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## A Facile hybrid 'flow and batch' access to substituted 3,4-dihydro-2*H*-benzo[*b*][1,4]oxazinones

Andrew J. S. Lin,<sup>a</sup> Cecilia Russell,<sup>a</sup> Jennifer R. Baker,<sup>a</sup> Shelby L. Frailey<sup>a,b</sup> Jennette A Sakoff<sup>c</sup> and Adam McCluskey<sup>a\*</sup>

**Electronic supporting information** 

## Contents

S2	Typical flow che	emistry synthetic detail
S3 -	– S63	<sup>1</sup> H and <sup>13</sup> C NMR spectra for $5 - 11$ , $12a-I$ , $13a-13I$ .
S64	– S93	Mass spectrum for products $5 - 11$ , $12a-l$ , $13a-13l$ .
S94	Ļ	Cytotoxicity activity result.

## N-Ethyl-4-hydroxy-3-nitrobenzamide (6)

Using a two pump equipped Vapourtec R4 system two separate reservoirs were charged with (a) a solution of 4-hydroxy-3benzoate (4, 0.92 mg, 5.0 mmol), COMU (2.6 g, 6.05 mmol) and iPr<sub>2</sub>NEt (1.76 mL, 10.1 mmol) in CH<sub>3</sub>CN (25 mL); and (b) a solution ethylamine (0.28 mL, 10.1 mmol of a 70% aqueous solution) in DMF (15 mL). These solutions were placed on the reagent shelf and the two pumps promised and set to flow at 0.5 mL.min<sup>-1</sup> (for a total flow rate of 1.0 mL.min<sup>-1</sup>). A single 10 mL PFA coil reactor was fitted to the Vapourtec R4 and the temperature equilibrated at 60 °C. Once equilibrated, pumping of both solutions commenced at a combined flow rate of 1 mL.min<sup>-1</sup> (residence time: 10 min) with a backpressure of 5 bar. Reagent pumping was continued, in this instance, for 30 minutes. The resulting product stream was adsorbed on SiO<sub>2</sub> and, then, subjected to column chromatography (SiO<sub>2</sub>) with a gradual elution of 0:1 to 1:0 v/v EtOAc/n-Hexane.

Concentration of the relevant fractions ( $R_f$ : 0.3 – 1:1 v/v EtOAc/n-Hexane) afforded a yellow solid (340 mg, 32 %). m.p. = 122–124°C;

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  10.75 (s, 1H), 8.51 (d, *J* = 2.2 Hz, 1H), 8.06 (dd, *J* = 8.8, 2.2 Hz, 1H), 7.22 (d, *J* = 8.8 Hz, 1H), 6.17 (s, 1H), 3.55 - 3.48 (m, 2H), 1.28 (t, *J* = 7.3 Hz, 3H);

<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 164.7, 157.2, 136.3, 133.1, 127.3, 123.8, 120.6, 35.4, 15.0;

Mass spectrum (ESI, +ve) *m*/z 211 [(M+H)+, 100%];

HRMS (ESI) calcd. for  $C_9H_{11}N_2O_4$  (M+H) 211.0713, found 211.0714;

 $IR \, \nu_{max} \, (cm^{-1}) \, 3301, \, 3279, \, 2996, \, 1626, \, 1615, \, 1520, \, 1418, \, 1250, \, 1240, \, 1162, \, 1076, \, 756, \, 658, \, 618, \, 582, \, 487, \, 425.$ 

## Ethyl 3-amino-4-hydroxybenzoate (7)

A single reagent bottle was charged with ethyl 4-hydroxy-3-nitrobenzoate (5, 130 mg, 0.62 mmol) in CH<sub>3</sub>CN (12 mL). This was positioned at the inlet end of the ThalesNano H-Cube Pro<sup>®</sup> pumping system. A single 30 mm 10% Pd/C CatCart<sup>®</sup> catalyst installed in line and the flow rate set to 3 mL.min<sup>-1</sup> (residence time: 4 min) with the catalyst bed equilibrated at 50 °C and the hydrogen pressure set at 50 bar. Once equilibrated, pumping was the CH<sub>3</sub>CN/5 solution commenced at 3 mL.min<sup>-1</sup> until the reagent reservoir was emptied. The resulting product stream was concentrated under reduced pressure to afforded a creamy white solid (110 mg, 98 %). m.p. 65.5–67.8 °C, R<sub>f</sub> 0.15 (1:9 v/v CH<sub>3</sub>OH/CH<sub>2</sub>Cl<sub>2</sub>);

<sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) δ 7.46 (d, *J* = 1.8 Hz, 1H), 7.43 (dd, *J* = 8.2, 1.8 Hz, 1H), 6.75 (d, *J* = 8.2 Hz, 1H), 4.32 (q, *J* = 7.1 Hz, 2H), 3.74 (broad s, 2H), 1.37 (t, *J* = 7.1 Hz, 3H);

<sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>) δ 167.2, 149.0, 134.4, 422.7, 122.0, 117.5, 114.5, 60.8, 14.4;

Mass spectrum (ESI, +ve) m/ $\chi$  265 [(M+iPrOH+Na+H)+, 100%], 182 [(M+H)+, 100%], (ESI, -ve) m/ $\chi$  180 [(M–H)-, 100%]; HRMS (ESI) calcd. for C<sub>9</sub>H<sub>10</sub>NO<sub>3</sub> (M–H)<sup>-</sup> 181.1885, found 181.0499; IR v<sub>max</sub> (cm<sup>-1</sup>) 3386, 3100, 2985, 1687, 1602, 1520, 1365, 1302, 1285, 1202, 1151, 1094, 1022, 893, 764, 637, 453.

<sup>1</sup>H and <sup>13</sup>C NMR spectra of product 5





<sup>1</sup>H and <sup>13</sup>C NMR spectra of product 6





 $^1\mathrm{H}$  and  $^{13}\mathrm{C}$  NMR spectra of product 7



7



<sup>1</sup>H and <sup>13</sup>C NMR spectra of product 8



9



<sup>1</sup>H and <sup>13</sup>C NMR spectra of product **10** 





<sup>1</sup>H and <sup>13</sup>C NMR spectra of product **12a** 











<sup>1</sup>H and <sup>13</sup>C NMR spectra of product **12c** 











<sup>1</sup>H and <sup>13</sup>C NMR spectra of product **12e** 





<sup>1</sup>H and <sup>13</sup>C NMR spectra of product **12f** 





<sup>1</sup>H and <sup>13</sup>C NMR spectra of product **12g** 





<sup>1</sup>H and <sup>13</sup>C NMR spectra of product **12h** 









<sup>1</sup>H and <sup>13</sup>C NMR spectra of product **12j** 












<sup>1</sup>H and <sup>13</sup>C NMR spectra of product 11





<sup>1</sup>H and <sup>13</sup>C NMR spectra of product 13a





<sup>1</sup>H and <sup>13</sup>C NMR spectra of product **13b** 





 $^1\text{H}$  and  $^{13}\text{C}$  NMR spectra of product 13c





 $^1\mathrm{H}$  and  $^{13}\mathrm{C}$  NMR spectra of product 13d





<sup>1</sup>H and <sup>13</sup>C NMR spectra of product **13e** 





 $^1\mathrm{H}$  and  $^{13}\mathrm{C}$  NMR spectra of product 13f





<sup>1</sup>H and <sup>13</sup>C NMR spectra of product **13g** 













<sup>1</sup>H and <sup>13</sup>C NMR spectra of product 13j







60









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Page 3 of 3

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Page 3 of 3

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Page 3 of 3

### Mass spectrum of product 13c

### **LCMS Report**



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Page 3 of 3

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### Cytotoxicity Assay Results

All test agents were prepared as stock solutions (30 mM) in DMSO and stored at -20°C. Cell lines used in the study included HT29 (colorectal carcinoma); U87, SJ-G2, SMA (glioblastoma); MCF-7 (breast carcinoma); A2780 (ovarian carcinoma); H460 (lung carcinoma); A431 (skin carcinoma); Du145 (prostate carcinoma); BE2-C (neuroblastoma); and MiaPaCa-2 (pancreatic carcinoma) together with one non-tumour derived normal breast cell line (MCF10A). All cell lines were incubated in a humidified atmosphere 5% CO2 at 37°C. The cancer cell lines were maintained in Dulbecco's modified Eagle's medium (DMEM; Trace Biosciences, Australia) supplemented with foetal bovine serum (10 %), sodium bicarbonate (10 mM), penicillin (100 IUmL<sup>-1</sup>), streptomycin (100 mgmL<sup>-1</sup>), and glutamine (4 mM). The non-cancer MCF10A cell line was maintained in DMEM:F12 (1:1) cell culture media, 5% heat inactivated horse serum, supplemented with penicillin (50 IUmL<sup>-1</sup>), streptomycin (50 mgmL<sup>-1</sup>), HEPES (20 mm), l-glutamine (2 mM), epidermal growth factor (20 ngmL<sup>-1</sup>), hydrocortisone (500 ngmL<sup>-1</sup>), cholera toxin (100 ngmL<sup>-1</sup>), and insulin (10 mgmL<sup>-1</sup>). Cytotoxicity was determined by plating cells in duplicate in medium (100 mL) at a density of 2500-4000 cells per well in 96-well plates. On day 0, when the cells were in logarithmic growth (24h after plating), medium (100 mL) with or without the test agent was added to each well. After 72h drug exposure, growth inhibitory effects were evaluated using the MTT (3-[4,5-dimethyltiazol-2yl]-2,5-diphenyl-tetrazolium bromide) assay and absorbance read at 540 nm. All analogues were initially tested for percentage growth inhibition at a fixed concentration of  $25\mu$ M. Those that showed appreciable inhibition underwent further examination to produce an eight-point dose-response curve from which a  $GI_{50}$  value was calculated, representing the drug concentration that inhibited cell growth by 50% based on the difference between the optical density values on day 0 and those at the end of drug exposure. All assays were conducted in duplicate and replicated on at least three separate occasions. Values represent the mean  $\pm$  standard error of the mean.

Table 1. Initial cytotoxicity screening of analogues 10, 11, 12a–12l, 13a–13l. Values given are percentage inhibition at 25 µM drug concentration.												
Compound	HT29	U87	MCF-7	A2780	H460	A431	Du145	BE2-C	SJ-G2	MIA	SMA	MCF10A
5	$11 \pm 10$	$21 \pm 1$	$27 \pm 4$	$70 \pm 0$	$26 \pm 3$	$21 \pm 3$	$17 \pm 4$	$74 \pm 5$	$25 \pm 3$	$37 \pm 4$	$31 \pm 5$	$16 \pm 6$
				85 ±								
12a	$23 \pm 8$	$35 \pm 1$	$32 \pm 4$	1.2	$50 \pm 2$	$25 \pm 1$	$30 \pm 4$	$82 \pm 4$	$31 \pm 1$	$64 \pm 3$	$54 \pm 3$	$33 \pm 7$
									$100 \pm$			
12b	$82 \pm 2$	$43 \pm 1$	$42 \pm 13$	$88 \pm 1$	$68 \pm 1$	>100	$59 \pm 2$	$96 \pm 3$	4	$68 \pm 2$	$72 \pm 1$	$73 \pm 6$
		10.0	10.0						51 ±			
12c	$20 \pm 2$	$18 \pm 3$	$13 \pm 6$	$84 \pm 2$	$26 \pm 6$	$24 \pm 4$	$26 \pm 1$	$55 \pm 7$	12	$33 \pm 1$	$58 \pm 2$	$28 \pm 3$
124	$7 \pm 4$	12 + 2	57 + 4	42 + 1	12 + 2	$11 \pm$	$22 \pm 0$	50 + 7	25   7	$24 \pm 2$	20 + 2	12 + 4
120	7 ± 4	$13 \pm 2$ 12 $\pm 1$	$3/\pm 4$ 12 $\pm 6$	$42 \pm 1$ 22 $\pm 1$	$12 \pm 2$ $10 \pm 4$	1.4 $10 \pm 4$	$23 \pm 0$ 21 ± 4	$30 \pm 7$	$33 \pm 7$	$24 \pm 2$ 15 $\pm 2$	$30 \pm 3$	$13 \pm 4$ 7 + 4
120	$5 \pm 0$	$13 \pm 1$	$13 \pm 0$	$23 \pm 1$	19 ± 4	$10 \pm 4$ 28 $\pm$	$21 \pm 4$	44 ± 9	$21 \pm 3$	$13 \pm 2$	$10 \pm 1$	/±4
12f	29 + 2	$26 \pm 1$	19 + 7	42 + 1	25 + 5	$\frac{20 \pm}{35}$	22 + 3	$46 \pm 6$	31 + 5	26 + 2	26 + 3	14 + 4
12g	$58 \pm 2$	$43 \pm 3$	$60 \pm 4$	$\frac{12 \pm 1}{88 \pm 1}$	$67 \pm 2$	>100	$55 \pm 4$	$92 \pm 4$	$92 \pm 5$	$20 \pm 2$ 67 ± 2	$20 \pm 3$ $75 \pm 2$	$69 \pm 6$
					•, =	20 ±			49 ±	•, _	,	
12h	$6 \pm 5$	$23 \pm 2$	$20 \pm 7$	$68 \pm 2$	$24 \pm 5$	3.0	$9 \pm 4$	$50 \pm 3$	10	$22 \pm 2$	$43 \pm 1$	$23 \pm 3$
12i	$34 \pm 2$	$15 \pm 2$	$13 \pm 6$	$42 \pm 2$	$14 \pm 5$	$23 \pm 3$	$22 \pm 1$	$49 \pm 9$	$29 \pm 3$	$28 \pm 2$	$27 \pm 2$	$9 \pm 4$
12j	$71 \pm 2$	$41 \pm 2$	$63 \pm 4$	$85 \pm 1$	$65 \pm 1$	>100	$60 \pm 1$	$99 \pm 3$	$97 \pm 4$	$69 \pm 2$	$76 \pm 1$	$73 \pm 6$
12k	$20 \pm 2$	$20 \pm 2$	$13 \pm 6$	$40 \pm 2$	$10 \pm 6$	$15 \pm 2$	$20 \pm 2$	$57 \pm 10$	$32 \pm 2$	$27 \pm 3$	$22 \pm 3$	$3 \pm 1$
											$100 \pm$	
121	$71 \pm 3$	$27 \pm 6$	ND	$86 \pm 1$	$77 \pm 2$	$93 \pm 1$	$56 \pm 7$	>100	>100	$50 \pm 4$	0	$79 \pm 4$
6	$10 \pm 5$	$15 \pm 1$	$18 \pm 6$	$25 \pm 3$	$12 \pm 2$	$10 \pm 2$	$17 \pm 3$	$7 \pm 5$	$19 \pm 3$	$19 \pm 1$	$19 \pm 4$	$6 \pm 2$
13a	$10 \pm 3$	$18 \pm 1$	$24 \pm 8$	$28 \pm 2$	$15 \pm 2$	$10 \pm 2$	$16 \pm 4$	$15 \pm 5$	$20 \pm 2$	$21 \pm 1$	$16 \pm 3$	9 ± 3
13b	$1 \pm 4$	$10 \pm 3$	$5\pm 5$	$43 \pm 2$	$21 \pm 2$	$10 \pm 0$	$20 \pm 4$	$11 \pm 3$	$25 \pm 9$	$24 \pm 6$	$17 \pm 3$	$24 \pm 3$
12	0.1.2	14 + 2	22 1 6	06 1 2	20 1 2	$20 \pm$	17 . 4	20 1 2	46 1 0	24 + 2	67 · 4	22 + 2
130	$9 \pm 2$	$14 \pm 3$	$22 \pm 6$	$86 \pm 2$	$29 \pm 2$	2.0	$1/\pm 4$	$28 \pm 3$	$46 \pm 8$	$24 \pm 3$	$5/\pm 4$	$33 \pm 3$
130	$5 \pm 5$	$10 \pm 2$	$15 \pm 3$	$39 \pm 3$	$10 \pm 2$	$12 \pm 1$	$20 \pm 0$	$11 \pm /$	$25 \pm 9$	$25 \pm 4$	$18 \pm 3$	$20 \pm 1$
13e 13f	$5 \pm 2$	$11 \pm 0$ $10 \pm 1$	8 ± 0 <0	$1/\pm 4$ 20 ± 3	$10 \pm 2$ 7 + 2	$0 \pm 4.3$ $1 \pm 4$	$8 \pm 3$ 7 + 3	0 + 1	$2 \pm 7$ $22 \pm 2$	$14 \pm 2$ 7 + 2	$0 \pm 4$ 7 + 2	$10 \pm 1$ 1 + 1
151	~0	10 ± 1	<0	$20 \pm 3$	1 ± 2	4 - 4	1 ± 5	$9 \pm 1$	$32 \pm 2$	1 ± 2	1 ± 2	1 - 1
13g	<0	$11 \pm 5$	<0	$57 \pm 5$	$28 \pm 3$	$5 \pm 3.2$	$9\pm5$	$16 \pm 11$	13	$26 \pm 5$	$27 \pm 2$	$21 \pm 5$
	-	-	-					-	33 ±			-
13h	$7 \pm 2$	$9 \pm 3$	$8 \pm 4$	$59 \pm 3$	$28 \pm 2$	$14 \pm 2$	$13 \pm 1$	$9\pm9$	10	$21 \pm 1$	$21 \pm 3$	$27 \pm 3$
13i	$7 \pm 2$	$13 \pm 1$	$19 \pm 7$	$19 \pm 4$	$9 \pm 1$	9 ± 1.9	$17 \pm 3$	$6 \pm 2$	$20 \pm 5$	$17 \pm 1$	$16 \pm 2$	6 ± 3
13j	$17 \pm 6$	$17 \pm 2$	$56 \pm 7$	$31 \pm 2$	$25 \pm 4$	$3\pm3$	$27 \pm 3$	$9\pm 2$	$26 \pm 1$	$24 \pm 3$	$14 \pm 1$	9 ± 5
13k	<0	$13 \pm 2$	$20 \pm 7$	$31 \pm 3$	$10 \pm 2$	$6 \pm 3.1$	$10 \pm 4$	$19 \pm 2$	$23 \pm 1$	$17 \pm 3$	$11 \pm 3$	6 ± 3
121	<0	-0	-0	20 1 2	20 + 4	-0	<0	20 + 6	$14 \pm 10$	-0	51 + 2	20 + 7
131	<0	<0	<0	$38 \pm 2$	$29 \pm 4$	<0	<0	$30 \pm 6$	19	<0	$51 \pm 3$	$30 \pm 7$