

Supplementary Material

Selective Complexation of Di-*n*-hexylammonium Salts by Tailed Porphyrin Host

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Experimental	S2
NMR and MS of OP1	S6
NMR and MS of OP2	S8
NMR and MS of OP3	S10
¹ H NMR titration of host OP with salt 4c	S12
Partial 2D NOESY and MS of the mixture of OP1 and 4c	S13
Partial 2D NOESY and MS of the mixture of OP2 and 4c	S14
Partial 2D NOESY and MS of the mixture of OP3 and 4c	S15
¹ H NMR Titration of host OP with salt 4b	S16
¹ H NMR of the mixture of OP and salt 4a	S18
UV-Vis titration of host OP with salt 4c	S19
References.....	S20

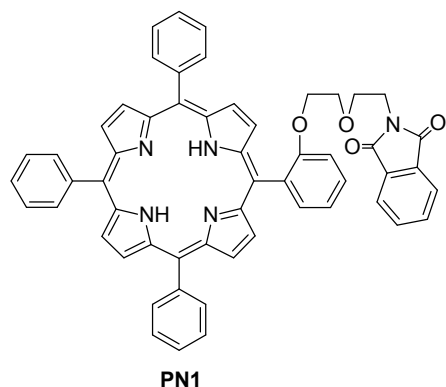
Experimental

Melting points are uncorrected. All reactions were performed under an atmosphere of dry nitrogen. The ^1H NMR and ^{13}C NMR spectra were recorded on 400 MHz spectrometers in the indicated solvents. Chemical shifts are expressed in parts per million (δ) using residual solvent protons as internal standards. Elemental analysis was carried out at the analytical center. Unless otherwise indicated, all starting materials were obtained from commercial suppliers and were used without further purification.

Determination of binding constants. Di-*n*-hexylammonium salts were added into the solution of tailed porphyrin. ^1H NMR titrations were carried out at the concentrations of 6.4 mM in CDCl_3 at 25°C. UV-Vis titrations were carried out at the concentration of 1×10^{-6} M in CH_2Cl_2 at 25°C. The 1:1 binding constants were calculated by using equation $K_{\text{assoc}} = \frac{[\text{host}]_c}{[\text{host}]_{uc}[\text{guest}]_{uc}}$, in which the concentration of complexed and uncomplexed host was defined by Δ value of chemical shift or absorbance.^{1,2}

Compound **OP-OH** and compound **3** was prepared according to the reference.^{3,4}

General procedure for the synthesis of PN. To the solution of **OP-OH** in DMF was added compound **3** (1.5 equiv.), potassium carbonate (10 equiv.). The reaction mixture was stirred at 120 °C for 2h. After removing DMF in vacuo, dichloromethane was added to the residue and washed with water, dried over anhydrous Na_2SO_4 . The solvent was evaporated, and the crude product was purified by column chromatography (DCM/MeOH, 100:1) to give the **PN** as a purple solid.



PN1: ^1H NMR (CDCl_3): δ -2.86 (s, 2H), 2.26 (t, 2H), 2.58 (t, 2H), 3.11 (t, 2H), 4.02 (t, 2H), 7.13 (t-d, 2H), 7.36 (m, 4H), 7.71-7.77 (m, 10H), 7.94 (d-d, 1H), 8.11 (d, 1H), 8.23 (d, 2H), 8.26 (m, 1H), 8.31 (m, 2H), 8.79-8.83 (m, 8H).

PN2: ^1H NMR (CDCl_3): δ -2.78 (s, 2H), 2.11 (t, 2H), 2.27 (t, 2H), 2.66 (t, 2H), 2.95 (t, 2H), 3.03 (t, 2H), 3.97 (t, 2H), 7.33 (d, 1H), 7.36 (t, 1H), 7.52 (m, 2H), 7.59 (m, 2H), 7.71-7.76 (m, 10H), 8.02 (d-d, 1H), 8.19 (m, 4H), 8.24 (d, 2H), 8.78-8.84 (m, 8H).

PN3: ^1H NMR (CDCl_3): δ -2.77 (s, 2H), 2.17 (t, 2H), 2.30 (t, 2H), 2.45 (t, 2H), 2.72 (t, 2H), 2.99 (t,

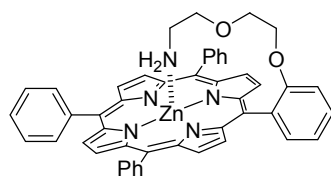
2H), 3.18 (t, 2H), 3.52 (t, 2H), 4.02 (t, 2H), 7.36 (m, 2H), 7.44 (m, 2H), 7.59 (m, 2H), 7.74-7.77 (m, 10H), 8.03 (d-d, 1H), 8.19 (d, 2H), 8.22 (d-d, 4H), 8.81-8.83 (m, 8H).

General procedure for the synthesis of OP. To the solution of **PN** in ethanol was added hydrazine hydrate (50 equiv.). The reaction mixture was stirred under reflux for 30 mins. After removing solvents by evaporation, dichloromethane was added to the residue and washed with diluted HCl, saturated NaHCO₃, brine, then dried over anhydrous MgSO₄. The solvent was evaporated to give the crude free base which was used without further purification. To the solution of free base in MeOH and CHCl₃ (v/v, 1:1) was added zinc acetate dihydrate (10 euqiv.). The reaction mixture was stirred at room temperature for 2h. After removing solvents by evaporation, the crude product was purified by column chromatography (CH₂Cl₂/MeOH, 20:1) to give the **OP** as a purple solid.

Freebase1: ¹H NMR (CDCl₃): δ -2.89 (s, 2H), 1.37 (t, 2H), 2.08 (t, 2H), 2.68 (t, 2H), 3.67 (t, 2H), 7.03 (m, 1H), 7.22 (m, 1H), 7.51 (m, 1H), 7.62 (m, 9H), 7.88 (m, 1H), 8.13 (m, 6H), 8.77-8.82 (m, 8H).

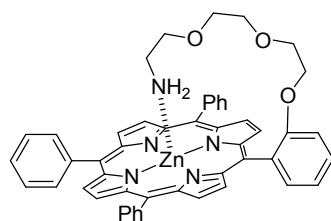
Freebase2: ¹H NMR (CDCl₃): δ -2.97 (s, 2H), 2.61 (t, 2H), 3.27 (t, 2H), 3.43 (t, 2H), 3.53 (t, 2H), 3.72 (t, 2H), 4.11 (t, 2H), 7.03 (m, 1H), 7.14 (m, 1H), 7.43 (m, 1H), 7.55 (m, 9H), 7.78 (m, 1H), 8.02 (m, 6H), 8.63-8.68 (m, 8H).

Freebase3: ¹H NMR (CDCl₃): δ -2.82 (s, 2H), 2.81 (t, 2H), 3.44 (t, 2H), 3.52 (t, 2H), 3.57 (t, 2H), 3.65 (t, 2H), 3.72 (t, 2H), 3.91 (t, 2H), 4.31 (t, 2H), 7.03 (m, 1H), 7.24 (m, 1H), 7.52 (m, 1H), 7.60 (m, 9H), 7.87 (m, 1H), 8.12 (m, 6H), 8.61-8.65 (m, 8H).



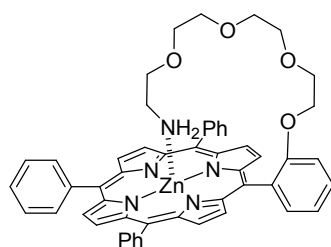
OP1

M.p. > 250 °C. ¹H NMR (CDCl₃): δ -4.13 (s, 2H), -1.60 (t, 2H), 0.56 (t, 2H), 2.48 (t, 2H), 3.61 (t, 2H), 7.14 (d-d, 1H), 7.60 (t-d, 1H), 7.72-7.84 (m, 10H), 8.25 (d-d, 2H), 8.37 (m, 2H), 8.43 (m, 2H), 8.65 (d-d, 1H), 8.96-8.99 (m, 8H). ¹³C NMR (CDCl₃): δ 35.8, 66.9, 68.3, 70.7, 115.5, 115.8, 120.6, 121.0, 126.4, 126.5, 126.7, 127.3, 127.4, 129.8, 131.5, 131.6, 131.7, 132.1, 133.2, 134.7, 134.9, 143.7, 143.8, 150.0, 150.1, 150.2, 150.3, 160.2. MS (MALDI): m/z 779.6 [M]⁺. Anal. Calcd for C₄₈H₃₇N₅O₂Zn: C, 73.80; H, 4.77; N, 8.96. Found: C, 73.92; H, 4.98; N, 8.63.



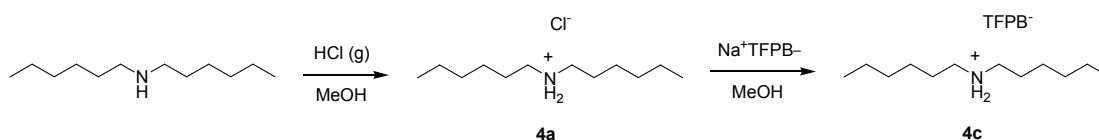
OP2

M.p. > 250 °C. ^1H NMR (CDCl_3): δ -4.10 (s, 2H), -1.78 (t, 2H), 0.48 (t, 2H), 2.13 (t, 2H), 2.71 (t, 2H), 3.06 (t, 2H), 4.04 (t, 2H), 7.27 (d-d, 1H), 7.41 (t-d, 1H), 7.74-7.76 (m, 10H), 8.16 (m, 3H), 8.25 (m, 2H), 8.32 (m, 2H), 8.85-8.88 (m, 8H). ^{13}C NMR (CDCl_3): δ 36.6, 67.2, 69.0, 69.1, 69.5, 69.6, 112.6, 116.4, 120.0, 120.4, 120.7, 126.3, 126.4, 126.5, 126.7, 127.3, 127.4, 129.5, 131.5, 131.6, 131.8, 131.9, 133.2, 134.6, 134.7, 134.8, 135.0, 136.0, 143.8, 149.9, 150.0, 150.1, 150.2, 150.6, 159.3. MS (MALDI): m/z 823.6 $[\text{M}]^+$. Anal. Calcd for $\text{C}_{50}\text{H}_{41}\text{N}_5\text{O}_3\text{Zn}$: C, 72.77; H, 5.01; N, 8.49. Found: C, 73.03; H, 5.34; N, 8.08.



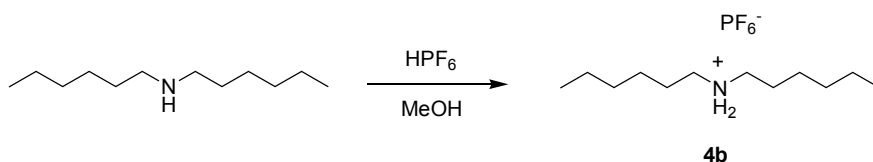
OP3

M.p. > 250 °C. ^1H NMR (CDCl_3): δ -3.87 (s, br, 2H), -1.97 (t, 2H), 0.73 (t, 2H), 2.14 (t, 2H), 2.68 (m, 4H), 2.75 (t, 2H), 3.18 (t, 2H), 4.07 (t, 2H), 7.31 (d-d, 1H), 7.35 (t-d, 1H), 7.70-7.77 (m, 10H), 7.99 (d-d, 1H), 8.17 (d-d, 2H), 8.21 (m, 2H), 8.31 (m, 2H), 8.81-8.85 (m, 8H). ^{13}C NMR (CDCl_3): δ 36.8, 68.0, 68.9, 69.3, 69.4, 70.0, 70.2, 70.9, 112.3, 116.4, 120.0, 120.4, 120.7, 126.4, 126.5, 126.6, 127.3, 127.4, 129.5, 131.6, 131.8, 133.2, 134.6, 134.7, 134.8, 134.9, 135.0, 136.1, 143.9, 149.9, 150.0, 150.1, 150.6, 158.9. MS (MALDI): m/z 867.7 $[\text{M}]^+$. Anal. Calcd for $\text{C}_{52}\text{H}_{45}\text{N}_5\text{O}_4\text{Zn}$: C, 71.84; H, 5.22; N, 8.06. Found: C, 72.11; H, 5.36; N, 7.73.



4a: To the solution of di-*n*-hexylamine (2 ml) in anhydrous methanol (15 ml) was bubbled dry HCl gas for 2.5 h. After removing methanol by evaporation, the residue was recrystallized from anhydrous $\text{CHCl}_3/\text{Et}_2\text{O}$ to give the **4a** as a white solid (3.46 g, 91%). ^1H NMR (CDCl_3): δ 0.85 (t, 6H), 1.32 (m, 12H), 1.85 (m, 4H), 2.88 (t, 4H), 9.44 (s, br, 2H).

4c: To the solution of **4a** (0.115 g, 0.52 mmol) in anhydrous methanol (3 ml) was added a solution of sodium tetrakis[3,5-bis(trifluoromethyl)phenyl]borate (0.594 g, 0.67 mmol) in anhydrous methanol (2 ml). The reaction mixture was stirred overnight at room temperature. After removing methanol by evaporation, the brown residue was washed with deionized water and filtered off, then dried under vacuum to give the **4c** as a brown solid (0.485 g, 89%). ^1H NMR (CDCl_3): δ 0.85 (t, 6H), 1.26 (m, 12H), 1.59 (m, 4H), 2.97 (q, 4H), 6.41 (s, br, 2H), 7.55 (s, 4H), 7.70 (s, 8H).



4b: To the solution of di-*n*-hexylamine (2ml) in anhydrous methanol (2 ml) was added a solution of hexafluorophosphoric acid (0.8 ml) in anhydrous methanol (2 ml). The reaction mixture was stirred at room temperature for 24h. After removing methanol by evaporation, the residue was recrystallized from anhydrous $\text{CHCl}_3/\text{Et}_2\text{O}$ to give the **4b** as a white solid (0.771 g, 22%). ^1H NMR (CDCl_3): δ 0.88 (t, 6H), 1.32 (m, 12H), 1.74 (m, 4H), 3.02 (q, 4H), 6.79 (s, br, 2H).

Preparation of single crystal of OP1: By evaporation of methanol in the outside jar, it diffused into the dichloromethane solution of **OP1** in the inside vial, which facilitated the crystallization of **OP1**. The apparatus shown in Fig. S1 was used.

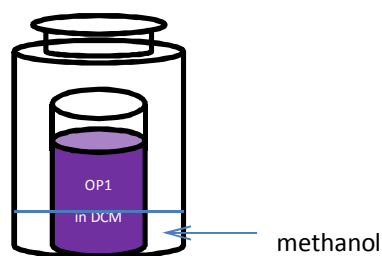


Fig. S1 The apparatus used for preparation of single crystal of **OP1**

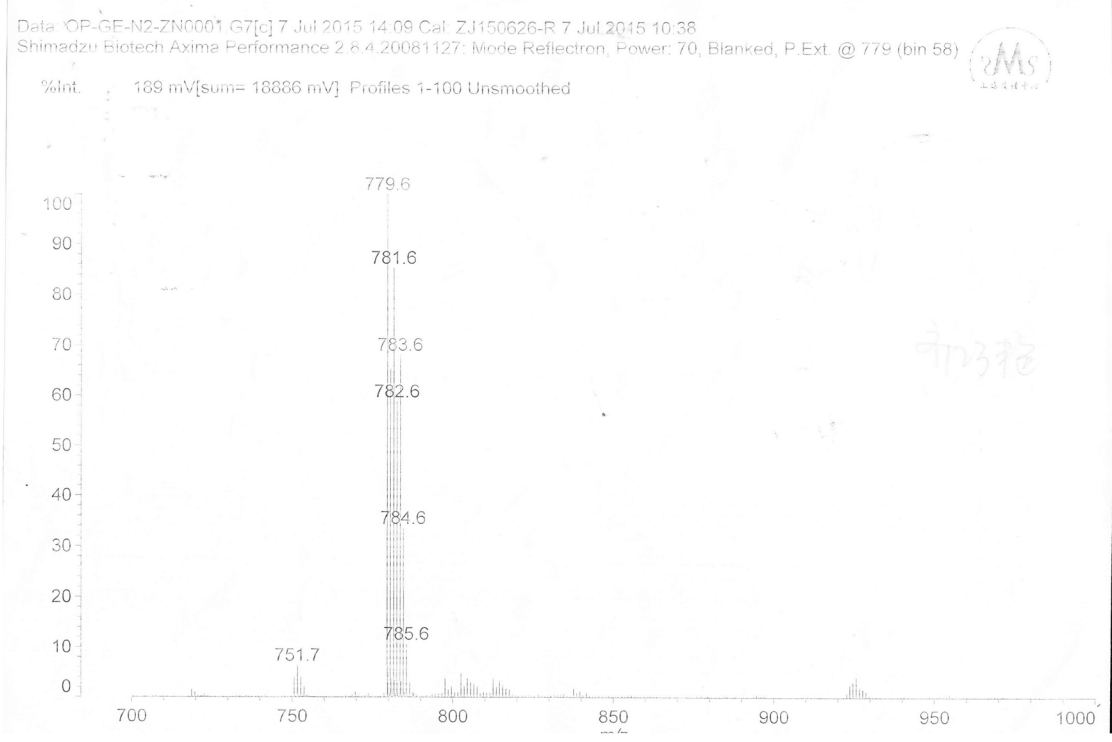


Figure S3. MALDI MS of OP1.

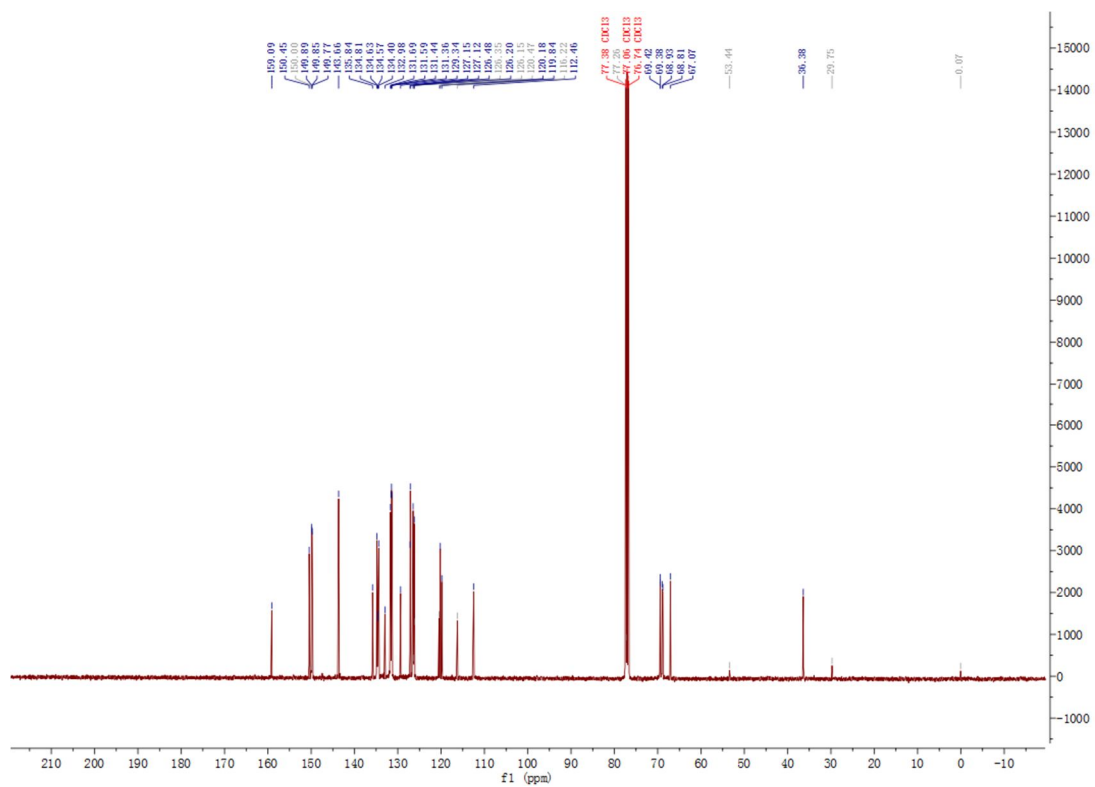
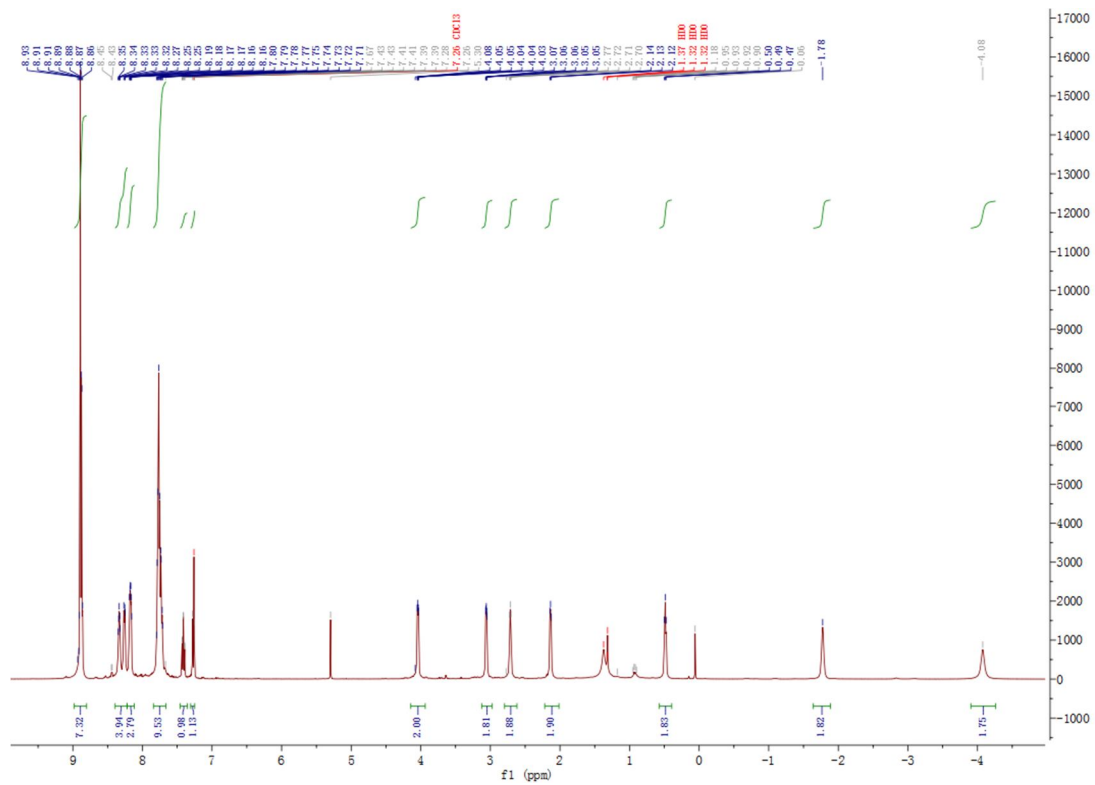


Figure S4. ¹H NMR and ¹³C NMR spectra of OP2.

Data: OP-GE-N3-ZN0001-G5[c] 7 Jul 2015 14:07 Cal: ZJ150626-R 7 Jul 2015 10:38
Shimadzu Biotech Axima Performance 2.8.4.20061127: Mode Reflectron, Power: 70, Blanked, P.Ext. @ 823 (bin 59)



%Int. 21 mV[sum= 2102 mV] Profiles 1-100 Unsmoothed

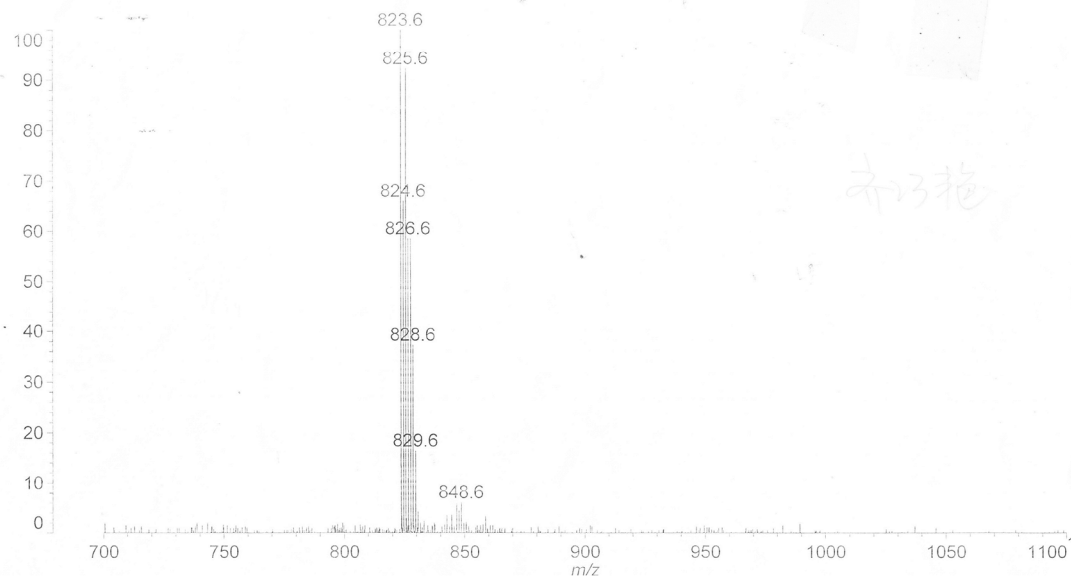


Figure S5. MALDI MS of **OP2**.

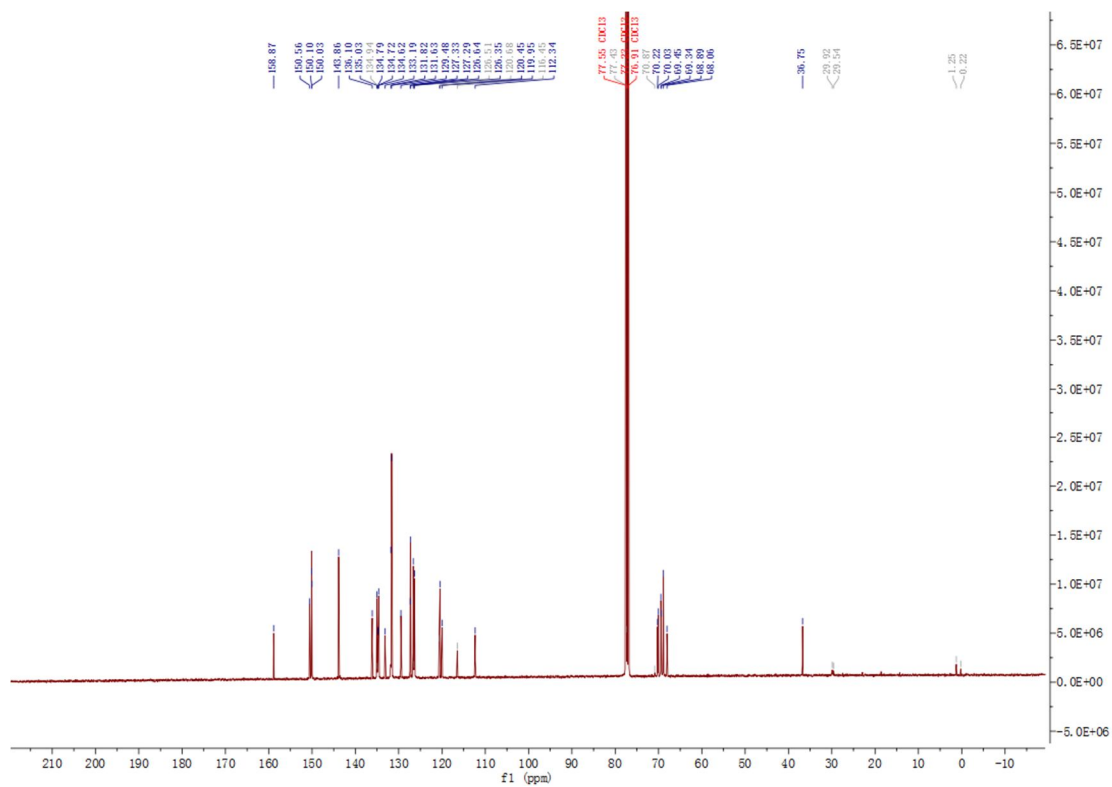
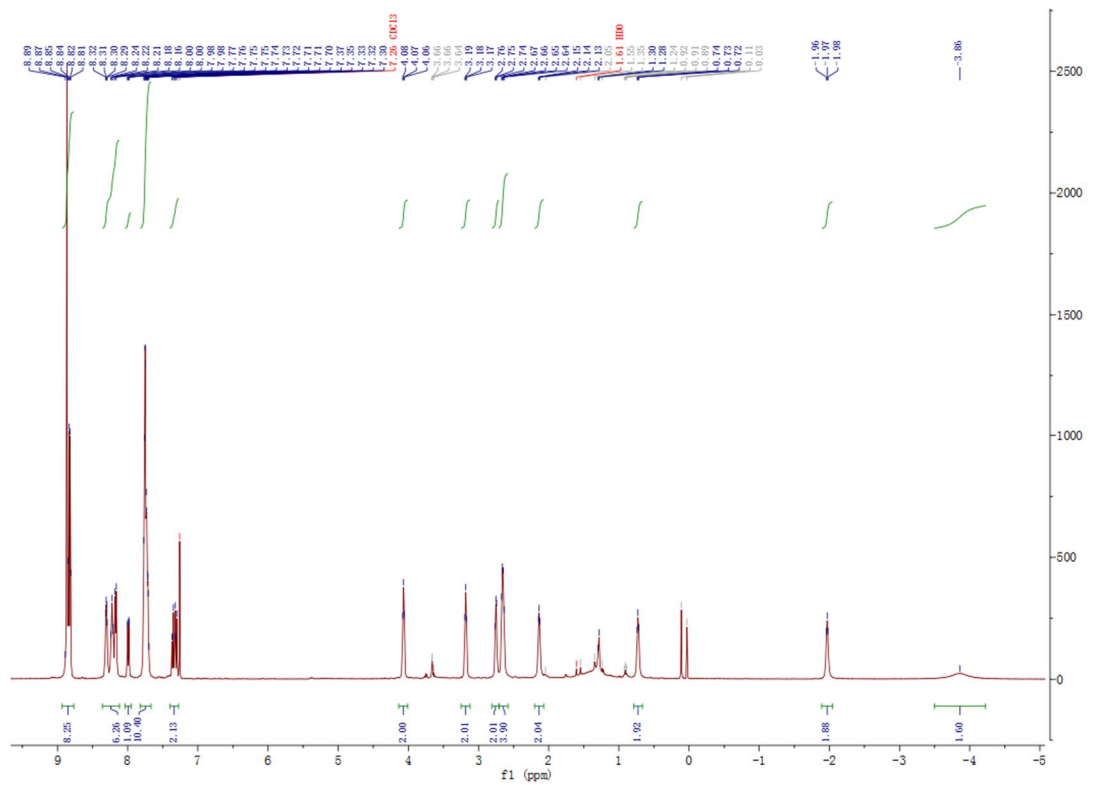


Figure S6. ¹H NMR and ¹³C NMR spectra of OP3.

Data: QP-GF-N4-ZN0001.G3[c] 7 Jul 2015 14:05 Cal: ZJ150626-R 7 Jul 2015 10:38
Shimadzu Biotech Axima Performance 2.8.4.20081127: Mode Reflectron; Power: 70, Blanked, P.Ext. @ 867 (bin 61)



%Int. 916 mV[sum= 91569 mV] Profiles 1-100 Unsmoothed

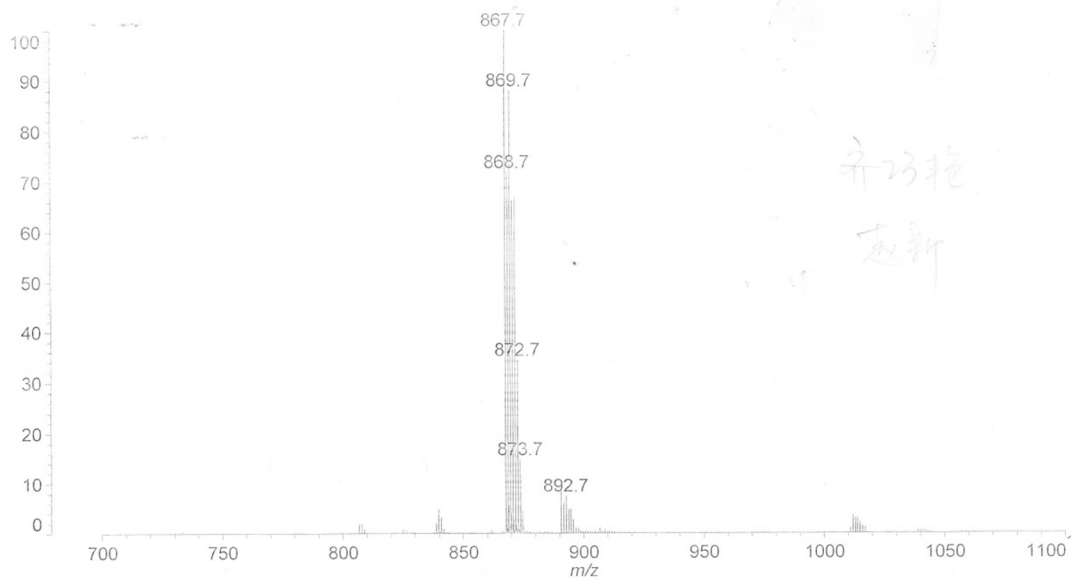


Figure S7. MALDI MS of OP3.

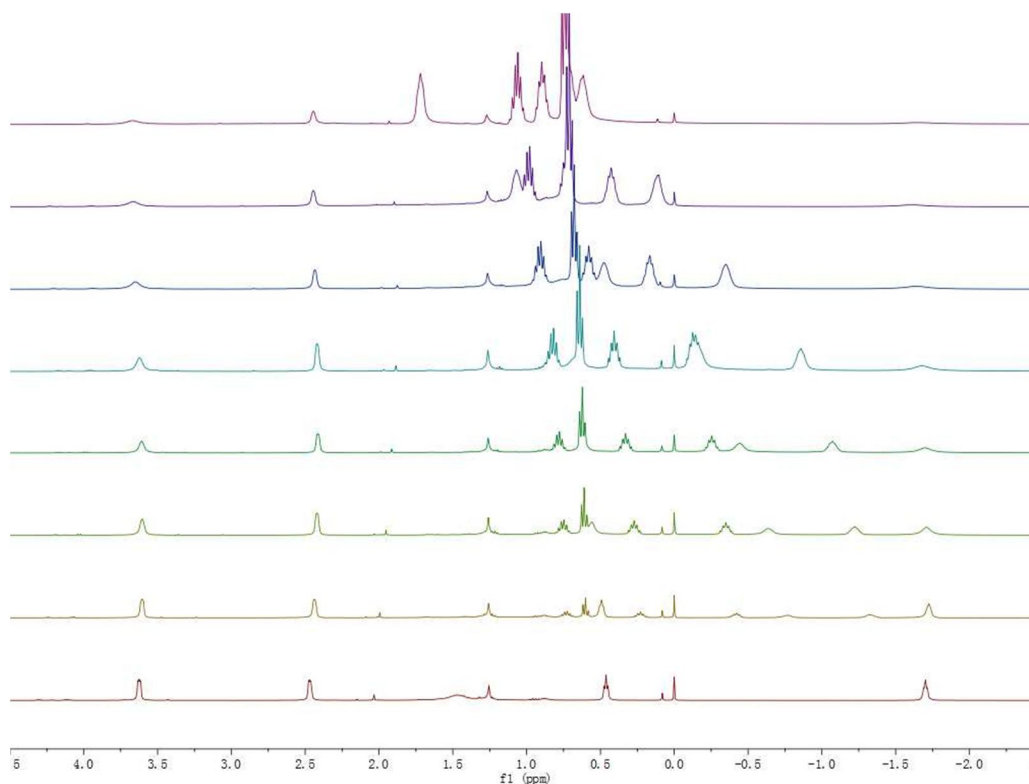


Figure S8. ^1H NMR titrations of host **OP1** (400 MHz, CDCl_3 , 298 K) with the increment of **4c**: (a) 0.00 equiv., (b) 0.25 equiv., (c) 0.50 equiv., (d) 0.75 equiv., (e) 1.00 equiv., (f) 1.50 equiv., (g) 2.00 equiv., (h) 3.00 equiv.

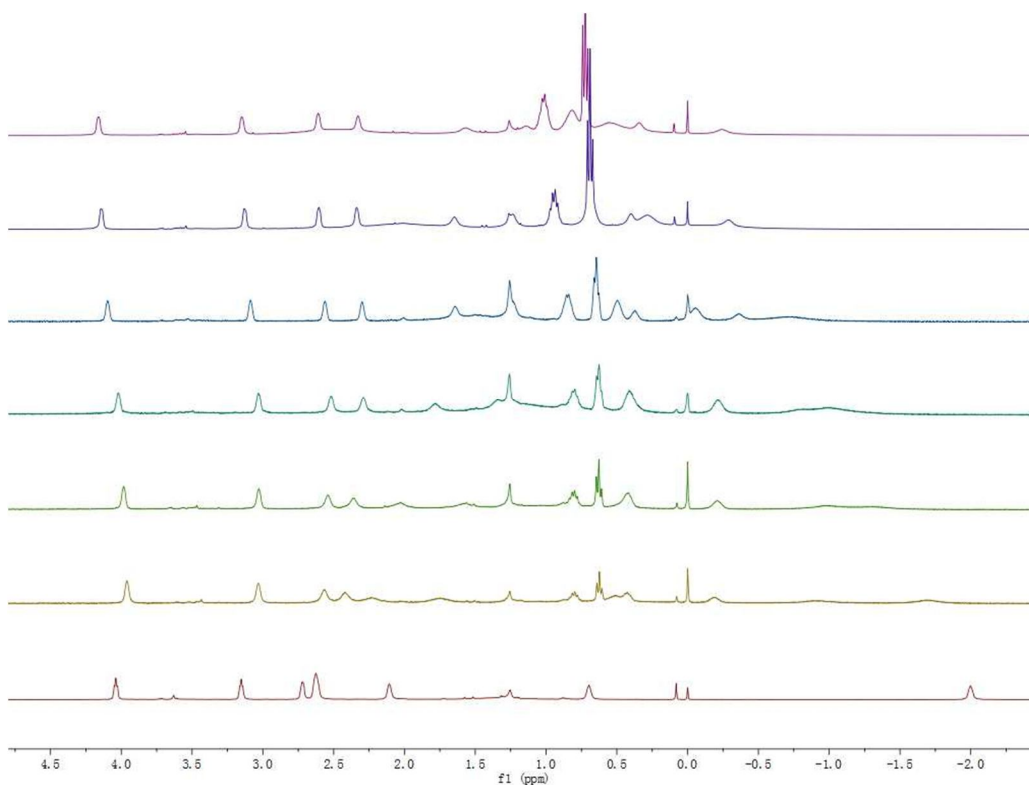


Figure S9. ^1H NMR titrations of host **OP3** (400 MHz, CDCl_3 , 298 K) with the increment of **4c**: (a) 0.00 equiv., (b) 0.25 equiv., (c) 0.50 equiv., (d) 0.75 equiv., (e) 1.00 equiv., (f) 1.50 equiv., (g) 2.00 equiv., (h) 3.00 equiv.

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T: ITMS + c ESI Full ms [200.00-2000.00]

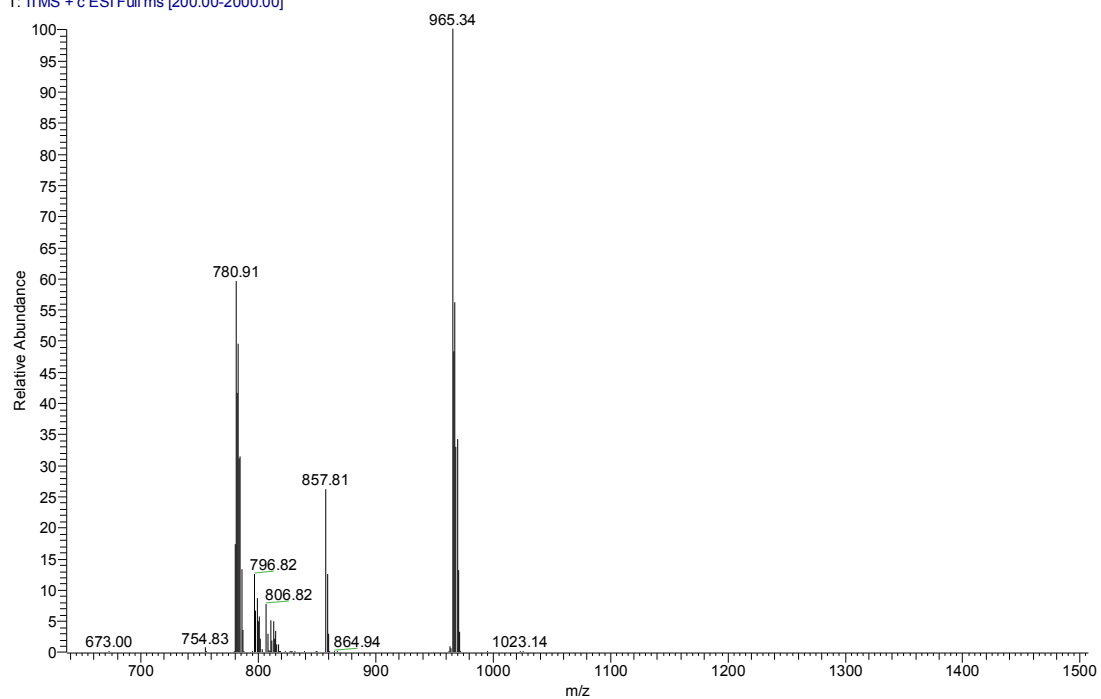


Figure S10. Mass spectrum of equimolar mixture of host **OP1** and **4c**.

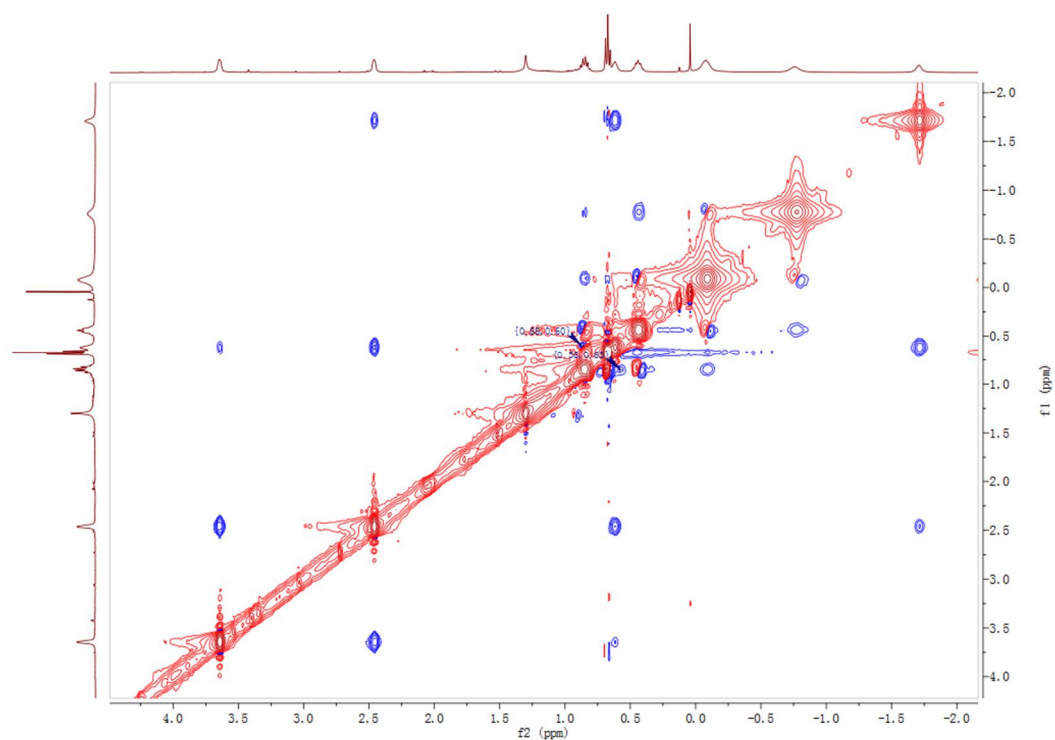


Figure S11. Partial 2D Noesy of equimolar mixture of host **OP1** and **4c**.

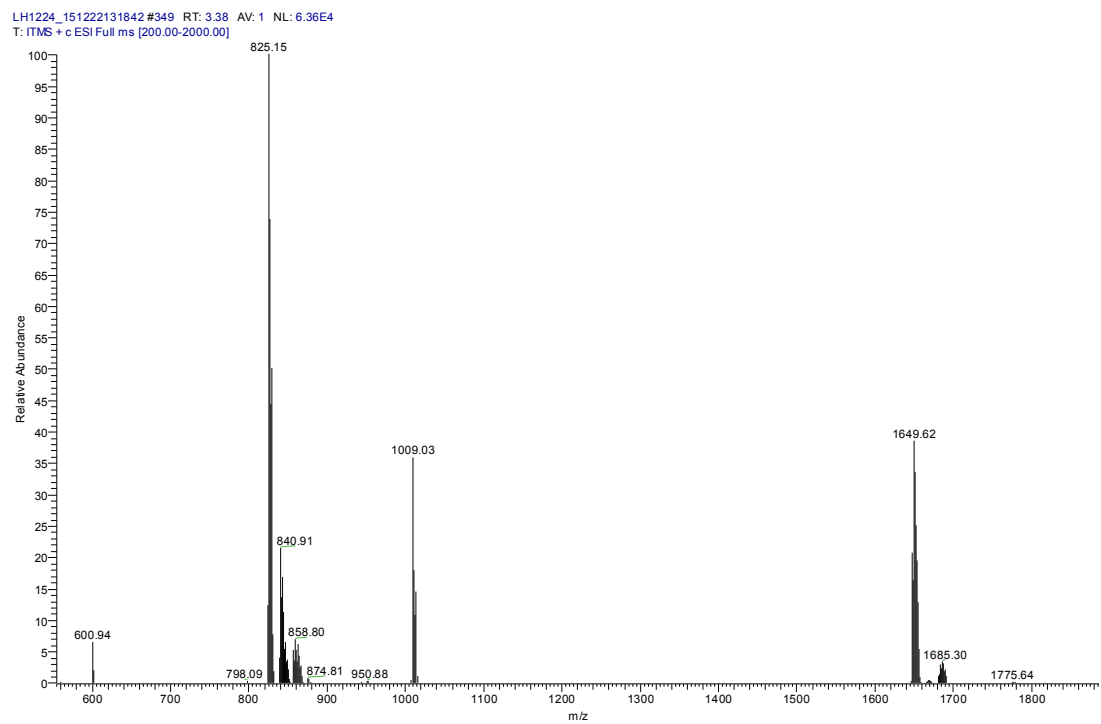


Figure S12. Mass spectrum of equimolar mixture of host **OP2** and **4c**.

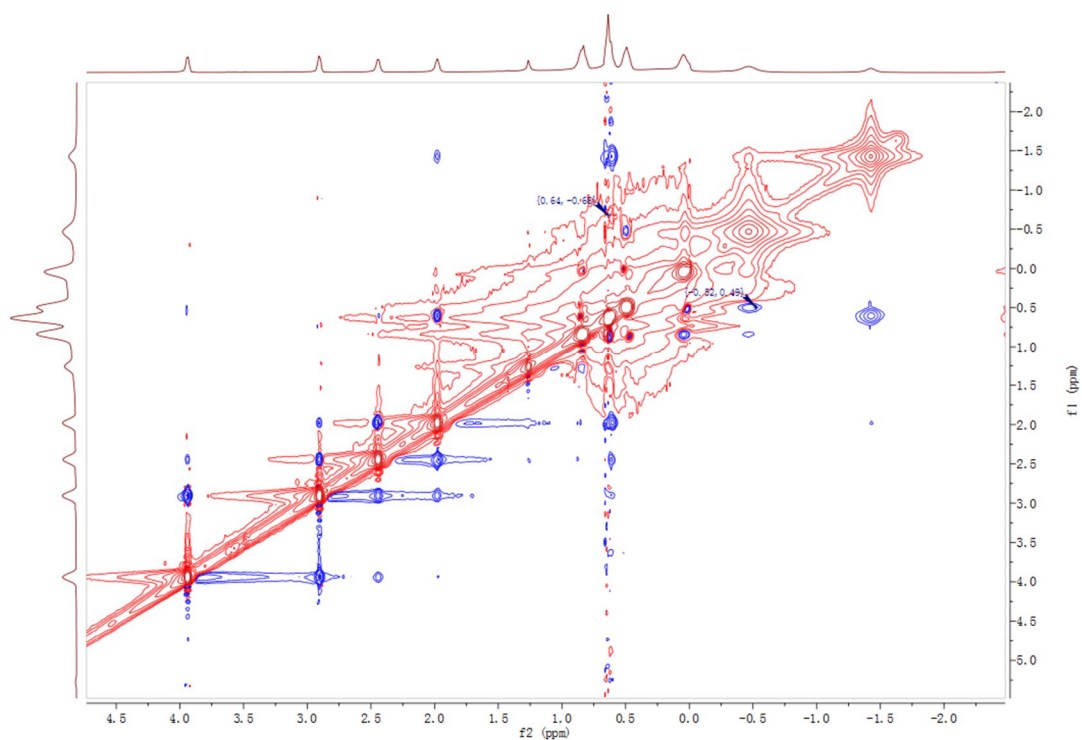


Figure S13. Partial 2D NMR of equimolar mixture of host **OP2** and **4c**.

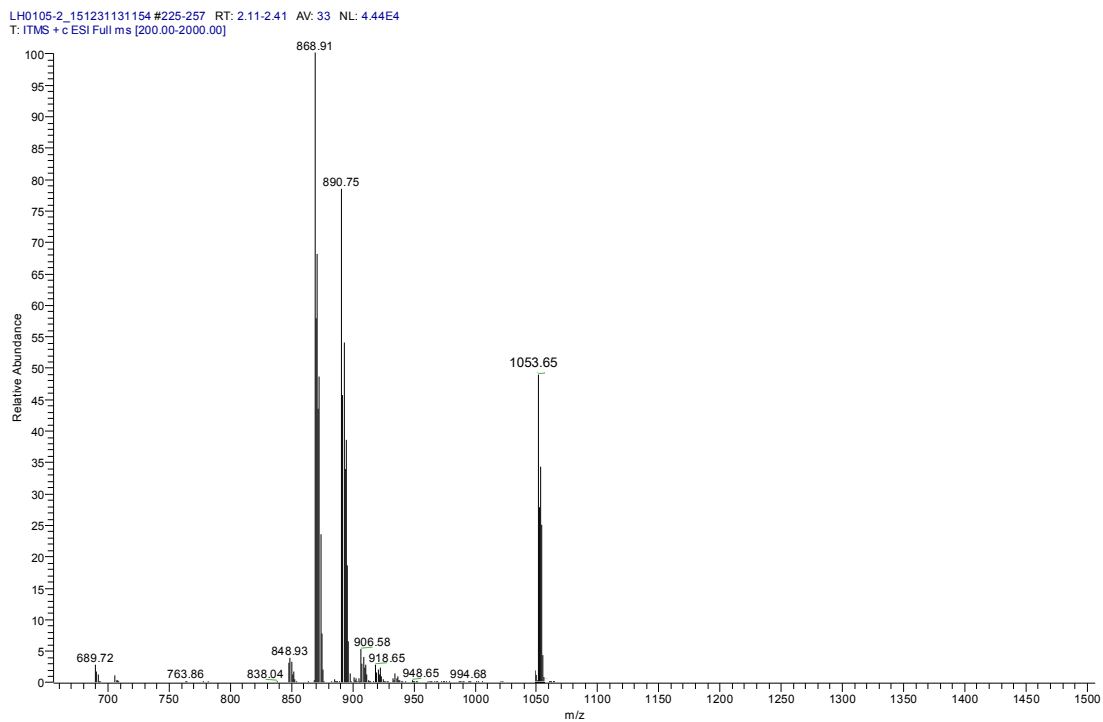


Figure S14. Mass spectrum of equimolar mixture of host **OP3** and **4c**.

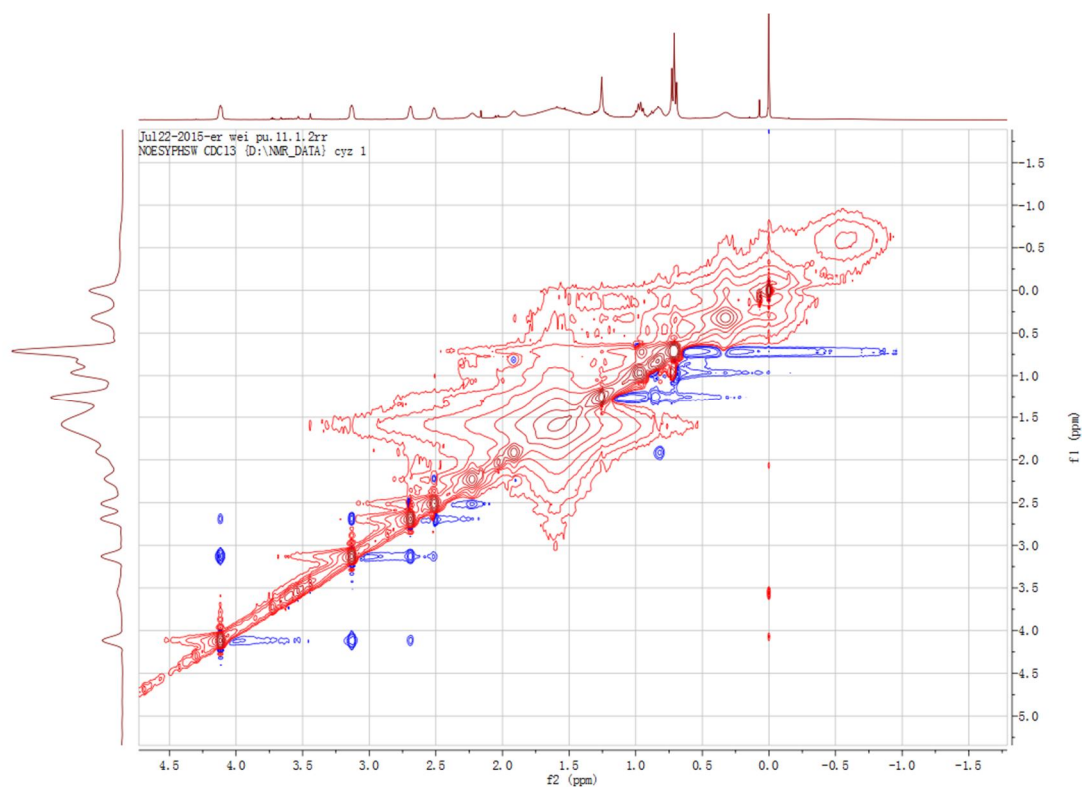


Figure S15. Partial 2D Noesy of equimolar mixture of host **OP3** and **4c**.

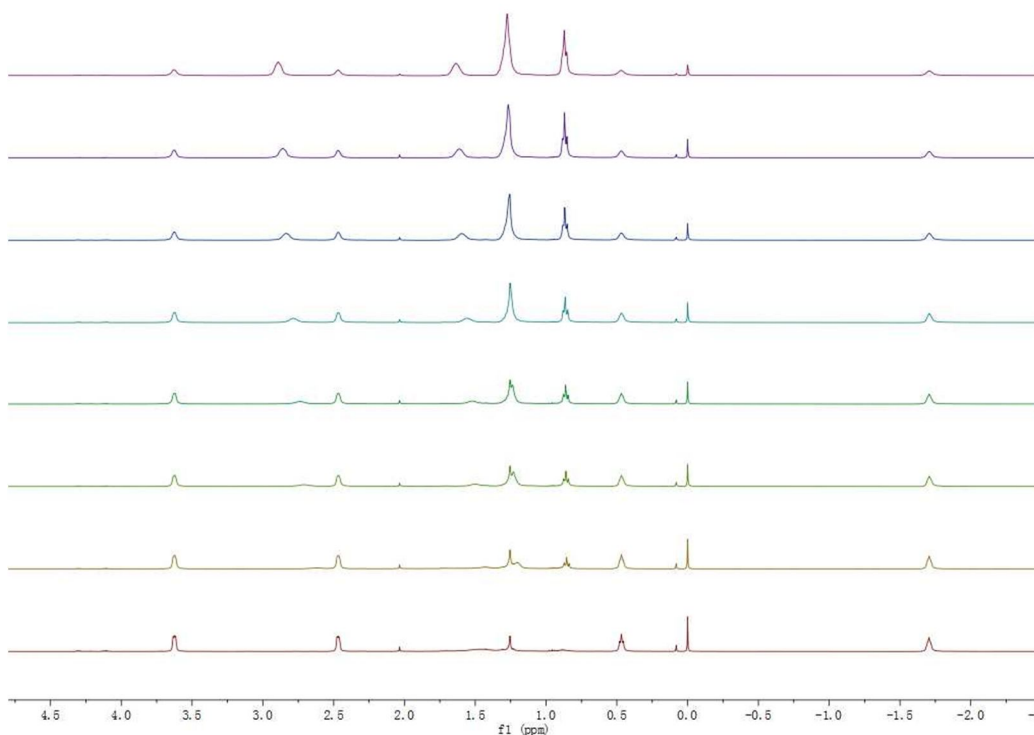


Figure S16. ^1H NMR titrations of host **OP1** (400 MHz, CDCl_3 , 298 K) with the increment of **4b**: (a) 0.00 equiv., (b) 0.25 equiv., (c) 0.50 equiv., (d) 0.75 equiv., (e) 1.00 equiv., (f) 1.50 equiv., (g) 2.00 equiv., (h) 3.00 equiv.

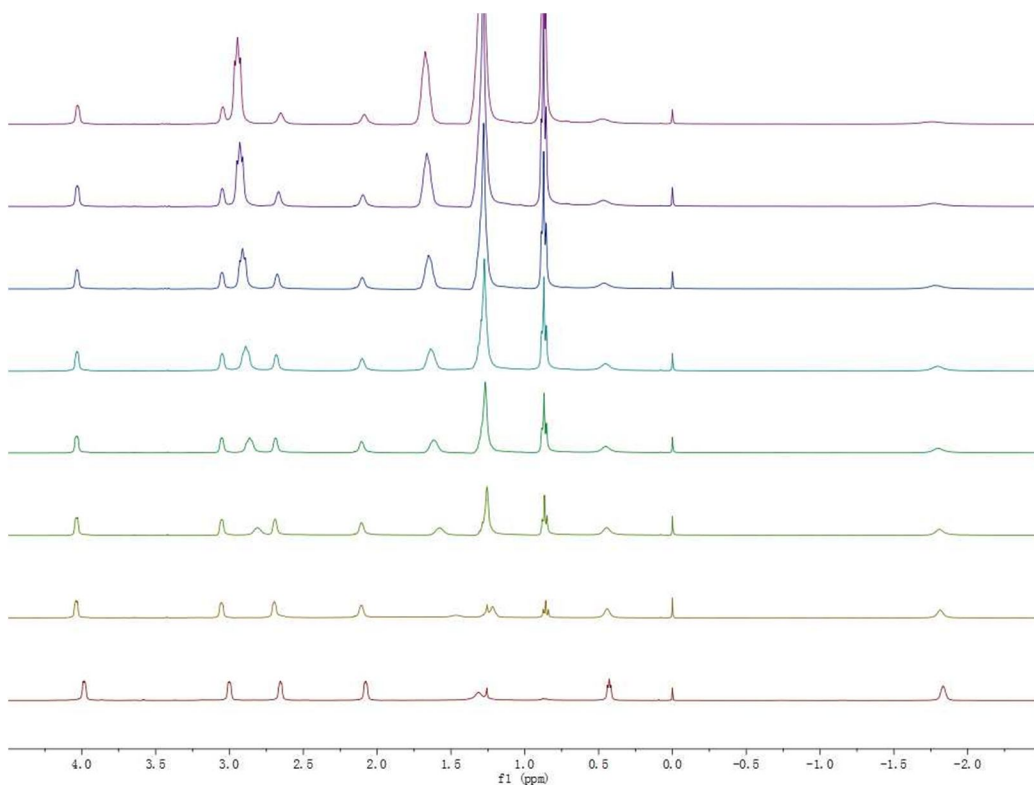


Figure S17. ^1H NMR titrations of host **OP2** (400 MHz, CDCl_3 , 298 K) with the increment of **4b**: (a) 0.00 equiv., (b) 0.25 equiv., (c) 0.50 equiv., (d) 0.75 equiv., (e) 1.00 equiv., (f) 1.50 equiv., (g) 2.00 equiv., (h) 3.00 equiv.

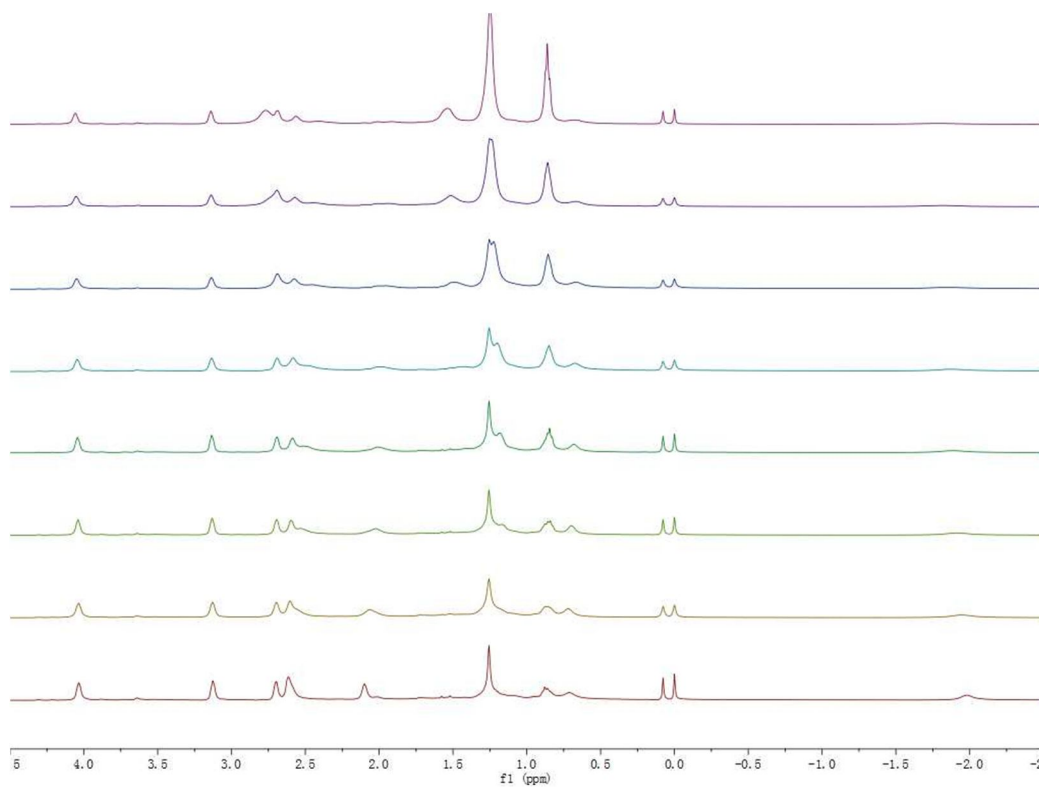
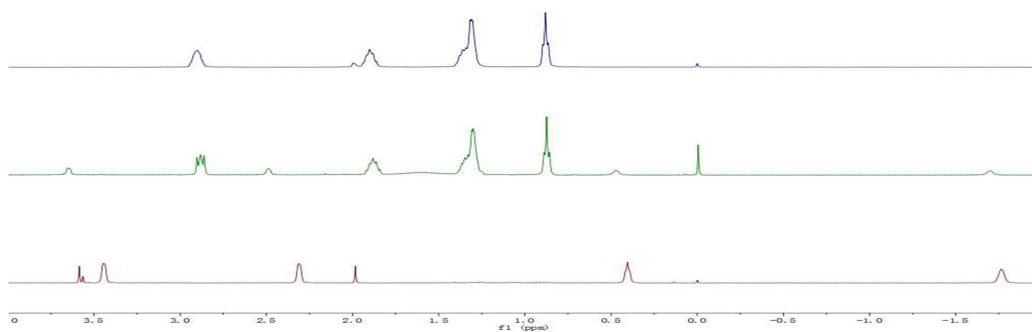
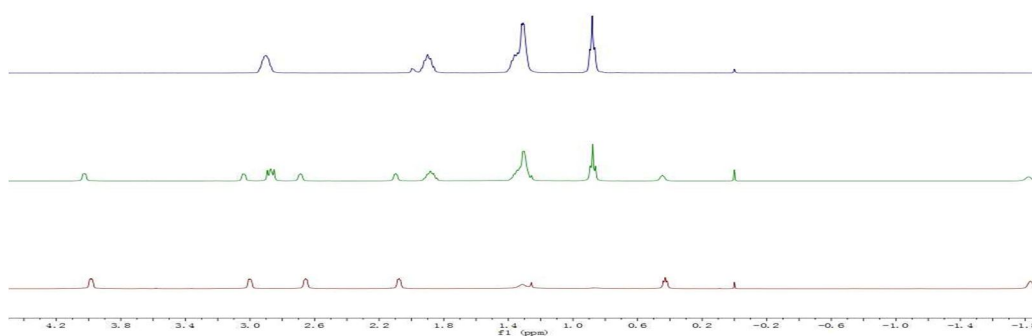


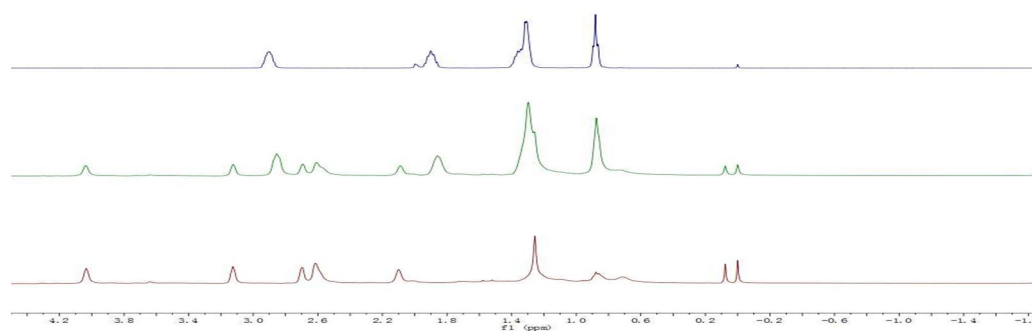
Figure S18. ¹H NMR titrations of host **OP3** (400 MHz, CDCl₃, 298 K) with the increment of **4b**: (a) 0.00 equiv., (b) 0.25 equiv., (c) 0.50 equiv., (d) 0.75 equiv., (e) 1.00 equiv., (f) 1.50 equiv., (g) 2.00 equiv., (h) 3.00 equiv.



OP1+4a



OP2+4a



OP3+4a

Figure S19. Partial ^1H NMR spectra of equimolar mixture of tailed porphyrin **OP** and **4a** (400 M Hz, CDCl_3 , 298 K).

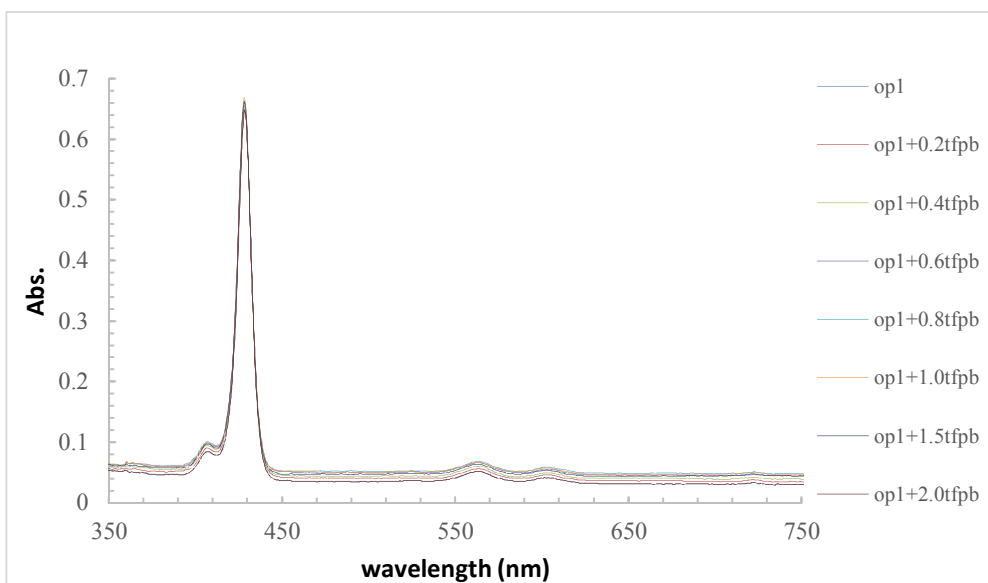
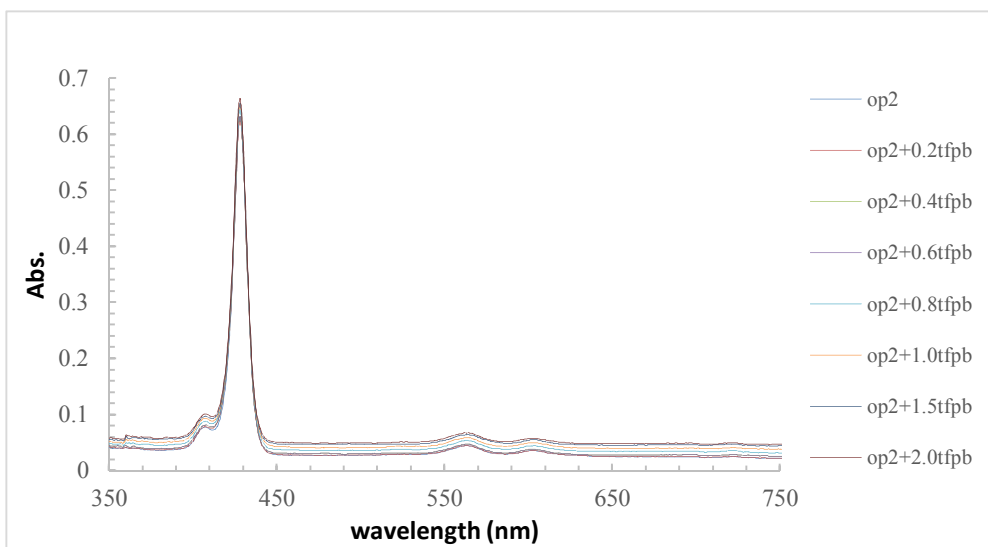
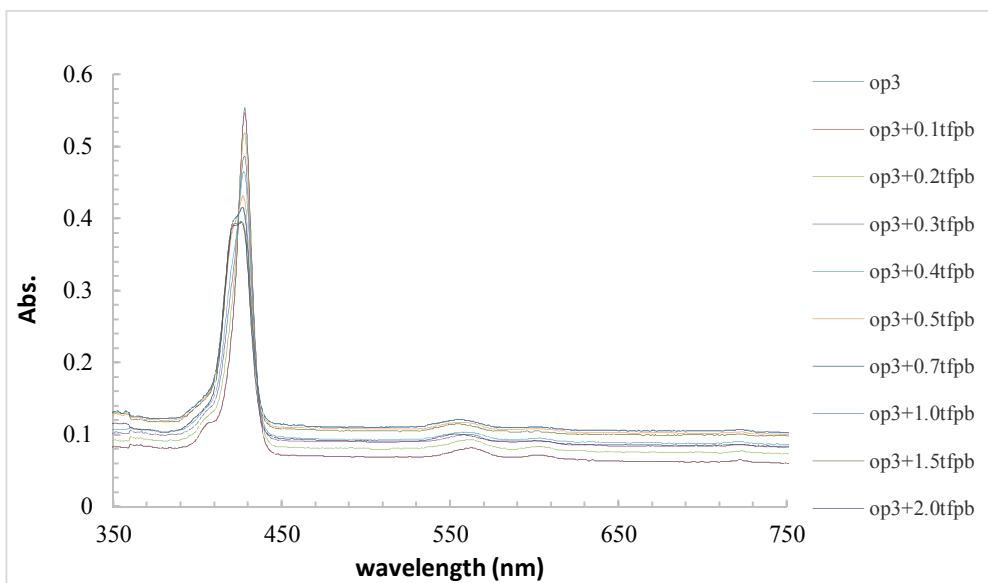


Figure S20. UV-Vis titration of host **OP** (1×10^{-6} mol/L in CH_2Cl_2 at 298 K) with the increment of **4c**.

References

- (1) Connors, K. A. *Binding Constants: The measurement of Molecular Complex Stability*; Wiley: New York, 1987.
- (2) Huang, F.; Gibson, H. W.; Bryant, W. S.; Nagvekar, D. S.; Fronczek, F. R. *J. Am. Chem. Soc.* **2003**, *125*, 9367-9371.
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- (4) Lukyanenko, N. G.; Kirichenko, T. I.; Scherbakov, S. V. *J. Chem. Soc., Perkin Trans. 1*, **2002**, 2347-2351.