

**Dual-responsive self-assembly of a bola-type supra-amphiphile  
constructed from a new pillar[6]arene-based recognition motif in  
water and its application in controlled release**

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**Electronic supplementary information (14 pages)**

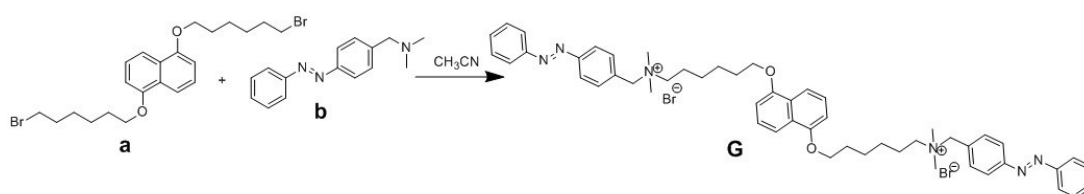
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## 1. Materials and methods

All reagents were commercially available and used as supplied without further purification. Compounds **WP6**,<sup>S1</sup> **a**,<sup>S2</sup> **b**,<sup>S3</sup> and **c**,<sup>S3</sup> were prepared according to published procedures. NMR spectra were recorded with a Bruker Avance DMX 500 spectrophotometer or a Bruker Avance DMX 400 spectrophotometer. Low-resolution electrospray ionization mass spectra were recorded with a Bruker Esquire 3000 Plus spectrometer. High-resolution mass spectrometry experiments were performed with a Bruker 7-Tesla FT-ICR mass spectrometer equipped with an electrospray source (Billerica, MA, USA). The melting points were collected on a SHPSIC WRS-2 automatic melting point apparatus. UV-vis spectra were taken on a Perkin-Elmer Lambda 35 UV-vis spectrophotometer. Isothermal titration calorimetric (ITC) measurements were performed on a VP-ITC micro-calorimeter (Microcal, USA). The determination of the critical aggregation concentration (CAC) values was carried out on a DDS-307 instrument. Transmission electron microscopy investigations were carried out on a JEM-1200EX instrument. Atomic force microscopy (AFM) experiments were performed on a Multi-Mode Nanoscope-IIIa Scanning Probe Microscope (Veeco Company, USA) in the tapping mode. Dynamic light scattering was carried out on a Malvern Nanosizer S instrument at room temperature.

## 2. Syntheses of **G** and **MG**

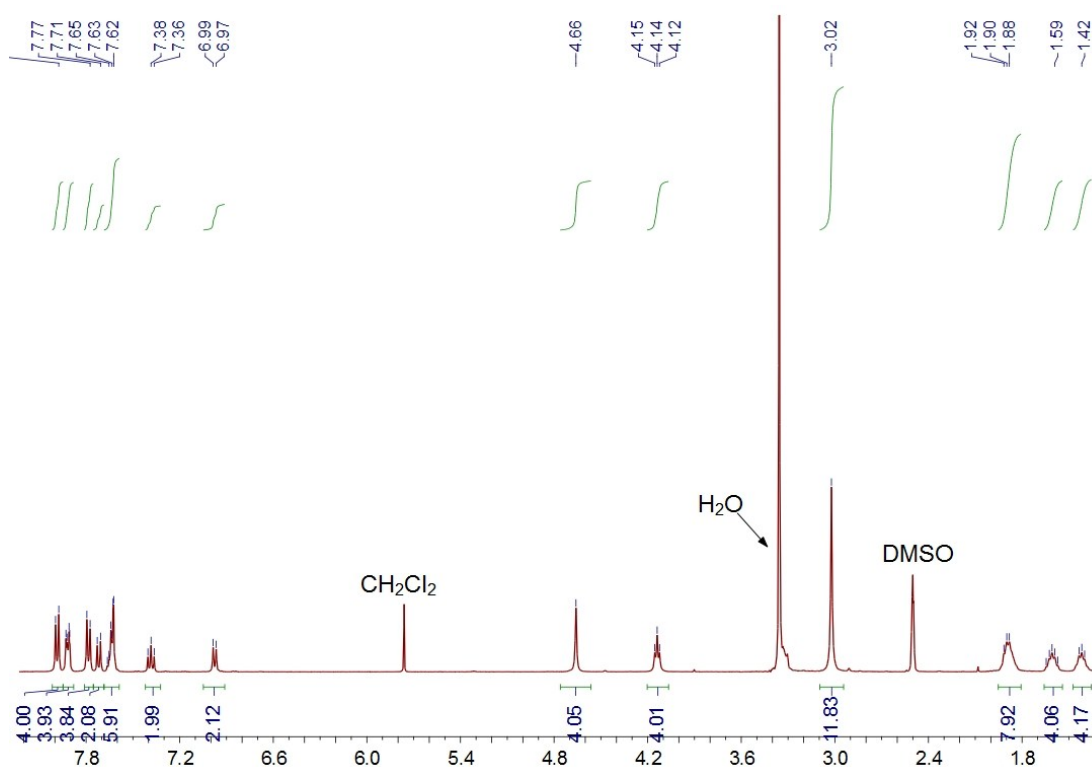
### 2.1. Synthesis of **G**



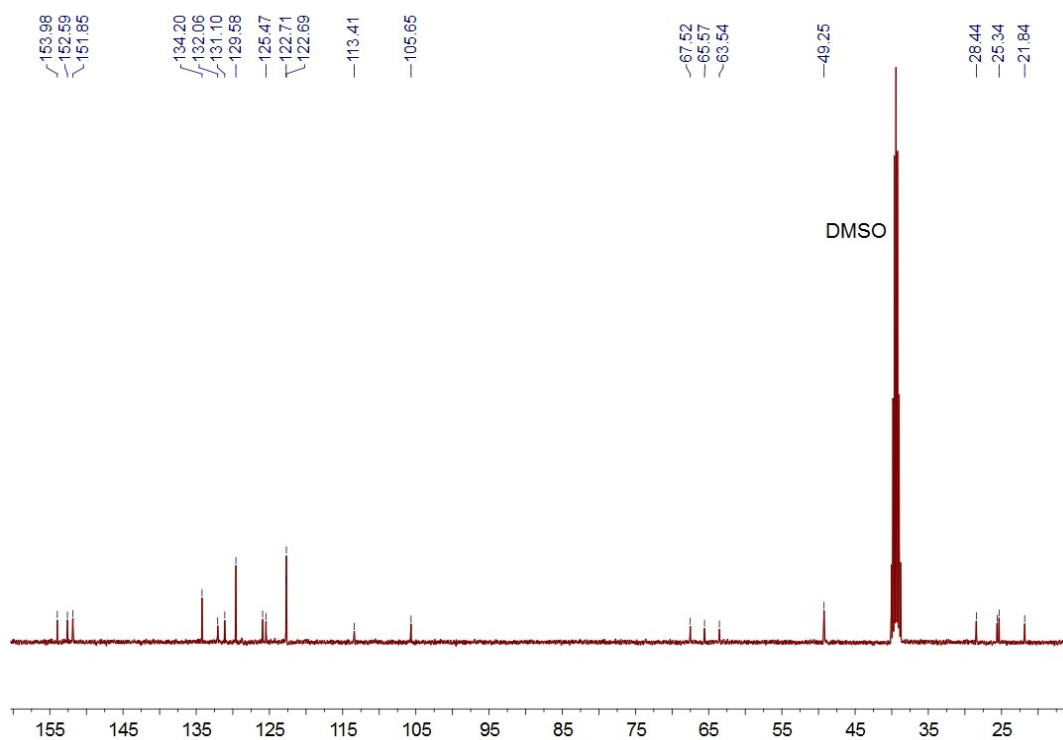
**Scheme S1** Synthetic route to **G**

A mixture of **a** (1.52 g, 3.12 mmol), **b** (2.00 g, 9.36 mmol) was stirred in acetonitrile at 82 °C under N<sub>2</sub> atmosphere for 24 h. The precipitated product **G** was collected by filtration, washed with acetonitrile and dried under vacuum to obtain **G** as an orange solid (2.17 g, 72%). Mp: 142.2–144.9 °C. The proton NMR spectrum of **G** is shown in Figure S1. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>, room temperature)  $\delta$  (ppm): 7.99 (d, *J* = 8.4 Hz, 4H), 7.92 (dd, 4H), 7.78 (d, *J* = 8.4 Hz, 4H), 7.72 (d, *J* = 8.4 Hz, 2H),

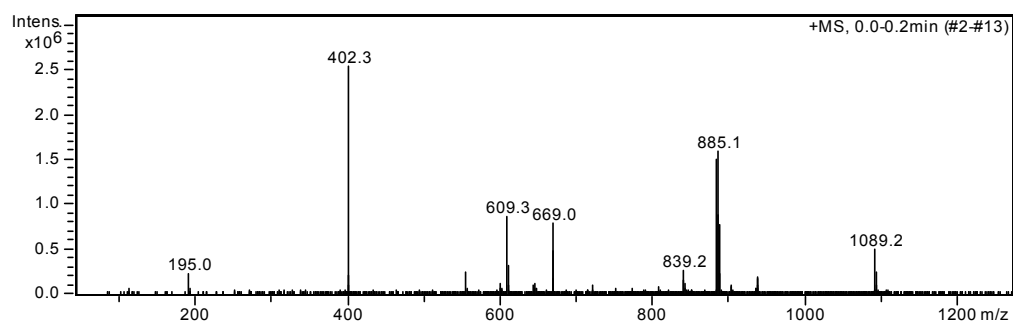
7.68–7.59 (m, 6H), 7.38 (t,  $J = 4.2$  Hz, 2H), 6.98 (d,  $J = 7.6$  Hz, 2H), 4.66 (s, 4H), 4.14 (t,  $J = 6.0$  Hz, 4H), 3.02 (s, 3H), 1.95–1.81 (m, 8H), 1.66–1.54 (m, 4H), 1.47–1.36 (m, 4H). The  $^{13}\text{C}$  NMR spectrum of **G** is shown in Figure S2.  $^{13}\text{C}$  NMR (100 MHz, DMSO- $d_6$ , room temperature)  $\delta$  (ppm): 153.98, 152.59, 151.85, 134.20, 132.06, 131.10, 129.58, 125.47, 122.71, 122.69, 113.41, 105.65, 67.52, 65.57, 63.54, 49.25, 28.44, 25.34, 21.84. LRESIMS is shown in Figure S3:  $m/z$  402.3  $[\text{M} - 2\text{Br}]^{2+}$  (100%). HRESIMS:  $m/z$  calcd for  $[\text{M} - 2\text{Br}]^{2+} \text{C}_{52}\text{H}_{64}\text{N}_6\text{O}_2$ , 402.2545, found 402.2544, error 0 ppm.



**Figure S1**  $^1\text{H}$  NMR spectrum (400 MHz, DMSO- $d_6$ , 298 K) of **G**.

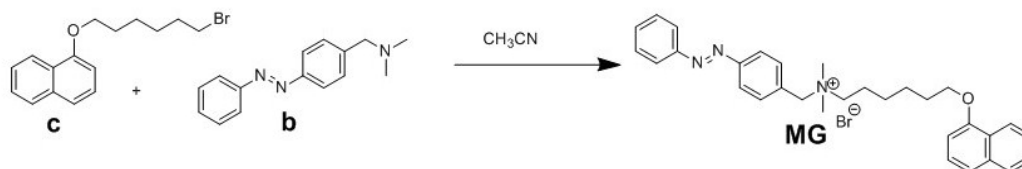


**Figure S2**  $^{13}\text{C}$  NMR spectrum (100 MHz,  $\text{DMSO-}d_6$ , 298 K) of **G**.



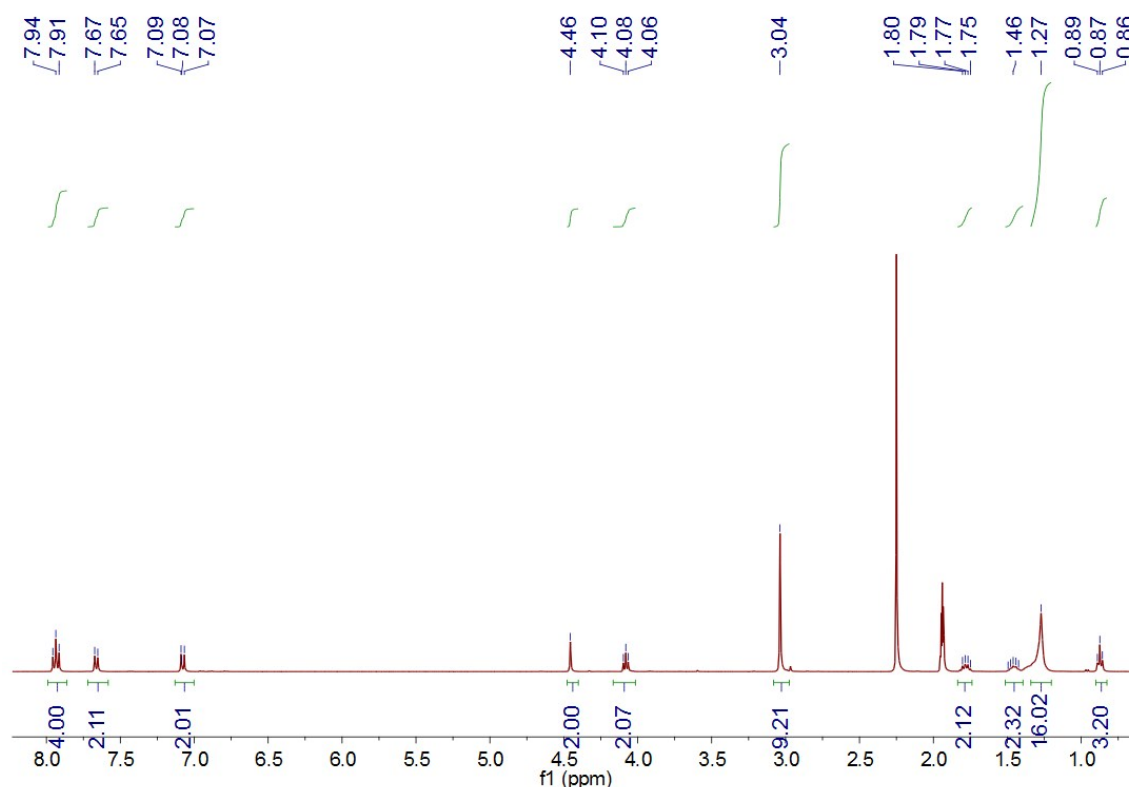
**Figure S3** Electrospray ionization mass spectrum of **G**. Main peak:  $m/z$  402.3 [ $\text{M} - 2\text{Br}$ ] $^{2+}$  (100%).

## 2.2. Synthesis of **MG**

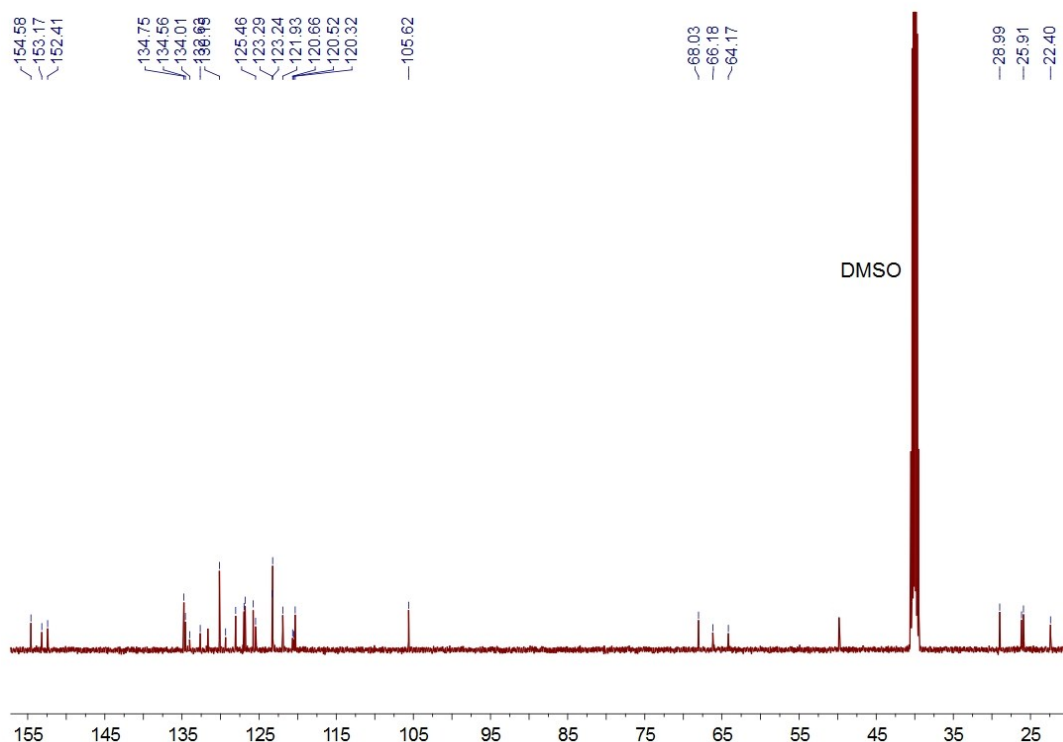


**Scheme S2** Synthetic route to **MG**.

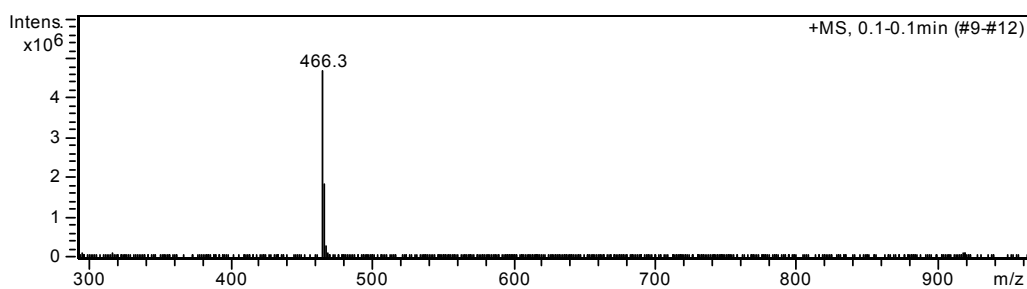
A mixture of **c** (1.25 g, 3.28 mmol), **b** (0.590 g, 3.28 mmol) was stirred in acetonitrile at 82 °C under N<sub>2</sub> atmosphere for 24 h. The precipitated product **MG** was collected by filtration, washed with acetonitrile and dried under vacuum to obtain **MG** as an orange solid (2.17 g, 72%). Mp: 175.3–177.1 °C. The <sup>1</sup>H NMR spectrum of compound **MG** is shown in Figure S3. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>, 298 K)  $\delta$  (ppm): 8.16 (dd, 1H), 7.99 (d, *J* = 8.4 Hz, 2H), 7.95–7.90 (m, 2H), 7.87 (dd, 1H), 7.79 (d, *J* = 8.4 Hz, 2H), 7.68–7.60 (m, 3 H), 7.55–7.48 (m, 2H), 7.48–7.39 (m, 2H), 6.97 (d, *J* = 6.8 Hz, 1H), 4.65 (s, 2H), 4.18 (t, *J* = 6.2 Hz, 2H), 3.02 (s, 6H), 1.99–1.79 (m, 4 H), 1.63–1.59 (m, 2 H), 1.52–1.35 (m, 2 H). The <sup>13</sup>C NMR spectrum of **MG** is shown in Figure S4. <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>, 298 K)  $\delta$  (ppm): 154.58, 153.17, 152.41, 134.75, 134.56, 134.01, 132.63, 130.15, 129.34, 128.04, 126.95, 126.81, 125.77, 125.46, 123.29, 123.24, 121.93, 120.66, 120.52, 120.32, 105.62, 68.03, 66.18, 64.17, 28.99, 26.16, 25.91, 22.40. LRESIMS is shown in Figure S5: *m/z* 466.3 [M – Br]<sup>+</sup> (100%). HRESIMS: *m/z* calcd for [M – Br]<sup>+</sup> C<sub>31</sub>H<sub>36</sub>N<sub>3</sub>O, 466.2858, found 466.2853, error –1 ppm.



**Figure S4** <sup>1</sup>H NMR spectrum (400 MHz, DMSO-*d*<sub>6</sub>, 298 K) of **MG**.



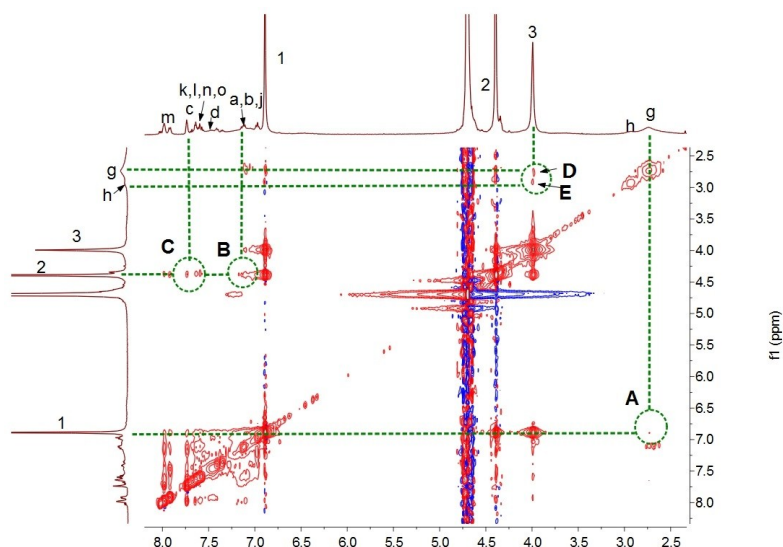
**Figure S5**  $^{13}\text{C}$  NMR spectrum (100 MHz,  $\text{DMSO-}d_6$ , 298 K) of **MG**.



**Figure S6** Electrospray ionization mass spectrum of **MG**. Main peak:  $m/z$  466.3  $[\text{M} - \text{Br}]^+$  (100%).

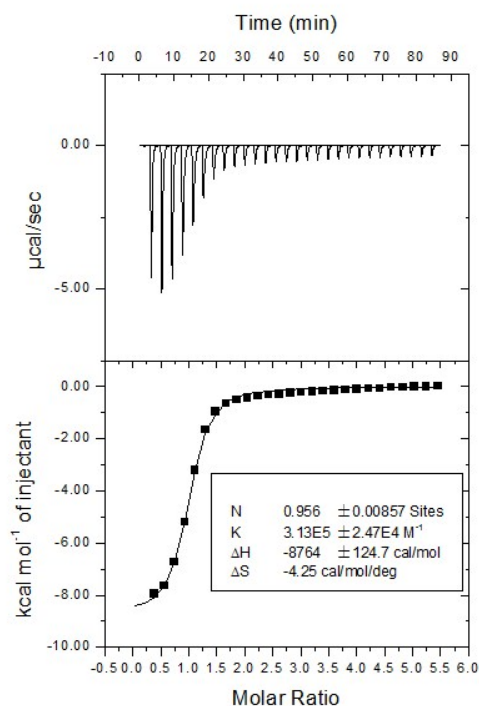
### 3. Partial 2 D NOESY spectra of an equimolar solution of *trans*-**MG** and **WP6**

2 D NOESY NMR experiment was employed to study the relative positions of the components in complex **WP6**⊃*trans*-**MG**. NOE correlation signals were observed between protons  $\text{H}_g$  of *trans*-**MG** and proton  $\text{H}_1$  of **WP6** (Figure S7, A), between protons  $\text{H}_a$ ,  $\text{H}_b$ , and  $\text{H}_c$  on the azobenzene unit of *trans*-**MG** and proton  $\text{H}_2$  of **WP6** (Figure S7, B and C), and between protons  $\text{H}_g$  and  $\text{H}_h$  of *trans*-**MG** and proton  $\text{H}_3$  of **WP6**.



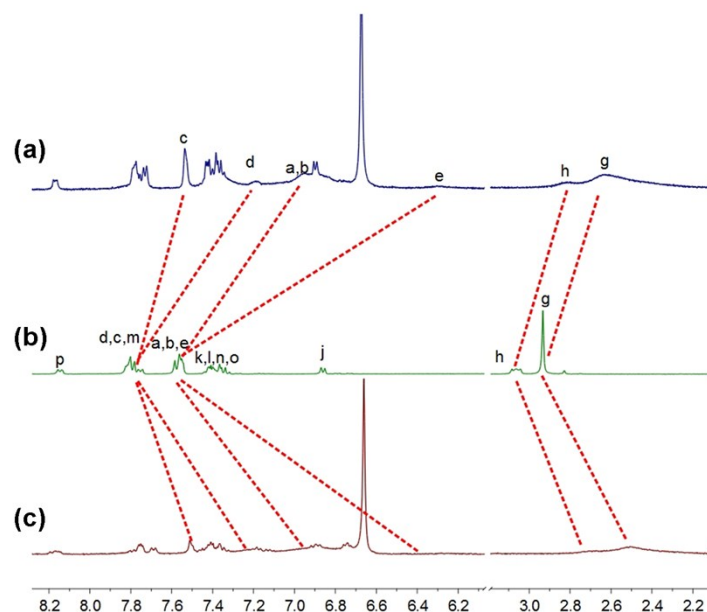
**Figure S7** Partial 2D NOESY spectra (500 MHz, 3:1 D<sub>2</sub>O/CD<sub>3</sub>CN, room temperature) of an equimolar mixture *trans*-MG and WP6 (2.50 mM).

#### 4. Isothermal titration calorimetry (ITC) experiment of WP6 and *trans*-G



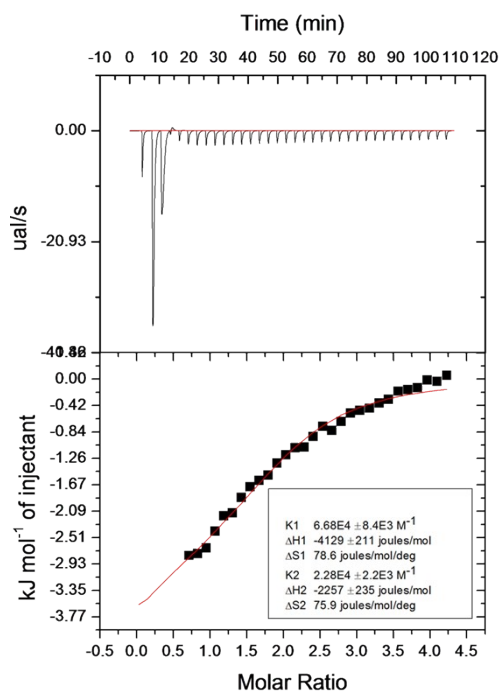
**Figure S8** Microcalorimetric titration of WP6 with *trans*-MG in water at 298.15 K. (Top) Raw ITC data for 29 sequential injections (10  $\mu$ L per injection) of a WP6 solution (2.00 mM) into a *trans*-MG solution (0.100 mM). (Bottom) Net reaction heat obtained from the integration of the calorimetric traces.

### 5. pH-Responsive complexation of WP6 $\rightarrow$ trans-MG



**Figure S9**  $^1\text{H}$  NMR spectra (500 MHz, 5:1  $\text{D}_2\text{O}/\text{CD}_3\text{CN}$ , room temperature): (a) WP6 $\rightarrow$ trans-MG (2.50 mM) when the solution pH was 7.0; (b) WP6 $\rightarrow$ trans-MG (2.50 mM) when the solution pH decreased to 5.0; (c) WP6 $\rightarrow$ trans-MG (2.50 mM) when the solution pH recovered to 7.0.

### 6. Isothermal titration calorimetry (ITC) experiment of WP6 and trans-G

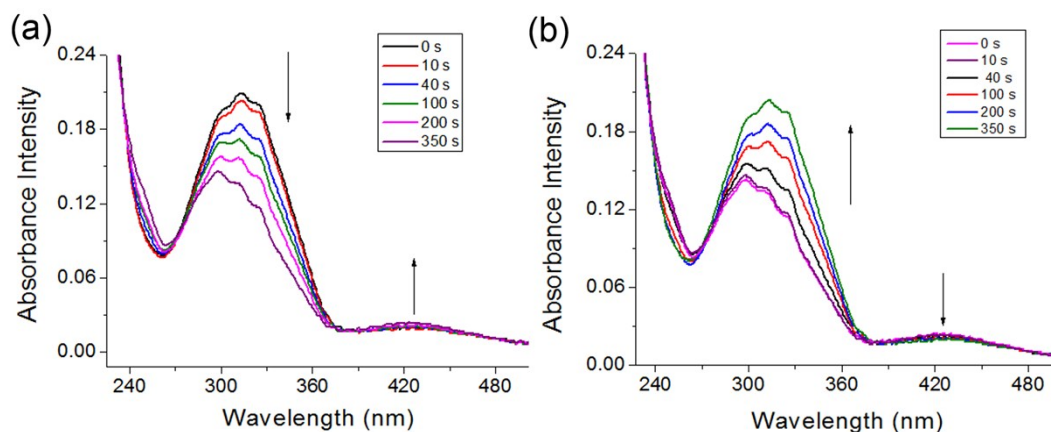


**Figure S10** Microcalorimetric titration of WP6 with trans-G in water at 298.15 K. (Top)

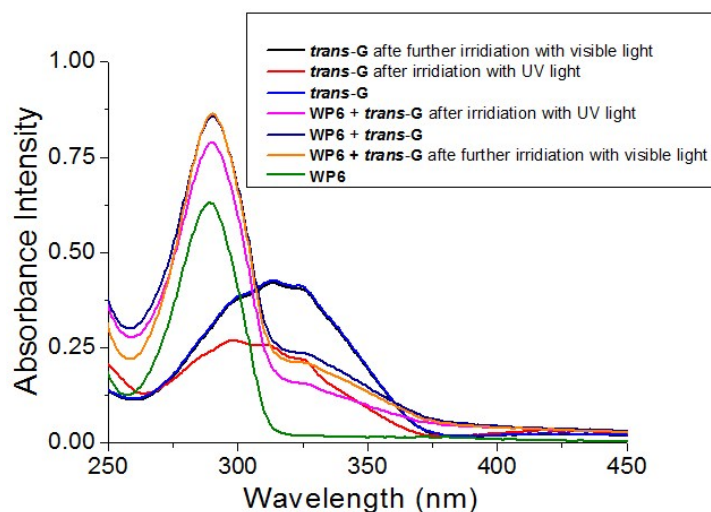


Raw ITC data for 29 sequential injections (10  $\mu\text{L}$  per injection) of a **WP6** solution (2.00 mM) into a *trans*-**G** solution (0.100 mM). (Bottom) Net reaction heat obtained from the integration of the calorimetric traces.

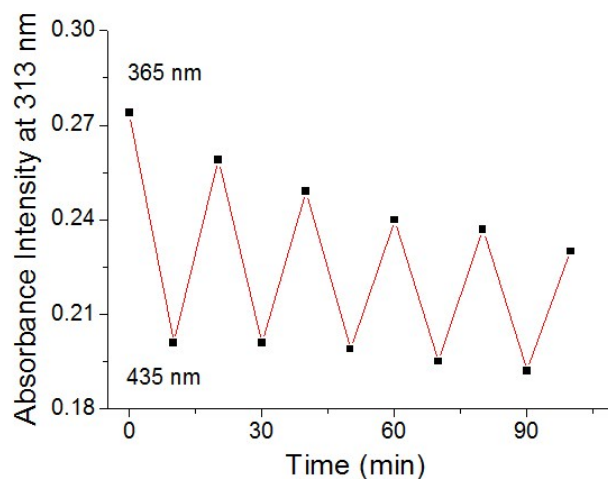
### 7. UV-vis absorption spectroscopy experiments



**Figure S11** UV-vis absorption spectra of *trans*-**G** in water under UV irradiation at 365 nm from 0 to 350 s (a) and upon visible irradiation at 435 nm from 0 to 350 s (b). The concentration of *trans*-**G** was  $2.50 \times 10^{-5}$  M.



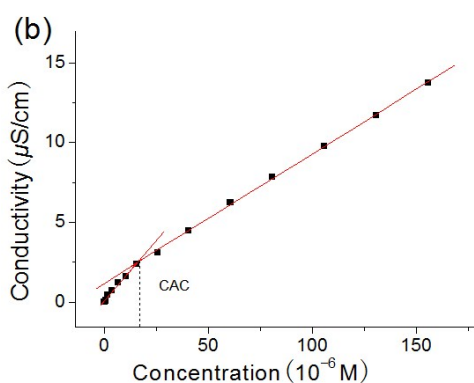
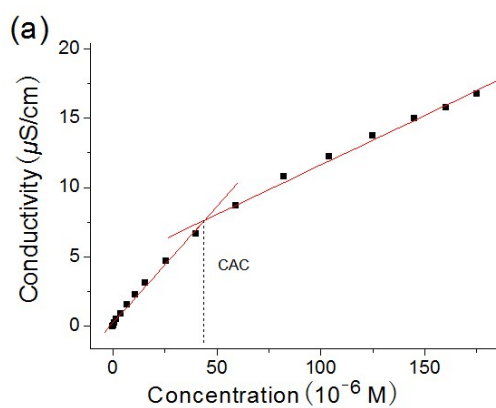
**Figure S12** (a) UV-vis spectra of  $2.50 \times 10^{-5}$  M *trans*-**G** (initial and after irradiation with UV light at 365 nm for 10 min) and solution of  $2.50 \times 10^{-5}$  M *trans*-**G** and  $5.00 \times 10^{-5}$  M **WP6** (initial, after irradiation with UV light at 365 nm for 10 min, and then after irradiation with visible light at 435 nm for 10 min)



**Figure S13** Changes of the absorbance at 313 nm of  $(\text{WP6})_2\supset\text{G}$  upon alternating irradiation with UV and visible light for 10 min.

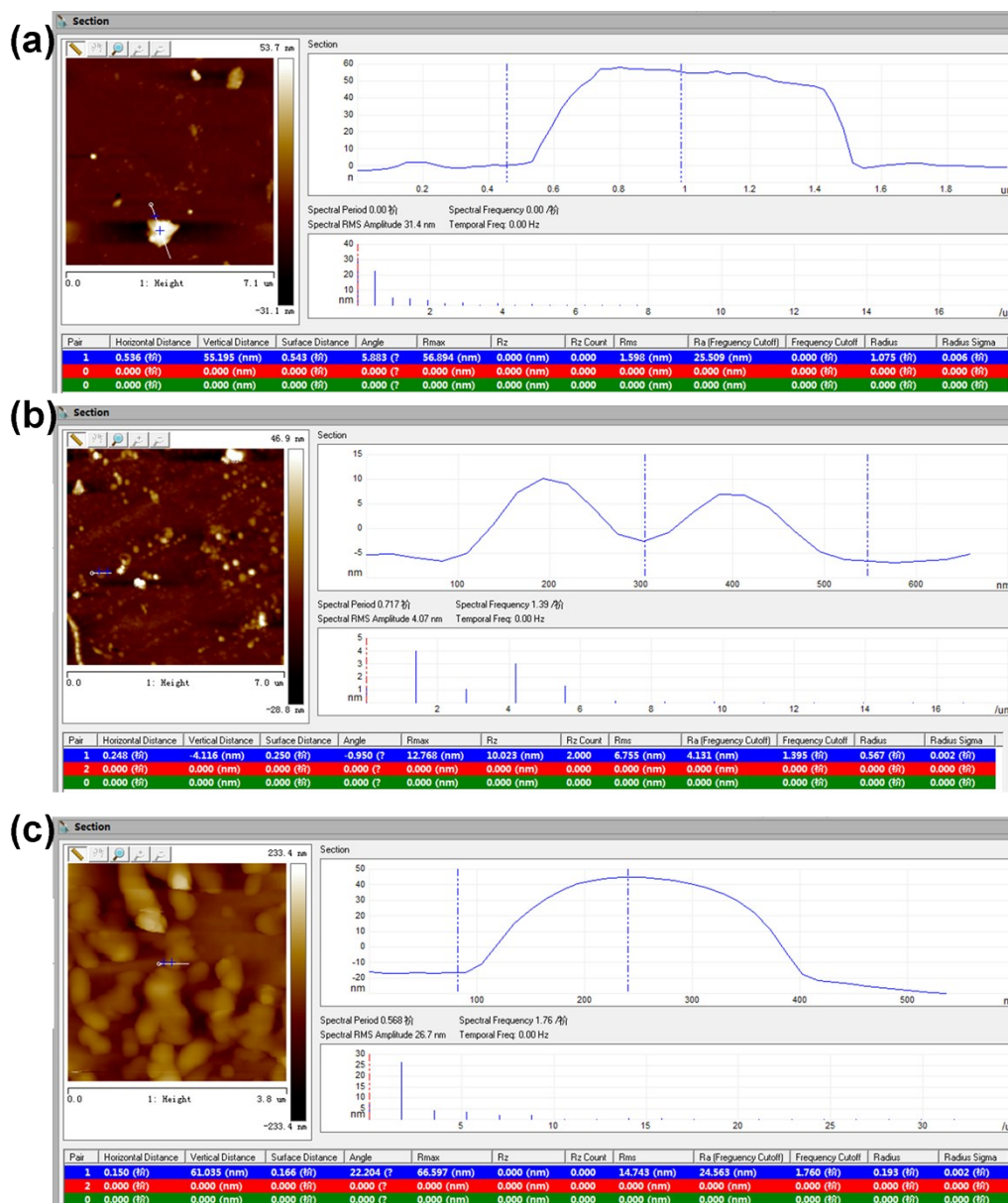
7. Critical aggregation concentration (CAC) determination of *trans-G* and the equimolar mixture of *trans-G* and *WP6*

To measure the CAC values of *trans-G* and the equimolar mixture of *trans-G* and *WP6*, the conductivities of the solutions at different concentrations of *trans-G* and the equimolar mixture of *trans-G* and *WP6* were determined. By plotting the conductivity versus the concentration, we estimated the CAC values of *trans-G* and the mixture of *trans-G* and *WP6* (1:2 in ratio).



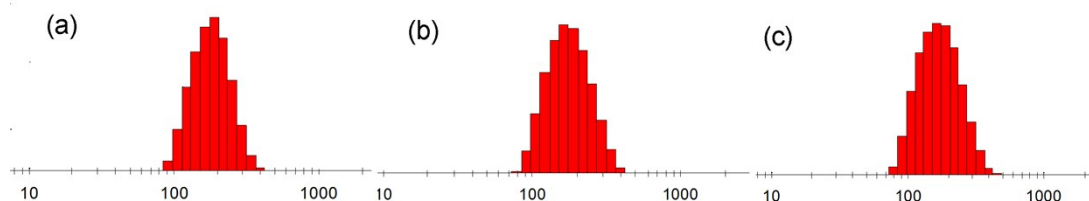
**Figure S14** (a) Concentration-dependent conductivity of *trans*-G. The critical aggregation concentration (CAC) was determined to be  $4.40 \times 10^{-5}$  M; (b) Concentration-dependent conductivity of the mixture of *trans*-G and WP6 (1:2 in ratio). The critical aggregation concentration (CAC) was determined to be  $1.69 \times 10^{-5}$  M of (WP6)<sub>2</sub>⊃G.

### 8. AFM results



**Figure S15** (a) AFM results of *trans*-G ( $2.50 \times 10^{-5}$  M) aggregates in water; (b) AFM results of WP6 ( $5.00 \times 10^{-5}$  M) and *trans*-G ( $2.50 \times 10^{-5}$  M) for both in water; (c) AFM results of the aggregates after irradiation with UV light at 365 nm of (b).

### 9. Dynamic light scattering (DLS) experiments



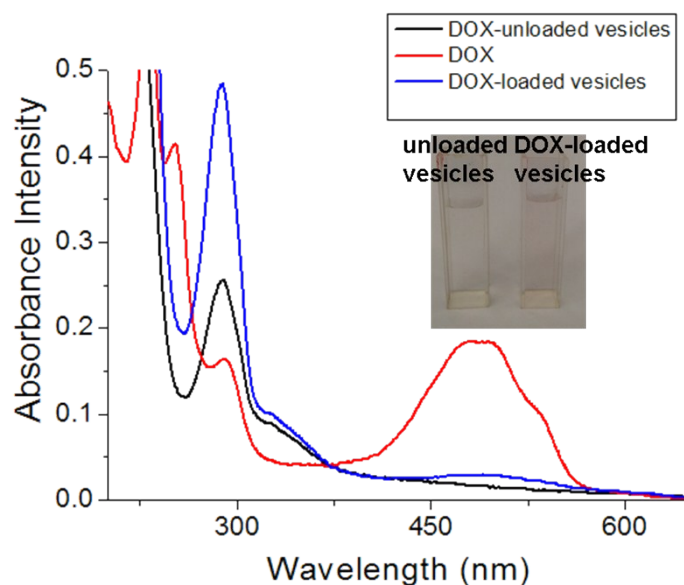
**Figure S16** (a) DLS result of **WP6** with **trans-G** ( $2.50 \times 10^{-4}$  M for both) in water; (b) DLS result of **WP6** with **trans-G** ( $2.50 \times 10^{-4}$  M for both) in water after further irradiating with visible light at 435 nm; (c) DLS result of **WP6** with **trans-G** ( $2.50 \times 10^{-4}$  M for both) in water after adding HCl solution and then adding NaOH solution.

#### 10. DOX·HCl encapsulation and controllable DOX·HCl release experiments

DOX·HCl loading experiment: DOX-loaded vesicles were prepared by adding a certain amount of DOX·HCl into a freshly prepared aqueous solution of **WP6** ( $5.00 \times 10^{-5}$  M) and **trans-G** ( $2.5 \times 10^{-5}$  M). The ultimate concentrations of DOX·HCl, **WP6**, and **trans-G** were 0.0500, 0.0500, and 0.02500 mM, respectively. And then the prepared DOX-loaded vesicles were purified by dialysis (molecular weight cutoff = 3500) in distilled water for several times until the water outside the dialysis tube exhibited negligible DOX·HCl fluorescence. As a result, DOX·HCl was successfully loaded into the vesicles constructed from **WP6**-**trans-G**-based bola-type supra-amphiphiles. The DOX·HCl encapsulation and loading efficiency were calculated by the following equations:<sup>S3</sup>

$$\text{Encapsulation Efficiency (\%)} = (m_{\text{DOX-loaded}}/m_{\text{DOX}})100$$

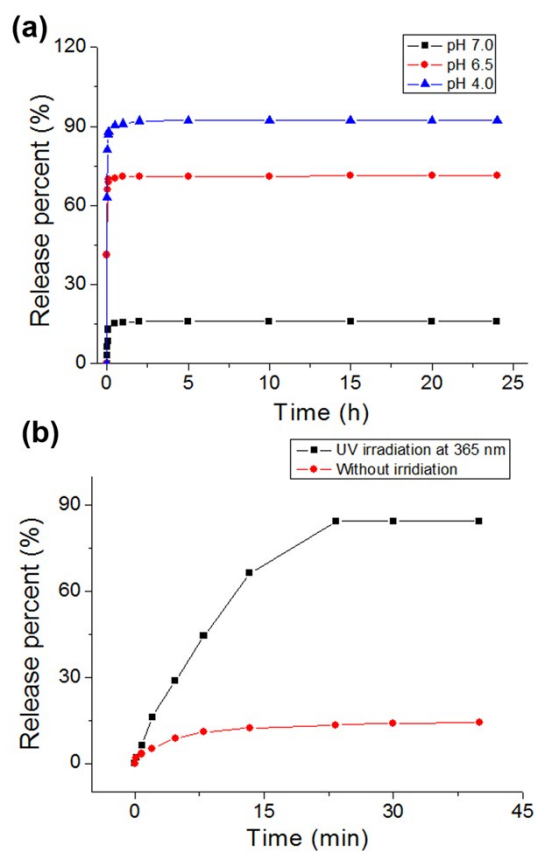
$m_{\text{DOX-loaded}}$  and  $m_{\text{DOX}}$  are mass of DOX·HCl encapsulated in vesicles and mass of DOX·HCl added, respectively. The mass of DOX·HCl was measured by a UV spectrophotometer at 490 nm and calculated as relative to a standard calibration curve in the concentrations from  $2.00 \times 10^{-6}$  to  $4.00 \times 10^{-5}$  M in water.



**Figure S17** UV-vis absorption spectra of DOX-loaded vesicles, DOX·HCl, unloaded vesicles in water. Inset: color change of DOX-loaded vesicle (right) compared with unloaded one (left).

Controllable DOX·HCl release by pH changes: 0.05 M tris-HCl (pH = 7.4), 0.2 M sodium acetate (pH = 4.0), and 0.1 M citrate (pH = 6.5) buffer solutions were used as the release media. In a typical release experiment, 1.60 mL of DOX-loaded vesicles was added into 8.40 mL of appropriate release medium at 37 °C. At selected time intervals, 3 mL of the release media was taken out for measuring the released DOX·HCl concentrations by the UV-vis absorption technique and fluorescence emission spectrometer and then was returned to the original release media. The concentration of DOX·HCl was determined by measurement of absorbance at 490 nm using a standard absorbance versus concentration curve constructed for DOX·HCl in the corresponding release buffer. By presenting the vesicles to very low pH (the solution of HCl, pH = 2.0), a nearly 100% release of DOX·HCl from DOX-loaded vesicles could be obtained.

Controllable DOX·HCl release by photo irradiation: 0.05 M tris-HCl (pH = 7.4) buffer solutions were used as DOX·HCl release media. 1.60 mL of DOX-loaded vesicles was added into 8.40 mL of release medium at 37 °C. Upon irradiation with UV light at 365 nm, 3 mL of the release media at selected time intervals was taken out for measuring the released DOX·HCl concentrations by the UV-vis absorption technique and fluorescence emission spectrometer and then was returned to the original release media. The concentration of DOX·HCl was determined by measurement of absorbance at 490 nm using a standard absorbance versus concentration curve constructed for DOX·HCl in 0.05 M tris-HCl (pH = 7.4) buffer solutions.



**Figure S18** (a) Release percentage of the DOX-loaded vesicles in the release media of different pH values by fluorescence emission spectroscopy; (b) Release percentage of the DOX-loaded vesicles under the irradiation with UV light at 365 nm from 0 to 40 min by fluorescence emission spectroscopy.

*References:*

- S1. D. Xia, G. Yu, J. Li and F. Huang, *Chem. Commun.*, 2014, **50**, 3606–3608.
- S2. S. De and S. Ramakrishnan, *Chem. Asian J.*, 2011, **6**, 149–156.
- S3. L. Peng, M. You, C. Wu, D. Han, I. Öçsoy, T. Chen, Z. Chen and W. Tan, *ACS Nano*, 2014, **8**, 2555–2561.
- S4. S. Chambert, F. Thomasson and J.-L. Décout, *J. Org. Chem.*, 2002, **67**, 1898–1904.