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Supplementary Information for

Ratiometric captopril assay based on the recovery of the Bi (III)-quenched vellow fluorescence of dually emitting carbon nanodots

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Materials

Ascorbic acid (AA), ethylene glycol, CH₃COOH, CH₃COONa, AgNO₃, NaCl, KCl, CoCl₂, Sr(NO₃)₂, NiCl₂, BaCl₂, MgSO₄, MnSO₄, Pb(NO₃)₂, ZnCl₂, CuCl₂, Hg(NO₃)₂, Cr(NO₃)₃, Bi(NO₃)₃, CAP, glucose, methionine, cysteine, norvaline, threonine, histidine, and arginine were purchased from Aladdin (Shanghai, China). DNA oligonucleotides used in this work were synthesized by Sangon Biotechnology Co., Ltd. (Shanghai, China). CAP tablets and human blood were purchased from a local pharmacy. The All of the chemicals were used as received without further purication. Ultrapure water was used throughout all of the experiments.

Apparatus

A Hitachi F-7000 fluorescence spectrophotometer (Tokyo, Japan) was used for recording fluorescence spectra with the slits (Ex/Em) of 10.0/10.0 nm. The photomultiplier tube (PMT) voltage was set at 400 V.

The calculation of limit of detection (LOD) of CAP

All experiments were repeated at least 3 times, and the standard deviation (SD) of experimental data was controlled within 0.25%. The limit of detection (LOD) was defined as the concentration of analyte that corresponds to three times the signal-to-noise ratio (S/N=3). Here, LOD was calculated according to the expression $LOD=3\sigma/K$, where σ was the standard deviation for the blank solution (n=10), and K was the slope of the calibration curve. In this paper, 3σ was about 0.0043; the corresponding linear regression equation was $I_{530}/I_{410}=0.354C+0.366$, and C was the concentration of p-NP, so the LOD= 0.0043/ 0.354=12 nM.



Fig. S1 Structure of captopril.



Fig. S2 Fluorescence spectra of blue emitters (a) and yellow emitters (b). The photographs of de-CDs under visible and UV light at 365 nm (c).



Fig. S3 Fluorescence responses to different concentrations of Bi^{3+} (a) and corresponding linear range (b). The concentrations of Bi^{3+} were 0.05, 0.1, 0.5, 1, 2, 3, 4, 5, 6, 7 μ M, respectively.



Fig. S4 Effect of probe concentration on the sensing assay for CAP. The concentrations of Bi^{3+} and CAP were 7 and 10 μ M, respectively.



Fig. S5 Influence of reaction time on the detection of CAP. The concentrations of de-CDs, Bi^{3+} and CAP were 0.054 mg mL⁻¹, 7 μ M and 10 μ M, respectively.



Fig. S6 Effect of reaction temperature on the determination of CAP. The concentrations of de-CDs, Bi^{3+} and CAP were 0.054 mg mL⁻¹, 7 μ M and 10 μ M, respectively.



Fig. S7 Effect of pH on the determination of CAP. The different pH values (pH 3.6, 4.0, 4.4 4.8, 5.2, 5.6) were adjusted by adding 0.2 M CH₃COOH-CH₃COONa. The concentrations of de-CDs, Bi^{3+} and CAP were 0.054 mg mL⁻¹, 7 μ M and 10 μ M, respectively.



Fig. S8 FT-IR spectra of CAP (red line), Bi³⁺ (blue line), and CAP in the presence of Bi³⁺ (black line).



Fig. S9 (a) Fluorescence spectra of de-CDs-Bi³⁺ with the addition of different concentrations of CAP (from bottom to top: 0, 0.1, 3, 7, 9, 11, 13, 15 μ M) in the human plasma and the corresponding linear range (b).

Number	Labeled amount	Found amount	Recovery
	(µM)	(µM)	(n = 3, %)
1	5	5.03	100.6
2	5	4.85	97.0
3	5	4.79	95.8

Table S1. Determination of CAP in tablets.

Samples	Found amount	Standard added	Found value	Recovery
	(µM)	(µM)	(µM)	(n = 3, %)
plasma	NF	0.2	0.195	97.5
	NF	3	2.90	96.7
	NF	10	10.05	100.5
urine	NF	0.1	0.095	95.0
	NF	3	3.12	104.0
	NF	10	10.21	102.1

Table S2. Determination of CAP in human plasma and urine.

NF: not found

References

 D. Y. Liu, F. Qu, X. E. Zhao and J. M. You, J. Phys. Chem. C, 2015, 119, 17979-17987.