Supporting Information

6-Bromo-7-hydroxy-3-methylcoumarin (mBhc) is an efficient multi-photon labile protecting group for thiol caging and three-dimensional chemical patterning

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Figure S1. Computational analysis of the conformational energies of Bhc-SCH₃ and mBhc-SCH₃ showing the lowest energy conformer in each case, determined using Gaussian. A plot of energy versus dihedral angle (for the exocyclic thioether bond) is shown below. The conformations shown above in line angle form have dihedral angles for the C3-C4-C4'-S bond of 0° while the conformations shown in ball and stick form have dihedral angles for the C3-C4-C4'-S bond of 90° (lowest energy conformers).



Figure S2. LC-MS analysis of a photolysis of **18** to probe for the formation of a disulfide-linked dimer. (A) EIC chromatogram (m/z = 647.45, calcd for $[M + 2H]^{2+}$ = 647.39) of a 7.5 µM solution of **18** after 60 s irradiation at 365 nm, and (B) is the corresponding mass spectrum. 0.5 mass unit difference between the peaks clearly shows the observed ion has a +2 charge thus corresponds to the free thiol containing peptide with the expected mass and not the disulfide. (C) and (D) are EIC chromatograms corresponding to the $[M+2H]^{2+}$ and $[M+3H]^{3+}$ ions of the possible formed disulfide. This data clearly shows the absence of any significant disulfide bond formation in the photolysis reaction.



Figure S3. Analysis of the photolysis of mBhc-NBD-Cysteamine. (A) Scheme showing photo-triggered uncaging of mBhc-protected cysteamine. (B) fluorescence HPLC traces representing time-course photolysis of mBhc-protected cysteamine. These results show clean conversion of the caged peptide to the free thiol with no appearance of any unexpected byproducts. Each peak was isolated and analyzed by ESI-MS confirming that the observed masses were in agreement with the calculated values. mBhc-OH produced in the photolysis reaction was also purified by preparative RP-HPLC and characterized by LC-MS and NMR (See Figure S5).



Figure S4. Analysis of the photolysis of a 5-Fam-labeled peptide containing a mBhc-protected cysteine. (A) Scheme showing photo-triggered uncaging of mBhc-protected 5-Fam labeled K-Ras peptide. (B) fluorescence HPLC traces representing the time-course of photolysis of 5-Fam-KKKSKTKC(mBhc)VIM. These results show clean conversion of caged peptide to the free compound with minimal byproduct formation. Each peak was isolated and analyzed by ESI-MS confirming that the observed masses were completely in agreement with the calculated values.



Figure S5. Analysis of a photolysis reaction of **18** showing the formation of the uncaged product and its subsequent enzymatic prenylation by PFTase. (A) EIC chromatogram (m/z = 520.57, calcd for $[M + 3H]^{3+}$ = 520.58) of a 7.5 μ M solution of **18** in a prenylation buffer containing PFTase with no irradiation (B) EIC chromatogram (m/z = 647.38, calcd for $[M + 2H]^{2+}$ = 647.39) of a 7.5 μ M solution of **18** after 5 min irradiation at 800 nm in prenylation buffer without any PFTase, indicates the generation of free peptide **19**, (C) EIC chromatogram (m/z = 499.97, calcd for $[M + 3H]^{3+}$ = 499.99) of a 7.5 μ M of **18** after 5 min irradiation at 800 nm in presence of PFTase indicates the formation of farnesylated peptide **19**.



Figure S6. Characterization of mBhc-OH generated in the photolysis reaction and isolated via preparatory RP-HPLC. (A) EIC chromatogram showing high purity of the sample and confirming the observed mass exactly matches with the calculated one $([M+H]^+$ cacd,obs = 284.978). (B) ¹HNMR of the isolated mBhc-OH obtained in deuterated acetone.











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ESI-MS data:



HR-MS of Compound 3a [M]¹⁻



HR-MS of Compound 16 [M+Na]⁺



HR-MS of Compound 17 [M+Na]⁺



HR-MS of Compound **18** [**M+2H**]⁺



HR-MS of Compound 5-Fam-KKKSKTKC(mBhc)VIM [M+2H]⁺



HR-MS of Compound mBhc-NBD-cysteamine [M+K]⁺

