Supplementary Information

A versatile post-synthetic method on solid support for the synthesis of RNA containing reduction-responsive modifications

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Figure S1: 121 MHz ³¹P-NMR spectrum (CD₃CN) of 2'-O-AcSM uridine phosphoramidite 1





Figure S2: 300 MHz ¹H-NMR and 75 MHz ¹³C-NMR spectra (CDCl₃) of 2a





Figure S3: 300 MHz ¹H-NMR and 75 MHz ¹³C-NMR spectra (CDCl₃) of 2b



Figure S4: 400 MHz ¹H-NMR and 100 MHz ¹³C-NMR spectra (CDCl₃) of 2c



Figure S5: 400 MHz ¹H-NMR and 100 MHz ¹³C-NMR spectra (CDCl₃) of 2d



Figure S6: 400 MHz ¹H-NMR and 100 MHz ¹³C-NMR spectra (DMSO-d₆) of 2e







Figure S7: 400 MHz ¹H-NMR, 100 MHz ¹³C-NMR and 160 MHz ¹⁹F-NMR spectra (CDCl₃) of **2f**







Figure S8: 400 MHz ¹H-NMR, 100 MHz ¹³C-NMR and 160 MHz ¹⁹F-NMR spectra (DMSO- d_6) of 2g















Figure S12: IEX-HPLC and MALDI-TOF MS analysis of purified RNA 4d



Figure S13: IEX-HPLC and MALDI-TOF MS analysis of purified RNA 4e











Figure S16: Melting curves of RNAs 3 and 4a-g with their complementary strand



Figure S17: CD spectroscopy curves of RNAs 3 and 4a-g with their complementary strand





Figure S18: MALDI-TOF MS spectra of RNAs 3 and 4a-g incubated with SVPDE



Figure S19 : IEX-HPLC and MALDI-TOF Mass spectra of reductive conversion of RNAs **4a** into unmodified RNA **3** after 1h incubation with 5.6 mM glutathione





Figure S20 : IEX-HPLC and MALDI-TOF Mass spectra of reductive conversion of RNAs 4a-g with their complementary strand into unmodified duplex after 1h incubation with 5.6 mM glutathione

2'-O-RSSM RNA duplexes	Purity of products RNAs
4a	94%
4 b	85%
4 c	92%
4d	94%
4e	95%
4f	70%
4g	93%

Table S1 : Purity of the product 2'OH RNAs duplex after reductive conversion of RNAs **4a-g** with their complementary strand into unmodified duplex after 1h incubation with 5.6 mM glutathione