Supporting information (New Journal of Chemistry)

Fluorescent probe for Lewisite Simulant

Doo-Hee Lee, Dong-Nam Lee, and Jong-In Hong*

Department of Chemistry, College of Natural Sciences, Seoul National University, Seoul 08826, Republic of Korea

Contents

General

Chemical structures

Synthesis of probe

pH profiles of 1

Limit of detection

Comparison of HCI titration with AsCI₃ titration

¹H NMR titration

Photo-physical property of 2

Photo-physical property of 4

AsCl₃ quantification in soil

LCt₅₀ and LD₅₀ of blister agents

Selected ¹H, ¹³C NMR and mass spectra

General

Materials

7-hydroxycoumarin, acetic anhydride, 1,3,5,7-tetraazaadamantane, triphenylmethanethiol, bis(2-chloroethyl)amine, sodium cyanoborohydride, trifluoroacetic acid (TFA), triisopropylsilane (TIPS), dichloromethane (DCM), methanol (MeOH), cyclohexane, hexane, ethyl acetate, tetrahydrofuran (THF) and CDCl₃ were purchased as reagent grade from Aldrich, Acros, Samchun, TCI and used as received. The used metal salts are Hg(OAc)₂, Zn(ClO₄)₂, AgNO₃, Cd(ClO₄)₂, Cu(ClO₄)₂, Fe(ClO₄)₂, Pb(ClO₄)₂.

Instruments

NMR characterization: ¹H and ¹³C NMR spectra were recorded by Advance 300 and 75 MHz Bruker spectrometer in chloroform- d_3 . Chemical shifts were expressed in parts per million (δ) and reported as s (singlet), d (doublet), t (triplet) and m (multiplet).

Fluorescence & UV–Vis experiment: Probe **2** was dissolved in THF to afford a concentration of 10 mM stock solution, which was diluted to 10 μ M with distilled water up. Analyte was added into 10 μ M of **2** in the presence of TCEP (12 μ M), and photophysical property of **2** was measured in real time. Fluorescence and UV–Vis absorbance were recorded on Jasco FP-6500 and Beckman DU 800 spectrophotometer, respectively.

Chemical structures



Chart S1. Chemical structures of probes, BAL, lewisite, and lewisite simulant

Synthesis of probe



Scheme S1. Synthetic procedure of probes

8-((bis(2-(tritylthio)ethyl)amino)methyl)-7-hydroxy-2H-chromen-2-one (8): To a solution of compound **6**¹ (1.99 g, 10 mmol) in methanol/DCM (100 mL/50 mL) was added compound **7**² (6.22 g, 10 mmol), and then a small amount of acetic acid was further added. Sodium cyanoborohydride (0.63 g, 10 mmol) was added dropwise to the ice-cooled resulting solution under stirring. After the solution was stirred for three days at room temperature, it was acidified by adding conc. HCl and then evaporated almost to dryness under reduced pressure. The residue was dissolved in saturated Na₂CO₃ and extracted with DCM. The fractions were combined, dried over anhydrous Na₂SO₄, and evaporated under reduced pressure to give amber-colored oil. The residue was further purified on a silica-gel column with hexane and ethyl acetate to provide compound **8** in 42% yield. ¹H NMR (CDCl₃, 300 MHz) δ = 2.13 (t, 4H), 2.26 (t, 4H), 3.61 (s, 2H), 6.17 (d, 1H), 6.70 (d, 1H), 7.20 (m, 31H), 7.62 (d, 1H).

8-((1,2,5-dithiazepan-5-yl)methyl)-7-hydroxy-2H-chromen-2-one (2): Compound **8** (3.34 g, 4.2 mmol) was deprotected by treatment with DCM:TFA:TIPS (50:47.5:2.5, v/v/v, 400 mL) during 1 h. Deprotection solution was evaporated under reduced pressure and residual TFA was removed by co-evaporation with cyclohexane (3 × 100 mL) and dried *in vacuo*. The residue was dissolved in DCM and dried over anhydrous Na₂SO₄ and evaporated under reduced pressure to give amber-colored oil. The residue was air-oxidized in methanol/water with Na₂CO₃ prior to the purification. The resulting residue was further purified on a silica-gel column with hexane and DCM to give a white solid in 33% yield. ¹H NMR (CDCl₃, 300 MHz) δ = 2.95 (t, 4H), 3.30 (t, 4H), 4.35 (s, 2H), 6.20 (d, 1H), 6.81 (d, 1H), 7.31 (d, 1H), 7.62 (d, 1H). ¹³C NMR (CDCl₃, 75 MHz) δ = 162.7, 161.0, 153.0, 144.3, 128.4, 113.9, 111.7, 111.5, 108.3, 56.6, 53.3, 38.0. HRMS: calculated for C₁₄H₁₆O₃NS₂ [M+H⁺]⁺ 310.0572; found 310.0581.

pH profiles of 1



Fig. S1 Changes in the fluorescence intensity (at 445 nm) of **1** (10 μ M) (red square) upon addition of 10 equiv. of AsCl₃ (blue square) in the presence of TCEP (12 μ M) in water (pH 2.8–10.11). The fluorescence intensity was obtained by excitation at 370 nm.



Limit of detection

Fig. S2 Linear range of fluorescence quenching of 1 (10 μ M) upon addition of AsCl₃ (0-1.0 equiv.) in the presence of TCEP (12 μ M) in water. The fluorescence intensity was obtained by excitation at 370 nm.



Comparison of HCI titration with AsCI₃ titration

Fig. S3 Changes in the fluorescence intensity ratio (F/F_0) of **2** upon addition of increasing amount of HCI (blue square) and AsCl₃ (red square): F = the intensity of **2** (λ_{em} = 445 nm) upon addition of HCI or AsCl₃, F_0 = the initial intensity.

¹H NMR titration



Fig. S4 ¹H NMR (300 MHz, CDCl₃) spectra of 3 upon the addition of AsCl₃ (0-1.0 equiv.).



Photo-physical property of 2

Fig. S5 UV–Vis absorbance (a) and fluorescence spectra (b) of **2** (10 μ M) upon addition of AsCl₃ (0–10 equiv.) in water. The inset of panels shows absorption ($\lambda_{abs} = 360$ nm) (a) and emission ($\lambda_{em} = 445$ nm) (b) intensity of **2** (10 μ M) upon AsCl₃ addition (0–10 equiv.). The fluorescent emission was monitored by excitation at 370 nm. (c) Fluorescence intensities at 455 nm (*F*₀/*F*) upon titration of probe **2** (blue square) and probe **2** + TCEP (red square) with AsCl₃ (0–1 equiv.).

Photo-physical property of 4



Fig. S6 Fluorescence (a) and UV–Vis absorption spectra (b) of **4** (10 μ M) upon addition of AsCl₃ (0–10 equiv.) in water. The fluorescent emission was monitored by excitation at 340 nm.

AsCl₃ quantification in soil

Experimental method: 2.0 g of soil was sprayed with AsCl₃ (0–5µL) in THF and allowed to stand for 2 h. Then, the soil sample was centrifuged with water (10 mL), and quantification was conducted with probe **2** (10 µM).³

LCt₅₀ and LD₅₀ of blister agents

Blister agent	LCt₅₀ Inhalation mg⋅min/m³	LD₅₀ Skin mg/kg
Lewisite (L)	1400	30
Sulfur mustard (HD)	1500	100
Nitrogen mustard (HN-1)	1500	No data available

Table S1. Lethal concentration and time (LCt $_{50}$) and lethal dose (LD $_{50}$) of lewisite, sulfur/nitrogen mustard⁴



Selected ¹H, ¹³C NMR and mass spectra



Fig. S9 (a) Experimental HRMS (TOF, MeOH) spectrum and (b) theoretical isotope distribution of compound **2** (calculated mass: 310.0572, measured mass: 310.0578, Formula: $[M+H^+]^+$, $C_{14}H_{16}O_3NS_{2^+}$). The theoretical isotope distribution was determined from the software, Isotope Distribution Calculator and Mass Spec Plotter.



Fig. S11 ¹³C NMR Spectrum of 2



Fig. S13 (a) Experimental HRMS (ESI, MeOH) spectrum and (b) theoretical isotope distribution of compound 3 (calculated mass: 242.0673, measured mass: 242.0668, Formula: $[M+H^+]^+$, $C_{11}H_{16}ONS_{2^+}$).

Reference

- K. S. Lee, H. J. Kim, G. H. Kim, I. Shin and J. I. Hong, *Org. Lett.*, 2008, **10**, 49-51.
 N. Ollivier, J. Dheur, R. Mhidia, A. Blanpain and O. Melnyk, *Org. Lett.*, 2010, **12**, 5238-5241.
 V. Kumar and E. V. Anslyn, *Chem. Sci.*, 2013, **4**, 4292-4297.
- 4. S. L. Hoenig, Compendium of Chemical Warfare Agents, Springer, New York, NY, 2007, pp1-46.