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Electronic Supplementary Information

Fig. S1. The influence of media and the intracellular environment on photoluminescence spectra obtained from a Raman Renishaw inVia microscope, 514 nm excitation wavelength. Spectra show NV PL of a) oxidized FND particles in water, medium and cells; b) FND-PEI complex in water, medium and cells; c) FND-PEI-DNA complex in water, medium and cells after 30, 60 and 120 min incubation.



Fig. S2. Visualization of red (FND) and green (FAM-labelled DNA) signal colocalization derived by Olympus software at the end of the experimental period. Representative images of a) extracellular and b) intracellular signal distribution in IC-21 cells incubated with FND-PEI-DNA complexes for 2 h. Prominent separation of the red signal derived from DNA-free FNDs is marked with an arrow.



Fig. S3. Confocal analysis of FND and FAM signal colocalization (detailed view). IC-21 macrophages incubated with FND complexes for 30 min. FND-PEI: cells incubated with 25 μ g/ml FND-PEI, DNA: cells transfected with X-tremeGENE HP transfection reagent and 165 ng of the 137-kb DNA fragment labelled with FAM, FND-PEI-DNA: cells transfected with 25 μ g/ml FND-PEI and 165 ng of the DNA. From the left: DAPI staining of nuclei; FAM signal; fluorescence of NV centers; merged fluorescence. Arrows point at cells that have internalized or are about to engulf nanodiamond particles and/or DNA oligonucleotide.





Fig. S4. Flow cytometry analysis of transfection efficiency of AlexaFluor 488-labeled oligonucleotide in IC-21 cells using FND-PEI-DNA complex or a commercial transfection system X-tremeGENE HP. These data are source data for Fig. 3. a) Representative histograms of Alexa Fluor 488 intensity measured using flow cytometer in IC-21 cells incubated with transfection mixture for 30, 60, and 120 minutes. AlexaFluor 488-labeled oligonucleotide was used to form the transfection complex with FND-PEI (FND-PEI-DNA) or X-tremeGENE (HP-DNA). The histograms show intensity of Alexa Fluor 488 signal (x-axis) and cell counts (y-axis). We observed a time-dependent increase of cells exhibiting AlexaFluor 488-positive signal in both FND-PEI-DNA and HP-DNA alternatives. However, this percentage was always higher for FND-PEI-DNA. b) Representative dot plots of Alexa Fluor 488 and Hoechst 33258 signals and appropriate percentage of events in each quadrant (Q I-IV). The dot plot analyses show distribution of cells positive or negative on Alexa Fluor 488 and/or on Hoechst 33258. The intensity of Alexa Fluor 488 signal is shown on x-axis and the intensity of Hoechst 33258 signal on y-axis. The appropriate percentage of cells in each quadrant (Q I-IV, counted clockwise, starting at the top left corner) is revealed. Here, Q I represent Hoechst 33258 positive cells; dead cells with membrane permeable for Hoechst dye without Alexa Fluor 488 signal. Q II represent double positive cells; cells positively transfected with Alexa Fluor 488-oligo but possessing permeable membrane (high Hoechst 33258 signal). Q III represent cells transfected with Alexa Fluor 488-oligo which are alive without Hoechst 33258 signal. Q IV represents double negative cells, with no signal either from Alexa Fluor 488 or Hoechst 33258.



Fig. S5. Detection of green fluorescent protein (GPF) signal 48 h after transfection of HT-29 cells with FND-PEI and X-tremeGENE HP transfection reagent, both complexed with the pGFP expression plasmid. FND-PEI: cells incubated with 25 µg/ml FND-PEI, GFP: cells transfected with X-tremeGENE HP transfection reagent and 2 µg pGFP, FND-PEI-GFP: cells transfected with 25 µg/ml FND-PEI and 2 µg pGFP. From the left: GFP signal; fluorescence of NV centers; merged fluorescence with bright field. Arrows point out cells that internalized nanodiamonds and/or plasmid DNA.