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Supporting information

Organocatalytic Knoevenagel Condensation by Chiral C₂-Symmetric Tertiary Diamines

Xiaoyu Gu^a, Yan Tang^a, Xiang Zhang^a, Zhibin Luo^b and Hongfei Lu^a*

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General Information:

Unless otherwise stated, all reagents were purchased from commercial suppliers and used without further purification. All solvents used in the reactions were distilled from appropriate drying agents prior to use. Reactions were monitored by thin layer chromatography on silica gel. Column chromatography was performed with silica gel 200-300 mesh. All ¹H NMR and ¹³C NMR spectra were recorded in CDCl₃ and CD₃OD solution and reported in ppm (δ). ¹H NMR spectra were referenced internally to the residual proton resonance in CDCl₃ (δ =7.26 ppm), or with tetramethylsilane (TMS, δ =0.00 ppm) as the internal standard. HPLC chromatograms of Knoevenagel condensates were obtained using a Shimadzu apparatus, LC-20AT Pump, SPD-10A UV/Vis Detector, SCL-10A System Controller, using a Chiralcel IA (4.6 mmØ×250 mmL, particle size 5µm) or a Chiralcel OD-H (4.6 mmØ×250 mmL, particle size 5µm).

1. Synthesis of the organocatalysts ^[1]:

1.1 General synthetic procedure for catalyst 1a:



(S, S)-1, 2-Diaminocyclohexane (11.4g, 0.1mol) was dissolved in formic acid 85% (40mL) and paraformaldehyde 95% (50mL) was added slowly at room temperature. The mixture was heated at reflux 6h. After cooling, the reaction mixture was made basic until PH=14 and extracted with ether. The combined organic layers were washed with brine, dried over Na_2SO_4 , filtered and concentrated under reduced pressure. The product **1a** was distilled to give a colorless liquid (12.8g 75%).

1.2 General synthetic procedure for other catalysts:



To take 1c as example for synthetic procedure. Drop-wise tert-butylacetyl chloride (14.8g, 0.11mol) was added to a cooled (0°C) solution of 1, 2-Diaminocyclohexane (5.7g, 0.05mol) in dry THF. The suspension was stirred in an ice bath for 30 min, then heated to 50°C. After stirring 12h at this temperature, the mixture was quenched with 1M aqueous sodium hydroxide until pH>12. The layers were separated and the organic phase was washed with brine, drived over Na₂SO₄ filtered and concentrated in vacuum. The residue was dried adequately under reduced pressure to give A as a white solid. This material was pure enough to be used in the next step without further purification. To a cooled suspension of A (9.3g, 0.03mol) and NaBH₄ (4.5 g, 0.12mol) in dry THF (60mL) at 0°C was added drop-wise a solution of BF₃THF (37.8g, 0.135mol) in 20mL of dry THF over 30 min. The reaction mixture was heated to reflux slowly and reacted for 72h. The suspension was cooled to room

temperature, poured into crushed ice and extracted with either. The combined organic layer was dried over Na_2CO_3 , filtered and concentrated to give the crude diamine B as a solid, which was methylated without purification. The dimine B (6.8g, 0.24mol) was dissolved in formic acid 85% (35mL), paraformaldehyde 95% (3.0g) was added and the mixture was refluxed 8h. After basification and extraction with ether, the organic layer was drived over Na_2SO_4 , filtered and concentrated. The crude product was purified by recrystallization with EtOH-HCl to give the desired products.

1.3 Spectral details of organocatalysts^[1]:

1a: ¹H NMR (500 MHz, CDCl₃) δ 2.37 (dd, J = 8.8, 5.7 Hz, 2H), 2.26 (s, 12H), 1.84–1.79 (m, 2H), 1.74–1.69 (m, 2H), 1.10 (dd, J = 16.3, 8.6 Hz, 4H). [α]²⁰_D = -62.9 (c 1.05, CHCl₃).

1b: ¹H NMR (500 MHz, CDCl₃) δ 2.35–2.32 (m, 2H), 2.30 (d, J = 4.5 Hz, 4H), 2.28 (s, 6H), 1.75 (d, J = 13.1 Hz, 2H), 1.68–1.61 (m, 2H), 1.22 (ddd, J = 21.0, 9.7, 3.3 Hz, 2H), 1.09–0.99 (m, 2H), 0.88 (s, 18H). [α]²⁰_D = +6.3 (c 1.03, CHCl₃).



1c: ¹H NMR (500 MHz, CDCl₃) δ 2.46–2.54 (m, 6H), 2.26 (s, 6H), 1.78–1.81 (m, 2H), 1.72–1.73 (m, 2H), 1.40 (m, 4H), 1.11–1.20 (m, 4H), 0.91 (s, 18H). [α]²⁰_D = -31.1 (c 1.02, CHCl₃).

1d: ¹H NMR (500 MHz, CDCl₃) δ 7.38 (d, J = 7.3 Hz, 4H), 7.27 (t, J = 7.4 Hz, 4H), 7.21 (t, J = 7.2 Hz, 2H), 3.74 (d, J = 13.3 Hz, 2H), 3.65 (d, J = 13.3 Hz, 2H), 2.62 (dd, J = 5.7, 3.3 Hz, 2H), 2.21 (s, 6H), 1.92 (d, J = 12.8 Hz, 2H), 1.73 (dd, J = 6.4, 2.5 Hz, 2H), 1.25 (dd, J = 8.4, 5.9 Hz, 2H), 1.12 (t, J = 9.5 Hz, 2H). [α]²⁰_p = +7.22 (c 1.02, CHCl₃).



1e: ¹H NMR (400MHz, CDCl₃) δ 7.90-7.74 (m, 2H), 7.59-7.37 (m, 3H), 6.97 (s, 1H), 4.15-4.01 (m, 1H), 3.13 (dd, *J*=8.5, 3.9Hz, 1H), 1.71-1.42 (m, 8H).



1f: ¹H NMR (300MHz, CDCl₃) δ 7.74-7.67 (m, 4H), 7.48-7.42 (m, 2H), 7.42-7.36 (m, 4H), 3.93 (s, 2H), 2.52-2.49 (m, 2H), 1.91 (d, *J*=12.9Hz, 2H), 1.74 (d, *J*=7.9Hz, 2H), 1.51 (d, *J*= 9.1Hz, 2H), 1.29 (t, *J*=9.8Hz, 2H).



Ph 1g: 1H NMR (400MHz, CDCl₃) δ 7.81 (d, J=7.1Hz, 2H), 7.46 (dt, J=14.5, 7.0Hz, 3H), 6.67
(s, 1H), 4.33 (s, 1H), 2.56-2.46 (m, 1H), 2.26 (s, 6H), 2.05 (d, J=2.9Hz, 2H), 1.81 (dd, J=
6.0, 2.9Hz, 1H), 1.48 (d, J=3.1Hz, 1H), 1.42-1.22 (m, 4H).

1.4 The crystal structure of compound 1c:



2. General procedure for the catalytic enantioselective Knoevenagel condensation reaction:

2.1 General procedure for racemates preparation:



The aldehyde **2** (1 mmol), malonate **3** (1.1 mmol) and piperidine catalyst (0.1mmol) were dissolved in EtOH (3mL) / HAc (0.1mL). After reflowing for 8h, the reaction mixture was poured to water and organic layer was extracted with ethyl acetate. The organic fractions were dried over anhydrous MgSO₄ and concentrated under reduced pressure. Purification by silica gel column chromatography afforded the racemic compound **4**.

2.2 General procedure, Spectral details for the asymmetric Knoevenagel condensation reaction:



The catalyst **1c** (0.01mmol) and aldehyde **2** (0.1mmol) were dissolved in CH₃CN (2.0mL). The reaction mixture was stirred for 10min and malonate **3** (1.0mmol) was added at the same temperature. After heating to 60° C and vigorous stirring for 168h, the reaction mixture was poured to water (3mL) and extracted with ethyl acetate (3 × 15mL). The organic fractions were dried over anhydrous MgSO₄ and concentrated under reduced pressure. Purification by flash column chromatography afforded **4a-l** as desired products.

 $\begin{array}{l} \textbf{4a}^{[2]:} \text{ colorless oil, yield: 75\%, }^{1}\text{H NMR} (300\text{MHz, CDCl}_{3}) \ \delta \ 7.37 (ddd, J=7.1, 4.3, 1.7\text{Hz}, 2\text{H}), 7.31 (s, 2\text{H}), 7.29-7.23 (m, 1\text{H}), 7.02 (d, J=10.8\text{Hz}, 1\text{H}), 4.37 (q, J=7.1\text{Hz}, 2\text{H}), 4.25 (q, J=7.1\text{Hz}, 2\text{H}), 3.92 (dq, J=10.8, 6.9\text{Hz}, 1\text{H}), 1.49 (d, J=6.9\text{Hz}, 3\text{H}), 1.38 (t, J=7.1\text{Hz}, 3\text{H}), 1.31 (t, J=7.1\text{Hz}, 3\text{H}). \text{HPLC: Daicel CHIRALCEL IA acetonitrile /methyl alcohol (99/1), flow rate: 0.4mL/min, <math>\lambda$ =210nm, (τ_{major} =19.567min, τ_{minor} =17.089min).

4b: colorless oil, yield: 73%, ¹H NMR (400MHz, CDCl₃) δ 7.35-7.30 (m, 2H), 7.28-7.25 (m, 3H), 6.93 (d, *J*=10.8Hz, 1H), 5.22 (p, *J*=6.3Hz, 1H), 5.07 (p, *J*=6.3Hz, 1H), 3.87 (dq, *J*=10.7, 6.9Hz, 1H), 1.45 (d, *J*=6.9Hz, 3H), 1.33 (dd, *J*=6.3, 4.4Hz, 6H), 1.29 -1.26 (m, 6H). ¹³C NMR (126MHz, CDCl₃) δ 150.81, 142.59, 128.72, 127.58, 127.12, 126.86, 68.94, 60.38, 39.46, 21.71, 20.23. HPLC: Daicel CHIRALCEL OD-H hexane/2-propanol (99/1), flow rate: 1mL/min, λ =220nm, (τ_{major} =6.085min, τ_{minor} =5.368min).

4c: colorless oil, yield: 74%, ¹H NMR (400MHz, CDCl₃) δ 7.25-7.19 (m, 2H), 7.03-6.98 (m, 2H), 6.86 (d, J=10.8Hz, 1H), 5.21 (m, 1H), 5.09-5.04 (m, 1H), 3.86 (dq, J=10.7, 6.9Hz, 1H), 1.43 (d, J=7.0Hz, 3H), 1.32 (t, J=6.0Hz, 6H), 1.26 (dd, J=6.2, 2.7Hz, 6H). ¹³C NMR (101MHz, CDCl₃) δ 164.02, 162.46, 148.99, 140.59, 130.78, 130.31, 127.88, 127.00, 119.73, 68.07, 37.84, 20.69, 19.11. HPLC: Daicel CHIRALCEL OD-H hexane/2-propanol (99/1), flow rate: 1mL/min, λ=220nm, (τ_{major}=4.726min, τ_{minor} =4.501min).



ÇOOiPr

4d: colorless oil, yield: 72%, ¹H NMR (400MHz, CDCl₃) δ 7.28 (dd, *J*=8.4, 6.3Hz, 2H), 7.19 (d, *J*=8.5Hz, 2H), 6.85 (d, *J*=10.7Hz, 1H), 5.21 (p, *J*=6.3Hz, 1H), 5.07 (p, *J*=6.3Hz, 1H), 3.86 (dq, *J*=10.7, 6.9Hz, 1H), 1.43 (d, *J*=7.0Hz, 3H), 1.32 (d, *J*=6.5Hz, 6H), 1.26 (d, *J*=6.7Hz, 6H). ¹³C NMR (101MHz, CDCl₃) δ 165.07, 163.50, 150.13, 141.06, 132.69, 128.84, 128.52, 127.96, 69.10, 38.81, 21.73, 20.18. HPLC: Daicel CHIRALCEL OD-H hexane/2-propanol (99/1), flow rate: 1mL/min, λ =220nm, (τ_{major} =5.209min, τ_{minor} =4.876min).

Br COOIPr 4e: 0 7.12

4e: colorless oil, yield: 74%, ¹H NMR (400MHz, CDCl₃) δ 7.46-7.42 (m, 2H), 7.16-7.12 (m, 2H), 6.85 (d, *J*=10.7Hz, 1H), 5.22 (dq, *J*=12.6, 6.3Hz, 1H), 5.10-5.04 (m, 1H), 3.84 (dq, *J*=10.8, 6.9Hz, 1H), 1.43 (d, *J*=7.0Hz, 3H), 1.33 (d, *J*=6.5Hz, 6H), 1.27-1.25 (m, 6H). ¹³C NMR (101MHz, CDCl₃) δ 165.12, 163.54, 150.48, 138.27, 138.24, 128.66, 128.58, 127.72, 115.61, 115.40, 69.04, 38.67, 21.71, 20.29. HPLC: Daicel CHIRALCEL OD-H hexane/2-propanol (99/1), flow rate: 1mL/min, λ =220nm, (τ_{major} =5.476min, τ_{minor} =5.009min).

4f: colorless oil, yield: 72%, ¹H NMR (400MHz, CDCl₃) δ 7.19-7.10 (m, 4H), 6.91 (d, *J*=10.8Hz, 1H), 5.22 (p, *J*=6.3Hz, 1H), 5.06 (p, *J*=6.3Hz, 1H), 4.12 (q, *J*=7.1Hz, 1H), 2.32 (s, 3H), 1.43 (d, *J*=6.9Hz, 3H), 1.33 (dd, *J*=6.3, 3.0Hz, 6H), 1.26-1.24 (m, 6H). ¹³C NMR (101MHz, CDCl₃) δ 165.23, 163.63, 151.07, 139.57, 129.40, 127.36, 127.00, 68.90, 39.10, 29.72, 21.73, 20.99, 20.23. HPLC: Daicel CHIRALCEL OD-H hexane/2-propanol (99/1), flow rate: 1mL/min, λ =220nm, (τ_{major} =5.334min, τ_{minor} =4.917min).

COOEt COOEt **4g**^[2]: colorless oil, yield: 72%, ¹H NMR (400MHz, CDCl₃) δ 7.25-7.20 (m, 2H), 7.01 (t, *J*=8.7Hz, 2H), 6.92 (d, *J*=10.8Hz, 1H), 4.33 (m, *J*=7.1Hz, 2H), 4.23 (m, *J*=7.1Hz, 2H), 3.87 (m, *J*=10.9Hz, 1H), 1.44 (d, *J*=7.0Hz, 3H), 1.35 (t, *J*=7.1Hz, 3H), 1.28 (t,

J=7.1Hz, 3H). HPLC: Daicel CHIRALCEL OD-H hexane/2-propanol (99/1), flow rate: 1mL/min, λ =220nm, (τ_{major} =7.244min, τ_{minor} =6.635min).

4h^[2]: colorless oil, yield: 72%, ¹H NMR (400MHz, CDCl₃) δ 7.31-7.26 (m, 2H), 7.21-7.18 (m, 2H), 6.91 (d, *J*=10.7Hz, 1H), 4.33 (q, *J*=7.1Hz, 2H), 4.23 (q, *J*=7.2 Hz, 2H), 3.87 (dq, *J*=10.7, 6.9Hz, 1H), 1.43 (d, *J*=6.9Hz, 3H), 1.34 (t, *J*=7.1Hz, 3H), 1.29 (d, *J*=7.3Hz, 3H). HPLC: Daicel CHIRALCEL OD-H hexane/2-propanol (99/1), flow rate: 1mL/min, λ =220nm, (τ_{major} =6.802min, τ_{minor} =6.385min).

4i: colorless oil, yield: 73%, ¹H NMR (400MHz, CDCl₃) δ 7.44 (d, *J*=8.5Hz, 2H), **5** COOET 7.14 (d, *J*=8.5Hz, 2H), 6.91 (d, *J*=10.8Hz, 1H), 4.32 (q, *J*=7.1Hz, 2H), 4.23 (dd, *J*=7.2, 2.1Hz, 2H), 3.86 (dd, *J*=10.7, 6.9Hz, 1H), 1.43 (d, *J*=6.9Hz, 3H), 1.36-1.33 (m, 3H), 1.32-1.29 (m, 3H). ¹³C NMR (101MHz, CDCl₃) δ 168.01, 150.92, 141.33, 131.33, 128.88, 127.66, 119.60, 61.77, 20.10, 16.46. HPLC: Daicel CHIRALCEL OD-H hexane/2-propanol (99/1), flow rate: 1mL/min, λ =220nm, (τ_{major} =7.977min, τ_{minor} =7.410min).

4 $\mathbf{j}^{[2]}$: colorless oil, yield: 70%, ¹H NMR (400MHz, CDCl₃) δ 7.20-7.09 (m, 4H), 6.97 (d, *J*=10.8Hz, 1H), 4.33 (q, *J*=7.1Hz, 2H), 4.23 (d, *J*=7.1Hz, 2H), 3.90-3.80 (m, 1H), 2.32 (s, 3H), 1.43 (d, *J*=6.9Hz, 3H), 1.34 (d, *J*=7.1Hz, 3H), 1.27 (d, *J*=1.6Hz, 3H). HPLC: Daicel CHIRALCEL OD-H hexane/2-propanol (99/1), flow rate: 1mL/min, λ =220nm, (τ_{major} =7.660min, τ_{minor} =6.718min).

4k^[2]: colorless oil, yield: 71%, ¹H NMR (400MHz, CDCl₃) δ 7.36-7.29 (m, 2H), 7.28 -7.20 (m, 3H), 7.04 (d, *J*=10.8Hz, 1H), 3.92-3.86 (m, 1H), 3.85 (s, 3H), 3.76 (d, *J*=3.8Hz, 3H), 1.45 (d, *J*=6.9Hz, 3H). HPLC: Daicel CHIRALCEL OD-H hexane/2-propanol (98/2), flow rate: 0.4mL/min, λ =230nm, (τ_{major} =13.397min, τ_{minor} =11.096min).

COOMe COOMe **41**: colorless oil, yield: 68%, ¹H NMR (400MHz, CDCl₃) δ 7.14 (s, 4H), 7.02 (d, *J*=10.9Hz, 1H), 4.12 (d, *J*=7.1Hz, 1H), 3.86 (s, 3H), 3.76 (s, 3H), 2.32 (s, 3H), 1.43 (d, *J*=6.9Hz, 3H). ¹³C NMR (101MHz, CDCl₃) δ 165.90, 153.10, 139.27, 136.63, 129.46, 126.96, 52.38, 39.27, 21.00, 20.24. HPLC: Daicel CHIRALCEL OD-H hexane/2-propanol (99/1), flow rate: 1mL/min, λ =220nm, (τ_{major} =10.320min, τ_{minor} =9.270min).

2.3 Chromatograms of racemic mixtures and condensates obtained by organocatalysts:



Chromatogram of racemic mixture of 4a



Chromatogram of 4a



Peak #	RetTime/ min	Height	Area	Area%
1	5.293	339525.844	1867389.875	51.562
2	6.026	284525.156	1754240.000	48.438
Total				100

Chromatogram of racemic mixture of **4b**



Chromatogram of 4b



Chromatogram of 4c



Chromatogram of 4d



Peak #	RetTime/ min	Height	Area	Area%
1	5.109	447233.813	2350263.000	50.664
2	5.418	405149.156	2288643.000	49.336
Total				100

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	Chromatogram	of rac	emic	mixture	of 4	e
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Peak #	RetTime/ min	Height	Area	Area%
1	5.009	322118.844	2623093.250	91.012
2	5.476	64789.281	259028.500	8.988
Total				100

Chromatogram of 4e



Peak #	RetTime/ min	Height	Area	Area%
1	4.909	227756.781	1482091.875	49.200
2	5.301	220025.875	1530298.375	50.800
Total				100

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Chromatogram of racemic mixture of 4f



4.5 4.55 4.6 4.65 4.7 4.75 4.8 4.85 4.9 4.95 5 5.05 5.1 5.15 5.2 5.25 5.3 5.35 5.4 5.45 5.5

Peak #	RetTime/ min	Height	Area	Area%
1	4.917	513338.500	4037205.750	96.418
2	5.334	45135.137	149984.797	3.582
Total				100

Chromatogram of 4f



Chromatogram of racemic mixture of 4g



Chromatogram of 4g



Peak #	RetTime/ min	Height	Area	Area%
1	6.427	556001.250	3577783.500	51.237
2	6.893	469305.219	3405034.250	48.763
Total				100

Chromatogram of racemic mixture of $\mathbf{4h}$



66.05.6.16.2.26.6.35.6.45.6.56.6.66.75.8.86.9.9577.05.7.15.2.25.2.35.4.45.5.55.6.65.7.75.8.85.9.958

Peak #	RetTime/ min	Height	Area	Area%
1	6.385	352630.125	2208220.000	87.323
2	6.802	30105.619	320612.500	12.677
Total				100

Chromatogram of 4h



Chromatogram of 4i



Peak #	RetTime/ min	Height	Area	Area%
1	6.827	514620.844	3727768.000	50.573
2	7.619	444577.500	3643328.500	49.427
Total				100

.

Chromatogram of racemic mixture of 4j



Peak #	RetTime/ min	Height	Area	Area%
1	6.718	273170.281	3334752.750	92.920
2	7.660	39300.637	254106.156	7.080
Total				100

Chromatogram of 4j



Chromatogram of 4k



Chromatogram of 41

Reference:

- [1] J. Kizirian, N. Cabello, L. Pinchard, J. Caille, A. Alexakis, *Tetrahedron*. 2005, 61, 8939-8946.
- [2] A. Lee, A. Michrowska, S. Sulzer-Mosse, B. List, Angew. Chem., Int. Ed. 2011, 50, 1707.



3. Copy of original ¹H NMR spectra of all products and ¹C NMR spectra of 4b-f, 4i and 4l









































