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1	Electronic supplementary information:
2	Substituted bidentate and ancillary ligands modulate bioimaging
3	properties of classical Re (I) tricarbonyl core with yeasts and bacteria
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5	Alexander Carreño <sup>1,2</sup> *, Alejandra E. Aros <sup>2,3</sup> , Carolina Otero <sup>4</sup> , Rubén Polanco <sup>5</sup> , Manuel
6	Gacitúa <sup>6</sup> , Ramiro Arratia-Pérez <sup>1,2</sup> , Juan A. Fuentes <sup>3**</sup>
7	
8	1. Center of Applied Nanosciences (CENAP), Universidad Andres Bello, República 275,
9	Santiago, Chile.
10	2. Núcleo Milenio de Ingeniería Molecular para Catálisis y Biosensores (MECB), ICM,
11	Chile.
12	3. Laboratorio de Genética y Patogénesis Bacteriana, Facultad de Ciencias Biológicas,
13	Universidad Andrés Bello, República 217, Santiago, Chile.
14	4. Center for Integrative Medicine and Innovative Science (CIMIS), Facultad de Medicina,
15	Universidad Andres Bello, Echaurren 183, Santiago, Chile.
16	5. Laboratorio de Bioquímica, Facultad de Ciencias Biológicas, Universidad Andres Bello,
17	República 217, Santiago, Chile.
18	6. Center of Applied Ecology and Sustainability (CAPES), Universidad Adolfo Ibáñez,
19	Peñalolén, Chile.
20	† Electronic supplementary information (ESI) available.
21	*Corresponding author for chemical experiments: acarreno@puc.cl
22	**Corresponding author for biological experiments: jfuentes@unab.cl
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26 Scheme S1. Alternative route to obtained classical rhenium complexes (C1 to C5).







52 Figure S3. FTIR spectrum for (*E*)-2-{[(3-aminopyridin-4-yl)imino]-methyl}-4,6-di-tert-

53	B butyl-phenol (L).		
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## 186 S.3. Biological methods and Supplementary Results

S.3.1. Minimum inhibitory concentration (MIC): All the compounds were evaluated for 187 their in vitro growth inhibitory activity against Gram-negative pathogen Salmonella enterica 188 189 serovar Typhimurium ATCC14028s (S. enterica), and the clinical yeasts Candida albicans and Cryptococcus spp. obtained from the Hospital Clínico of the Universidad de Chile, 190 Santiago, Chile. Minimum inhibitory concentration (MIC) was obtained by broth dilution. 191 192 The MIC is defined as the lowest concentration of the tested compounds at which no growth of the strain was observed after incubation. S. enterica was previously grown in Luria-Bertani 193 broth (Bacto peptone, 10 g/l; Bacto yeast extract, 5 g/l; NaCl, 5 g/l) at 37 °C with shaking to 194  $OD_{600} = 1.4$  (stationary phase). Yeasts were previously cultured in Sabouraud agar (Bacto 195 peptone, 10 g/L; glucose, 40 g/L; agar, 15 g/L; pH 5.6) at 30 °C. Further dilutions of 196 microorganisms (bacteria and yeasts; 0.5 McFarland) were performed with Bacto Tryptic 197 Soy broth (pancreatic digest casein 17.0 g/l, papaic digest of soybean 3.0 g/l, dextrose 2.5 198 g/l, sodium chloride 5.0 g/l, dipotassium phosphate 2.5 g/l). Stock solutions of the tested 199 200 compounds were prepared in dimethyl sulfoxide (DMSO) for C1, C2, C3, C4, C5, C6, and K1 (ketoconazole, control for yeasts); or in ethanol 95% for Cam (chloramphenicol, control 201 for bacteria). The concentration range of the compounds tested was between  $1.042 \,\mu g/ml$  and 202 203 133.33  $\mu$ g/ml. The inoculated wells were then incubated at 37 °C for 24 h and 48 h (bacteria), or at 30 °C for 48 h and 72 h (yeasts). As control, DMSO or ethanol 95% without any tested 204 compound, were included when necessary. Only when the inhibition effects of the tested 205 compounds were distinguishable from DMSO alone (in the case of C1, C2, C3, C4, C5, C6, 206 and K1), or from ethanol 95% alone (in the case of Cam), the compound was considered to 207 exhibit antimicrobial activity. The MIC values of the tested compounds were obtained as 208 µg/ml. All the experiments were performed in triplicate. Results are shown in Table S1. 209

S.3.2. MTT assay performed in HeLa cells: Cells were cultured in Dulbecco's Modified 210 Eagle's Medium (DMEM) containing 10% fetal bovine serum (FBS), 2 mM L-glutamine, 211 100 units/mL penicillin and 100 µg/mL streptomycin. Cells were maintained in 75 cm<sup>2</sup> flasks 212 in a 5% CO2-humidified atmosphere at 37 °C. Passage takes place every 2–3 days. All cell 213 culture ingredients were purchased from Sigma-Aldrich. Toxicity of the respective 214 complexes was determined using the 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium 215 216 bromide (MTT) cell viability assay after 24 h of incubation with C1, C2, C3, C4, C5, or C6. MTT is a yellow compound that, when reduced by functioning mitochondria, produces 217 purple formazan crystals that can be measured spectrophotometrically. For this purpose, 218 219 MTT (Sigma-Aldrich) was dissolved in phosphate buffered saline (PBS) to a concentration of 5 mg/mL and further diluted in culture medium (1:11). Cells were incubated with this 220 MTT-solution for 3 h under normal culture conditions. Afterwards 155  $\mu$ L of the solution 221 were rejected and 90 µL of DMSO were added. To completely dissolve the formazan salts 222 plates were incubated for 10 min on a shaker and afterwards quantified by measuring the 223 absorbance at 535 nm with a ELISA microplate reader. Cell viability was calculated as 224 percentage of surviving cells compared to untreated control cells. As shown in Figure S16, 225 toxicity of complexes tested is undistinguishable from the vehicle (i.e. DMSO) alone. 226

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229 Table S1. Minimal inhibition concentration (µg/ml) of tested compounds determined after

230	48 and 72 h for yeasts,	and after 24 and	1 48 h for bacteria.
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	Yeasts				Bacteria			
	Candida albicans		Cryptococcus spp.		S. enterica <sup>a</sup>			
	48 h	72 h	48 h	72 h	24 h	48 h		
C1	-	-	-	-	-	-		
C2	-	-	-	-	-	-		
С3	-	-	-	-	-	-		
C4	-	-	-	-	-	-		
C4	-	-	-	-	-	-		
C5	-	-	-	-	-	-		
C6	-	-	-	-	-	-		
K1 <sup>b</sup>	_d	_d	<1.042	<1.042	ND	ND		
Cam <sup>c</sup>	ND	ND	ND	ND	3.125	± 3.125	±	
					0.156	0.156		

- : The inhibition was indistinguishable from the vehicle alone (i.e. DMSO for C1 to C6, and

- 232 **K1**; Ethanol for **Cam**)
- 233 ND: Not determined
- 234 a: Salmonella enterica serovar Typhimurium ATCC14028s
- <sup>235</sup> <sup>b</sup>: Ketoconazole (antifungal compound)
- 236 <sup>c</sup>: Chloramphenicol (antibacterial/antibiotic)
- <sup>237</sup> <sup>d</sup>: The *Candida albicans* strain tested is resistant to ketoconazole
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Figure S15. MTT Assay in HeLa cells. Cells were incubated 24 h with the complexes prior to measuring cell viability. Red line: Rhenium complex C1 (A), C2 (B), C3 (C), C4 (D), C5 (E), or C6 (F) (200  $\mu$ g/ml + DMSO 50% v/v, 100  $\mu$ g/ml + DMSO 25% v/v, 50  $\mu$ g/ml + DMSO 12.5% v/v, or 25  $\mu$ g/ml + DMSO 6.3% v/v); Blue line: Vehicle alone (i.e. DMSO) (50% v/v, 25% v/v, 12.5% v/v, or 6.3% v/v, respectively). Medium alone was used to set 100% viability. C- corresponded to DMSO 100% v/v. The figure shows a representative experiment.





Cryptococcus spp.

Figure S16. Fluorescence confocal microscopy images showing *Cryptococcus spp.* (yeasts) under confocal microscopy. The "Red channel" corresponded to excitation of 405 nm and emission collected in a range of 555 to 625 nm. In all the cases, microorganisms were observed fresh, immobilized with 1% agarose, using a 100X objective. White bars represent 5 μm. DMSO alone was used to set the detection threshold.