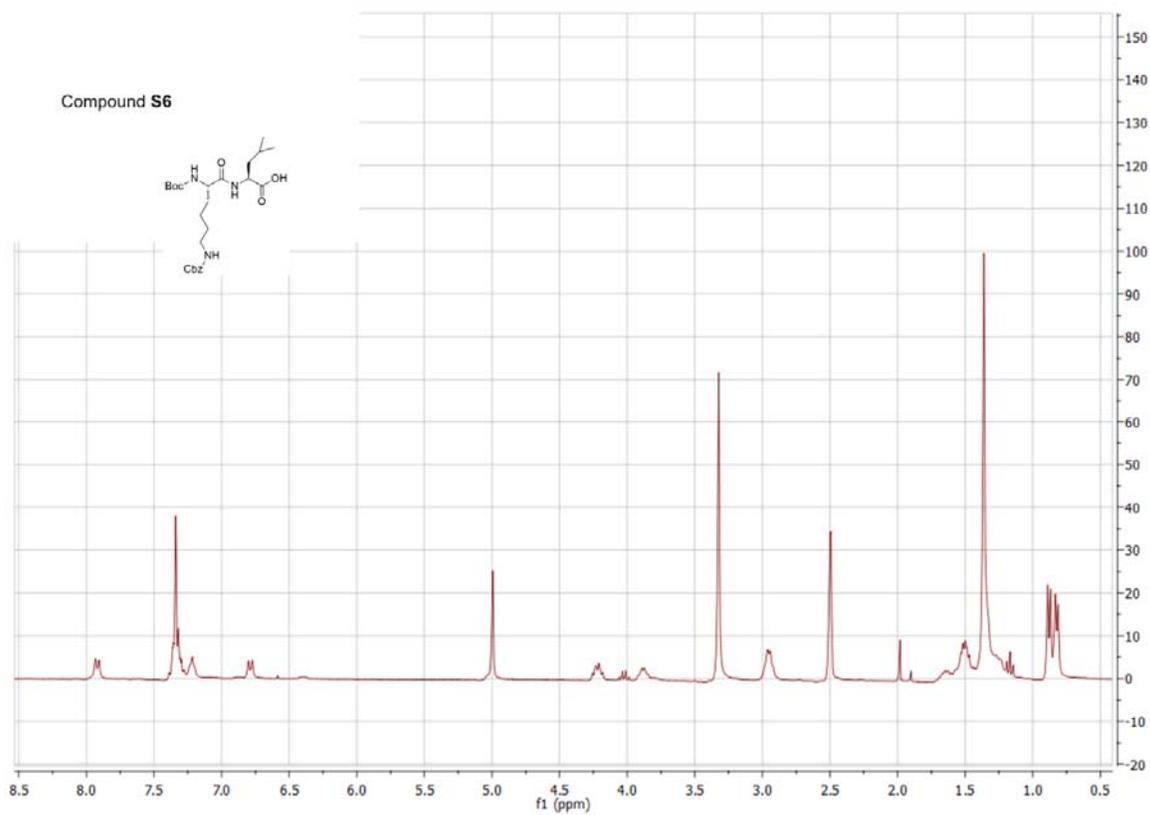
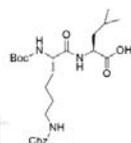


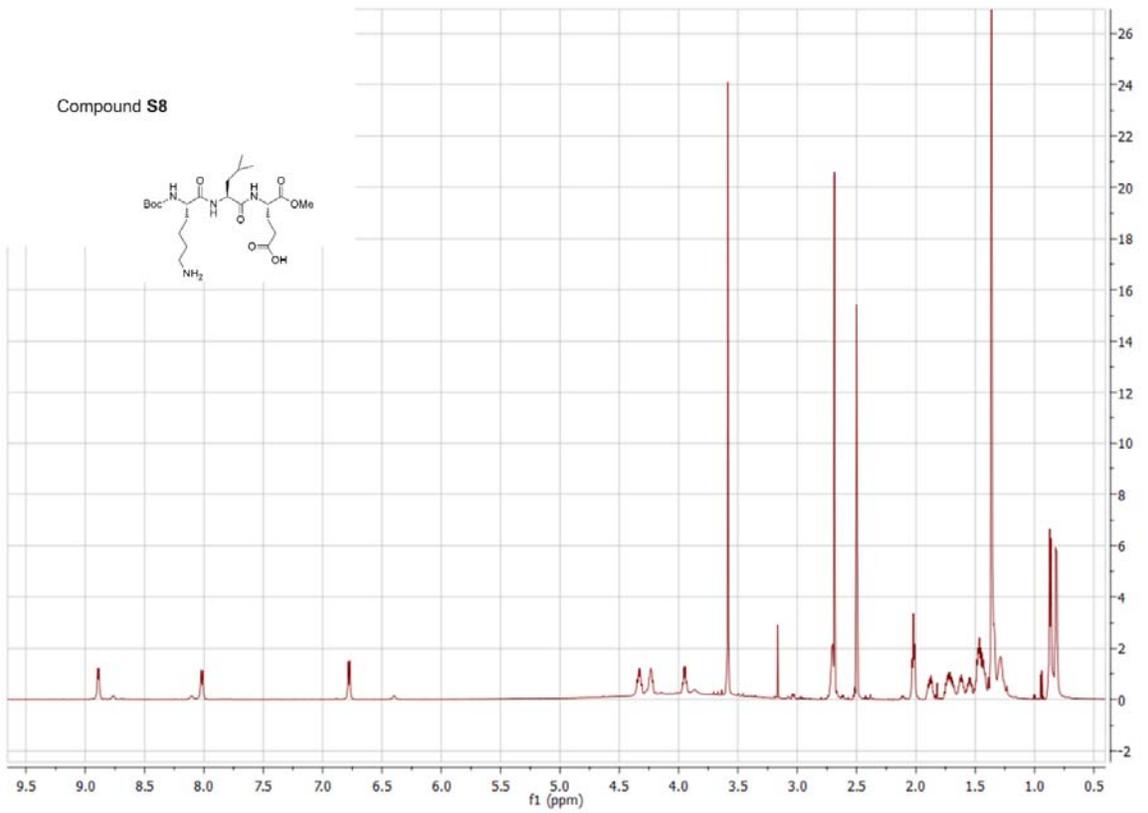
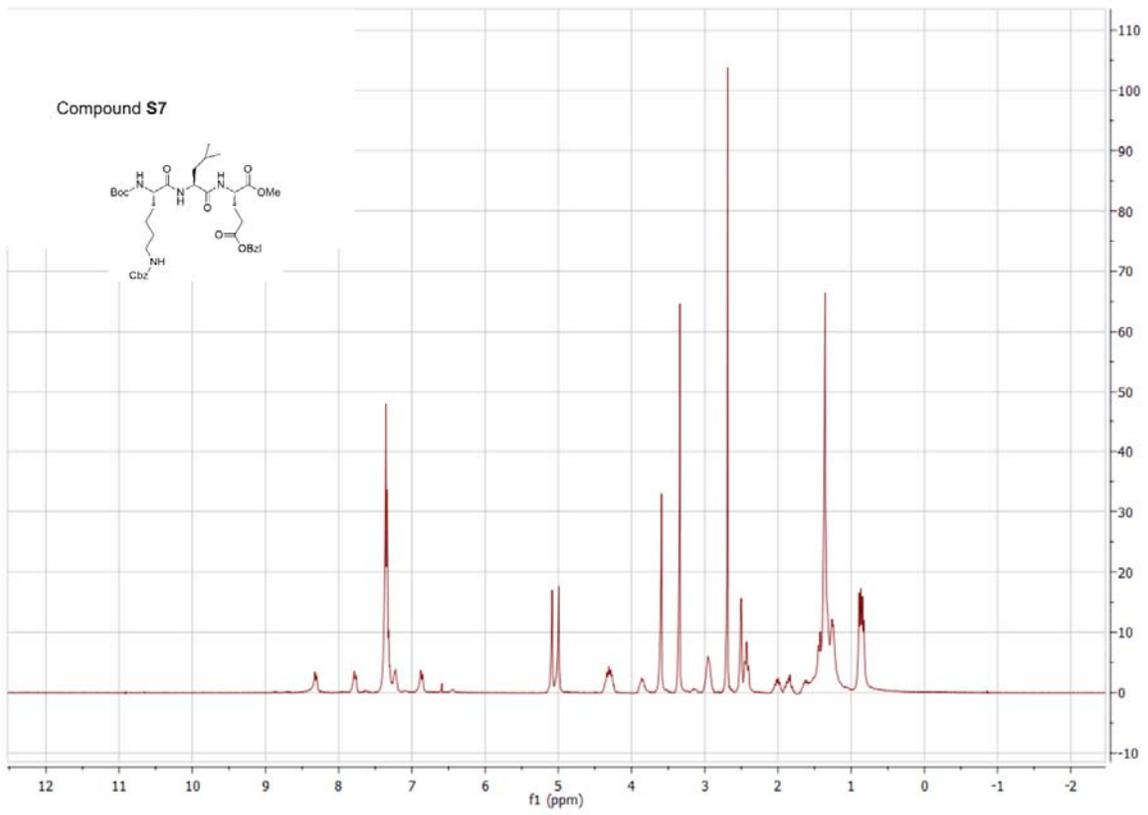


Compound S5

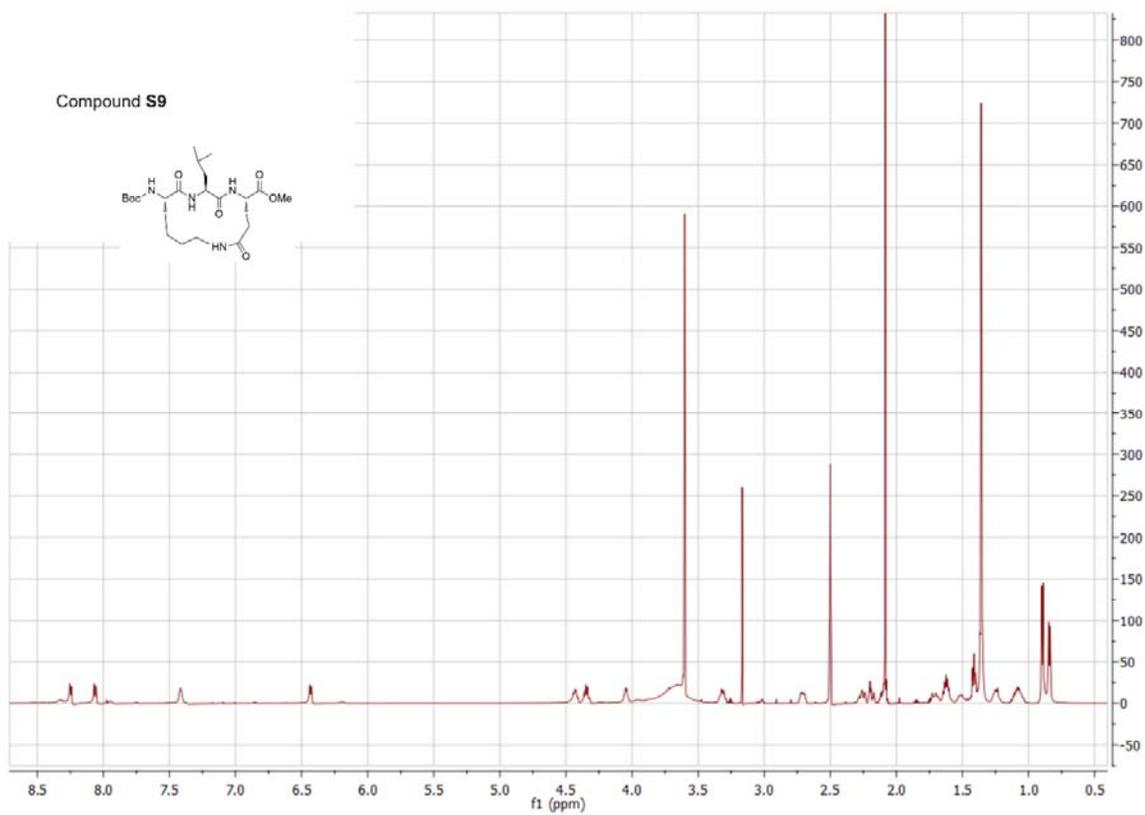


Compound S6

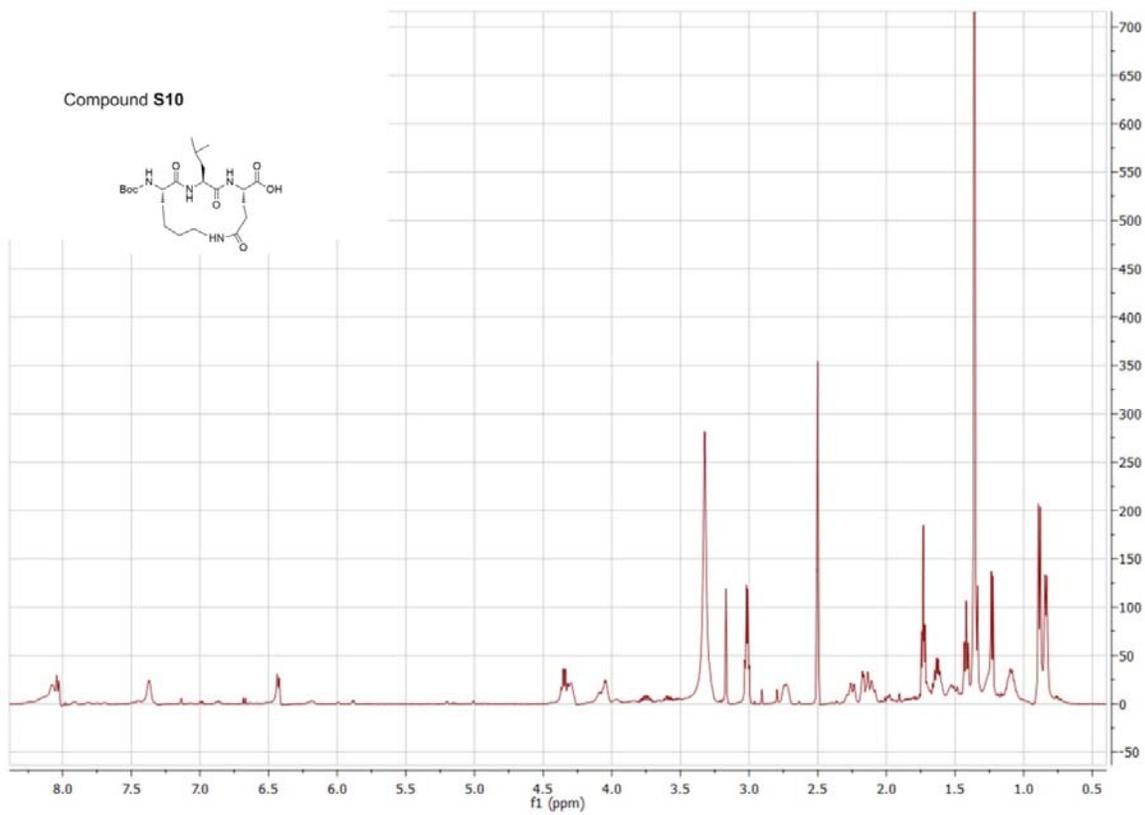


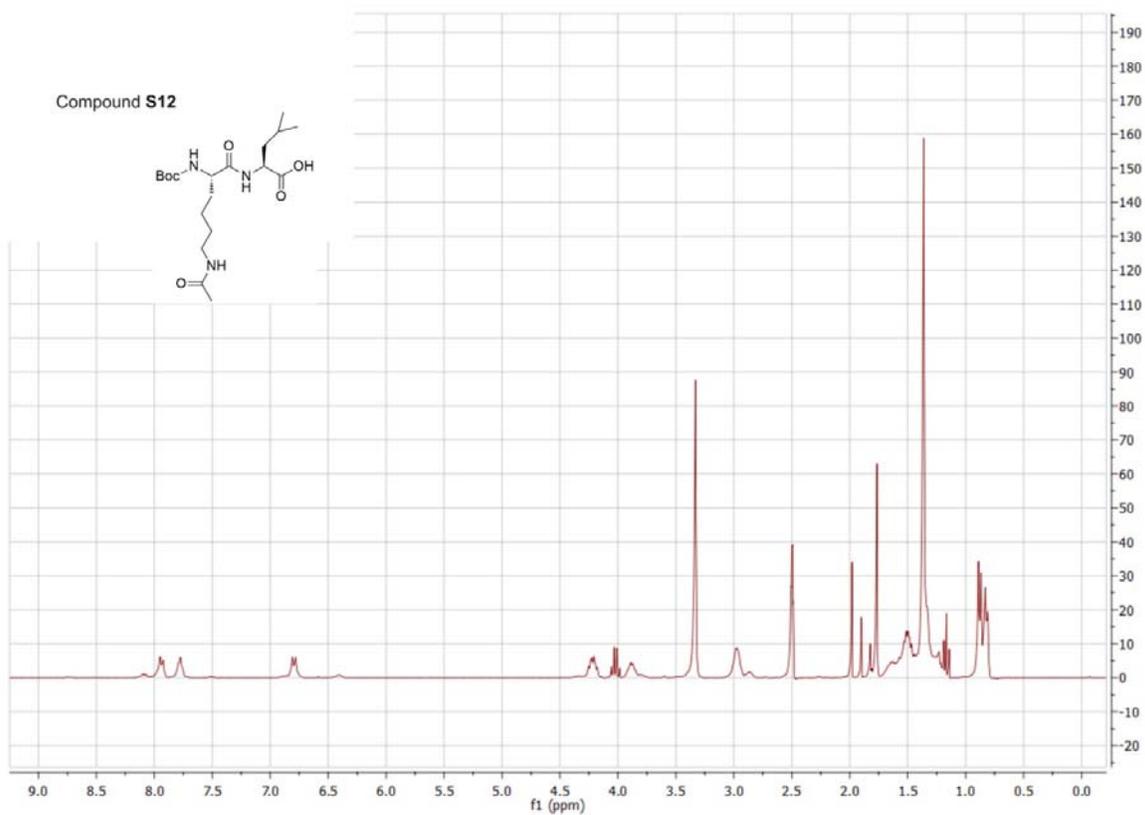
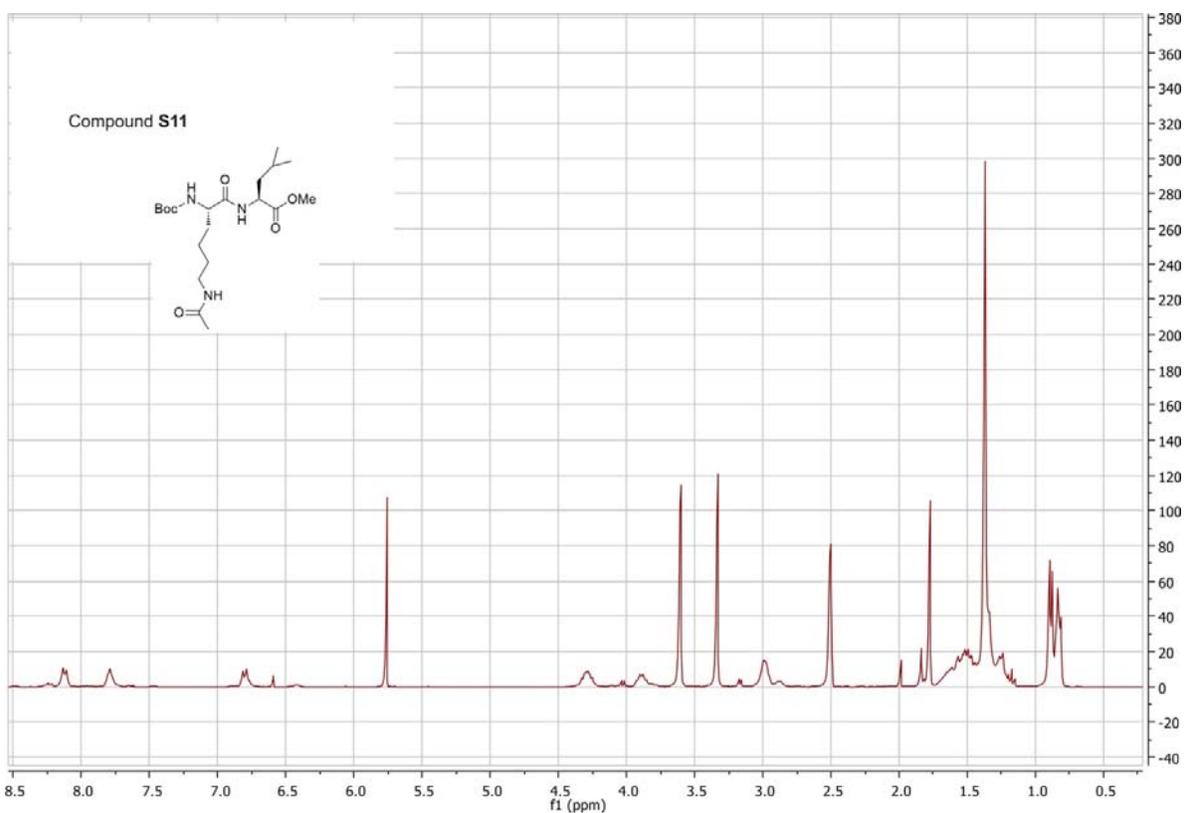


Compound S9

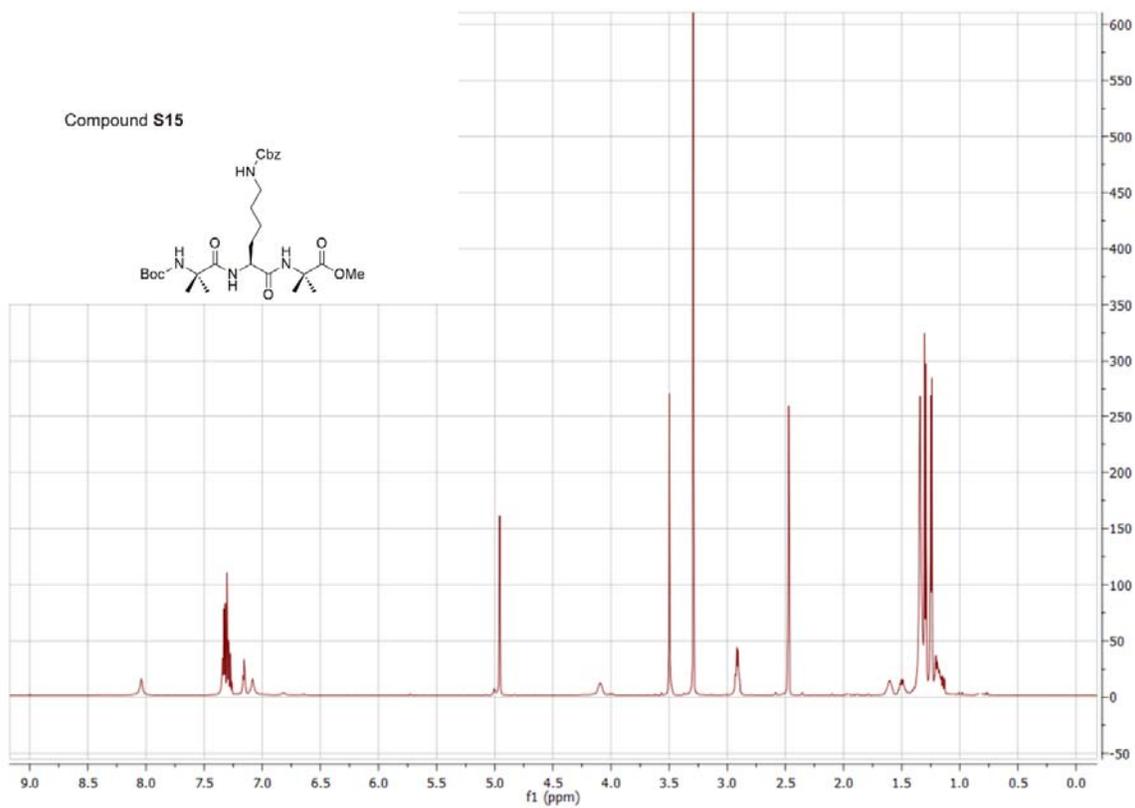
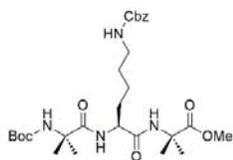


Compound S10

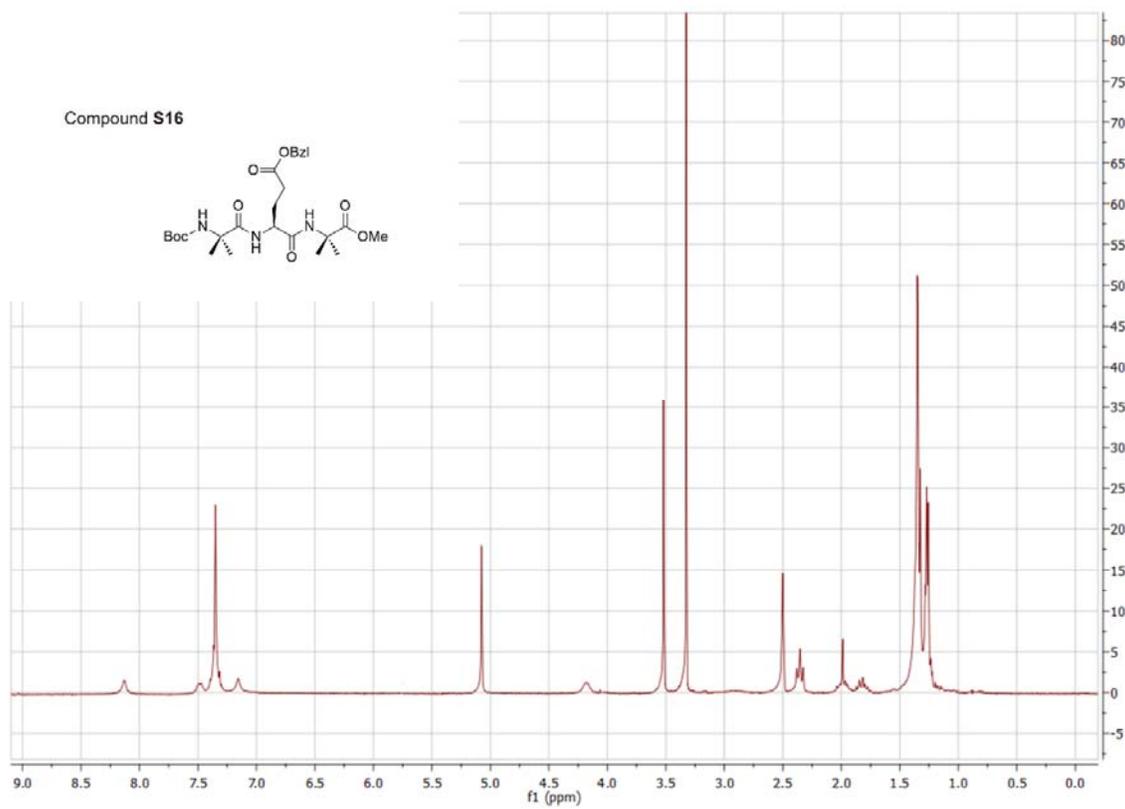
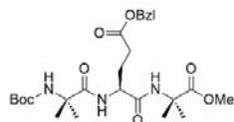


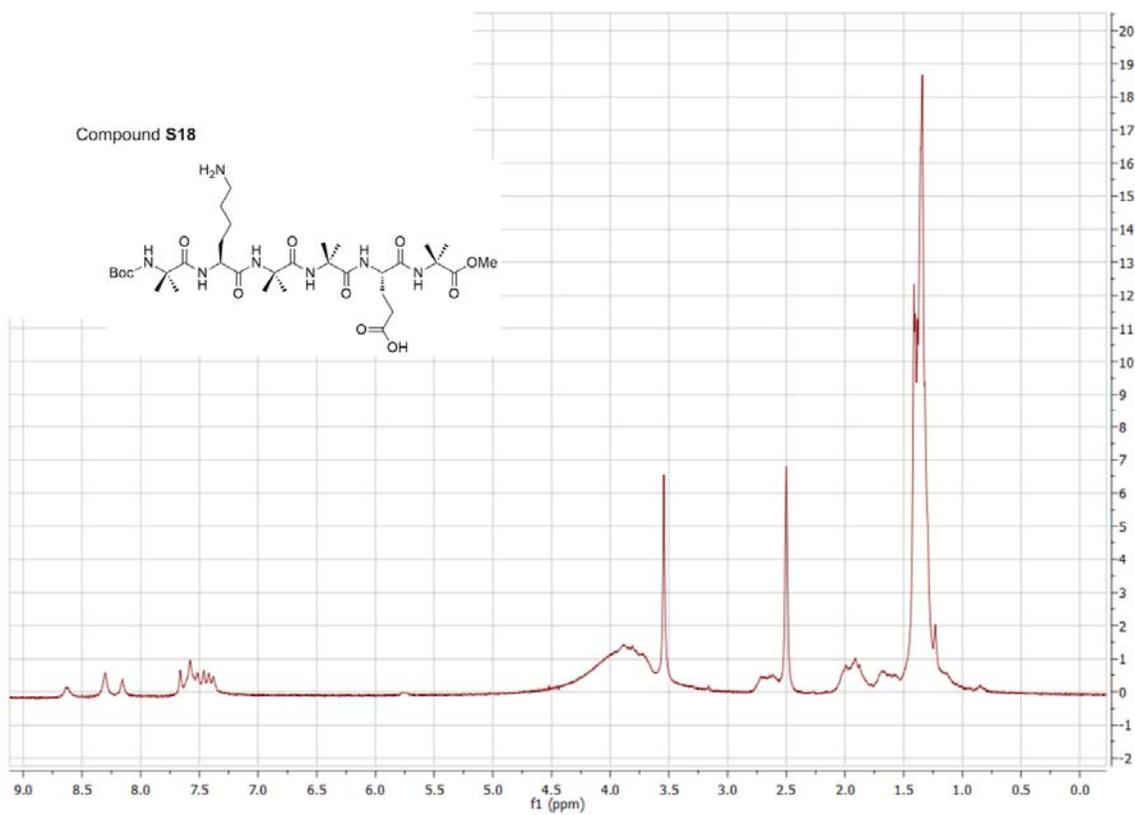
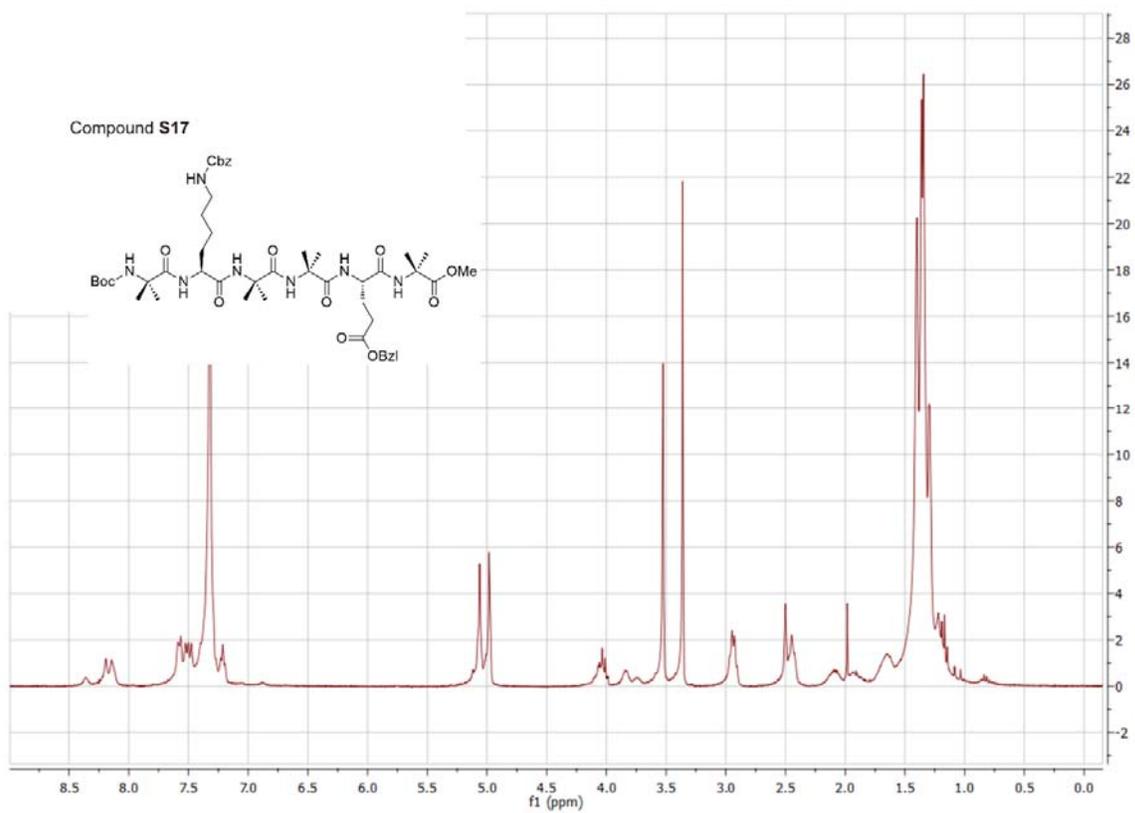


Compound S15



Compound S16





10. References

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