

Electronic Supplementary Information

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DNA Nanomachine Based Regenerated Sensing Platform: a Novel Electrochemiluminescence Resonance Energy Transfer Strategy for Ultra-high Sensitive Detection of MicroRNA from Cancer Cells

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14 **Table 1.** *The oligonucleotide sequences applied in the proposed work.*

Name	Sequence (5' to 3')
A	TGCCTTGTAAGAATCAAATGCTTCGGAT
B	SH- GGTCGCTCTTACGGCATTTCGACGACGGCTCGGAGAAGAGAT
C	ATCAGACTGATGTTGAACTCATATCCGAAGCATTCCAGGT- SH
DNA H1	GCATCCATCAGACTGATGTTGAACTCACTATARGGAAGAGA TG TAGCTTATCAGACTGATGTTGATCAACATCAGTCTGATAG CTA
DNA H2	TAGCTTATCAGACTGATGTTGAACTCACTATGTCTATGCGTT TCATATATTCTCCGAGCCGGTCGAAATAGTGAGTTCAACATC

	AGTCTGATGGATGC
DNA H3	ATCAGACTGATGTTGAACTCACTATATCTCTTCTCCGAGCCG GTGCGAAATAGTGAGTTCAACATCAGTCTGATAAGCTA
miRNA-21	UAG CUU AUC AGA CUG AUG UUG A
miRNA-141	UAA CAC UGU CUG GUA AAG AUGG
miRNA-155	UUA AUG CUA AUC GUG AUA GGG GU
One-base mismatched miRNA-21	UCG CUU AUC AGA CUG AUG UUG A

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2 **The buffer and detection solution used in this work.**

3 TAE/Mg²⁺ buffer (40 mM Tris, 20 mM acetic acid, 2 mM EDTA and 12.5 mM
4 magnesium acetate, pH 8.0) was used to dilute DNA sequences in this work. RNA
5 hybridization buffer (Tris-HCl containing 0.2 mM NaCl, 10 mM MgCl₂ and 1 mM
6 EDTA, pH 8.0) was introduced to dilute miRNA-21. Thiolated DNA strand used in the
7 paper was activated with 10 mM TCEP and stored at 4 °C for 2 h before introducing to
8 the sensing platform. Phosphate buffered solution (PBS, pH 7.4) was employed for the
9 preparation of AF488. CdSe@ZnS QDs were diluted by boric acid buffer (pH 8.4).
10 Finally, Ferricyanide solutions (Fe(CN)₆^{3/4-}, 5 mM, pH 7.4) which were employed for
11 CV measurements were achieved by dissolving potassium ferricyanide and potassium
12 ferrocyanide with PBS buffer (pH 7.4). All the hairpin hybridizations were heated to
13 95 °C for 2 min before using and cool down to room temperature over 1h. Double
14 distilled water was employed throughout.

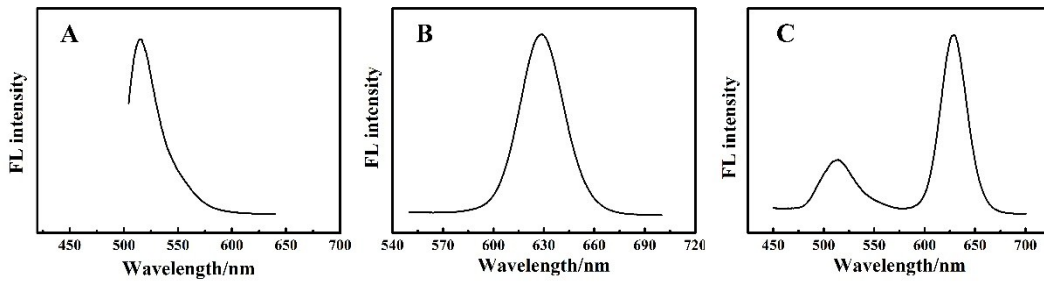
15 **Analytical apparatus and measurements.**

16 Cyclic voltammetric (CV) and ECL detections were performed with a CHI 660E

1 electrochemistry workstation (Shanghai CH Instruments, China) and a MPI-E ECL
2 analyzer (Xi'an Remax Electronic Science & Technology Co), respectively. The
3 measurements were carried out with a three-electrode system with glassy carbon
4 electrode (GCE) as the working electrode, platinum wire as the counter electrode and
5 Ag/AgCl (sat. KCl) as the reference electrode. Additionally, the voltage of the
6 photomultiplier tube (PTM) was set at 800 V, and the potential scanning ranged from
7 -2 to 0 V.

8 **The fluorescence response of AF488 and QDs**

9 As shown in Fig. S1, In order to further understand the mechanism of the proposed
10 ERET system, the fluorescence spectrum of AF488, QDs and the composite of AF488
11 and QDs were carried out to further confirm the luminescent properties for the
12 clarification. As showed in the following figure A and B, the luminescent emission
13 wavelength of AF488 was measured to be about 515 nm and the luminescent emission
14 of QDs was about to be 630 nm. Importantly, the composite of AF488 and QDs
15 exhibited a dual-emission at 515 nm and 630 nm with the excited wavelength of 496
16 nm. 496 nm was the excited wavelength of donor-AF488, while the composite of
17 AF488 and QDs presented a dual-emission spectrum at the excitation of 496 nm,
18 demonstrating the energy was transferred from AF488 to QDs successfully. The
19 obtained result was accordance with the ECL investigation, which suggested the
20 mechanism of the proposed ERET system comprehensively.



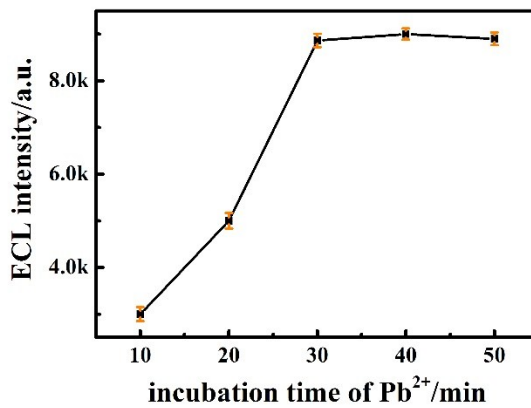
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2 **Fig. S1** The fluorescence response of AF488 (A), QDs (B) and the composite of AF488

3 and QDs (C).

4 **The optimization of incubation time of Pb²⁺**

5 In order to make this study more convincing, we optimized the incubation time of
 6 Pb²⁺. As shown in Fig. S2, the ECL intensity increased with the increasing incubation
 7 time of Pb²⁺ and reached a platform after the incubation time was longer than 30 min.
 8 So the optimal incubation time of Pb²⁺ incubation was 30 min.



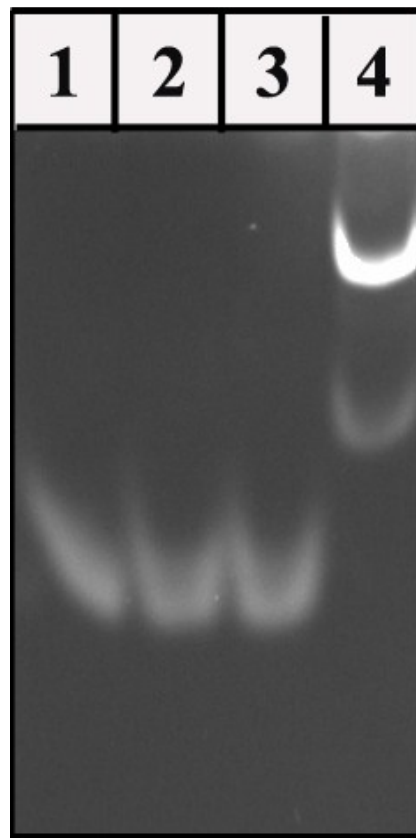
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10 **Fig. S2** The optimization of the Pb²⁺ incubation time of the proposed sensor.

11 **Polyacrylamide gel electrophoresis (PAGE) characterization**

12 In order to characterize the successful synthesis of the DNA tweezer,
 13 polyacrylamide gel electrophoresis (PAGE) was employed and the result was shown as
 14 follows. The distinct bands from lanes 1-3, respectively, correspond to the three
 15 assembly strands (DNA-A, DNA-B and DNA-C). As can be seen, the mixture of the

1 three assembly strands (lane 4) showed a band with lower mobility accompanied by the
2 disappearance of the corresponding bands in lanes 1-3, indicating the successful
3 formation of the DNA tweezer.
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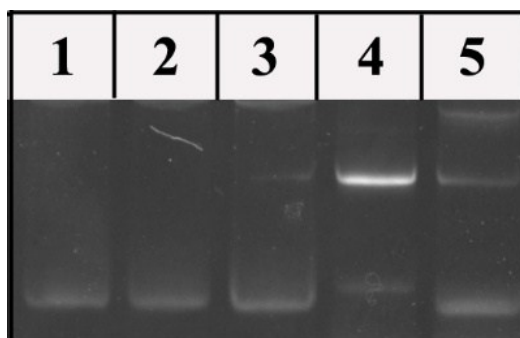


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6 **Fig. S3** PAGE (16%) analysis of assembly of the DNA tweezer. The concentration
7 of the strands was 2 μ M respectively.

8 **PAGE performance of the formation and cleaving of “Y” structure**

9 As shown in Fig. S4, the formation of “Y” structure and production of reporter
10 DNA were investigated by PAGE. Lane 1-3 represented hairpin structure of H1, H2
11 and H3, respectively. A band with low mobility was observed with the aid of target
12 miRNA, suggesting the successful formation of “Y” structure. When the “Y” structure
13 was added with Pb²⁺, the band of lane 4 disappeared and the band of lane 1-3 showed

1 up, which demonstrated that the “Y” structure was cleaved by the DNAzyme with the
2 generation of numerous reporter DNA.



3

4 **Fig. S4** PAGE (16%) analysis of the formation of “Y” structure and reporter DNA.

5 Lane 1: H1; Lane 2: H2; Lane 3: H3; Lane 4: H1, H2, H3 and target miRNA; Lane 5:

6 the “Y” structure was cleaved by Pb^{2+} . The concentration of the strands was $2\mu M$

7 respectively.

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