

Supplementary Information for:  
**Bioconjugatable, PEGylated Hydroporphyrins for Photochemistry and  
Photomedicine. Narrow-Band, Red-Emitting Chlorins**

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**1. Click Reaction Conditions**

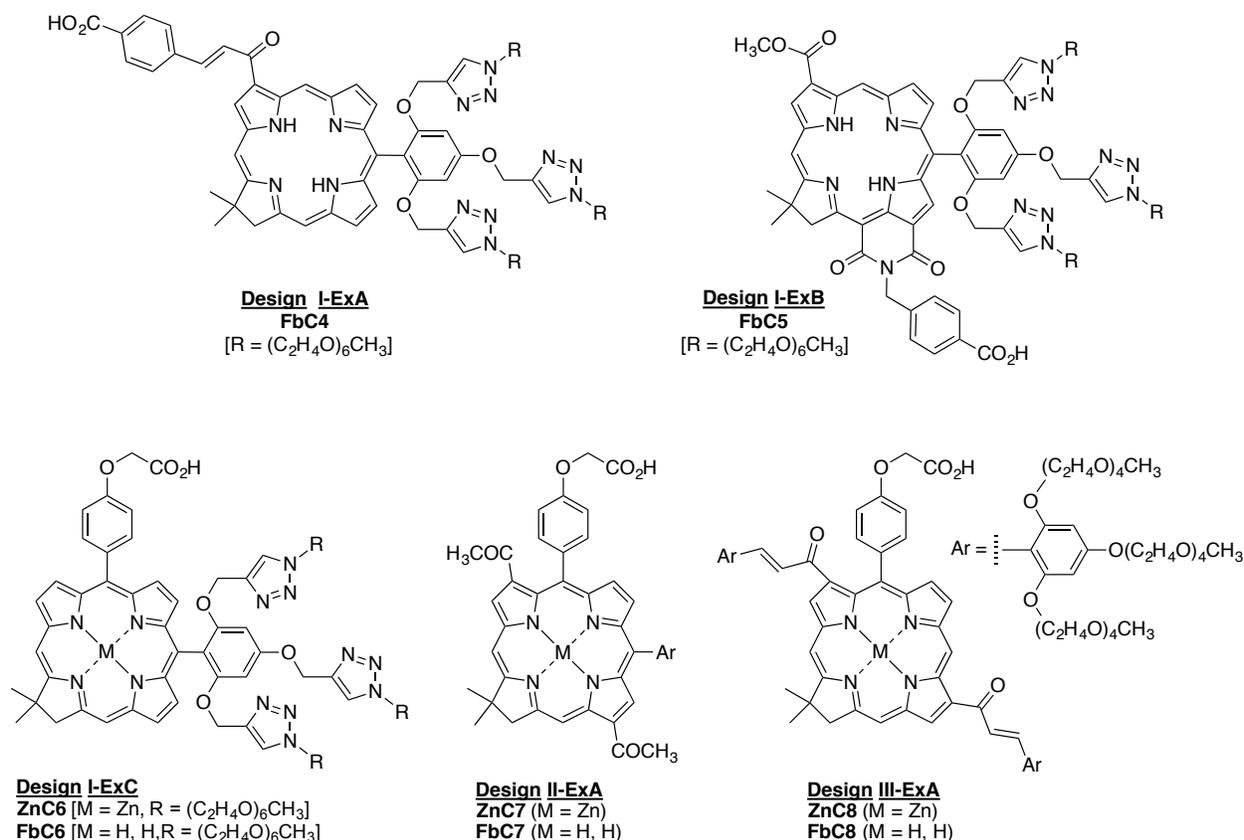
**Table S1.** Conditions for click reaction of chlorin **6** with PEG-azide **7** to form **ZnC1**.

<b>Entry</b>	<b>Cu (I) amount</b>	<b>Solvent</b>	<b>Temp., Time</b>	<b>Result<sup>a</sup></b>
1	0.03 equiv	Butanol/H <sub>2</sub> O (2:1)	RT, 16 h	<b>6</b>
2	0.03 equiv	CH <sub>3</sub> CN/H <sub>2</sub> O (2:1)	RT, 16 h	<b>6</b>
3	0.1 equiv	CH <sub>3</sub> CN/H <sub>2</sub> O (1:1)	40 °C, 36 h	mono-, di-, tri-derivatized chlorins, minor <b>6</b>
4	0.2 equiv	CH <sub>3</sub> CN/H <sub>2</sub> O (2:1)	40 °C, 16 h	Same as entry 3
5	1 equiv	DMSO/H <sub>2</sub> O (4:1)	40 °C, 16 h	<b>ZnC1</b>

<sup>a</sup>Results are based on TLC analysis and MALDI-MS.

**2. Exploratory Molecular Designs**

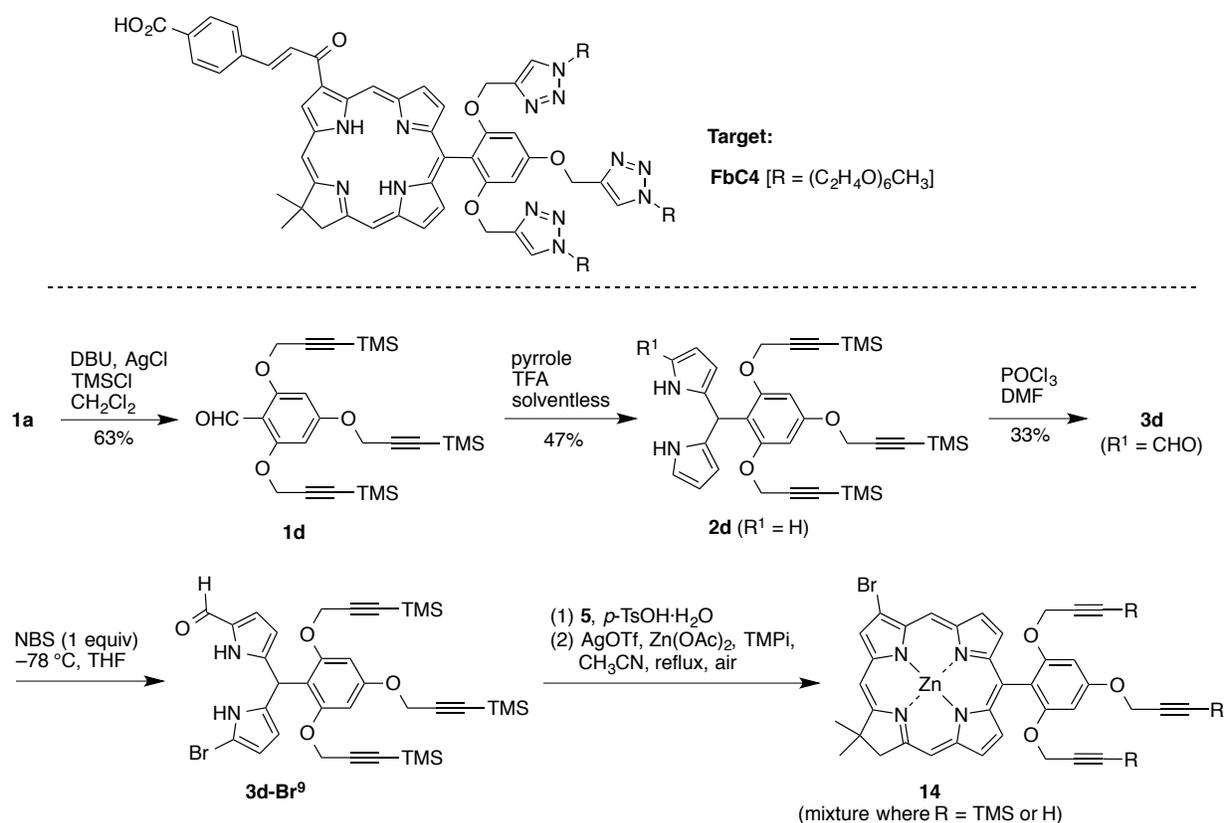
Several targets that were pursued are shown in Chart S1. The limitations of each design with regard to syntheses, and the synthetic procedures and characterization, are provided herein. Much promising chemistry was accomplished, which potentially can be exploited in other synthetic routes. The chemistry segues naturally from that described in the body of the paper, hence the numbering system is continued without use of “S” designation as is customary for compounds in the Supplementary Information section.



**Chart S1.** Attempted target chlorins.

**Design I-ExA.** In this attempted design (Scheme S1), the chalcone moieties would be incorporated at the 3-position to achieve wavelength-tunable and bioconjugatable chlorins. Here, chalcone formation entails an aldol condensation of the 3-acetylchlorin and 4-formylbenzoic acid under basic conditions. The synthesis started by protection of **1a** with TMSCl in the presence of DBU and a catalytic amount of AgCl, thereby affording the TMS-protected tris(propargyloxy)benzaldehyde **1d** in 63% yield.<sup>S1,S2</sup> The condensation of **1d** and pyrrole with InCl<sub>3</sub> unexpectedly gave TMS-cleaved dipyrromethane **2a** (not shown) instead of the corresponding dipyrromethane **2d**. TMS cleavage may happen when NaOH powder was added to quench the reaction; the use of NaOH addition is essential for dipyrromethane formation with InCl<sub>3</sub>.<sup>60</sup> TFA was then used as catalyst,<sup>S3</sup> and 0.1 N aqueous NaOH was added to quench the reaction, which gave **2d** in 47% yield.

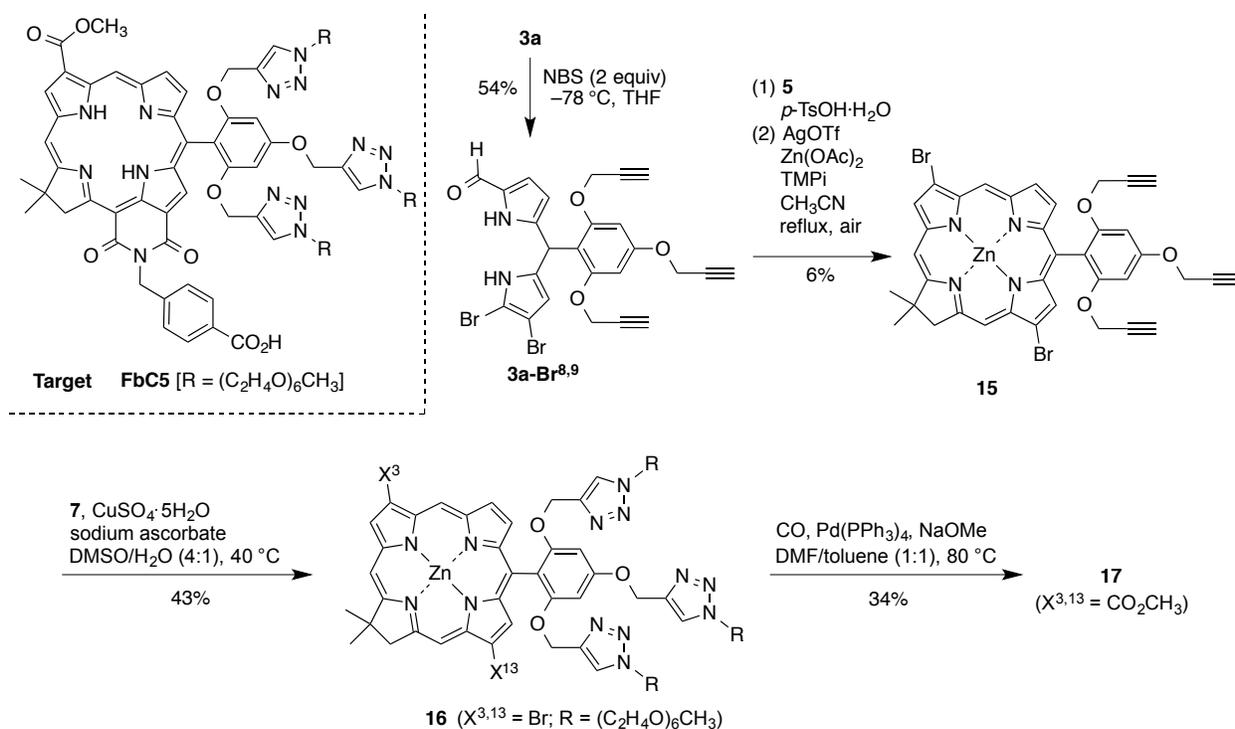
Vilsmeier formylation of TMS-protected dipyrromethane **2d** with POCl<sub>3</sub>/DMF afforded 1-formyldipyrromethane **3d** in 33% yield. Bromination of **3d** with 1 equiv of NBS at -78 °C afforded **3d-Br**<sup>9</sup>, which was used in the next step without further purification. The chlorin-forming reaction was carried out under standard conditions.<sup>62</sup> However, the reaction provided a mixture of zinc chlorins wherein 1–3 TMS protecting groups were cleaved, and no desired chlorin **14** was observed. The presence of the base TMPi might cause TMS deprotection during the oxidative cyclization. If successful, a Stille coupling reaction would replace the 3-bromo group with an acetyl moiety, but the synthesis was ceased here to avoid competing Sonogashira side reactions of the unprotected alkyne groups.



**Scheme S1.** Attempted synthesis of TMS-protected tris(propargyl)chlorin.

**Design I-ExB.** To avoid competing Sonogashira reaction with unprotected alkyne groups, an alternative strategy is to carry out the click reaction before Pd-catalyzed reactions. This strategy was employed to synthesize a target chlorin-imide (Scheme S2). The target chlorin-imide was expected to exhibit  $Q_y$  absorption  $\sim 705$  nm, which could fill the spectral window between that of synthetic chlorins (603–687 nm) and bacteriochlorins (707–792 nm).<sup>S4</sup> In this design, a bioconjugatable tether would be introduced at the imide ring at the stage of ring closure, and water-solubility achieved by click reaction between the alkynes and **7**. Bromination<sup>S5</sup> of **3a** with 2 equiv of NBS at  $-78$  °C afforded **3a-Br**<sup>8,9</sup> in 54% yield. Note: **3a-Br**<sup>8,9</sup> was unstable and decomposed quickly at room temperature; hence, the bromination was conducted immediately prior to the synthesis of the chlorin.

The chlorin-forming reaction was carried out under standard conditions,<sup>62</sup> which afforded zinc chlorin **15** in 6% yield. Treatment of **15** and **7** with a stoichiometric amount of Cu(I) in DMSO/H<sub>2</sub>O (4:1) at 40 °C gave the zinc triazole-PEG-chlorin **16** in 43% yield. The dibromochlorin **16** was subjected to carbonylation with CO in DMF/toluene (1:1) in the presence of a stoichiometric amount of Pd(PPh<sub>3</sub>)<sub>4</sub>. Subsequent treatment of the resulting acylpalladium intermediate with NaOMe in methanol afforded chlorin–diester **17** in 34% yield.

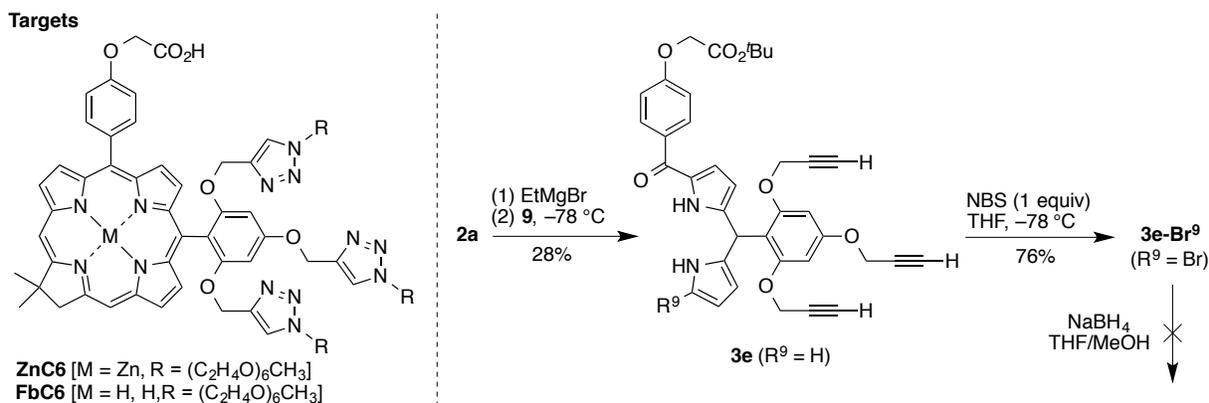


**Scheme S2.** Synthesis approach toward a chlorin-imide.

The successful formation of **17** provided an attractive methodology to synthesize water-soluble chlorins wherein only moderate derivatization is required after chlorin formation. For example, the target chlorin-chalcone **FbC4** (Scheme S1, upper panel) would be synthesized in a similar way: after preparation of the alkyne-unprotected chlorin **14**, a click reaction could be carried out first, followed by Stille coupling reaction to introduce the acetyl group. The last step would entail aldol condensation between the acetylchlorin and 4-formylbenzoic acid to incorporate a bioconjugatable tether. However, for the synthesis of the chlorin-imide, the limitation of this strategy came from the lengthy elaboration of the chlorin. Two more steps (regioselective 15-bromination, Pd-mediated carbonylation followed by ring closure to form the imide) remain to form the target chlorin-imide from **17**, but a total yield of **17** from Eastern half **3a** was less than 1%. At the same time, PEG groups were carried through all of the steps for chlorin derivatization, which complicated purification.

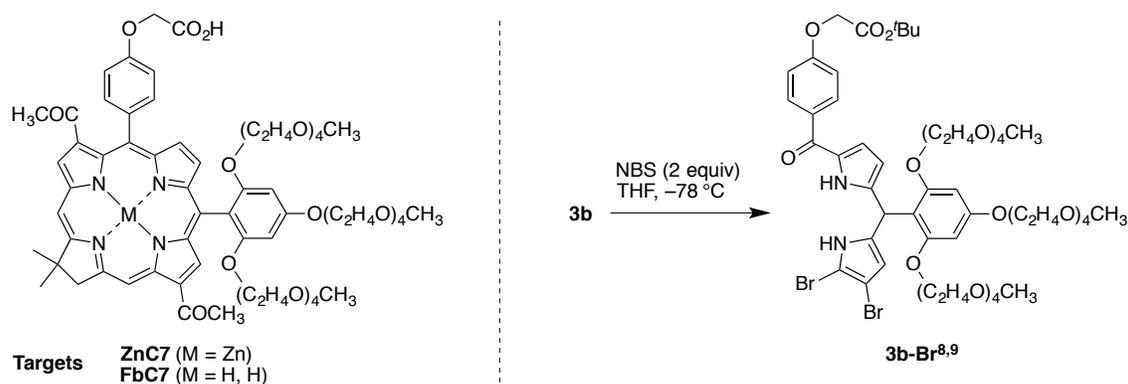
**Design I-ExC.** Target chlorins **ZnC6** and **FbC6** were pursued on the basis of the success of **ZnC1** and **FbC1**. The same PEGylation strategy was adopted with the only difference the requirement to introduce a bioconjugable tether at the 5-position. Acylation of dipyrromethane **2a** with thioester **9** afforded monoacyldipyrromethane **3e** in 28% yield along with recovery of unreacted dipyrromethane **2a** in 27% yield (Scheme S3). Acyldipyrromethane **3e** was brominated selectively at the remaining free  $\alpha$ -position by treatment<sup>70</sup> with NBS in THF at -78 °C, affording **3e-Br<sup>9</sup>** in 76% yield. The reduction of bromoacyldipyrromethane **3e-Br<sup>9</sup>** under the standard conditions<sup>62,66</sup> with excess NaBH<sub>4</sub> in THF/MeOH (4:1) was inefficient, as only the starting material **3e-Br<sup>9</sup>** was observed by TLC analysis, and none of the desired dipyrromethanecarbinol was detected. Unlike the analogous **3a-Br<sup>9</sup>**, which was unstable, compound **3e-Br<sup>9</sup>** was sufficiently stable for routine handling. The low conversion yield upon acylation of **2a** to **3e** and the failure of subsequent reduction might be attributed to the reactive

terminal alkynyl groups. One workaround would entail the protection of terminal alkynyl groups and removal thereof prior to the click reaction, but this was not investigated.



**Scheme S3.** Synthesis of 5-substituted chlorin precursors.

**Design II-ExA.** Another attempted design (for **ZnC7** and **FbC7**) entailed introduction of auxochromes at the 3- and 13-positions of PEGylated chlorins, which would be synthesized via pre-installation of the PEG moieties. Bromination of **3b** with 2 equiv of NBS at  $-78\text{ }^{\circ}\text{C}$  afforded the desired compound **3b-Br<sup>8,9</sup>**, but a tribrominated impurity (not shown) could not be removed (Scheme S4). The inability to purify dipyrromethane **3b-Br<sup>8,9</sup>** caused this approach to be discontinued.

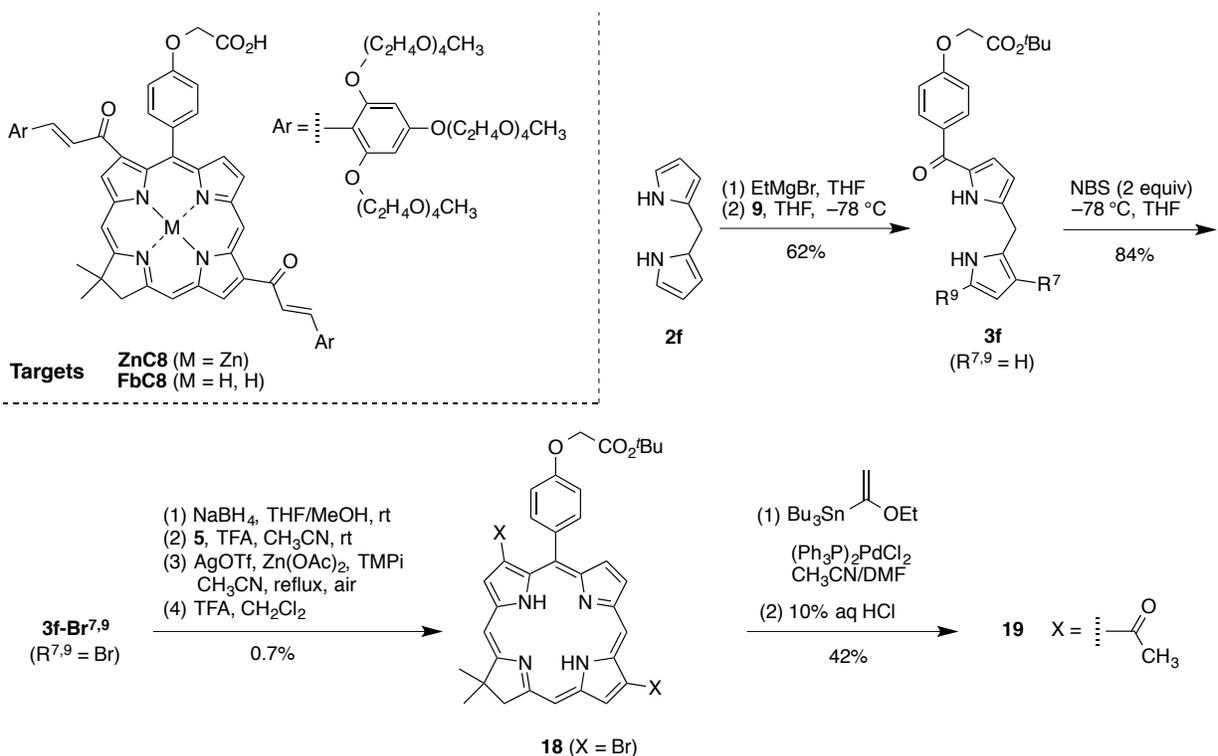


**Scheme S4.** Synthesis of 5-substituted chlorin precursors.

**Design III-ExA.** A chalcone design was employed here by derivatization of the 3,12-positions of the chlorin to achieve wavelength tunability and water solubility. The 5-position was assigned to the bioconjugatable tether. Thus, dipyrromethane<sup>60</sup> **2f** was acylated with thioester **9** to form monoacyldipyrromethane **3f** in 62% yield (Scheme S5). Acyldipyrromethane **3f** was brominated selectively at the 7,9-positions by treatment with 2 equiv of NBS in THF at  $-78\text{ }^{\circ}\text{C}$ ,<sup>S5</sup> affording **3f-Br<sup>7,9</sup>** in 84% yield. Attempts to reduce bromoacyldipyrromethane **3f-Br<sup>7,9</sup>** under the standard conditions<sup>62</sup> with excess NaBH<sub>4</sub> in

THF/MeOH (4:1) were unsuccessful: starting material **3f-Br**<sup>7,9</sup> was observed by TLC analysis after reduction, and the yield of the chlorin-forming reaction to produce **18** was a meager 0.7%.

Stille coupling<sup>69</sup> of 3,12-dibromochlorin **18** with tributyl(1-ethoxyvinyl)tin and a catalytic amount of (Ph<sub>3</sub>P)<sub>2</sub>PdCl<sub>2</sub> in CH<sub>3</sub>CN/DMF (3:2) followed by acidic hydrolysis gave 3,12-diacetylchlorin **19** in 42% yield. The expected Q<sub>y</sub> band of **19** is ~690 nm in toluene, on the basis of the known 5-unsubstituted chlorins that contain 3,12- or 3,13-diacetyl groups.<sup>70,S6</sup> However, the Q<sub>y</sub> band of **19** appeared at 671 nm in toluene. The failure to achieve the desired wavelength along with abysmally low yield of chlorin formation prompted us to abandon this strategy and avoid a chlorin design with a 5-substituent.



**Scheme S5.** Synthesis of a diacetylchlorin with a bioconjugatable tether at the 5-position.

### Experimental Procedures and Characterization

**2,4,6-Tris(3-trimethylsilylpropargyloxy)benzaldehyde (1d).** Following a trimethylsilyl protection procedure,<sup>S1,S2</sup> DBU (23.80 g, 156.5 mmol) was added to a mixture of benzaldehyde **1a** (7.00 g, 26.1 mmol) and AgCl (2.20 g, 15.7 mmol) in anhydrous CH<sub>2</sub>Cl<sub>2</sub> (50 mL). The reaction mixture was then heated under reflux, whereupon chlorotrimethylsilane (25.50 g, 234.9 mmol) was added dropwise. After refluxing with stirring for 36 h, the mixture was allowed to cool to room temperature. The mixture was diluted with hexanes, and then washed with aqueous NaHCO<sub>3</sub>, 2 M HCl, and water. The resulting organic phase was dried (Na<sub>2</sub>SO<sub>4</sub>), concentrated, and chromatographed [silica, hexanes/ethyl acetate (9:1)] to afford a light yellow oil (8.0 g, 63%): <sup>1</sup>H NMR δ 0.13 (s, 18H), 0.15 (s, 9H), 4.68 (s, 2H), 4.70 (s, 4H), 6.35 (s, 2H), 10.31 (s, 1H); <sup>13</sup>C NMR δ -0.40, 56.9, 57.7, 93.6, 93.7, 94.1, 98.8, 99.15, 110.3; ESI-MS obsd 485.20011, calcd 485.19942 [(M + H)<sup>+</sup>, M = C<sub>25</sub>H<sub>36</sub>O<sub>4</sub>Si<sub>3</sub>].

**5-[2,4,6-Tris(3-trimethylsilylpropargyloxy)phenyl]dipyrromethane (2d).** Following a reported procedure,<sup>60,S7</sup> a solution of benzaldehyde **1d** (6.00 g, 12.4 mmol) and pyrrole (33.2 g, 49.5 mmol) was degassed with a stream of argon for 10 min, and then TFA (95.0  $\mu$ L, 1.24 mmol) was added. The solution was stirred for 20 min, and then diluted with CH<sub>2</sub>Cl<sub>2</sub>. The organic phase was washed (0.1 N aqueous NaOH, H<sub>2</sub>O), dried (Na<sub>2</sub>SO<sub>4</sub>), concentrated and chromatographed [silica, hexanes/ethyl acetate (20:1)] to afford a light yellow oil (3.5 g, 47%): <sup>1</sup>H NMR  $\delta$  0.19 (s, 9H), 0.20 (s, 18H), 4.50 (s, 2H), 4.63 (s, 4H), 5.93 (s, 1H), 6.09–6.10 (m, 4H), 6.40 (s, 2H), 6.63–6.64 (m, 2H), 8.57 (s, 2H); <sup>13</sup>C NMR  $\delta$  –0.12, –0.03, 32.8, 57.2, 58.6, 92.8, 93.3, 94.8, 96.3, 99.9, 100.2, 106.3, 108.0, 115.1, 116.4, 116.7, 133.0, 158.2; ESI-MS obsd 601.27373, calcd 601.27325 [(M + H)<sup>+</sup>, M = C<sub>33</sub>H<sub>44</sub>N<sub>2</sub>O<sub>3</sub>Si<sub>3</sub>].

**1-Formyl-5-[2,4,6-tris(3-trimethylsilylpropargyloxy)phenyl]dipyrromethane (3d).** The Vilsmeier reagent was prepared by treatment of dry DMF (2.5 mL) with POCl<sub>3</sub> (542  $\mu$ L, 5.80 mmol) at 0 °C and stirring of the resulting mixture for 10 min under argon. In a separate flask, a solution of **2d** (3.50 g, 5.80 mmol) in DMF (30 mL) was treated with the freshly prepared Vilsmeier reagent at 0 °C. The resulting mixture was stirred at 0 °C for 1.5 h. The reaction mixture was treated with saturated aqueous NaOAc (~25 mL) for 2 h. CH<sub>2</sub>Cl<sub>2</sub> was added to reaction mixture, then the organic phase was washed (water, brine), dried (Na<sub>2</sub>SO<sub>4</sub>), concentrated, and chromatographed [silica, hexanes/ethyl acetate (9:1)] to afford an orange oil (1.2 g, 33%): <sup>1</sup>H NMR  $\delta$  0.19 (s, 18H), 0.20 (s, 9H), 4.52 (s, 2H), 4.54 (s, 2H), 4.65 (s, 2H), 5.90–5.92 (m, 1H), 6.07 (s, 1H), 6.12–6.16 (m, 2H), 6.40 (s, 2H), 6.70–6.72 (m, 1H), 6.83–6.85 (m, 1H), 8.98 (s, 1H), 9.06 (s, 1H), 9.33 (s, 1H); <sup>13</sup>C NMR  $\delta$  –0.08, 33.1, 57.2, 58.2, 93.4, 93.7, 95.6, 99.58, 99.69, 108.0, 109.6, 112.6, 117.7, 122.3, 130.1, 131.6, 144.6, 157.1, 158.7, 178.1; ESI-MS obsd 629.26867, calcd 629.26816 [(M + H)<sup>+</sup>, M = C<sub>34</sub>H<sub>44</sub>N<sub>2</sub>O<sub>4</sub>Si<sub>3</sub>].

**Zn(II)-3-Bromo-10-[2,4,6-tris(3-trimethylsilylpropargyloxy)phenyl]-18,18-dimethylchlorin (14).** Following a standard procedure,<sup>S8</sup> a solution of **3d** (1.2 g, 1.9 mmol) in anhydrous THF (19 mL) was treated with NBS (340 mg, 1.9 mmol) under argon at –78 °C. The reaction mixture was stirred for 1 h at –78 °C, after which the ice bath was removed and the reaction mixture was allowed to warm to room temperature. Upon reaching 0 °C, hexanes and water were added. The mixture was extracted with ethyl acetate. The organic layer was dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated to afford 9-bromo-1-formyl-5-[2,4,6-tris(3-trimethylsilylpropargyloxy)phenyl]dipyrromethane (**3d-Br**<sup>9</sup>) as a yellow oil, which was used in the next step without further purification: <sup>1</sup>H NMR  $\delta$  0.19 (s, 18H), 0.20 (s, 9H), 4.47–4.61 (m, 4H), 4.65 (s, 2H), 5.91–5.93 (m, 1H), 5.98 (s, 1H), 6.03–6.08 (m, 2H), 6.40 (s, 2H), 6.84–6.86 (m, 1H), 8.91 (s, 1H), 9.00 (s, 1H), 9.35 (s, 1H).

Following a general procedure,<sup>62</sup> a solution of **5** (511 mg, 1.90 mmol) and **3d-Br**<sup>9</sup> (1.90 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (50 mL) was treated with a solution of *p*-TsOH·H<sub>2</sub>O (1.80 g, 9.50 mmol) in methanol (12.6 mL) under argon. The reaction mixture immediately turned red. The mixture was stirred for 30 min under argon, then treated with 2,2,6,6-tetramethylpiperidine (2.40 mL, 14.0 mmol) and concentrated to dryness. The resulting brown solid was suspended in acetonitrile (190 mL) followed by the successive addition of 2,2,6,6-tetramethylpiperidine (8.10 mL, 47.5 mmol), Zn(OAc)<sub>2</sub> (5.20 g, 28.5 mmol), and AgOTf (1.40 g, 5.70 mmol). The resulting suspension was refluxed for 22 h exposed to air. The crude mixture was filtered through a silica pad with CH<sub>2</sub>Cl<sub>2</sub>. The crude product was a mixture of chlorins with TMS cleaved and partially cleaved.

**1-{4-[2-(*tert*-Butoxy)-2-oxoethoxy]phenoxy}-5-[2,4,6-tris(propargyloxy)phenyl]dipyrromethane (3e).** Following a general procedure,<sup>67</sup> EtMgBr (0.9 M solution in THF, 2.66 mL, 2.39 mmol) was added dropwise to a solution of **2a** (459 mg, 1.19 mmol) in THF (1.2 mL) over a 5 min period. The solution was stirred at room temperature for 10 min before cooling to  $-78\text{ }^{\circ}\text{C}$ . A solution of **9** (412 mg, 1.19 mmol) in THF (1.2 mL) was added, and the mixture was stirred at  $-78\text{ }^{\circ}\text{C}$  for 30 min. The cooling bath was removed and the reaction mixture was allowed to warm to room temperature for 1 h before saturated aqueous  $\text{NH}_4\text{Cl}$  was added. The aqueous layer was removed and washed with  $\text{CH}_2\text{Cl}_2$ . The combined organic extract was washed with brine, concentrated, and chromatographed [silica, hexanes/ethyl acetate (3:1)] to recover starting material **2a** (126 mg) as the first fraction followed by the title compound as a yellow oil (203 mg, 28%):  $^1\text{H NMR}$   $\delta$  1.44 (s, 9H), 2.53–2.57 (m, 3H), 4.57 (s, 2H), 4.59 (d,  $J = 2.4$  Hz, 4H), 4.68 (d,  $J = 2.4$  Hz, 2H), 5.92–5.95 (m, 1H), 6.10 (s, 1H), 6.12–6.13 (m, 2H), 6.43 (s, 2H), 6.70–6.72 (m, 1H), 6.73–6.76 (m, 1H), 6.93 (d,  $J = 9$  Hz, 2H), 7.84 (d,  $J = 9$  Hz, 2H), 8.83 (s, 1H), 9.27 (s, 1H);  $^{13}\text{C NMR}$   $\delta$  28.15, 33.0, 56.2, 57.2, 65.7, 76.2, 76.4, 78.1, 82.8, 95.7, 107.6, 108.0, 109.3, 114.2, 117.5, 119.9, 129.8, 130.4, 131.0, 132.3, 142.5, 156.8, 158.2, 160.7, 167.7, 182.8; ESI-MS obsd 619.24297, calcd 619.24388 [(M + H)<sup>+</sup>, M =  $\text{C}_{37}\text{H}_{34}\text{N}_2\text{O}_7$ ].

**9-Bromo-1-{4-[2-(*tert*-butoxy)-2-oxoethoxy]phenyl}-5-[2,4,6-tris(propargyloxy)-phenyl]dipyrromethane (3e-Br<sup>9</sup>).** Following a general procedure,<sup>70</sup> a solution of **3e** (77.0 mg, 0.124 mmol) in anhydrous THF (2.5 mL) was treated with NBS (22.1 mg, 0.124 mmol) under argon at  $-78\text{ }^{\circ}\text{C}$ . The reaction mixture was stirred for 1 h at  $-78\text{ }^{\circ}\text{C}$ , after which the ice bath was removed, and the reaction mixture was allowed to warm to room temperature. Upon reaching  $0\text{ }^{\circ}\text{C}$ , hexanes and water were added. The mixture was extracted with ethyl acetate. The organic layer was dried ( $\text{Na}_2\text{SO}_4$ ), concentrated, and chromatographed [silica, hexanes/ethyl acetate (3:1)] to afford an orange oil (67 mg, 76%):  $^1\text{H NMR}$   $\delta$  1.48 (s, 9H), 2.56–2.57 (m, 3H), 4.56 (s, 2H), 4.62 (d,  $J = 2.4$  Hz, 4H), 4.68 (d,  $J = 2.4$  Hz, 2H), 5.95–5.96 (m, 1H), 6.02 (s, 1H), 6.03–6.04 (m, 2H), 6.42 (s, 2H), 6.73–6.75 (m, 1H), 6.93 (d,  $J = 9$  Hz, 2H), 7.84 (d,  $J = 9$  Hz, 2H), 8.80 (s, 1H), 9.38 (s, 1H);  $^{13}\text{C NMR}$   $\delta$  28.11, 33.2, 56.1, 57.1, 65.6, 76.3, 76.6, 77.8, 78.2, 82.8, 95.5, 96.8, 109.4, 110.0, 112.3, 114.2, 119.7, 130.0, 131.0, 132.0, 132.1, 141.4, 156.6, 158.4, 160.7, 167.6, 182.9; ESI-MS obsd 697.15421, calcd 697.15439 [(M + H)<sup>+</sup>, M =  $\text{C}_{37}\text{H}_{33}\text{BrN}_2\text{O}_7$ ].

**8,9-Dibromo-1-{4-[2-(*tert*-butoxy)-2-oxoethoxy]phenyl}-5-[2,4,6-tris(3,6,9,12-tetra-oxatridecyloxy)phenyl]dipyrromethane (3b-Br<sup>8,9</sup>).** A solution of **3b** (1.14 g, 1.06 mmol) in anhydrous THF (10 mL) was treated with NBS (377.3 mg, 2.12 mmol) under argon at  $-78\text{ }^{\circ}\text{C}$ . The reaction mixture was stirred for 1 h at  $-78\text{ }^{\circ}\text{C}$ , after which the ice bath was removed and the reaction mixture was allowed to warm. Upon reaching  $-20\text{ }^{\circ}\text{C}$ , hexanes (20 mL) and water (20 mL) were added. The mixture was extracted with ethyl acetate. The organic layer was dried ( $\text{Na}_2\text{SO}_4$ ), concentrated, and chromatographed [silica,  $\text{CH}_2\text{Cl}_2/\text{CH}_3\text{OH}$  (97:3)] to afford the title compound mixed with tribromodipyrromethane in a ratio of 5:1 on the basis of  $^1\text{H NMR}$  analysis (1000 mg, impure).

**1-{4-[2-(*tert*-Butoxy)-2-oxoethoxy]phenyl}dipyrromethane (3f).** Following a general procedure,<sup>67</sup> EtMgBr (1 M solution in THF, 6.2 mL, 6.2 mmol) was added dropwise to a solution of **2f** (0.45 g, 3.1 mmol) in THF (3.4 mL) over a 5 min period. The solution was stirred at room temperature for 10 min before cooling to  $-78\text{ }^{\circ}\text{C}$ . A solution of **9** (1.07 mg, 3.10 mmol) in THF (3.4 mL) was added, and the mixture was stirred at  $-78\text{ }^{\circ}\text{C}$  for 30 min. The cooling bath was removed, and the reaction mixture was allowed to warm to room temperature for 1 h before saturated aqueous  $\text{NH}_4\text{Cl}$  was added. The aqueous layer was removed and washed with  $\text{CH}_2\text{Cl}_2$ .

The combined organic layer was washed with brine, concentrated, and chromatographed [silica, hexanes/ethyl acetate (4:1)] to recover starting material **2f** as the first fraction followed by the title compound, which upon concentration gave a white solid (734 mg, 62%): mp 120–122 °C; <sup>1</sup>H NMR δ 1.52 (s, 9H), 4.09 (s, 2H), 4.59 (s, 2H), 6.066–6.069 (m, 1H), 6.12–6.15 (m, 2H), 6.62–6.64 (m, 1H), 6.80–6.82 (m, 1H), 6.95 (d, *J* = 8.4 Hz, 2H), 7.86 (d, *J* = 8.4 Hz, 2H), 8.63 (s, 1H), 10.17 (s, 1H); <sup>13</sup>C NMR δ 27.9, 49.1, 83.0, 106.3, 106.4, 108.1, 108.2, 109.6, 109.7, 114.2, 117.6, 121.5, 127.9, 130.1, 131.1, 131.9, 139.6, 160.8, 167.8, 183.8; ESI-MS obsd 381.18073, calcd 381.18008 [(M + H)<sup>+</sup>, M = C<sub>22</sub>H<sub>24</sub>N<sub>2</sub>O<sub>4</sub>].

**Zn(II)-3,13-Dibromo-10-[2,4,6-tris(propargyloxy)phenyl]-18,18-dimethylchlorin (15).** Following a standard procedure,<sup>S8</sup> a solution of **3a** (948 mg, 2.30 mmol) in anhydrous THF (23 mL) was treated with NBS (818 mg, 4.60 mmol) under argon at –78 °C. The reaction mixture was stirred for 1 h at –78 °C, after which the ice bath was removed. Upon reaching –20 °C, hexanes (20 mL) and water (20 mL) was added. The mixture was extracted with ethyl acetate. The organic layer was dried (Na<sub>2</sub>SO<sub>4</sub>), concentrated and chromatographed [silica, hexanes/ethyl acetate (4:1)] to afford 8,9-dibromo-1-formyl-5-[2,4,6-tris(propargyloxy)phenyl]dipyrrromethane (**3a-Br**<sup>8,9</sup>) as a yellow oil (710 mg, 54%): <sup>1</sup>H NMR δ 2.57 (t, *J* = 2.4 Hz, 2H), 2.58 (t, *J* = 2.4 Hz, 1H), 4.63 (d, *J* = 2.4 Hz, 4H), 4.70 (d, *J* = 2.4 Hz, 2H), 5.89–5.91 (m, 1H), 6.10 (d, *J* = 3.0 Hz, 1H), 6.21 (s, 1H), 6.43 (s, 2H), 6.85–6.87 (m, 1H), 9.21 (s, 1H), 9.24 (s, 1H), 9.43 (s, 1H). Dibromodipyrrromethane **1e** in CDCl<sub>3</sub> solution darkened and decomposed, and hence was used directly in the chlorin-forming process.

Following a general procedure,<sup>62</sup> a solution of **5** (161 mg, 0.600 mmol) and **3a-Br**<sup>8,9</sup> (342 mg, 0.600 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (16 mL) was treated with a solution of *p*-TsOH·H<sub>2</sub>O (570. mg, 3.00 mmol) in methanol (4 mL) under argon. The reaction mixture immediately turned red. The mixture was stirred for 30 min under argon, then treated with 2,2,6,6-tetramethylpiperidine (0.76 mL, 4.5 mmol) and concentrated to dryness. The resulting brown solid was suspended in acetonitrile (60 mL) followed by the successive addition of 2,2,6,6-tetramethylpiperidine (2.6 mL, 15.0 mmol), Zn(OAc)<sub>2</sub> (1.65 g, 9.00 mmol), and AgOTf (463 mg, 1.80 mmol). The resulting suspension was refluxed for 22 h exposed to air. The crude mixture was filtered through a silica pad with CH<sub>2</sub>Cl<sub>2</sub>. The crude product was chromatographed [silica, hexanes/CH<sub>2</sub>Cl<sub>2</sub> (1:2)] to afford a green solid (28 mg, 6%): ESI-MS obsd 795.96584, calcd 795.96576 [M<sup>+</sup>, M = C<sub>37</sub>H<sub>26</sub>Br<sub>2</sub>N<sub>4</sub>O<sub>3</sub>Zn].

**Zn(II)-3,13-Dibromo-10-[2,4,6-tris(2,5,8,11,14,17-hexaoxonadecyl-1H-1,2,3-triazol-4-ylmethoxy)phenyl]-18,18-dimethylchlorin (16).** The Cu(I) catalyst was prepared by treatment of CuSO<sub>4</sub>·H<sub>2</sub>O (15 mg, 0.060 mmol) and sodium ascorbate (24.0 mg, 0.120 mmol) with 600 μL of deionized H<sub>2</sub>O under argon. The reaction mixture turned brown immediately and was stirred until homogeneous. In a separate vial, **15** (24 mg, 0.030 mmol) and **7** (87 mg, 0.27 mmol) in DMSO (1.2 mL) were treated with freshly prepared Cu (I) catalyst (300 μL) under argon. The reaction mixture was stirred at 40 °C for 16 h. Upon cooling to room temperature, H<sub>2</sub>O was added and organic phase was extracted with CH<sub>2</sub>Cl<sub>2</sub>. The extract was washed (brine), dried (Na<sub>2</sub>SO<sub>4</sub>), concentrated, and chromatographed [silica, CH<sub>2</sub>Cl<sub>2</sub> /CH<sub>3</sub>OH (93:7)] to afford a green solid (23 mg, 43%): ESI-MS obsd 1760.54352, calcd 1760.54355 [(M + H)<sup>+</sup>, M = C<sub>76</sub>H<sub>107</sub>Br<sub>2</sub>N<sub>13</sub>O<sub>21</sub>Zn].

**Zn(II)-3,13-Bis(methoxycarbonyl)-10-[2,4,6-tris(2,5,8,11,14,17-hexaoxonadecyl-1H-1,2,3-triazol-4-ylmethoxy)phenyl]-18,18-dimethylchlorin (17).** Samples of **16** (10 mg, 0.0057 mmol) and Pd(PPh<sub>3</sub>)<sub>4</sub> (13.2 mg, 0.0114 mmol) were dried in a Schlenk flask under high

vacuum for 1 h. The solvent DMF/toluene (1:1) was purged with argon (30 min) and subsequently with CO (30 min). The reaction flask was filled with CO gas and DMF/toluene [0.6 mL, (1:1)]. CO gas was bubbled through the stirred reaction mixture for 2 h. Then excess NaOMe (6.2 mg, 0.11 mmol) in MeOH (285  $\mu$ L) was added to the reaction mixture. The latter was stirred at 80  $^{\circ}$ C for 5 min before being cooled to room temperature. A saturated aqueous solution of NH<sub>4</sub>Cl was added, and the mixture was extracted with CH<sub>2</sub>Cl<sub>2</sub>. The organic layer was separated, filtered through Celite (CH<sub>2</sub>Cl<sub>2</sub>), and concentrated. Column chromatography [silica, CH<sub>2</sub>Cl<sub>2</sub>/CH<sub>3</sub>OH (7:93)] afforded a green oil (3.0 mg, 34%): MALDI-MS obsd 1742.0881, calcd 1742.7158 [(M + Na)<sup>+</sup>, M = C<sub>80</sub>H<sub>113</sub>N<sub>13</sub>O<sub>25</sub>Zn].

### 3,12-Dibromo-5-[4-(carboxymethoxy)phenyl]-18,18-dimethylchlorin (**18**).

Following a standard procedure,<sup>S5</sup> a solution of **3f** (734 mg, 1.93 mmol) in anhydrous THF (38 mL) was treated with NBS (686 mg, 3.86 mmol) under argon at  $-78^{\circ}$ C. The reaction mixture was stirred for 1 h at  $-78^{\circ}$ C, after which the cooling bath was removed and the reaction mixture was allowed to warm. Upon reaching  $-20^{\circ}$ C, hexanes (38 mL) and water (38 mL) were added. The mixture was extracted with ethyl acetate. The organic layer was dried (Na<sub>2</sub>SO<sub>4</sub>), concentrated, and chromatographed [silica, hexanes/ethyl acetate (4:1)] to afford 7,9-dibromo-1-[4-(2-(*tert*-butoxy)-2-oxoethoxy)phenyl]dipyrrromethane (**3f-Br**<sup>7,9</sup>) as a light yellow solid (878 mg, 84%): <sup>1</sup>H NMR  $\delta$  1.49 (s, 9H), 4.05 (s, 2H), 4.59 (s, 2H), 6.00 (d, *J* = 3 Hz, 1H), 6.27–6.29 (m, 1H), 6.87–6.89 (m, 1H), 6.98 (d, *J* = 9 Hz, 2H), 7.87 (d, *J* = 9 Hz, 2H), 10.45 (s, 1H), 11.52 (s, 1H).

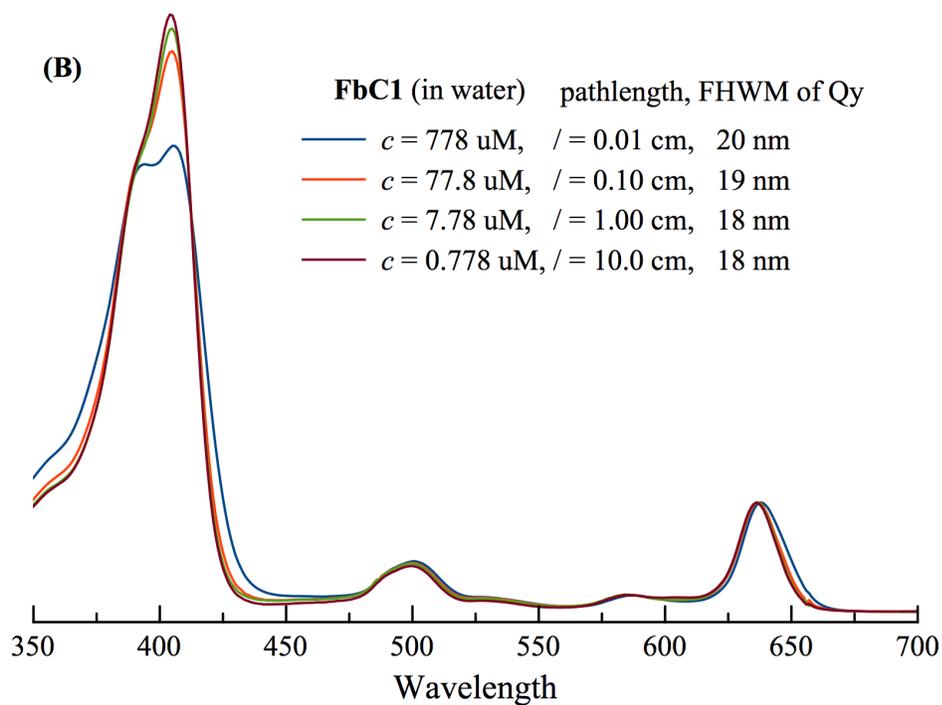
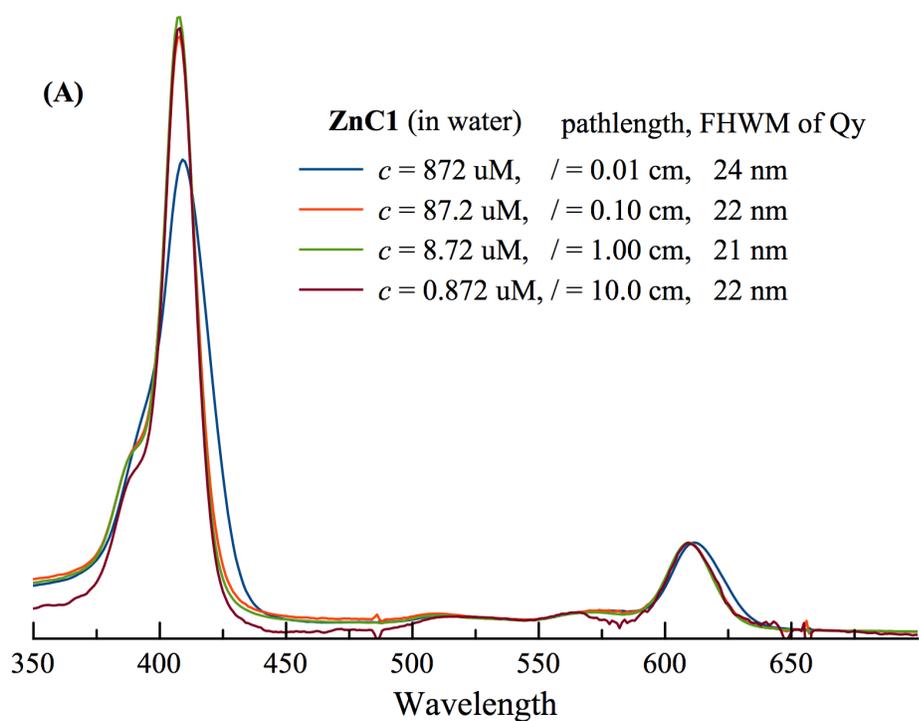
Following a general procedure,<sup>S9</sup> a solution of **3f-Br**<sup>7,9</sup> (878 mg, 1.63 mmol) in anhydrous THF (26 mL) and anhydrous methanol (6.6 mL) was treated with NaBH<sub>4</sub> (616 mg, 16.3 mmol) at room temperature for 30 min. The reaction mixture was quenched by the addition of saturated aqueous NH<sub>4</sub>Cl, and then extracted with ethyl acetate. The combined organic phase was washed with brine, dried (Na<sub>2</sub>SO<sub>4</sub>), and concentrated. The resulting oil was dissolved in CH<sub>3</sub>CN (16 mL) containing **5** (439 mg, 1.63 mmol) and TFA (125  $\mu$ L, 1.63 mmol). The resulting mixture was stirred at room temperature for 30 min and then diluted with CH<sub>3</sub>CN. Sample of 2,2,6,6-tetramethylpiperidine (4.16 mL, 24.5 mmol), Zn(OAc)<sub>2</sub> (4.49 g, 24.5 mmol) and AgOTf (1.26 g, 4.89 mmol) were added successively. The resulting suspension was refluxed for 22 h exposed to air. The crude mixture was filtered through a silica pad with ethyl acetate. The filtrate was concentrated, dissolved in anhydrous CH<sub>2</sub>Cl<sub>2</sub> (100 mL), and treated dropwise with TFA (150  $\mu$ L). After 10 min, saturated aqueous NaHCO<sub>3</sub> was added slowly. The organic layer was dried (Na<sub>2</sub>SO<sub>4</sub>), concentrated and chromatographed [silica, hexanes/CH<sub>2</sub>Cl<sub>2</sub> (1:1)] to afford a green solid (8.0 mg, 0.7%): <sup>1</sup>H NMR  $\delta$   $-2.12$  (br s, 1H),  $-1.52$  (br s, 1H), 1.61 (s, 6H), 2.00 (s, 6H), 4.51 (s, 2H), 4.79 (s, 2H), 7.22 (d, *J* = 8.8 Hz, 2H), 7.84 (d, *J* = 8.8 Hz, 2H), 8.50 (d, *J* = 4.4 Hz, 1H), 8.75 (s, 1H), 8.77 (s, 1H), 8.85 (s, 1H), 8.90 (d, *J* = 4.4 Hz, 1H), 8.96 (s, 1H), 9.82 (s, 1H); <sup>13</sup>C NMR  $\delta$  28.3, 29.8, 31.3, 40.3, 46.7, 51.5, 66.2, 82.7, 94.7, 96.2, 105.6, 113.1, 116.6, 118.8, 121.8, 124.2, 128.7, 130.1, 132.1, 132.7, 133.2, 134.6, 138.6, 139.3, 151.4, 154.4, 158.4, 164.8, 168.3, 174.8; ESI-MS obsd 703.09084, calcd 703.09139 [(M + H)<sup>+</sup>, M = C<sub>34</sub>H<sub>32</sub>Br<sub>2</sub>N<sub>4</sub>O<sub>3</sub>];  $\lambda_{\text{abs}}$  (toluene) 401, 647 nm.

**3,12-Diacetyl-5-[4-(carboxymethoxy)phenyl]-18,18-dimethylchlorin (**19**).** Following a procedure for Stille coupling of chlorins,<sup>70</sup> a mixture of **18** (8.0 mg, 0.011 mmol), tributyl(1-ethoxyvinyl)tin (20.  $\mu$ L, 0.055 mmol), and (Ph<sub>3</sub>P)<sub>2</sub>PdCl<sub>2</sub> (1.6 mg, 0.0022 mmol) was stirred in CH<sub>3</sub>CN/DMF [600  $\mu$ L (3:2)] under argon for 4 h at 83  $^{\circ}$ C in a Schlenk line. The reaction mixture was treated with 10% aqueous HCl (2 mL) at room temperature for 20 min.

CH<sub>2</sub>Cl<sub>2</sub> was added. The organic layer was separated, washed (saturated aqueous NaHCO<sub>3</sub>, water, and brine), dried (Na<sub>2</sub>SO<sub>4</sub>), concentrated, and chromatographed [silica, CH<sub>2</sub>Cl<sub>2</sub>/ethyl acetate (49:1)] to afford a brown oil (3.2 mg, 42%): <sup>1</sup>H NMR δ -1.97 (br s, 1H), -1.56 (br s, 1H), 1.61 (s, 6H), 2.04 (s, 6H), 2.17 (s, 3H), 3.27 (s, 3H), 4.57 (s, 2H), 4.79 (s, 2H), 7.24 (d, *J* = 8.4 Hz, 2H), 8.02 (d, *J* = 8.4 Hz, 2H), 8.61 (d, *J* = 4.4 Hz, 1H), 8.89 (s, 1H), 8.95 (s, 1H), 8.98 (s, 1H), 9.03 (d, *J* = 4.4 Hz, 1H), 9.27 (s, 1H), 10.81 (s, 1H); <sup>13</sup>C NMR δ 28.4, 29.9, 30.1, 31.3, 31.9, 47.0, 51.4, 66.1, 82.9, 96.6, 98.4, 109.3, 113.8, 121.2, 125.5, 126.5, 132.1, 133.1, 133.6, 135.1, 136.6, 137.0, 137.7, 143.3, 153.4, 155.5, 158.8, 163.6, 168.1, 175.4, 197.2, 201.3; ESI-MS obsd 631.29117, calcd 631.29150 [(M + H)<sup>+</sup>, M = C<sub>38</sub>H<sub>38</sub>N<sub>4</sub>O<sub>5</sub>]; λ<sub>abs</sub> (toluene) 430, 671 nm.

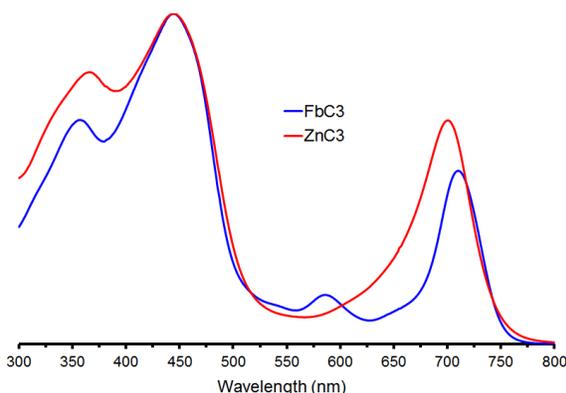
### 3. Absorption Spectra

The spectral properties of the PEGylated chlorins **ZnC1** and **FbC1** are shown in Figure S1. **ZnC1** exhibited almost unchanged spectroscopic properties over a concentration range of 1000-fold, indicating high solubility of the chlorin in water. On the other hand, at higher concentrations, **FbC1** exhibited no significant broadening in the Q<sub>y</sub> band, but the shape of the B bands changed at concentrations ~800 μM showing some degree of aggregation.



**Figure S1.** Absorption versus concentration of **ZnCl** and **FbCl** over a range of 1000-fold. The spectra are normalized at the Q<sub>y</sub> band; the FWHM of the Q<sub>y</sub> band is shown in the inset.

Absorption spectra of **ZnC3** and **FbC3** in neat water at  $\sim 5 \mu\text{M}$  are shown in Figure S2. Obvious broadening of the spectra is evident.



**Figure S2.** Absorption spectra of PEGylated chlorins in  $\text{H}_2\text{O}$ .

#### 4. Purification of PEGylated Compounds

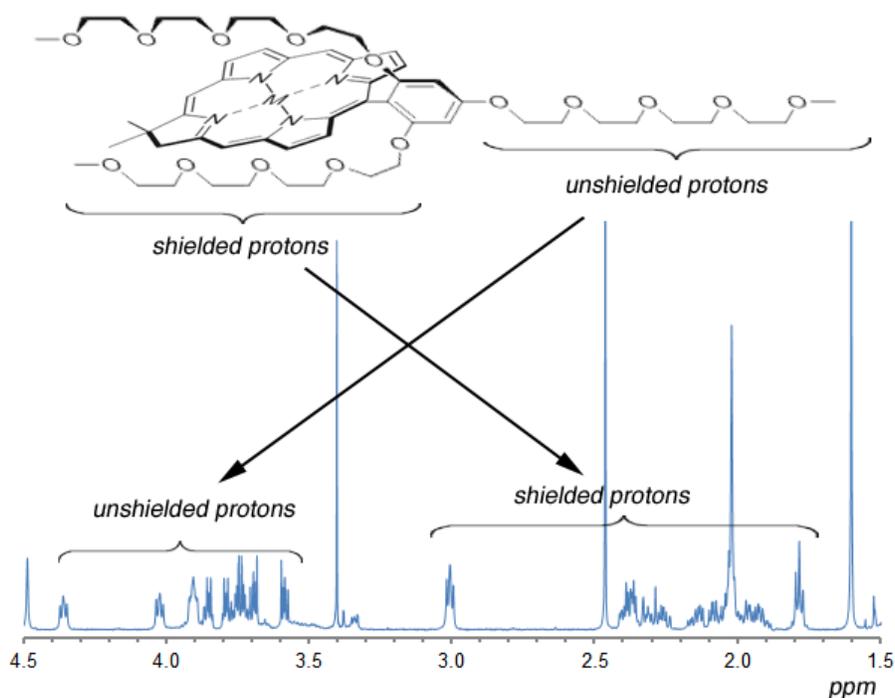
One essential issue arising from the pre-installation route concerns the purification process. During flash chromatography, PEGylated compounds possessing high polarity tended to bind tightly to silica gel. When the crude mixture containing the desired product was mixed with PEG-containing impurities, the resolution of the flash chromatography process was compromised. A typical flash chromatography (3 cm dia.  $\times$  15 cm) procedure entails: (1) pack the column and load the crude mixture with  $\text{CH}_2\text{Cl}_2$  (or ethyl acetate); (2) elute with one column volume of  $\text{CH}_2\text{Cl}_2$  (or ethyl acetate) to remove impurities while the PEGylated precursors remained bound or near the top of the column; and (3) elute with  $\text{CH}_2\text{Cl}_2$  with a gradual increase of 0.5–5% MeOH (or EtOH with ethyl acetate<sup>S10</sup>) to remove PEG-containing impurities and isolate the desired product. The PEGylated precursors **1b**, **1b'**, and **2b** generally were obtained with good purity following the above procedure. For acylation of **2b**, the crude mixture often contained  $\sim 20\%$  of unreacted starting material **2b**. Isolation of **3b** with good purity required prolonged elution (2–3 h) in step 3.

The purification of the PEGylated chlorins proceeded in the same manner as employed for separation of multiporphyrin arrays.<sup>75</sup> A general purification procedure for the crude PEGylated chlorin is as follows, which afforded the PEGylated chlorins **ZnC1**, **10**, and **FbC3** in good purity:

- (1) Silica chromatography (2 cm dia.  $\times$  10 cm) was eluted with  $\text{CH}_2\text{Cl}_2$  (0.5–5% MeOH).
- (2) Size-exclusion chromatography (4 cm dia.  $\times$  40 cm; Bio Beads S-X3, 200–400 mesh; flow rate:  $\sim 1.5$  ml/min) was eluted with toluene (HPLC grade).
- (3) Flash silica chromatography (2 cm dia.  $\times$  4 cm) was eluted with  $\text{CH}_2\text{Cl}_2$  (0.5–10% MeOH, HPLC grade).

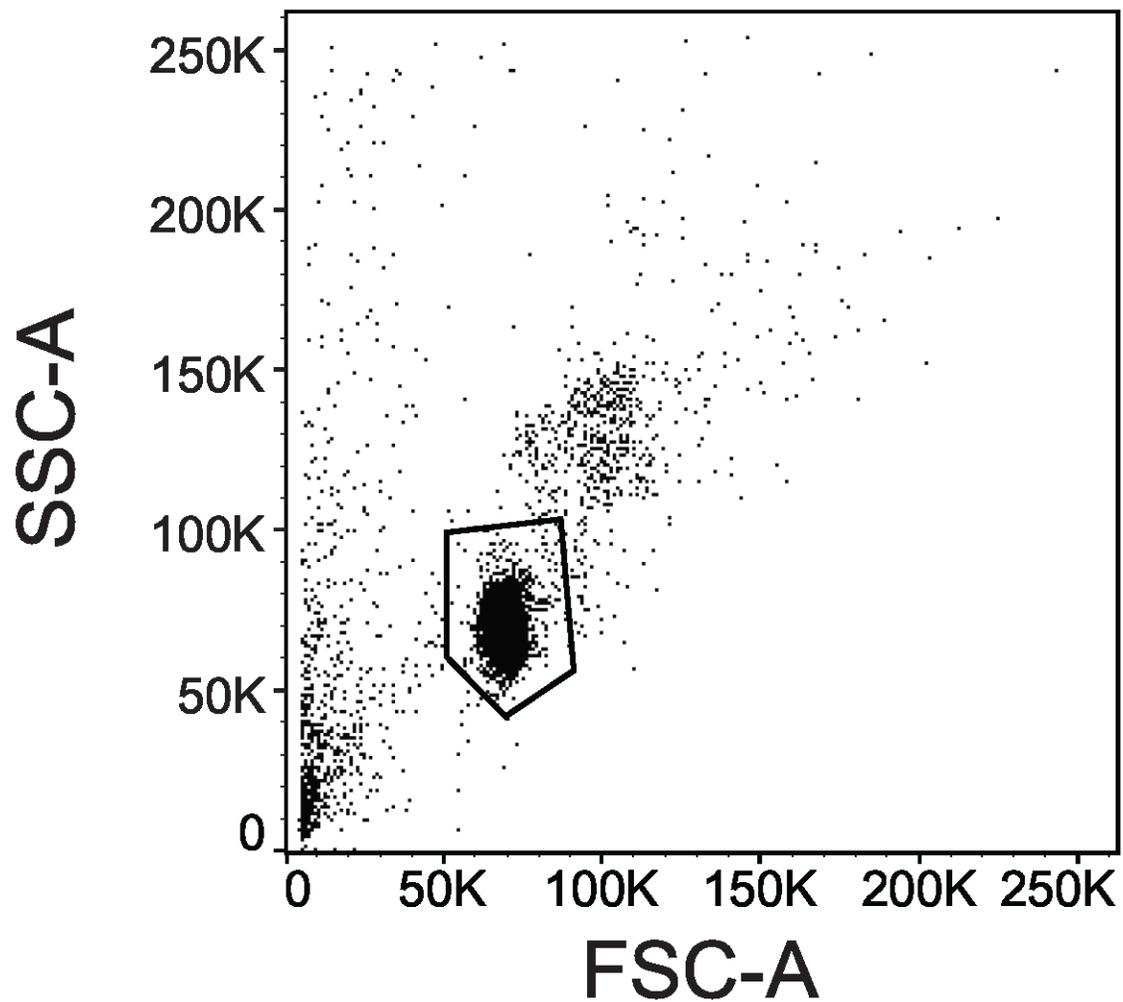
### 5. 3-Dimensional Conformation

The conformation of the 2,4,6-trisubstituted aryl motif was examined by  $^1\text{H}$  NMR spectroscopy. Protons on the PEG chain typically resonate at 3.4–4.6 ppm, depending on the terminal substitution groups. Indeed, the PEGylated precursors **1b**, **1b'**, **2b**, and **3b** exhibited a complicated resonance pattern in this region in the  $^1\text{H}$  NMR spectra. No significant difference was found between the resonances of protons from the *ortho*- and *para*-PEG chains in the non-macrocyclic precursors. However, for the PEGylated chlorins a significant difference was observed. The protons of the *para*-PEG chain resonated similarly to that of a typical PEG-chain, while the protons of the *ortho*-PEG chains resonated at a higher frequency, ranging from 1.7–3.2 ppm. A  $^1\text{H}$  NMR spectrum of PEGylated chlorins is demonstrated by **10** and shown in Figure S3. The *ortho*-PEG chains are conformationally restricted to above and below the macrocycle plane and thus are magnetically more shielded due to the aromatic ring-current effect. Although detailed analyses and assignments of PEG-protons do not relate to our main interests, the different resonance regions clearly show that the desired geometric design was achieved.



**Figure S3.**  $^1\text{H}$  NMR spectrum (in  $\text{CDCl}_3$  at room temperature) of **10** and illustrative representation of shielded and unshielded protons. Note: the scheme only represents the relative orientation of the chlorin and the PEG moiety and does not reflect the actual size and conformation.

## 6. Flow Cytometry Data



**Figure S4.** Dot plot of side-scattered light (SSC) versus forward-scattered light (FSC), which provide a measure of granularity and size, respectively. Selection of beads for subsequent analyses was achieved by gating around the major population of single beads, as shown.

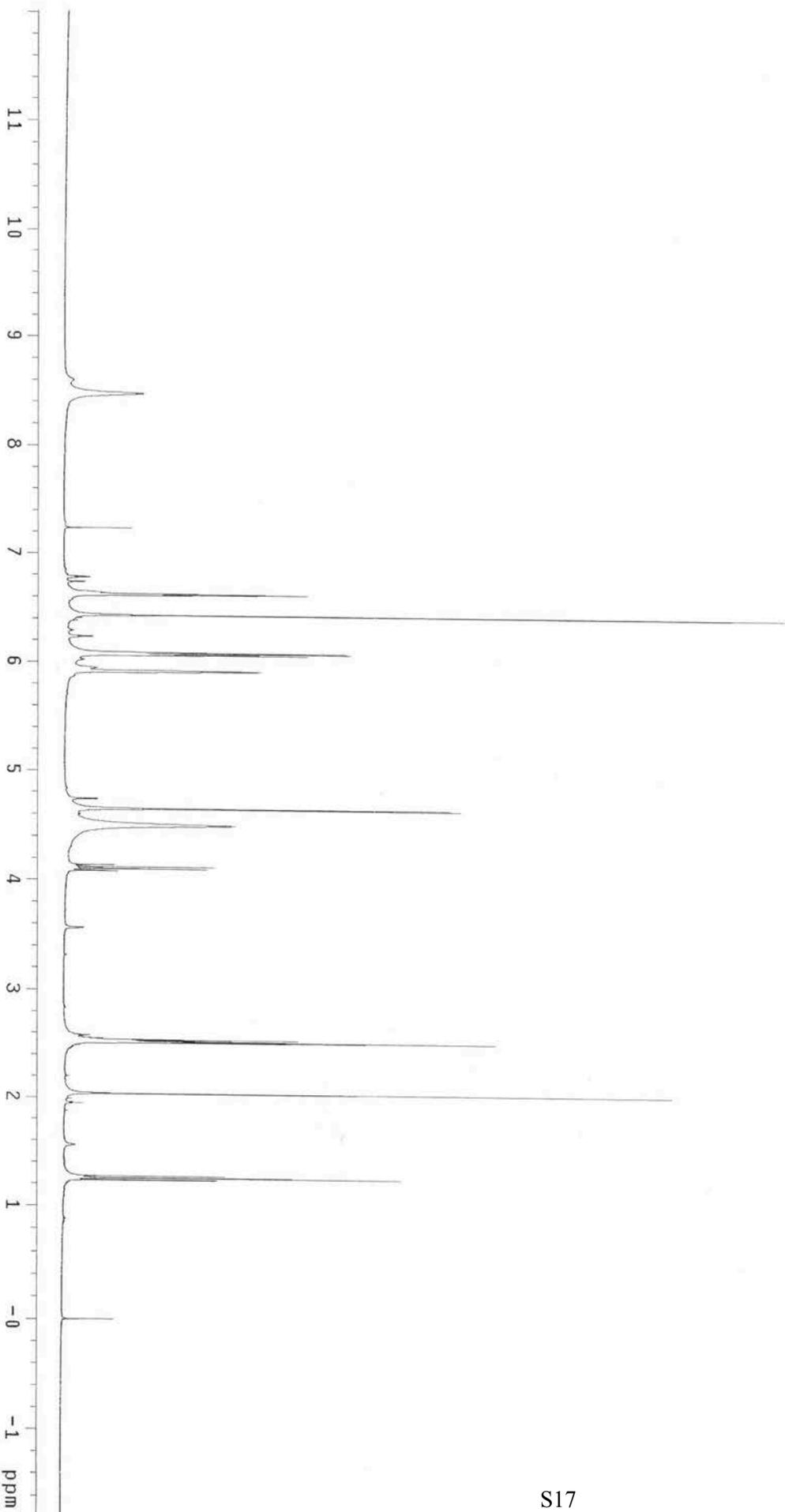
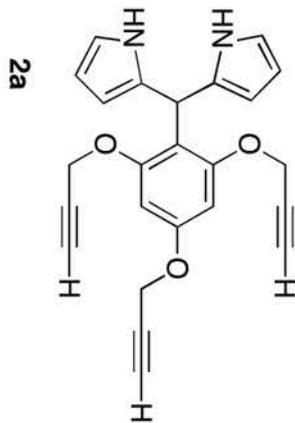
## 7. References

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- S5 O. Mass, M. Ptaszek, M. Taniguchi, J. R. Diers, H. L. Kee, D. F. Bocian, D. Holten and J. S. Lindsey, *J. Org. Chem.*, 2009, **74**, 5276–5289.
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- S7 B. J. Littler, M. A. Miller, C.-H. Hung, R. W. Wagner, D. F. O’Shea, P. D. Boyle and J. S. Lindsey, *J. Org. Chem.*, 1999, **64**, 1391–1396.
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- S9 H. L. Kee, C. Kirmaier, Q. Tang, J. R. Diers, C. Muthiah, M. Taniguchi, J. K. Laha, M. Ptaszek, J. S. Lindsey, D. F. Bocian and D. Holten, *Photochem. Photobiol.*, 2007, **83**, 1110–1124.
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## 8. Spectra

STANDARD 1H OBSERVE

Pulse Sequence: s2pu1  
Solvent: CDCl3  
Ambient temperature  
F1file: 031814\_DPM  
Mercury-400BB "ncsummerc400"  
Relax. delay 1.000 sec  
Pulse: 35.0 degrees  
Acq. time 1.993 sec  
Width 6006.0 Hz  
48 repetitions  
OBSERVE H1, 400.1245763 MHz  
DATA PROCESSING  
FT size 32768  
Total time 3 min, 4 sec



Pulse Sequence: s2pu1

Solvent: CDCl3

Ambient temperature

F1file: 031714\_benzaldehyde\_C13

Mercury-400BB "hcsuner400"

Pulse 81.2 degrees

Acq. time 1.139 sec

Width 25000.0 Hz

4070 repetitions

OBSERVE C13, 100.6113782 MHz

DECUPLE H1, 400.1266027 MHz

Power 49 dB

continuously on

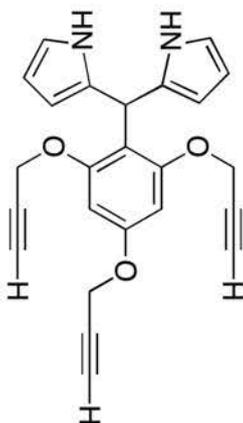
WALTZ-16 modulated

DATA PROCESSING

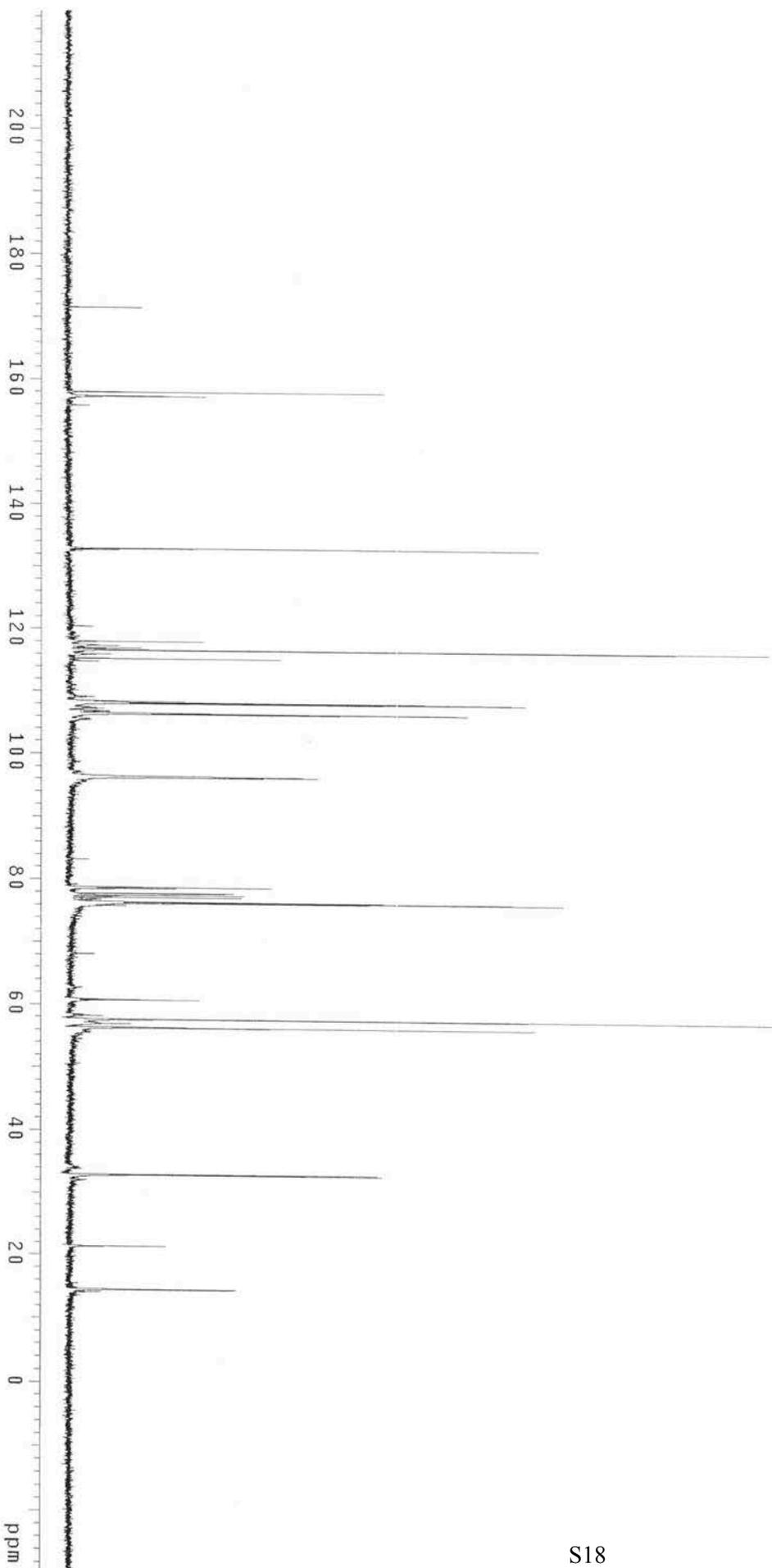
Line broadening 1.0 Hz

FT size 65536

Total time 43 hr, 20 min, 10 sec



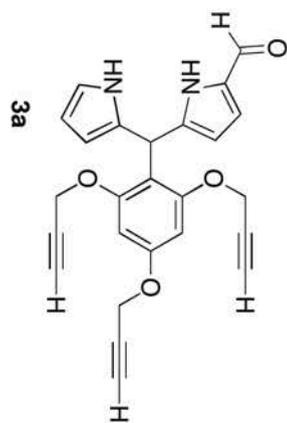
2a



STANDARD 1H OBSERVE

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Ambient temperature  
Mercury-400BB "hcsuimerc400"

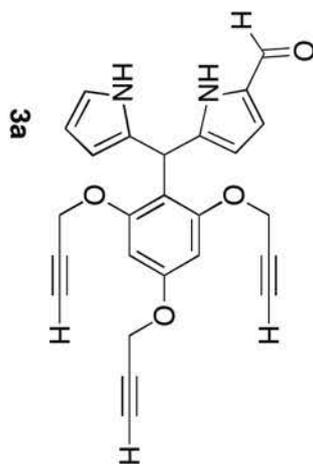
Relax. delay 1.000 sec  
Pulse -35.0 degrees  
Acq. time 1.993 sec  
Width 6006.0 Hz  
36 repetitions  
OBSERVE H1, 400.1245655 MHz  
DATA PROCESSING  
F1 size 32768  
Total time 4 min, 6 sec



13C OBSERVE

Pulse Sequence: s2pu1  
Solvent: CDCl3  
Ambient temperature  
Mercury-400BB "ncstumerc400"

Pulse: 81.2 degrees  
Acq. time: 1.199 sec  
Width: 23000.0 Hz  
382 repetitions  
OBSERVE: C13, 100.6114189 MHz  
DECUPLE: H1, 400.1266027 MHz  
Power: 44.0 dB  
continuously on  
WALTZ-16 modulated  
DATA PROCESSING  
Line Broadening: 1.0 Hz  
F1 size: 65536  
Total time: 4 hr, 20 min, 1 sec



STANDARD 1H OBSERVE

Pulse Sequence: s2pu1

Solvent: CDCl3

Ambient temperature

Mercury-300B8 "ncsummerc638"

Relax. delay 1.000 sec

Pulse 50.4 degrees

Acq. time 1.995 sec

Width 4506.5 Hz

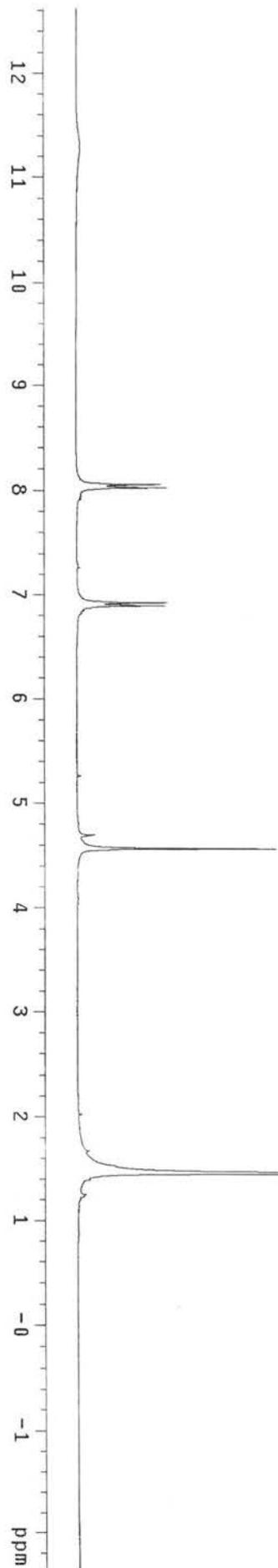
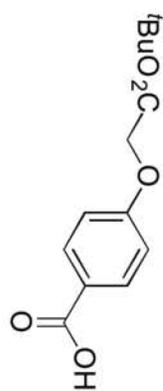
28 repetitions

OBSERVE H1, 300.1683396 MHz

DATA PROCESSING

FT size 32768

Total time 0 min, 0 sec



13C OBSERVE

Pulse Sequence: szpu1

Solvent: CDCl3

Ambient temperature

Mercury-300BB "ncsummerc638"

Pulse 23.3 degrees

Acq. time 1.815 sec

Width 18761.7 Hz

538 repetitions

OBSERVE C13, 75.4773853 MHz

DECOUPLE H1, 300.1698746 MHz

Power 36 dB

continuously on

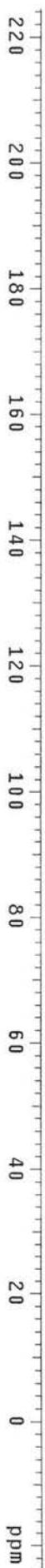
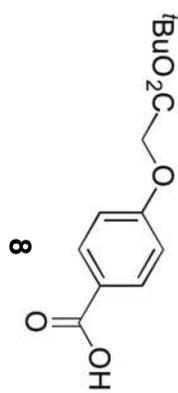
WALTZ-16 modulated

DATA PROCESSING

Line broadening 1.0 Hz

FT size 131072

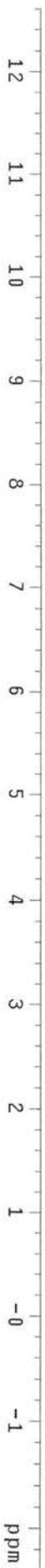
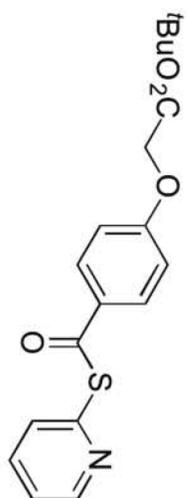
Total time 4 hr, 25 min, 3 sec



STANDARD 1H OBSERVE

Pulse Sequence: s2pu1  
Solvent: CDCl3  
Ambient temperature  
Mercury-40088 "ncsummerc400"

Relax. delay 1.000 sec  
Pulse 35.0 degrees  
Acq. time 1.993 sec  
Width 6006.0 Hz  
32 repetitions  
OBSERVE H1, 400.1245620 MHz  
DATA PROCESSING  
FT size 32768  
Total time 2 min, 3 sec

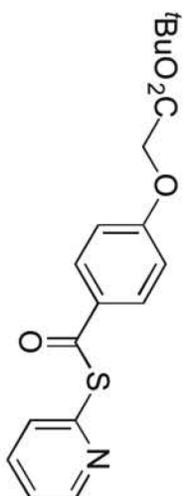


13C OBSERVE

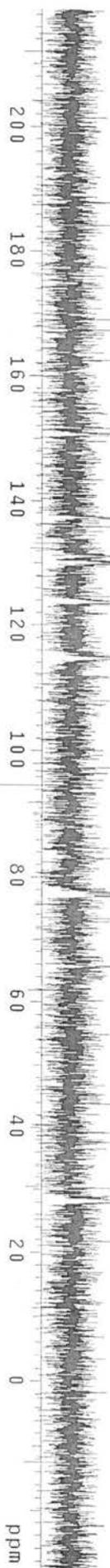
Pulse Sequence: szput

Solvent: CDCl3  
Ambient temperature  
File: 050614\_mukaiyama\_13C  
Mercury-400BB "ncsummerc400"

Pulse 81.2 degrees  
Acq. time 1.199 sec  
Width 25000.0 Hz  
696 repetitions  
OBSERVE C13 100.6113899 MHz  
DECUPLE H1 400.1256027 MHz  
Power 44 dB  
continuously on  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
FT size 65536  
Total time 4 hr, 20 min, 1 sec

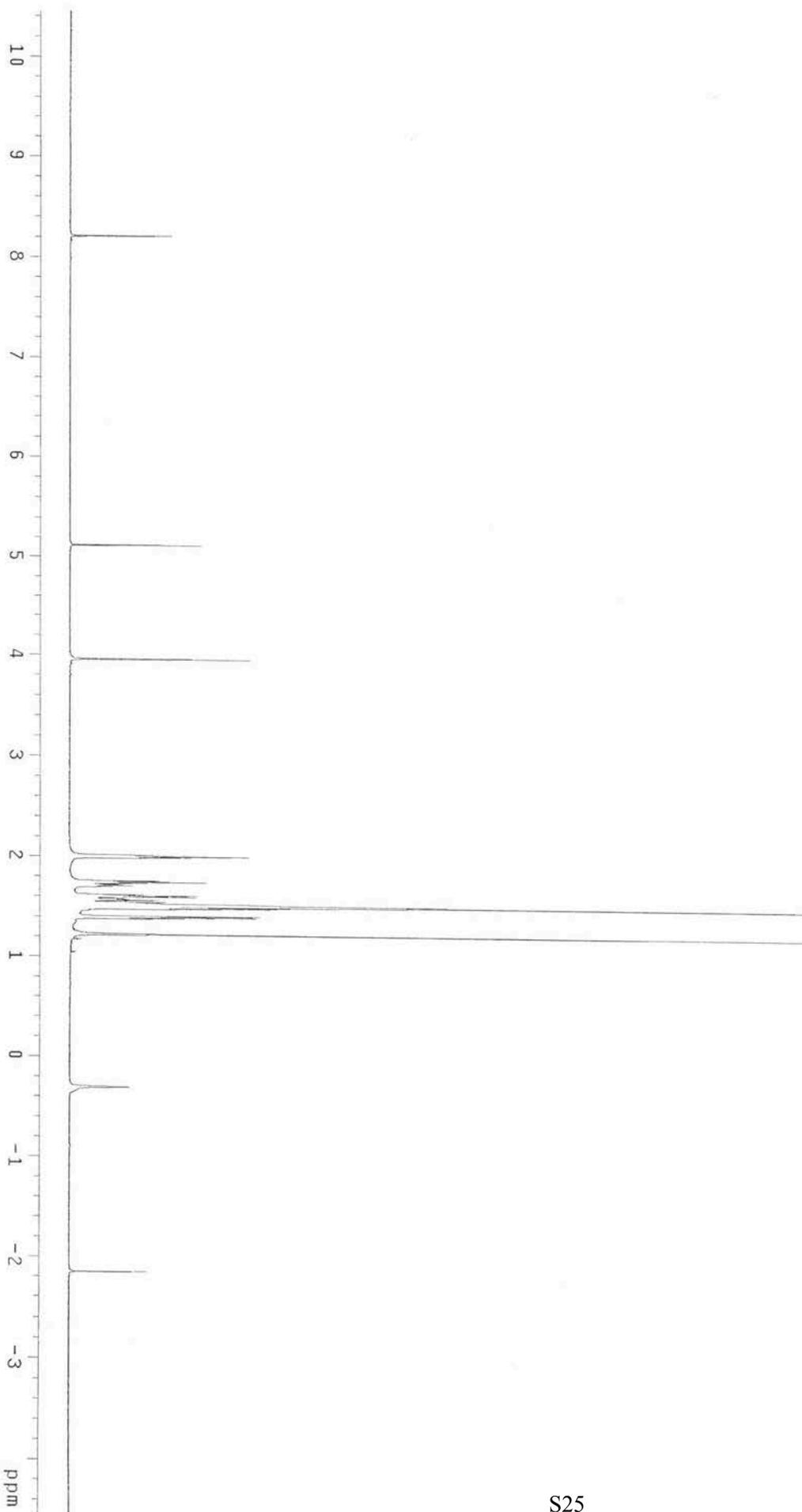
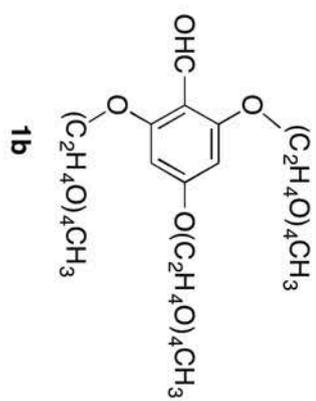


9



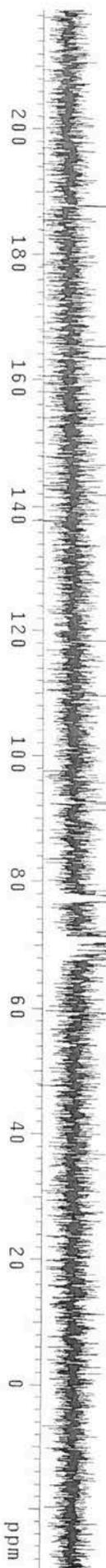
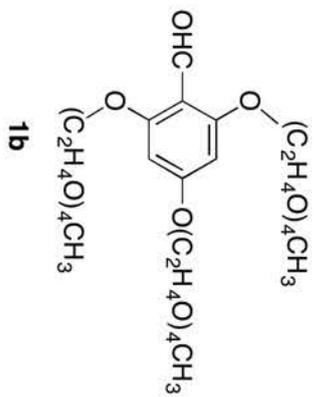
STANDARD 1H OBSERVE

Pulse Sequence: s2pu1  
Solvent: CDCl3  
Ambient temperature  
File: PEG-1  
Mercury-1000B "hcsuser/c100"  
Relax. delay 1.000 sec  
Pulse 35.0 degrees  
Acq. time 1.993 sec  
Width 6006.0 Hz  
128 repetitions  
OBSERVE H1, 400.1254230 MHz  
DATA PROCESSING  
FT size 32768  
Total time 6 min, 40 sec



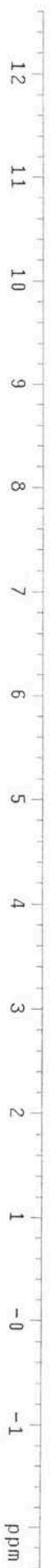
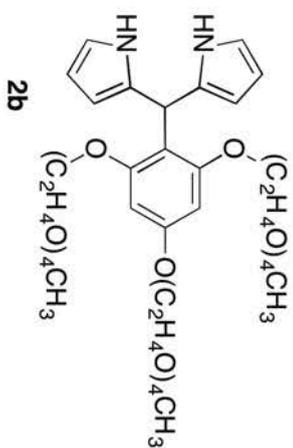
Pulse Sequence: s2pu1  
 Solvent: CDCl3  
 Ambient temperature  
 File: PEG-I-C  
 Mercury-400BB "ncsumberc400"

Pulse 81.2 degrees  
 Acq. time 1.199 sec  
 Width 23000.0 Hz  
 5120 repetitions  
 OBSERVE C13, 100.6113782 MHz  
 DECOUPLE H1, 400.1266027 MHz  
 Power 44 db  
 continuously on  
 WALTZ-16 modulated  
 DATA PROCESSING  
 Line broadening 1.0 Hz  
 FT size 65536  
 Total time 2 hr, 5 min, 8 sec



STANDARD 1H OBSERVE

Pulse Sequence: s2pu1  
Solvent: CDCl3  
Ambient temperature  
File: PEG2  
Mercury-400DB "ncsumerc400"  
Relax. delay 1.000 sec  
Pulse 35.0 degrees  
Acq. time 1.993 sec  
Width 6006.0 Hz  
256 repetitions  
OBSERVE H1, 400.1245645 MHz  
DATA PROCESSING  
FT size 32768  
Total time 13 min, 20 sec



13C OBSERVE

Pulse Sequence: s2pul

Solvent: CDCl3

Ambient temperature

File: PEG2-C  
Mercury-400BB "ncsumerc400"

Pulse 81.2 degrees

Acq. time 1.199 sec

Width 25000.0 Hz

5120 repetitions

OBSERVE C13, 100.6113782 MHz

DECUPLE H1, 400.1266027 MHz

Power 44 dB

continuously on

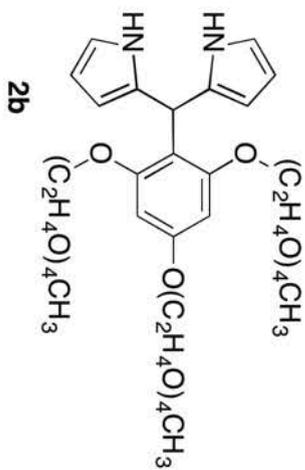
WALTZ-16 modulated

DATA PROCESSING

Line broadening 1.0 Hz

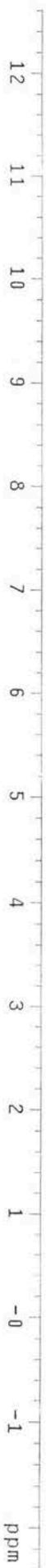
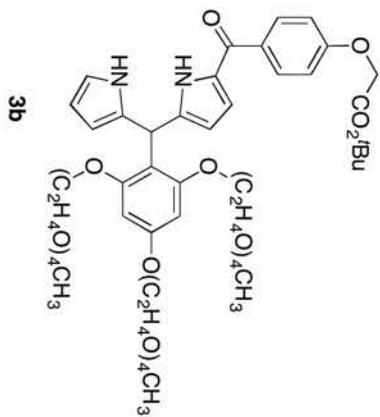
FT size 65536

Total time 2 hr, 5 min, 8 sec



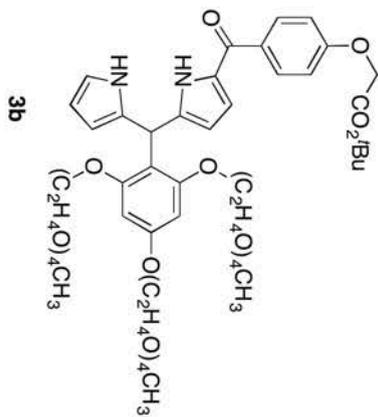
STANDARD 1H OBSERVE

Pulse Sequence: s2pu1  
Solvent: CDCl3  
Ambient temperature  
File: PEG3  
Mercury-400BB "ncsunmerc400"  
Relax. delay 1.000 sec  
Pulse 35.0 degrees  
Acq. time 1.993 sec  
Width 6006.0 Hz  
128 repetitions  
OBSERVE H1, 400.1245609 MHz  
DATA PROCESSING  
FT size 32768  
Total time 6 min, 40 sec



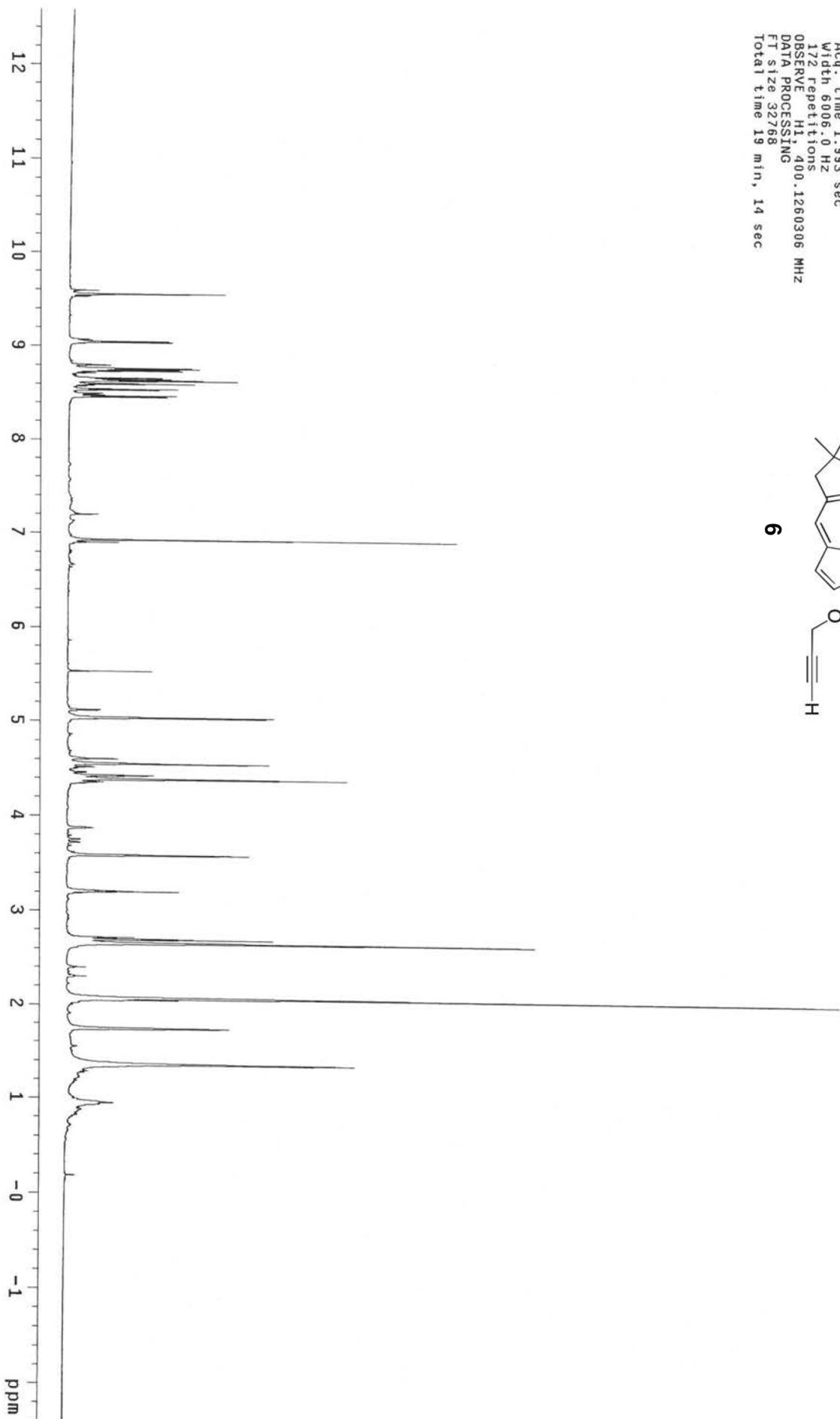
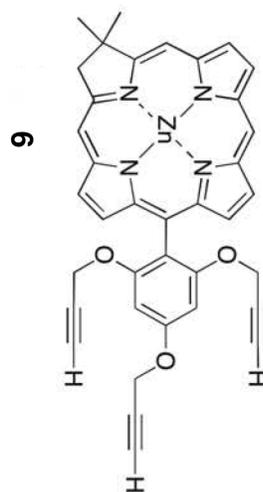
Pulse Sequence: s2pu1  
 Solvent: CDCl3  
 Ambient temperature  
 File: PEG3-c  
 Mercury-40088 "ncsummerc400"

Pulse: 81.2 degrees  
 Acq. time: 1.199 sec  
 Width: 25000.0 Hz  
 SIZO: repelitions  
 OBSERVE: C13, 100.6113782 MHz  
 DECOUPLE: H1, 400.1266027 MHz  
 Power: 44 dB  
 continuously on  
 WALTZ-16 modulated  
 DATA PROCESSING  
 Line broadening: 1.0 Hz  
 F1 size: 85536  
 Total time: 2 hr, 5 min, 8 sec



STANDARD 1H OBSERVE

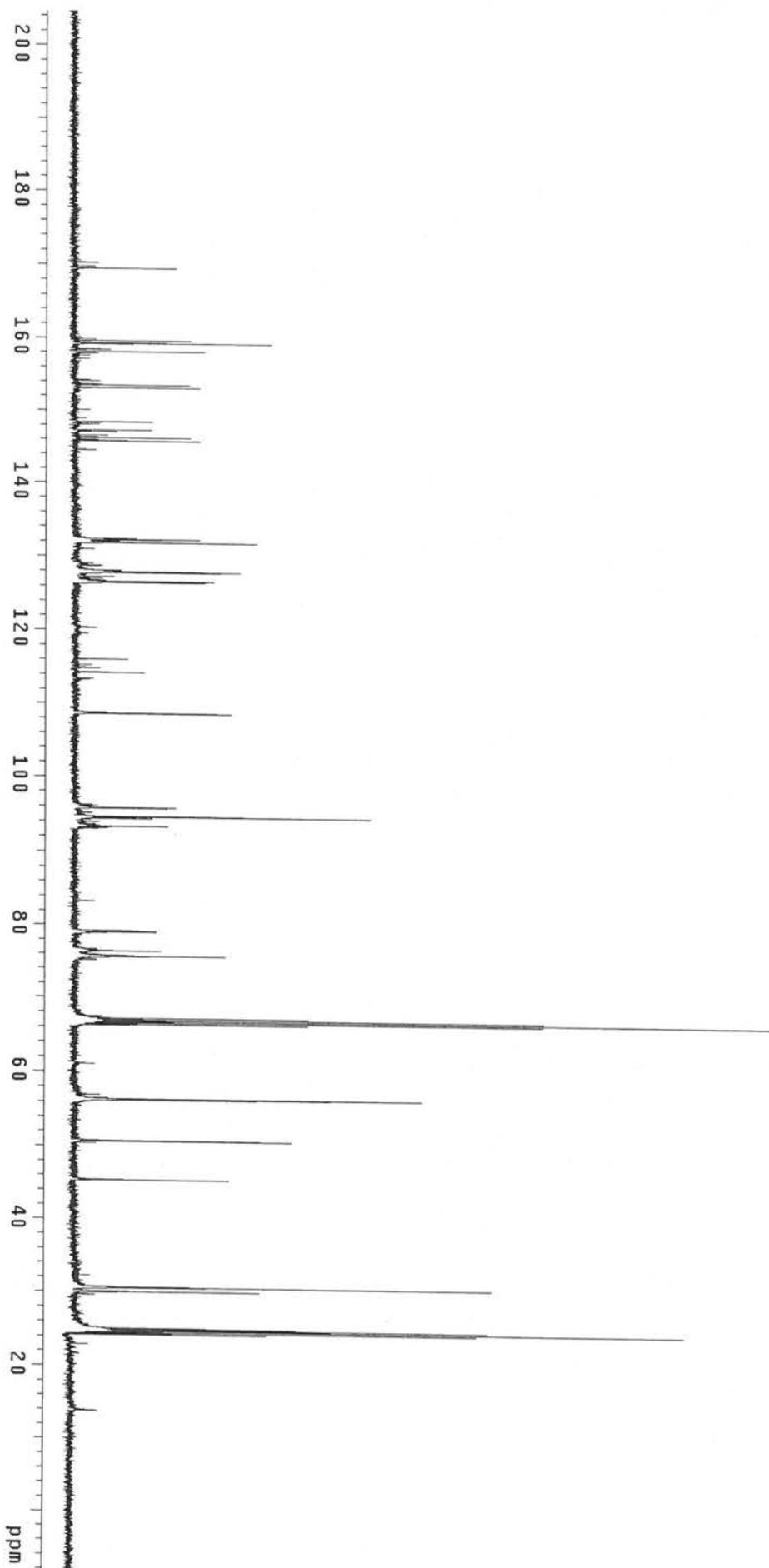
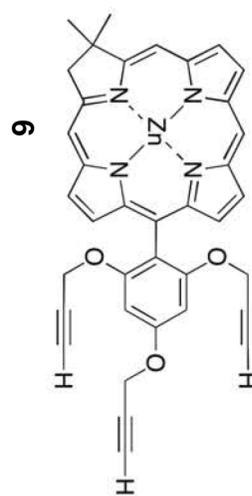
Pulse Sequence: s2pu1  
Solvent: THF  
Ambient temperature  
Mercury-400BB "ncsummerc400"  
Relax. delay 1.000 sec  
Pulse 35.0 degrees  
Acq. time 1.993 sec  
Width 6006.0 Hz  
172 repetitions  
OBSERVE H1, 400.1260306 MHz  
DATA PROCESSING  
F1 size 32768  
Total time 19 min, 14 sec



13C OBSERVE

Pulse Sequence: szpul  
Solvent: thf  
Ambient temperature  
Mercury-400BB "ncsummerc400"

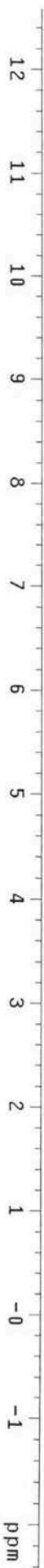
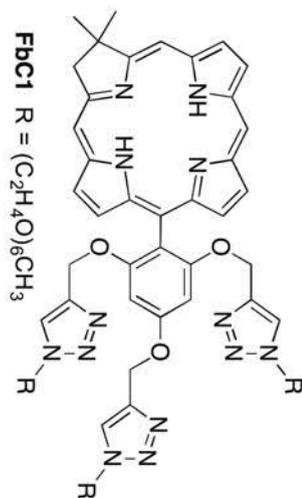
Pulse 81.2 degrees  
Acq. time 1.199 sec  
Width 25000.0 Hz  
5696 repetitions  
OBSERVE C13, 100.6117464 MHz  
DECUPLE H1, 400.1280672 MHz  
Power 44 dB  
Continuously on  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
FT size 65536  
Total time 43 hr, 20 min, 10 sec





STANDARD 1H OBSERVE

Pulse Sequence: s2pu1  
Solvent: THF  
Ambient temperature  
Mercury-400BB "ncsumerc400"  
Relax. delay 1.000 sec  
Pulse 45.0 degrees  
Acq. time 1.995 sec  
Width 6006.0 Hz  
64 repetitions  
OBSERVE H1, 400.1260343 MHz  
DATA PROCESSING  
FT size 32768  
Total time 4 min, 6 sec

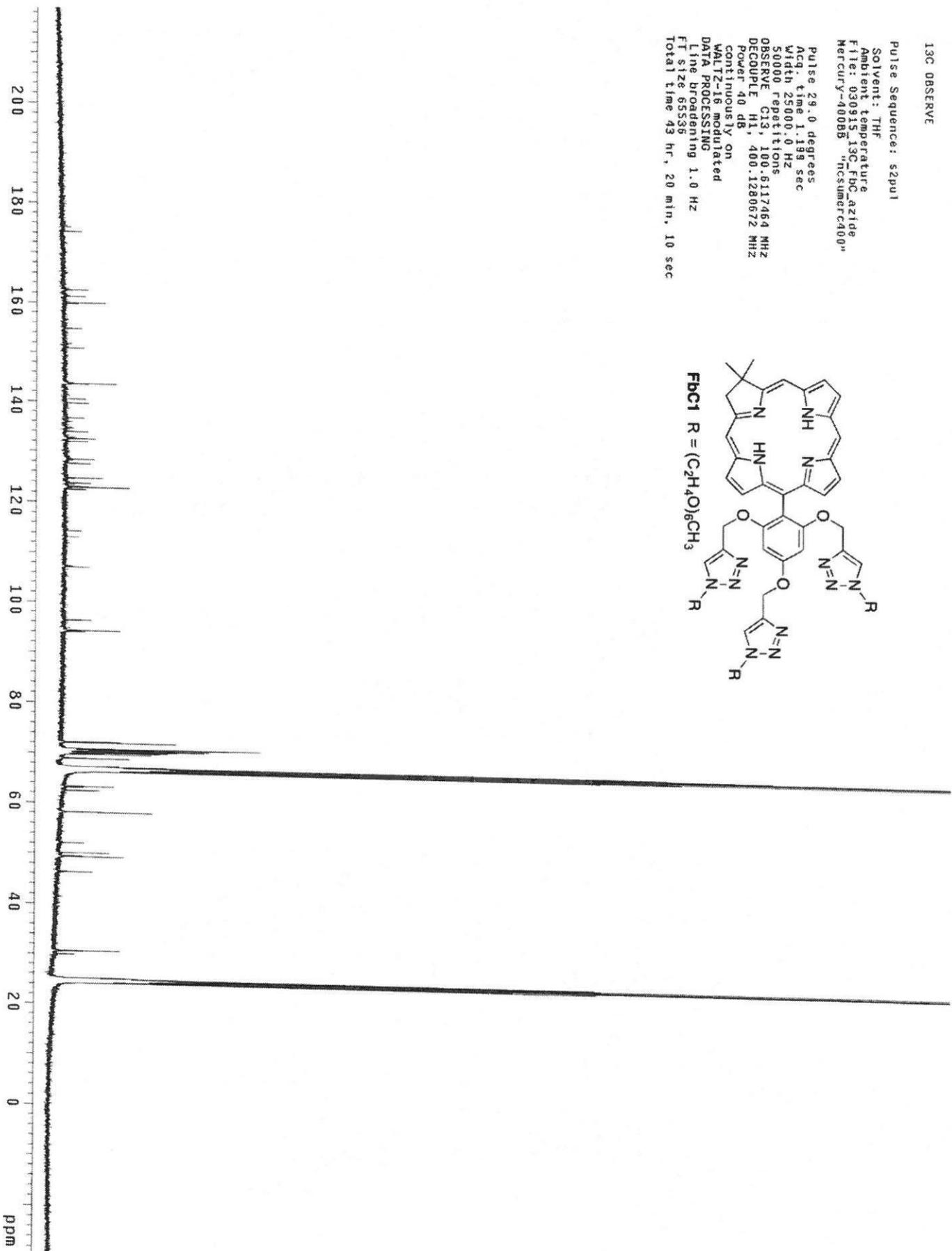
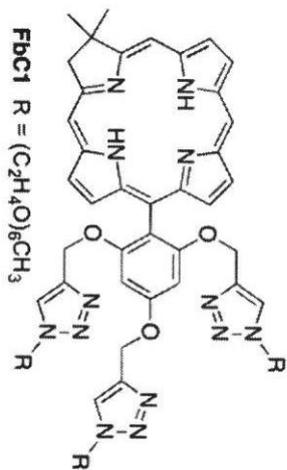


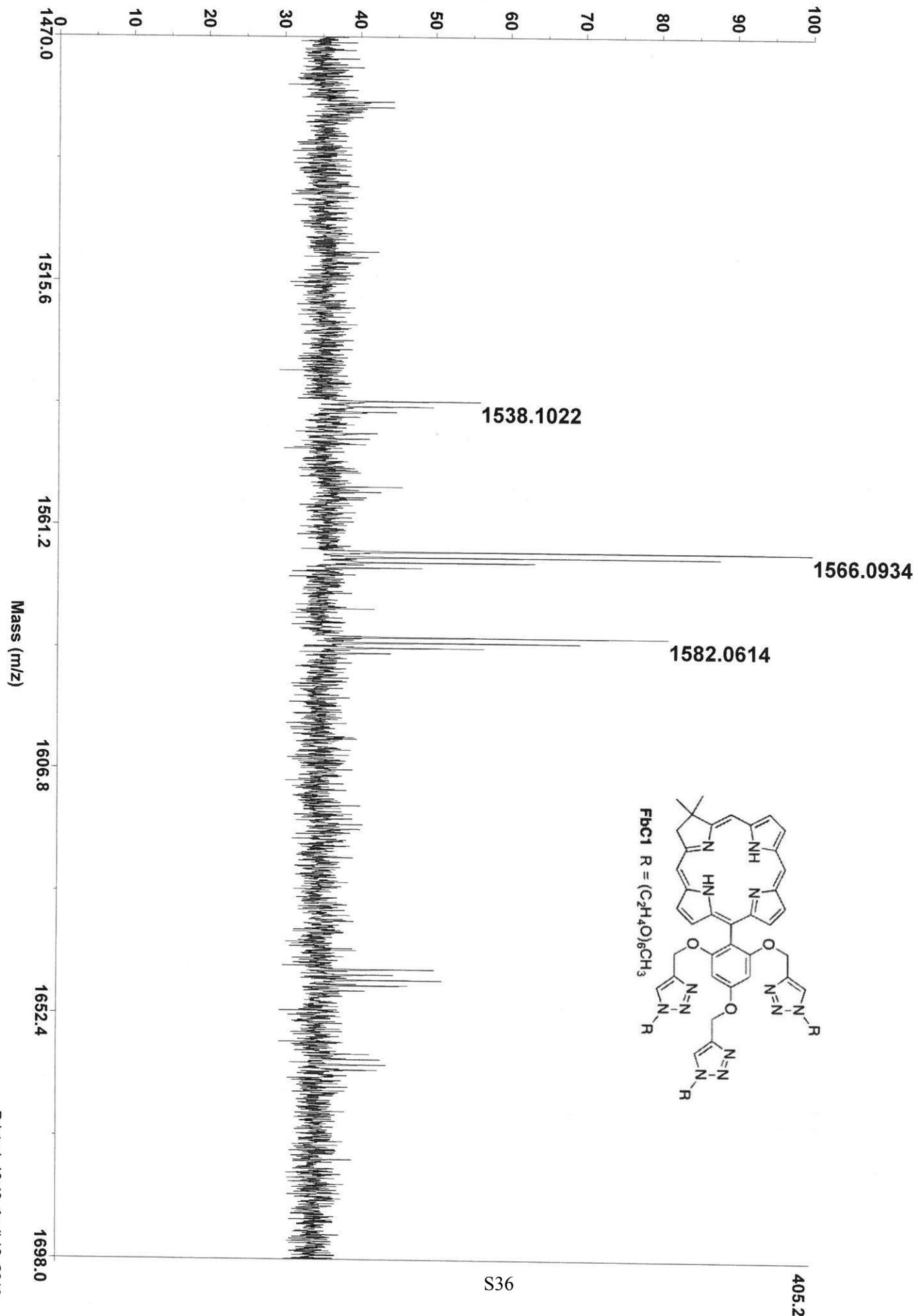
13C OBSERVE

Pulse Sequence: s2pu1

Solvent: THF  
Ambient temperature  
File: 030915\_13C\_FbC\_Azide  
Mercury-400BS "ncsummerc400"

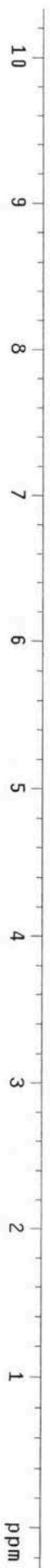
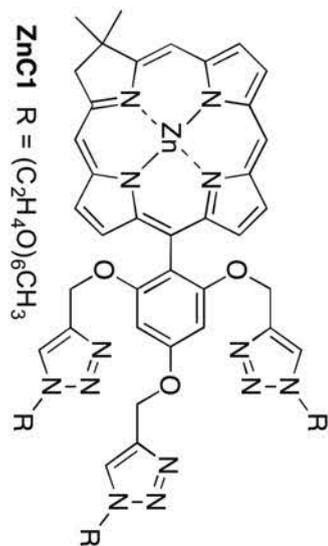
Pulse 29.0 degrees  
Acq. time 1.199 sec  
Width 25000.0 Hz  
50000 repetitions  
OBSERVE C13, 100.6117464 MHz  
DECOUPLE H1, 400.1280672 MHz  
Power 40 dB  
continuously on  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
Ft size 65535  
Total time 43 hr, 20 min, 10 sec





STANDARD 1H OBSERVE

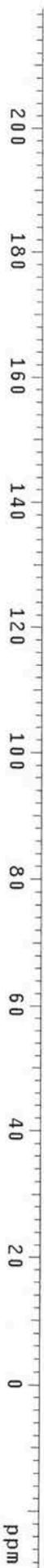
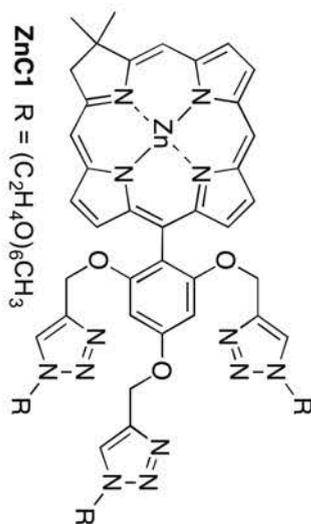
Pulse Sequence: s2pu1  
Solvent: THF  
Ambient temperature  
Mercury-40088 "ncsmerc400"  
Relax. delay 1.000 sec  
Pulse time 1.991 sec  
Acq. time 1.991 sec  
Width 6006.0 Hz  
64 repetitions  
OBSERVE H1, 400.126789 MHz  
DATA PROCESSING  
FT size 32768  
Total time 4 min, 6 sec

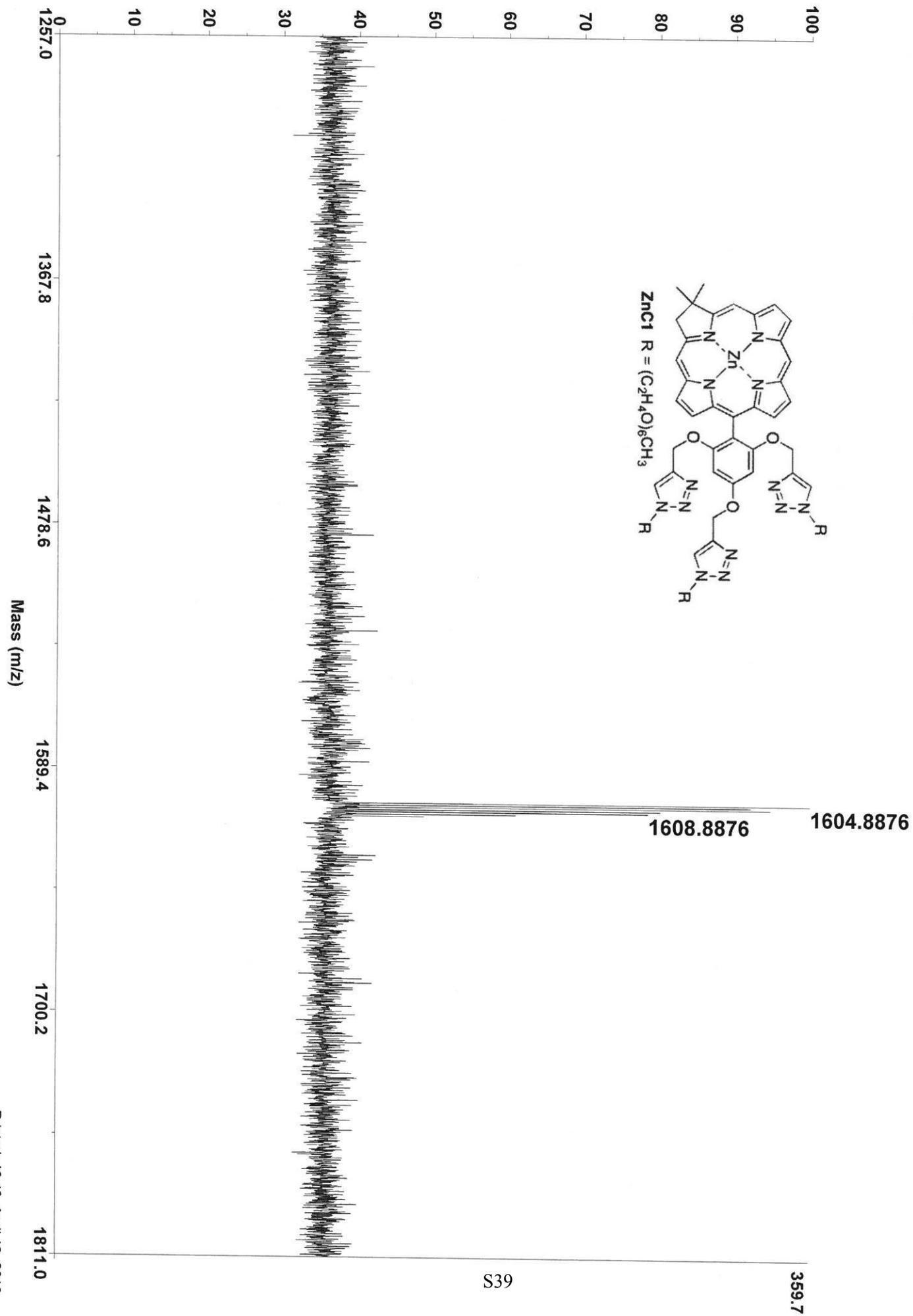
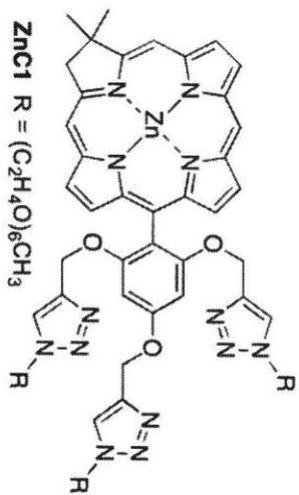


Pulse Sequence: szpu1

Solvent: THF  
 Ambient temperature  
 File: 030515\_13C\_Click\_LXn  
 Mercury-400BB "ncsmmerc400"

Pulse 29, 0 degrees  
 Acq. time 1.199 sec  
 Width 23000.0 Hz  
 20000 repetitions  
 OBSERVE C13, 100.6117464 MHz  
 DECOUPLE H1, 400.1280672 MHz  
 Power 40 dB  
 continuously on  
 WALTZ-16 modulated  
 DATA PROCESSING  
 Line broadening 1.0 Hz  
 FT size 65536  
 Total time 17 hr, 20 min, 4 sec





STANDARD IN OBSERVE

Pulse Sequence: zgpg30

Solvent: CDCl3

Ambient Temperature

F1 file: psg-znc-l-bu-tong01

Mercury-400DBB "NucTimerC400"

Relax. delay: 5.000 sec

Pulse: 35.0 degrees

Acq. time: 1.395 sec

Width: 8009.0 Hz

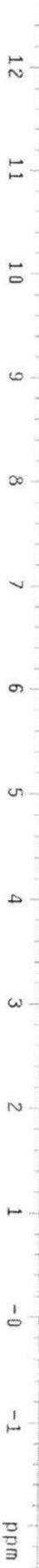
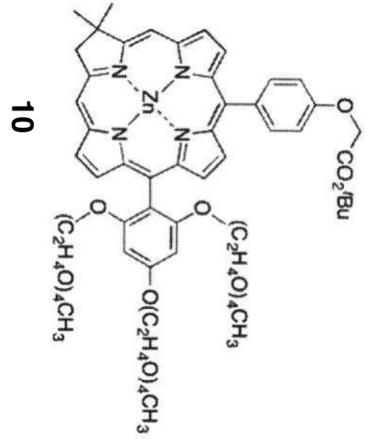
128 repetitions

OBSERVE: H1, 400.1245653 MHz

DATA PROCESSING

FT size: 32768

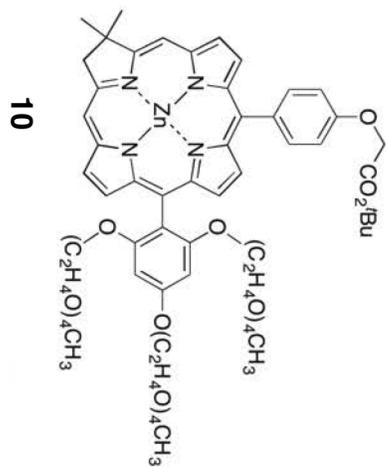
Total time: 15 min, 12 sec



13C OBSERVE

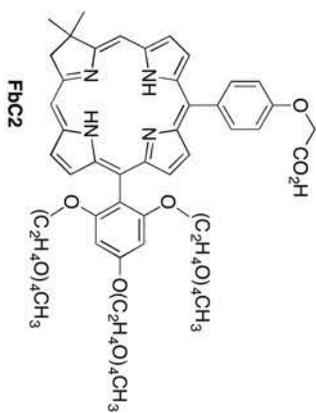
Pulse Sequence: szpul  
Solvent: CDCl3  
Ambient Temperature  
File: peg-znc-t-bu-13c  
Mercury-400BB "ncsummerc100"

Pulse 81.2 degrees  
Acq. time 1.199 sec  
Width 25000.0 Hz  
40960 repetitions  
OBSERVE C13, 100.6114037 MHz  
DECUPLE H1, 400.1256027 MHz  
Power 44 dB  
continuously on  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
FT size 65536  
Total time 16 hr, 41 min, 6 sec



STANDARD 1H OBSERVE

Pulse Sequence: s2pu1  
 Solvent: CDCl3  
 Ambient temperature  
 File: peg-fbc-fa-10mgD1-2  
 Mercury-400BB "hcsimmerc400"  
 Relax. delay 5.000 sec  
 Pulse: 35.0 degrees  
 Acq. time 1.993 sec  
 Width 6006.0 Hz  
 256 repetitions  
 OBSERVE H1, 400.1245656 MHz  
 DATA PROCESSING  
 Line broadening 0.5 Hz  
 FI size 32768  
 Total time 30 min, 24 sec



Pulse Sequence: s2pu1

Solvent: CDCl3

Ambient temperature

File: peg-fbc-fa-13C-2

Mercury-40088 "msumerca400"

Pulse: 81.2 degrees

Acq. Time: 1.199 sec

Width: 25000.0 Hz

51200 repetitions

OBSERVE: G13, 100.6114037 MHz

DECUPLE: H1, 400.1266027 MHz

Power: 44 dB

continuously on

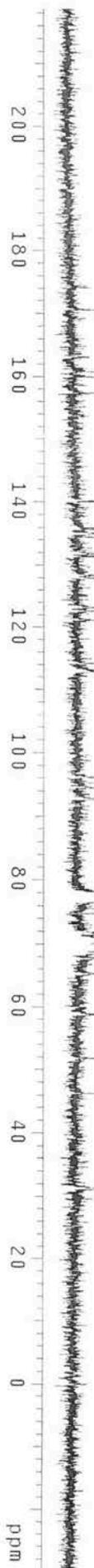
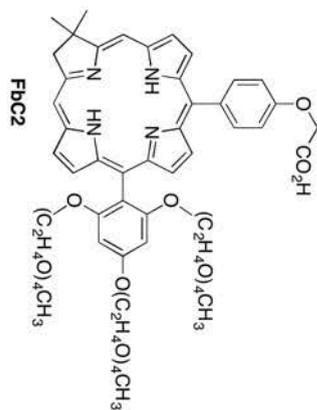
WALTZ-16 modulated

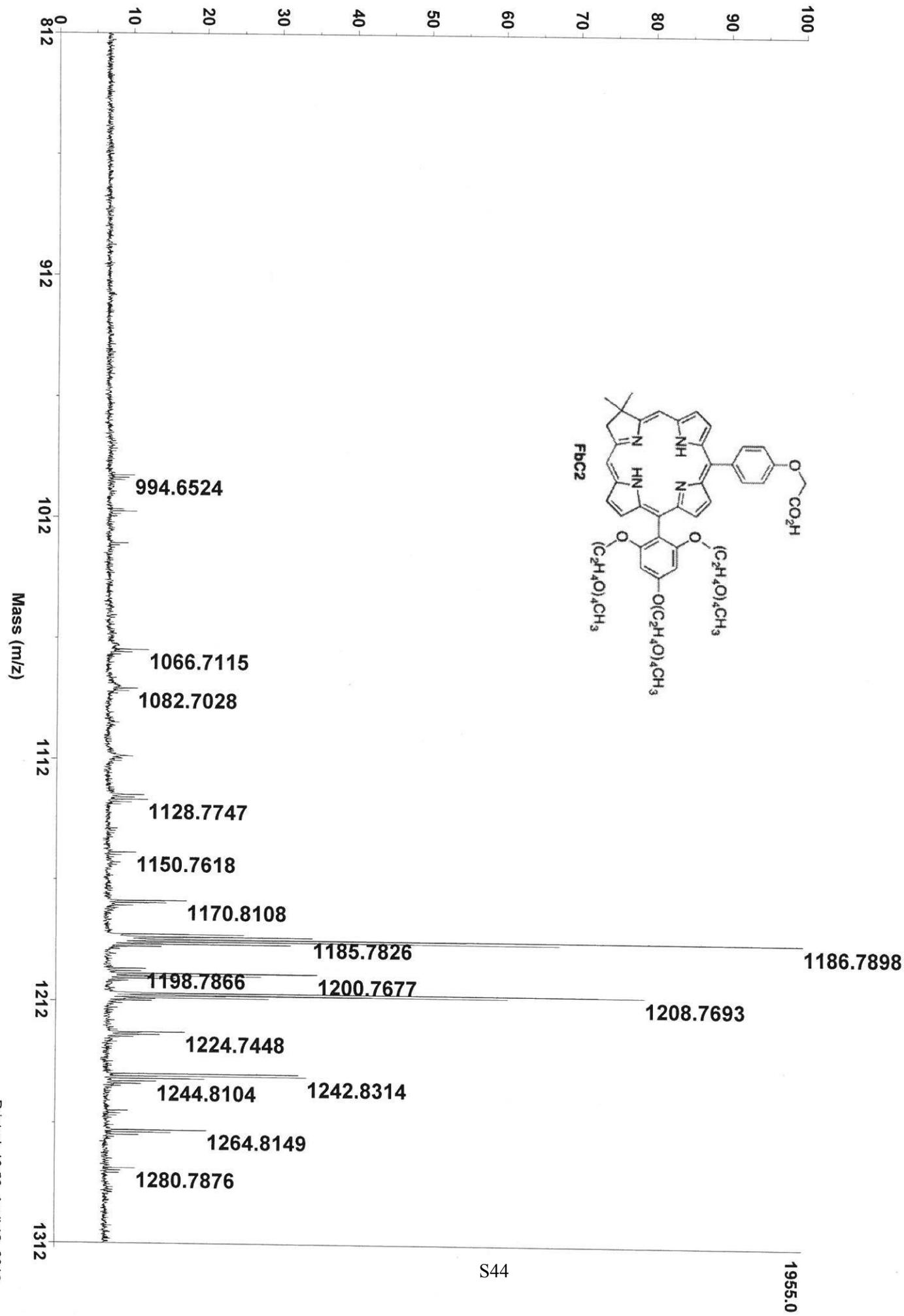
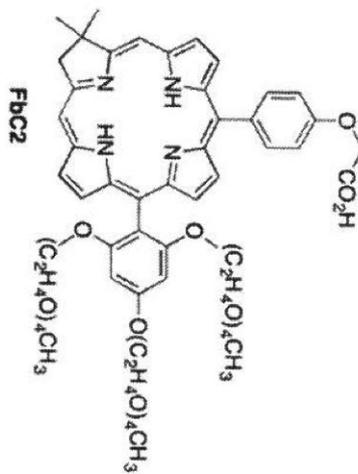
DATA PROCESSING

Line broadening: 1.0 Hz

FT size: 65536

Total time: 20 hr, 51 min, 22 sec



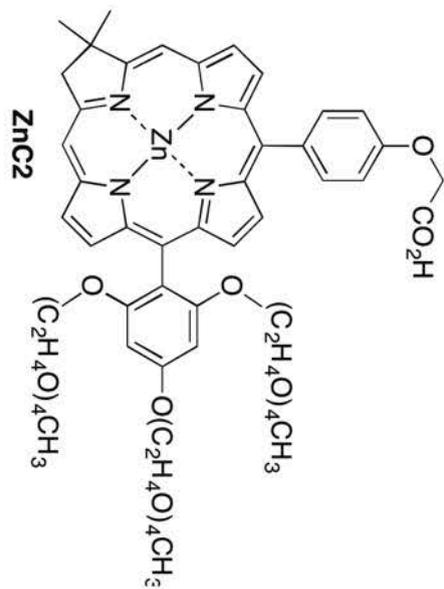


STANDARD 1H OBSERVE

Pulse Sequence: s2pu1

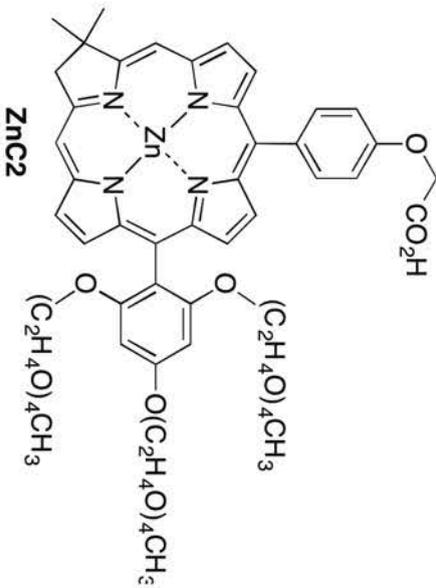
Solvent: CDCl3  
Ambient temperature  
File: peg-znc-fa-10mg01  
Mercury-30088 "hcnsmc638"

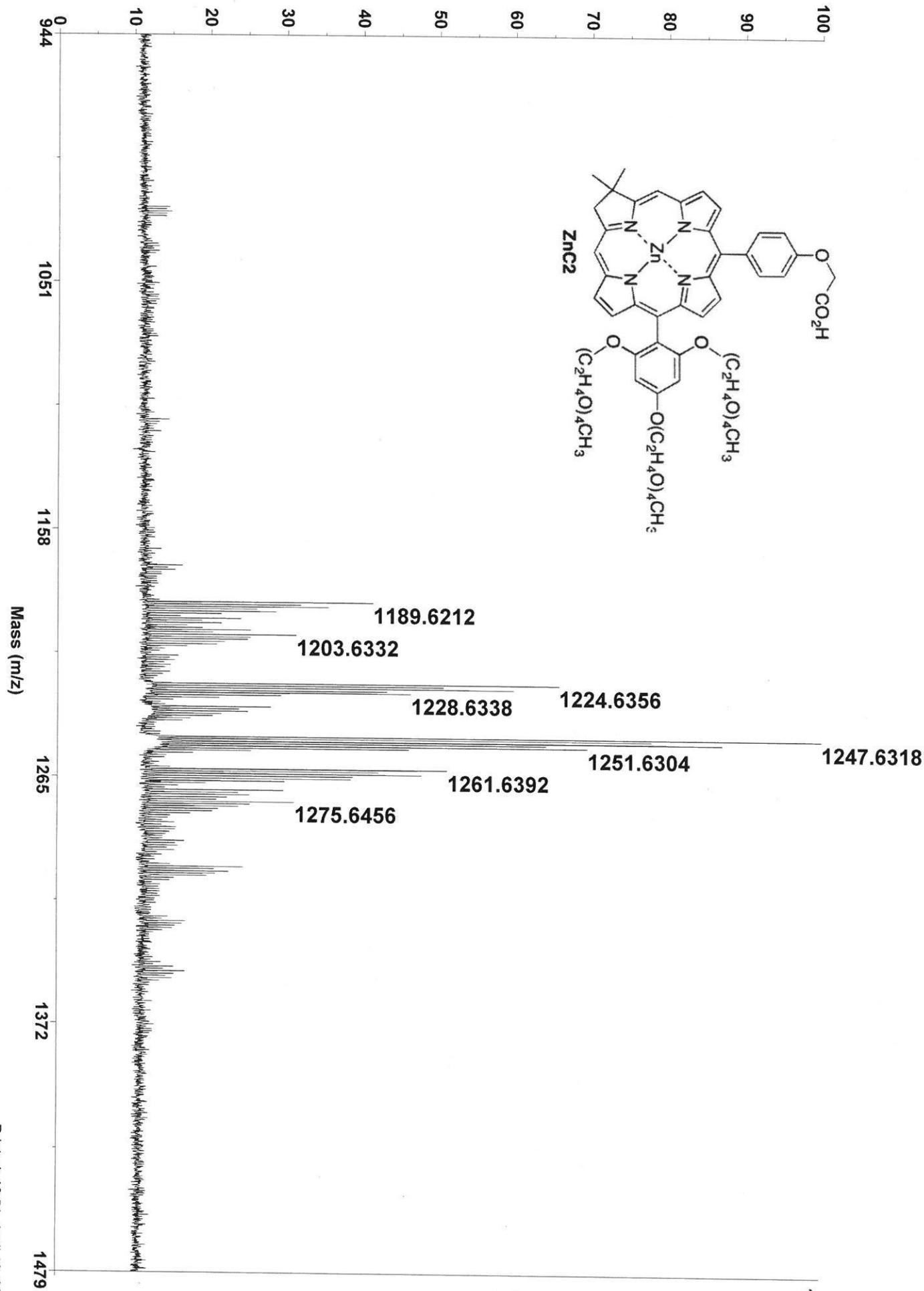
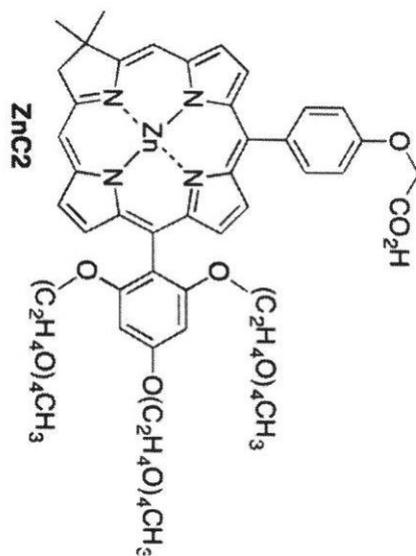
Relax. delay 5.000 sec  
Pulse 86.4 degrees  
Acq. time 1.995 sec  
Width 4506.5 Hz  
256 repetitions  
OBSERVE H1 300.1683379 MHz  
DATA PROCESSING  
F1 size 32768  
Total time 0 min, 0 sec



13C OBSERVE

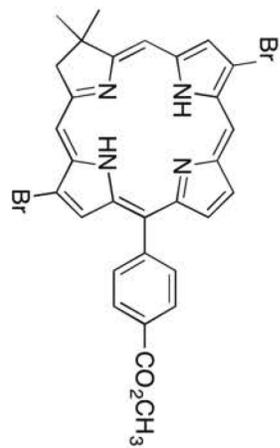
Pulse Sequence: szpul  
Solvent: CDCl3  
Ambient temperature  
File: peg-znc-fa-c  
Mercury-300BB "nsummerc638"  
Pulse 23.3 degrees  
Acq. time 1.815 sec  
Width 18761.7 Hz  
40960 repetitions  
OBSERVE C13, 75.4773710 MHz  
DECUPLE H1, 300.1698746 MHz  
Power 36 dB  
continuously on  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
FT size 131072  
Total time 24 hr, 1 min, 52 sec





STANDARD 1H OBSERVE

Pulse Sequence: s2pu1  
Solvent: CDCl3  
Ambient Temperature  
Mercury-400DB "nucsummerc400"  
Relax. delay: 1.000 sec  
Pulse: 78.7 degrees  
Acq. time: 1.993 sec  
Width: 6006.0 Hz  
64 repetitions  
OBSERVE: H1, 400.1245785 MHz  
DATA PROCESSING  
F1 size: 32768  
Total time: 9 min, 6 sec



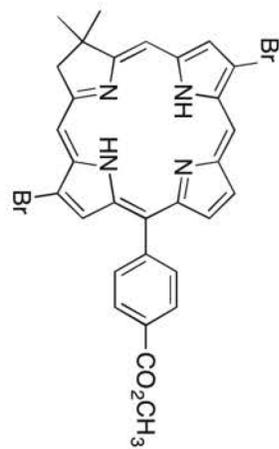
12



13C OBSERVE

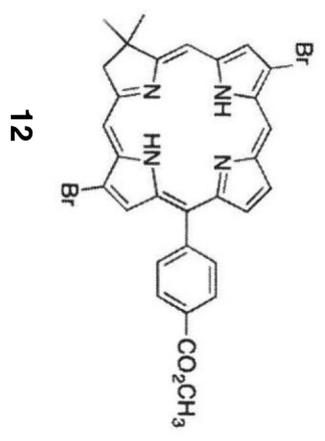
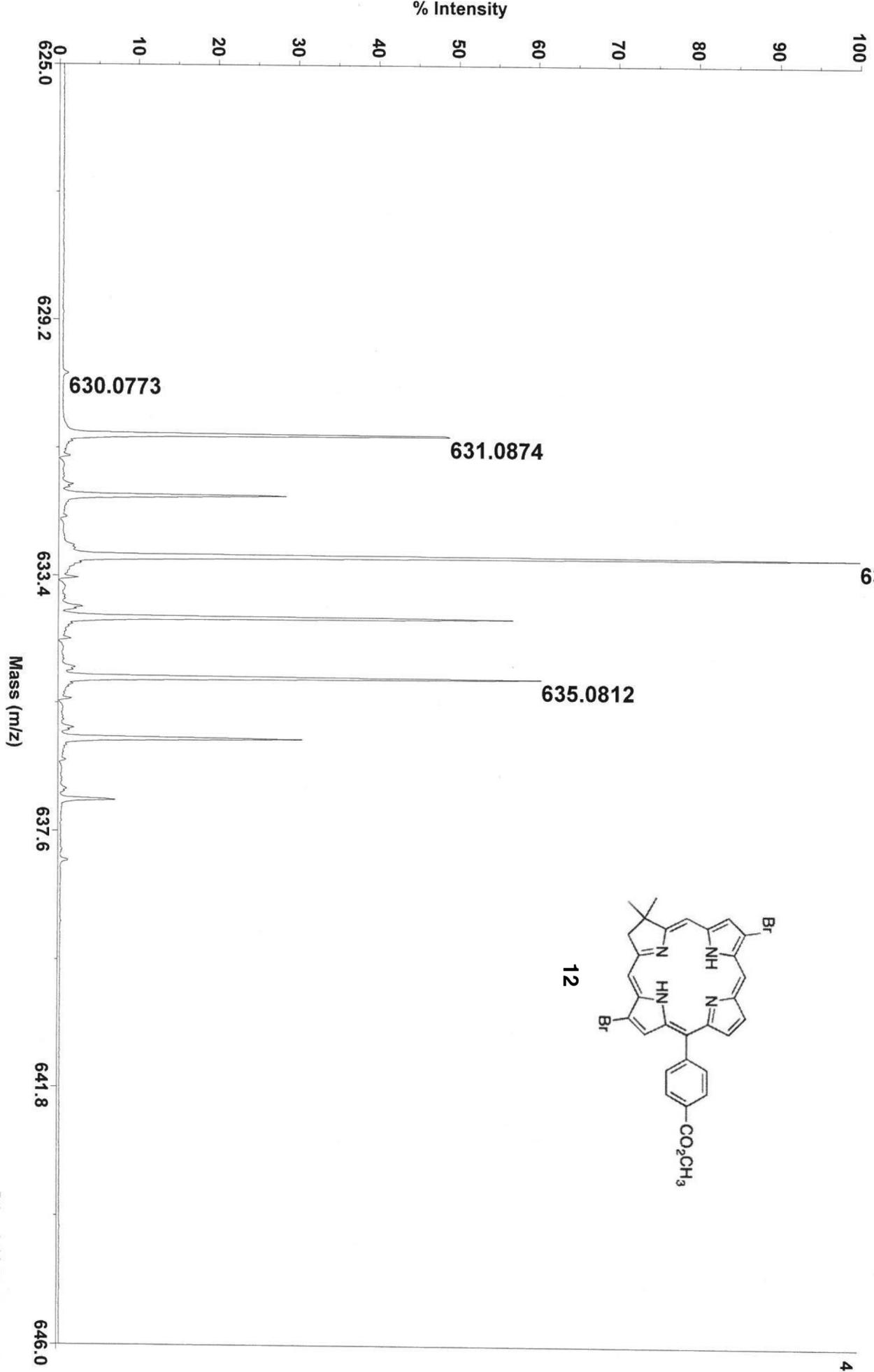
Pulse Sequence: s2pu1  
Solvent: CDCl3  
Ambient temperature  
Mercury-400BB "ncsmerc400"

Pulse 46.9 degrees  
Acq. time 1.199 sec  
Width 25000.0 Hz  
6880 repetitions  
OBSERVE C13, 100.6113806 MHz  
DECOUPLE H1, 400.1266027 MHz  
Power 39 dB  
continuously on  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
FT size 65536  
Total time 8 hr, 40 min, 2 sec



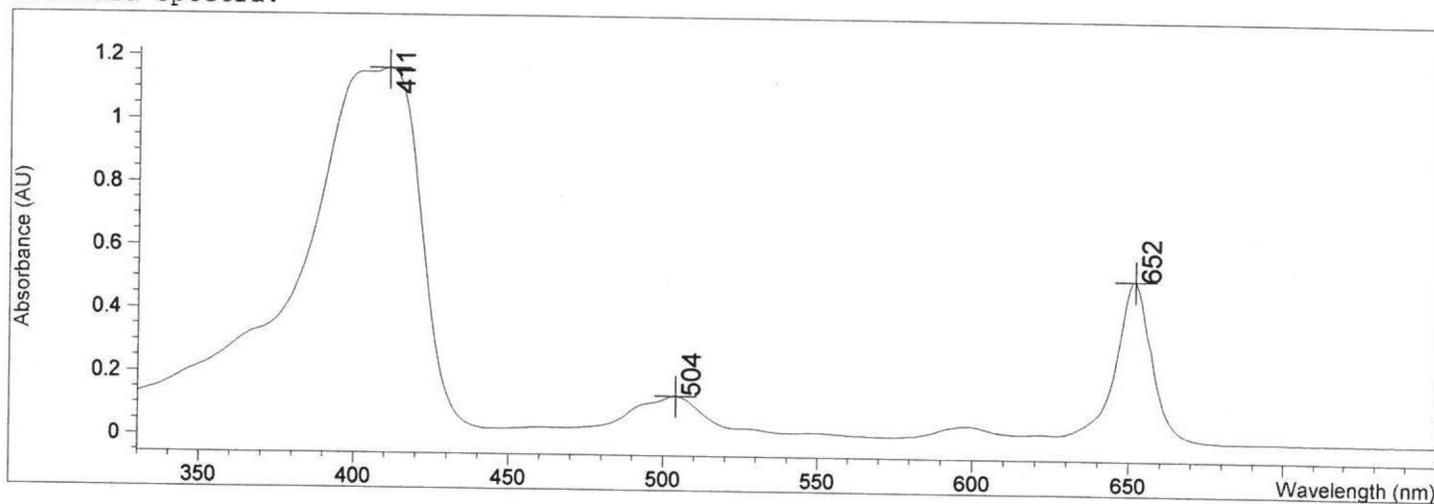
12



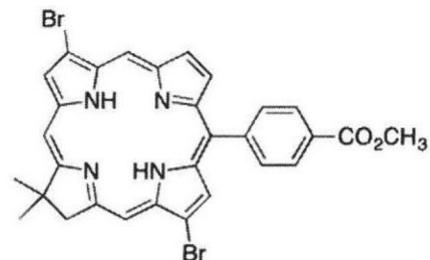


Method file : <method not saved>  
Information : Default Method  
Data File : C:\CHEM32\1\DATA\MENGRAN\110414\_DIBROMO\_PH\_COOME\_TOLUENE.SD  
Created : 11/4/14 11:39:10

## Overlaid Spectra:



#	Name	Peaks (nm)	Abs (AU)
1		411.0	1.16240
1		652.0	0.51388
1		504.0	0.13340

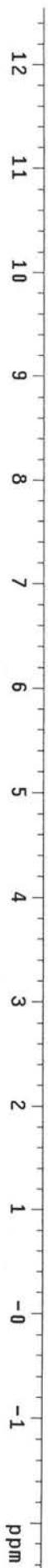
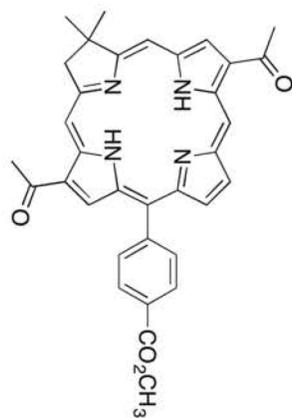
**12**

Report generated by : jon

\*\*\* End Spectrum/Peak Report \*\*\*

STANDARD 1H OBSERVE

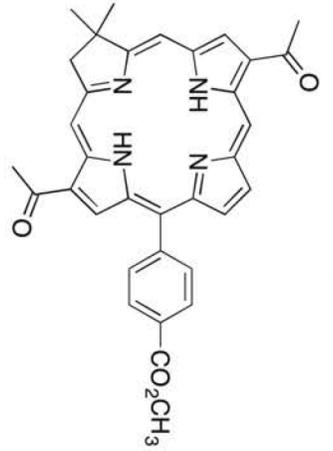
Pulse Sequence: szput  
Solvent: CDCl3  
Ambient temperature  
Mercury-400BB "ncsummerc400"  
Relax. delay 1.000 sec  
Pulse 78.7 degrees  
Acq. time 1.993 sec  
Width 6006.0 Hz  
32 repetitions  
OBSERVE H1 400.1245843 MHZ  
DATA PROCESSING  
F1 size 32768  
Total time 2 min, 3 sec



13C OBSERVE

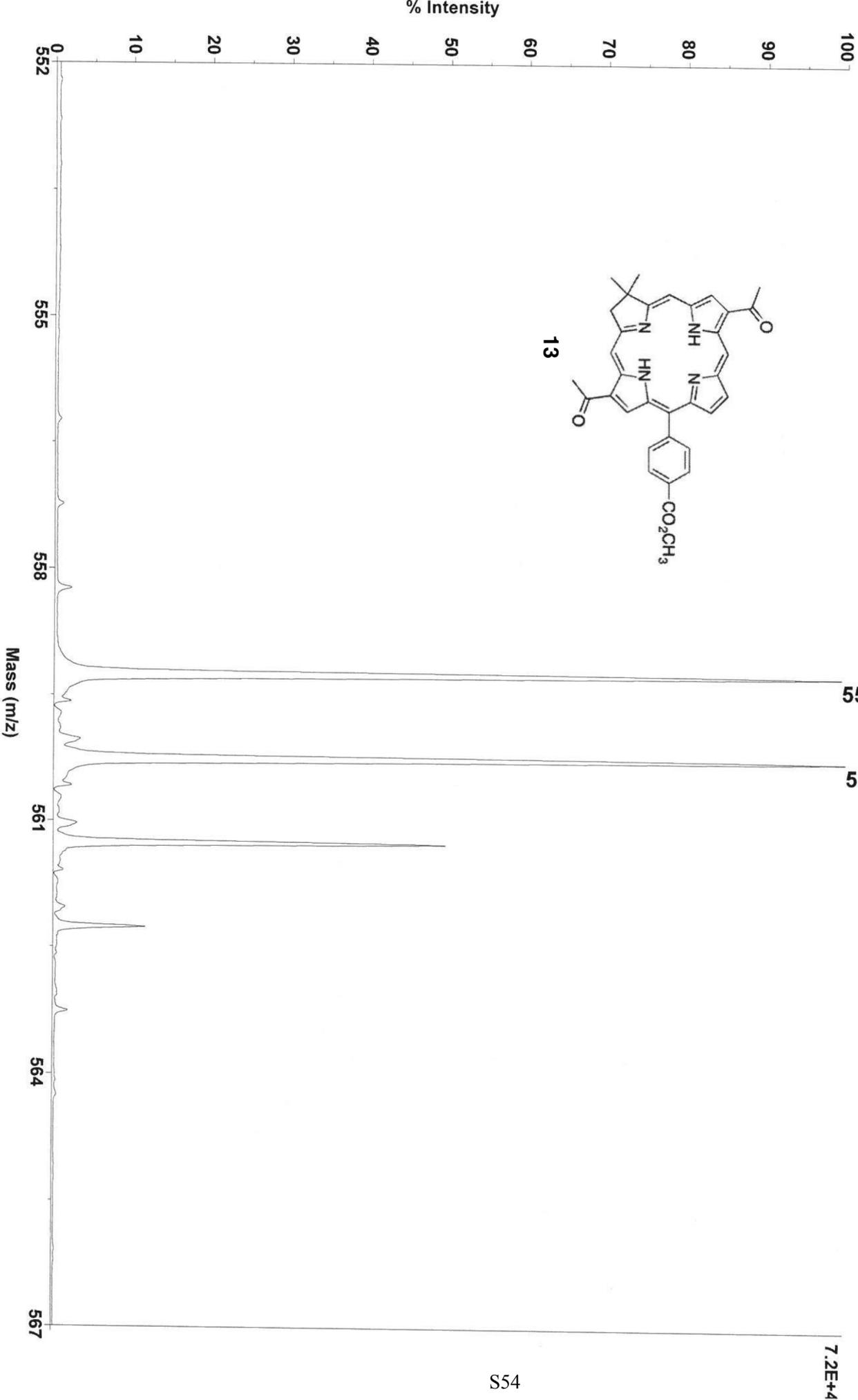
Pulse Sequence: szpu1  
Solvent: CDCl3  
Ambient temperature  
Mercury-400BB "ncsummerc400"

Pulse 46.9 degrees  
Acq. time 1.199 sec  
Width 25000.0 Hz  
3028 repetitions  
OBSERVE C13, 100.6113859 MHz  
DECUPLE H1, 400.1266027 MHz  
Power 39 dB  
continuously ON  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
FT size 65536  
Total time 8 hr, 40 min, 2 sec



13



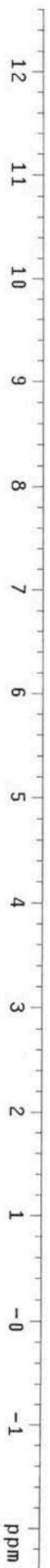
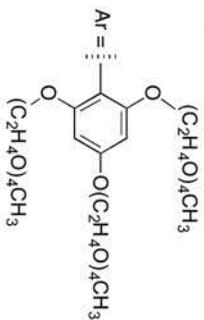
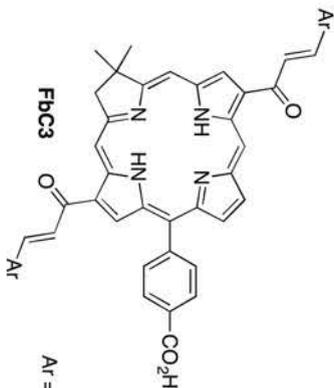


STANDARD 1H OBSERVE

Pulse Sequence: s2pu1

Solvent: CDCl3  
Ambient temperature  
File: 030015\_chlorin\_chalcone  
Mercury-400BB "ncsusercd400"

Relax. delay 1.000 sec  
Pulse 45.0 degrees  
Acq. time 1.995 sec  
Width 6006.0 Hz  
64 repetitions  
OBSERVE H1, 400.1245620 MHz  
DATA PROCESSING  
FT size 32768  
Total time 4 min, 6 sec



13C OBSERVE

Pulse Sequence: szpu1

Solvent: CDCl3

Ambient temperature

Mercury-400BB "ncsunc400"

Pulse 29.0 degrees

Acq. time 1.19 sec

Width 25000.0 Hz

50000 repetitions

OBSERVE C13, 100.6113937 MHz

DECOUPLE H1, 400.1266027 MHz

Power 40 dB

continuously on

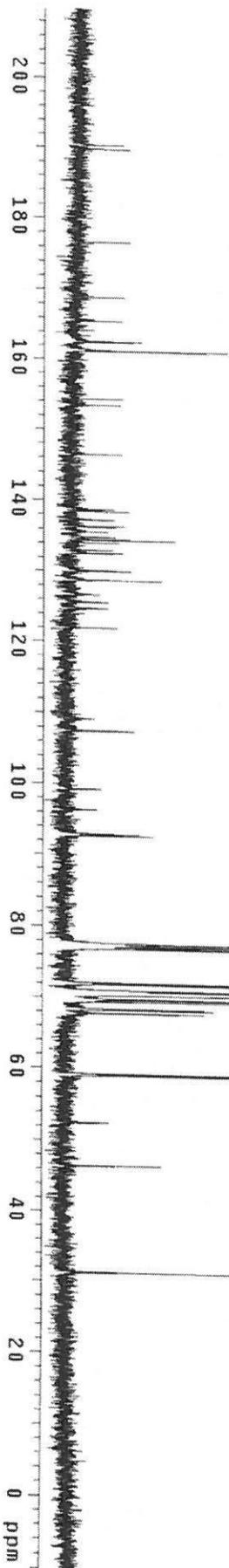
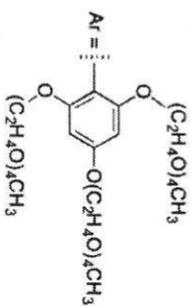
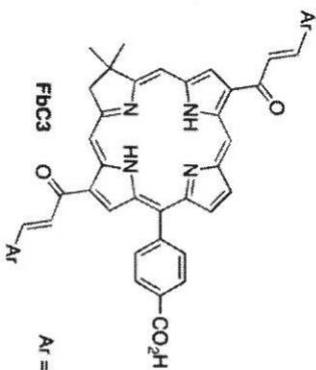
WALTZ-16 modulated

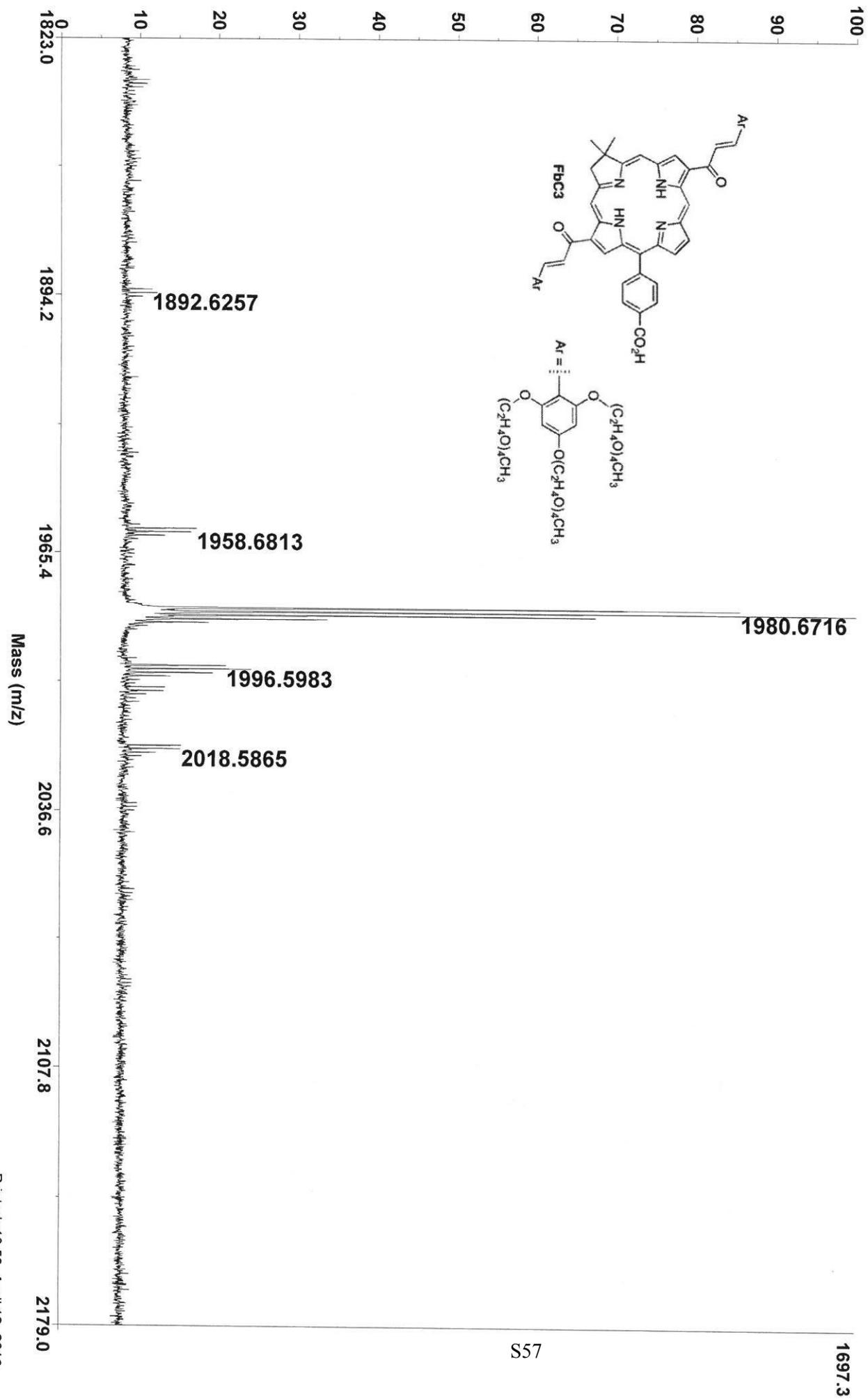
DATA PROCESSING

Line broadening 1.0 Hz

FI size 65536

Total time 43 hr, 20 min, 10 sec







STANDARD 1H OBSERVE

Pulse Sequence: szpu1

Solvent: CDCl3

Ambient temperature

Mercury-400BB "ncsuserc400"

Relax. delay 1.000 sec

Pulse 78.7 degrees

Acq. time 1.993 sec

Width 6006.0 Hz

256 repetitions

OBSERVE H1, 400.1245642 MHz

DATA PROCESSING

FT size 32768

Total time 13 min, 20 sec

