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Synthesis and Evaluation of Benzothiazole-triazole and Benzothiadiazole-triazole Scaffolds as Potential Molecular **Probes for Amyloid-β Aggregation**

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Scheme S1. Synthesis of compound 4 and 5. Reagents and conditions: a) Br₂, 48% HBr, reflux, 1 h;

Synthesis of 4-bromobenzo[*c*][1,2,5]thiadiazole (4).¹ The title compound was synthesized as previously reported in the literature.² Br₂ (0.7 mL, 13.66 mmol) was added dropwise, during 20 min, to a stirred solution of 2,1,3-benzothiadiazole (4) (2.00 g, 14.76 mmol) and aqueous HBr (48%, 12 mL). The slurry was then refluxed setpoint: 120 °C) for 1 hour. After cooling, 200 mL of ice and 100 mL of saturated NaHSO₃ solution were poured into the reaction mixture. The slurry was filtered off and the solid was recrystallized from ethanol to give compound **5** as a beige powder (0.87 g, 27%). ¹H NMR (CDCl₃): 7.95-7.97 (dd, 1H), 7.82-7.84 (dd, 1H), 7.45-7.49 (dd, 1H).

Synthesis of 4,7-Dibromobenzo[*c*]-1,2,5-thiadiazole (5).²⁻³ The title compound was synthesized as previous reported in the literature.^{1,2} 2,1,3-Benzothiadiazole (4) (5.05 g, 36.91 mmol) was added to a stirred solution of aqueous HBr (48%, 30 mL). Br₂ (5.7 mL 110.73 mmol) was added slowly to the mixture and the reaction was refluxed for 2 hours at 130 °C. The final slurry was allowed to reach room temperature and was then poured into a solution of ice in saturated NaHSO₃. The resulting mixture was stirred for 10 minutes and the precipitation was filtered off. The crude product was recrystallized from water (35 mL, reflux for 30 min), filtered off, and washed with isopropanol (10 mL) to give a greenish/beige powder (5.28 g, 49%). ¹H NMR (400 MHz, CDCl₃) δ 7.70 (s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 153.1, 132.5, 114.0. HRMS (EI+) [M]⁺ Calcd for C₆H₂Br₂N₂S, 291.8305. Found, 291.8294.



Figure S1. Absorption (blue line) and emission spectra (red line) for L1 (a), L2 (b) and L3 (c) in 5% DMSO in phosphate buffered saline (PBS, 10 mM phosphate, 140 mM NaCl, 2.7 mM KCl, pH 7.4).



Figure S2. Emission spectra at 50 μ M (5% DMSO in PBS buffer) for L1 (a), L2 (b) and L3 (c) at different pH. pH 2 and 4: Glycine HCl buffer (50 mM); pH 6 and 8: phosphate buffer (25 mM); and pH 10 and 12: Glycine OH buffer (50 mM).



Figure S3. Concentration-absorbance curves for L1, L2 and L3 using known concentrations (20, 40, 60, 80, 100 and 250 μ M) in 5% DMSO in PBS buffer, pH 7.4. Linear curve fitting (y = mx + b) afforded R² = 0.9335, 0.9955 and 0.9985 for L1, L2 and L3, respectively.



Figure S4. The absorbance-integrated fluorescence intensity relationship measured in 5% DMSO/PBS buffer, pH 7.4; a) L1 (λ_{ex} = 310 nm), b) L2 ((λ_{ex} = 350 nm), c) L3 ((λ_{ex} = 400 nm).



Figure S5. Emission spectra for L1 (a), L2 (b) and L3 (c) at two different excitation wavelengths in 5% DMSO in phosphate buffered saline (PBS, 10 mM phosphate, 140 mM NaCl, 2.7 mM KCl, pH 7.4). Blue line represents the emission profile from excitation at a wavelength that is close to the absorption maxima for each compound, whereas the red line represents excitation at 405 nm, i.e. the excitation wavelength that were used in the fluorescent microscopy study.



Figure S6. ¹H-NMR (top) and ¹³C-NMR (bottom) spectra of L1 in DMSO-*d*₆.



Figure S7. ¹H-NMR (top) and ¹³C-NMR (bottom) spectra of L2 in CD₃OD.



Figure S8. ¹H-NMR (top) and ¹³C-NMR (bottom) spectra of L3 in CD₃OD and DMSO- d_6 , respectively.

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