

***Modelling flavoenzymatic charge transfer events: development of catalytic
indole deuteration strategies.***

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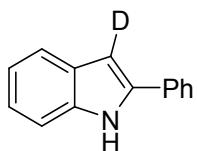
I GENERAL INFORMATION

All reagents were purchased from commercial suppliers: Acros Organics, Alfa Aesar or Sigma Aldrich and used without further purification. Flash chromatography was performed on chromatography grade, silica, 60 Å particle size 35-70 micron from Sigma Aldrich using the solvent system as stated. ^1H and ^{13}C NMR was performed on Brüker Avance 250 (^1H 250 MHz) Brüker Avance 300 (^1H 300 MHz and ^{13}C 75 MHz), Brüker Avance 400 (^1H 400 MHz and ^{13}C 100 MHz), Brüker Avance 500 (^1H 500 MHz and ^{13}C 125 MHz) or Agilent ProPulse 500 (^1H 500 MHz and ^{13}C 125 MHz) as stated. Chemical shifts are reported in parts per million (ppm) relative to tetramethylsilane (TMS) ($\delta = 0.00$). Coupling constants are reported in Hertz (Hz) and signal multiplicity is denoted as singlet (s), doublet (d), triplet (t), quartet (q), quintet (quin.), sextet (sex.), septet (sept.), multiplet (m), and broad (br). High resolution mass spectrometry electrospray (ESI) was performed on a Brüker μTOF using electrospray ionisation (ESI) in either positive or negative ionisation. Infra-red spectroscopy was carried out using a Perkin Elmer Spectrum RX FT-IR system. All NMR data of partially deuterated compounds were compared to the authentic starting materials obtained from Sigma-Aldrich.

II EXPERIMENTAL SECTION

General Procedure for flavin-catalysed deuteration of indole (General procedure 1)

To a small vial (5 mL) of 0.5 mL CD₃OD was added 0.1 mmol of indole, followed by 3 mol % (1 mg) **5a**. The vial was shaken and left to stand at room temperature for one minute or alternatively capped and left to stand at room temperature for 15 minutes. The reaction was then quenched with diethyl ether before being run through a pipette column with silica gel washed with a 1:2 mixture of petroleum ether/ethyl acetate (2% triethylamine). Solvent was removed by rotary evaporation.



3-deutero-2- phenylindole (7a)

General procedure 1 (1 minute) using indole (12 mg). Full characterisation was performed on this compound and is shown below.

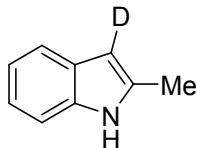
¹H NMR (500 MHz, CDCl₃) δ_H 7.70 (d, *J* = 7.5 Hz, 2H), 7.66 (d, *J* = 7.6 Hz, 1H), 7.47 Hz (apparent t, *J* = 7.5 Hz, 2H), 7.35 (t, *J* = 7.5 Hz, 1H), 7.22 (apparent t, *J* = 7.6 Hz, 1H), 7.15 (apparent t, *J* = 7.6 Hz, 1H).

¹³C NMR (125 MHz, CDCl₃) δ_C 138.0, 137.0, 132.5, 129.3, 127.9, 125.3, 122.5, 120.8, 120.4, 111.0, 100.2 (d, *J*_{C,D} = 75 Hz)

²D NMR (75 MHz, CD₃OD) δ_D 6.81 (s)

IR ν_{max} (neat) 3443, 3048, 2564, 1668, 1456, 1445, 1349 cm⁻¹

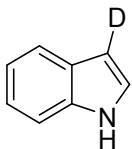
HRMS (ESI, -ve) Calcd. for C₁₄H₉ND, 193.0876. Found: 193.0887 (M-H)⁻



3-deutero-2-methylindole (7b)

General procedure 1 (15 minutes) using 2-methylindole (13 mg).

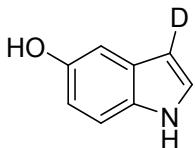
¹H NMR (250 MHz, CDCl₃) δ_H 7.87 (br s, 1H), 7.56 – 7.48 (m, 1H), 7.33 – 7.25 (m, 1H), 7.17 – 7.02 (m, 2H), 6.22 (s, 0.06 H), 2.44 (s, 3H).



3-deuteroindole (7c)

General procedure 1 (15 minutes) using indole (12 mg).

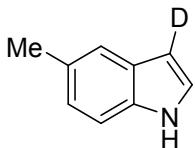
¹H NMR (300 MHz, CDCl₃) δ_H 7.69 (d, *J* = 7.9 Hz, 1 H), 7.44 (d, *J* = 8.0, 1 H), 7.22-7.10 (m, 4 H), 6.58 (s, 0.16 H).



3-deutero-5-hydroxyindole (7d)

General procedure 1 (1 minute) using 5-hydroxyindole (13 mg).

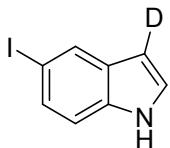
¹H NMR (250 MHz, CDCl₃) δ_H 7.25 (t, *J* = 4.3 Hz, 3 H), 7.19 (s, 1 H), 7.00 (d, *J* = 2.4 Hz, 1 H), 6.78 (dd, *J* = 8.7, 2.5 Hz, 1 H), 6.43 (d, *J* = 2.4 Hz, 0.17 H).



3-deutero-5-methylindole (7e)

General procedure 1 (15 minutes) using 5-methylindole (13 mg)

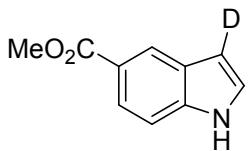
¹H NMR (250 MHz, CDCl₃) δ_H 8.05 (br s, 1 H), 7.44 (s, 1 H), 7.28 (m, 1 H), 7.17 (s, 1 H), 7.03 (d, *J* = 8.26 Hz, 1 H), 6.48 (d, *J* = 2.5 Hz, 0.17 H), 2.46 (s, 3 H).



3-deutero-5-iodoindole (7f)

General procedure 1 (15 minutes) using 5-iodoindole (24 mg).

¹H NMR (300 MHz, CDCl₃) δ_H 8.19 (br s, 1 H), 7.99 (s, 1 H), 7.44 (dd, *J* = 8.5, 1.9 Hz, 1 H), 7.18 (d, *J* = 8.5 Hz, 2 H), 6.48 (d, *J* = 1.9 Hz, 0.35 H).



3-deutero-methylindole-5-carboxylate (7g)

General procedure 1 (1 minute) using methyl indole-5-carboxylate (18 mg).

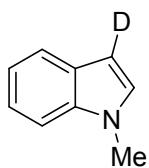
¹H NMR (250 MHz, CDCl₃) δ_H 8.42 (s, 1 H), 7.91 (d, 8.7 Hz, 1H), 7.41 (d, *J* = 8.7 Hz, 1 H), 7.27 (m, 2 H), 6.65 (m, 0.94 H), 3.93 (s, 3 H)



3-deutero-7-methoxyindole (7j)

General procedure 1 (15 minutes) using 7-methoxyindole (15 mg)

¹H NMR (250 MHz, CDCl₃) δ_H 8.18 (br s, 1 H), 7.16-7.01 (m, 3 H), 6.67 (s, 0.67 H), 6.53 (m, 1 H), 3.97 (s, 3 H)



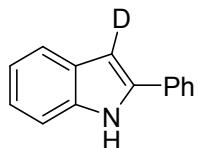
1-methylindole (7k)

General procedure 1 (15 minutes) using 1-methylindole (12 μL).

¹H NMR (250 MHz, CDCl₃) δ_H 7.64 (d, *J* = 8.04 Hz, 1 H), 7.35-7.05 (m, 4 H), 6.49 (d, *J* = 2.5 Hz, 0.54 H), 3.80 (s, 3 H).

General Procedure for acetyl chloride-catalysed deuteration of indole

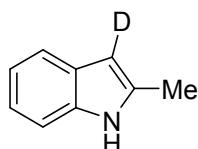
To a small vial (5 mL) of 0.5 mL CD₃OD was added 0.1 mmol of indole, followed by 10 µL of acetyl chloride solution (30 µL acetyl chloride in 1 mL CD₃OD). The vial was shaken and left to stand at room temperature for one minute or alternatively capped and left to stand at room temperature for 15 minutes. The reaction was then quenched with diethyl ether before being run through a pipette column with silica gel washed with a 1:2 mixture of petroleum ether/ethyl acetate (2% triethylamine). Solvent was removed by rotary evaporation.



3-deutero-2-phenylindole (7a)

General procedure 2 (1 minute) using 2-phenylindole (19 mg).

¹H NMR (250 MHz, CDCl₃) δ_H 8.37 (br s, 1 H), 7.66 (m, 3 H), 7.48-7.10 (m, 6 H), 6.84 (s, 0.07 H).

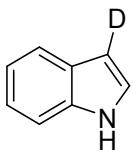


3-deutero-2-methylindole (7b)

General procedure 1 (15 minutes) using 2-methylindole (13 mg).

¹H NMR (250 MHz, CDCl₃) δ_H 7.56 – 7.48 (m, 1H), 7.33 – 7.25 (m, 1H), 7.17 – 7.02 (m, 2H), 6.15 (br s, 0.17 H), 2.44 (s, 3H).

NB. In this case it is possible, due to the broad signal the NH has been deuterated and shifted, and we do not observe residual C3. We assumed the ‘worst’ case that the broad peak is C3 proton.



3-deuteroindole (7c)

General procedure 2 (15 minute) using indole (12 mg). Full characterisation was performed on this compound and is shown below.

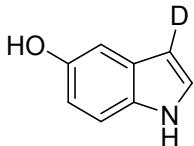
¹H NMR (500 MHz, CDCl₃) δ_H 7.67 (d, *J* = 8.0 Hz, 1H), 7.41 (d, *J* = 8.0 Hz, 1H), 7.24 – 7.19 (m, 2H), 7.16 – 7.12 (m, 1H).

¹³C NMR (125 MHz, CDCl₃) δ_C 135.7, 127.9, 124.0, 122.1, 120.8, 119.9, 111.1, 102.5 (t, *J_{C,D}* = 26 Hz).

²D NMR (75 MHz, CD₃OD) δ_D 6.48 (apparent s)

IR ν_{max} (neat) 3413, 2922, 2852, 1455, 1324 cm⁻¹

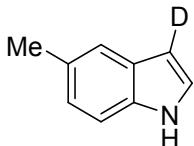
HRMS (ESI, -ve) Calcd. for C₈H₅ND 117.0563. Found: 117.0572 (M-H)⁻



3-deutero-5-hydroxyindole (7d)

General procedure 2 (1 minute) using 5-hydroxyindole (13 mg).

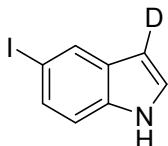
¹H NMR (250 MHz, CDCl₃) δ_H 7.27-7.24 (apparent d, *J* = 8.5 Hz, 2 H), 7.19 (s, 1 H), 7.04 (apparent d, *J* = 7.0 Hz, 1 H), 6.78 (dd, *J* = 8.5, 2.4 Hz, 1 H), 6.43 (d, *J* = 2.4 Hz, 0.18 H).



3-deutero-5-methylindole (7e)

General procedure 2 (1 minute) using 5-methylindole (13 mg)

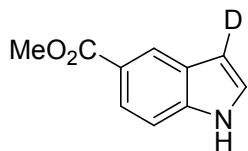
¹H NMR (250 MHz, CDCl₃) (br s, 1 H), 7.44 (s, 1 H), 7.28 (m, 1 H), 7.17 (s, 1 H), 7.03 (d, *J* = 8.26 Hz, 1 H), 6.48 (m, 0.57 H), 2.46 (s, 3 H).



5-iodoindole (7f)

General procedure 2 (15 minutes) using 5-iodoindole (24 mg).

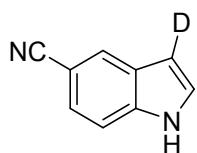
¹H NMR (250 MHz, CDCl₃) δ_H 7.99 (s, 1 H), 7.44 (dd, *J* = 8.5, 2.0 Hz, 1 H), 7.18 (d, *J* = 8.5 Hz, 2 H), 6.48 (d, *J* = 2.0 Hz, 0.19 H).



3-deutero-methylindole-5-carboxylate (7g)

General procedure 2 (15 minutes) using methyl indole-5-carboxylate (18 mg).

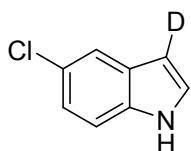
¹H NMR (250 MHz, CDCl₃) δ_H 8.42 (s, 1 H), 7.91 (d, *J* = 8.7 Hz, 1H), 7.41 (d, *J* = 8.7 Hz, 1 H), 7.27 (m, 2 H), 6.65 (m, 0.19 H), 3.93 (s, 3 H)



3-deutero-5-cyanoindole (7h)

General procedure 2 (15 minutes) using 5-cyanoindole (14 mg)

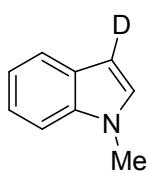
¹H NMR (250 MHz, CDCl₃) δ_H 8.01 (s, 1 H), 7.48-7.35 (m, 3 H), 6.65 (s), 0.16 H).



3-deutero-5-chloroindole (7i)

General procedure 2 (15 minutes) using 5-chloroindole (15 mg).

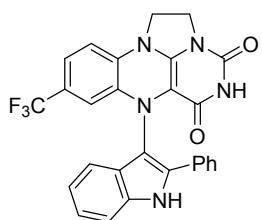
¹H NMR (250 MHz, CDCl₃) δ_H 7.54 (d, *J* = 2.2 Hz, 1H), 7.28 – 7.02 (m, 3H), 6.42 (d, *J* = 2.2 Hz, 0.06 H).



3-deutero-1-methylindole (7k)

General procedure 2 (1 minute) using 1-methylindole (13 µL) with 3 mol % acetyl chloride.

¹H NMR (250 MHz, CDCl₃) δ_H 7.64 (d, *J* = 8.1 Hz, 1 H), 7.35 (d, *J* = 8.1 Hz, 1 H), 7.22 (apparent d, *J* = 8.1 Hz, 1 H), 7.15-7.06 (m, 2 H), 6.50 (m, 0.27 H), 3.81 (s, 3 H).



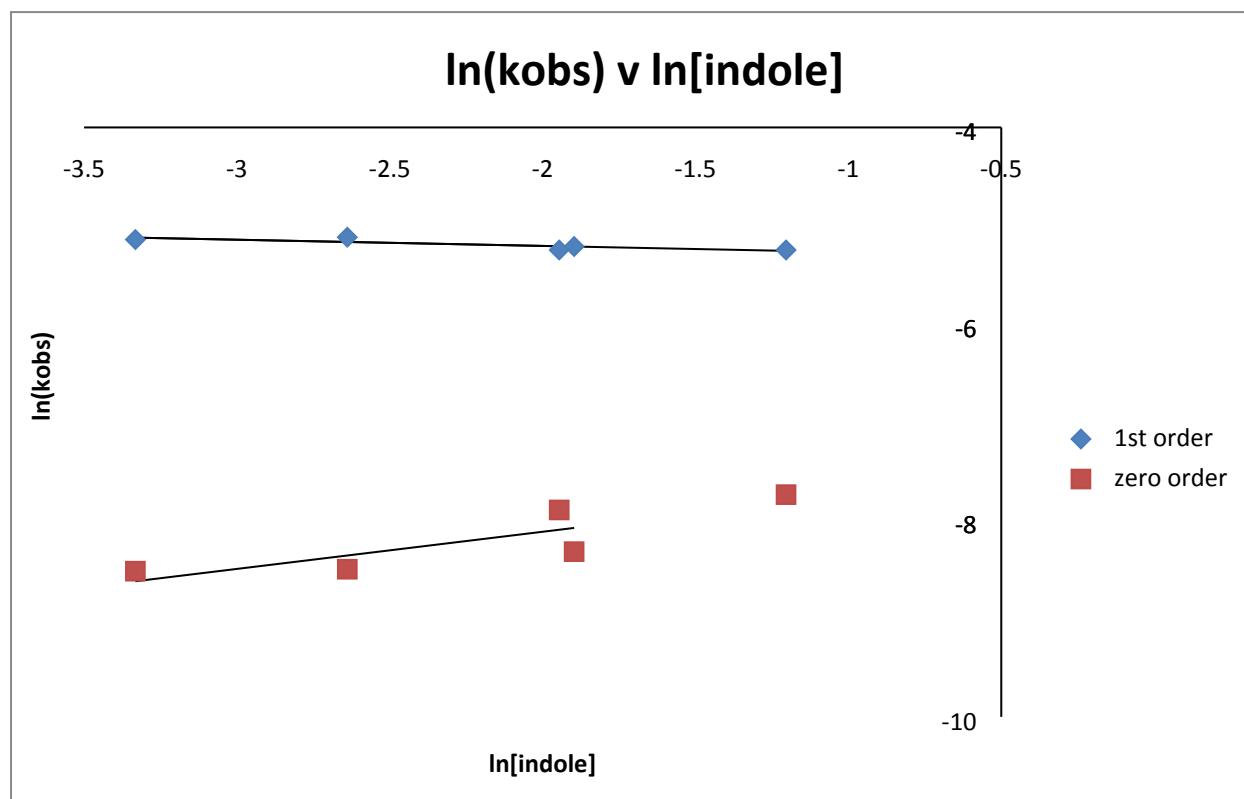
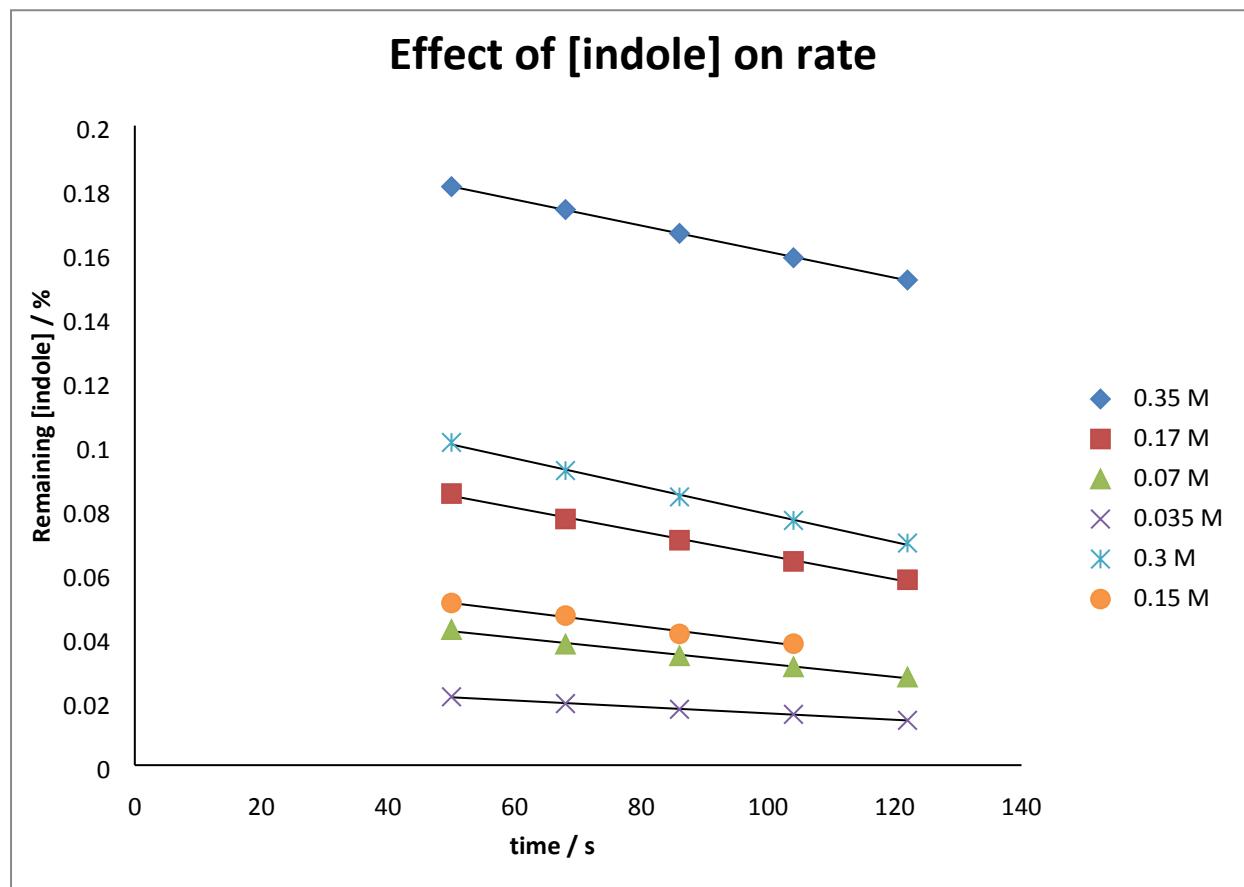
8-(2-phenyl-1H-indol-3-yl)-5-(trifluoromethyl)-1,8,11,13-tetraazatetracyclo[7.6.1.0^{2,7.0^{13,16]}]hexadeca-2,4,6,9(16)-tetraene-10,12-dione (9)}

Flavin **5a** (17 mg, 50 µmol) was dissolved in methanol (0.5 mL) and 2-phenylindole **6a** (10 mg, 50 µmol) was added, followed by water (100 µL). After six days, the yellow crystals were washed and decanted with 5:1 MeOH/water (5 x 1 mL), the product was isolated by column chromatography (100% CH₂Cl₂ → 100% EtOAc + 2% AcOH). **9** was isolated as a yellow solid (1 mg, 4%). ¹H NMR (300 MHz, CD₃CN) δ_H 9.67 (s, 9.67, 1H), 8.09 (d, *J* = 7.1 Hz, 2H), 7.95 (d, *J* = 8.0 Hz, 1H), 7.58 – 7.36 (m, 4H), 7.19 (t, *J* = 7.8 Hz, 1H), 7.09 (t, *J* = 7.8 Hz, 1H), 6.96 – 6.89 (m, 1H), 6.50 (d, *J* = 8.0 Hz, 1H), 6.26- 6.22 (m, 1H), 4.15 – 4.04 (m, 2H), 4.02 – 3.70 (br, 2H). ¹³C NMR (125 MHz, d⁶-Acetone) δ_C 147.5, 139.0, 138.2, 136.1, 135.0, 132.2, 128.9, 128.5, 128.1, 127.7, 125.5, 125.2 (m), 122.9, 122.2, 120.3, 119.7 (q, *J*_{C,F} = 4 Hz), 119.6, 117.9, 113.8 (q, *J*_{C,F} = 4 Hz), 112.7, 111.7, 111.6, 98.3, 45.9, 43.0 (not all C,F coupling resolvable). ¹⁹F NMR (471 Hz, d⁶-Acetone) δ_C -63.4. IR ν_{max} (neat) 3221, 3060, 1718, 1654, 1499, 1331, 1304, 1118. MP 199 – 204 °C. HRMS (ESI, -ve) *m/z* calcd. for C₂₇H₁₄F₃N₅O₂ 500.1329, found: 500.1331 (M-H)⁻

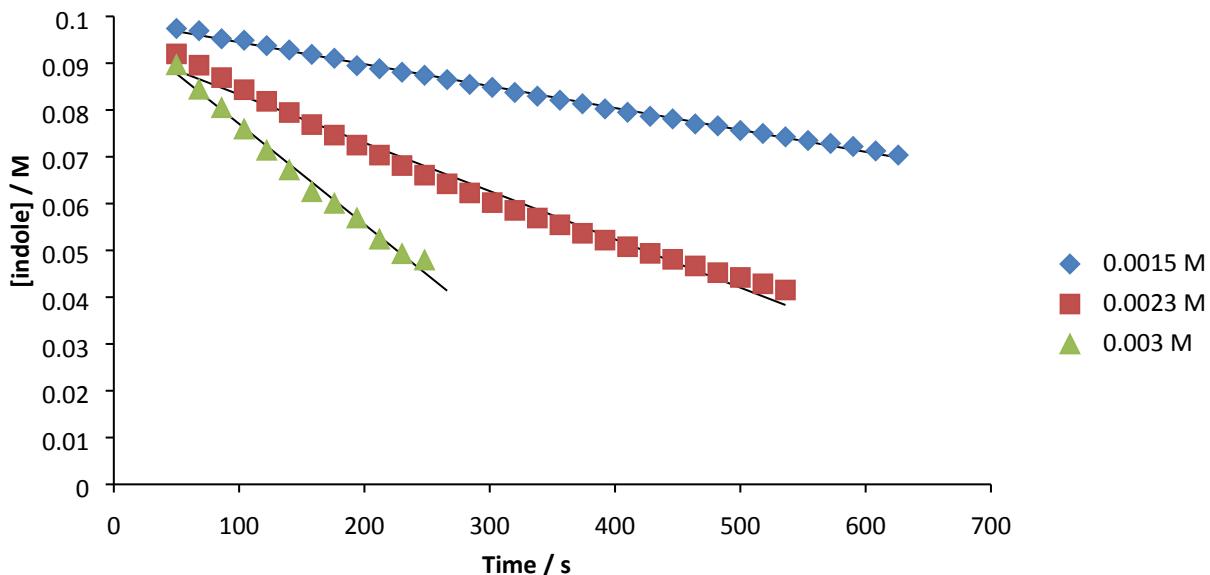
Kinetic experiments

General procedure for kinetic experiments was as those for isolation of deuteroindoles except the contents were placed in an NMR tube and multiple ^1H NMR spectra were automatically gathered without delay using an automation procedure, and ratios of deuteration were measured by integration of the C3 proton relative to another peak. Indole was used for flavin experiments, and 5-chloroindole for those using AcCl.

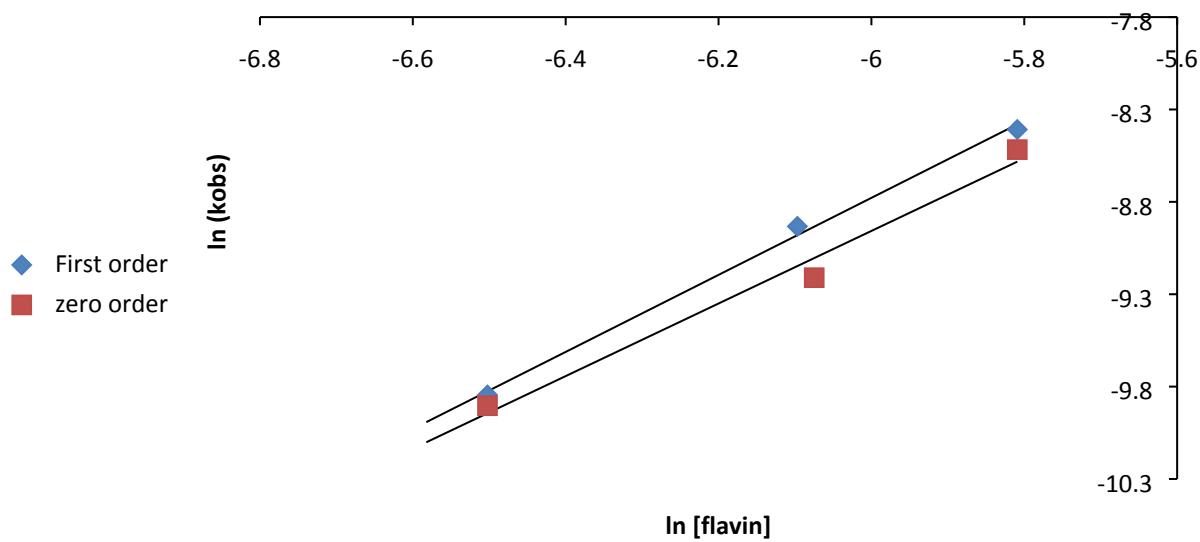
Unless deviated from, standard conditions were 0.1 mmol indole, 0.5 mL MeOD, 3 mol% flavin and 1.5 mol% AcCl.

Method A – flavin catalysis

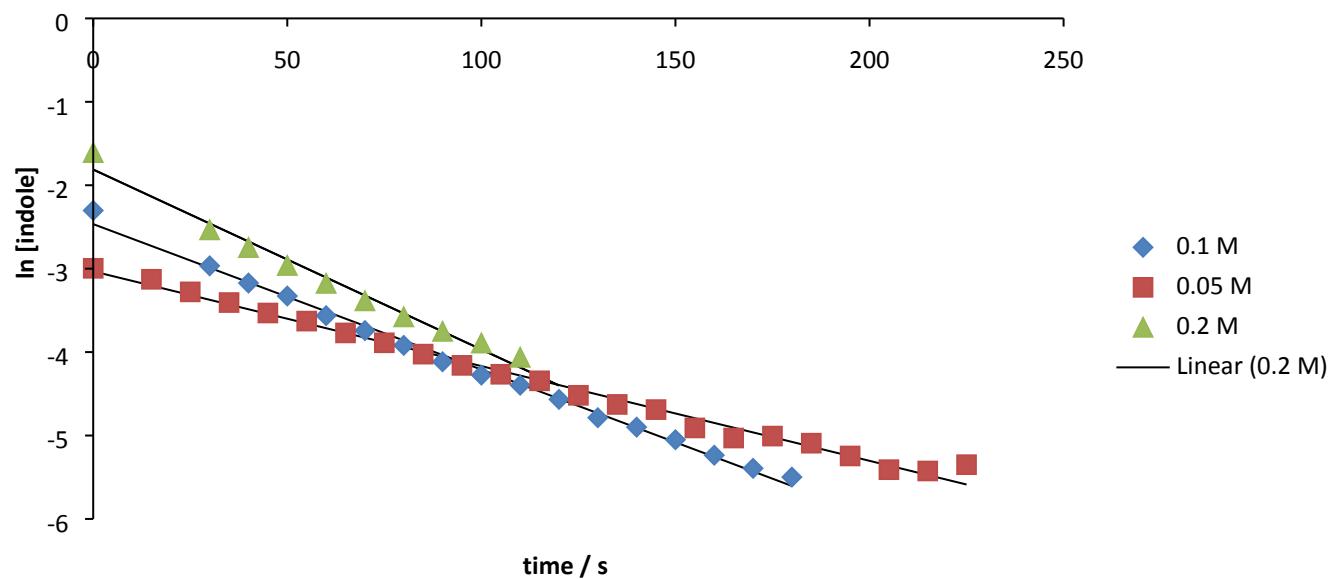
[Indole] v Time - Varying [flavin]



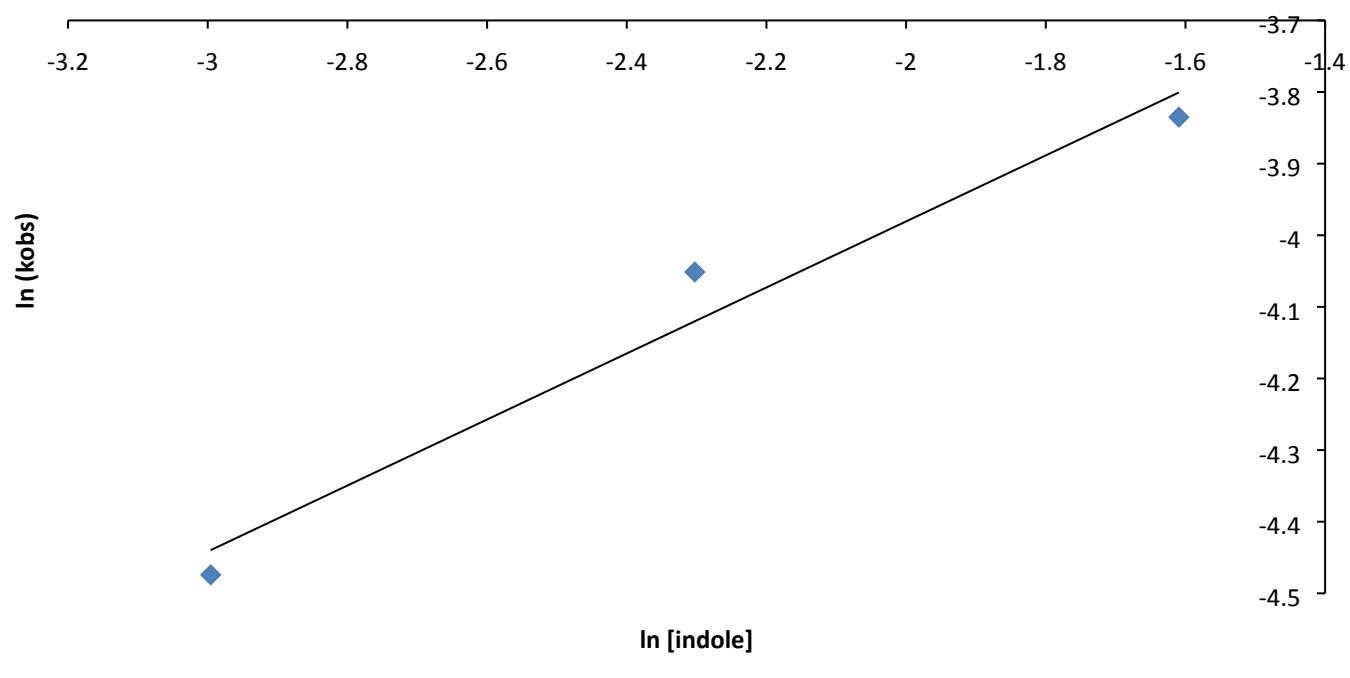
$\ln(k_{\text{obs}})$ v $\ln[\text{flavin}]$



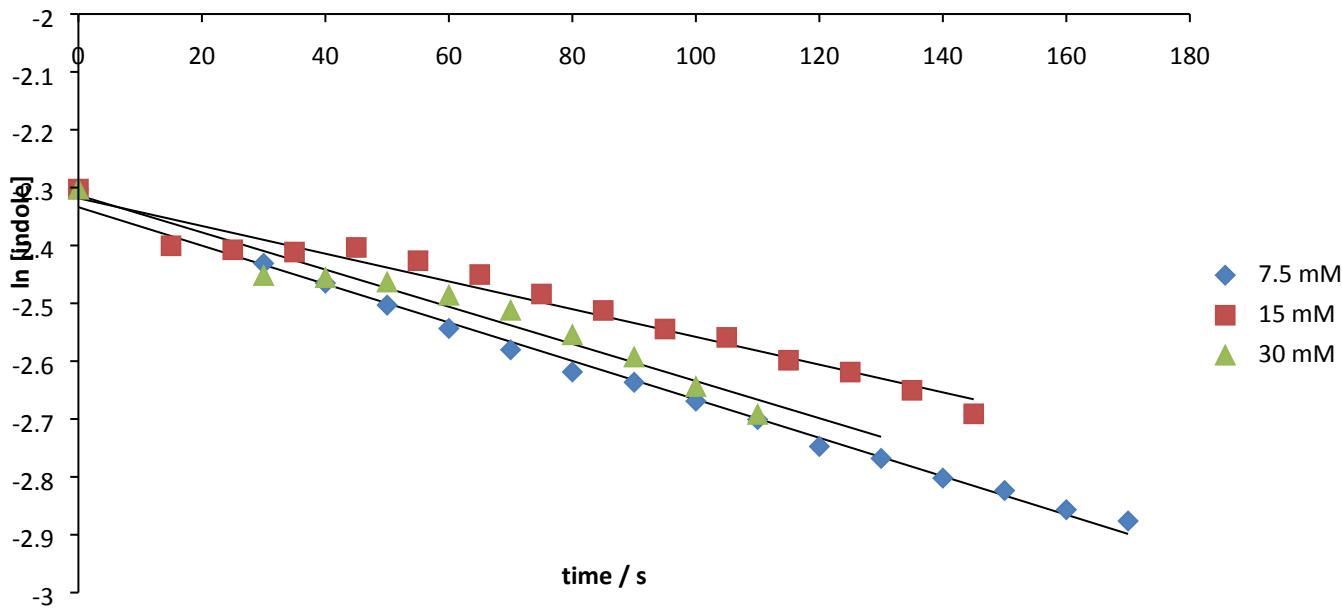
ln [indole] v time, varying [indole] - AcCl conditions



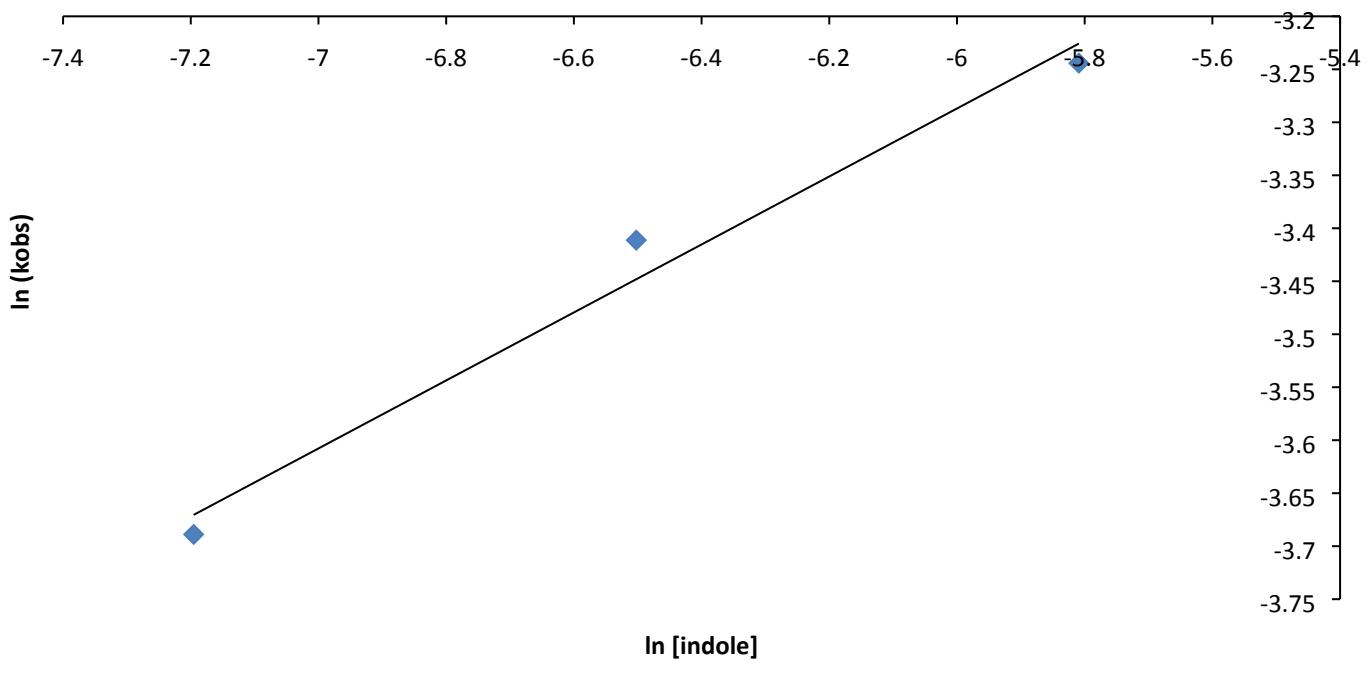
ln(k_{obs}) v ln[indole]

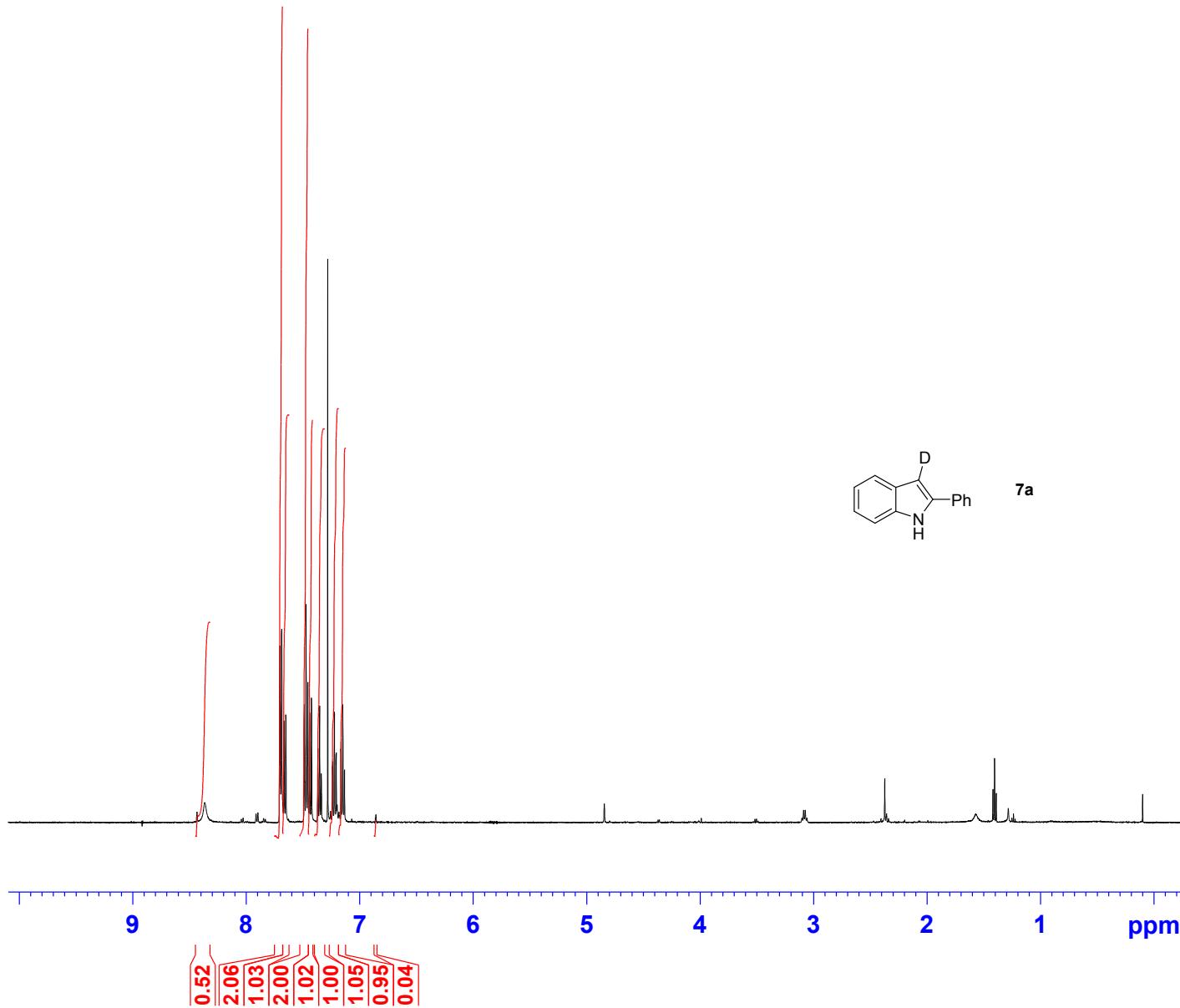


ln [indole] v time, varying [AcCl] - AcCl conditions



ln(k_{obs}) v ln[AcCl]



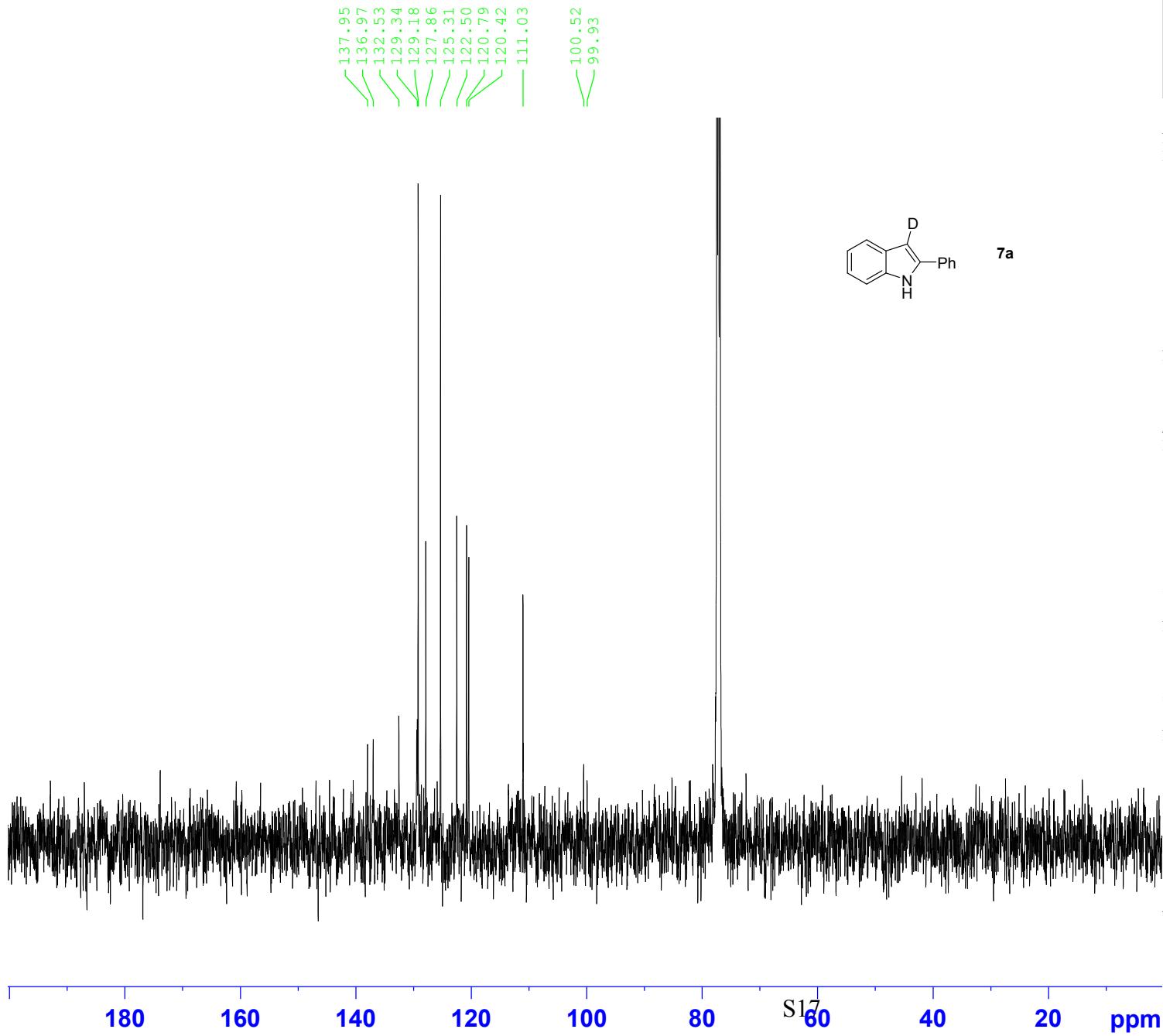
METHOD A

Current Data Parameters
NAME AM1415 r
EXPNO 1
PROCNO 1

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PULPROG zg30
TD 65536
SOLVENT CDC13
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DS 0
SWH 10330.578 Hz
FIDRES 0.157632 Hz
AQ 3.1719425 sec
RG 456
DW 48.400 usec
DE 6.50 usec
TE 298.0 K
D1 1.0000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 500.1330885 MHz
NUC1 1H
P1 9.50 usec
PLW1 27.19599915 W

F2 - Processing parameters
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LB 0 Hz
GB 0
PC 1.00



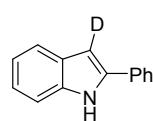
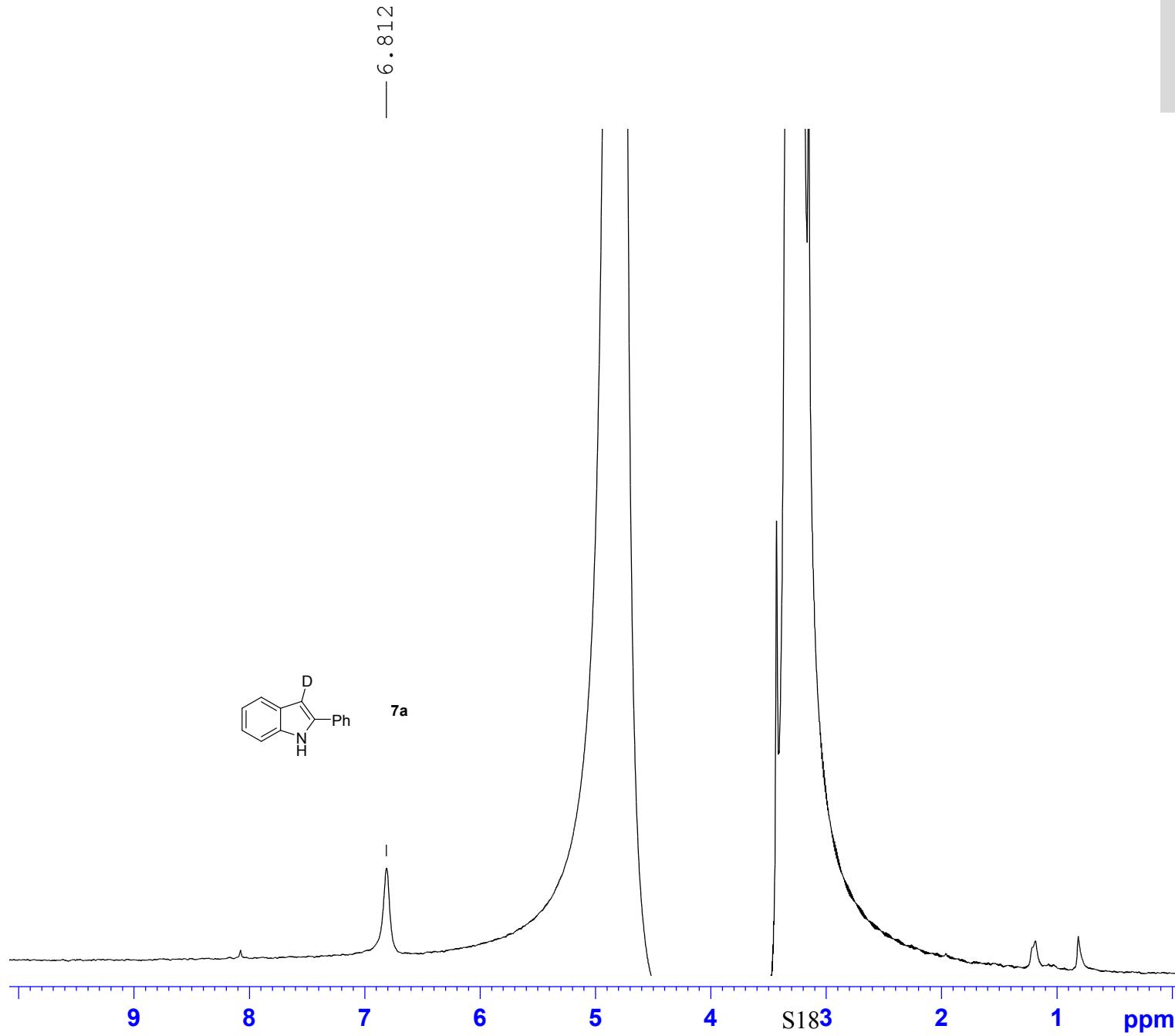
Current Data Parameters
 NAME AM1415 r
 EXPNO 2
 PROCNO 1

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 TD 65536
 SOLVENT CDCl₃
 NS 166
 DS 4
 SWH 29761.904 Hz
 FIDRES 0.454131 Hz
 AQ 1.1010048 sec
 RG 912
 DW 16.800 usec
 DE 6.50 usec
 TE 298.0 K
 D1 6.0000000 sec
 D11 0.0300000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 125.7703643 MHz
 NUC1 ¹³C
 P1 7.50 usec
 PLW1 63.6660037 W

===== CHANNEL f2 =====
 SFO2 500.1320005 MHz
 NUC2 ¹H
 CPDPRG[2] waltz16
 PCPD2 80.00 usec
 PLW2 27.19599915 W
 PLW12 0.34419000 W
 PLW13 0.22028001 W

F2 - Processing parameters
 SI 32768
 SF 125.7577707 MHz
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 SSB 0
 LB 4.00 Hz
 GB 0
 PC 1.40



7a

Current Data Parameters
NAME AM843 rerun
EXPNO 2
PROCNO 1

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F2 - Acquisition Parameters
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INSTRUM        spect
PROBHD        5 mm PABBO BB/
PULPROG       zg2h
TD             4096
SOLVENT        MeOD
NS              528
DS                 0
SWH            1151.543 Hz
FIDRES       0.281138 Hz
AQ            1.7784832 sec
RG             3.56
DW            434.200 usec
DE              6.50 usec
TE             298.0 K
D1            0.05000000 sec
D11           0.03000000 sec
TD0                  1

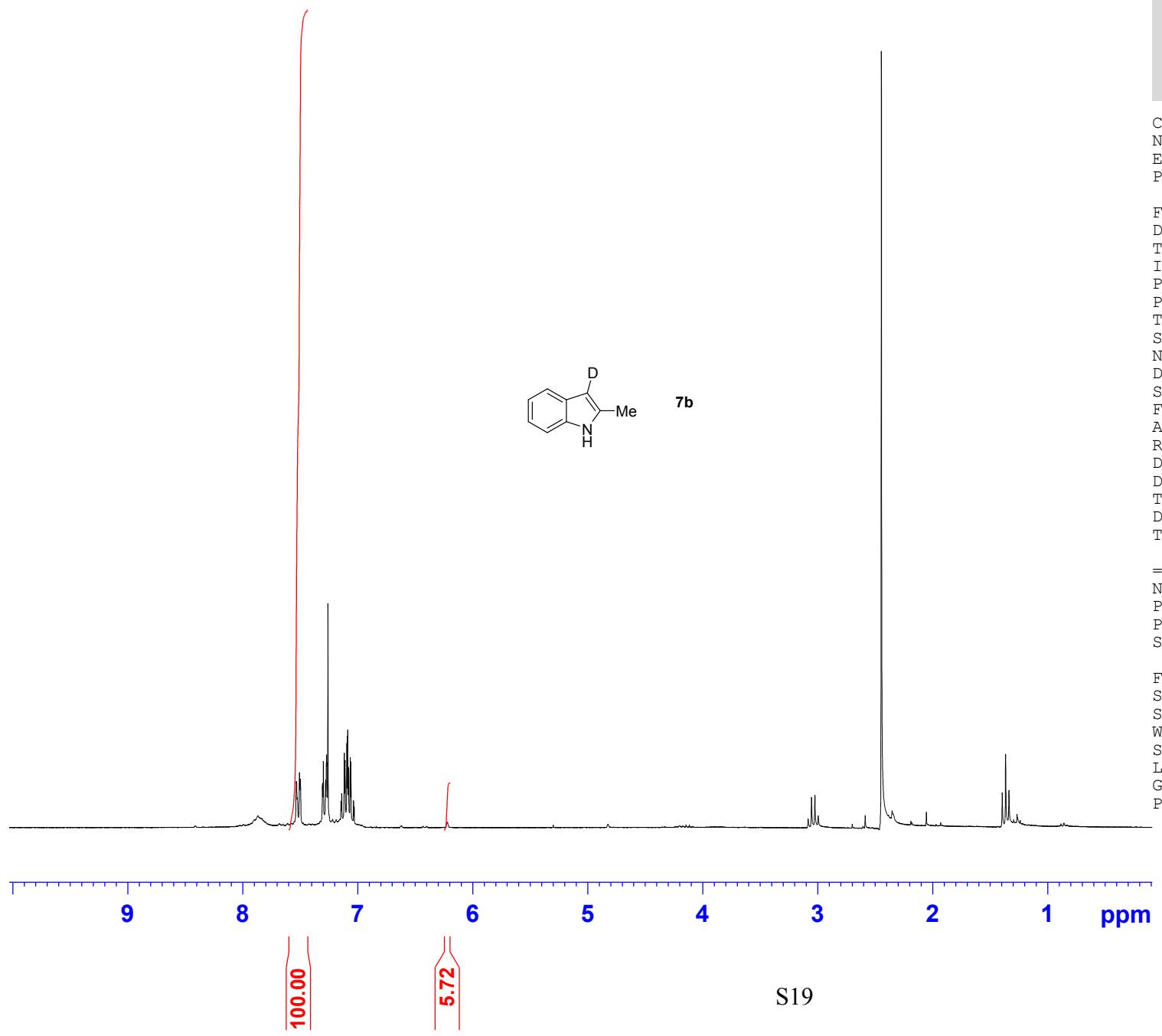
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SFO1 76.7732534 MHz
NUC1 2H
P1 230.00 usec
PLW1 2.79320002 W

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F2 - Processing parameters
SI           4096
SF          76.7730000 MHz
WDW          EM
SSB          0
LB           1.00 Hz
GB          0
PC          1.00

```



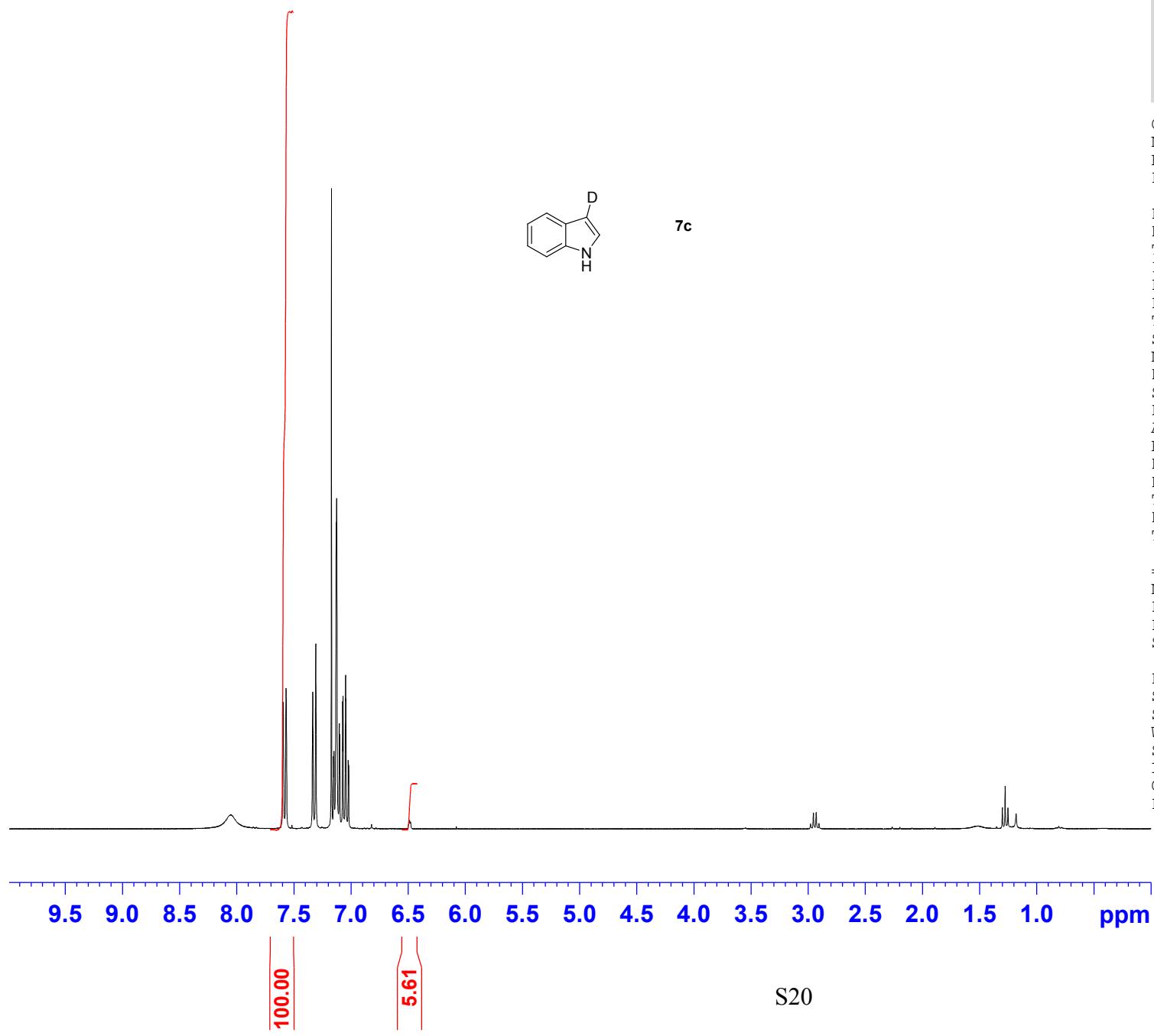
Current Data Parameters
 NAME 02272015
 EXPNO 40
 PROCNO 1

F2 - Acquisition Parameters
 Date 20150227
 Time 18.01
 INSTRUM spect
 PROBHD 5 mm DUL 1H-13
 PULPROG zg30
 TD 32768
 SOLVENT CDCl₃
 NS 32
 DS 2
 SWH 5175.983 Hz
 FIDRES 0.157958 Hz
 AQ 3.1653888 sec
 RG 228.1
 DW 96.600 usec
 DE 6.00 usec
 TE 300.0 K
 D1 1.00000000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 11.00 usec
 PL1 0 dB
 SFO1 250.1315450 MHz

F2 - Processing parameters
 SI 32768
 SF 250.1300077 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

AM1014



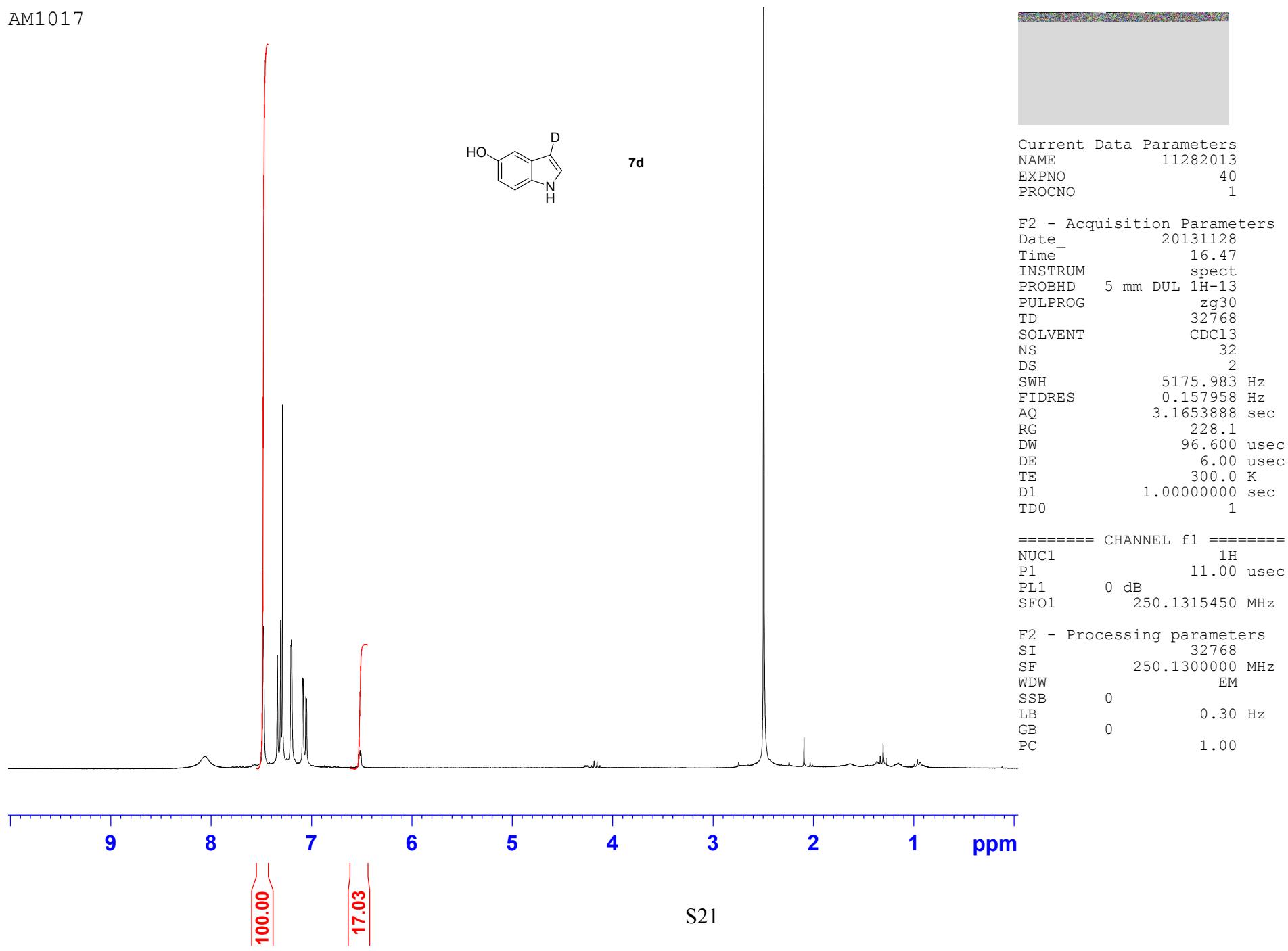
Current Data Parameters
NAME Nov28-2013
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20131128
Time_ 18.30
INSTRUM av300
PROBHD 5 mm BBO BB-1H
PULPROG zg30
TD 32768
SOLVENT CDCl₃
NS 32
DS 0
SWH 6172.839 Hz
FIDRES 0.188380 Hz
AQ 2.6542079 sec
RG 512
DW 81.000 usec
DE 6.00 usec
TE 294.0 K
D1 1.00000000 sec
TD0 1

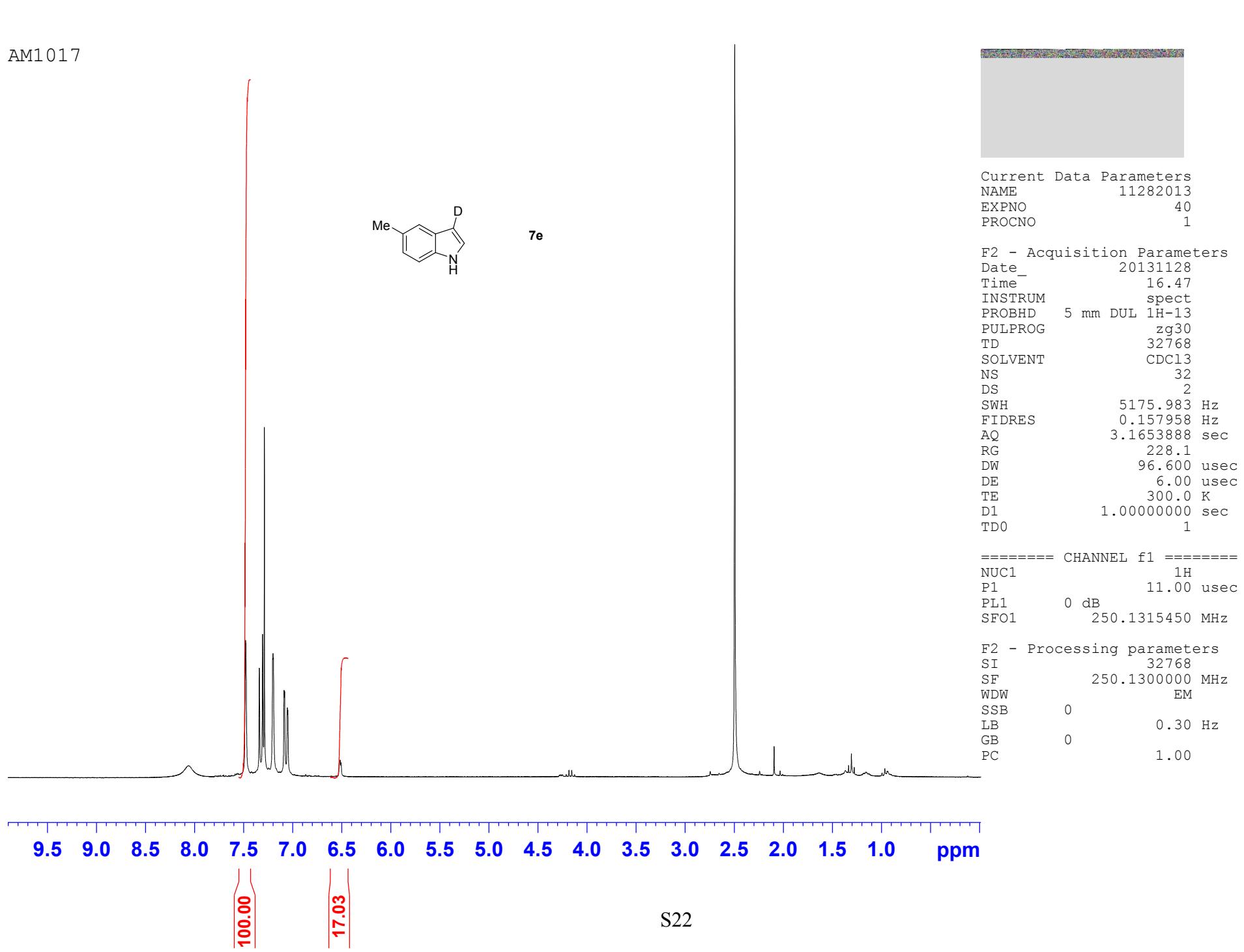
===== CHANNEL f1 =====
NUC1 1H
P1 9.40 usec
PL1 -1.50 dB
SFO1 300.2218540 MHz

F2 - Processing parameters
SI 32768
SF 300.2200284 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

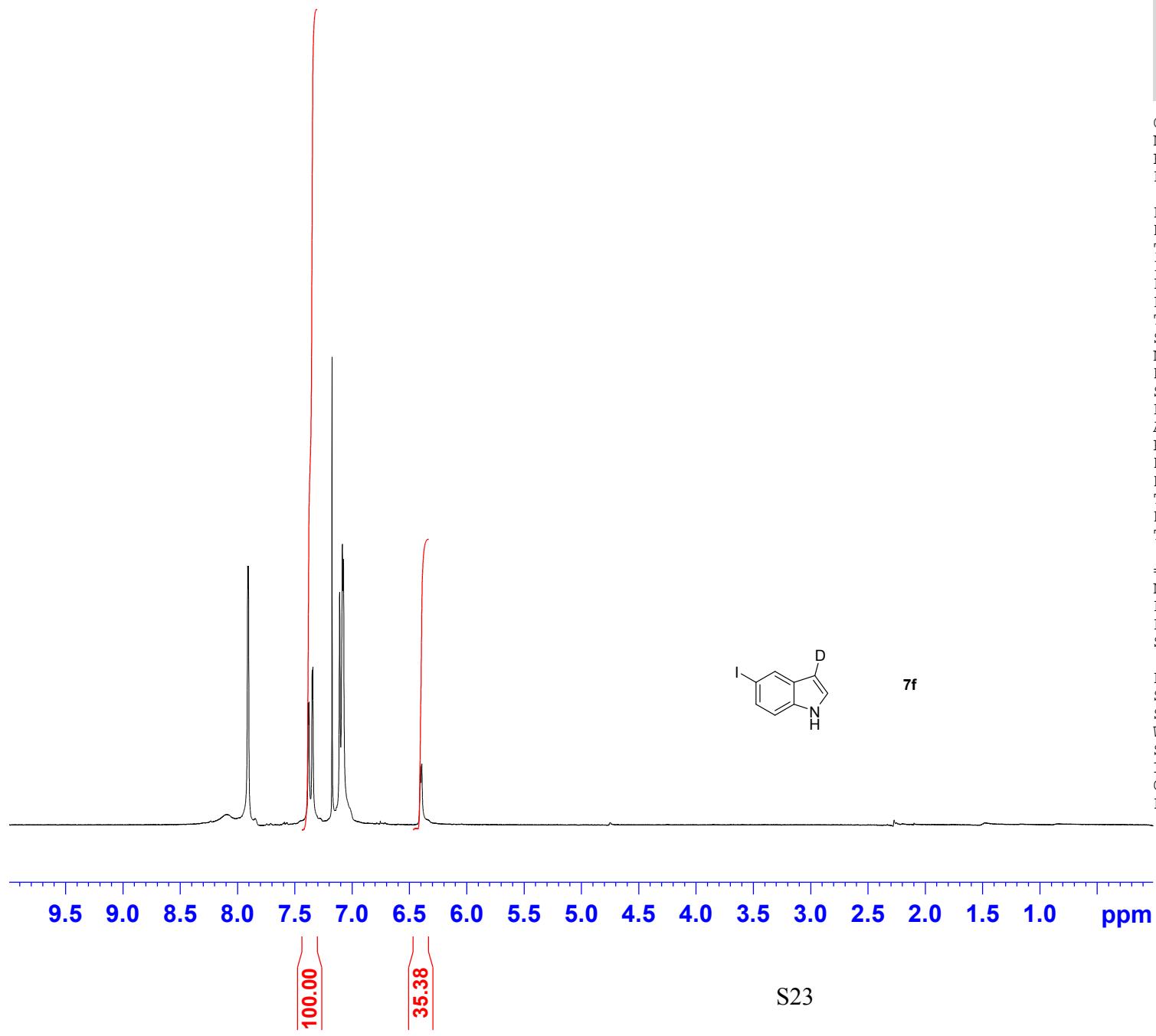
AM1017



AM1017



AM1437



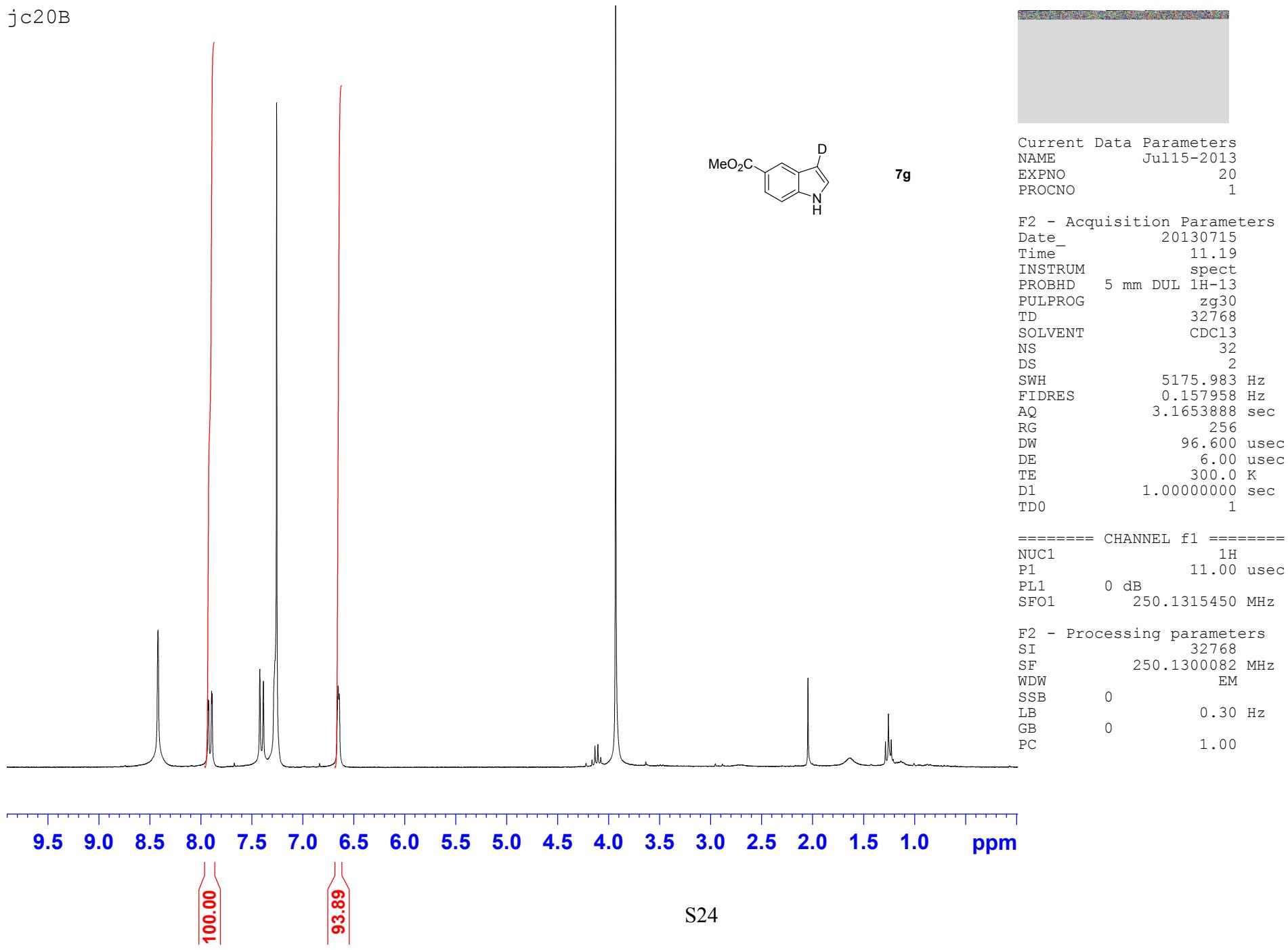
Current Data Parameters
NAME 03022015
EXPNO 40
PROCNO 1

F2 - Acquisition Parameters
Date_ 20150302
Time_ 15.47
INSTRUM spect
PROBHD 5 mm DUL 1H-13
PULPROG zg30
TD 32768
SOLVENT CDCl₃
NS 32
DS 2
SWH 5175.983 Hz
FIDRES 0.157958 Hz
AQ 3.1653888 sec
RG 228.1
DW 96.600 usec
DE 6.00 usec
TE 300.0 K
D1 1.0000000 sec
TDO 1

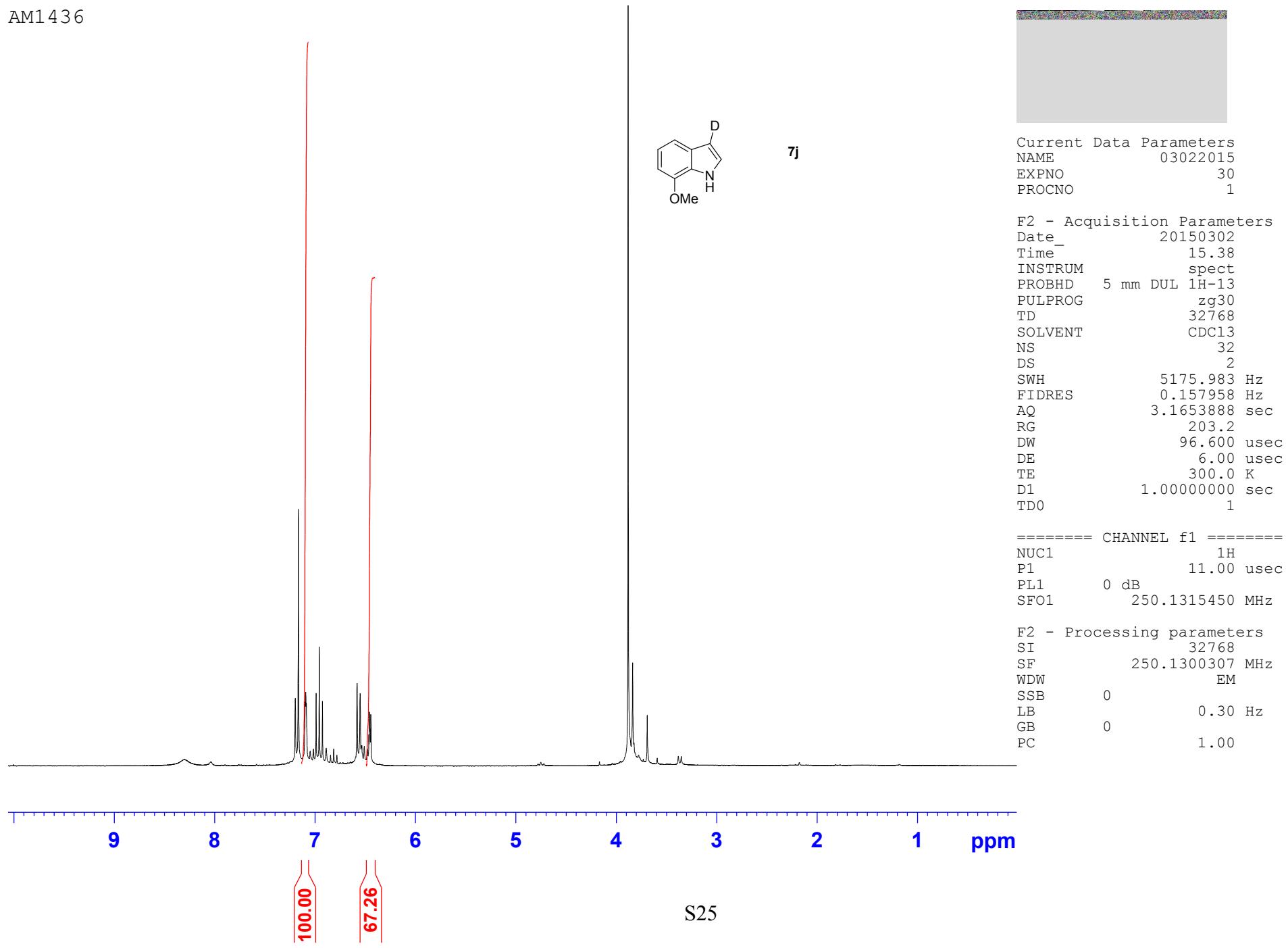
===== CHANNEL f1 =====
NUC1 1H
P1 11.00 usec
PL1 0 dB
SFO1 250.1315450 MHz

F2 - Processing parameters
SI 32768
SF 250.1300286 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

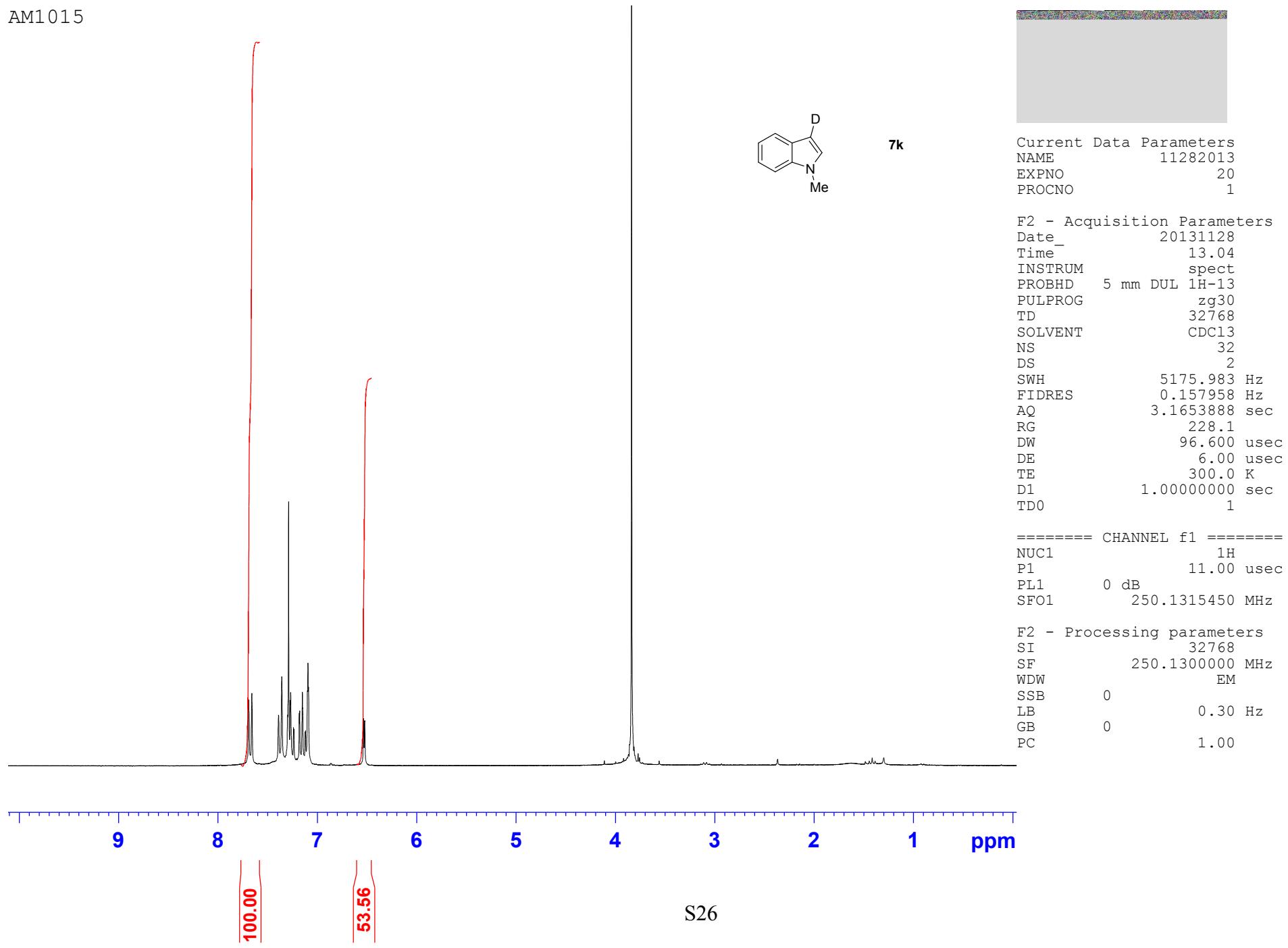
jc20B



AM1436



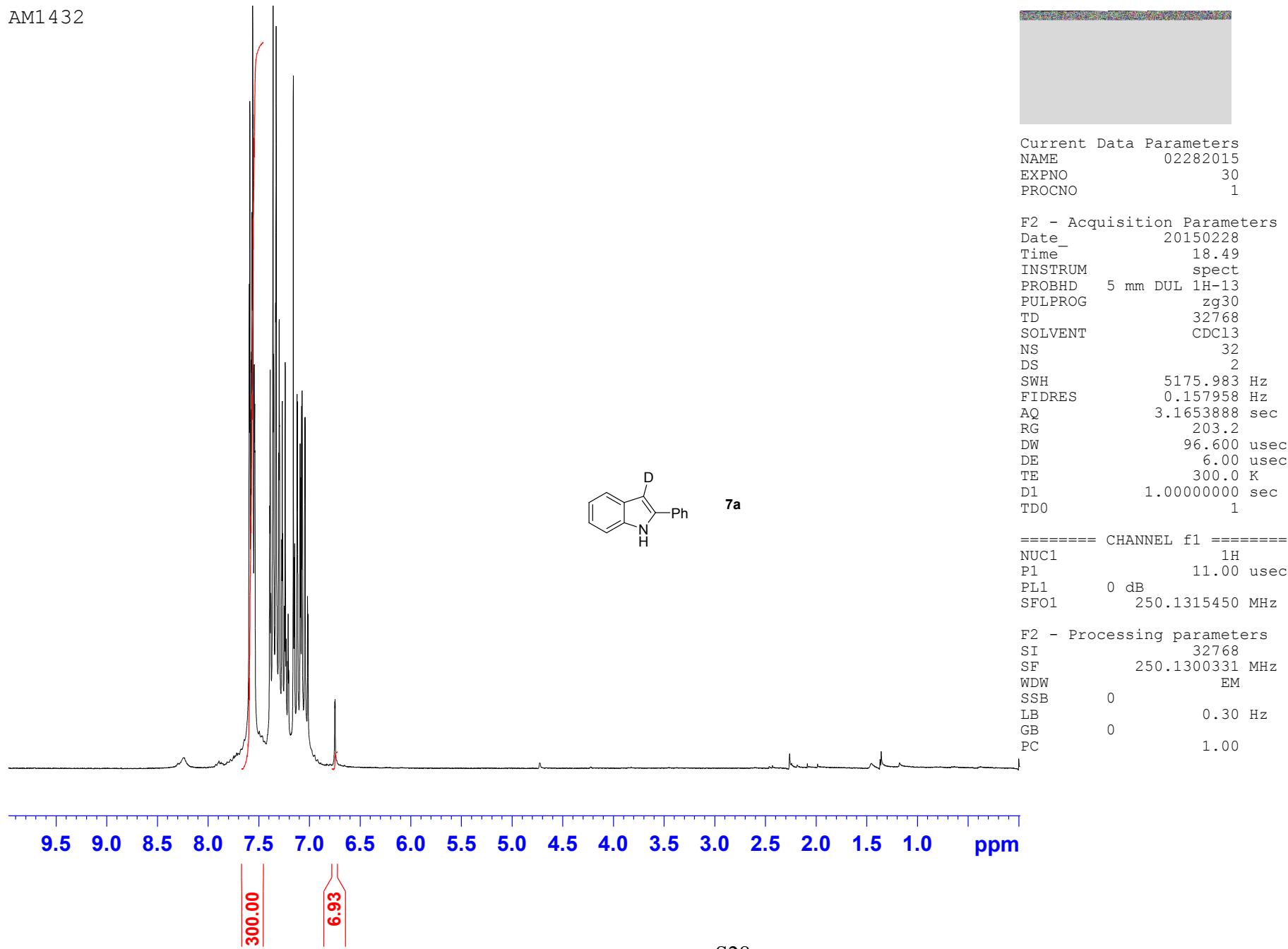
AM1015



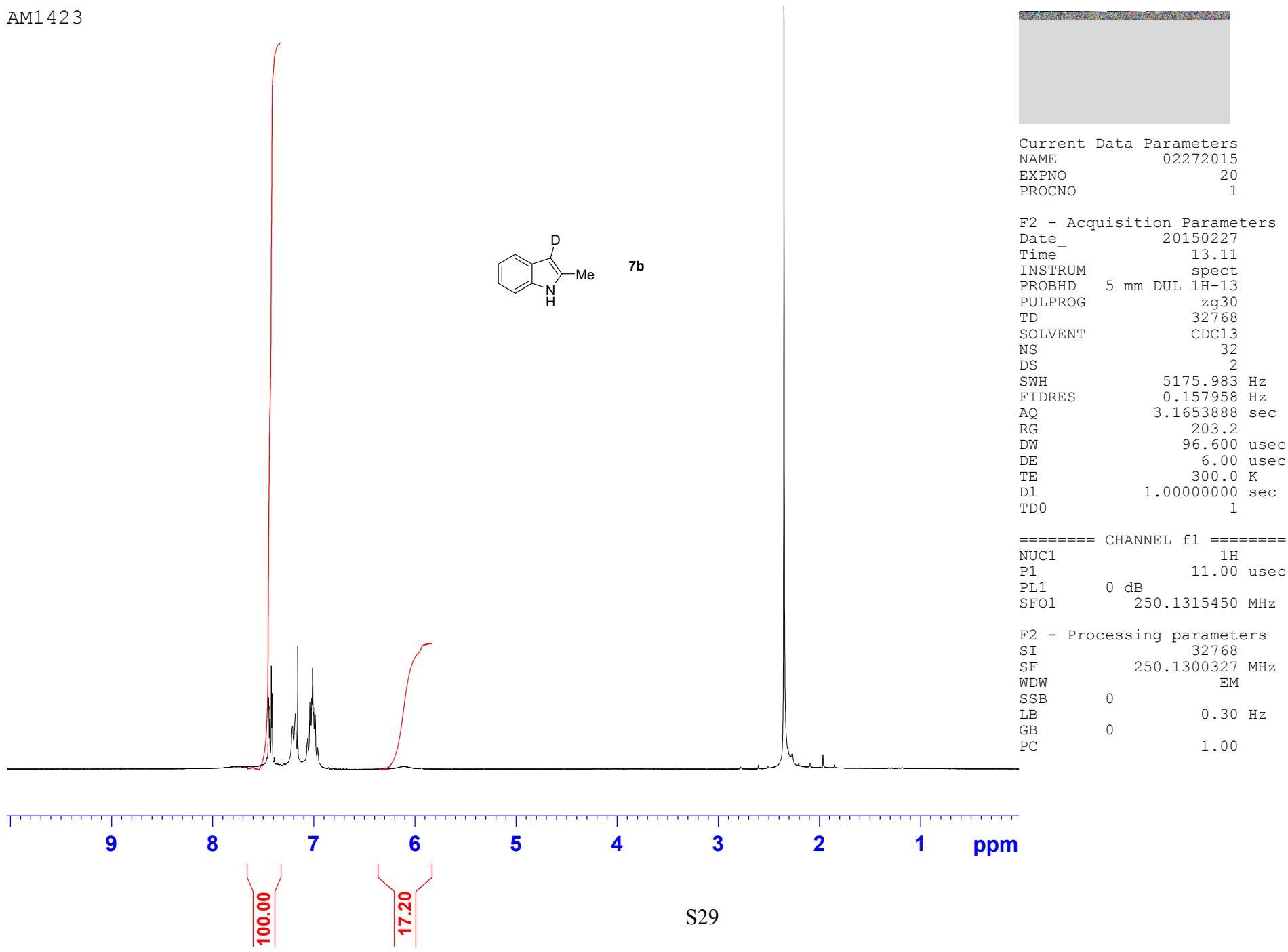
S26

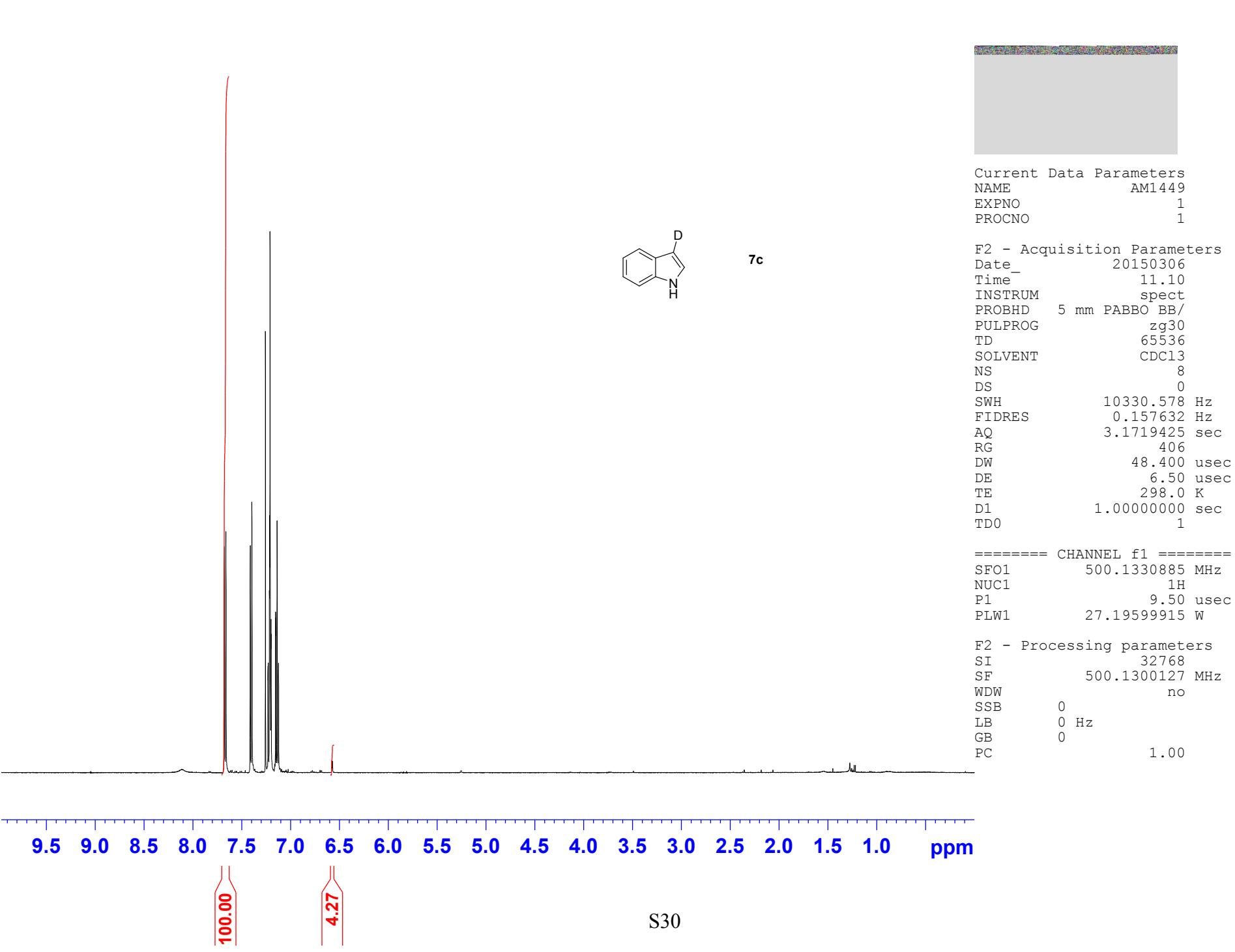
METHOD B

AM1432



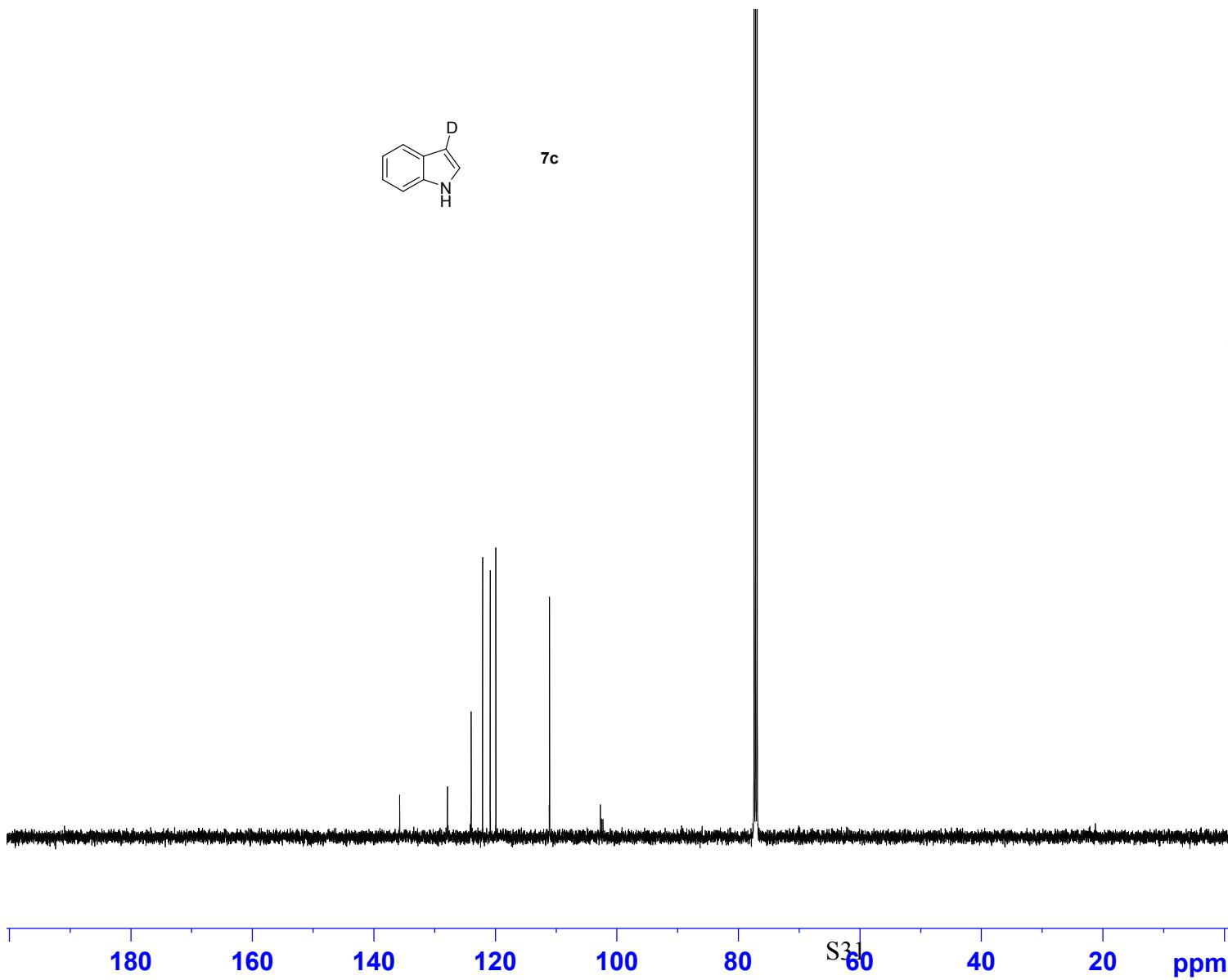
S28







7c



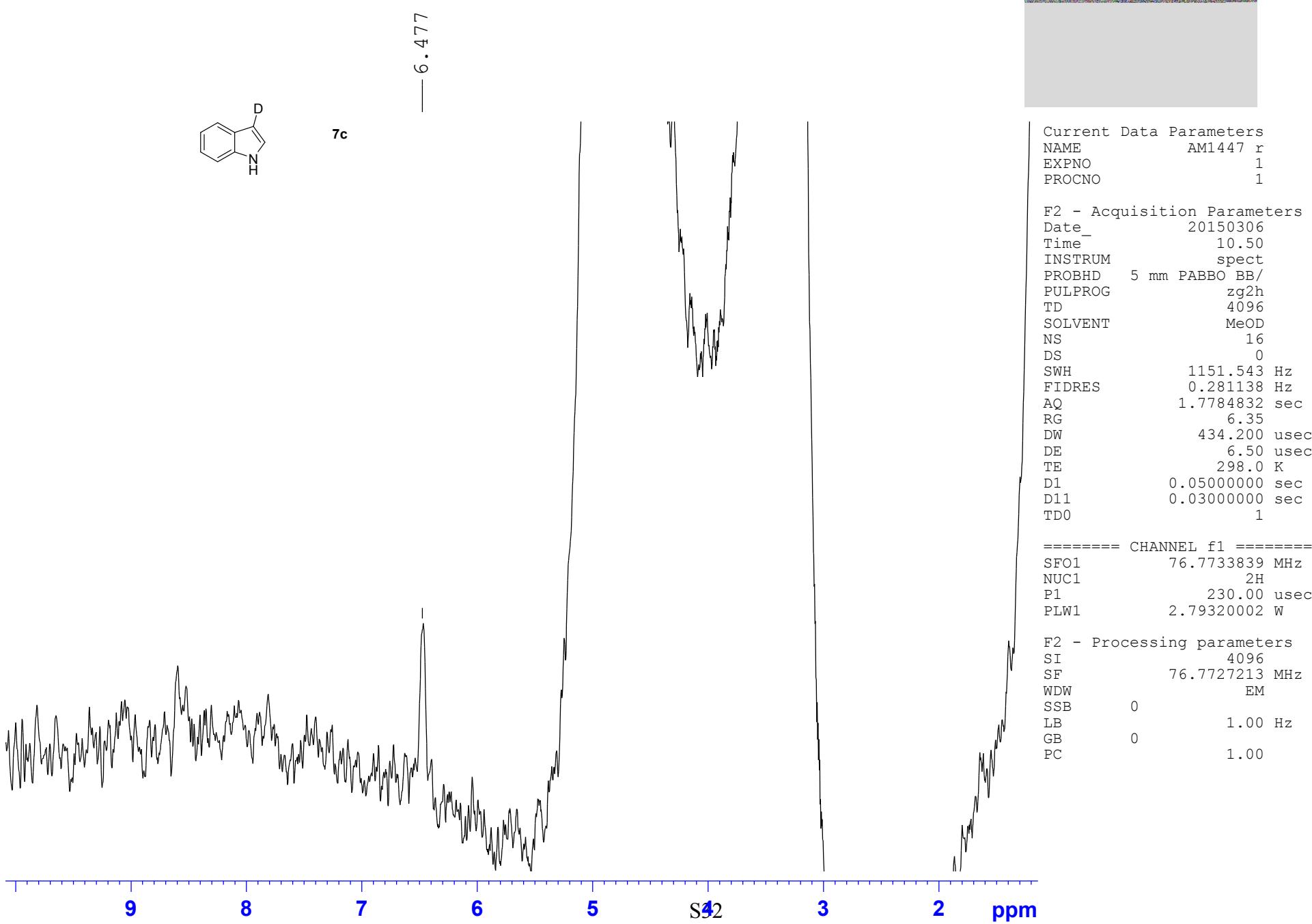
Current Data Parameters
NAME AM1449
EXPNO 2
PROCNO 1

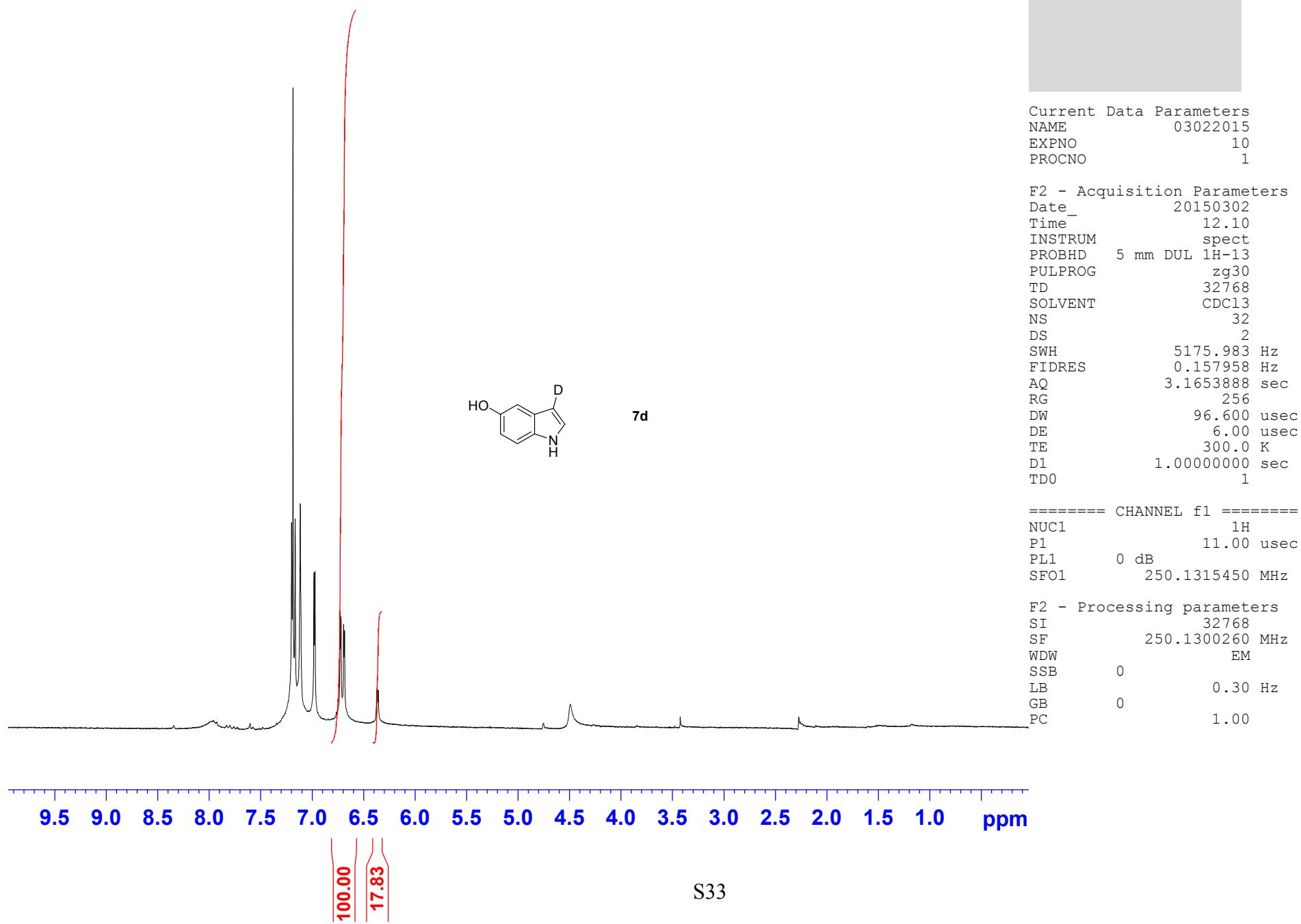
F2 - Acquisition Parameters
Date_ 20150306
Time 11.35
INSTRUM spect
PROBHD 5 mm PABBO BB/
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 204
DS 4
SWH 29761.904 Hz
FIDRES 0.454131 Hz
AQ 1.1010048 sec
RG 1820
DW 16.800 usec
DE 6.50 usec
TE 298.0 K
D1 5.0000000 sec
D11 0.03000000 sec
TD0 1

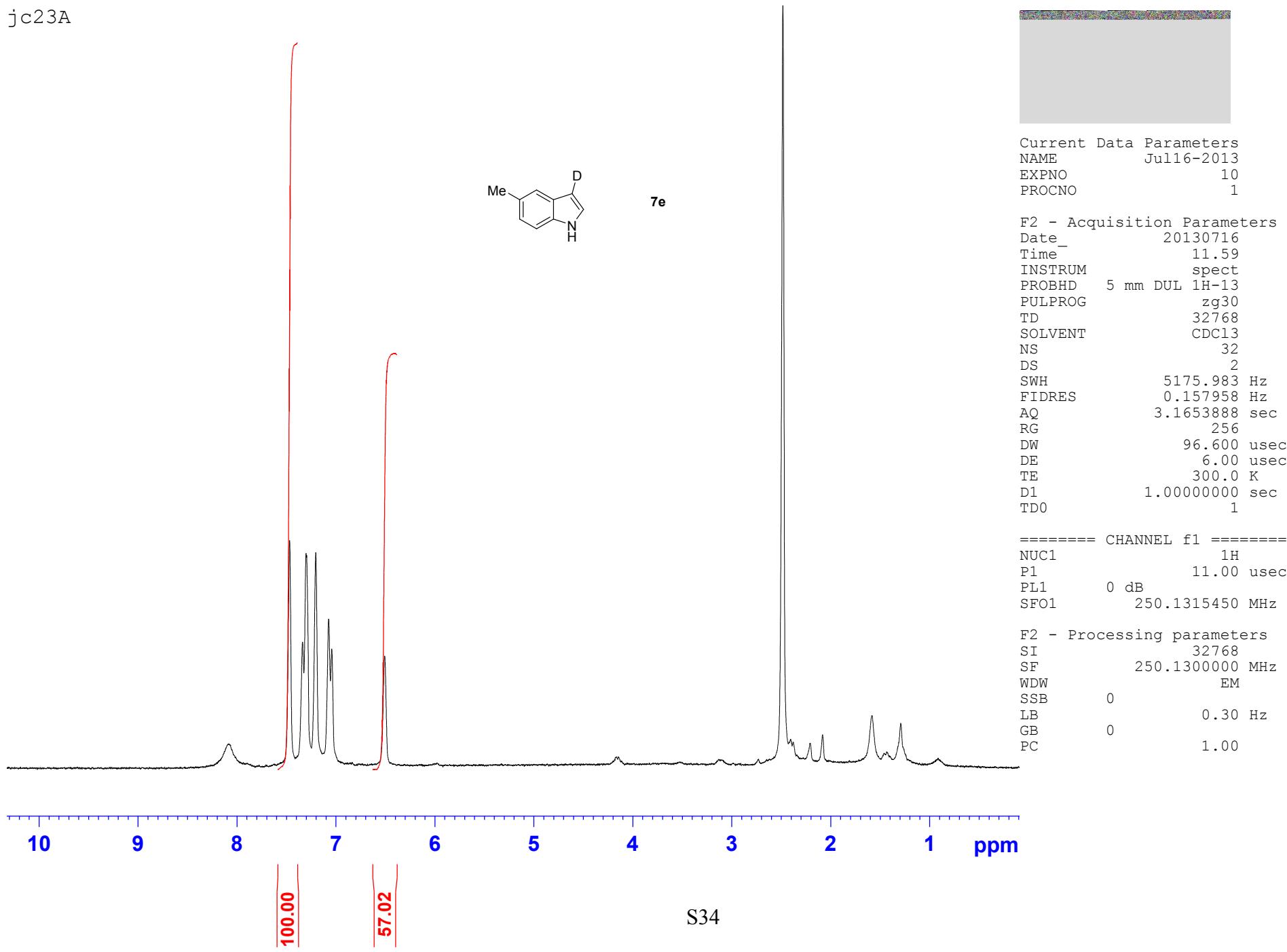
===== CHANNEL f1 =====
SFO1 125.7703643 MHz
NUC1 13C
P1 7.50 usec
PLW1 63.66600037 W

===== CHANNEL f2 =====
SFO2 500.1320005 MHz
NUC2 1H
CPDPRG[2] waltz16
PCPD2 80.00 usec
PLW2 27.19599915 W
PLW12 0.34419000 W
PLW13 0.22028001 W

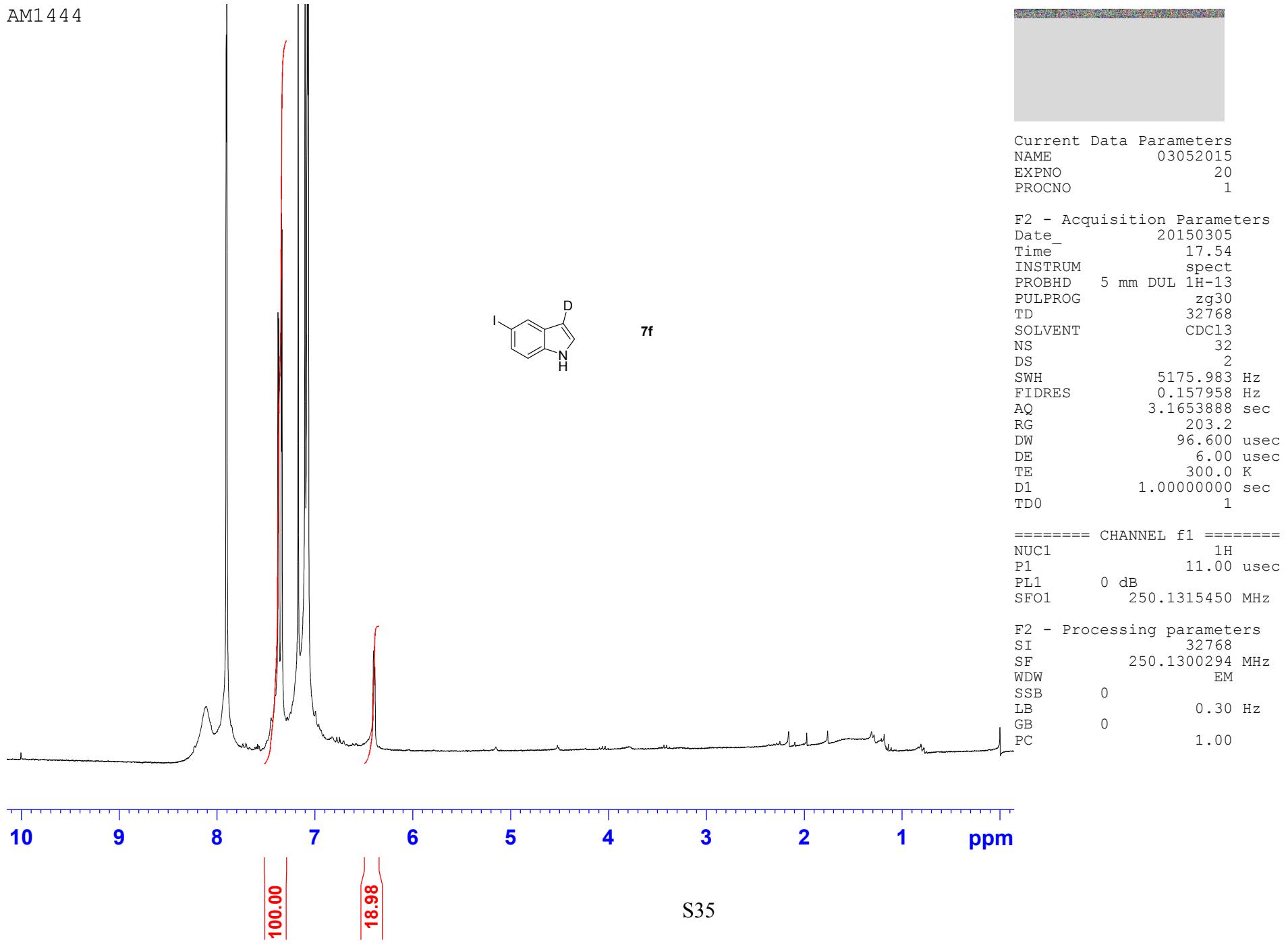
F2 - Processing parameters
SI 32768
SF 125.7577752 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



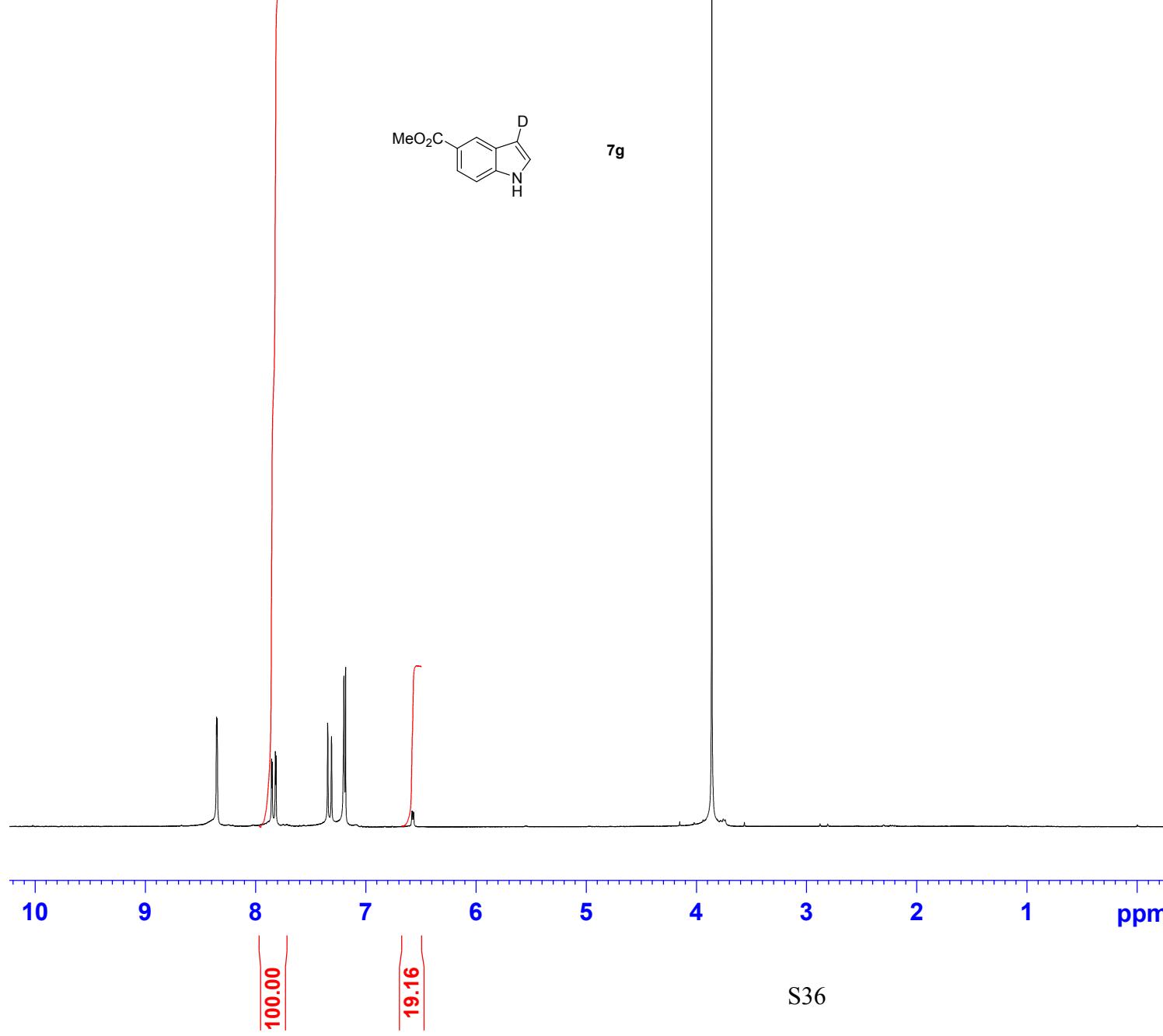




AM1444



AM1428



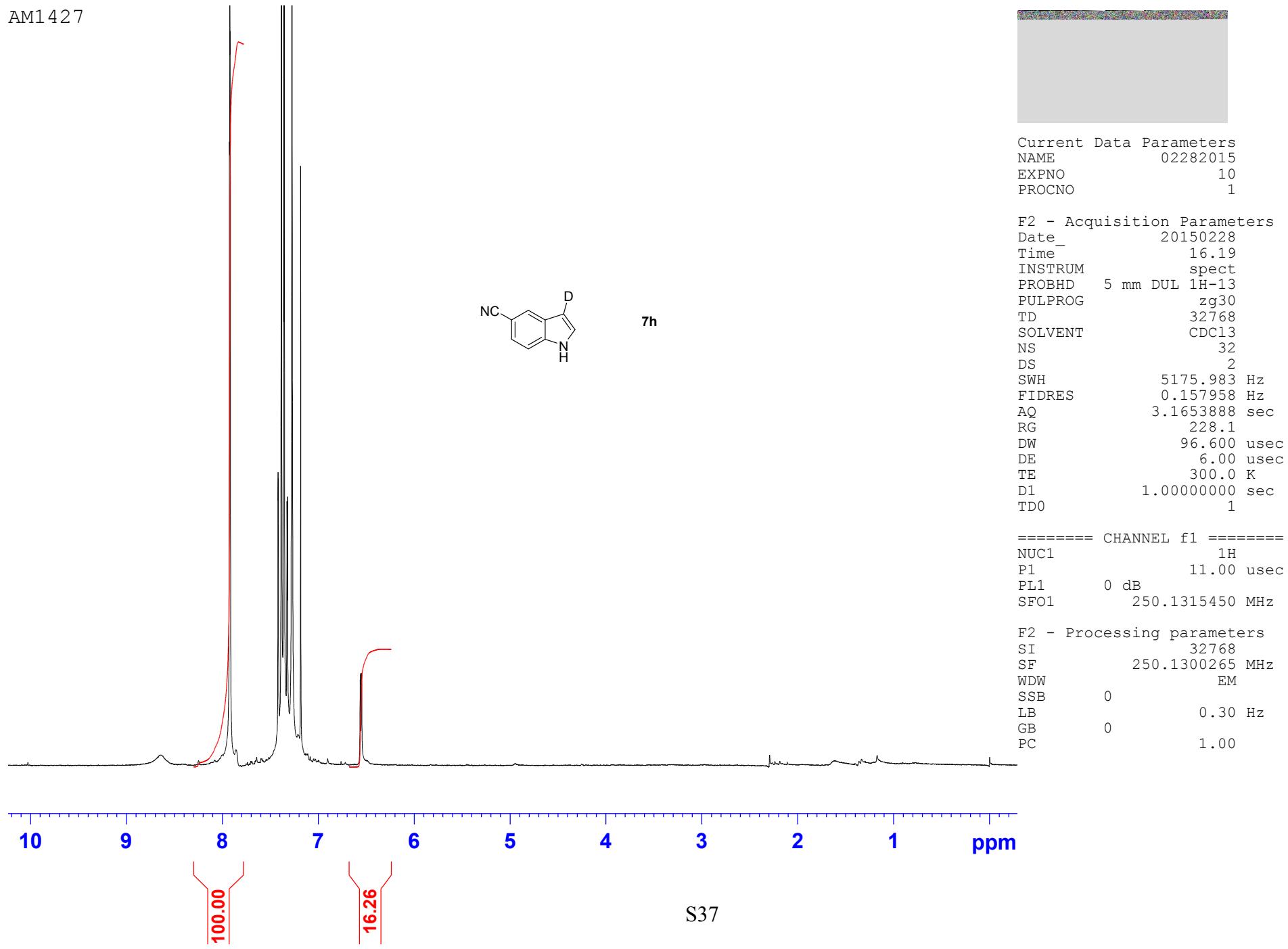
Current Data Parameters
NAME 02282015
EXPNO 20
PROCNO 1

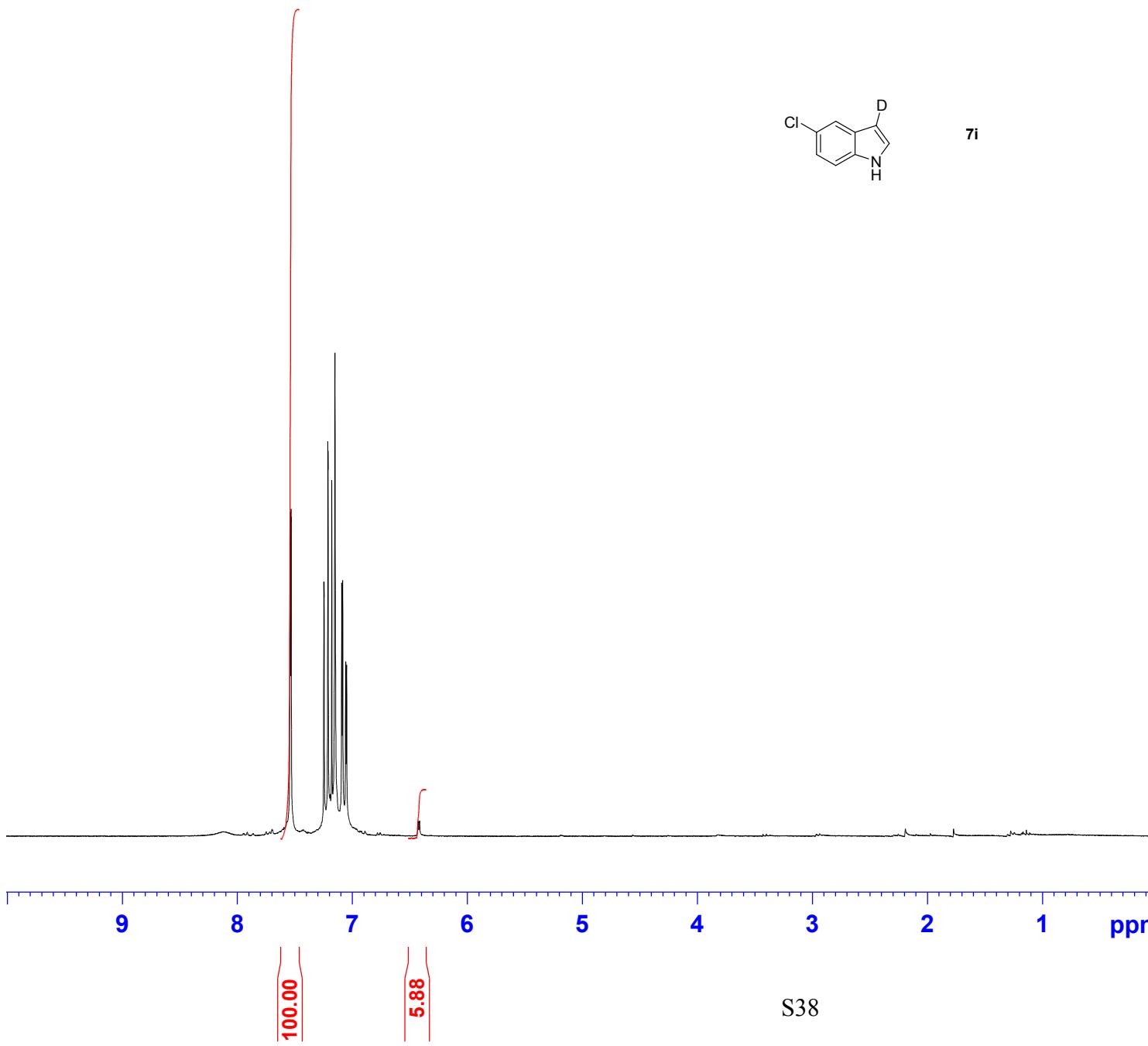
F2 - Acquisition Parameters
Date 20150228
Time 16.27
INSTRUM spect
PROBHD 5 mm DUL 1H-13
PULPROG zg30
TD 32768
SOLVENT CDCl₃
NS 32
DS 2
SWH 5175.983 Hz
FIDRES 0.157958 Hz
AQ 3.1653888 sec
RG 228.1
DW 96.600 usec
DE 6.00 usec
TE 300.0 K
D1 1.00000000 sec
TDO 1

===== CHANNEL f1 =====
NUC1 1H
P1 11.00 usec
PL1 0 dB
SFO1 250.1315450 MHz

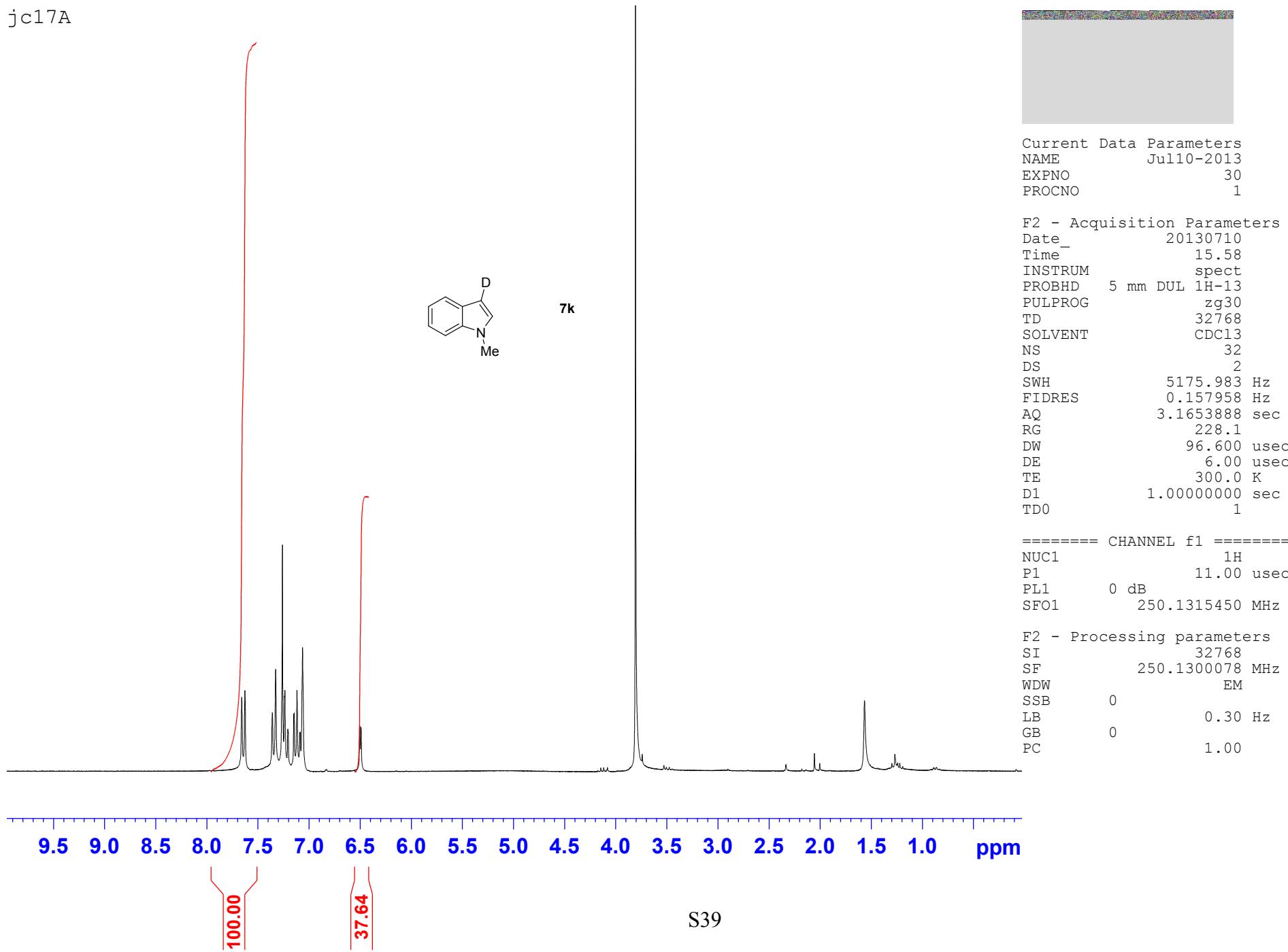
F2 - Processing parameters
SI 32768
SF 250.1300266 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

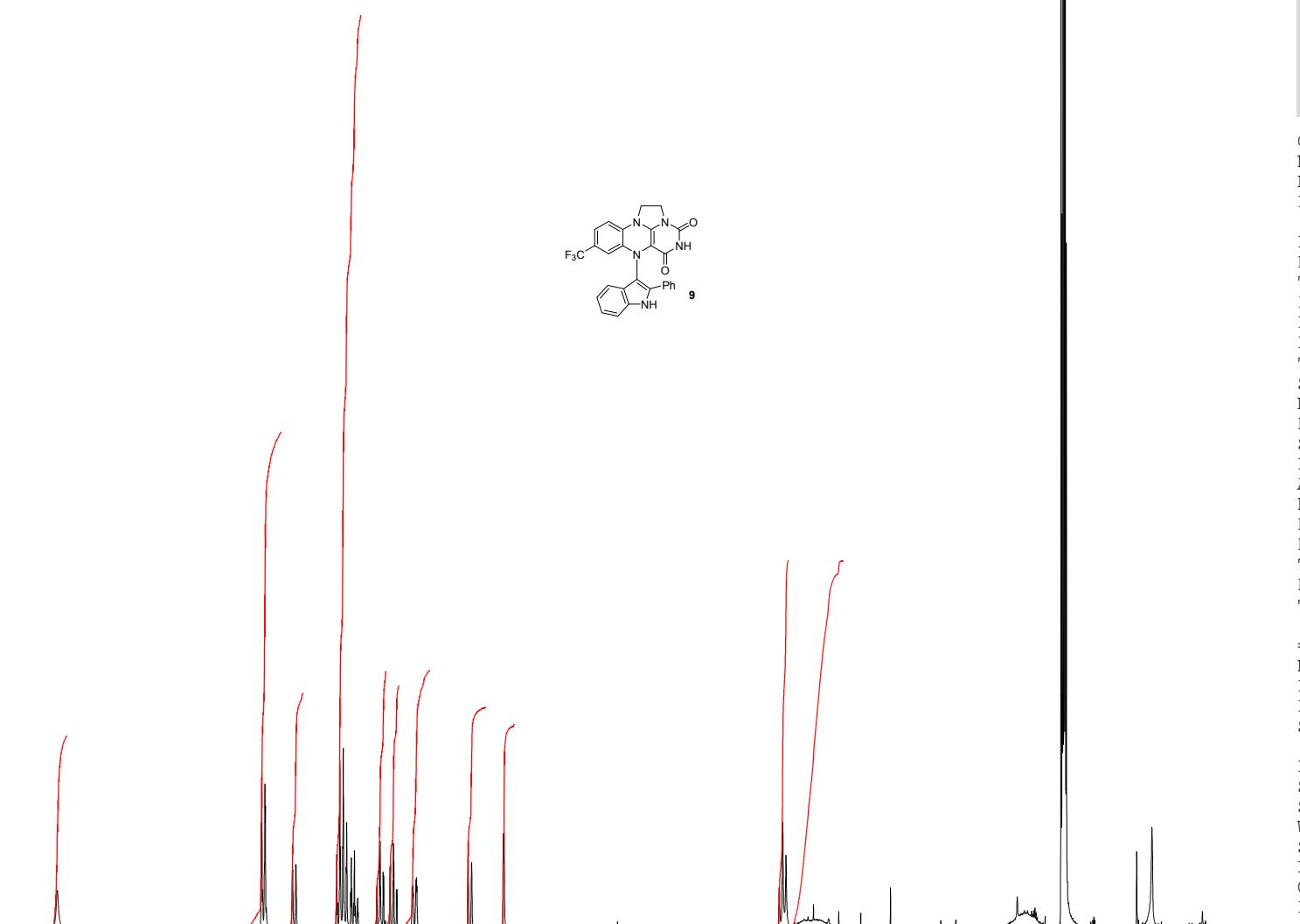
AM1427





jc17A





Current Data Parameters

NAME Mar25-2015
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters

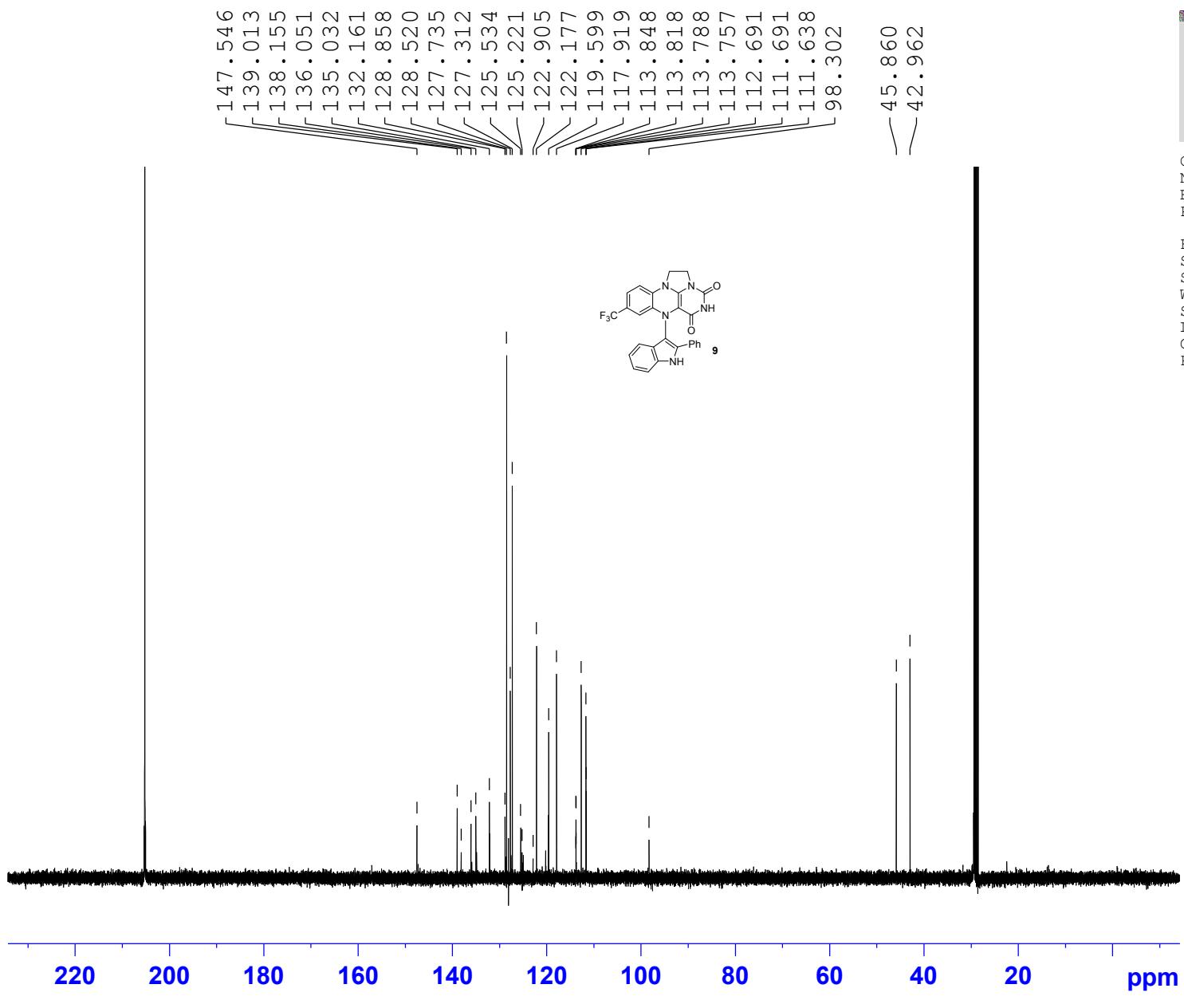
Date 20150325
Time 17.55
INSTRUM av300
PROBHD 5 mm BBO BB-1H
PULPROG zg30
TD 32768
SOLVENT CD3CN
NS 32
DS 0
SWH 6172.839 Hz
FIDRES 0.188380 Hz
AQ 2.6542079 sec
RG 574.7
DW 81.000 usec
DE 6.00 usec
TE 673.2 K
D1 1.0000000 sec
TD0 1

===== CHANNEL f1 =====

NUC1 1H
P1 9.40 usec
PL1 -1.50 dB
SFO1 300.2218540 MHz

F2 - Processing parameters

SI 32768
SF 300.2200023 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



Current Data Parameters
NAME AM1460.2 CARBON_01.fid
EXPNO 1
PROCNO 1

F2 - Processing parameters
SI 65536
SF 125.7422525 MHz
WDW no
SSB 0
LB 0 Hz
GB 0
PC 1.00

AM1410-2
25 March 2015
19F, Bruker 500

