

Screening of Neu5Ac α (2-6)Gal Isomers Preferences of Siglec with Sialic acid Microarray.

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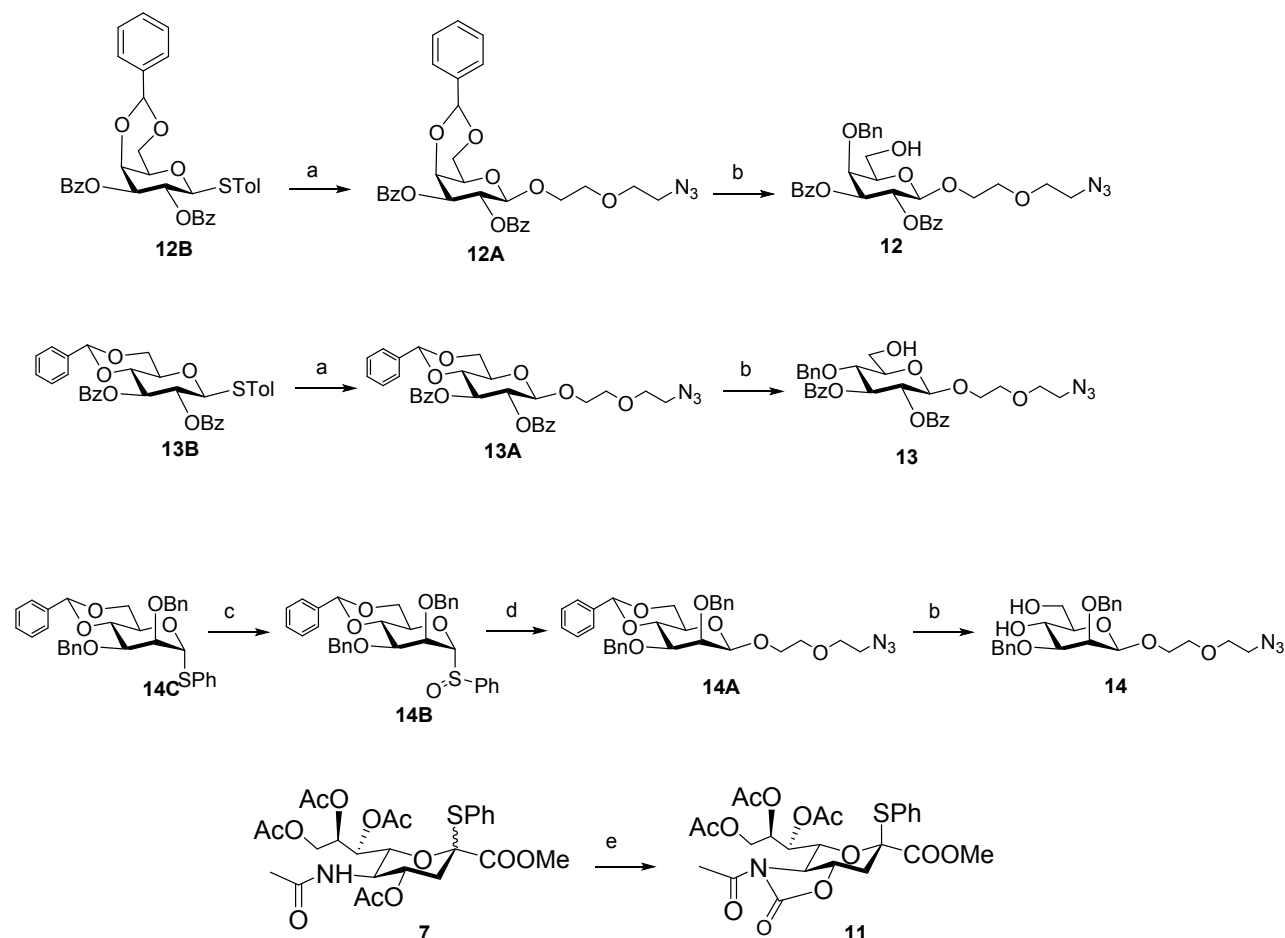
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1. General information. All chemicals were reagent grade and used as supplied except where noted. Analytical thin layer chromatography (TLC) was performed on Merck silica gel 60 F254 plates (0.25 mmol). Compounds were visualized by UV irradiation or dipping the plate in CAM/ninhydrin solution followed by heating. Column chromatography was carried out using force flow of the indicated solvent on Fluka Kiesel gel 60 (230–400 mesh). ¹H and ¹³C NMR spectra were recorded on Jeol 400 MHz, with cryo probe using residual solvents signals as an internal reference (CDCl₃ δ_H , 7.26 ppm, δ_C 77.3 ppm and CD₃OD δ_H 3.31 ppm, δ_C 49.0 ppm). The chemical shifts (δ) are reported in ppm and coupling constants (J) in Hz. UV-visible measurements were performed with Evolution 300 UV-visible spectrophotometer (Thermo Fisher Scientific, USA). Fluorescence spectra were recorded in FluoroMax-4 spectrofluorimeter (Horiba Scientific, U.S.A.).

2. Synthesis of disaccharide library



Scheme S1 : Reagents and conditions: (a) 2-azidoethoxyethanol, NIS/TfOH, -40°C , DCM; (b) PhBCl_2 , Et_3SiH , -78°C , DCM; (c) mCPBA, -78°C , DCM; (d) 2-azidoethoxyethanol, Tf_2O , TTBP, -78°C , DCM; (e) (i) MsOH , MeOH ; (ii) NPCC, NaHCO_3 , $\text{CH}_3\text{CN}:\text{H}_2\text{O}$ (2:1); (iii) AC_2O /Pyridine; (iv) AcCl , DIPEA, DCM.

Synthetic procedure for comp 12A : A solution of **12B** (500 mg, 1.08 mmol, 1.0 equiv), 2-azidoethoxyethanol (1.28 mmol, 1.2 equiv), and activated 4\AA powdered molecular sieves (1.5 gm) in anhydrous dichloromethane (5 ml) was stirred for 2h at room temperature under an argon atmosphere, and then cooled to -40°C followed by addition of NIS (290 mg, 1.28 mmol, 1.2

equiv) and TfOH (9.58 μ l, 0.108 mmol, 0.1 equiv). The reaction mixture was stirred at -40°C for 0.5 h to 3 h until the disappearance of donor on TLC, then quenched with triethylamine (81 μ l, 0.81 mmol, 0.75 equiv) and warmed to room temperature. The mixture was diluted with dichloromethane, filtered through celite, washed with 20% aqueous Na₂S₂O₃ solution, dried over Na₂SO₄, and concentrated under reduced pressure. The residue was purified by column chromatography on silica gel eluting with ethylacetate/hexane (1:1) system to afford the compound **12A** (450 mg, 70%). ¹H-NMR (400 MHz, CDCl₃) δ 8.04–7.92 (m, 4H), 7.55–7.43 (m, 4H), 7.43–7.29 (m, 7H), 5.86 (dd, J = 10.4, 8.0 Hz, 1H), 5.54 (s, 1H), 5.37 (dd, J = 10.4, 3.6 Hz, 1H), 4.86 (d, J = 8.0 Hz, 1H), 4.58 (dd, J = 3.6, 1.6 Hz, 1H), 4.39 (dd, J = 12.4, 1.2 Hz, 1H), 4.13 (dd, J = 11.3, 1.6 Hz, 1H), 4.04 (dt, J = 11.3, 3.6 Hz, 1H), 3.79 (ddd, J = 11.3, 7.4, 3.6 Hz, 1H), 3.68 (s, 1H), 3.60 (dt, J = 7.4, 3.6 Hz, 2H), 3.45 (t, J = 5.1 Hz, 2H), 3.04 (m, 2H). ¹³C-NMR (400 MHz, CDCl₃) δ 165.68, 165.27, 136.82, 133.37, 133.19, 130.08, 130.04, 129.99, 129.90, 129.87, 129.46, 129.40, 129.12, 128.57, 128.49, 128.40, 128.44, 128.38, 128.28, 126.20, 101.95, 101.57, 78.90, 72.56, 72.07, 70.44, 70.18, 69.81, 68.72, 66.68, 50.60. Maldi-ToF m/z calc'd for C₃₁H₃₁N₃O₉Na (M+Na⁺): 612.1958; found: 612.1951.

Synthetic procedure for comp 12: A solution of compound **12A** (300 mg, 0.509 mmol) and activated 4Å powdered molecular sieves 1.5 g were stirred 1h at room temperature. The mixture was cooled to -78°C and then to the stirred solution Et₃SiH (259 μ l, 1.62 mmol) and PhBCl₂ (147 μ l, 1.38 mmol) were added successively. After being stirred for 1 h at -78°C, Et₃N (1 ml) and MeOH (1 ml) were added successively, and the mixture was diluted with CHCl₃ and washed with aqueous NaHCO₃, dried over MgSO₄, filtered and concentrated. The crude product was purified by silica gel column hexane:ethylacetate (1:1) to afford comp **12** (260 mg, 86%). ¹H NMR (400 MHz, CDCl₃) δ 8.00–7.94 (m, 4H), 7.54–7.47 (m, 2H), 7.36 (dd, J = 10.5, 4.8 Hz, 4H), 7.28 – 7.21 (m, 5H), 5.84 (dd, J = 10.5, 7.9 Hz, 1H), 5.34 (dd, J = 10.5, 3.0 Hz, 1H), 4.79 (d, J = 7.9 Hz, 1H), 4.78 (d, J = 11.7 Hz, 1H), 4.48 (d, J = 11.7 Hz, 1H), 4.13 (d, J = 2.5 Hz, 1H), 3.99 (ddd, J = 11.4, 4.5, 3.3 Hz, 1H), 3.86 (dd, J = 11.4, 6.9 Hz, 1H), 3.81 – 3.69 (m, 2H), 3.64 – 3.53 (m, 3H), 3.46 (t, J = 5.1 Hz, 2H), 3.14 – 2.99 (m, 2H), 1.76 (bs, 1H). ¹³C NMR (400 MHz, CDCl₃) δ 166.11, 165.43, 137.30, 133.60, 133.24, 129.98, 129.98, 129.78, 129.78, 129.72, 129.02, 128.62, 128.61, 128.60, 128.59, 128.44, 128.38, 128.23, 128.08, 127.95, 101.65, 77.31, 75.03, 74.81, 74.78, 73.32, 70.56, 70.15, 69.35, 61.79, 50.64. Maldi-ToF m/z calc'd for C₃₁H₃₃N₃O₉Na (M+Na⁺): 614.2114; found: 614.2120.

Synthetic procedure for comp 13A: Compound was synthesized from **13B** by following synthetic procedure mentioned above **13A** (420 mg, 65%). **¹H-NMR** (400 MHz, CDCl₃) δ 7.99–7.91 (m, 4H), 7.55–7.27 (m, 11H), 5.78 (t, *J* = 9.6 Hz, 1H), 5.53 (s, 1H), 5.47 (dd, *J* = 9.6, 7.9 Hz, 1H), 4.89 (d, *J* = 7.9 Hz, 1H), 4.42 (dd, *J* = 10.5, 4.9 Hz, 1H), 3.99 (ddd, *J* = 11.3, 4.9, 3.5 Hz, 1H), 3.92 (t, *J* = 8.4 Hz, 1H), 3.90–3.85 (m, 1H), 3.79–3.73 (m, 1H), 3.69 (td, *J* = 9.6, 4.9 Hz, 1H), 3.62–3.52 (m, 2H), 3.48–3.39 (m, 2H), 3.12–3.00 (m, 2H). **¹³C-NMR** (400 MHz, CDCl₃) δ 165.68, 165.27, 136.82, 133.37, 133.19, 130.08, 129.99, 129.90, 129.87, 129.54, 129.46, 129.40, 129.22, 129.12, 128.57, 128.49, 128.38, 128.32, 128.28, 126.20, 101.95, 101.57, 78.90, 72.56, 72.07, 70.44, 70.18, 69.81, 68.72, 66.68, 50.60. Maldi-ToF *m/z* calc'd for C₃₁H₃₁N₃O₉Na (M+Na⁺): 612.1958; found: 612.1955.

Synthetic procedure of comp 13: Compound synthesized from **13A** by same procedure mentioned for 12 to give **13** (240 mg, 80%). **¹H-NMR** (400 MHz, CDCl₃) δ 7.96–7.89 (m, 4H), 7.53–7.45 (m, 2H), 7.39–7.32 (m, 4H), 7.18–7.11 (m, 5H), 5.73 (t, *J* = 9.6 Hz, 1H), 5.33 (dd, *J* = 9.6, 8.0 Hz, 1H), 4.80 (d, *J* = 8.0 Hz, 1H), 4.59 (s, 2H), 4.01–3.90 (m, 3H), 3.81 (dd, *J* = 11.2, 3.9 Hz, 1H), 3.74 (ddd, *J* = 11.2, 4.8, 3.2 Hz, 1H), 3.63–3.54 (m, 3H), 3.45 (m, 2H), 3.08 (m, 2H), 2.01(bs, 1H). **¹³C NMR** (400 MHz, CDCl₃) δ 165.78, 165.38, 137.22, 133.30, 133.28, 129.85, 129.84, 129.82, 129.47, 129.44, 128.47, 128.40, 128.38, 128.37, 128.35, 128.34, 128.32, 128.30, 128.28, 128.07, 101.31, 77.33, 75.55, 74.96, 74.91, 72.20, 70.46, 70.19, 69.69, 61.59, 50.61. Maldi-ToF *m/z* calc'd for C₃₁H₃₃N₃O₉Na (M+Na⁺): 614.2114; found: 614.2120.

Synthesis of compound 14A: To a stirred solution of sulfoxide **14B** (500 mg, 0.900 mmol, 1eq) and TTBP (468 mg, 2.1 eq) in CH₂Cl₂(10 ml) at -78⁰C was added Tf₂O (151 μl, 1eq) and, 5 min later, the solution of the glycosyl acceptor (78 mg, 1.2 eq) in CH₂Cl₂ (2 ml) drop wise were added. The reaction mixture was stirred at -78 ⁰C for 2 h and then allowed to warm to 0 ⁰C over 2 h and maintained for further 0.5 h before quenching with saturated aqueous NaHCO₃, washing with brine, drying, concentrating, and purified by chromatography on silica gel Hexane:Ethylacetate to give compound **14A** (320 mg, 64%); **¹H-NMR** (400 MHz, CDCl₃) δ 7.51–7.44 (m, 2H), 7.40–7.25 (m, 13H), 5.61 (s, 1H), 4.96 (d, *J* = 12.2 Hz, 1H), 4.87 (d, *J* = 12.2

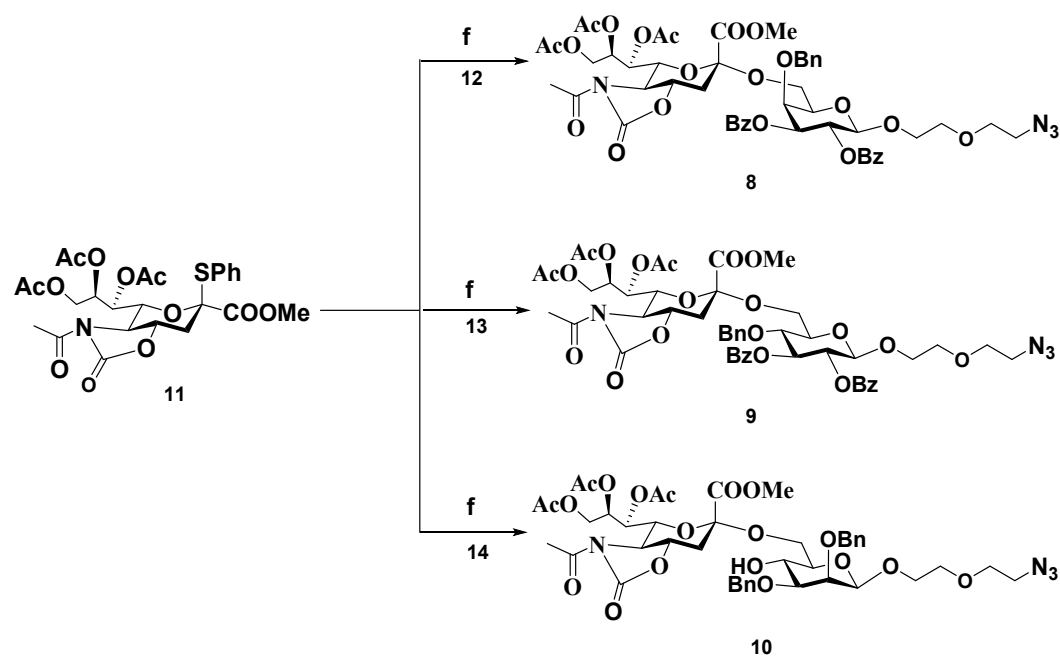
Hz, 1H), 4.66 (d, $J = 12.5$ Hz, 1H), 4.57 (d, $J = 12.5$ Hz, 1H), 4.54 (s, 1H), 4.29 (dd, $J = 10.4$, 4.9 Hz, 1H), 4.19 (t, $J = 9.6$ Hz, 1H), 4.05–3.99 (m, 1H), 3.98 (d, $J = 3.0$ Hz, 1H), 3.92 (t, $J = 10.3$ Hz, 1H), 3.74–3.62 (m, 5H), 3.58 (dd, $J = 9.6$, 3.1 Hz, 1H), 3.33 (dt, $J = 9.6$, 5.4 Hz, 3H). ^{13}C -NMR (400 MHz, CDCl_3) δ 138.56, 138.39, 137.64, 134.56, 129.84, 129.09, 128.93, 128.72, 128.63, 128.42, 128.38, 128.27, 128.18, 128.05, 127.86, 127.65, 127.60, 126.14, 102.09, 101.50, 81.48, 78.67, 74.92, 73.29, 71.09, 70.69, 70.30, 69.19, 67.50, 63.10, 50.82. Maldi-ToF m/z calc'd for $\text{C}_{31}\text{H}_{35}\text{N}_3\text{O}_7\text{Na}$ ($\text{M}+\text{Na}^+$): 584.2372; found: 584.2380.

Synthesis of compound 14: A solution of compound **14A** (350 mg 534 μmol) was dissolved in dichloromethane (7 ml) and stirred for 1 h at room temperature. The mixture was cooled to 0°C and then catalytic amount of *p*-TsOH were added to it. Allowed this reaction to stir at RT till complete disappearance of starting material on TLC. Then quenched the mixture by TEA and diluted with DCM followed by extraction with sat. NaHCO_3 . The organic layer were concentrated and purified by silica gel column chromatography (DCM:MeOH) to afford colourless syrup (250 mg, 85%) ^1H NMR (400 MHz, CDCl_3) δ 7.49–7.41 (m, 2H), 7.36–7.24 (m, 8H), 4.96 (d, $J = 12.3$ Hz, 1H), 4.80 (d, $J = 12.3$ Hz, 1H), 4.56 (s, 1H), 4.51 (d, $J = 11.8$ Hz, 1H), 4.33 (d, $J = 11.8$ Hz, 1H), 4.10–4.03 (m, 1H), 4.02–3.98 (m, 1H), 3.97–3.91 (m, 1H), 3.89–3.80 (m, 1H), 3.78–3.67 (m, 5H), 3.39 (ddd, $J = 9.6$, 5.5, 4.4 Hz, 2H), 3.37–3.29 (m, 2H), 2.75 (d, $J = 3.8$ Hz, 1H), 2.55 (s, 1H), 2.03 (s, 1H). ^{13}C NMR (400 MHz, CDCl_3) δ 138.52, 137.66, 128.56, 128.54, 128.36, 128.34, 128.20, 128.18, 127.92, 127.79, 127.77, 127.56, 101.97, 81.41, 75.94, 74.26, 73.41, 71.10, 70.66, 70.20, 69.08, 67.37, 62.91, 50.74. Maldi-ToF m/z calc'd for $\text{C}_{26}\text{H}_{28}\text{O}_5\text{S}$ ($\text{M}+\text{Na}^+$): 496.2059; found: 496.2061.

Synthesis of compound 11: A stirred solution of **7** (5.00 g, 8.57 mmol, 1.00 equiv) in methanol (80 ml) was treated with methanesulfonic acid (1.68 ml, 25.7 mmol, 3.0 equiv) at room temperature, and then refluxed under Ar for 24 h. After being cooled to room temperature, the reaction mixture was quenched with excess triethylamine, and then concentrated. The concentrate and NaHCO_3 (3.60 g, 42.8 mmol, 5.0 equiv) were dissolved in CH_3CN (30 ml) and H_2O (60 ml) and cooled to 0°C . To the vigorously stirred mixture was added 4-nitrophenylchloroformate (4.32 g, 21.4 mmol, 2.5 equiv) in CH_3CN (30 ml) slowly through a

dropping funnel, after which stirring was continued for 3 h at 0°C. The resulting mixture was extracted with ethylacetate and the combined extracts were washed with brine, and then dried over Na₂SO₄ and concentrated. The residue was purified by silica gel column chromatography, eluting with EtOAc then EtOAc/MeOH from 10/1 to 5/1 to give the title residue as white foam. A solution of residue (2.67 gm) in pyridine (20 ml) was treated with Ac₂O (24 ml) and stirred at room temperature overnight, then concentrated under reduced pressure. The residue was dissolved in anhydrous CH₂Cl₂, treated with EtN(*i*-Pr)₂ (11.6 ml, 66.8 mmol, 10 equiv), and cooled to 0°C before acetyl chloride (3.87 ml, 53.4 mmol, 8 equiv) was added dropwise, then the mixture stirred at 0°C for 1 h. After warming to room temperature, the resulting solution was poured into saturated aqueous NaHCO₃ solution, the organic layer was separated, the aqueous layer was extracted twice with CH₂Cl₂, and the combined organic phase was washed with brine, dried over Na₂SO₄, and concentrated. The residue was purified by column chromatography on silica gel eluting with EtOAc/Hex (1:1) to give donor **1** as a yellowish solid (2.8 g, 82%), which can be further purified by recrystallization from EtOAc/Et₂O/Hex to afford white needle crystals. **¹H-NMR** (400 MHz, CDCl₃) δ 7.55–7.32 (m, 5H), 5.57 (t, *J* = 2.3 Hz, 1H), 5.01 (dt, *J* = 8.2, 2.3 Hz, 1H), 4.90 (dd, *J* = 9.1, 2.5 Hz, 1H), 4.80 (ddd, *J* = 12.8, 11.4, 2.5 Hz, 1H), 4.38 (dd, *J* = 12.1, 2.5 Hz, 1H), 3.91 (dd, *J* = 12.1, 8.3 Hz, 1H), 3.77 (dd, *J* = 11.3, 9.2 Hz, 1H), 3.64 (s, 3H), 2.92 (dd, *J* = 13.0, 3.7 Hz, 1H), 2.54 (s, 3H), 2.35 (t, *J* = 13.0 Hz, 1H), 2.16 (s, 3H), 2.10 (s, 3H), 1.97 (s, 3H). **¹³C-NMR** (400 MHz, CDCl₃) δ 172.46, 171.24, 170.38, 169.72, 167.80, 153.58, 136.77, 136.70, 130.20, 129.23, 129.20, 128.18, 88.35, 75.72, 75.11, 73.85, 72.69, 62.92, 59.67, 52.79, 36.05, 24.76, 21.17, 20.82, 20.73. Maldi-ToF *m/z* calc'd for C₂₅H₂₉NO₁₂SNa (M+Na⁺): 590.1308; found: 590.1307.

Glycosylation of donor and acceptor:



Scheme S2: Reagents and Conditions: (f) NIS/TfOH, -40°C, DCM.

General procedure for glycosylation. A solution of donor **11** (200 mg, 0.11 mmol, 1.0 equiv), acceptor (1.2 equiv), and activated 4 Å powdered molecular sieves (216 mg, 2.0 g/mmol) in anhydrous dichloromethane (2 ml) was stirred overnight under an argon atmosphere, and then cooled to -40°C followed by addition of NIS (172 mg, 0.26 mmol, 2.4 equiv) and TfOH (9.5 μ l, 0.11 mmol, 1.0 equiv). The reaction mixture was stirred at -40°C for 20 min to 2 h until the disappearance of the donor on TLC, then quenched with triethylamine (22.6 μ l, 0.16 mmol, 1.5 equiv) and warmed to room temperature. The mixture was diluted with dichloromethane, filtered through celite, washed with 20% aqueous Na₂S₂O₃ solution, dried over Na₂SO₄, and concentrated under reduced pressure. The residue was purified by column chromatography on silica gel eluting with THF/Hex system to afford the sialic acid disaccharide.

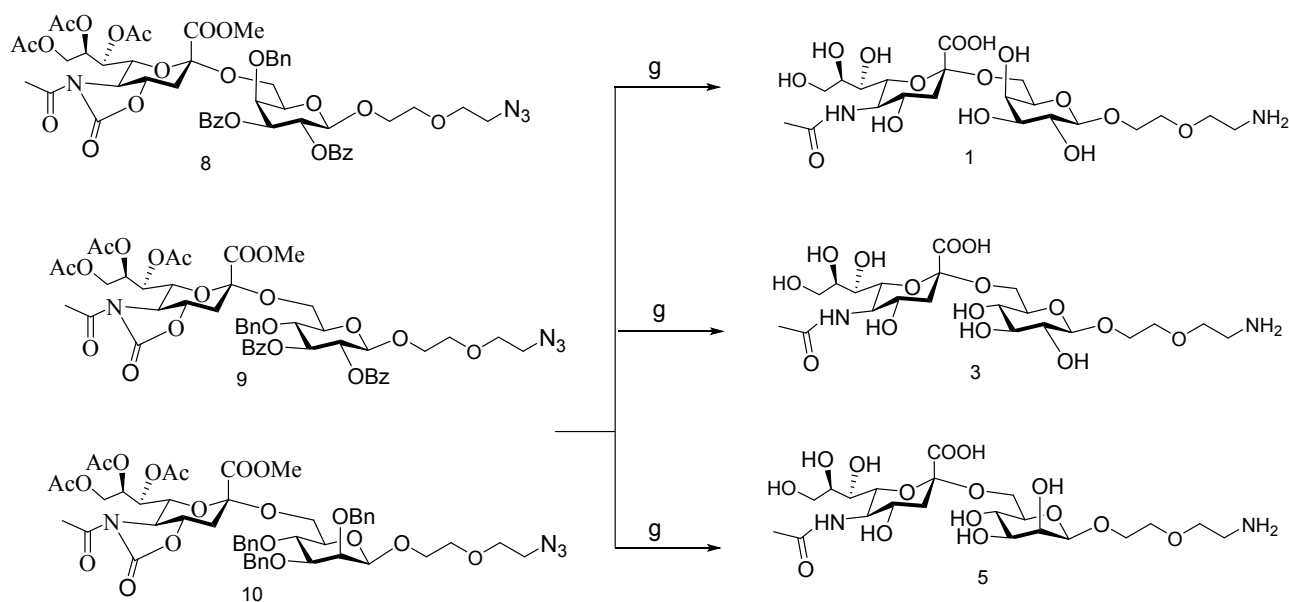
Compound 8: According to general procedure, donor **11** (200 mg) and acceptor **12** (250 mg , 1.2eq) to afford **8** (270 mg , 73%). **¹H-NMR** (400 MHz, CDCl₃) δ 7.52–7.46 (m, 3H), 7.39–7.31 (m, 6H), 7.30–7.25 (m, 3H), 7.18–7.15 (m, 3H), 5.81 (dd, *J* = 10.5, 8.0 Hz, 1H), 5.58 (dd, *J* = 8.0, 1.6 Hz, 1H), 5.43 (td, *J* = 7.1, 3.1 Hz, 1H), 5.35 (dd, *J* = 10.5, 3.1 Hz, 1H), 4.83 (d, *J* = 8.0 Hz, 1H), 4.70–4.60 (m, 3H), 4.41 (dd, *J* = 12.3, 3.0 Hz, 1H), 4.22 (d, *J* = 3.0 Hz, 1H), 4.06 (dd, *J* = 12.3, 7.0 Hz, 1H), 4.03–3.97 (m, 2H), 3.96–3.85 (m, 3H), 3.81–3.74 (m, 2H), 3.71(s, 3H), 3.58 (m, 2H), 3.47 (t, *J* = 5.1Hz, 2H), 3.08–2.96 (m, 2H), 2.85 (dd, *J* = 12.1, 3.5 Hz, 1H), 2.49 (s, 3H), 2.19 (s, 3H), 2.13 (s, 3H), 2.05 (t, *J* = 12.1 Hz, 1H), 2.00 (s, 3H). **¹³C-NMR** (400 MHz, CDCl₃) δ 172.03, 170.79, 170.14, 170.09, 168.39, 165.67, 165.36, 153.74, 137.44, 133.25, 133.20, 129.98, 129.98, 129.82, 129.78, 129.72, 129.47, 129.02, 128.62, 128.61, 128.60, 128.59, 128.43, 128.33, 128.23, 127.99, 101.21, 99.47, 75.52, 75.36, 75.13, 74.81, 74.60, 74.45, 71.98, 71.63, 70.34, 70.15, 69.53, 68.54, 64.53, 63.10, 59.19, 53.08, 50.60, 36.76, 24.79, 21.28, 20.88, 20.79. HRMS *m/z* calc'd for C₅₀H₅₆N₄O₂₁ (M+Na⁺): 1071.3314; found:1071.3311.

Compound 9: According to general procedure, donor **11**(200 mg) and acceptor **13** (250 mg , 1.2 eq) to afford **9** (255 mg, 69%). **¹H-NMR** (400 MHz, CDCl₃) δ 7.51–7.46 (m, 4H), 7.37–7.32 (m 5H), 7.15 – 7.08 (m, 6H), 5.69–5.60 (m, 2H), 5.48 (ddd, *J* = 9.0, 6.3, 3.0 Hz, 1H), 5.35–5.28 (m, 2H), 4.72 (dd, *J* = 9.4, 3.9 Hz, 2H), 4.66 (dd, *J* = 9.4, 1.6 Hz, 1H), 4.54 (d, *J* = 10.8 Hz, 1H), 4.35 (dd, *J* = 12.3, 2.9 Hz, 1H), 4.22 (dd, *J* = 11.4, 4.4 Hz, 1H), 4.05 (dd, *J* = 12.3, 6.4 Hz, 1H), 4.02–3.94(m, 2H), 3.90 (dd, *J* = 12.3, 6.6 Hz, 1H), 3.74 (s, 3H), 3.73 – 3.64 (m, 3H), 3.57 (m, 2H), 3.45–3.43 (m, 2H), 3.07–3.04 (m, 2H), 3.01 (dd, *J* = 12.0, 3.4 Hz, 1H), 2.49 (s, 3H), 2.13 (s, 3H), 2.03 (s, 3H), 2.03 (t, *J* = 12.5 Hz, 1H), 1.90 (s, 3H). **¹³C-NMR** (400 MHz, CDCl₃) δ 172.03, 170.79, 170.14, 170.09, 168.39, 165.67, 165.36, 153.74, 137.44, 133.25, 133.20, 129.85, 129.84, 129.82, 129.47, 129.44, 129.47, 129.44, 128.47, 128.43, 128.40, 128.38, 128.37, 128.33, 128.30, 127.99, 101.21, 99.47, 75.52, 75.36, 75.13, 74.81, 74.60, 74.45, 71.98, 71.63, 70.34, 70.15, 69.53, 68.54, 64.53, 63.10, 59.19, 53.08, 50.60, 36.76, 24.79, 21.28, 20.88, 20.79. HRMS *m/z* calc'd for C₅₀H₅₆N₄O₂₁ (M+Na): 1071.3314; found:1071.3322.

Compound 10: According to general procedure, donor **11** (200 mg) and acceptor **14** (238 mg ,1.2eq) to afford **10** (230 mg , 64%). **¹H NMR** (400 MHz, CDCl₃) δ 7.45–7.20 (m, 10H), 5.61 (dd, *J* = 8.2, 1.2 Hz, 1H), 5.46–5.32 (m, 1H), 5.05 (dd, *J* = 10.5, 3.1 Hz, 1H), 5.00 (s, 1H), 4.93 (d, *J* = 12.3 Hz, 1H), 4.78 (d, *J* = 12.3 Hz, 1H), 4.58 (d, *J* = 9.3 Hz, 1H), 4.51 (d, *J* = 11.3 Hz,

1H), 4.42 (d, $J = 11.3$ Hz, 1H), 4.34 (dd, $J = 12.3, 3.2$ Hz, 1H), 4.14 (dd, $J = 9.2, 4.5$ Hz, 1H), 4.10 – 4.01 (m, 4H), 3.98–3.90 (m, 3H), 3.78 (s, 3H), 3.76–3.52 (m, 4H), 3.39–3.23 (m, 4H), 3.03 (s, 1H), 2.88 (dd, $J = 12.3, 3.6$ Hz, 1H), 2.48 (s, 3H), 2.25 (s, 3H), 2.11 (s, 3H), 2.05 (t, dd, $J = 12.4$ Hz, 1H), 2.02 (s, 3H). ^{13}C NMR (400 MHz, CDCl_3) δ 172.12, 170.76, 170.16, 169.81, 168.99, 151.45, 139.06, 138.05, 129.42, 128.39, 128.33, 128.29, 128.25, 128.09, 127.70, 127.43, 126.82, 125.52, 102.08, 98.69, 80.76, 75.61, 75.51, 75.01, 73.99, 73.71, 71.58, 70.46, 69.11, 68.97, 66.42, 64.35, 62.78, 61.41, 59.05, 54.26, 53.00, 50.74, 36.48, 24.70, 21.19, 20.89, 20.85. HRMS m/z calc'd for $\text{C}_{43}\text{H}_{54}\text{N}_4\text{O}_{19}$ ($\text{M}+\text{Na}$): 953.3279; found: 953.3292.

Global deprotection of comp 8, 9, 10 :



Scheme S3: Reagents and Conditions g) i) LiOH, EtOH:H₂O (3:1), 80°C, 12h; ii) NaHCO₃, Ac₂O, H₂O, rt; iii) H₂/Pd(OH)₂, MeOH:H₂ (1:1), 12h.

LiOH (30 eq) was added to a stirred solution of protected oligosaccharide (1 eq) in ethanol water (3:1) 5 ml at room temperature. After stirring at 80°C for 12h, the reaction mixture cooled to room temperature and carefully neutralized by IR-120H⁺ resin to pH-7, diluted with methanol, filtered and concentrated. Crude residue was dissolved in water and NaHCO₃ (15 eq) were added to it. Cooled the reaction mixture to 0°C and Acetic-anhydride (10 eq) was added. The reaction were monitored by TLC (ethylacetate:methanol:water 7:2:1), after 3h, LiOH (10 eq)

were added and stirred for another 5-6h at room temperature, then reaction mixture carefully neutralized by IR-120H⁺resin to pH-7, diluted concentrated and purified by reverse-phase column chromatography(Bond Elu-C18).The Pd(OH)₂ (1 mmole) was added to the above residue in methanol water(1:1) 4ml. The reaction mixture was hydrogenalysed under H₂ atmosphere for 12h, filtered and concentrated. The residue was purified by reverse phase column chromatography (Bond Elu-C18) to give deprotected sialic acid analogues.

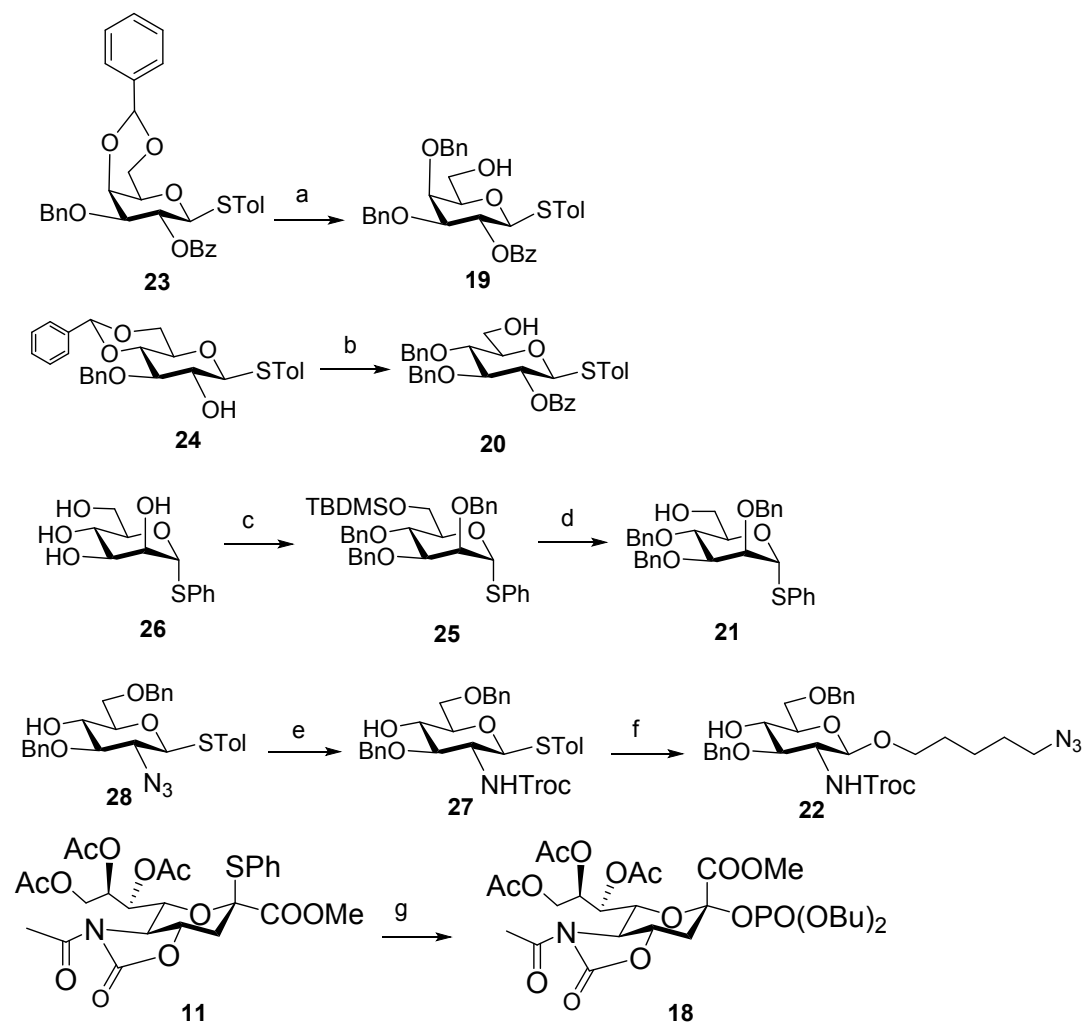
Compound 1: Synthesized from **8** by following general deprotection protocol to yield 36 mg (36%). **¹H-NMR** (400 MHz, D₂O) δ 4.33 (d, J = 7.7 Hz, 1H), 3.97 (dd, J = 11.8, 7.1 Hz, 1H), 3.83 (t, J = 8.7 Hz, 2H), 3.76 (dd, J = 11.5, 8.7 Hz, 4H), 3.72–3.64 (m, 5H), 3.63–3.45 (m, 7H), 3.42 (dd, J = 16.3, 8.3 Hz, 1H), 3.28 (t, J = 5.0 Hz, 1H), 3.15–3.10 (m, 1H), 2.63 (dd, J = 12.5, 3.6 Hz, 1H), 1.93 (s, 3H), 1.58 (t, J = 12.0 Hz, 1H). **¹³C-NMR** (400 MHz, D₂O) δ 175.02, 173.42, 103.01, 100.41, 73.44, 72.59, 72.46, 71.72, 71.42, 70.74, 68.59, 68.22, 68.17, 63.44, 62.60, 60.77, 60.37, 51.84, 40.15, 39.74, 22.03. HRMS m/z calc'd for C₂₁H₃₈N₂O₁₅ (M+H): 559.2350; found: 559.2352.

Compound 3: Synthesized from **9** by following general deprotection protocol to yield 20 mg (20%). **¹H-NMR** (400 MHz, MeOD) δ 4.34 (d, J = 7.8 Hz, 1H), 4.06 (dd, J = 10.4, 3.5 Hz, 1H), 3.95 (t, J = 9.5 Hz, 1H), 3.93–3.88 (m, 1H), 3.88–3.60 (m, 10 H), 3.58–3.48 (m, 2H), 3.38 (dd, J = 16.2, 6.8 Hz, 2H), 3.27–3.11 (m, 4H), 2.76 (dd, J = 12.0, 3.7 Hz, 1H), 2.04 (s, 3H), 1.66 (t, J = 12.0 Hz, 1H). **¹³C-NMR** (400 MHz, D₂O) δ 174.90, 173.86, 102.37, 99.79, 75.92, 75.03, 72.92, 71.84, 69.60, 69.18, 68.77, 68.31, 68.12, 67.57, 66.38, 62.67, 61.02, 51.75, 39.73, 39.15, 22.17. HRMS m/z calc'd for C₂₁H₃₈N₂O₁₅ (M+H): 559.2350; found: 559.2352.

Compound 5: Synthesized from **10** by following general deprotection protocol to yield 27 mg (25%). **¹H-NMR** (400 MHz, MeOD) δ 4.57 (d, J = 0.8 Hz, 1H), 4.11–4.02 (m, 2H), 3.94–3.81 (m, 6H), 3.81–3.59 (m, 10H), 3.55-3.45 (m, 4H), 2.85 (dd, J = 12.4, 4.3 Hz, 1H), 2.03 (s, 3H), 1.66 (t, J = 12.4 Hz, 1H). **¹³C-NMR** (400 MHz, D₂O) δ 175.11, 173.80, 102.52, 99.35, 75.92, 75.03, 72.92, 71.84, 69.60, 69.18, 68.77, 68.31, 68.12, 67.57, 66.38, 62.67, 61.02, 51.75, 39.73, 39.15, 21.84. HRMS m/z calc'd for C₂₁H₃₈N₂O₁₅ (M+H): 559.2350;found: 557.2352.

3. Synthesis of trisaccharide library of compounds:

Synthesis of donor and acceptors



Scheme S4 : Reagents and conditions: a) PhBCl₂, Et₃SiH, -78°C, DCM ; b) i) BzCl/Py; ii) PhBCl₂, Et₃SiH, -80°C, DCM ; c) i) TBDMSO, Imidazole, DMF; ii) BnBr, NaH, DMF; d) p-TSA, MeOH:DCM; e) i) 1,3-dithiopropene, DCM, 35°C ; ii) TrocCl, NaHCO₃, THF; f) 5-Azidopentanol, NIS/TfOH, -40°C, DCM; g) Dibutylphosphate, NIS/TfOH, DCM:ACN, -20°C.

Synthetic procedure:

Compound 19: A solution of compound **23** (600 mg, 1.08 mmol) and activated 4Å powdered molecular sieves 1.1 g in dichloromethane were stirred 1h at room temperature. The mixture was cooled to -78°C and then to the stirred solution Et₃SiH (552 µl, 3.46 mmol) and PhBCl₂ (265 µl, 2.05 mmol) were added successively. After being stirred for 1 h at -78°C, Et₃N (1 ml) and MeOH (1 ml) were added successively, and the mixture was diluted with CHCl₃ and washed with aqueous NaHCO₃, dried over MgSO₄, filtered and concentrated. The crude product was purified by silica gel column hexane:ethylacetate (2:1) to afford white solid (545 mg, 91%). ¹H-NMR (400 MHz, CDCl₃) δ 8.06 (d, *J* = 7.6 Hz, 2H), 7.63 (t, *J* = 7.4 Hz, 1H), 7.49 (t, *J* = 7.6 Hz, 2H), 7.42 – 7.31 (m, 7H), 7.27–7.14 (m, 5H), 7.06 (d, *J* = 7.8 Hz, 2H), 5.70 (t, *J* = 9.7 Hz, 1H), 5.02 (d, *J* = 11.8 Hz, 1H), 4.74 (d, *J* = 9.7 Hz, 1H), 4.67 (t, *J* = 10.8 Hz, 2H), 4.57 (d, *J* = 12.2 Hz, 1H), 3.94 (s, 1H), 3.93–3.85 (m, 1H), 3.72 (dd, *J* = 9.0, 2.2 Hz, 1H), 3.59 (dd, *J* = 9.0, 5.2 Hz, 1H), 3.56–3.50 (m, 1H), 2.31 (s, 3H), 1.67 (dd, *J* = 8.5, 3.7 Hz, 1H). ¹³C-NMR (400 MHz, CDCl₃) δ 165.28, 138.11, 137.85, 137.43, 133.08, 132.71, 132.69, 130.08, 129.90, 129.88, 129.58, 129.51, 128.47, 128.44, 128.45, 128.41, 128.40, 128.39, 128.37, 128.35, 127.92, 127.86, 127.84, 127.79, 127.77, 87.25, 81.32, 78.99, 77.23 74.07, 72.18, 70.54, 62.20, 21.14; HRMS *m/z* calc'd for C₃₄H₃₄O₆SNa (M+Na⁺). 593.1973; found: 593.1971.

Compound 20: To solution of compound **24** (700 mg, 1.5 mmol) in dry pyridine (10 ml) benzoyl chloride (440 ml, 3.7mmol) were added dropwise. Allowed the reaction mixture to stir for 12h at room temperature. After completion quenched the reaction by methanol 2ml and diluted with DCM. Extracted with dilute HCl (5%), dried over Na₂SO₄ and concentrated. Crude residue was purified through silica gel column to afford the white solid. This white solid (575 mg, 1.03 mmol) dissolved in dry dichloromethane (10 ml) containing 4Å M.S. and stirred at room temperature for 1h. Reaction mixture was cooled to -78°C and then Et₃SiH (533 µl, 3.32 mmol) and PhBCl₂ (253 µl, 1.95 mmol) were added successively. After being stirred for 1 h at -78°C, Et₃N (1 ml) and MeOH (1 ml) were added successively, and the mixture was diluted with CHCl₃ and washed with aqueous NaHCO₃, dried over MgSO₄, filtered and concentrated. The crude product was purified by silica gel column hexane:ethylacetate (1:1) to afford white solid(530 mg, 88%). ¹H-NMR (400 MHz, CDCl₃) δ 8.08 (dd, *J* = 8.3, 1.3 Hz, 2H), 7.65 – 7.60 (m, 1H), 7.49 (dd, *J* = 8.2, 4.7 Hz, 2H), 7.40–7.29 (m, 7H), 7.18–7.09 (m, 7H), 5.26 (dd, *J* = 9.9,

9.2 Hz, 1H), 4.87 (d, J = 11.0 Hz, 1H), 4.77 (dd, J = 10.5, 6.1 Hz, 2H), 4.68 (d, J = 11.0 Hz, 2H), 3.94 (ddd, J = 12.0, 6.0, 2.6 Hz, 1H), 3.88 (t, J = 9.0 Hz, 1H), 3.79–3.73 (m, 1H), 3.73–3.67 (m, 1H), 3.51 (ddd, J = 9.7, 4.7, 2.6 Hz, 1H), 2.35 (s, 3H), 1.99 (t, J = 6.8 Hz, 1H). $^{13}\text{C-NMR}$ (400 MHz, CDCl_3) δ 165.20, 138.44, 137.75, 137.60, 133.32, 133.31, 133.25, 129.88, 129.86, 129.85, 129.75, 129.73, 128.54, 128.52, 128.50, 128.45, 128.30, 128.28, 128.12, 128.10, 128.08, 128.05, 128.03, 127.70, 127.72, 86.34, 84.06, 79.56, 77.51, 75.34, 75.19, 72.55, 62.07, 21.16. HRMS m/z calc'd for $\text{C}_{34}\text{H}_{34}\text{O}_6\text{SNa}$ ($\text{M}+\text{Na}$). 593.1971; found: 593.1973.

Compound 25 : The solution of compound **26** (1.2gm, 4.41 mmol) in dry DMF (10 ml) cooled to -20°C and then imidazole (0.6 gm, 8.82 mmol) were added to it. After 10 minutes TBDMSCl (0.79 gm, 5.29 mmol) were added portion wise and allowed the reaction mixture to stir for 2h. After completion of reaction, evaporated the solvent and then diluted with DCM and extracted with 1N HCl. The organic layer dried over Na_2SO_4 . This crude residue was purified by flash silica gel column chromatography in DCM:MeOH (9:1) to afford as colorless syrup (0.7 gm, 40%). This syrup was dissolved in dry THF (10 ml) followed by addition of tetrabutylammonium iodide (2 gm, 5.44 mmol). Cooled this reaction mixture to 0°C and then sodium hydride (60%, 450 mg) was added. After 5 minutes benzyl bromide (1.0 ml, 9 mmol) was added dropwise and allowed the reaction mixture to stir for 12h at room temperature. After completion, reaction was quenched with methanol and diluted with ether. The organic layer was extracted with NaHCO_3 , brine and dried over Na_2SO_4 , concentrated. Purification by silica gel column chromatography (Ethylacetate:Hexane) afforded compound 25 as oily syrup (0.9 gm, 76%). $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ 7.36 (m, 20H), 5.59 (d, J = 1.5 Hz, 1H), 4.98 (d, J = 10.9 Hz, 1H), 4.74–4.62 (m, 5H) 4.13 (ddd, J = 9.6, 4.8, 1.7 Hz, 1H), 4.05 (d, J = 9.3 Hz, 1H), 4.01 (dd, J = 3.1, 1.6 Hz, 1H) 3.97–3.87 (m, 3H), 0.92 (s, 9H), 0.09 (s, 3H), 0.08 (s, 3H). $^{13}\text{C-NMR}$ (400 MHz, CDCl_3) δ 138.67, 138.28, 138.02, 134.80, 131.50, 131.48, 128.92, 128.90, 128.43, 128.41, 128.39, 128.37, 128.33, 128.31, 128.03, 128.01, 127.87, 127.86, 127.84, 127.82, 127.69, 127.64, 127.64, 127.24, 85.61, 80.16, 76.54, 75.21, 74.91, 74.23, 72.15, 71.79, 62.70, 25.97, 25.97, 25.97, 18.38, -5.12, -5.30. HRMS m/z calc'd for $\text{C}_{39}\text{H}_{48}\text{O}_5\text{SSiNa}$ ($\text{M}+\text{Na}^+$). 679.2889; found: 679.2891.

Compound 21 Compound **25** (0.750 gm, 1.14 mmol) was dissolved in dry methanol and dichloromethane mixture (10 ml). To it catalytic amount of *p*-TSOH (110 mg) were added and stirred this reaction mixture for 2h at room temperature. Quenched the reaction mixture with triethylamine and extracted with sat. NaHCO₃, brine and dried the organic layer over Na₂SO₄, concentrated. Purification of mixture by silica-gel column chromatography (Ethylacetate:Hexane) yielded compound **21** as colourless syrup (550 mg, 85%). **¹H-NMR** (400 MHz, CDCl₃) δ 7.41-7.25 (m, 20H), 5.50 (d, *J* = 1.6 Hz, 1H), 4.95 (d, *J* = 10.9 Hz, 1H), 4.67-4.59 (m, 2H), 4.64 (d, *J* = 5.1 Hz, 1H), 4.61 (d, *J* = 11.7 Hz, 1H), 4.15-4.09 (m, 1H), 4.04 (t, *J* = 9.4 Hz, 1H), 3.99 (dd, *J* = 2.9, 1.8 Hz, 1H), 3.88 (dd, *J* = 9.2, 3.1 Hz, 1H), 3.83 (dd, *J* = 8.9, 2.9 Hz, 1H), 3.79 (dd, *J* = 8.7, 4.4 Hz, 1H), 1.75 (bs, 1H). **¹³C NMR** (400 MHz, CDCl₃) δ 138.31, 138.12, 137.84, 133.94, 131.87, 131.85, 129.13, 129.12, 128.48, 128.47, 128.46, 128.45, 128.09, 128.07, 127.95, 127.93, 127.86, 127.85, 127.83, 127.81, 127.80, 127.78, 127.67, 127.65, 86.06, 80.12, 76.43, 75.32, 74.79, 73.25, 72.38, 72.26, 62.24. HRMS *m/z* calc'd for C₃₃H₃₄O₅S (M+Na⁺). 565.2024; found: 565.2025.

Compound 28: was synthesized by following synthetic reported protocol (C. H. Wang, K. Tony, K. Mong, C. Y. Huang, *J. Org. Chem.*, **2003**, 68, 2135-2142). **¹H-NMR** (400 MHz, CDCl₃) δ 7.50-7.40 (m, 2H), 7.39-7.21 (m, 11H), 7.07 (d, *J* = 7.9 Hz, 1H), 4.88 (d, *J* = 11.0 Hz, 1H), 4.81 (d, *J* = 11.0 Hz, 1H), 4.59 (d, *J* = 11.8 Hz, 1H), 4.54 (d, *J* = 11.8 Hz, 1H), 4.36 (d, *J* = 9.9 Hz, 1H), 3.78 (dd, *J* = 10.4, 4.9 Hz, 1H), 3.73 (dd, *J* = 10.4, 4.4 Hz, 1H), 3.60 (t, *J* = 9.0 Hz, 1H), 3.43 (dt, *J* = 9.4, 4.4 Hz, 1H), 3.35 (t, *J* = 9.0 Hz, 1H), 3.30-3.23 (m, 1H), 2.72 (s, 1H), 2.32 (s, 3H). **¹³C NMR** (400 MHz, CDCl₃) δ 138.86, 137.92, 137.79, 134.30, 129.87, 128.80, 128.72, 128.66, 128.57, 128.40, 128.34, 128.30, 128.22, 127.93, 127.95, 127.78, 127.80, 127.11, 86.25, 84.69, 78.04, 75.58, 73.83, 72.03, 70.39, 64.42, 21.26. HRMS *m/z* calc'd for C₂₇H₂₉N₃O₄S (M+Na⁺). 514.1776; found: 514.1769.

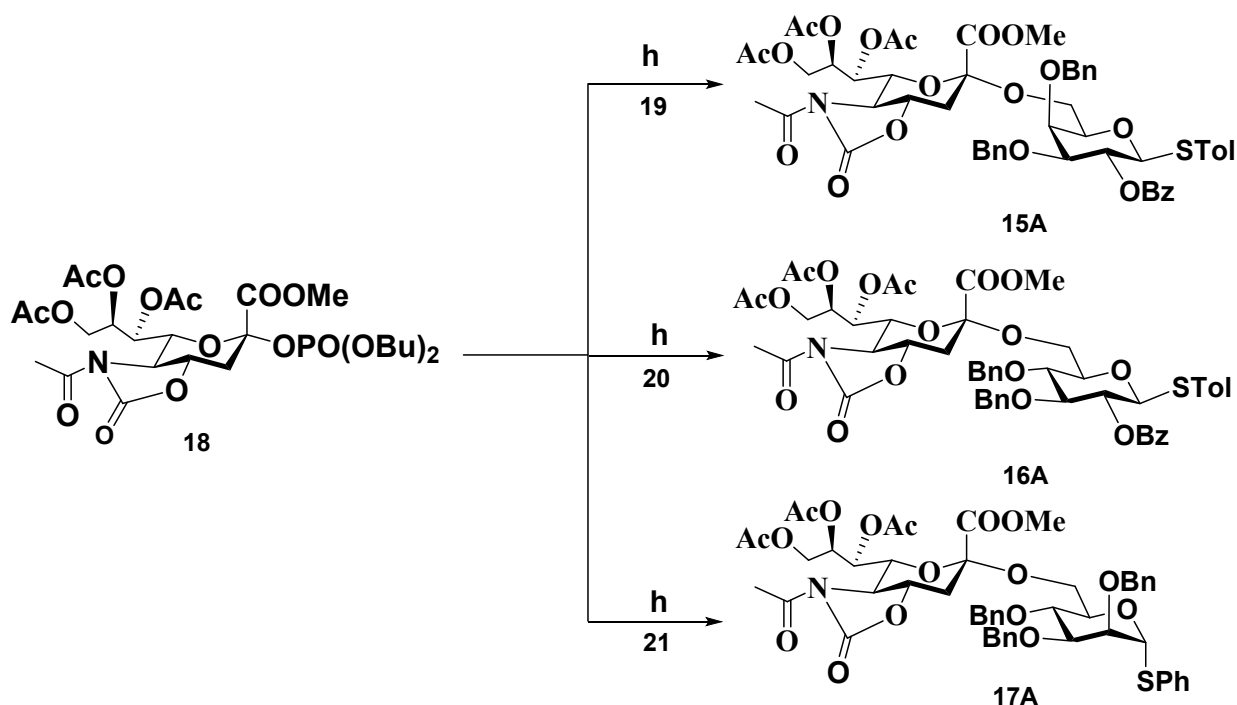
Compound 27. To compound **28** (2.5 gm, 5.09 mmol) in MeOH:DCM (1:1) was added 1,3 dithiopropene (2.5 ml, 25.45 mmol) and TEA (2.45 ml, 25.45 mmol) under Ar atmosphere. Reaction mixture was stirred at 35°C for 18h. Then coevaporated with toluene, concentrated and purified by flash column chromatography (Ethylacetate:Hexane) to afford oily syrup. This syrup was dissolved in THF and NaHCO₃ (0.818 gm, 10.18 mmol) was added to it. Cooled the reaction

mixture to 40°C and Troc-chloride (1 ml, 7.63 mmol) added dropwise. Stirred this reaction mixture at room temperature under argon for 6h. Filtered the reaction mixture and filtrate was concentrated. The residue was dissolved in DCM and washed with water, brine and the organic layer was dried over Na₂SO₄ and concentrated. The residue was purified on silica gel column chromatography (Ethylacetate:Hexane) to afford compound **27** as white solid (2.4 gm, 76%). **¹H-NMR** (400 MHz, CDCl₃) δ 7.42-7.24 (m, 12H), 7.04 (d, *J* = 7.9 Hz, 2H), 5.14 (d, *J* = 8.1 Hz, 1H), 4.88 (d, *J* = 10.2 Hz, 1H), 4.75 (m, 4H), 4.58 (d, *J* = 11.8 Hz, 1H), 4.53 (d, *J* = 11.8 Hz, 1H), 3.77 (dd, *J* = 4.7, 3.0 Hz, 2H), 3.67 (t, *J* = 9.3, 10.2 Hz, 2H), 3.45-3.55 (m, 1H), 3.35 (dd, *J* = 9.1, 4.2 Hz, 1H), 2.81 (s, 1H), 2.29 (s, 3H). **¹³C-NMR** (400 MHz, CDCl₃) δ 153.91, 138.31, 138.08, 137.80, 133.30, 133.28, 129.81, 129.79, 128.68, 128.66, 128.56, 126.54, 128.26, 128.24, 128.11, 127.95, 127.93, 127.85, 127.83, 95.57, 86.18, 81.91, 77.99, 74.83, 74.55, 73.83, 73.00, 70.64, 56.15, 21.22. HRMS *m/z* calc'd for C₃₀H₃₂NCl₃O₆SNa (M+Na⁺); 662.0913 found: 662.0910.

Compound 22: A solution of **27** (2.3 gm, 3.59 mmol, 1.0 equiv), 5-azidopentanol (4.31 mmol, 1.2 equiv), and activated 4Å powdered molecular sieves (1.5 gm) in anhydrous dichloromethane (12 ml) was stirred for 2h at room temperature under an argon atmosphere, and then cooled to -40°C followed by addition of NIS (0.96 gm, 4.31 mmol, 1.2 equiv) and TfOH (32 μl, 0.359 mmol, 0.1 equiv). The reaction mixture was stirred at -40°C for 0.5 h to 3 h until the disappearance of the donor on TLC, then quenched with triethylamine (270μl, 2.69 mmol, 0.75 equiv) and warmed to room temperature. The mixture was diluted with dichloromethane, filtered through Celite, washed with 20% aqueous Na₂S₂O₃ solution, dried over Na₂SO₄, and concentrated under reduced pressure. The residue was purified by column chromatography on silica gel eluting with Ethylacetate/Hexane (1:1) system to afford the compound **22** (2.1 gm, 91%). **¹H-NMR** (400 MHz, CDCl₃) δ 7.44-7.33 (m, 10H), 5.15 (s, 1H), 4.85-4.70 (m, 3H), 4.72 (d, *J* = 11.7 Hz, 2H), 4.64 (d, *J* = 12.0 Hz, 1H), 4.58 (d, *J* = 12.0 Hz, 1H), 3.91-3.84 (m, 2H), 3.83-3.75 (m, 2H), 3.72 (t, *J* = 8.3 Hz, 1H), 3.55-3.43 (m, 2H), 3.27 (t, *J* = 6.8 Hz, 3H), 2.83 (s, 1H), 1.66-1.57 (m, 4H), 1.43 (m, 2H). **¹³C-NMR** (400 MHz, CDCl₃) δ 153.96, 138.19, 137.59, 128.58, 128.56, 128.51, 128.49, 128.16, 128.18, 127.97, 127.92, 127.80, 127.78, 100.29, 95.53, 80.24, 77.23, 74.44, 73.77, 73.50, 73.49, 70.65, 69.50, 57.54, 51.31, 29.04, 28.56, 23.19. HRMS *m/z* calc'd for C₂₈H₃₅N₄Cl₃O₇S Na (M+Na⁺); 667.1468; found: 667.1467.

Compound 18A solution of **11** (3 gm, 4.62 mmol, 1.0 equiv), dibutylphosphate (6.94 mmol, 1.5 equiv), and activated 4Å powdered molecular sieves (2.2 gm) in anhydrous dichloromethane (20 ml) and acetonitrile (10 ml) was stirred for 2h at room temperature under an argon atmosphere, and then cooled to -20°C followed by addition of NIS (2 gm, 9.28 mmol, 2 equiv) and TfOH (200 μ l, 2.31 mmol, 0.5 equiv). The reaction mixture was stirred at -20°C for 12 h until the disappearance of the donor on TLC, then quenched with triethylamine (81 μ l, 0.81 mmol, 0.75 equiv) and warmed to room temperature. The mixture was diluted with dichloromethane, filtered through celite, washed with 20% aqueous Na₂S₂O₃ solution, dried over Na₂SO₄, and concentrated under reduced pressure. The residue was purified by column chromatography on silica gel eluting with Ethylacetate/Hexane (1:1) system to afford the compound 18 (2 gm, 67%). **α -isomer** **¹H-NMR** (400 MHz, CDCl₃) δ 5.69 (dd, J = 7.5, 1.5 Hz, 1H), 5.33 (ddd, J = 7.5, 6.2, 2.8 Hz, 1H), 4.77 (dd, J = 9.5, 1.5 Hz, 1H), 4.42 (dd, J = 12.3, 2.8 Hz, 1H), 4.26-4.05 (m, 6H), 3.91-3.83 (m, 4H), 3.02 (dd, J = 12.2, 4.0 Hz, 1H), 2.74 (t, J = 12.7 Hz, 1H), 2.51 (s, 3H), 2.16 (s, 3H), 2.12 (s, 3H), 2.05 (s, 3H), 1.74-1.64 (m, 4H), 1.43 (dq, J = 14.7, 7.4 Hz, 4H), 0.95 (t, J = 7.4 Hz, 6H). **¹³C-NMR** (400 MHz, CDCl₃) δ 171.84, 170.62, 169.99, 169.89, 167.30, 167.23, 153.52, 98.22, 98.15, 74.19, 71.48, 69.81, 68.16, 68.10, 68.05, 67.99, 62.51, 58.32, 53.46, 35.91, 35.87, 32.14, 32.10, 32.07, 24.63, 20.96, 20.79, 20.74, 18.63, 18.60, 13.56. HRMS m/z calc'd for C₂₇H₄₂NO₁₆P (M+Na) 690.2139; found: 690.2134.

Glycosylation of Donor and Acceptors:



Scheme 2: Reagents and condition: h) TMSOTf, M.S 4Å, DCM, -78°C.

General procedure of glycosylation : The mixture of donor **18** (0.899 mmol, 1eq), acceptor (0.674 mmol, 0.75 eq) and activated 4Å powdered molecular sieves (1.5 gm) in anhydrous dichloromethane (10 ml) and was stirred for 3h at room temperature under an argon atmosphere, and then cooled to -78°C followed by addition of TMSOTf (165 μ l, 0.899 mmol, 1 equiv). The reaction mixture was stirred at -78°C for 1-3 h until the disappearance of the donor on TLC, then quenched with triethylamine warmed to room temperature. The mixture was diluted with dichloromethane, filtered through celite, washed brine solution, dried over Na₂SO₄, and concentrated under reduced pressure. The residue was purified by column chromatography on silica gel eluting with Ethylacetate/Hexane (1:1) system to afford the sialic acid disaccharide analogues.

Compound 15A According to general procedure donor **18** and acceptor **19** to yield as white solid (700 mg, 75%). ¹H-NMR (400 MHz, CDCl₃) δ 8.05 (d, J = 7.6 Hz, 2H), 7.61 (t, J = 7.4 Hz, 1H), 7.48 (t, J = 7.6 Hz, 2H), 7.41 (d, J = 7.9 Hz, 2H), 7.33 (dt, J = 7.8, 3.6 Hz, 5H), 7.24 – 7.14 (m, 5H), 7.03 (d, J = 7.8 Hz, 2H), 5.66 (t, J = 9.7 Hz, 1H), 5.60 (d, J = 7.0 Hz, 1H), 5.46 (ddd, J = 8.7, 6.7, 3.1, Hz, 1H), 5.02 (d, J = 11.3 Hz, 1H), 4.77 (d, J = 11.3 Hz, 1H), 4.71 (J =

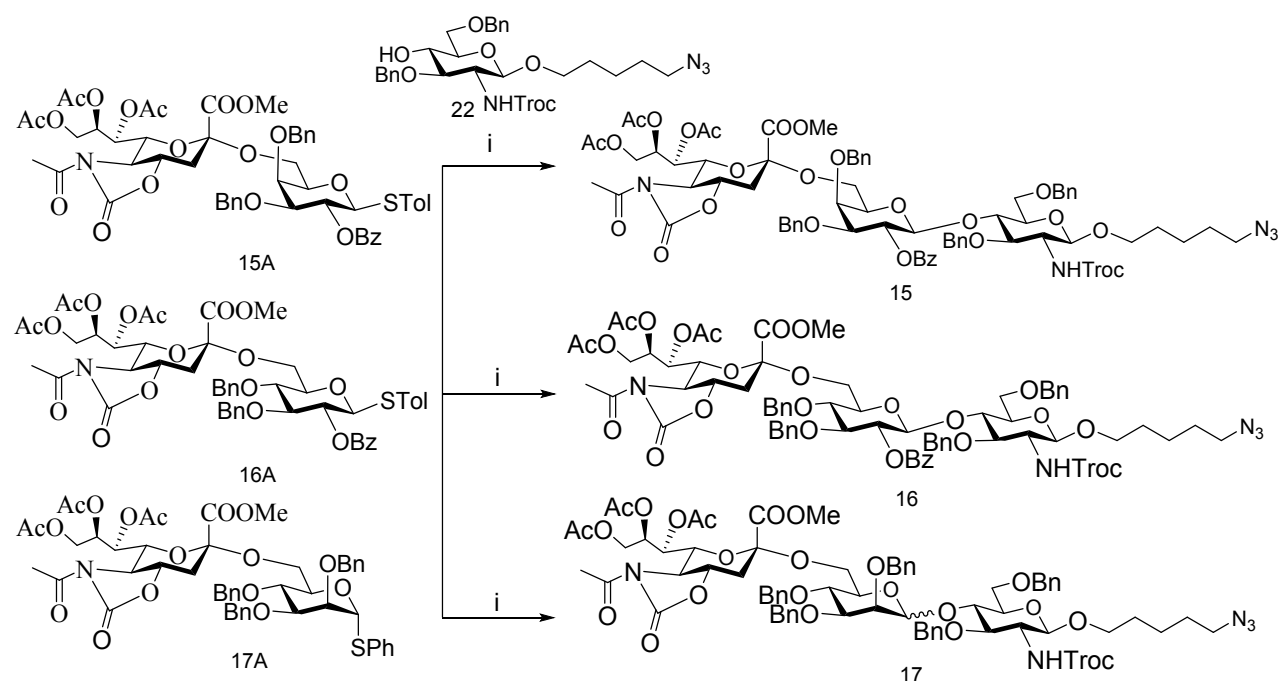
9.4 Hz, 1H), 4.66 (d, J = 11.3 Hz, 1H), 4.62 (d, J = 9.4 Hz, 1H), 4.54 (d, J = 12.2 Hz, 1H), 4.42 (dd, J = 12.2, 2.3 Hz, 1H), 4.13-4.04 (m, 2H), 4.04-3.98 (m, 2H), 3.79-3.71 (m, 3H), 3.69 (s, 3H), 3.66-6.59 (m, 1H), 2.90 (dd, J = 12.0, 3.3 Hz, 1H), 2.52 (s, 3H), 2.31 (s, 3H), 2.15 (s, 3H), 2.09 (t, J = 12.5 Hz, 1H), 2.09 (s, 3H), 2.03 (s, 3H). ^{13}C -NMR (400 MHz, CDCl_3) δ 172.04, 170.79, 170.17, 169.99, 168.28, 165.21, 153.68, 138.61, 137.99, 137.60, 133.01, 132.64, 130.17, 129.89, 129.87, 129.69, 129.44, 129.42, 128.33, 128.32, 128.29, 128.27, 128.16, 128.14, 127.66, 127.64, 127.62, 127.55, 127.58, 127.53, 127.35, 99.21, 86.85, 81.23, 76.85, 75.86, 74.93, 74.07, 72.65, 72.02, 71.95, 70.41, 69.51, 63.85, 62.95, 59.11, 53.13, 36.43, 24.73, 21.15, 21.11, 20.87, 20.77. HRMS m/z calc'd for $\text{C}_{53}\text{H}_{57}\text{NO}_{18}\text{S}$ (M+Na) 1050.3193; found: 1050.3191.

Compound 16A According to general procedure donor **18** and acceptor **20** gave as white solid (765 mg, 83%). ^1H -NMR (400 MHz, CDCl_3) δ 8.04 (t, J = 6.3 Hz, 2H), 7.60 (t, J = 7.4 Hz, 1H), 7.47 (t, J = 7.7 Hz, 2H), 7.40 (t, J = 7.2 Hz, 2H), 7.38-7.30 (m, 5H), 7.17-7.06 (m, 7H), 5.65 (dd, J = 8.6, 1.4 Hz, 1H), 5.51 (ddd, J = 8.8, 6.3, 2.8 Hz, 1H), 5.22 (dd, J = 9.9, 9.2 Hz, 1H), 4.80 (d, J = 10.5 Hz, 1H), 4.78 (d, J = 10.5 Hz, 1H), 4.73-4.65 (m, 3H), 4.64-4.60 (m, 1H), 4.38 (dd, J = 12.4, 2.8 Hz, 1H), 4.24 (dd, J = 11.0, 4.5 Hz, 1H), 4.08-4.00 (m, 2H), 3.82 (m, 4H), 3.77 (m, 1H), 3.71 (t, J = 10.2 Hz, 2H), 3.58 (dd, J = 8.6, 3.4 Hz, 1H), 3.01 (dd, J = 12.0, 3.4 Hz, 1H), 2.52 (s, 3H), 2.35 (s, 3H), 2.17 (s, 3H), 2.13 (t, J = 12.3 Hz, 1H), 2.07 (s, 3H), 1.87 (s, 3H). ^{13}C -NMR (400 MHz, CDCl_3) δ 171.86, 170.65, 170.05, 170.03, 168.37, 165.16, 153.66, 138.11, 138.01, 137.70, 133.16, 132.90, 132.88, 129.92, 129.85, 129.83, 129.48, 129.46, 129.44, 129.31, 128.46, 128.43, 128.41, 128.24, 128.22, 128.20, 128.18, 127.99, 127.96, 127.95, 127.63, 99.31, 86.34, 84.05, 78.01, 77.27, 75.31, 75.24, 75.11, 75.00, 72.24, 71.54, 68.61, 64.53, 62.92, 59.18, 53.09, 36.68, 24.70, 21.20, 21.16, 20.80, 20.62. HRMS m/z calc'd for $\text{C}_{53}\text{H}_{57}\text{NO}_{18}\text{S}$ (M+Na) 1050.3193; found: 1050.3195.

Compound 17A: According to general procedure donor **18** and acceptor **21** to afford as white solid (830 mg, 90%) ^1H NMR (400 MHz, CDCl_3) δ 7.49 (dd, J = 6.4, 3.0 Hz, 2H), 7.33 (m, 18H), 5.62 (dd, J = 8.3, 1.5 Hz, 1H), 5.55 (d, J = 1 Hz, 1H), 5.47 (ddd, J = 8.5, 6.0, 2.9 Hz, 1H), 4.93 (d, J = 10.4 Hz, 1H), 4.78 (d, J = 10.4 Hz, 1H), 4.70 (dd, J = 13.1, 6.9 Hz, 2H), 4.67-4.60 (m, 3H), 4.30 (dt, J = 9.1, 6.3 Hz, 2H), 4.21 (dd, J = 10.8, 6.6 Hz, 1H), 4.07-3.99 (m, 3H), 3.96 (t, J = 9.3 Hz, 1H), 3.88 (dd, J = 9.3, 3.0 Hz, 1H), 3.73-3.68 (m, 2H), 3.68 (s, 3H), 2.92 (dd, J = 12.0, 3.5 Hz, 1H), 2.51 (s, 3H), 2.14 (t, J = 12.6 Hz, 1H), 2.12 (s, 3H), 2.02 (s, 3H), 2.01 (s, 3H).

^{13}C -NMR (400 MHz, CDCl_3) δ 171.85, 170.60, 169.99, 169.82, 168.71, 153.74, 138.63, 138.13, 137.85, 133.95, 132.54, 132.52, 128.92, 128.43, 128.41, 128.40, 128.39, 128.37, 128.30, 128.28, 128.13, 128.10, 127.90, 127.88, 127.83, 127.81, 127.80, 127.79, 127.72, 127.70, 99.41, 85.73, 80.01, 76.13, 75.34, 75.14, 75.05, 74.77, 72.21, 72.13, 71.93, 71.55, 68.66, 65.43, 62.76, 59.16, 52.90, 36.48, 24.72, 21.12, 20.86, 20.77. HRMS m/z calc'd for $\text{C}_{52}\text{H}_{57}\text{NO}_{17}\text{SNa}$ ($\text{M}+\text{Na}$). 1022.3244; found: 1022.3245.

Synthesis of trisaccharide from disaccharide:



Scheme S5: Reagents and condition: i) NIS/TfOH, M.S. 4Å, DCM or ACN, -40°C.

General Glycosylation Procedure : A solution of donor **15A/16A/17A** (500 mg, 0.488 mmol, 1.0 equiv), acceptor **22** (0.585 mmol, 1.2 equiv), and activated 4Å powdered molecular sieves (976 mg, 2.0 g/mmol) in anhydrous dichloromethane (10 ml) was stirred overnight under an argon atmosphere, and then cooled to -40°C followed by addition of NIS (109 mg, 0.488 mmol, 1.0 equiv) and TfOH (6.46 μl , 0.048 mmol, 0.1 equiv) dissolved in ether. The reaction mixture was stirred at -40°C for 20 min to 2 h until the disappearance of the donor on TLC and warmed to room temperature. The mixture was diluted with dichloromethane, filtered through

celite, washed with sat. Na₂S₂O₃ (3:1) 20 ml solution, dried over Na₂SO₄, and concentrated under reduced pressure. The residue was purified by column chromatography on silica gel eluting with Ethylacetate/Hexane system to afford the sialic acid trisaccharide.

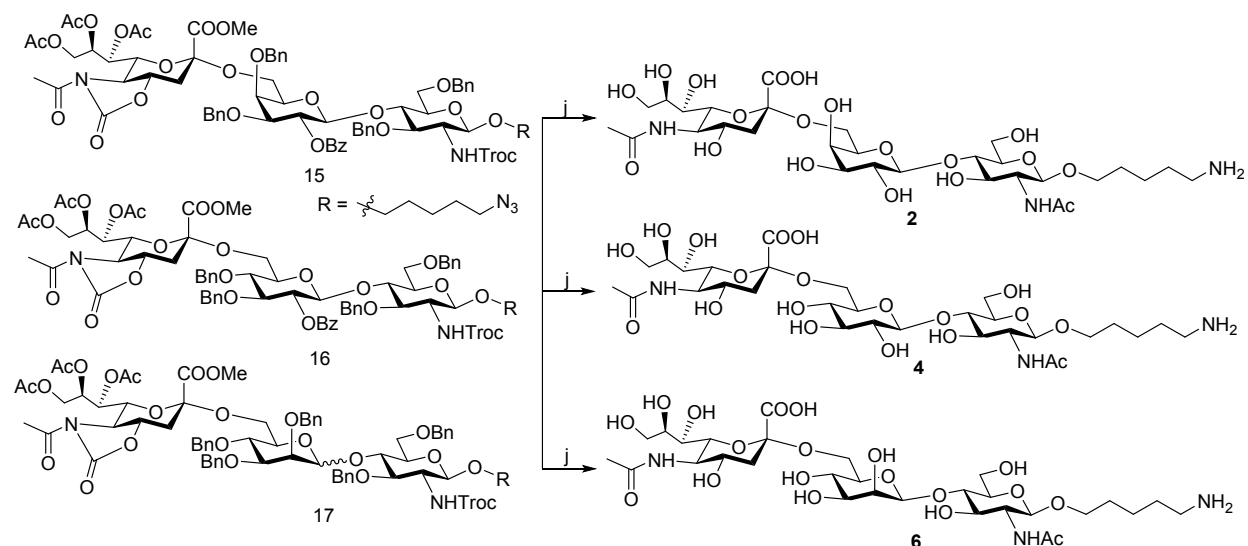
Compound 15 Synthesized from donor **15A** and acceptor **22** by following general glycosylation procedure to afford as white solid (430 mg, 57%) **¹H-NMR** (400 MHz, CDCl₃) δ 7.99 (d, *J* = 7.7 Hz, 2H), 7.61 (t, *J* = 7.4 Hz, 1H), 7.47 (t, *J* = 7.7 Hz, 2H), 7.39 (d, *J* = 7.4 Hz, 2H), 7.37-7.33 (m, 3H), 7.33-7.22 (m, 10H), 7.21-7.14 (m, 5H), 5.70-5.63 (m, 1H), 5.57 (s, 1H), 5.48 (d, *J* = 6.5 Hz, 1H), 5.39 (td, *J* = 6.9, 2.3 Hz, 1H), 5.09 (d, *J* = 11.4 Hz, 1H), 4.97 (d, *J* = 11.1 Hz, 1H), 4.75-4.63 (m, 5H), 4.60 (d, *J* = 12.2 Hz, 1H), 4.52 (d, *J* = 12.2 Hz, 2H), 4.47-4.39 (m, 2H), 4.33 (d, *J* = 12.2 Hz, 2H), 4.05 (dd, *J* = 12.2, 7.2 Hz, 1H), 4.01-3.91 (m, 4H), 3.87 (d, *J* = 6.3 Hz, 2H), 3.76 (m, 2H), 3.68-3.65 (m, 1H), 3.64 (s, 3H), 3.60 (d, *J* = 3.1 Hz, 1H), 3.56 (dd, *J* = 10.1, 2.1 Hz, 1H), 3.45 (dd, *J* = 12.9, 7.1 Hz, 1H), 3.34 (m, 3H), 3.24 (t, *J* = 6.9 Hz, 2H), 2.73 (dd, *J* = 12.3, 3.7 Hz, 1H), 2.51 (s, 3H), 2.17 (t, *J* = 12.6 Hz, 1H), 2.13 (s, 3H), 2.03 (s, 3H), 2.02 (s, 3H), 1.62-1.50 (m, 4H), 1.38 (m, 2H). **¹³C-NMR** (400 MHz, CDCl₃) δ 172.19, 170.85, 170.31, 169.85, 168.09, 165.17, 153.98, 153.80, 138.75, 138.44, 138.29, 137.57, 133.20, 133.17, 135.15, 129.87, 129.78, 129.76, 129.77, 128.46, 128.44, 128.42, 128.39, 128.40, 128.37, 128.36, 128.34, 128.32, 128.30, 128.03, 127.75, 127.71, 127.69, 127.68, 127.63, 127.60, 127.52, 127.32, 100.50, 99.75, 99.23, 95.65, 79.82, 77.24, 76.03, 75.57, 74.91, 74.64, 74.35, 74.26, 74.15, 73.45, 73.30, 72.80, 72.40, 72.16, 71.89, 69.96, 69.34, 68.47, 63.86, 62.97, 58.83, 57.20, 53.12, 51.31, 35.54, 29.00, 28.53, 24.74, 23.15, 21.13, 20.84, 20.73. HRMS *m/z* calc'd for C₇₄H₈₄Cl₃N₅O₂₅ (M+Na). 1570.4418; found: 1570.4411.

Compound 16 Synthesized from donor **16A** and acceptor **22** by following general glycosylation procedure to afford as white solid (340 mg, 45%) **¹H-NMR** (400 MHz, CDCl₃) δ 7.93 (d, *J* = 7.4 Hz, 2H), 7.59 (t, *J* = 7.4 Hz, 1H), 7.48-7.30 (m, 17H), 7.18-7.05 (m, 5H), 5.59 (dd, *J* = 9.0, 1.4 Hz, 1H), 5.48 (ddd, *J* = 8.9, 6.2, 2.7 Hz, 1H), 5.26 (dd, *J* = 9.4, 8.2 Hz, 1H), 4.98-4.88 (m, 4H), 4.80 (d, *J* = 12.9 Hz, 2H), 4.75 (d, *J* = 10.5 Hz, 1H), 4.71 (d, *J* = 3.4 Hz, 2H), 4.68 (s, 2H), 4.66 (d, *J* = 1.4 Hz, 1H), 4.58 (d, *J* = 11.2 Hz, 2H), 4.40 (d, *J* = 12.2 Hz, 2H), 4.33 (dd, *J* = 12.2, 2.7 Hz, 1H), 4.30-4.25 (m, 3H), 4.01 (dd, *J* = 11.5, 6.2 Hz, 2H), 3.95 (dd, *J* = 12.5, 6.2 Hz, 2H), 3.86 (t, *J* = 9.4 Hz, 1H), 3.73 (s, 3H), 3.68-3.62 (m, 3H), 3.56-3.47 (m, 4H), 3.25 (m, 4H), 2.73

(dd, $J = 12.0, 3.3$ Hz, 1H), 2.49 (s, 3H), 2.18 (s, 3H), 2.07 (s, 3H), 2.07 (t, $J = 12.3$ Hz, 1H), 1.60-1.48 (m, 4H), 1.41-1.26 (m, 2H). $^{13}\text{C-NMR}$ (400 MHz, CDCl_3) δ 171.90, 170.69, 170.00, 169.94, 168.38, 165.03, 154.03, 153.65, 138.92, 138.17, 137.94, 137.84, 133.24, 129.71, 129.63, 128.61, 128.48, 128.31, 128.27, 128.20, 128.15, 128.07, 127.96, 127.71, 127.83, 127.68, 127.58, 127.56, 127.53, 100.93, 99.46, 99.26, 95.70, 82.17, 78.27, 77.23, 75.72, 75.14, 74.95, 74.92, 74.53, 74.45, 74.09, 73.85, 73.65, 73.48, 71.39, 69.18, 68.32, 67.94, 64.37, 63.96, 62.93, 59.11, 57.11, 53.11, 51.30, 36.52, 28.96, 28.51, 24.67, 23.14, 21.18, 20.79, 20.34. HRMS m/z calc'd for $\text{C}_{74}\text{H}_{84}\text{Cl}_3\text{N}_5\text{O}_{25}$ (M+Na). 1570.4418; found: 1570.4411.

Compound 17 Synthesized from donor **17A**(0.502 mmol) and acceptor **22**(0.603 mmol) by following general glycosylation procedure in DCM:ACN (1:1) solvent to afford as white solid (410 mg, 53%); $\alpha:\beta$ (1:3). **β -isomer** $^1\text{H-NMR}$ (500 MHz, CDCl_3) δ 7.47-7.21(m, 25H) 5.65 (dd, $J = 8.6, 1.4$ Hz, 1H), 5.50 (ddd, $J = 10.5, 5.3, 2.4$ Hz, 1H), 5.35 (m, 1H), 5.24 (d, $J = 7.0$ Hz, 1H), 4.90 (d, $J = 10.4$ Hz, 1H), 4.83-4.87 (m, 1H), 4.76 (d, $J = 7.8$ Hz, 2H), 4.73 (d, $J = 3.9$ Hz, 1H), 4.70 (d, $J = 3.2$ Hz, 1H), 4.68 (d, $J = 1.8$ Hz, 1H), 4.64 (d, $J = 2.9$ Hz, 1H), 4.61 (s, 1H), 4.58 (t, $J = 10.6$ Hz, 1H), 4.46 (d, $J = 12.1$ Hz, 1H), 4.35 (d, $J = 12.1$ Hz, 1H), 4.31 (dd, $J = 12.4, 2.9$ Hz, 1H), 4.29 (d, $J = 2.8$ Hz, 1H), 4.23 (dt, $J = 7.5, 2.8$ Hz, 1H), 4.04 (dd, $J = 6.2, 3.6$ Hz, 1H), 4.02-3.96 (m, 2H), 3.94 (dd, $J = 9.3, 4.2$ Hz, 2H), 3.91-3.80 (m, 4H), 3.79-3.68 (m, 3H), 3.64 (d, $J = 9.8$ Hz, 1H), 3.61 (s, 3H), 3.58 (d, $J = 9.8$, 1H), 3.55-3.49 (m, 1H), 3.45-3.38 (m, 1H), 3.30 (t, $J = 6.9$ Hz, 2H), 2.93 (dd, $J = 7.3, 1.8$ Hz, 1H), 2.88 (dd, $J = 12.0, 3.5$ Hz, 1H), 2.54 (s, 3H), 2.15 (s, 3H), 2.07 (t, $J = 12.5$ Hz, 1H), 2.05 (s, 3H), 1.98 (s, 3H), 1.69-1.59 (m, 4H) 1.45-1.41 (m, 2H). $^{13}\text{C-NMR}$ (400 MHz, CDCl_3) δ 171.65, 170.43, 169.78, 169.55, 168.44, 153.74, 153.53, 138.41, 138.19, 138.13, 137.96, 137.59, 137.43, 133.77, 132.36, 131.69, 128.76, 128.74, 128.72, 128.70, 128.66, 128.64, 128.60, 128.59, 128.32, 128.16, 127.97, 127.80, 127.72, 127.60, 127.54, 127.46, 127.38, 127.25, 127.13, 127.08, 126.85, 101.46, 99.65, 99.20, 95.30, 85.55, 80.74, 79.35, 75.66, 75.07, 74.92, 74.75, 74.52, 74.38, 74.16, 73.60, 73.09, 72.18, 71.99, 71.27, 69.54, 69.26, 68.31, 65.41, 65.24, 62.52, 58.93, 57.63, 52.66, 51.14, 36.34, 29.18, 28.87, 24.53, 23.23, 22.52, 20.95, 20.58. HRMS m/z calc'd for $\text{C}_{74}\text{H}_{86}\text{Cl}_3\text{N}_5\text{O}_{24}$ (M+Na). 1556.4626; found: 1556.4624.

Global Deprotection of Sialoside:



Scheme3: Reagents and Conditions:(j) (i) LiOH, EtOH:H₂O (3:1), 80°C, 12h; (ii) NaHCO₃, Ac₂O, H₂O, rt; (iii) H₂/Pd (OH)₂, MeOH:H₂O (1:1).

General Deprotection Procedure: Oligosaccharide (1eq) in ethanol water (3:1) 5ml at room temperature. After stirring at 80°C for 12h, the reaction mixture cooled to room temperature and carefully neutralized by IR-120H⁺ resin to pH-7, diluted with methanol, filtered and concentrated. Crude residue was dissolved in water and NaHCO₃ (15 eq) were added to it. Cooled the reaction mixture to 0°C and Acetic-anhydride (10 eq) was added. The reaction were monitored by TLC (ethylacetate:methanol:water 7:2:1), after 3h, LiOH (10 eq) were added and stirred for another 5-6h at room temperature, then reaction mixture carefully neutralized by IR-120H⁺ resin to pH-7, diluted concentrated and purified by reverse-phase column chromatography (Bond Elu-C18). The Pd(OH)₂ (1 mmole) was added to the above residue in methanol water (1:1) 4ml. The reaction mixture was hydrogenolyzed under H₂ atmosphere for 12h, filtered and concentrated. The residue was purified by reverse phase column chromatography (Bond Elu-C18) to give deprotected sialic acid analogues.

Compound 2 Synthesized from **15** by following general deprotection protocol to afford white solid (50 mg, 50%). ¹H-NMR (400 MHz, D₂O) δ 4.47 (d, *J* = 7.3 Hz, 1H), 4.36 (d, *J* = 7.9 Hz, 1H), 3.97-3.87 (m, 3H), 3.86-3.79 (m, 4H), 3.78-3.67 (m, 4H), 3.64 (dd, *J* = 6.3, 3.9 Hz, 3H), 3.61-3.51 (m, 4H), 3.46 (m, 3H), 2.96-2.88 (m, 2H), 2.58 (dd, *J* = 12.4, 4.7 Hz, 1H), 1.97 (s, 3H), 1.94 (s, 3H), 1.66-1.57 (m, 3H), 1.56-1.48 (m, 2H), 1.37-1.29 (m, 2H). ¹³C-NMR (400

MHz, D₂O) δ 174.68, 174.42, 173.50, 103.45, 100.91, 100.13, 80.74, 74.46, 73.68, 72.53, 72.43, 72.41, 71.69, 70.71, 68.39, 68.35, 68.21, 63.35, 62.64, 60.36, 54.87, 51.87, 42.57, 40.06, 39.33, 28.04, 26.34, 25.01, 22.27, 22.03. HRMS m/z calc'd for C₃₀H₅₁N₅O₁₉(M+H). 760.3351; found: 760.3358.

Compound 4 Synthesized from **16** by following general deprotection protocol to afford white solid (40 mg, 35%). **¹H-NMR** (400 MHz, D₂O) δ 4.43 (d, J = 8.2 Hz, 1H), 4.35 (d, J = 8.0 Hz, 1H), 3.92-3.87 (m, 1H), 3.85-3.74 (m, 5H), 3.71 (dd, J = 10.0, 3.4 Hz, 1H), 3.67-3.56 (m, 4H), 3.56-3.48 (m, 4H), 3.42 (d, J = 9.2 Hz, 2H), 3.39-3.34 (m, 1H), 3.03-2.96 (m, 2H), 2.93 (dd, J = 7.8, 5.3 Hz, 2H), 2.69 (dd, J = 7.9, 4.8 Hz, 1H), 2.60 (dd, J = 12.3, 4.3 Hz, 1H), 1.96 (s, 3H), 1.95 (s, 3H), 1.61-1.50 (m, 4H), 1.49-1.45 (m, 1H), 1.35 -1.21(m, 2H). **¹³C-NMR** (400 MHz, D₂O) δ 174.94, 174.39, 173.44, 102.96, 100.92, 100.01, 80.69, 75.35, 74.44, 74.24, 72.96, 72.48, 71.68, 70.00, 69.78, 68.32, 68.18, 63.19, 62.59, 60.28, 54.92, 51.85, 42.72, 39.94, 39.04, 28.06, 25.27, 23.04, 22.24, 22.01. HRMS m/z calc'd for C₃₀H₅₁N₅O₁₉(M+H). 760.3351; found: 760.3358.

Compound 6 Synthesized from **17** by following general deprotection protocol to afford white solid (35 mg, 30%). **¹H-NMR** (400 MHz, D₂O) δ 5.06 (d, J = 0.8 Hz, 1H), 4.37 (d, J = 8.2 Hz, 1H), 3.90 (m, 1H), 3.82 (dd, J = 10.7, 4.4 Hz, 2H), 3.79-3.72 (m, 2H), 3.73 (d, J = 2.4 Hz, 1H), 3.70 (d, J = 3.5 Hz, 1H), 3.66 (dd, J = 7.2, 2.2 Hz, 1H), 3.63 (d, J = 3.7 Hz, 2H), 3.59 (d, J = 1.6 Hz, 2H), 3.57 (d, J = 1.6 Hz, 2H), 3.55 (d, J = 4.7 Hz, 2H), 3.53 (d, J = 7.6 Hz, 2H), 3.51-3.48 (m, 2H), 3.46 (d, J = 9.0 Hz, 2H), 2.88-2.83 2.86 (m, 1H), 2.62 (dd, J = 12.5, 4.7 Hz, 1H), 1.91(s, 6H), 1.63-1.50 (m, 4H), 1.50-1.45 (m, 1H), 1.35-1.22 (m, 2H). **¹³C-NMR** (400 MHz, D₂O) δ 181.55, 175.05, 173.58, 101.70, 100.96, 100.17, 74.64, 74.03, 72.61, 72.34, 71.61, 70.60, 70.20, 69.90, 68.26, 66.61, 66.53, 63.31, 62.62, 60.89, 55.74, 51.89, 51.91, 40.08, 39.36, 27.94, 26.35, 26.32, 22.06, 22.01. HRMS m/z calc'd for C₃₀H₅₁N₅O₁₉(M+H). 760.3351; found: 760.3344.

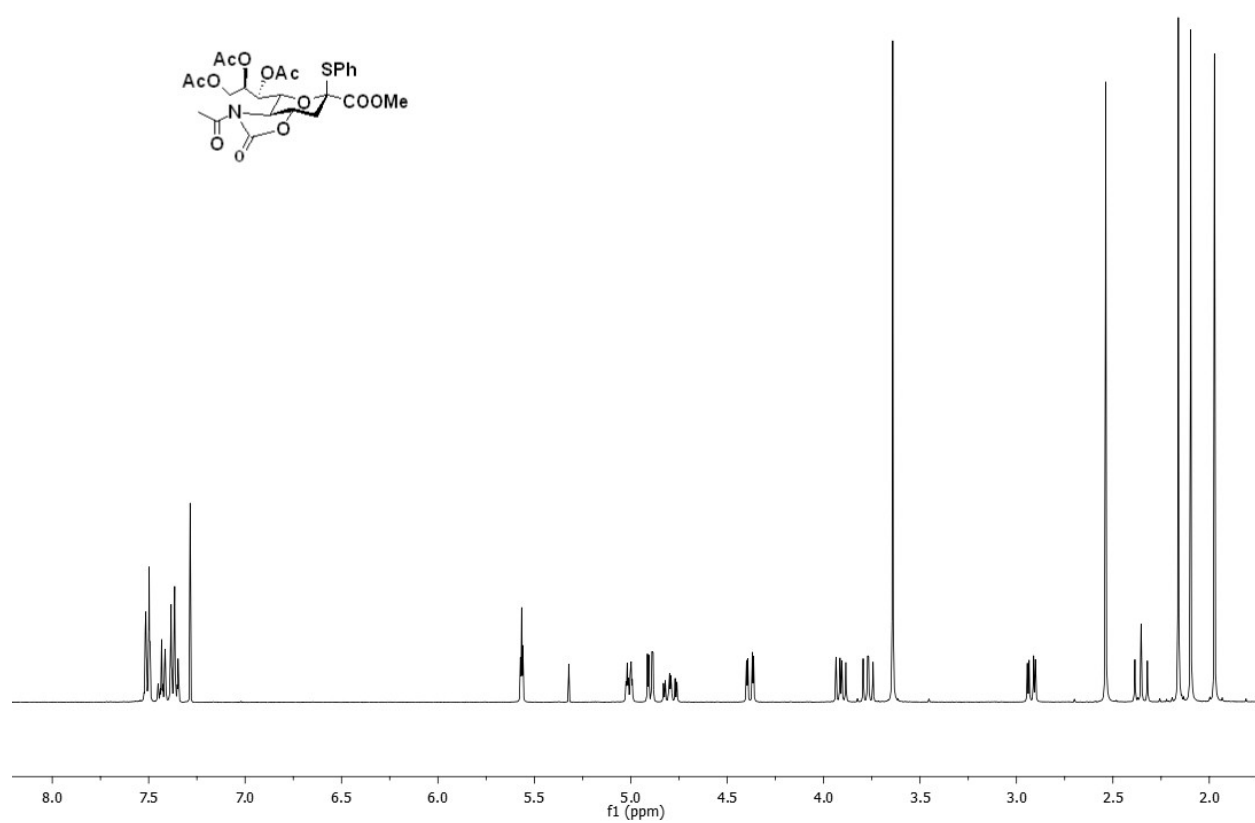
4. Glycanmicroarraydetails

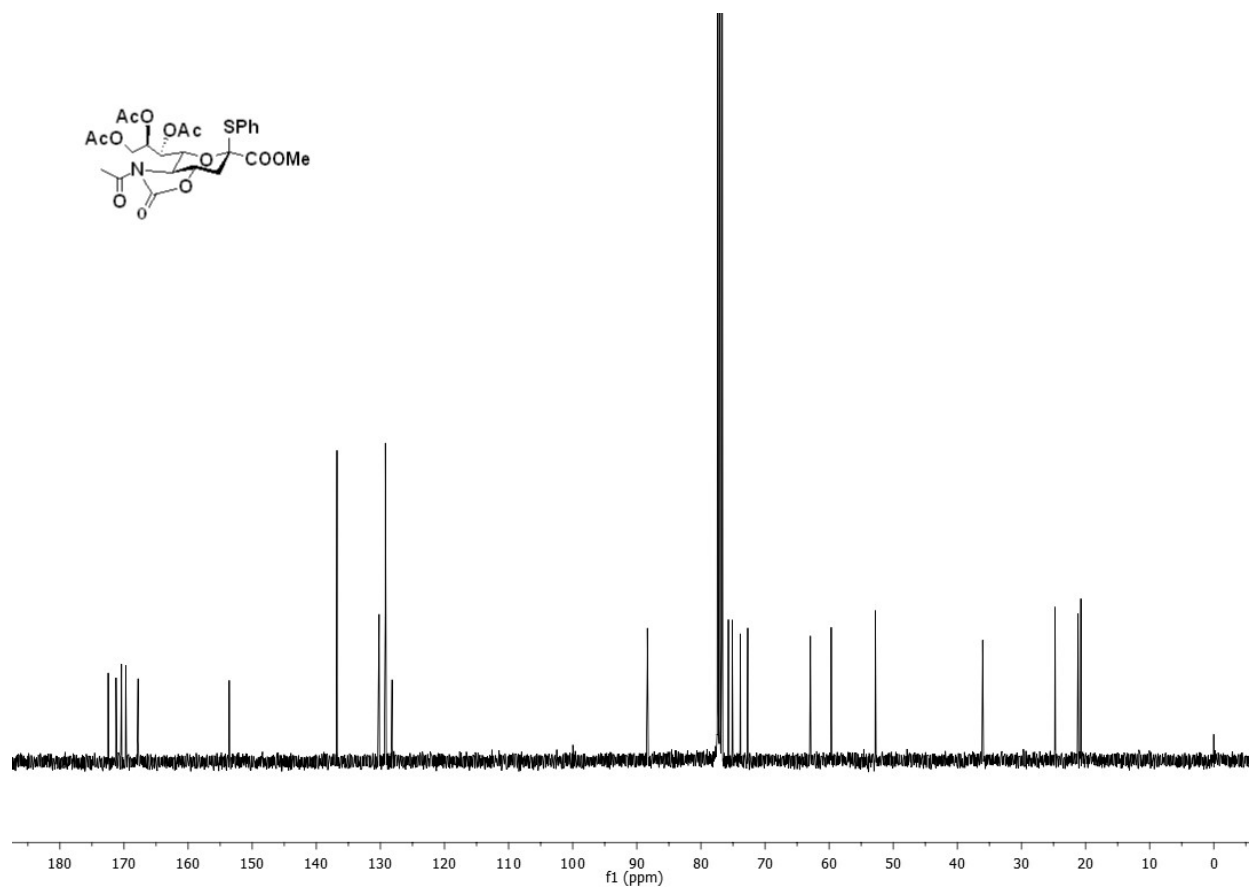
Sialoglycan microarray fabrication. Arrays were printed on Epoxide-derivatized Corning slides as described for Array 1 in Ref. {Padler-Karavani et al., 2012, J BiolChem} with some modifications. Arrays were fabricated with NanoPrint LM 60Microarray Printer (Arrayit, CA) on epoxide-derivatized slides (Corning) with 16 sub-array blocks on each slide. Glycoconjugates were distributed into one 384-well source plates using 4 replicate wells per sample and 8 μ l per well. Each glycoconjugate was prepared at 100 μ M in an optimized print buffer (300 mM phosphate buffer, pH 8.4). The arrays were printed with four 946MP3 pins (5 μ m tip, 0.25 μ l sample channel, \sim 100 μ m spot diameter; Arrayit, CA) with spot to spot spacing of 225 μ m. The humidity level in the arraying chamber was maintained at about 66% during printing. Printed slides were left on arrayer deck over-night, allowing humidity to drop to ambient levels (40-45%). Next, slides were packed, vacuum-sealed and stored in a desiccant chamber at room temperature(RT) until used.

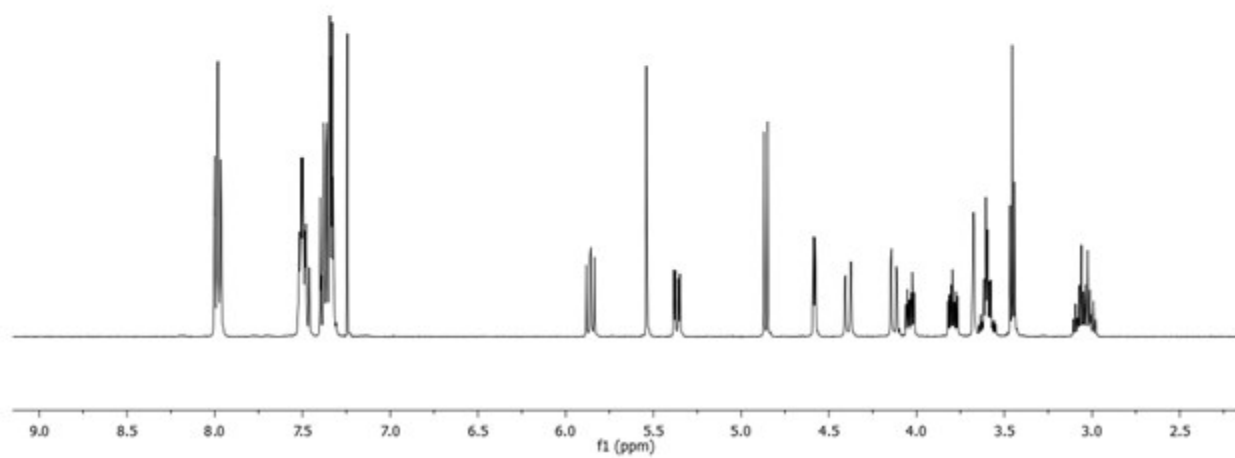
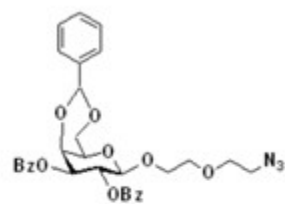
Microarray binding assay. Slides were developed {23520510} and analyzed {Padler-Karavani et al., 2012, J BiolChem} as previously described. Slides were rehydrated with dH₂O and incubated for 30 min in a staining dish with 50°C pre-warmed 0.05 M ethanolamine in 0.1 M Tris, pH 9.0 to block the remaining reactive epoxy groups on the slide surface, then washed with 50°C pre-warmed dH₂O. Slides were centrifuged at 200 \times g for 3 min then fitted with ProPlate™ Multi-Array 16-well slide module (Invitrogen) to divide into the sub-arrays (blocks). Slides were washed with PBST(PBS pH 7.4, 0.1% Tween-20), aspirated and blocked with 200 μ l/sub-array of blocking buffer (PBS pH 7.4, 1% ovalbumin;PBS/OVA) for 1 hour at RT with gentle shaking. Next, the blocking solution was aspirated and 200 μ l/ block of primary detectionwas added (Siglec-human IgGFc chimeras; R&D). Primary detections were incubated with gentle shaking for 2 hours at RT. Slides were washed three times with PBST then with PBS for 5 min/wash with shaking. Bound antibodies were detected by incubating with secondary detection diluted in PBS, 200 μ l/block at RT for 1 hour: Cy3-anti-human-IgG (Jackson; 0.4 μ g/ml). Slides were washed three times with PBST then with PBS 5 min/wash followed by removal from ProPlate™ Multi-Array slide module and immediately dipping slide in a staining dish with dH₂O for 10 min with shaking, then centrifuged at 200 \times g for 5 min. Dry slides were vacuum-sealed and stored in dark until scanning.

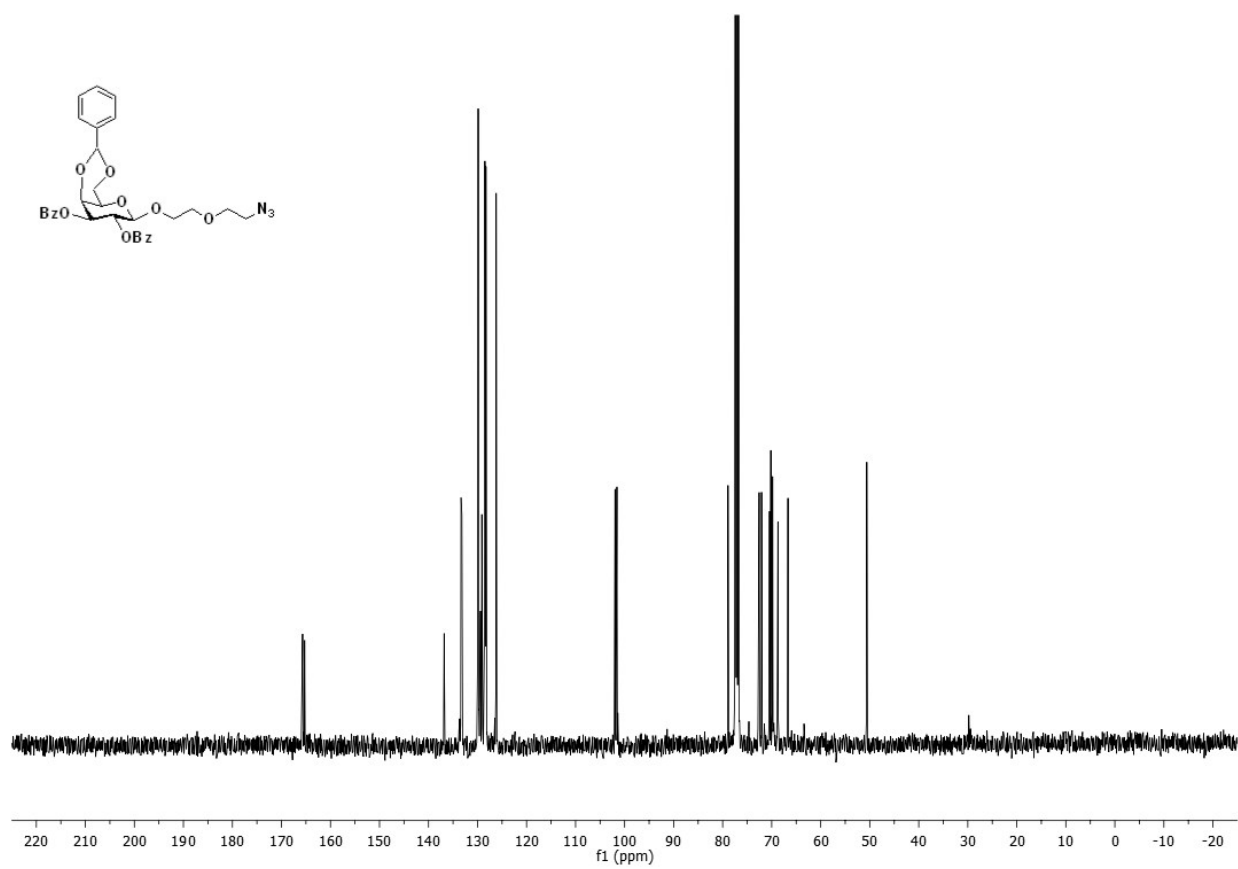
Array slide processing. Processed slides were scanned and analyzed as described {Padler-Karavani et al., 2011, Cancer Res, 71, 3352-63; Padler-Karavani et al., 2012, J BiolChem} at 10 μm resolution with a Genepix 4000B microarray scanner (Molecular Devices) using 350 gain. Image analysis was carried out with Genepix Pro 6.0 analysis software (Molecular Devices). Spots were defined as circular features with a variable radius as determined by the Genepix scanning software. Local background subtraction was performed. Data was analyzed by Excel using pivot tables.

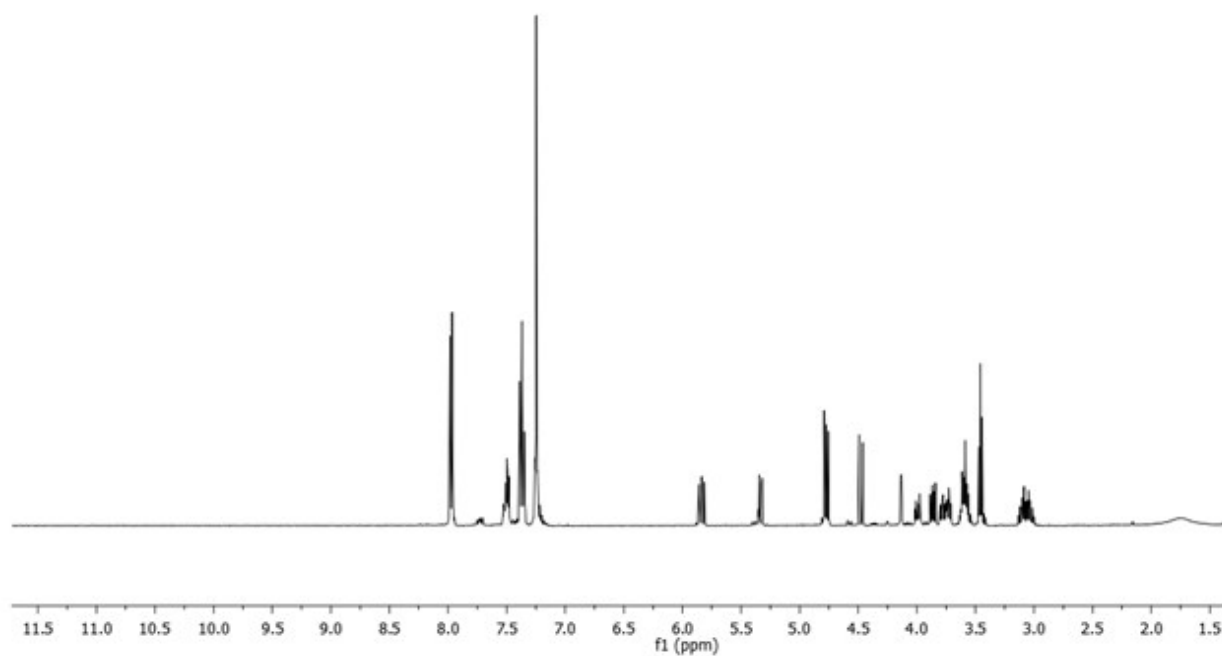
5 ^1H , ^{13}C -NMR, Mass Spectrometry Data of all the Compounds

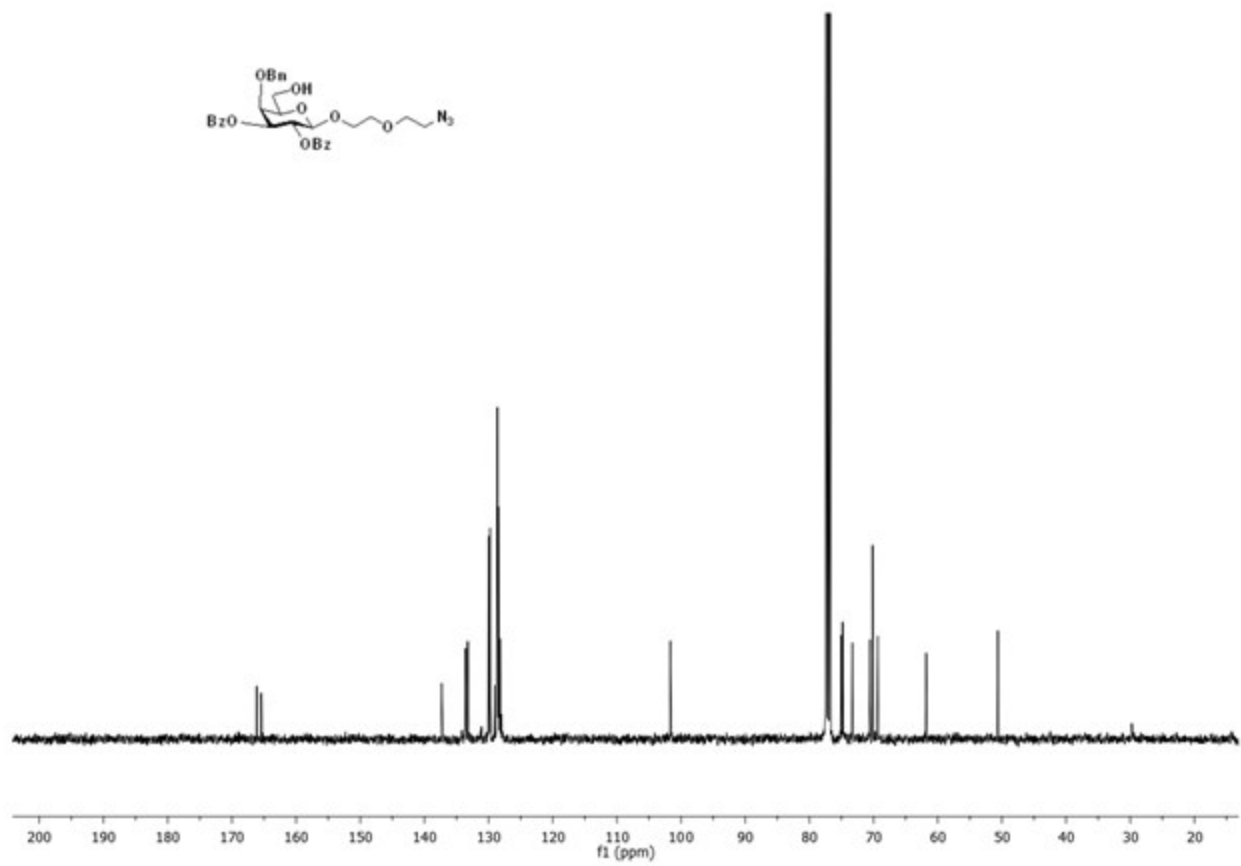
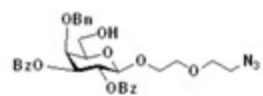








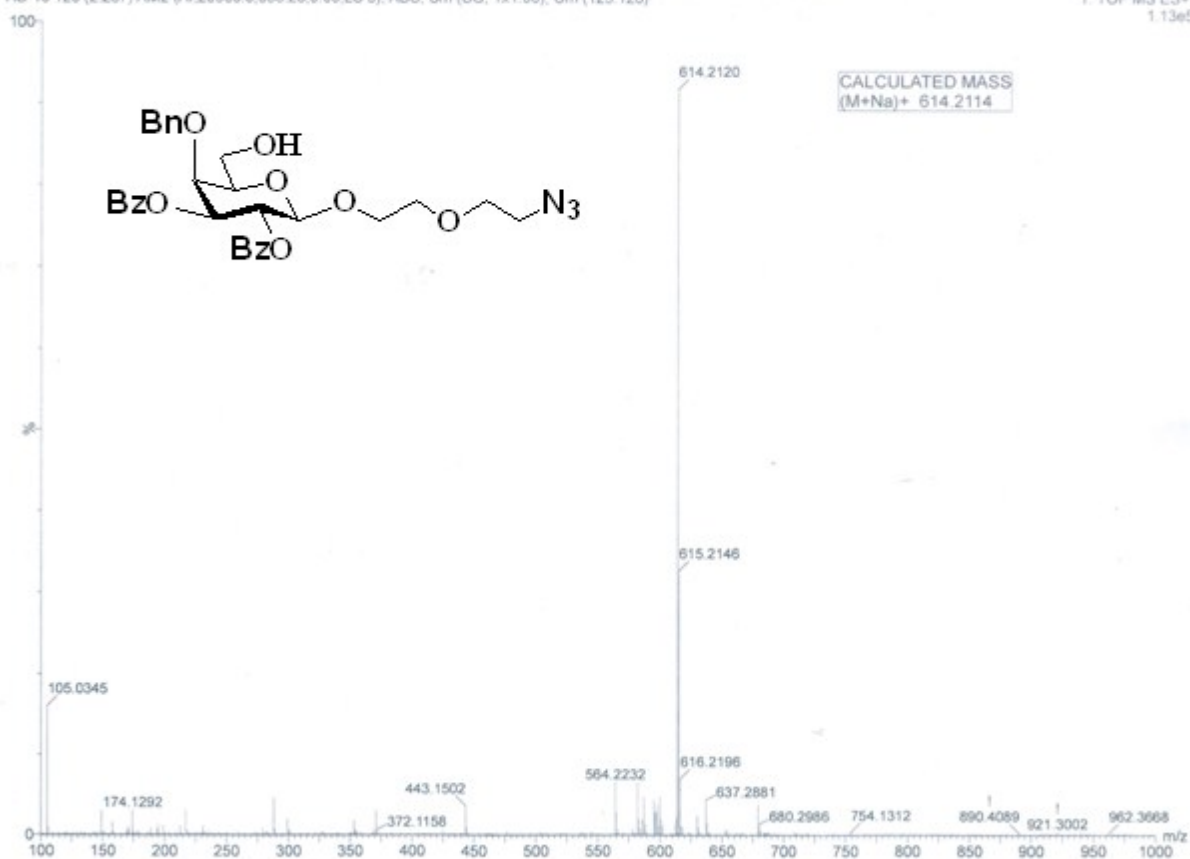


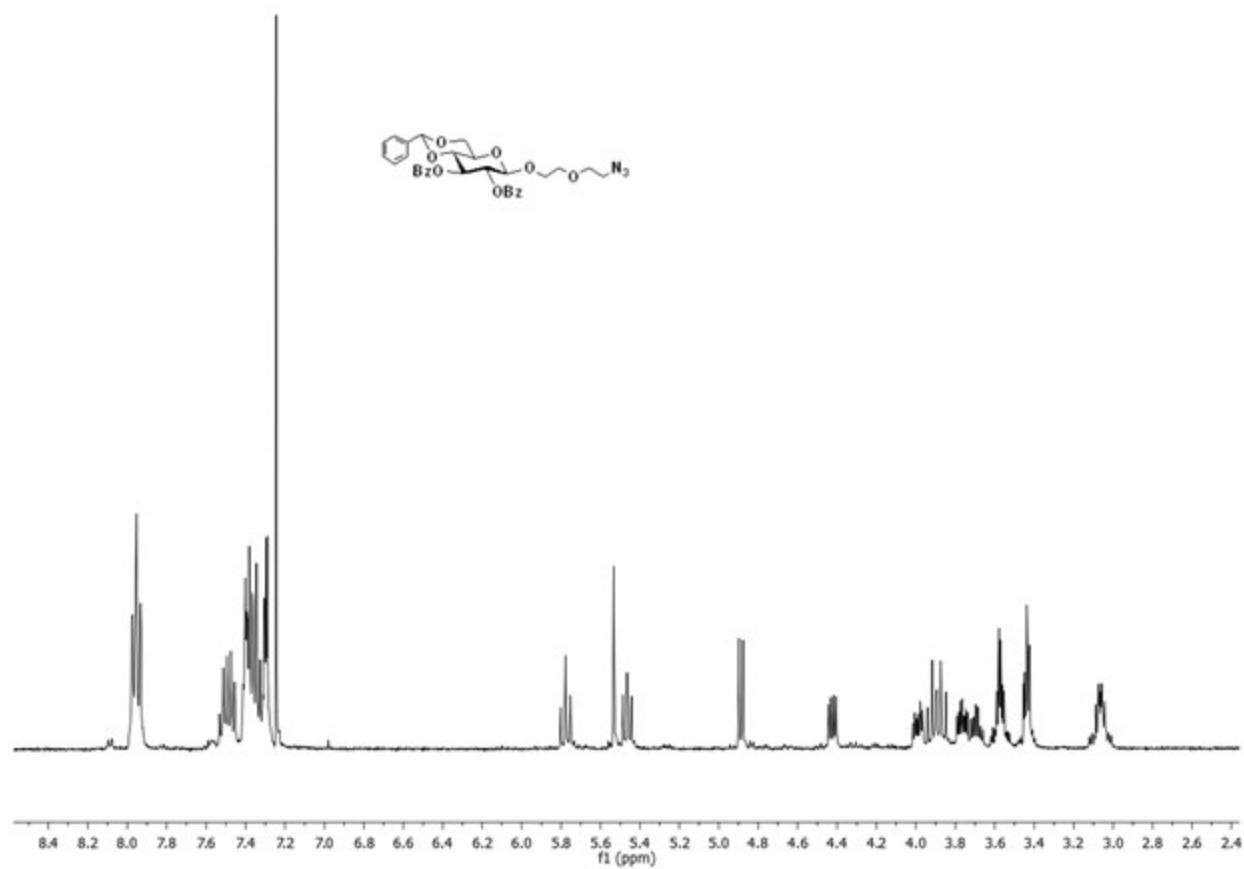


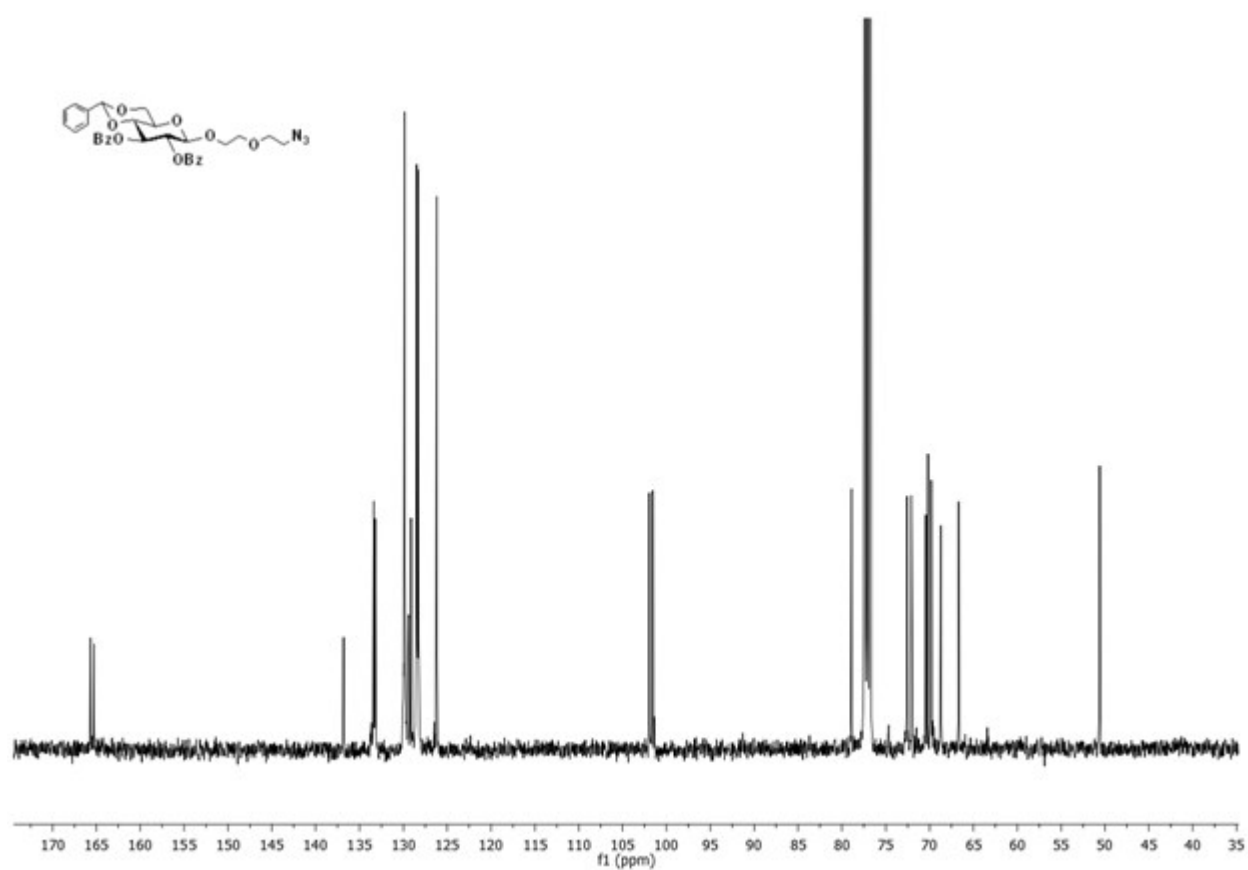
RD 15

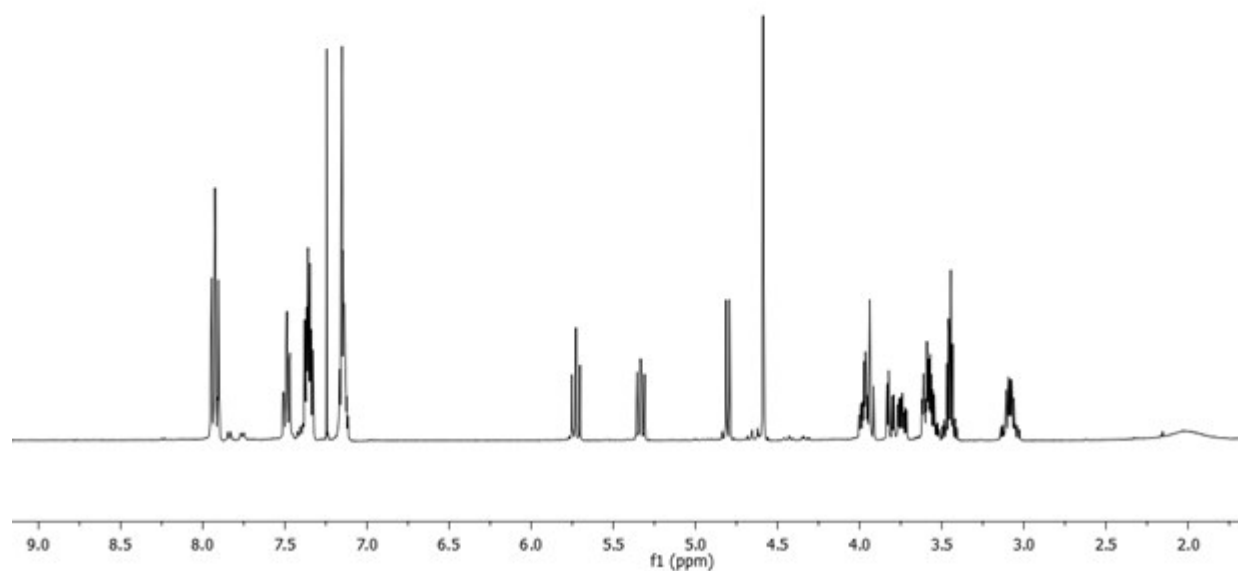
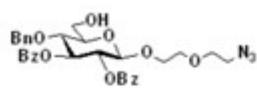
IISER PUNE

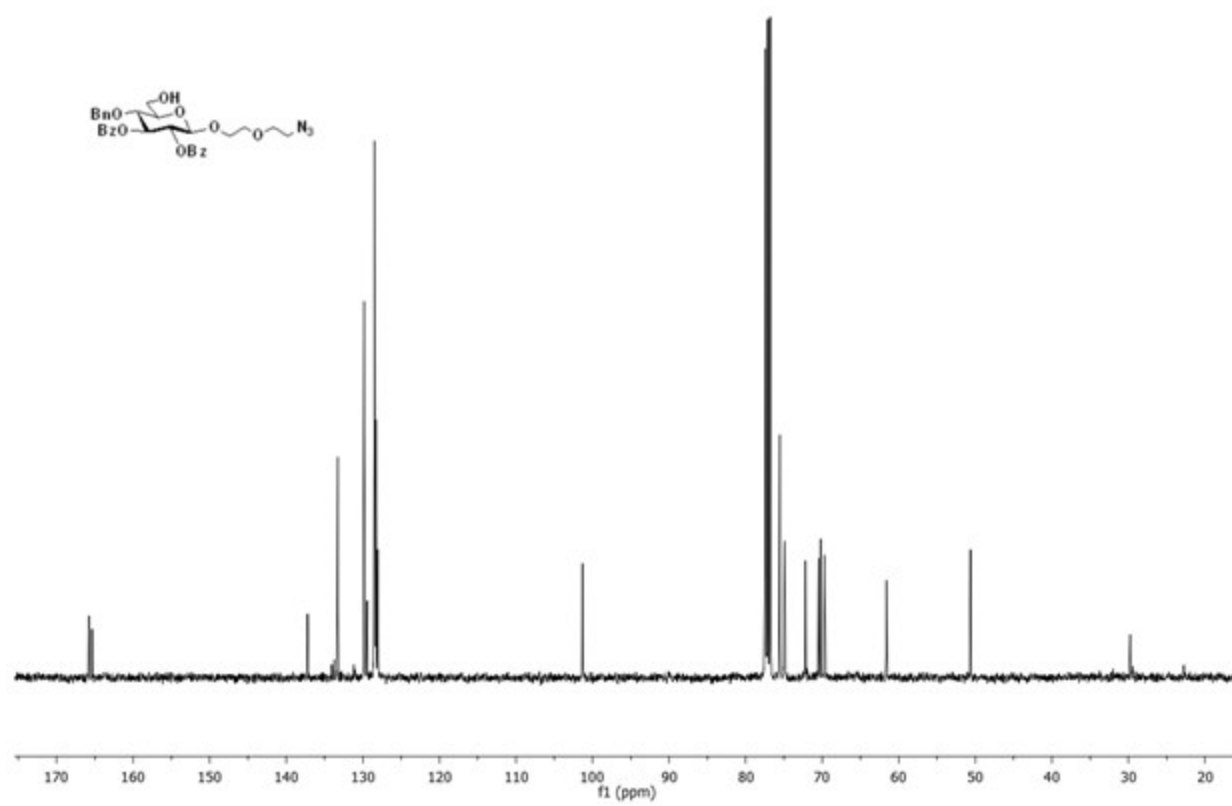
RD 15 125 (2.287) AM2 (Ar.20000.0.556.28.0.00.LS.3); ABS: Sm (SG, 1x1.00); Cm (123.126)

1. TOF MS ES+
1.13e5





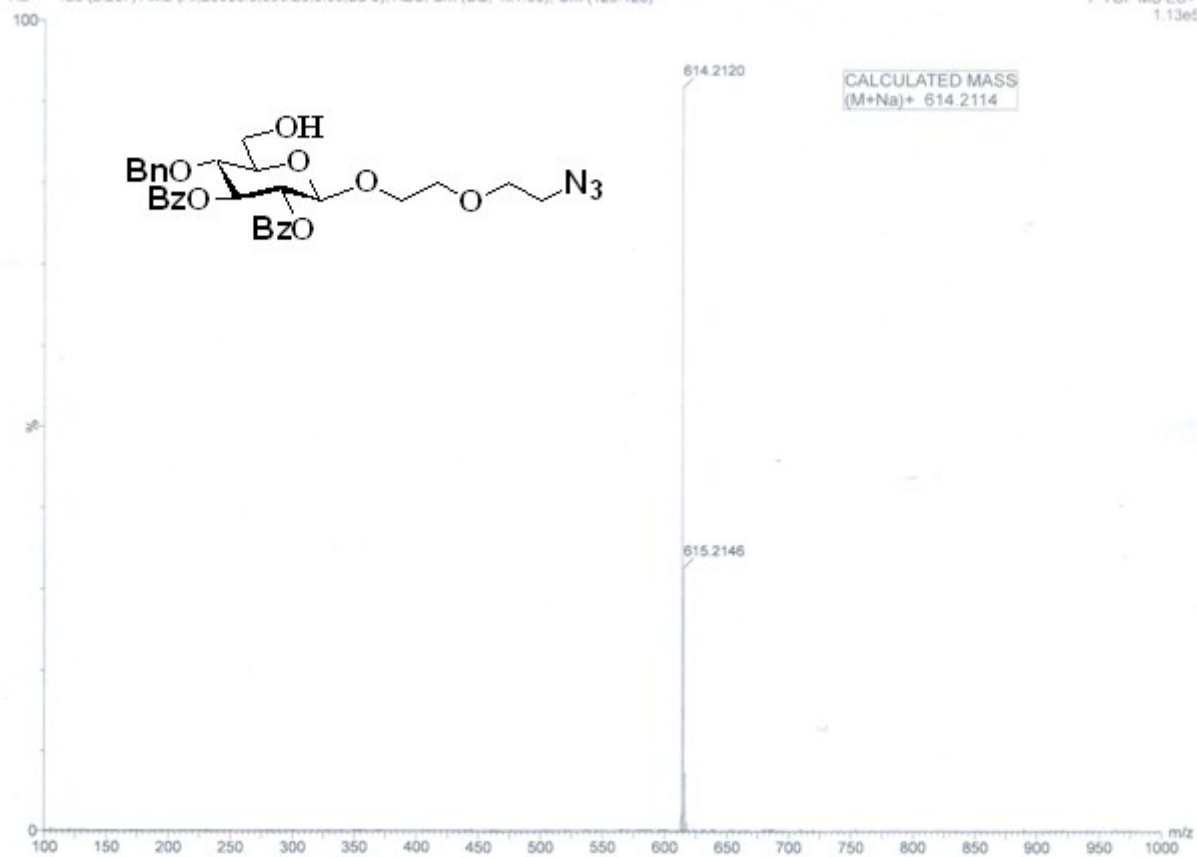


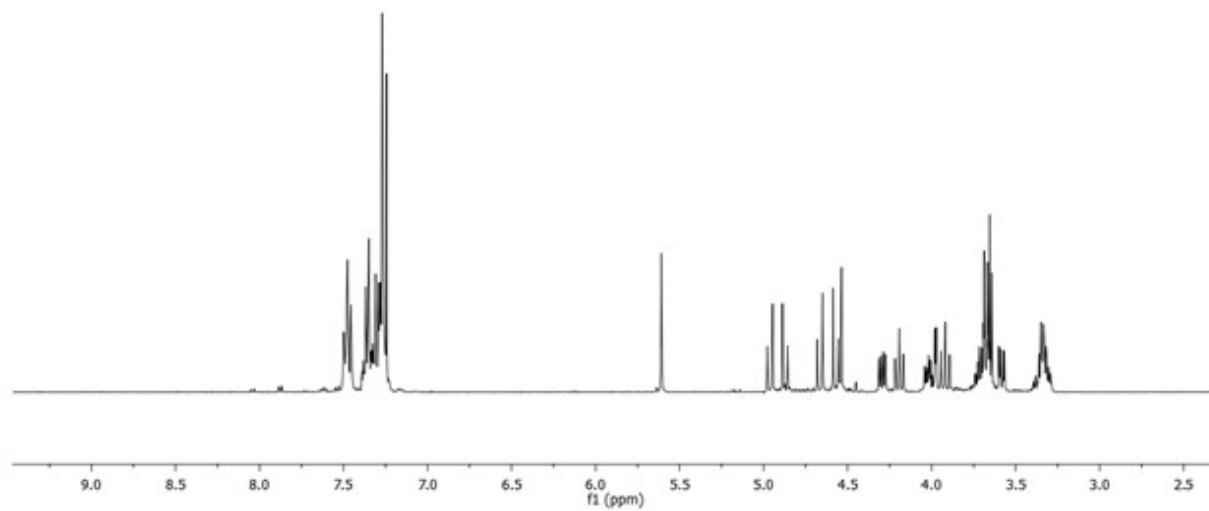
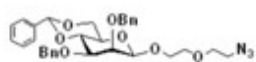


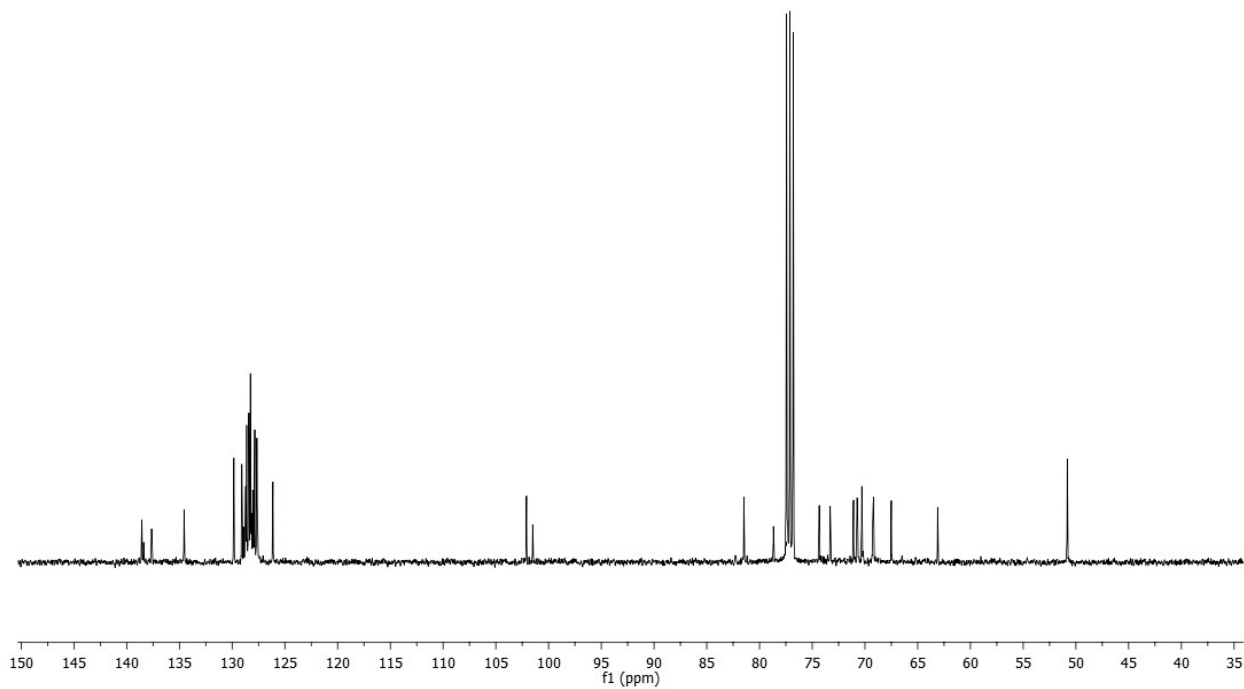
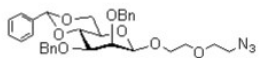
RD
RD 125 (2.287) AM2 (Ar.20000.0.556.28.0.00.LS.3); ABS; Sm (SG, 1x1.00); Cm (123.126)

IISER PUNE

1: TOF MS ES+
1.13e5





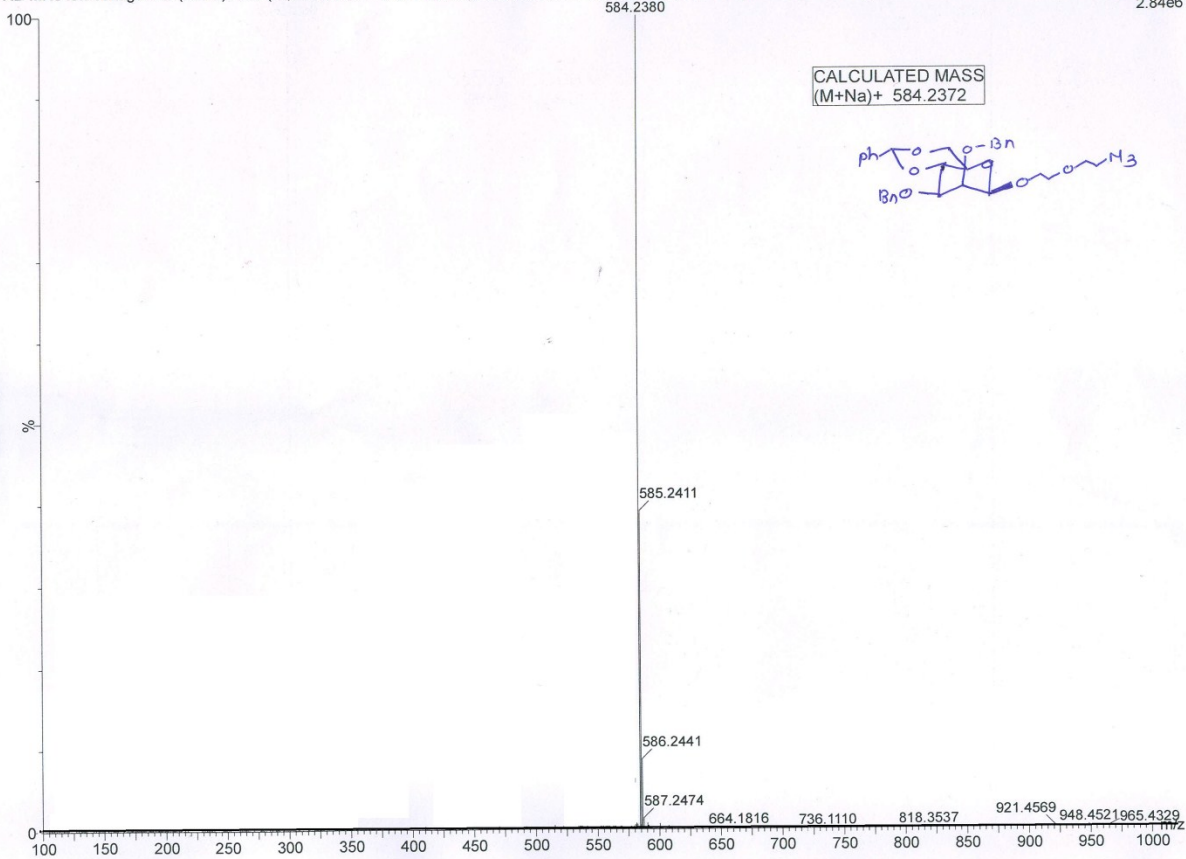


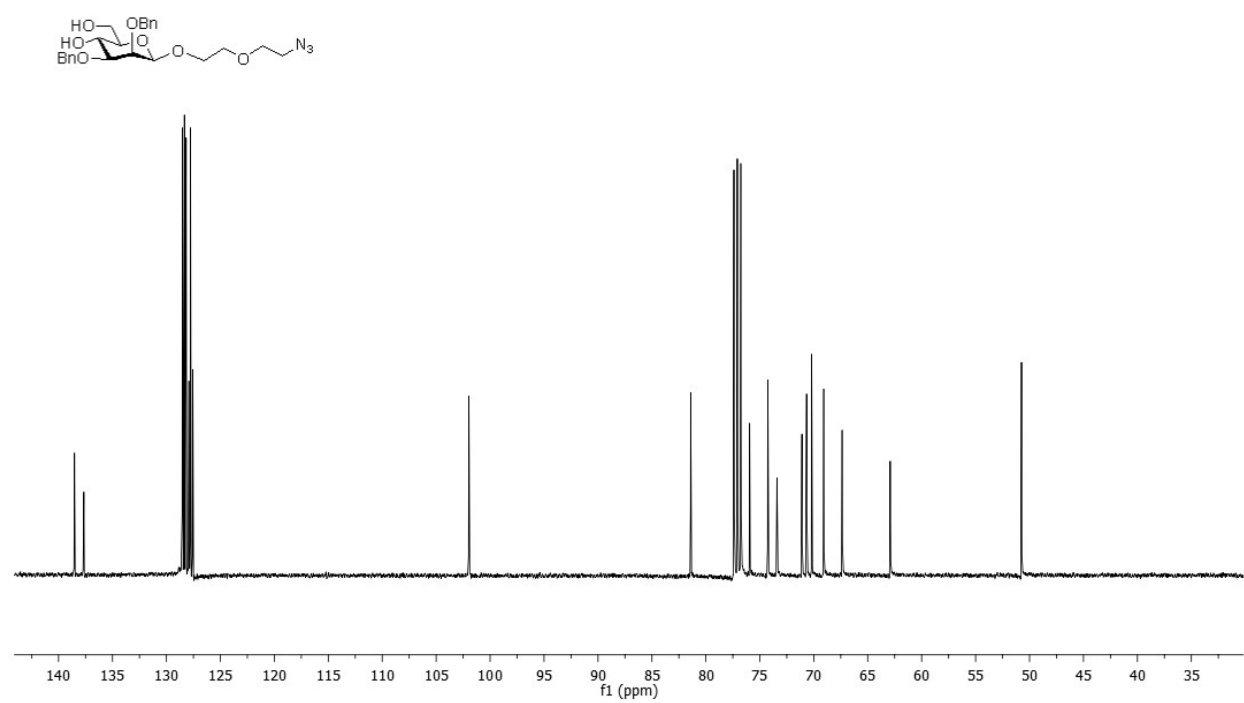
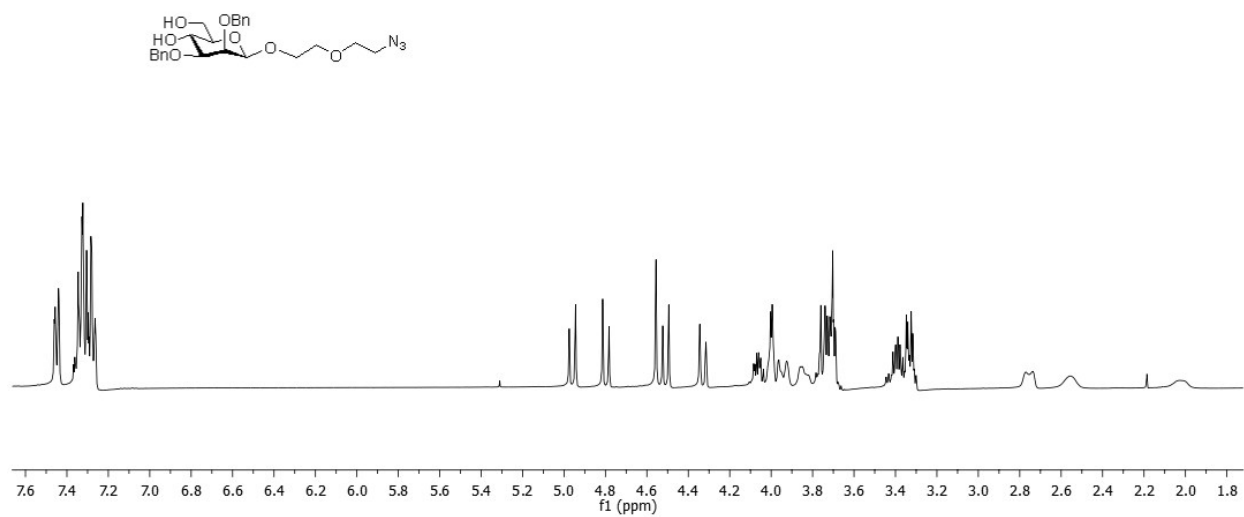
RD MN3 low voltage

IISER PUNE

RD MN3 low voltage 146 (2.685) AM2 (Ar,20000.0,556.28,0.00,LS 3); ABS; Sm (SG, 1x1.00); Cm (145:149-(128:143+152:170))

1: TOF MS ES+
2.84e6



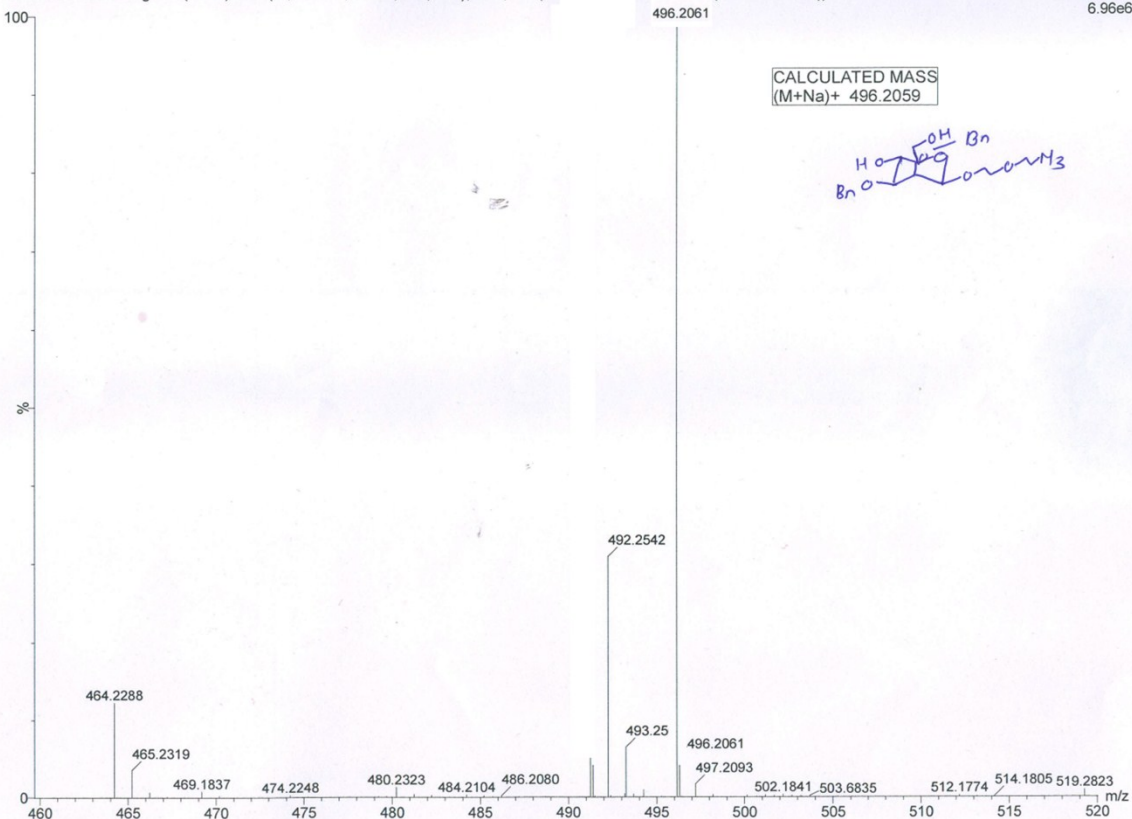


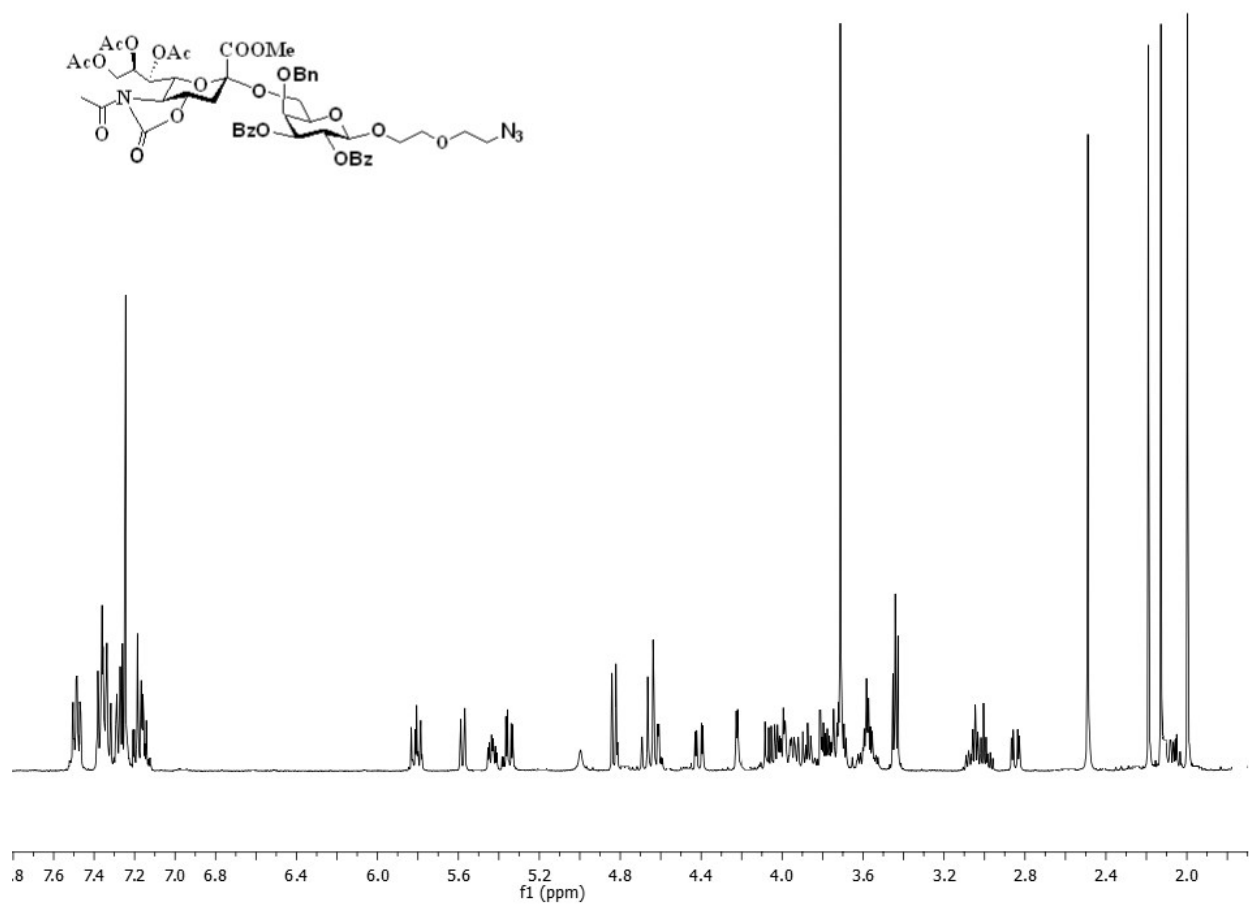
RD MN3OH low voltage

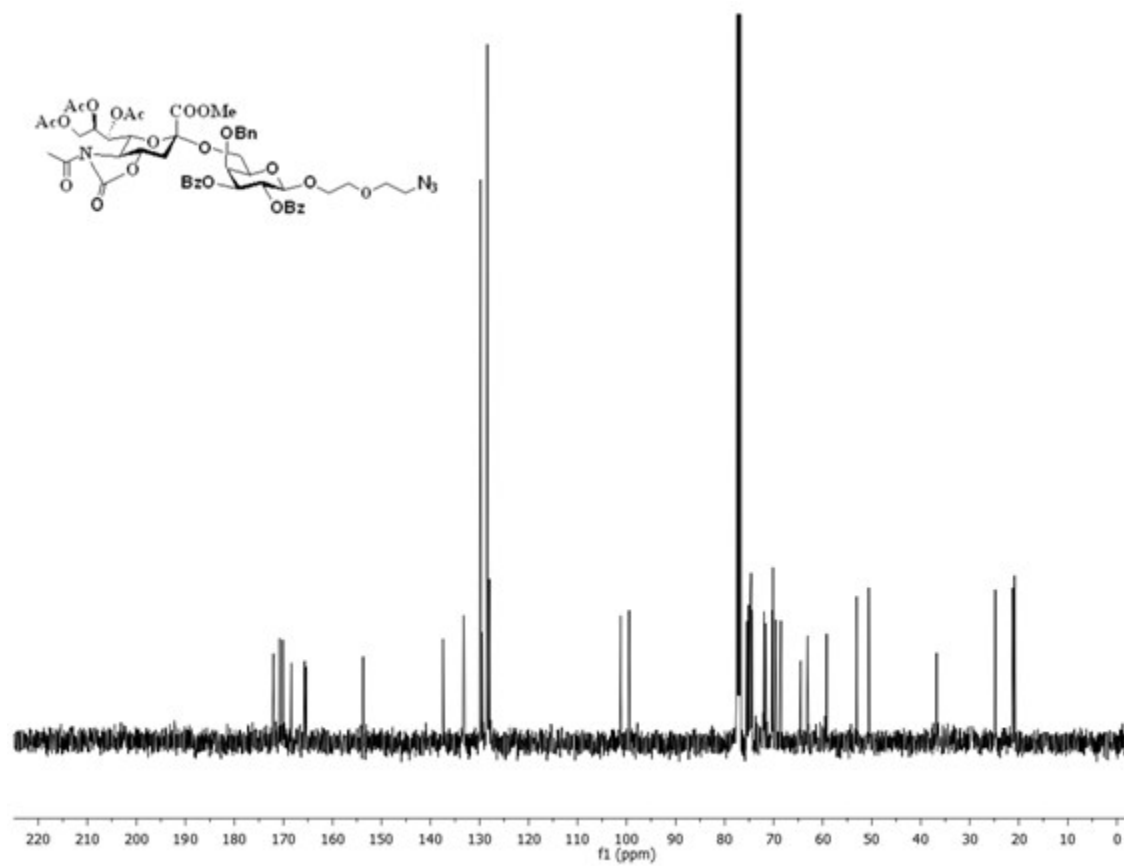
IISER PUNE

RD MN3OH low voltage 96 (1.772) AM2 (Ar,20000.0,556.28,0.00,LS 3); ABS; Sm (SG, 1x1.00); Cm /94:102-(78:92+108:127))

1: TOF MS ES+
6.96e6



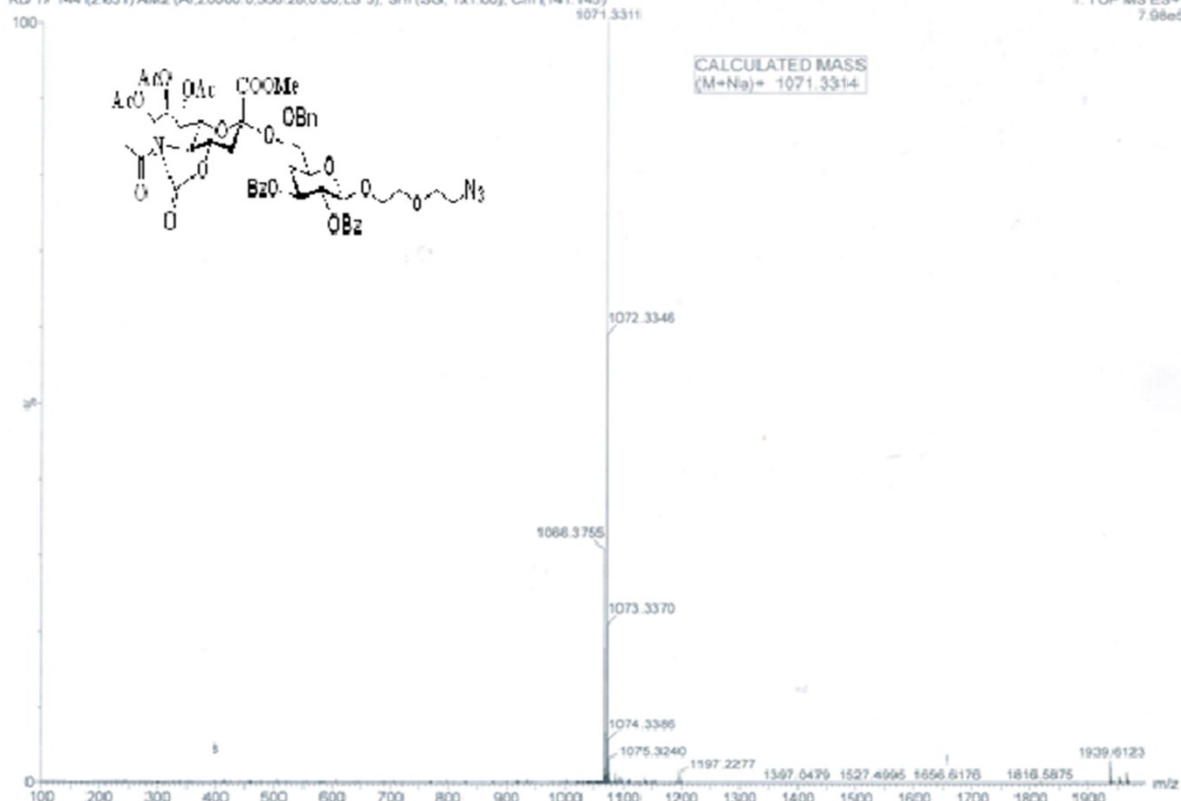


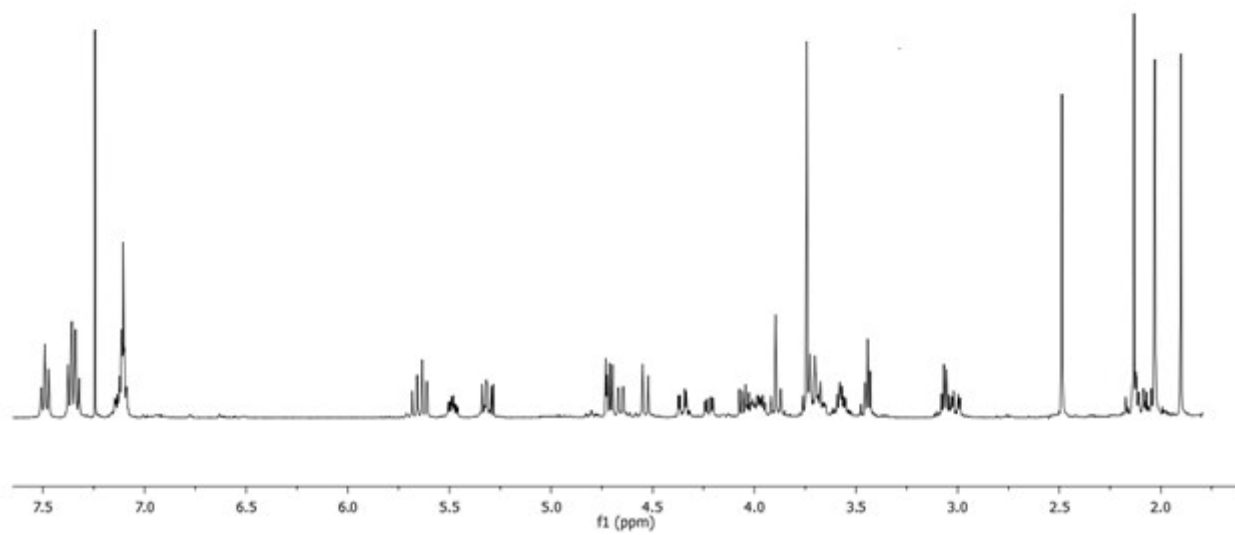
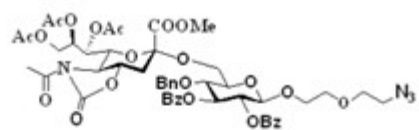


RD 17

RD 17 144 (2.631) AM2 (Ar:20000,0.556,28,0.00,LS 3): Sm (SG, 1x1.00); Cm (141:145)

IISER PUNE

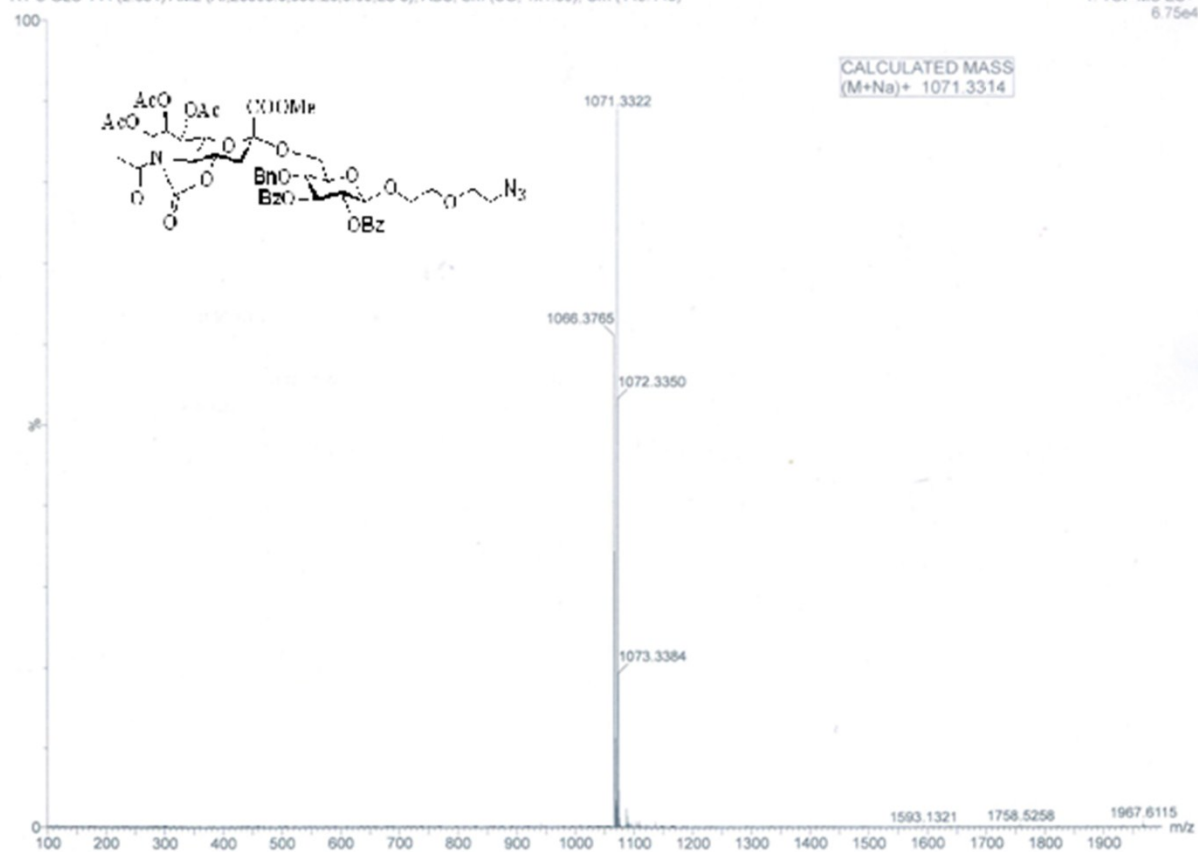
1: TOF MS ES+
7.98e5

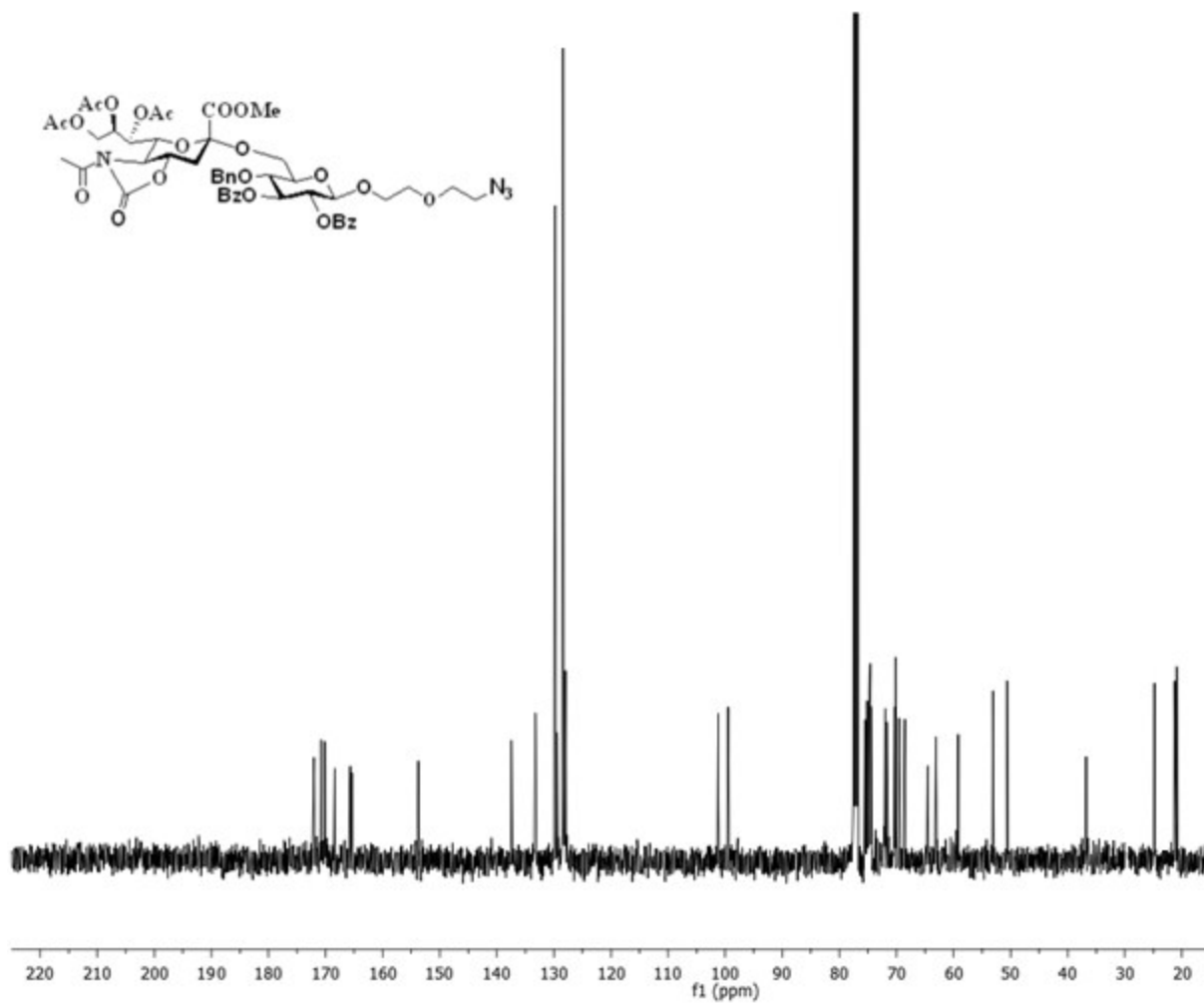


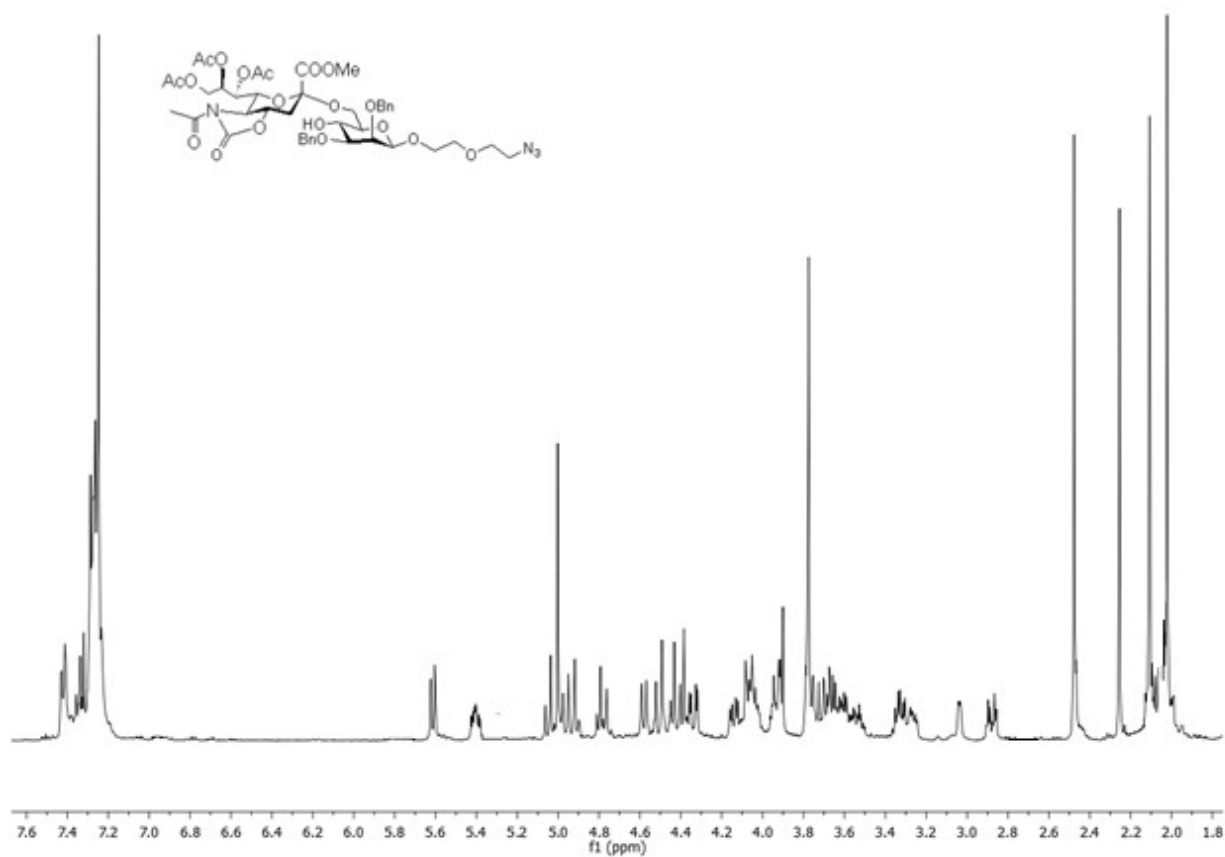
RY S GLU 144 (2.631) AM2 (Ar.20000 0.556 28.0 00,LS 3); ABS; Sm (SG, 1x1.00); Cm (143-145)

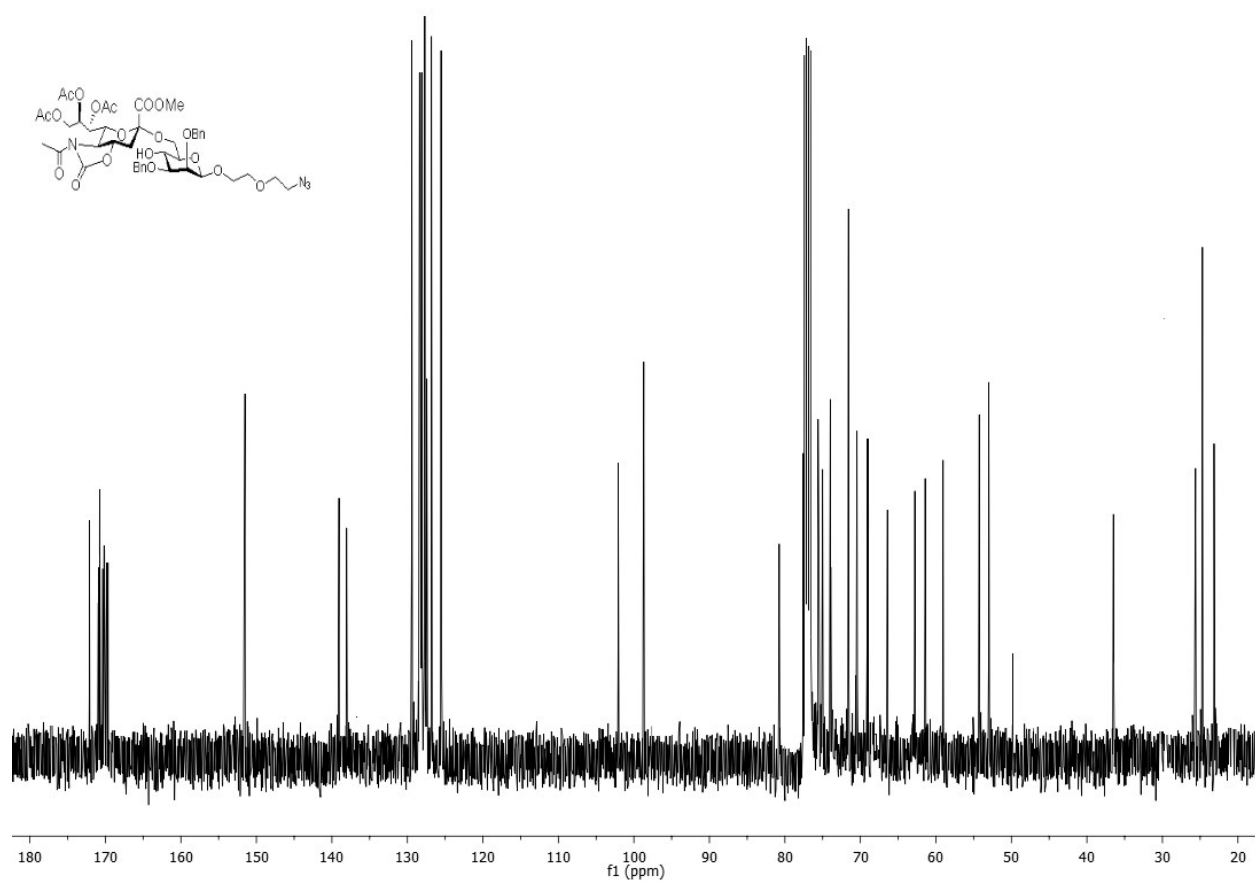
1.00); Cm (143:145)

1: TOF MS ES+
6.75e4









RD_SiM high voltage 80

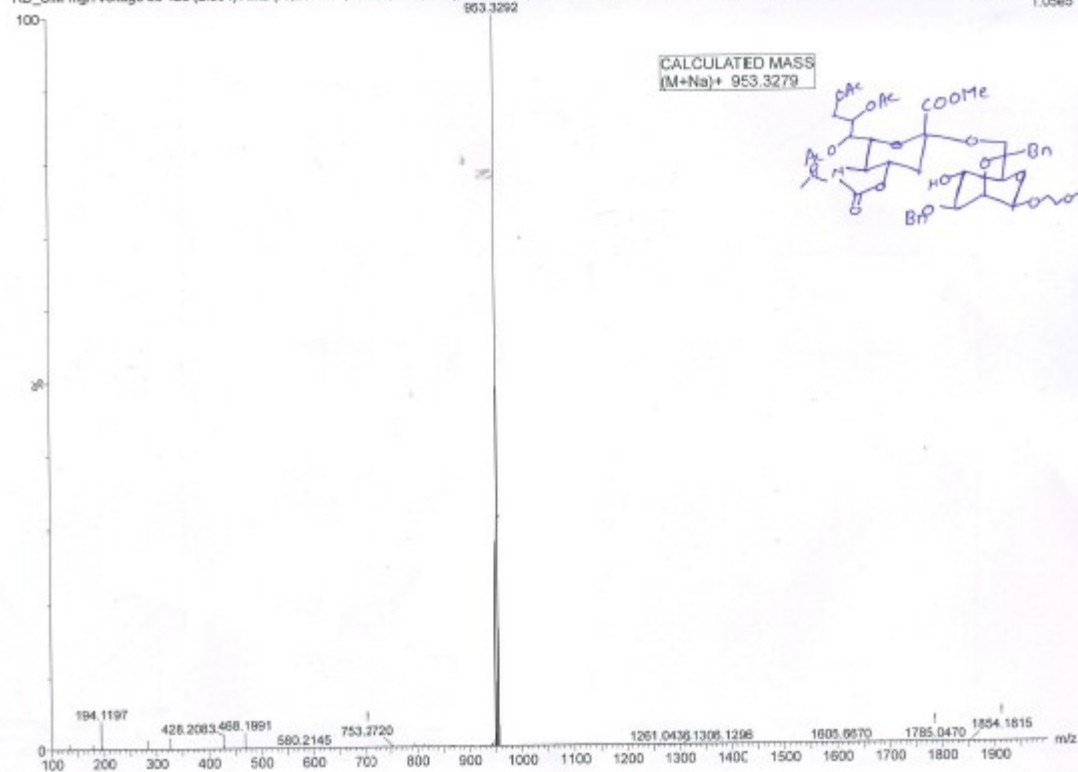
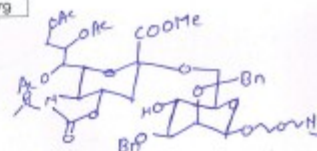
IIISER PUNE

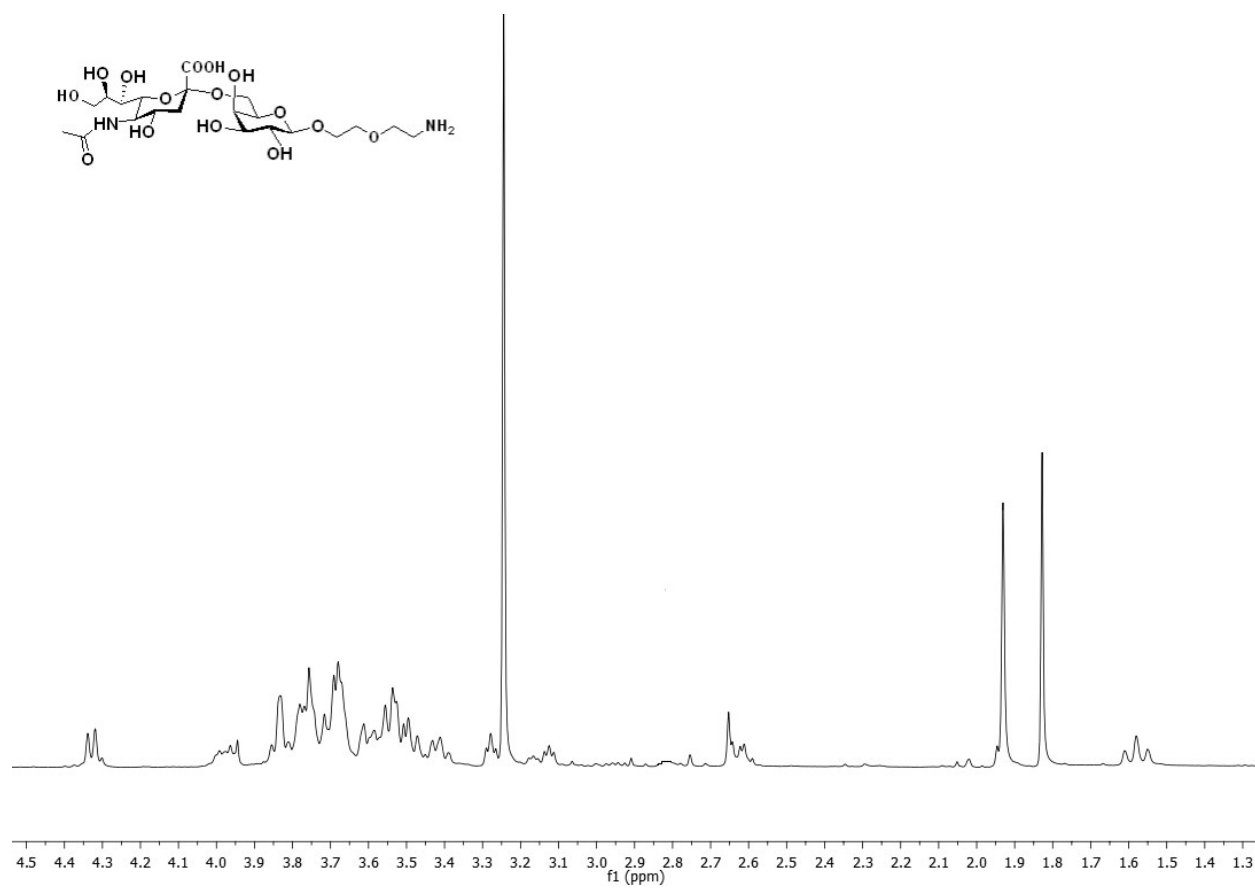
1: TOF MS ES+
1.05e5

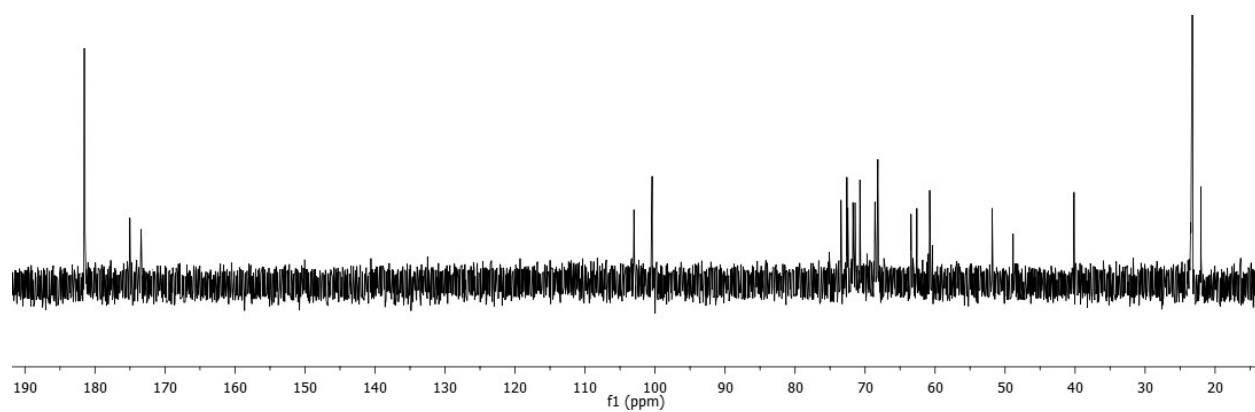
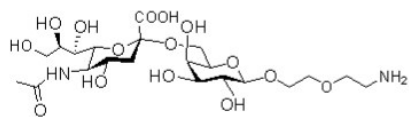
RD_SiM high voltage 80 125 (2.304) AM2 (Ar, 20000.0, 556.28, 0.00, 1.5 3); ABS; Sm (SG, 1x1.00); Cm (123:128-(112:12+131:144))

953.3292

CALCULATED MASS
(M+Na)+ 953.3279





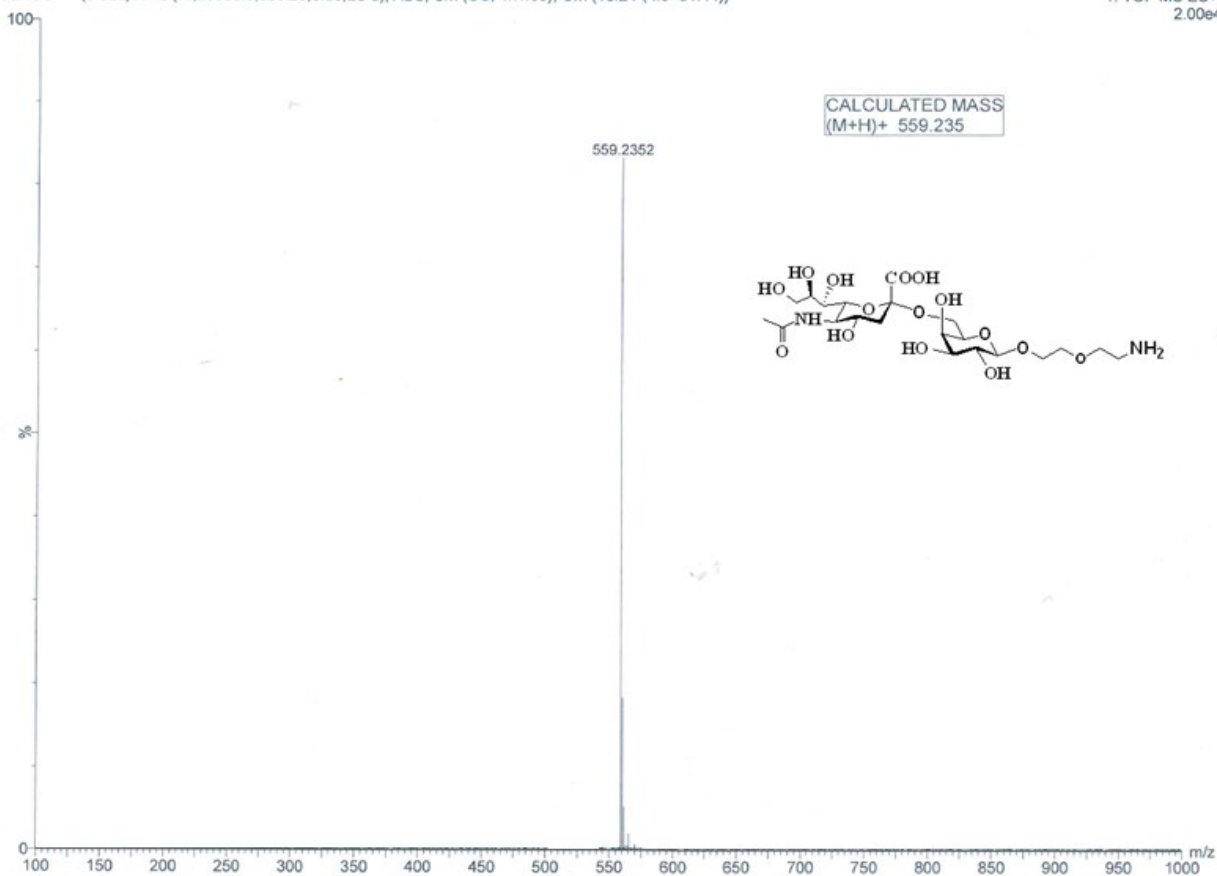


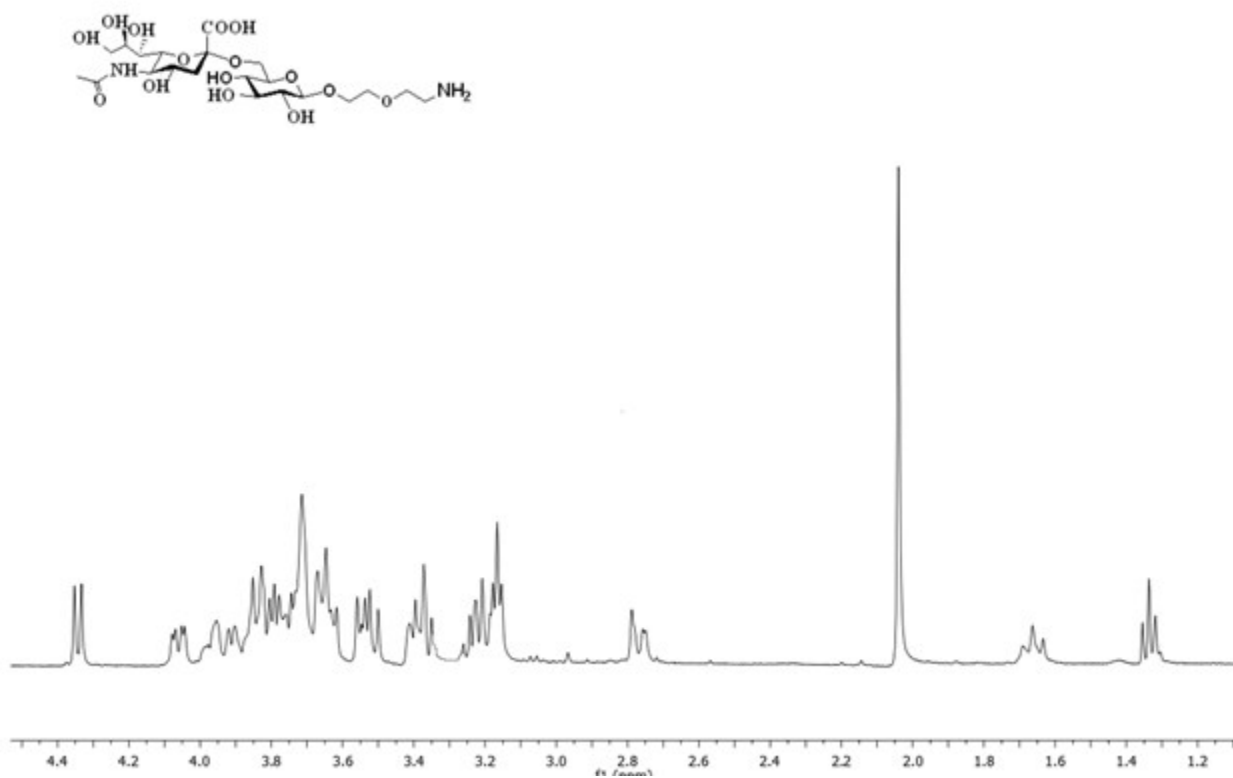
RD SGalcutuie

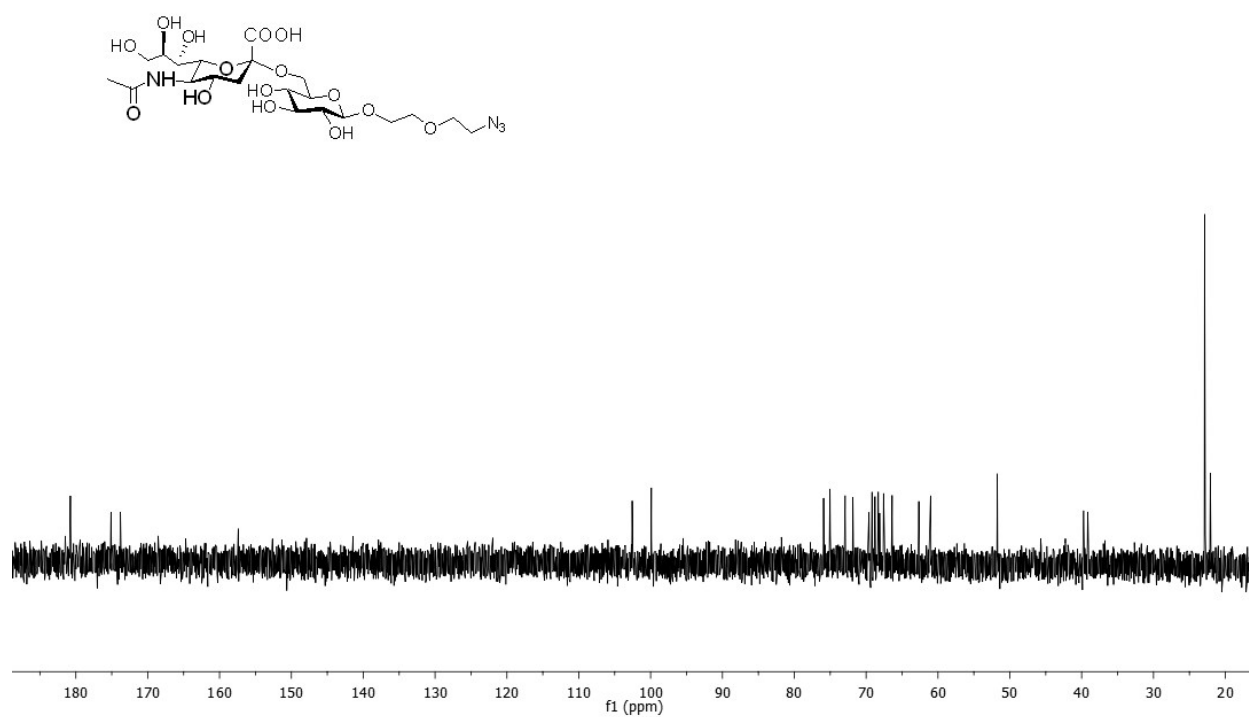
IISER PUNE

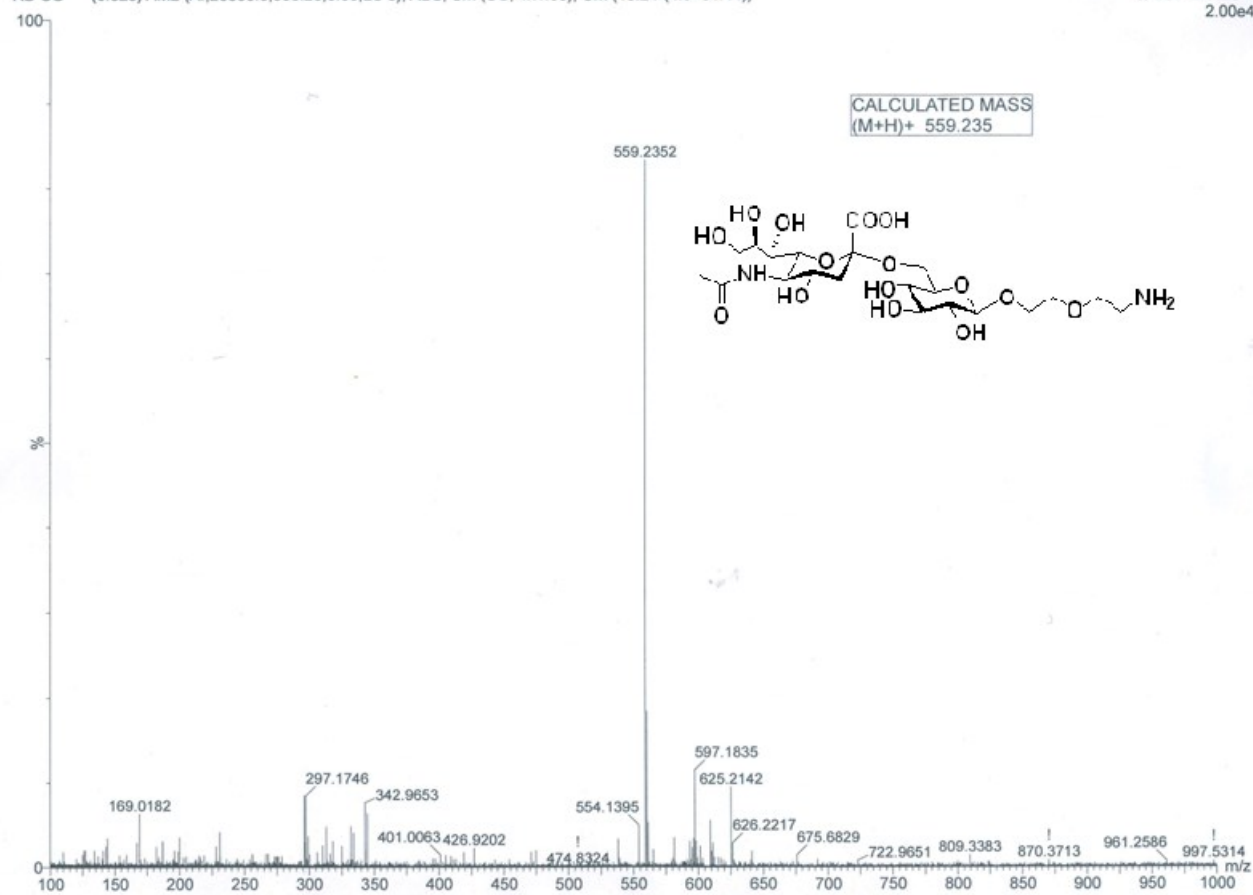
RD SG (0.325) AM2 (Ar,20000.0,556.28,0.00,LS 3); ABS; Sm (SG, 1x1.00); Cm (13.24-(4.9+31.44))

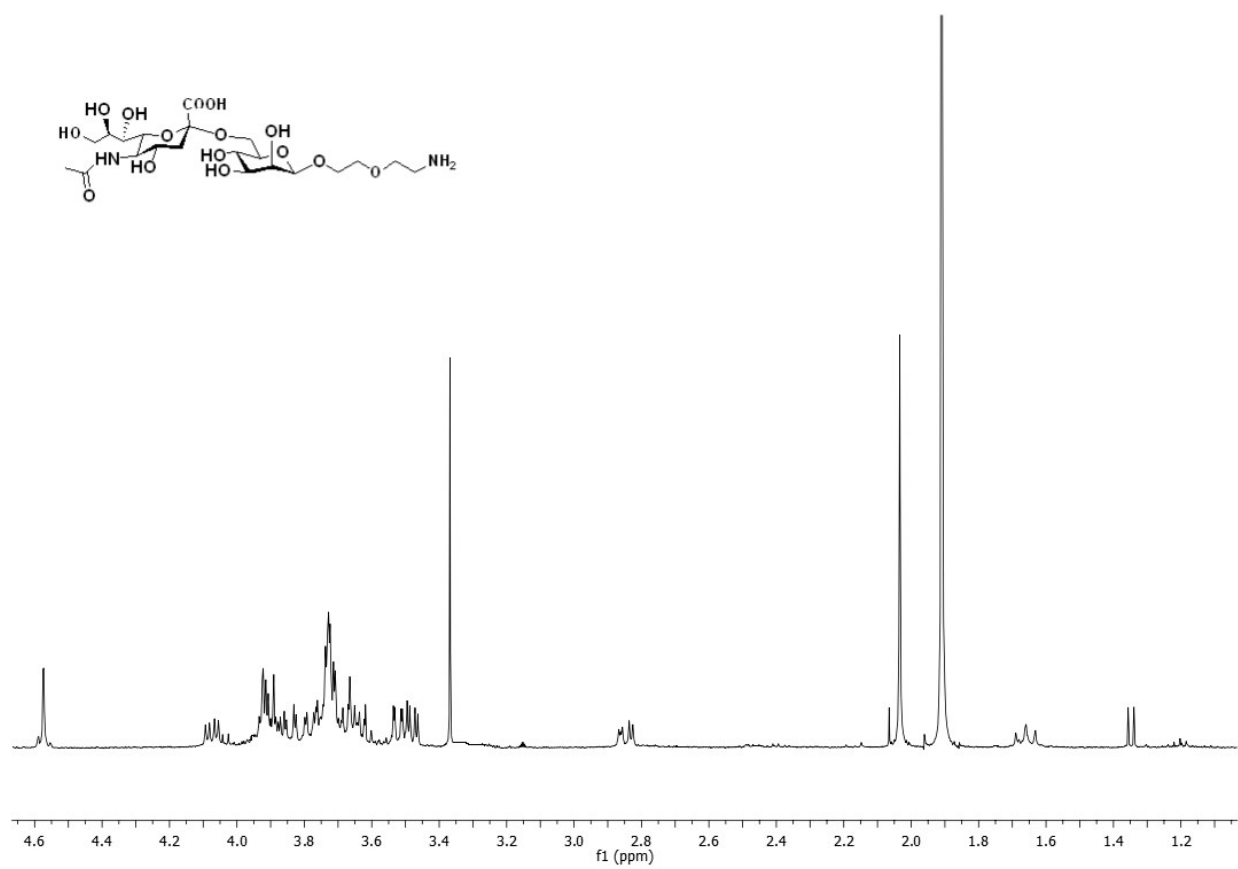
1: TOF MS ES+
2.00e4

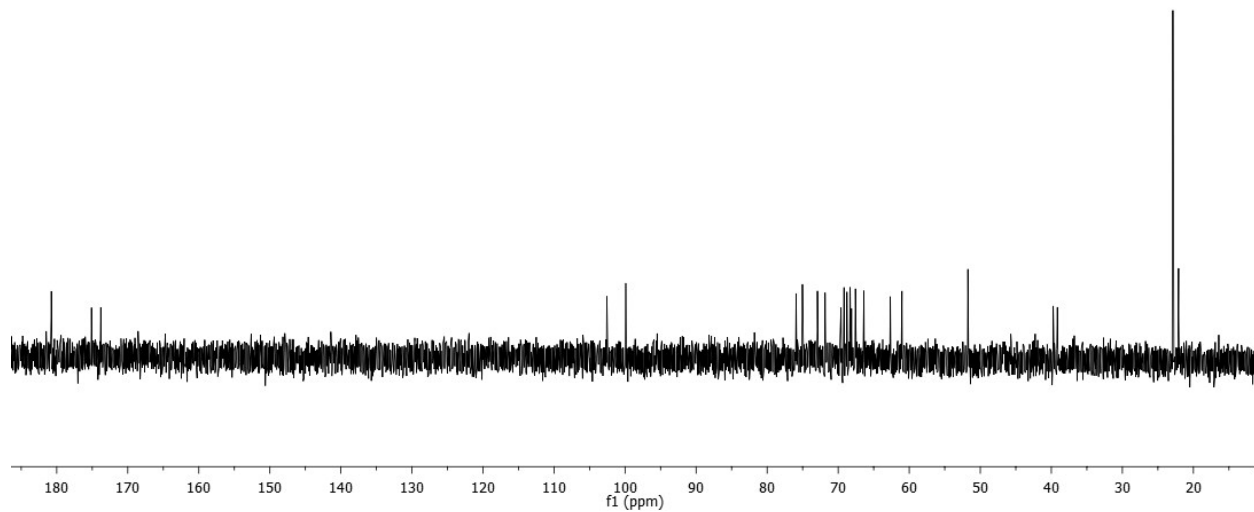
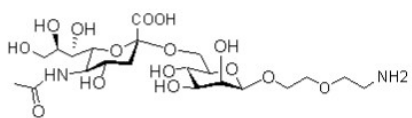






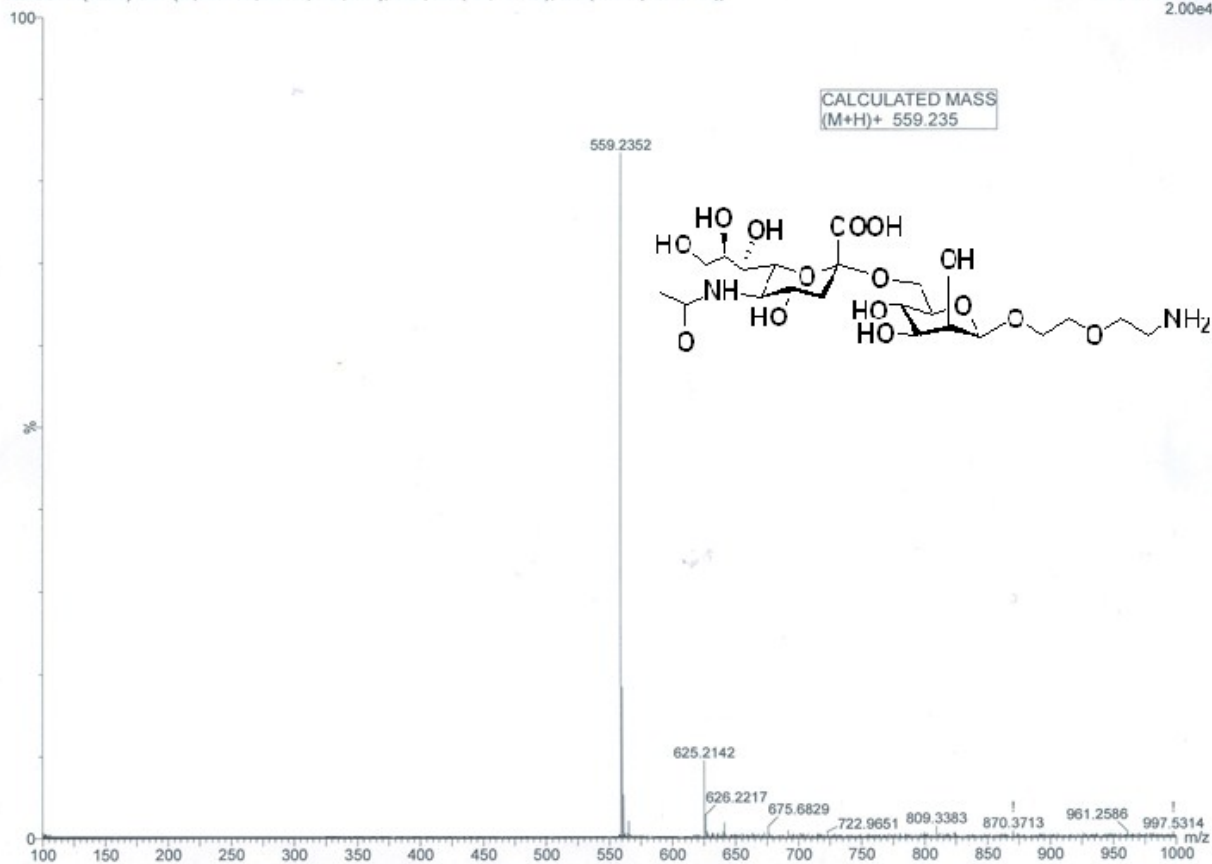




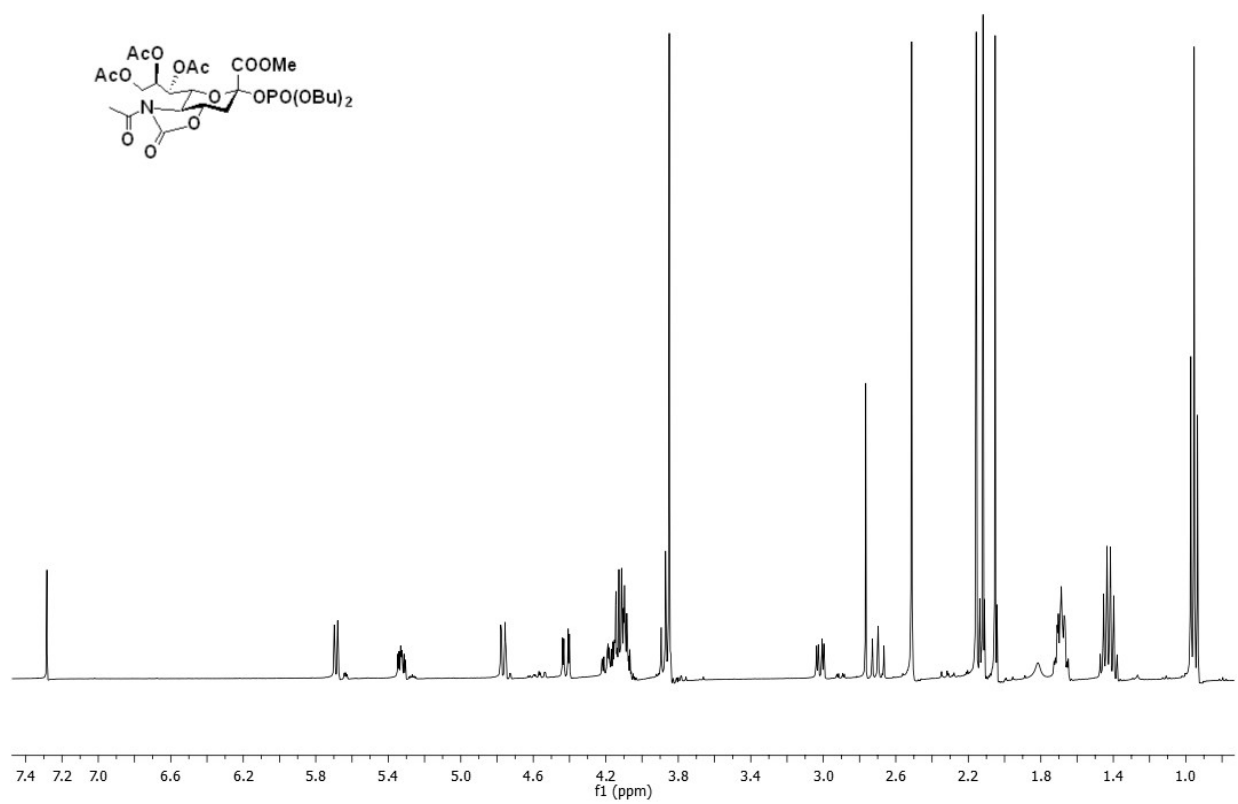


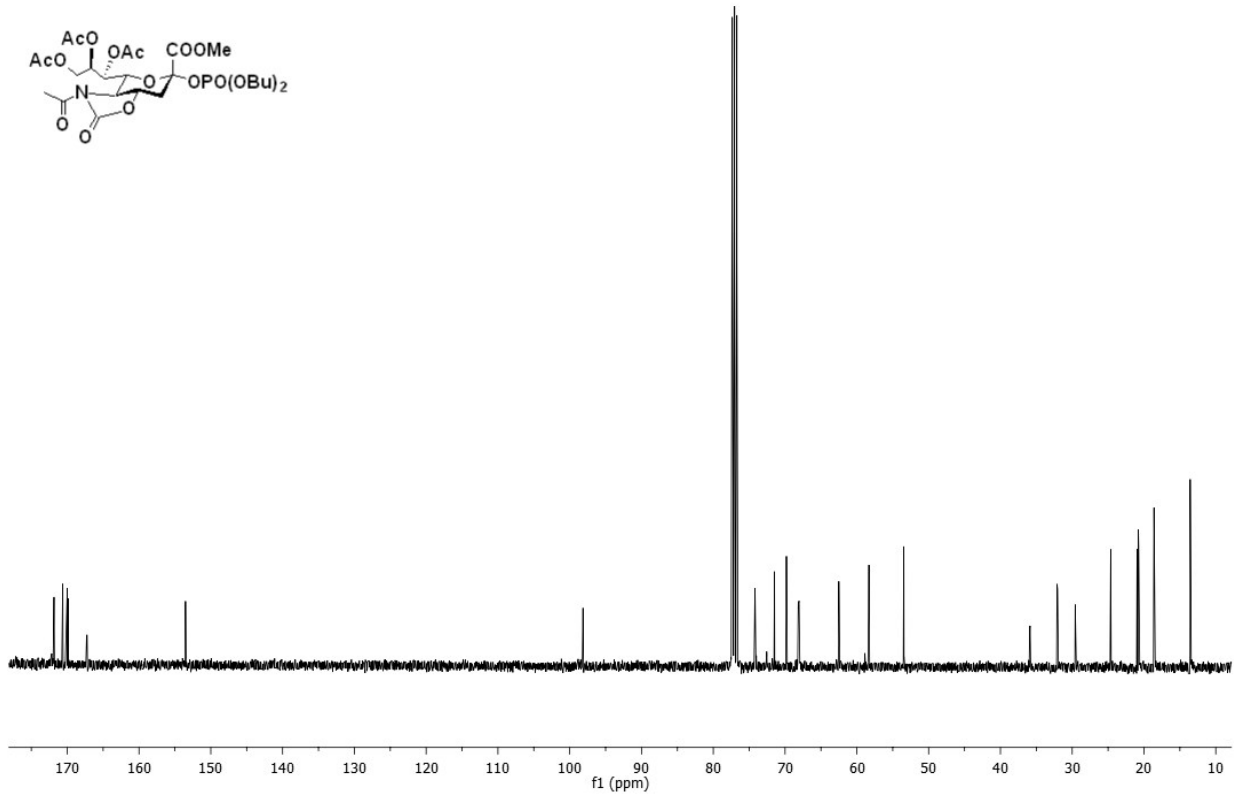
IISER PUNE
RD SG 17 (0.325) AM2 (Ar,20000.0,556.28,0.00,LS 3); ABS; Sm (SG, 1x1.00); Cm (13:24-(4.9+31:44))

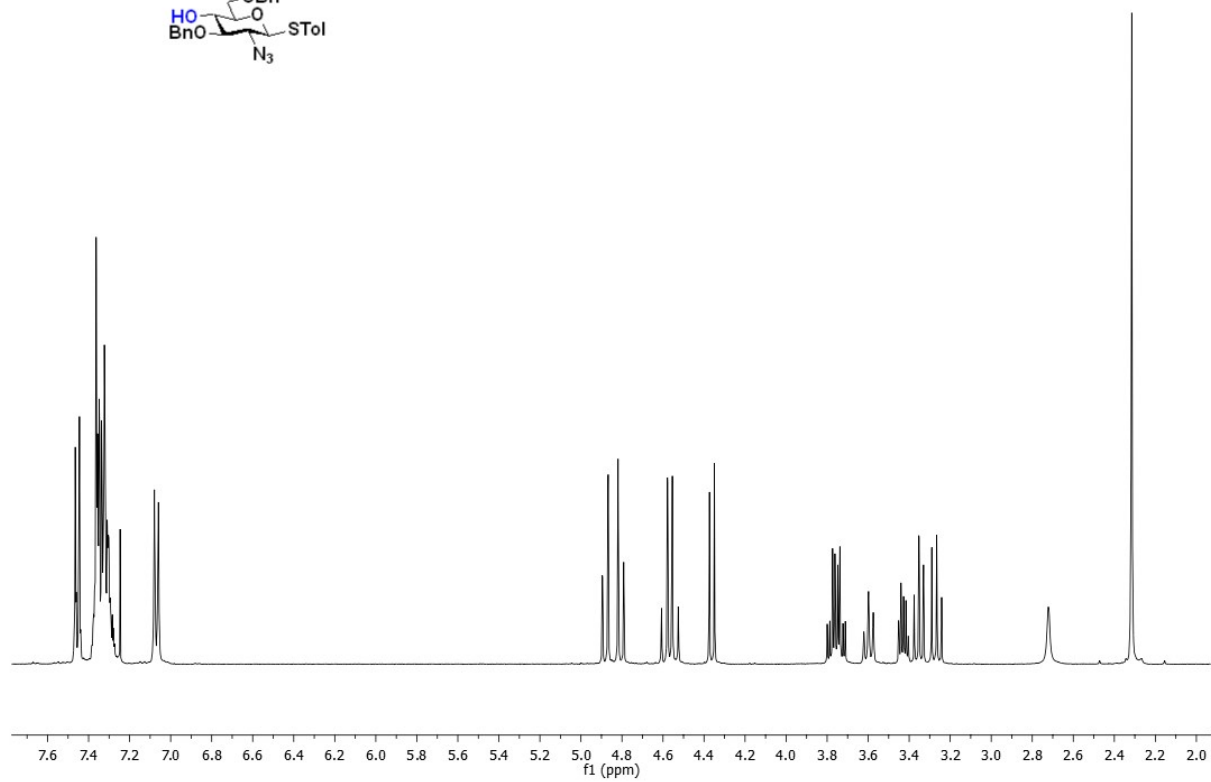
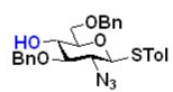
1: TOF MS ES+
2.00e4

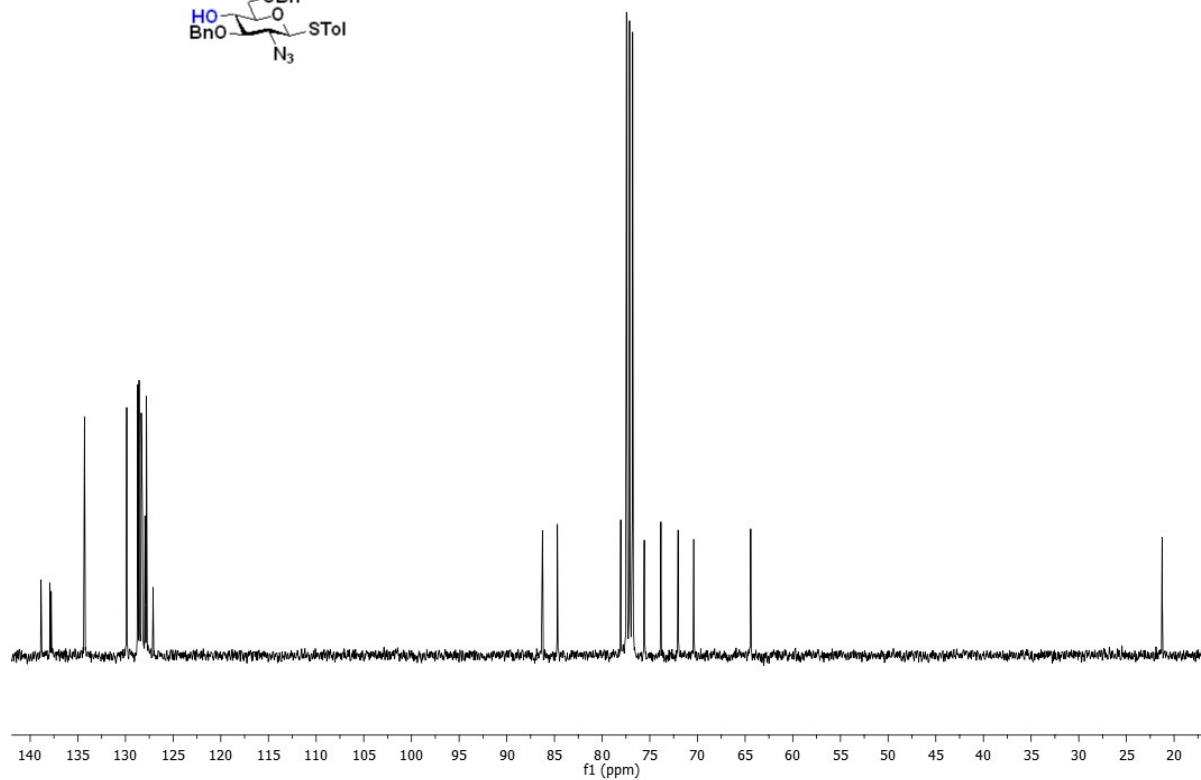
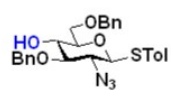


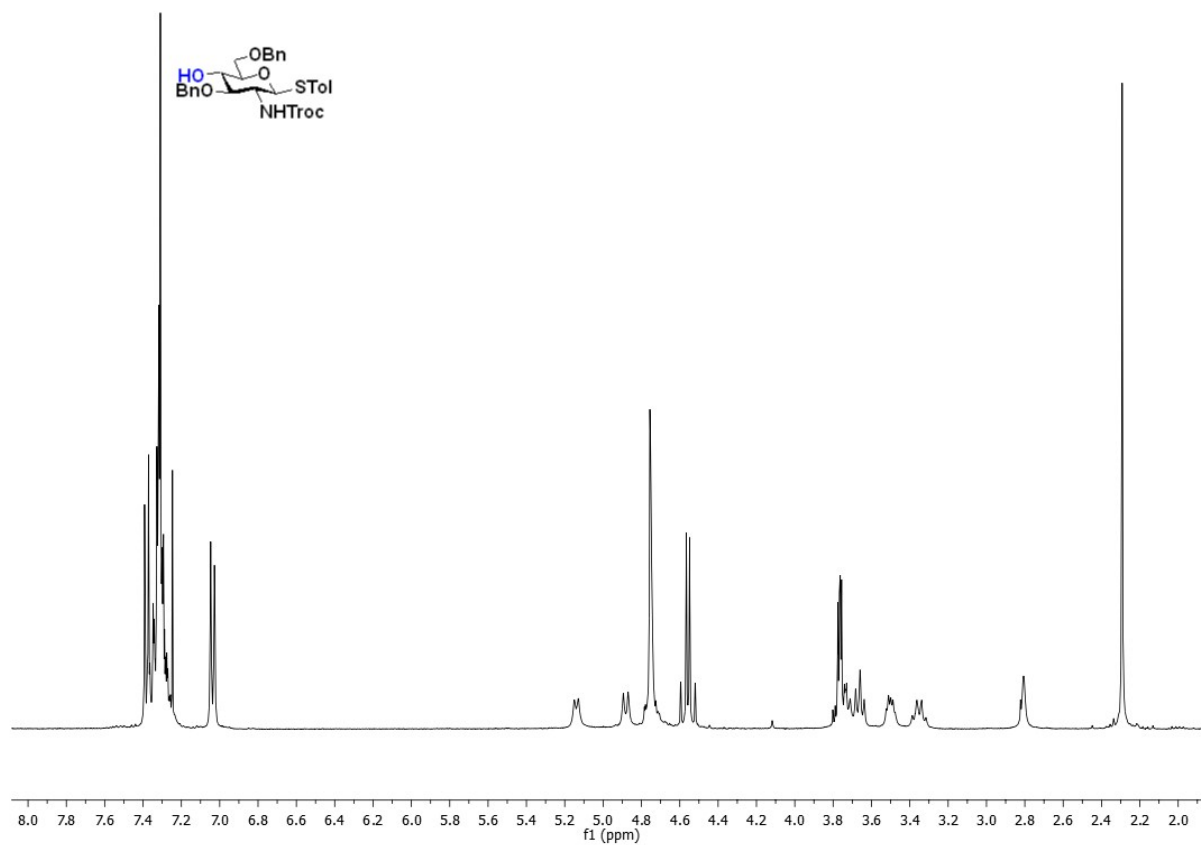
NMR For Trisaccharide Library

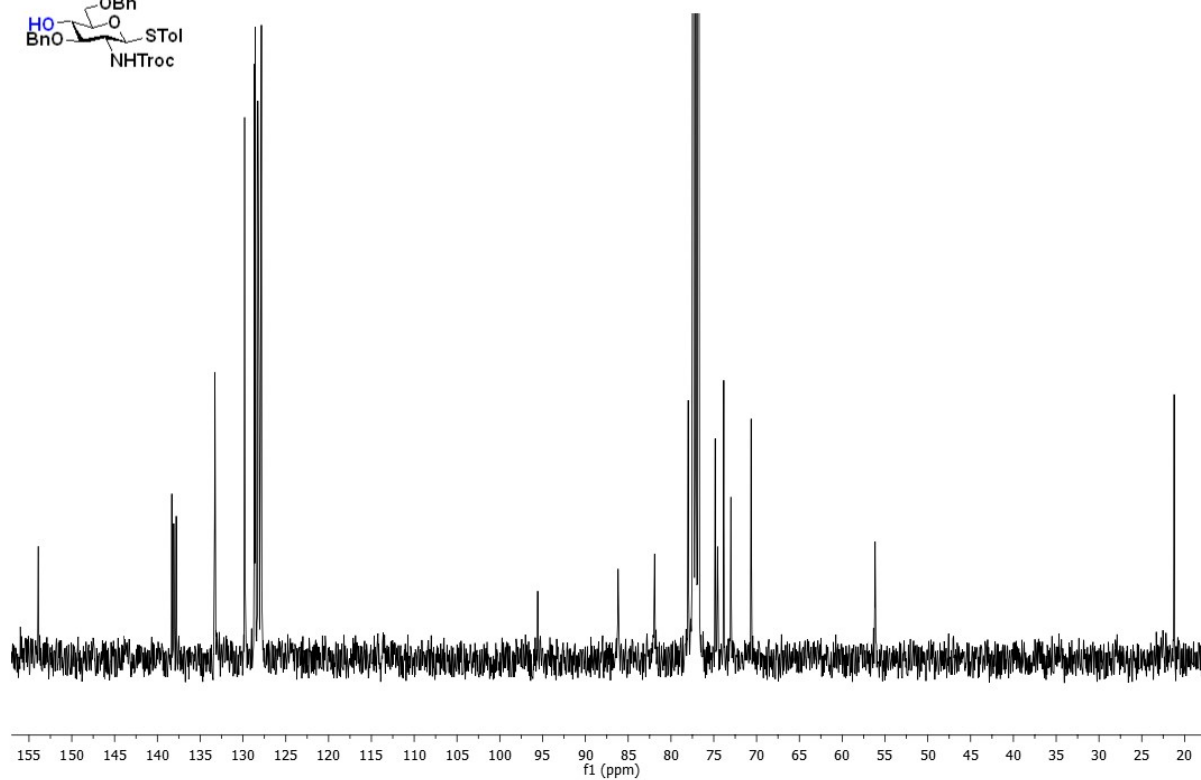
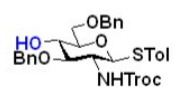










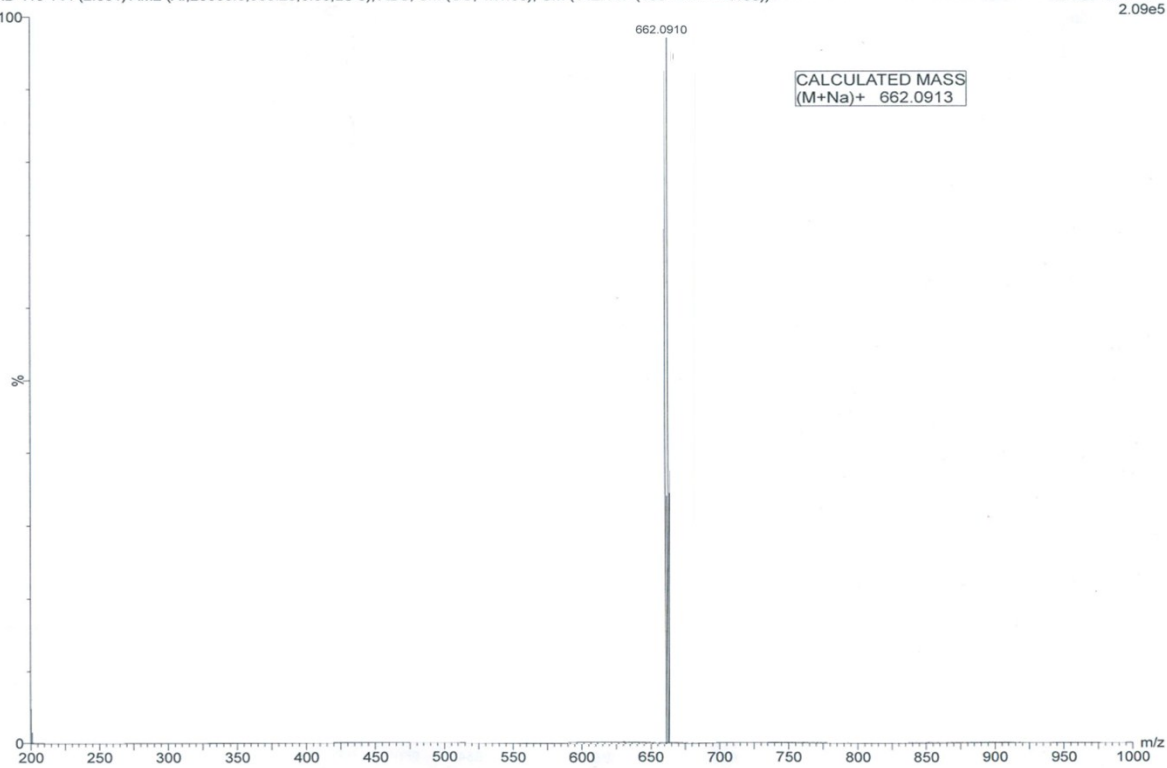


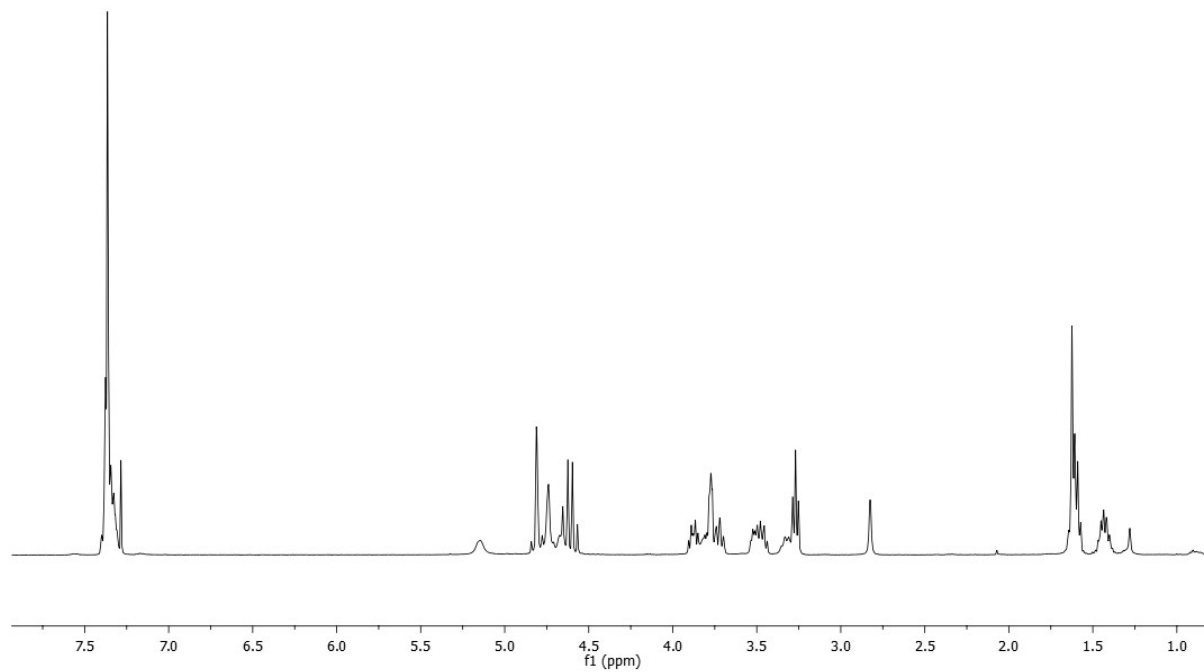
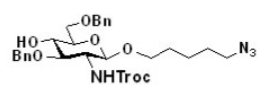
RD 110

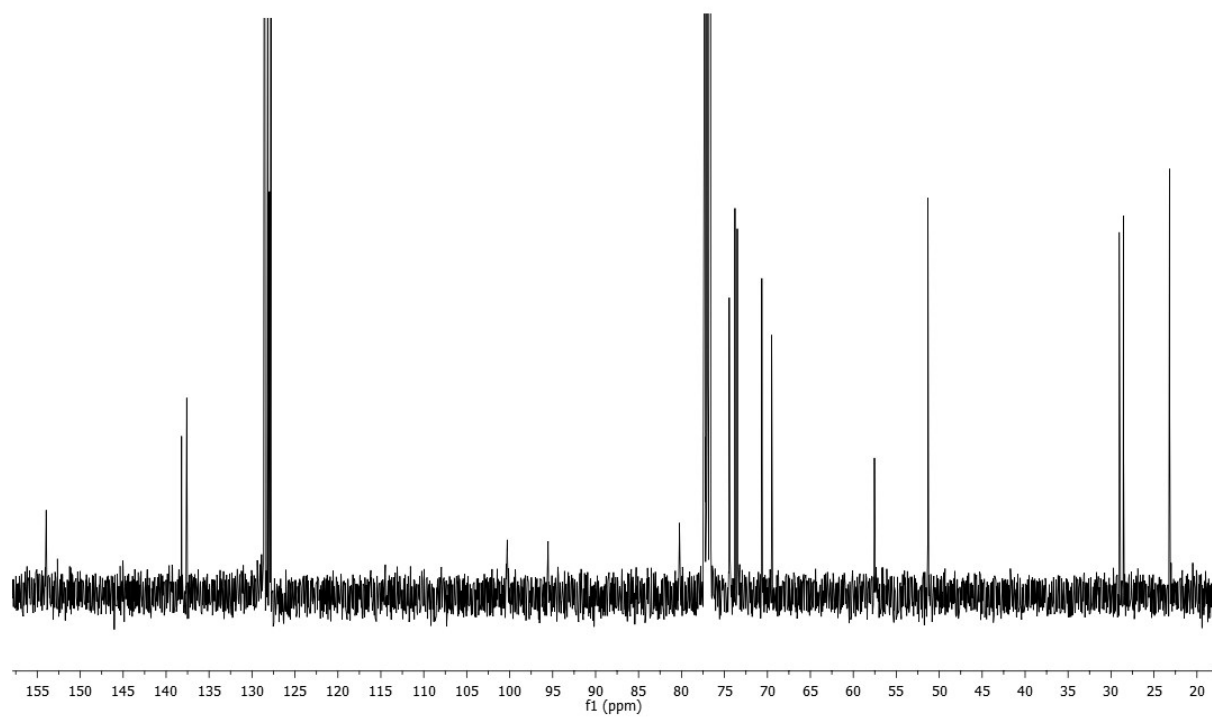
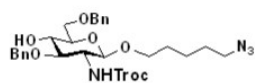
IISER PUNE

RD 110 144 (2.631) AM2 (Ar,20000,0,556.28,0,00,LS 3); ABS; Sm (SG, 1x1.00); Cm (142:147-(135:140+14R:163))

1: TOF MS ES+
2.09e5





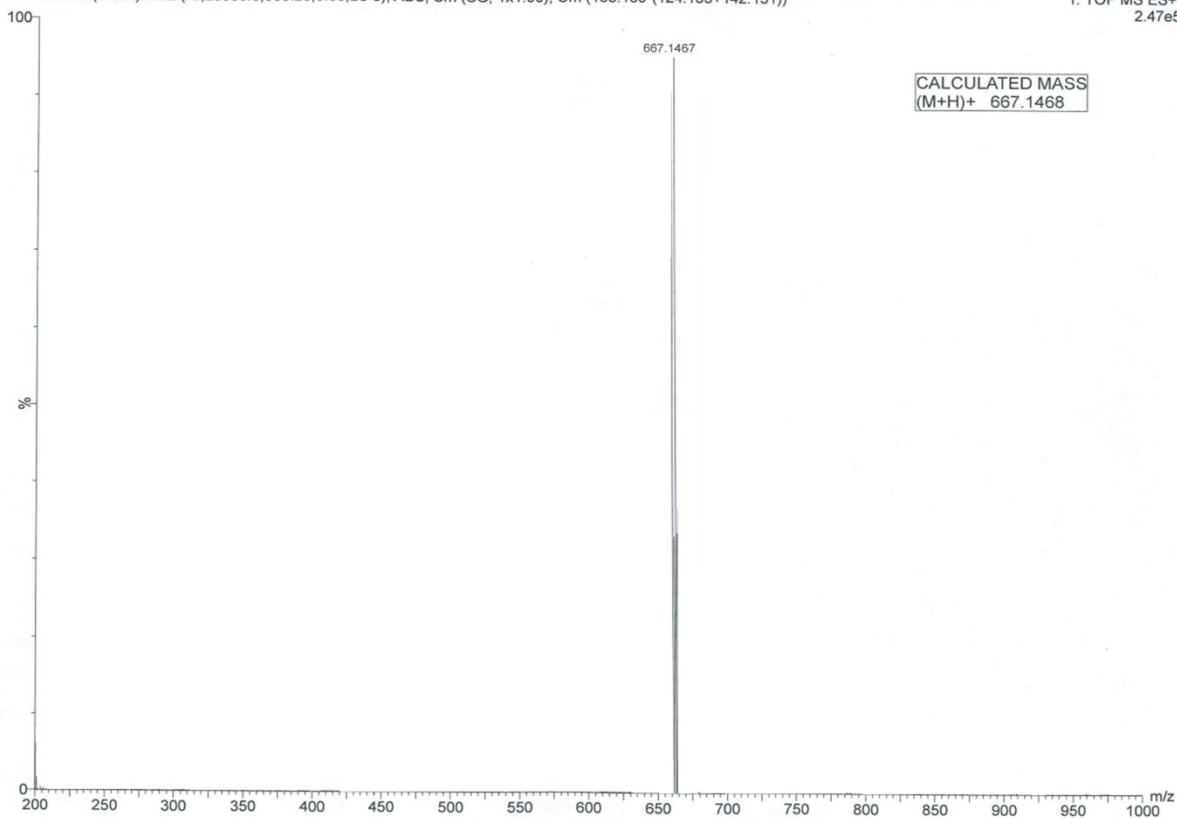


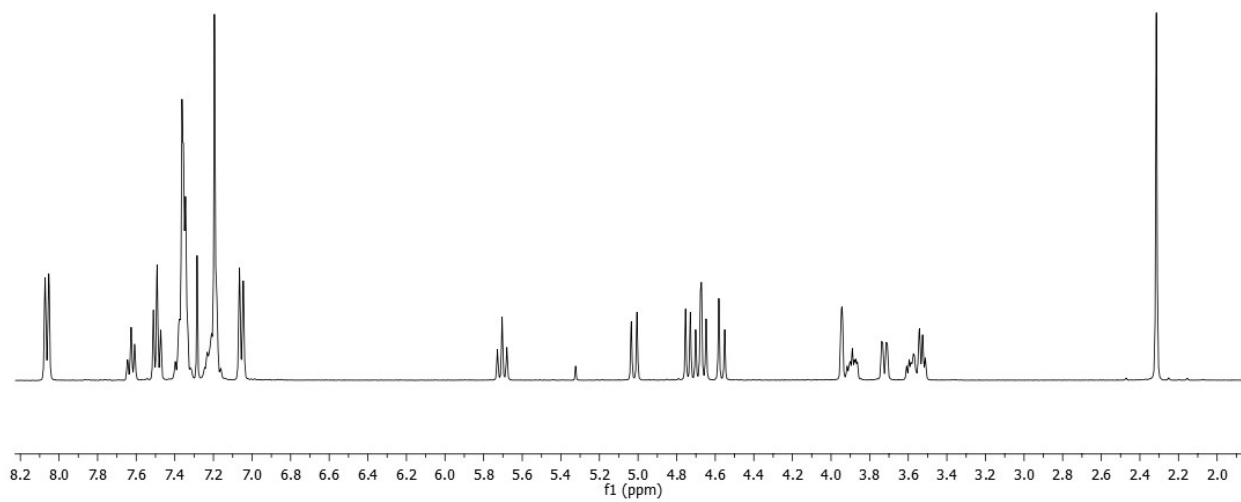
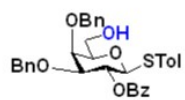
RD 22B

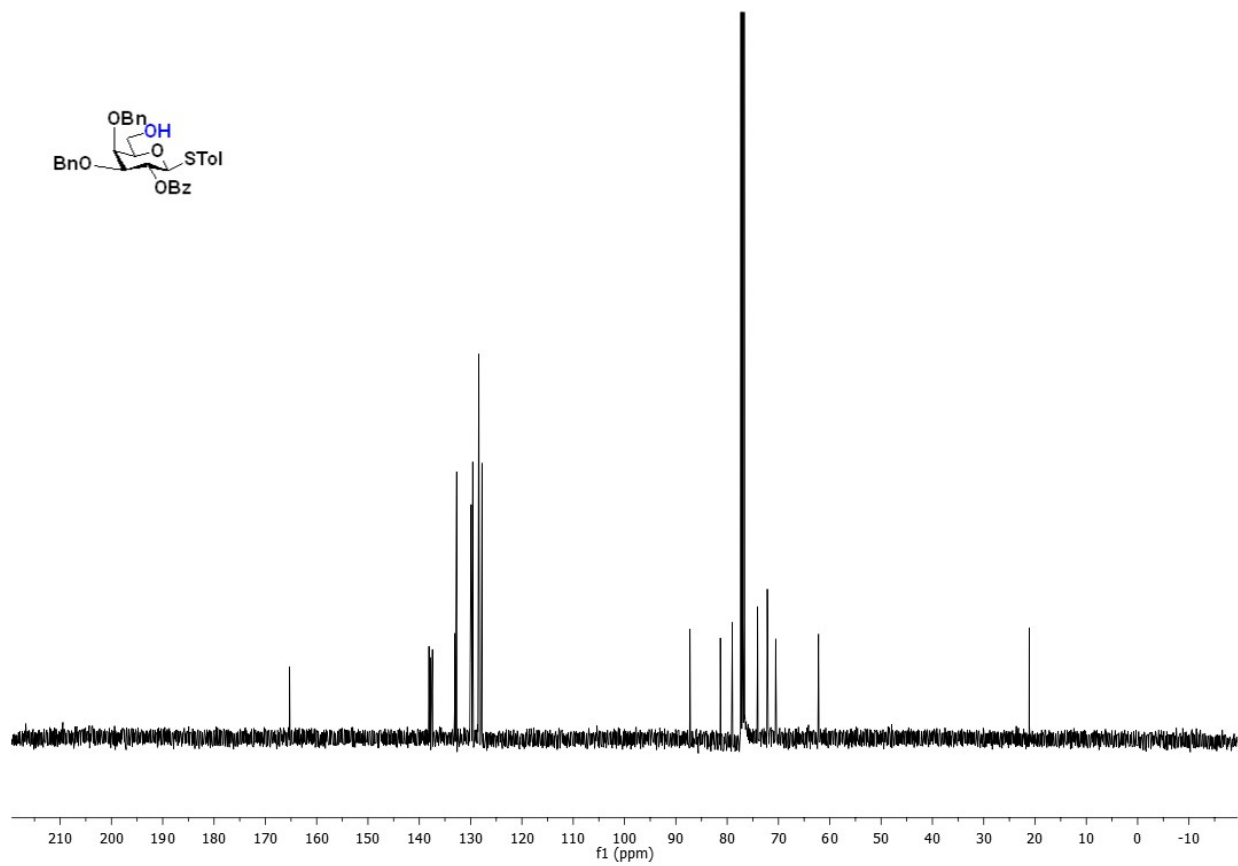
IISER PUNE

RD 22B 136 (2.494) AM2 (Ar,20000.0,556.28,0.00,LS 3); ABS; Sm (SG, 1x1.00); Cm (135:139-(124:133+142:151))

1: TOF MS ES+
2.47e5



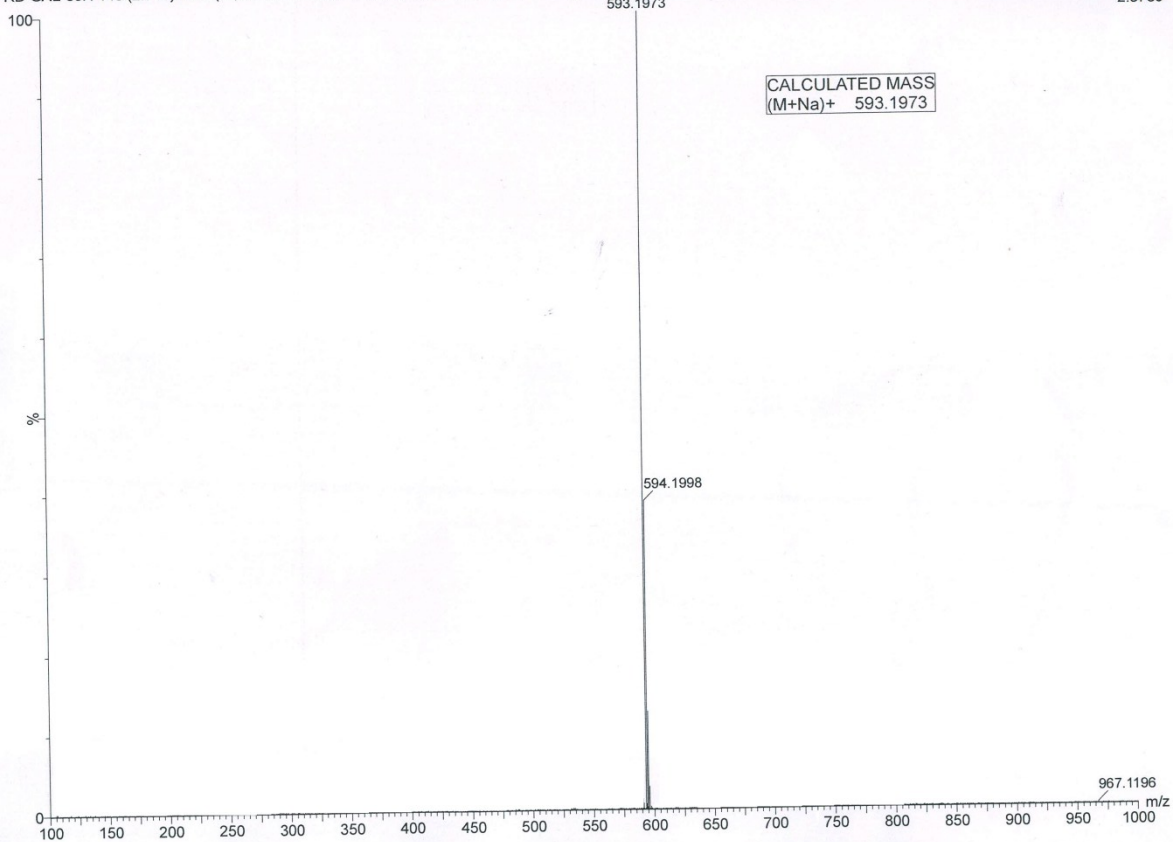


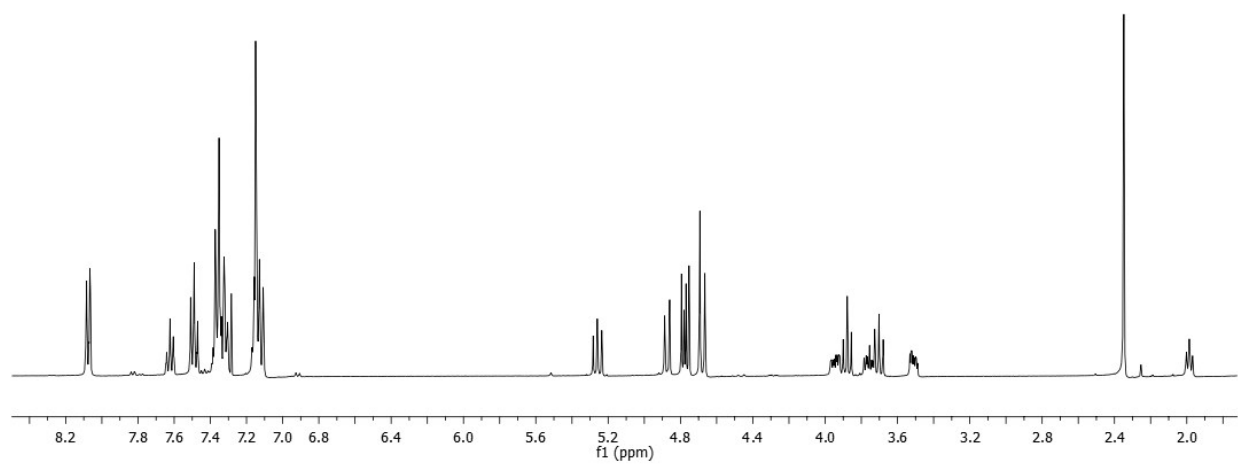
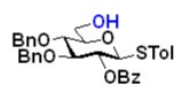


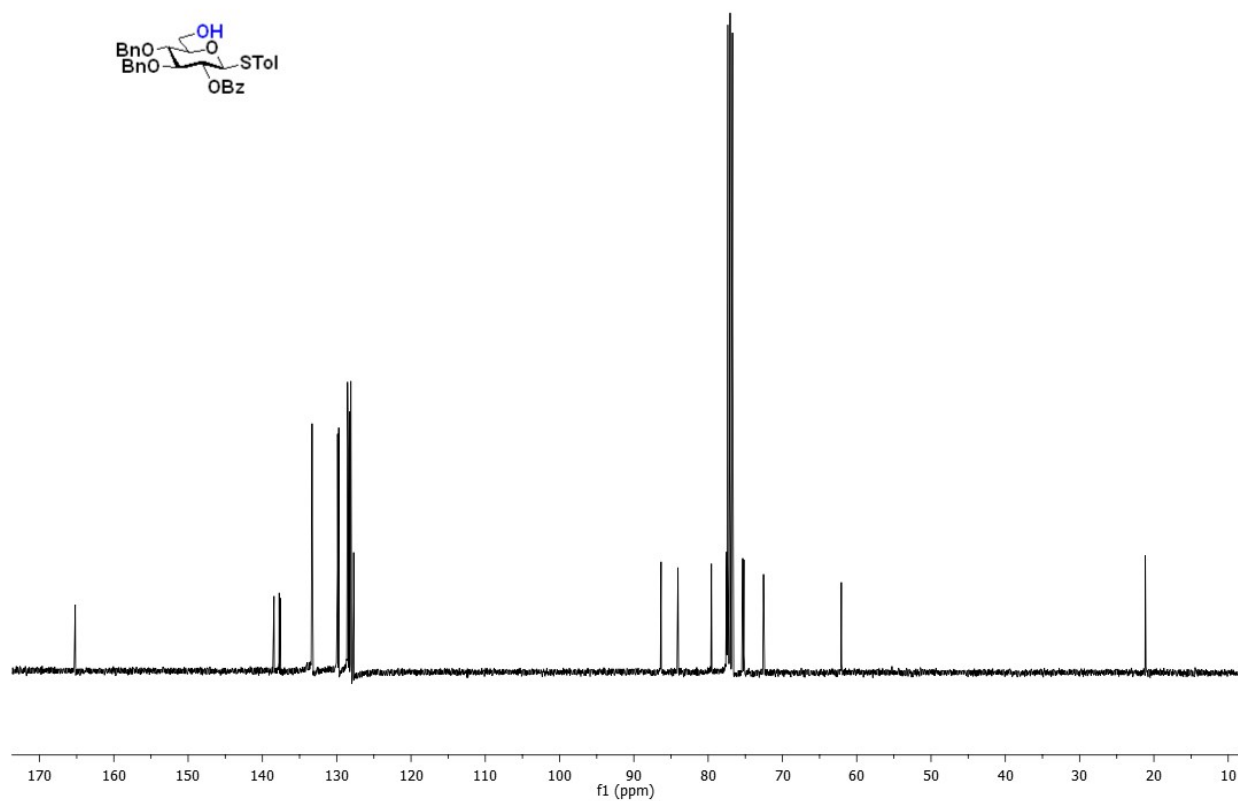
RD GAL-60H
RD GAL-60H 148 (2.719) AM2 (Ar, 20000.0, 556.28, 0.00, LS 3); ABS; Sm (SG, 1x1.00); Cm (145:161-(132:141+170:186))

IISER PUNE

1: TOF MS ES+
2.57e5





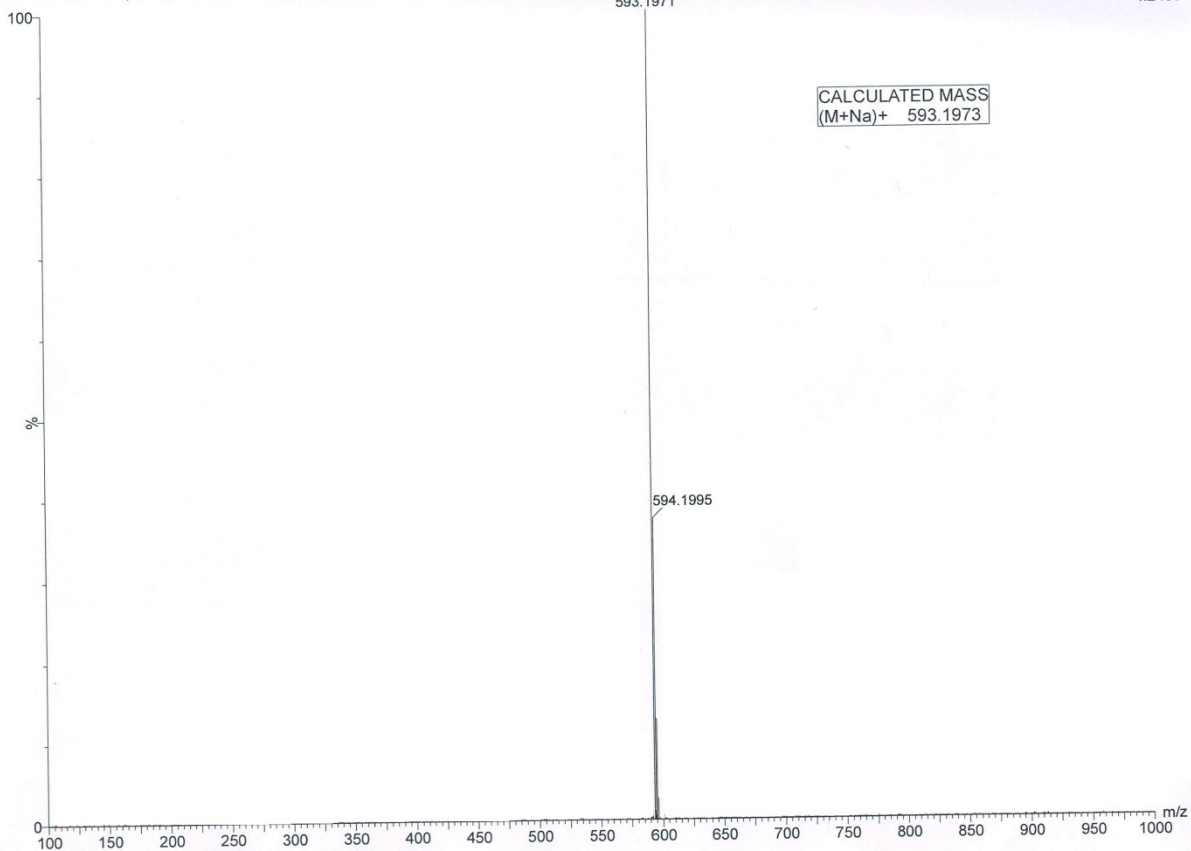


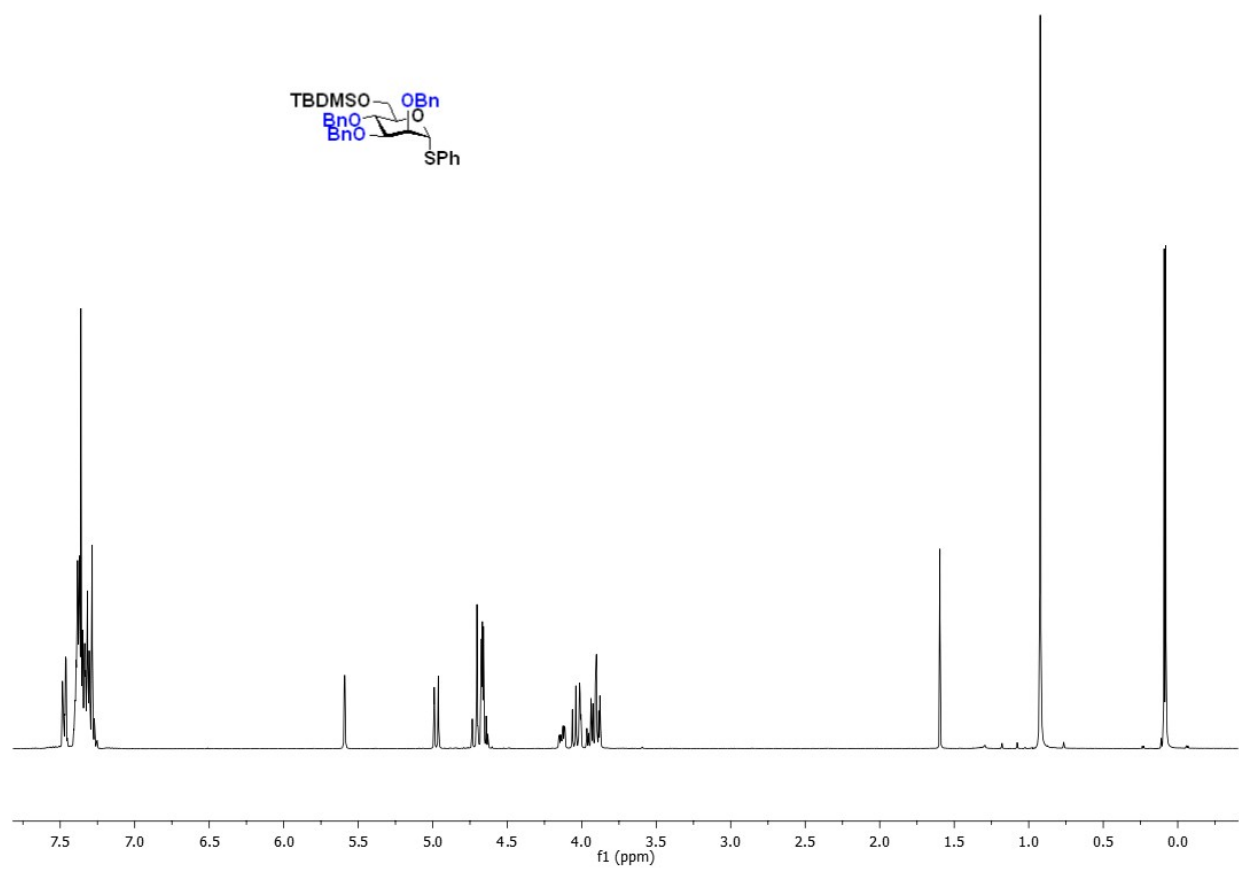
RD GLU60H

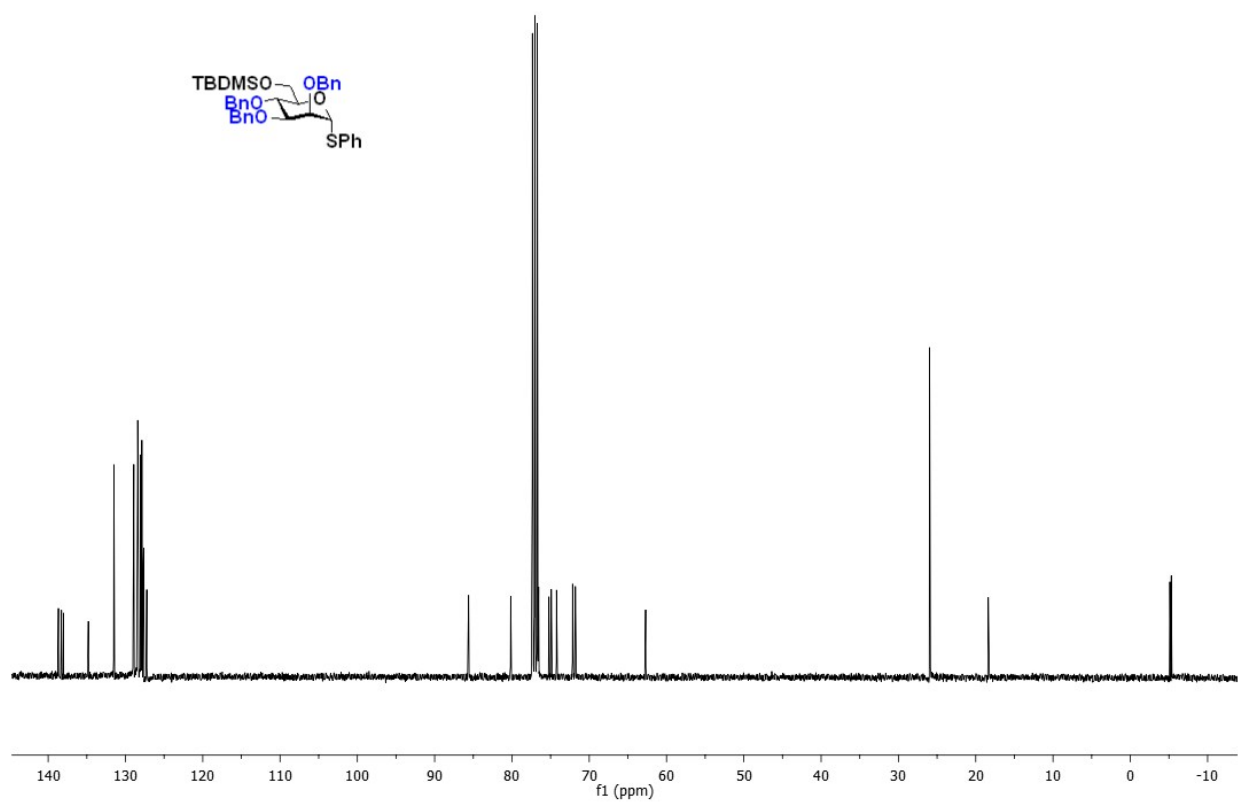
IISER PUNE

1: TOF MS ES+
4.24e5

RD GLU60H 148 (2.719) AM2 (Ar,20000.0,556.28,0.00,LS 3); ABS; Sm (SG, 1x1.00); Cm (145:158-(129:141+164:183))





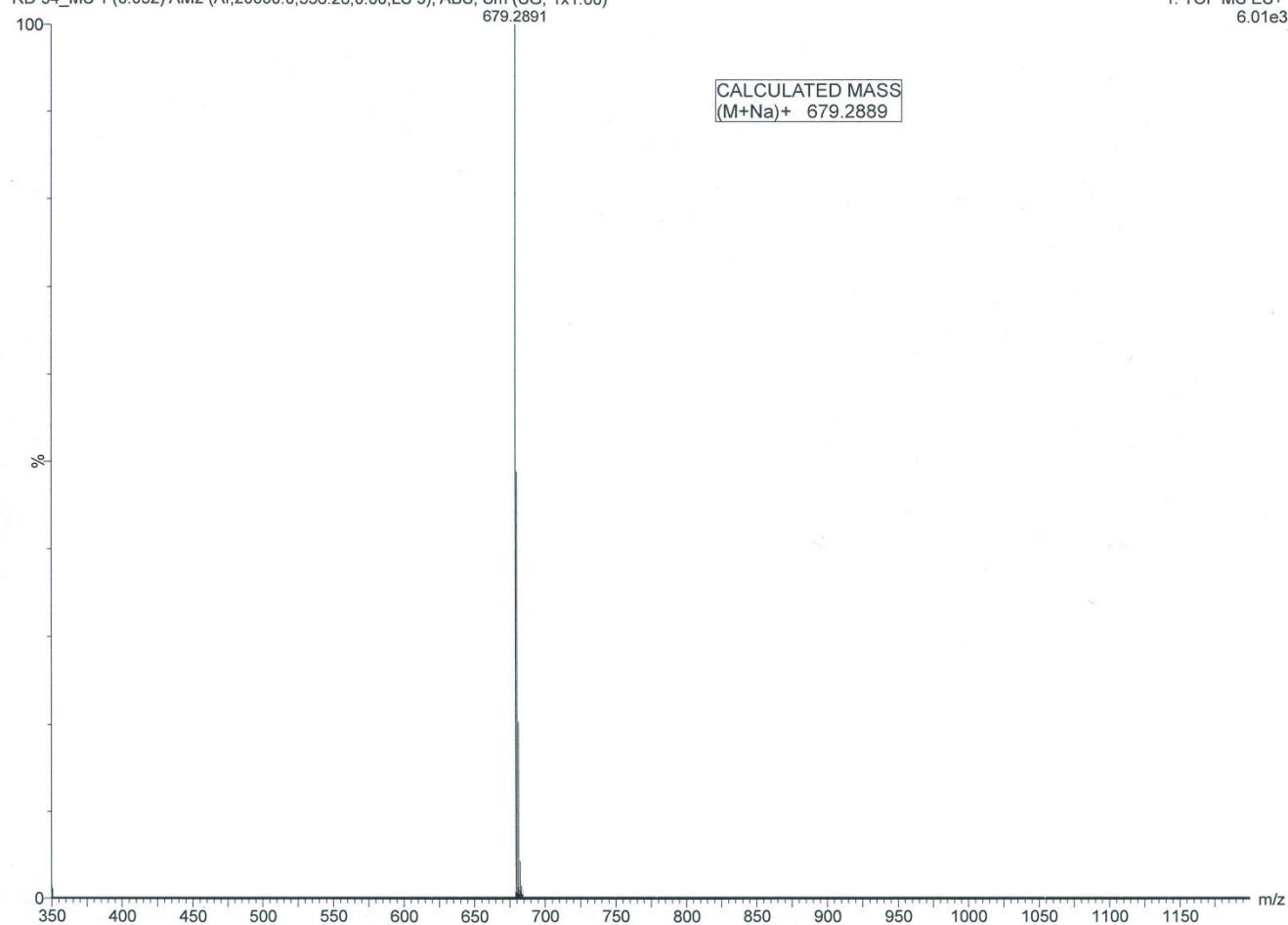


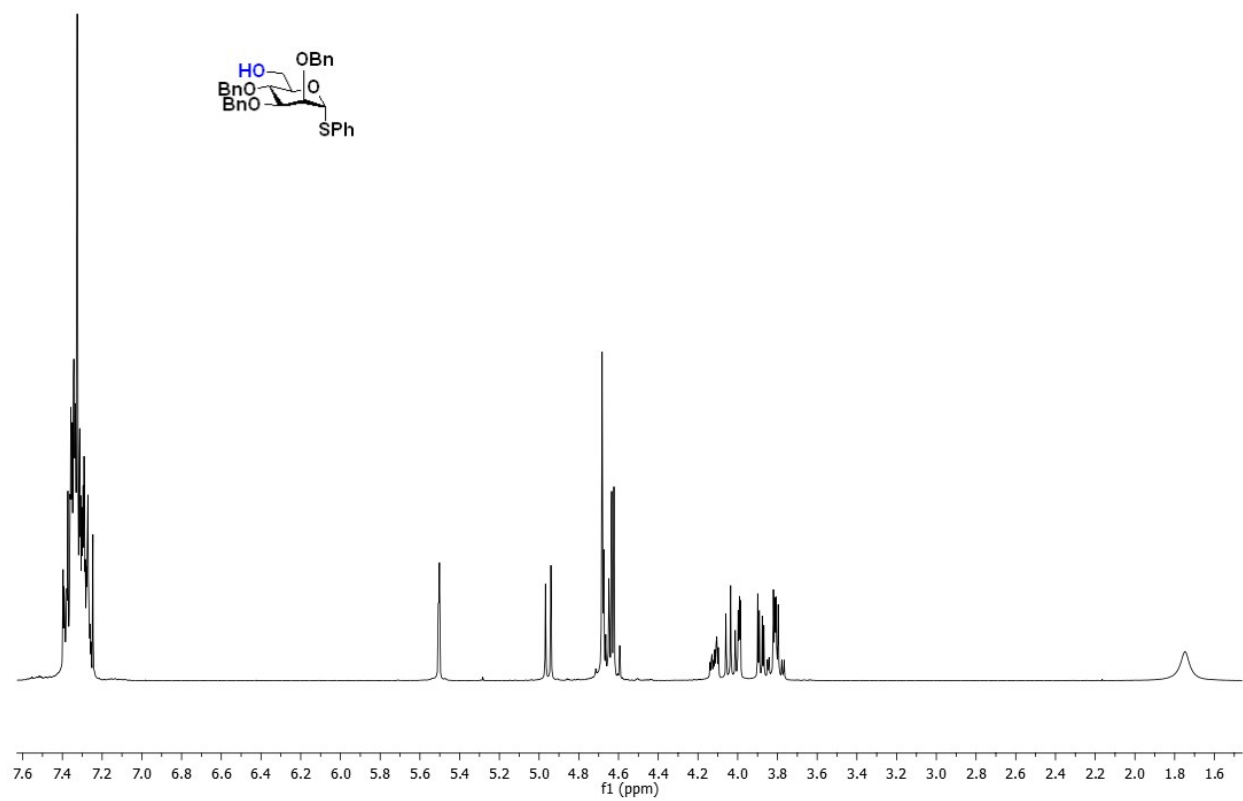
RD 34_MS

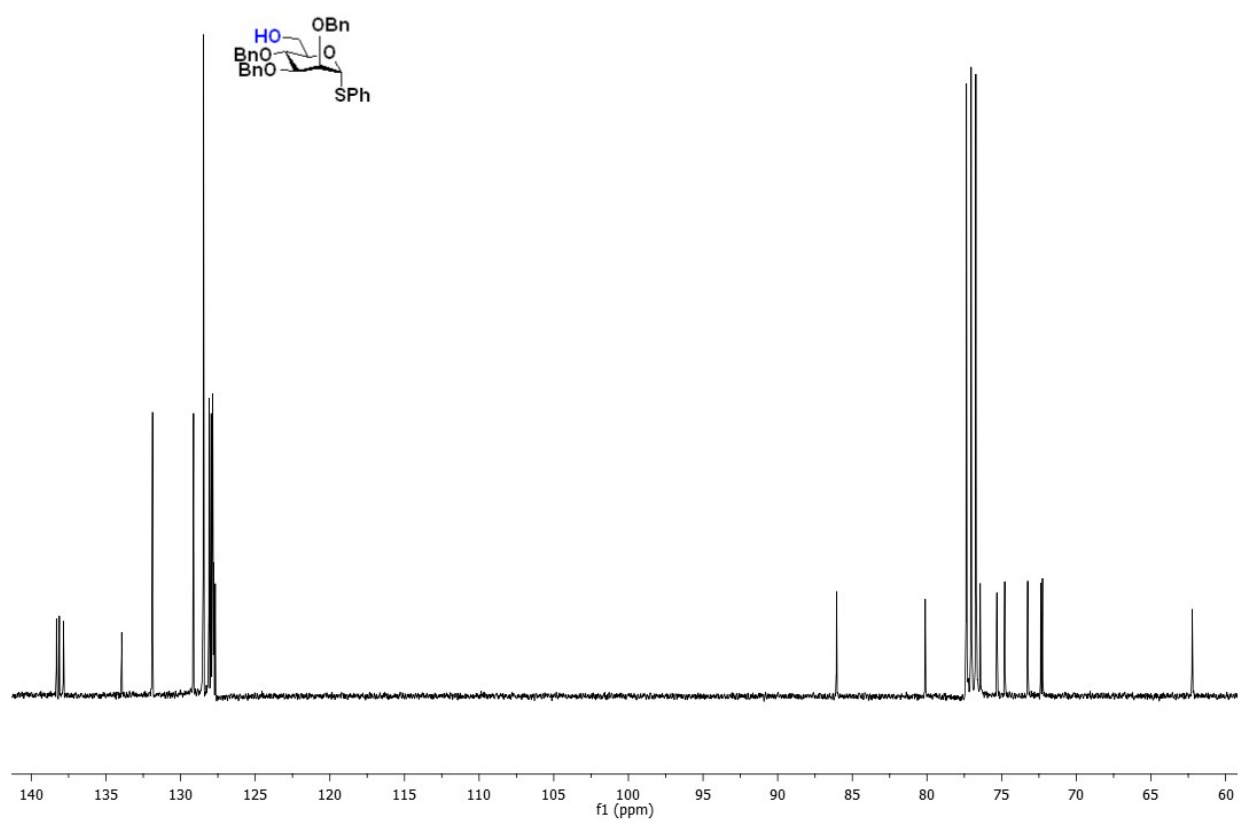
IISER PUNE

RD 34_MS 1 (0.052) AM2 (Ar,20000.0,556.28,0.00,LS 3); ABS; Sm (SG, 1x1.00)

1: TOF MS ES+
6.01e3





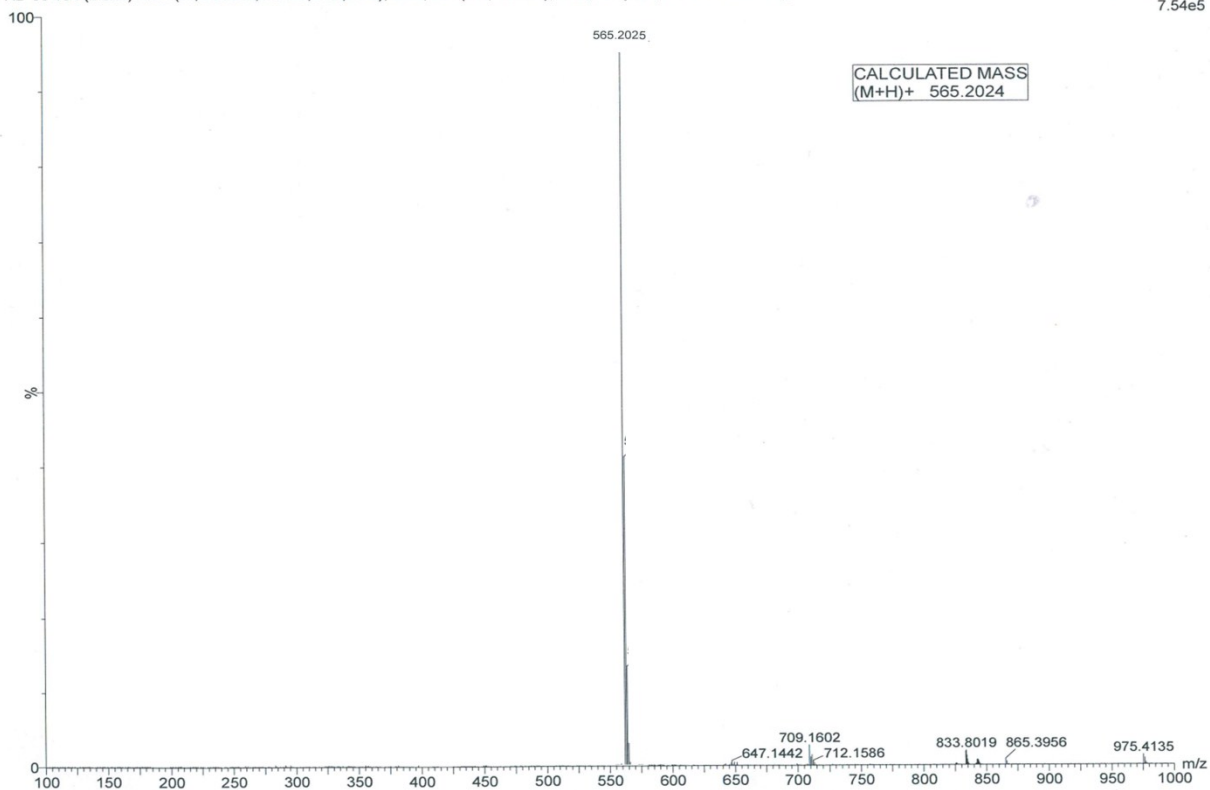


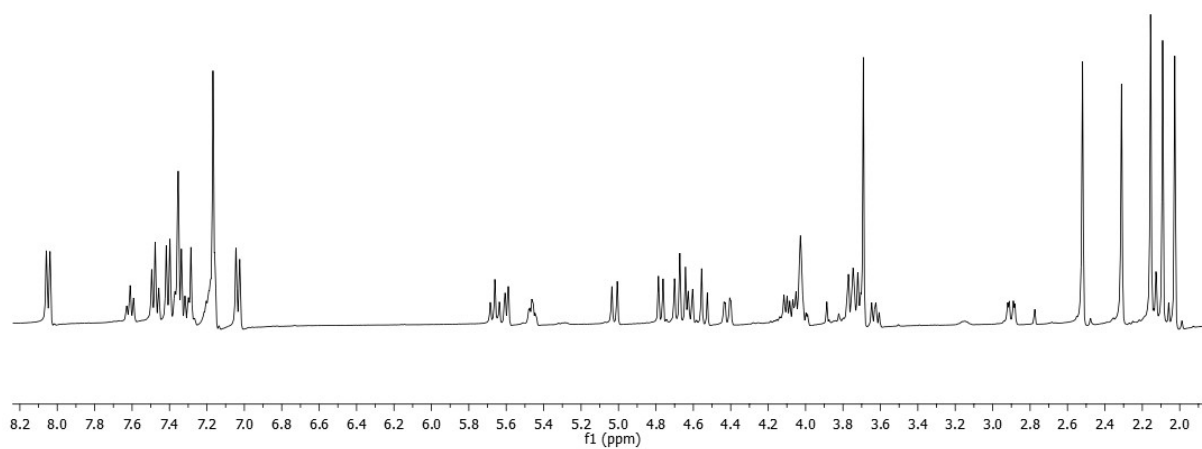
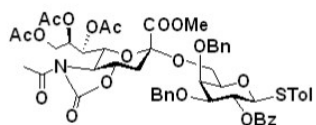
RD 38

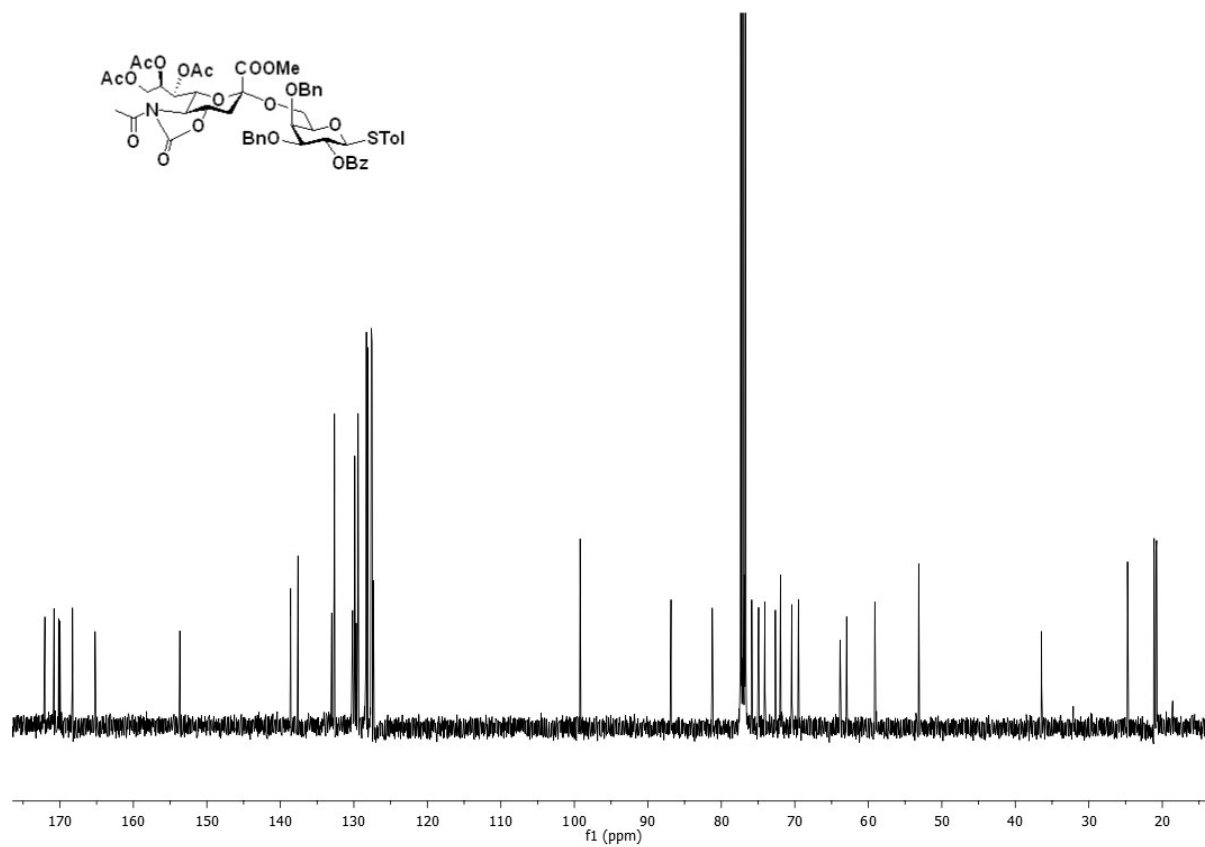
IISER PUNE

RD 38 154 (2.821) AM2 (Ar,20000.0,556.28,0.00,LS 3); ABS; Sm (SG, 1x1.00); Cm (153:159-(138:151+163:175))

1: TOF MS ES+
7.54e5





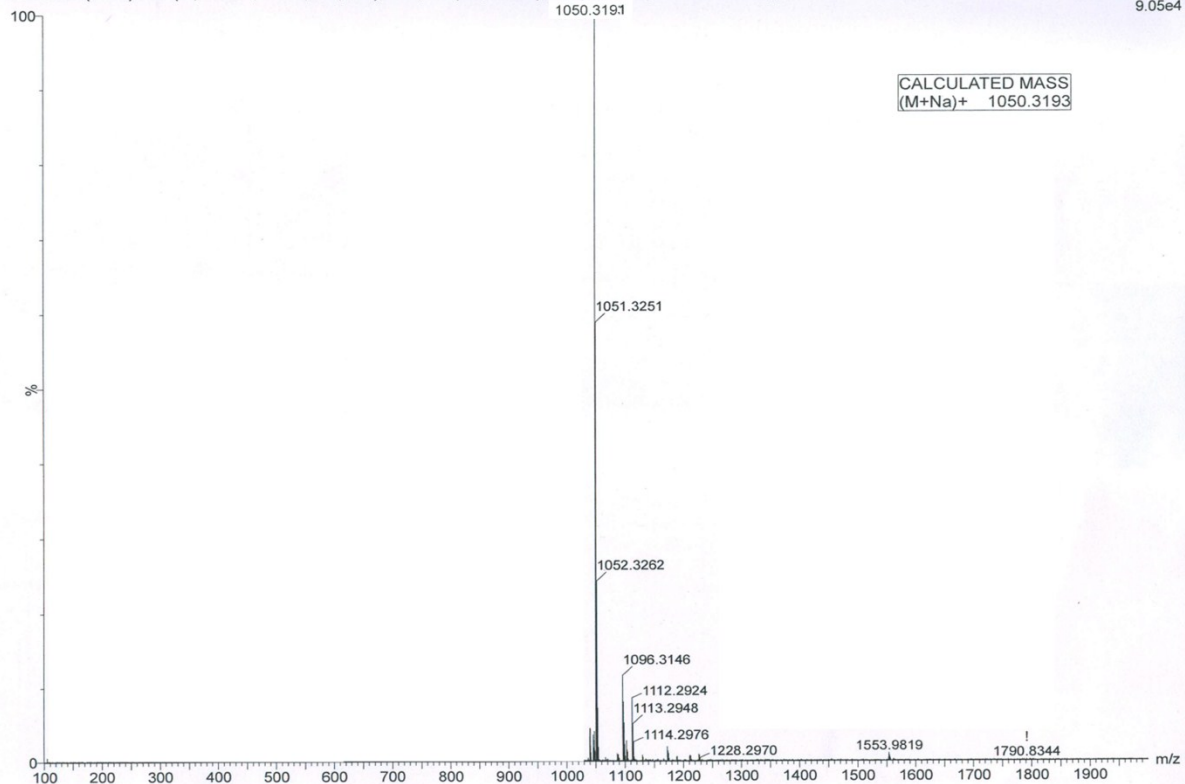


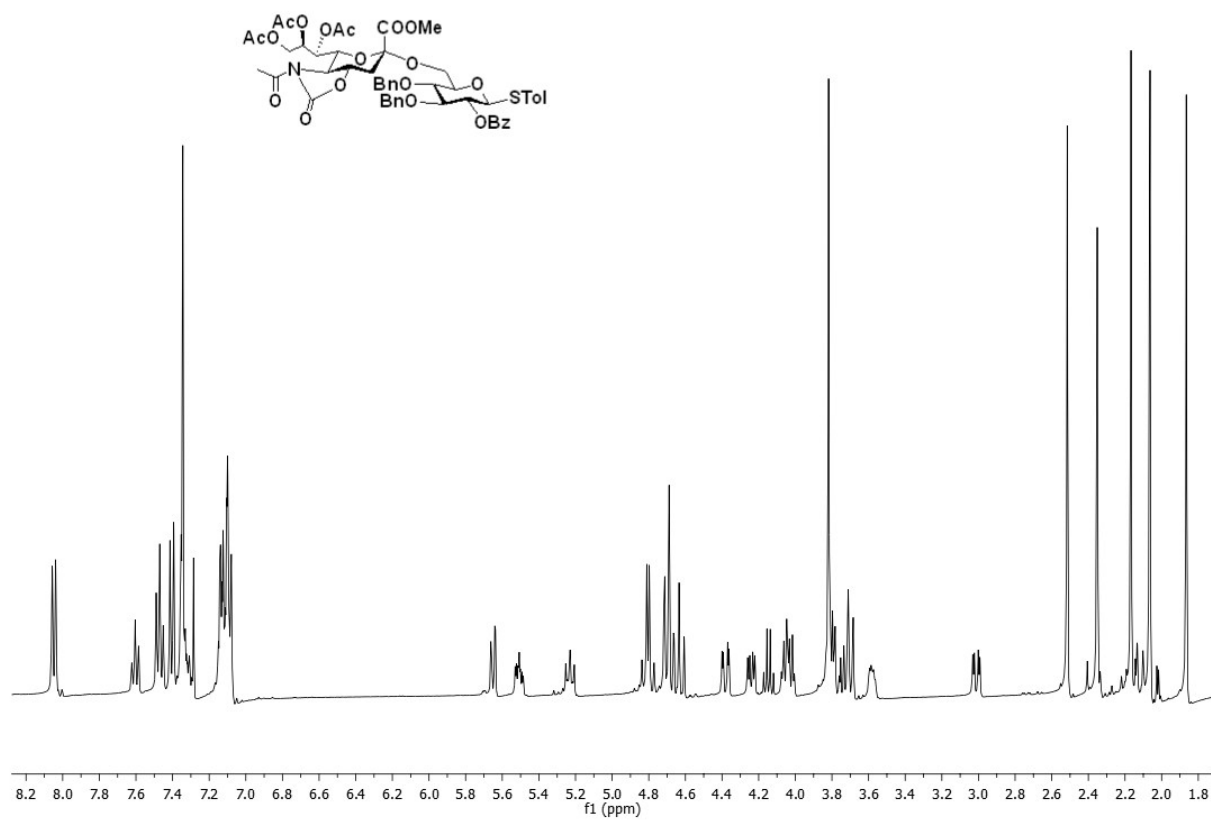
RD-26

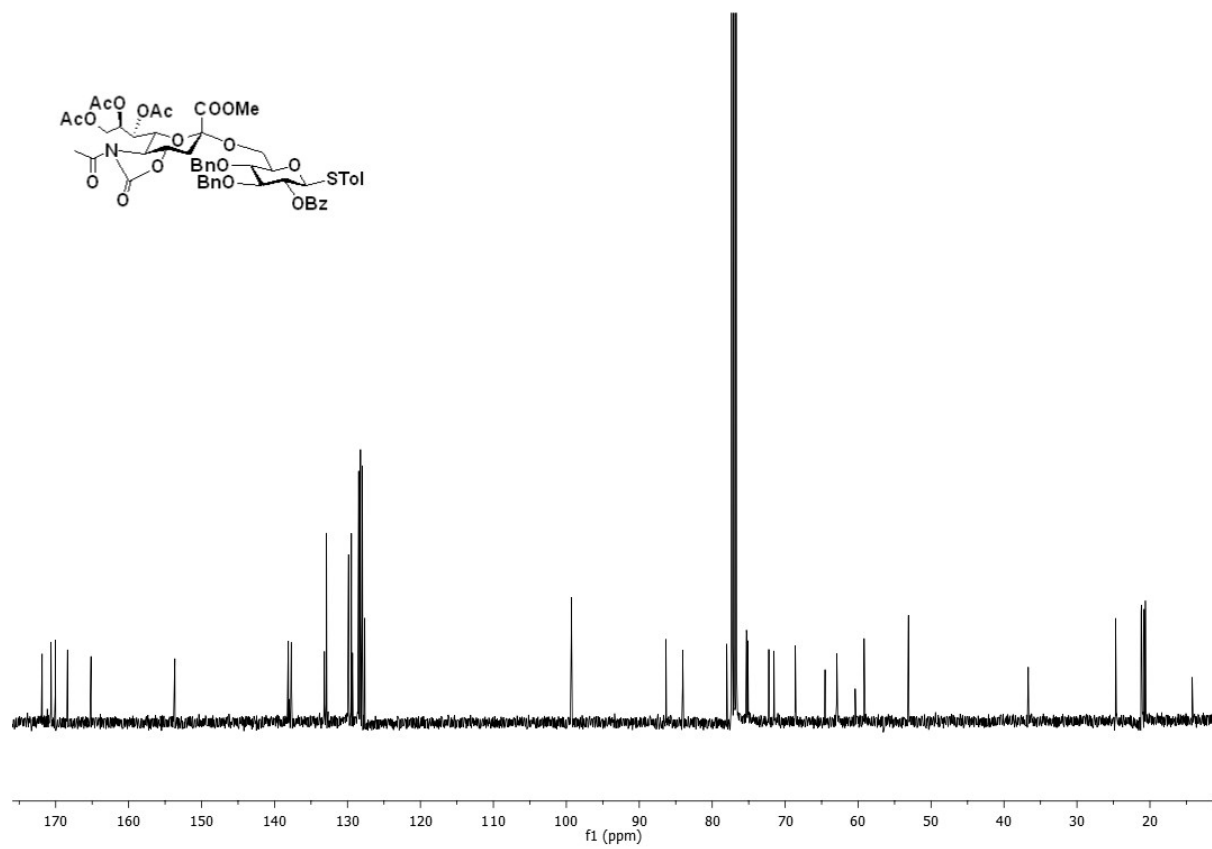
IISER PUNE

RD-26 152 (2.787) AM2 (Ar,20000.0,556.28,0.00,LS 3); ABS; Sm (SG, 1x1.00); Cm (148:160-(138:146+166:179))

1: TOF MS ES+
9.05e4





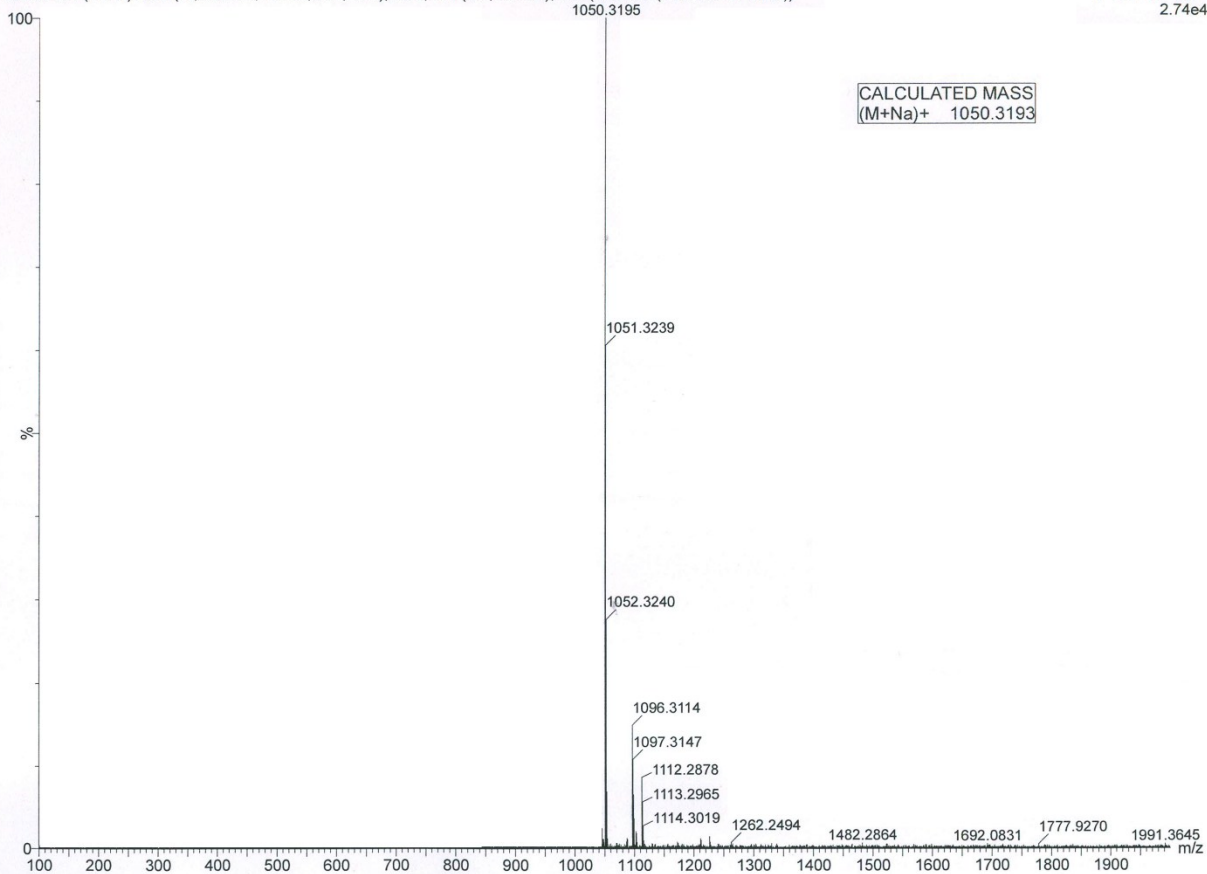


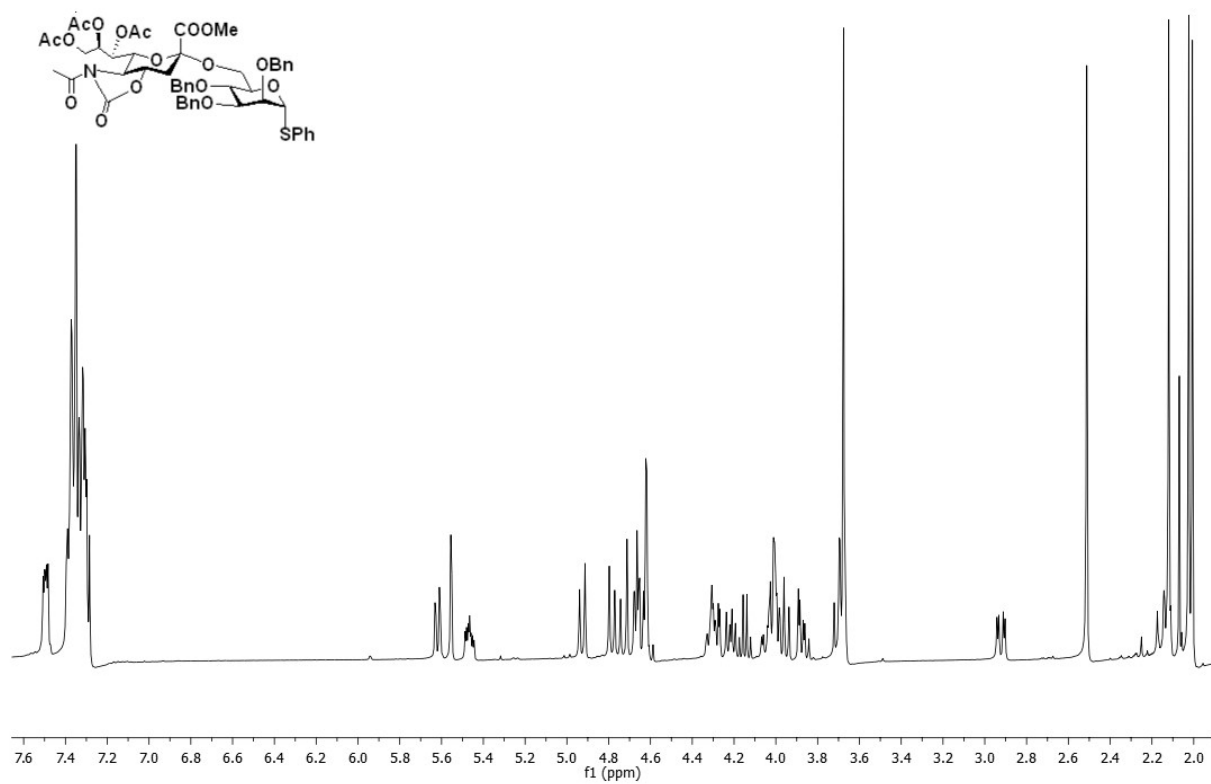
RD 39

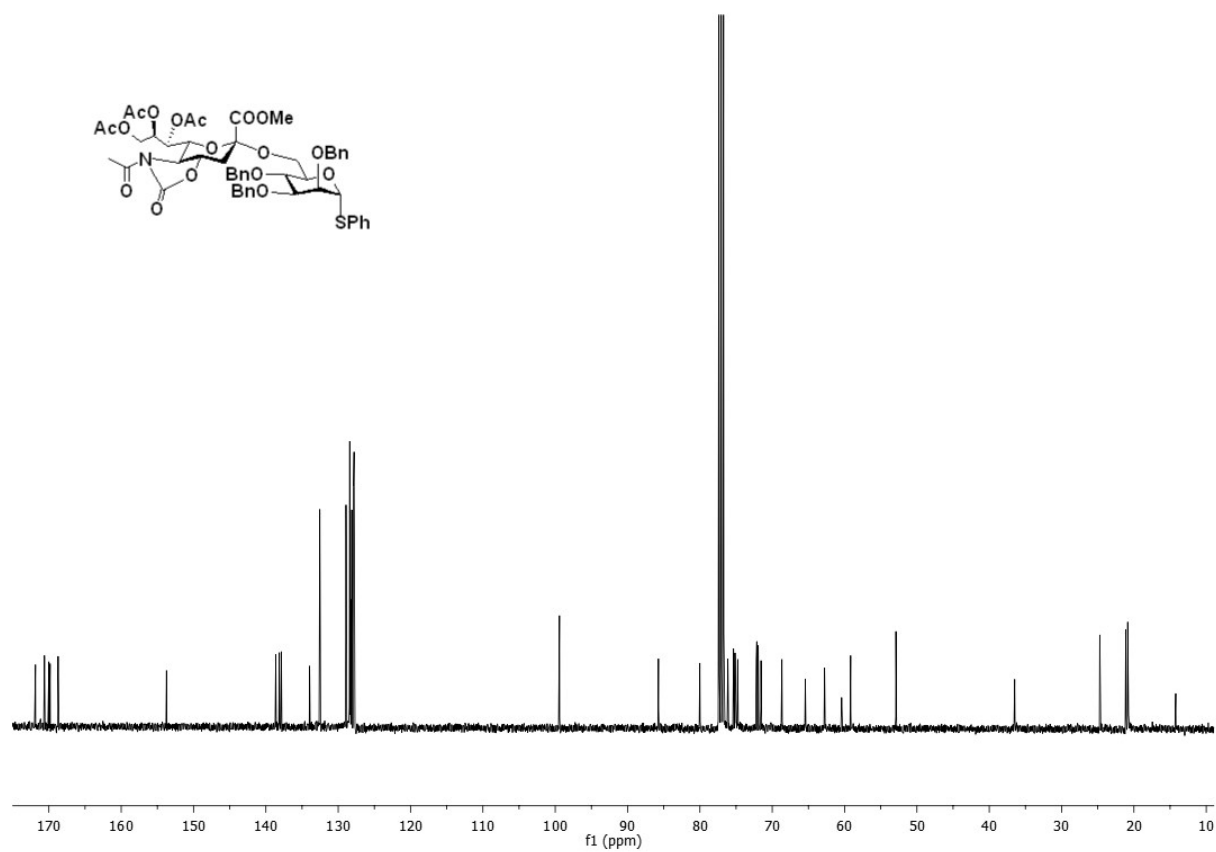
IISER PUNE

RD 39 162 (2.958) AM2 (Ar,20000.0,556.28,0.00,LS 3); ABS; Sm (SG, 1x1.00); Cm (160:168-(149:158+174:184))

1: TOF MS ES+
2.74e4





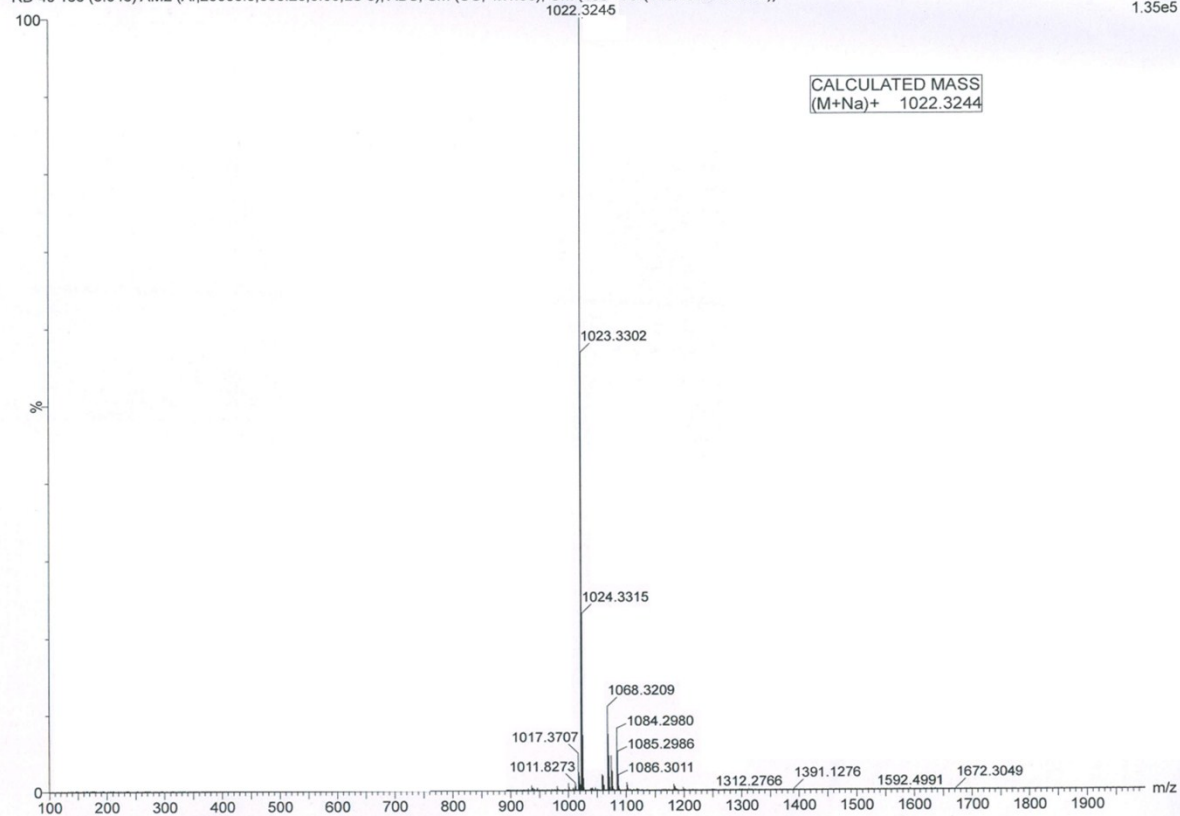


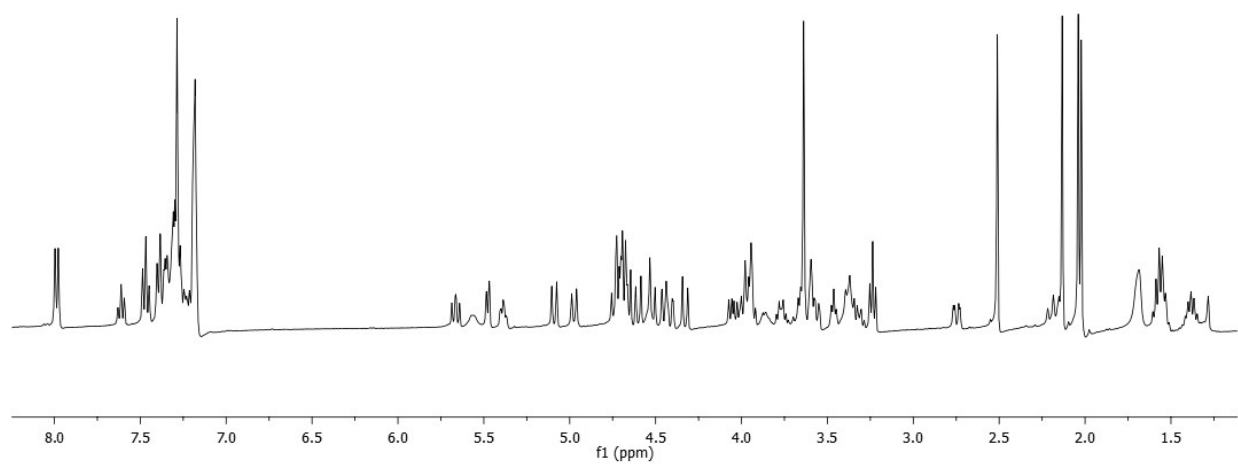
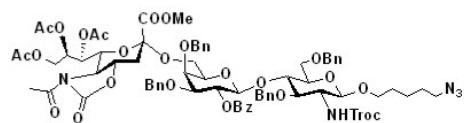
RD 40

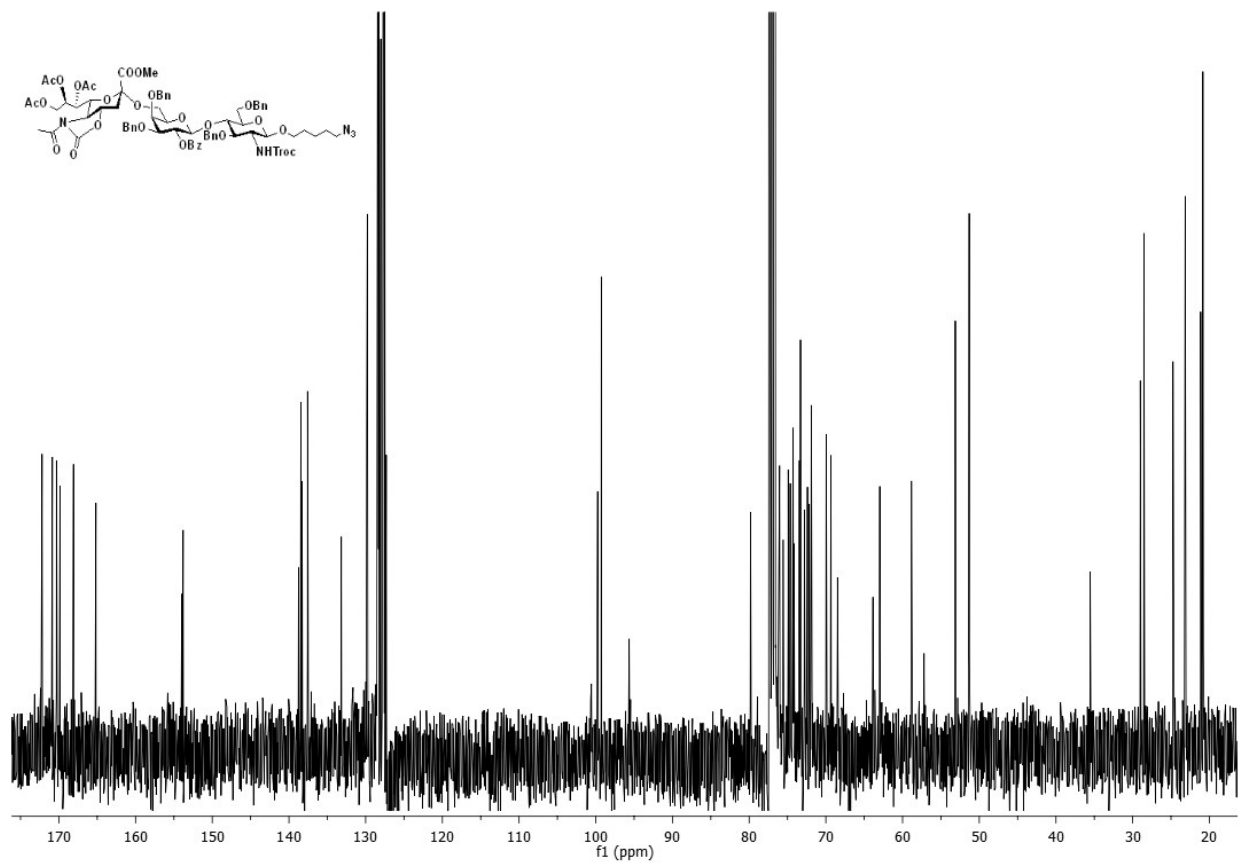
IISER PUNE

RD 40 166 (3.046) AM2 (Ar,20000.0,556.28,0.00,LS 3); ABS; Sm (SG, 1x1.00); Cm (163:175-(145:160+180:192))

1: TOF MS ES+
1.35e5





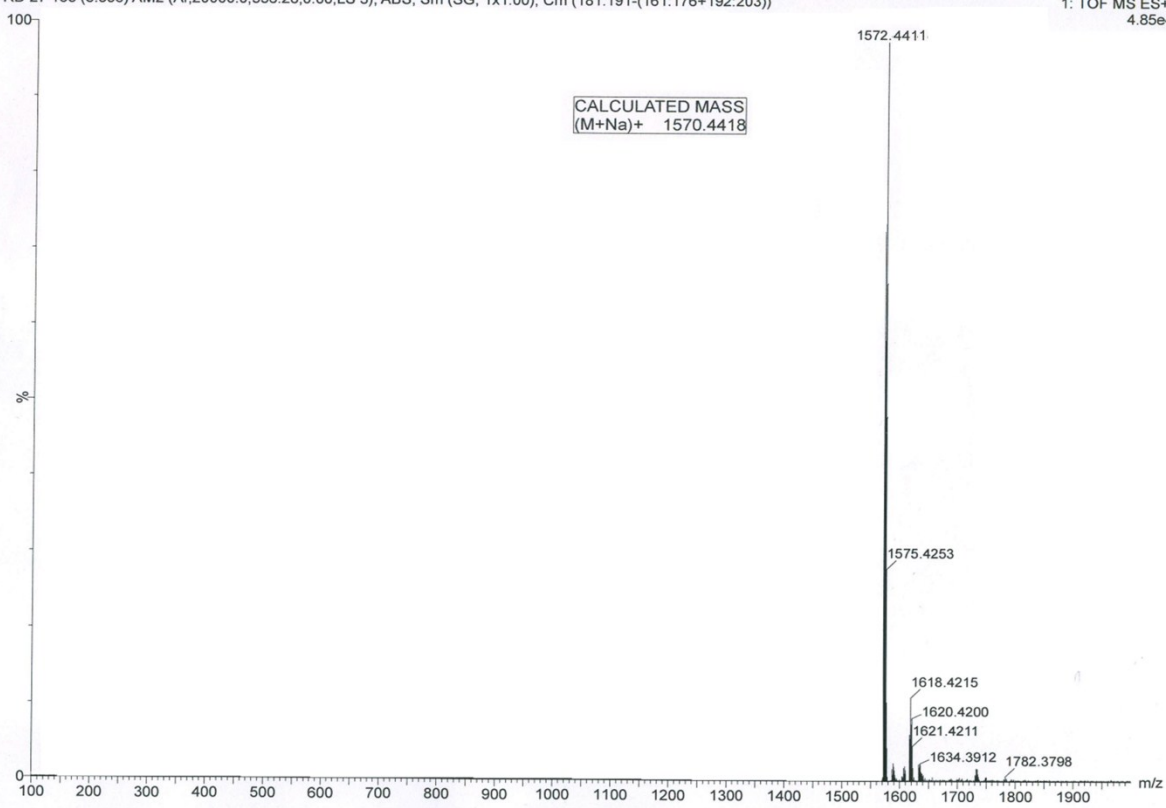


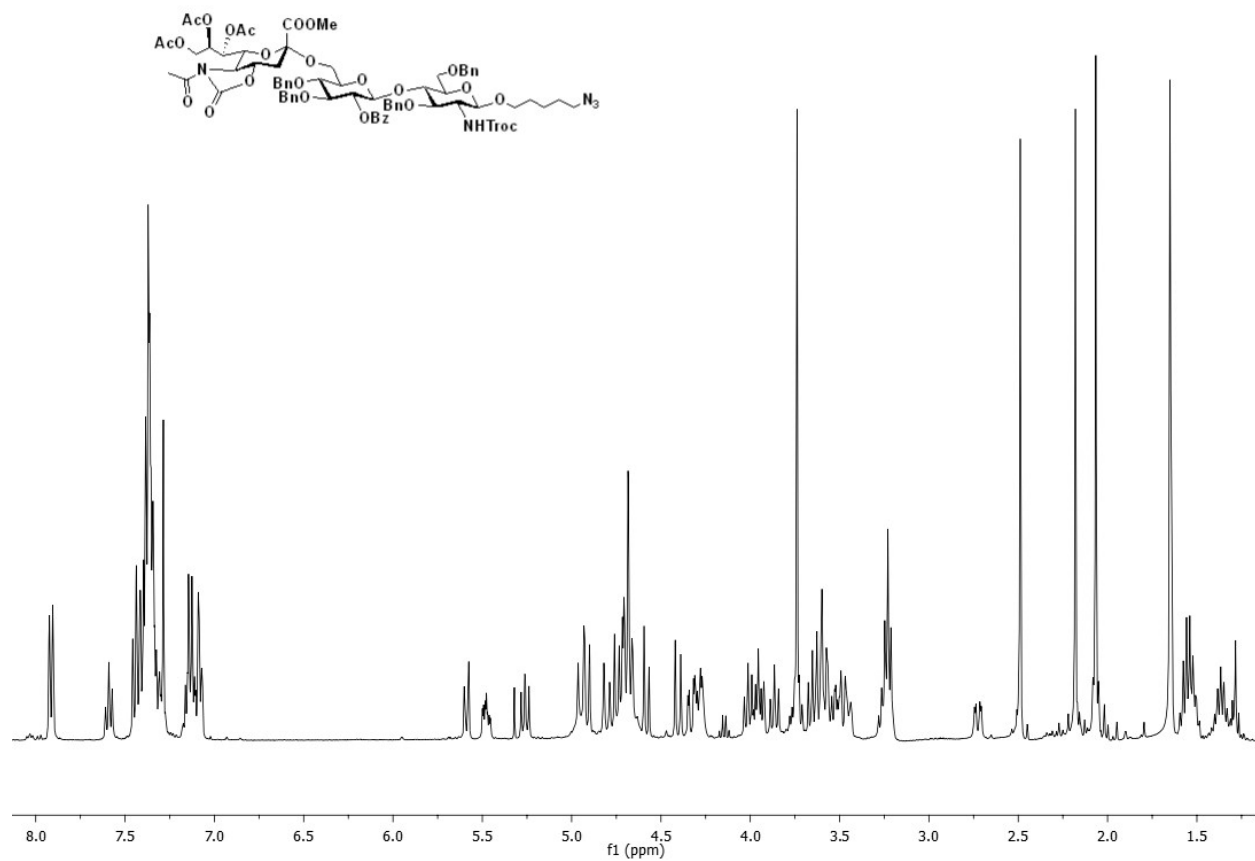
RD 27

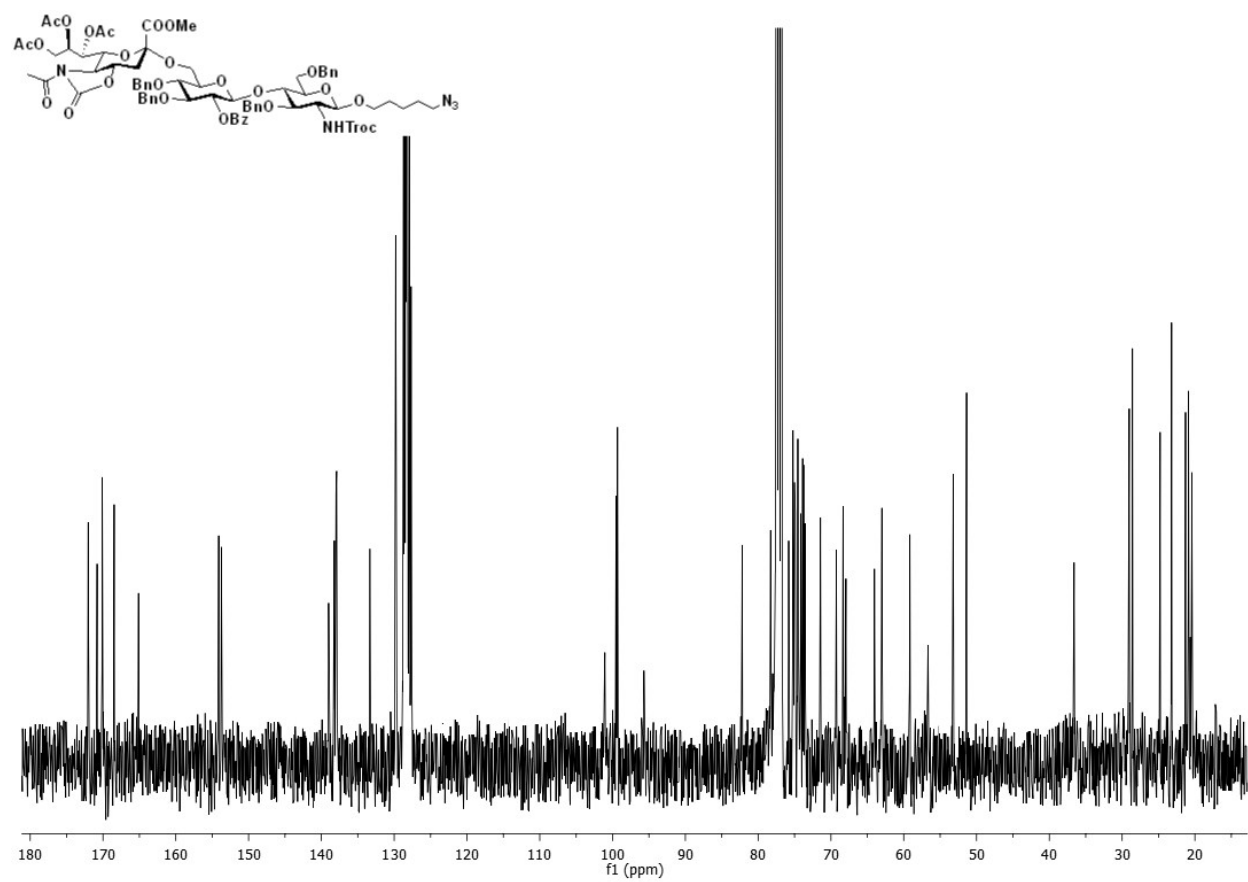
IISER PUNE

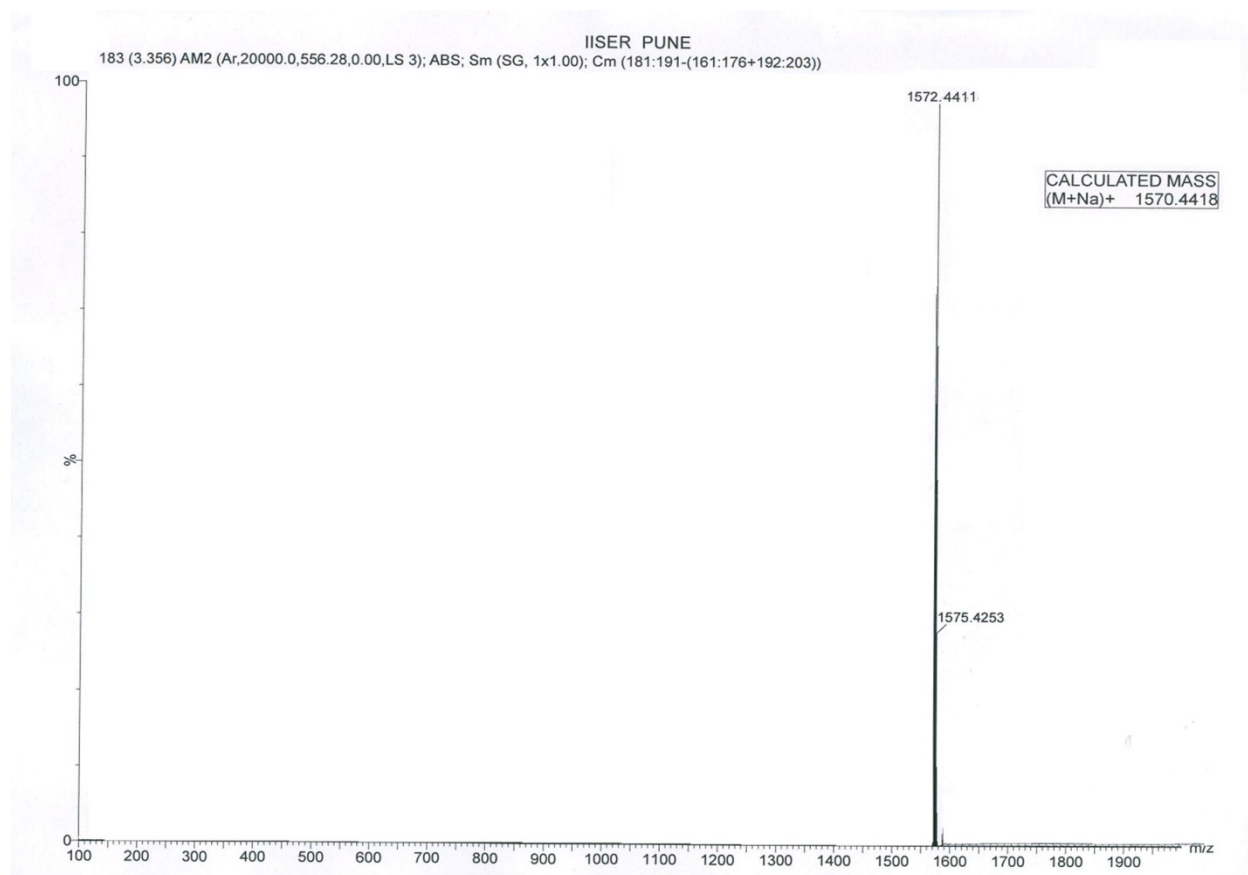
RD 27 183 (3.356) AM2 (Ar,20000.0,556.28,0.00,LS 3); ABS; Sm (SG, 1x1.00); Cm (181:191-(161:176+192:203))

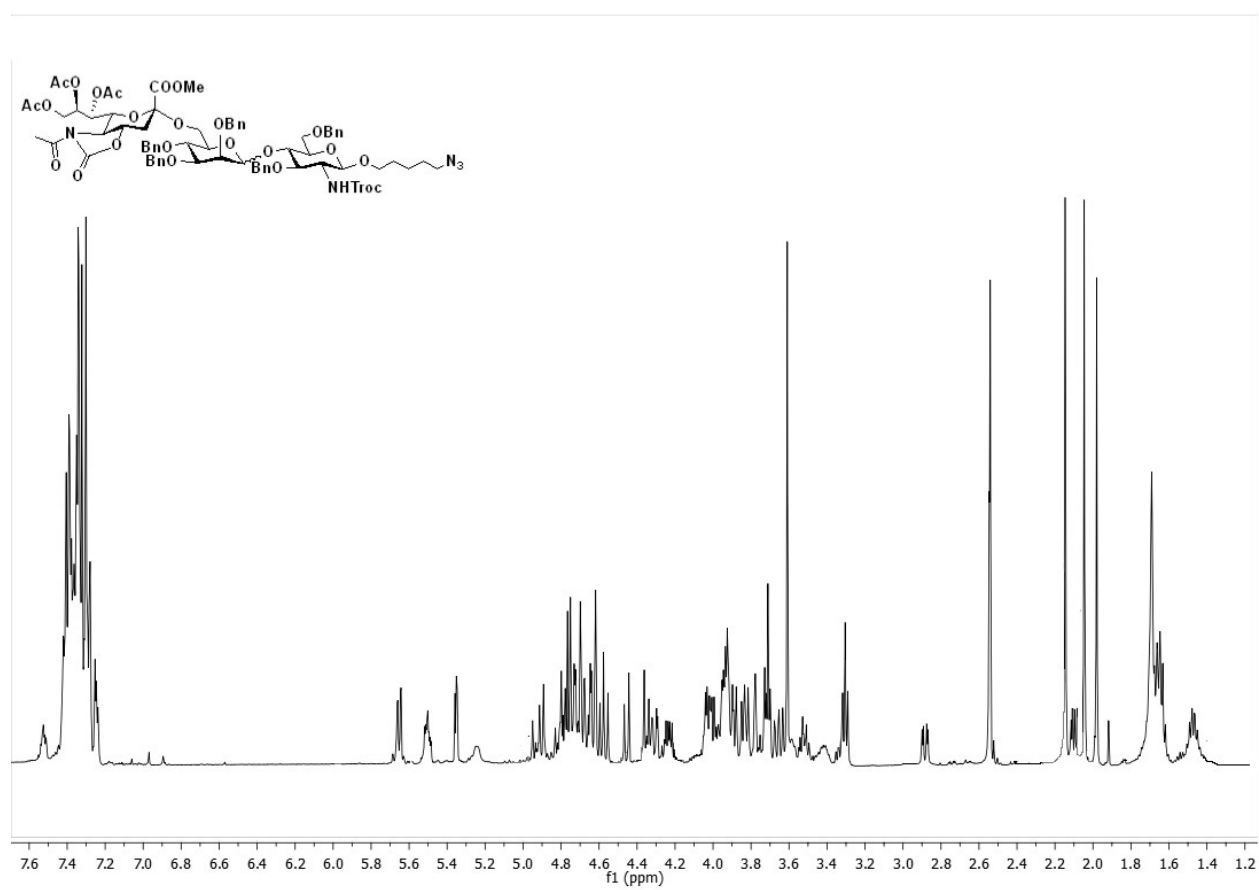
1: TOF MS ES+
4.85e4

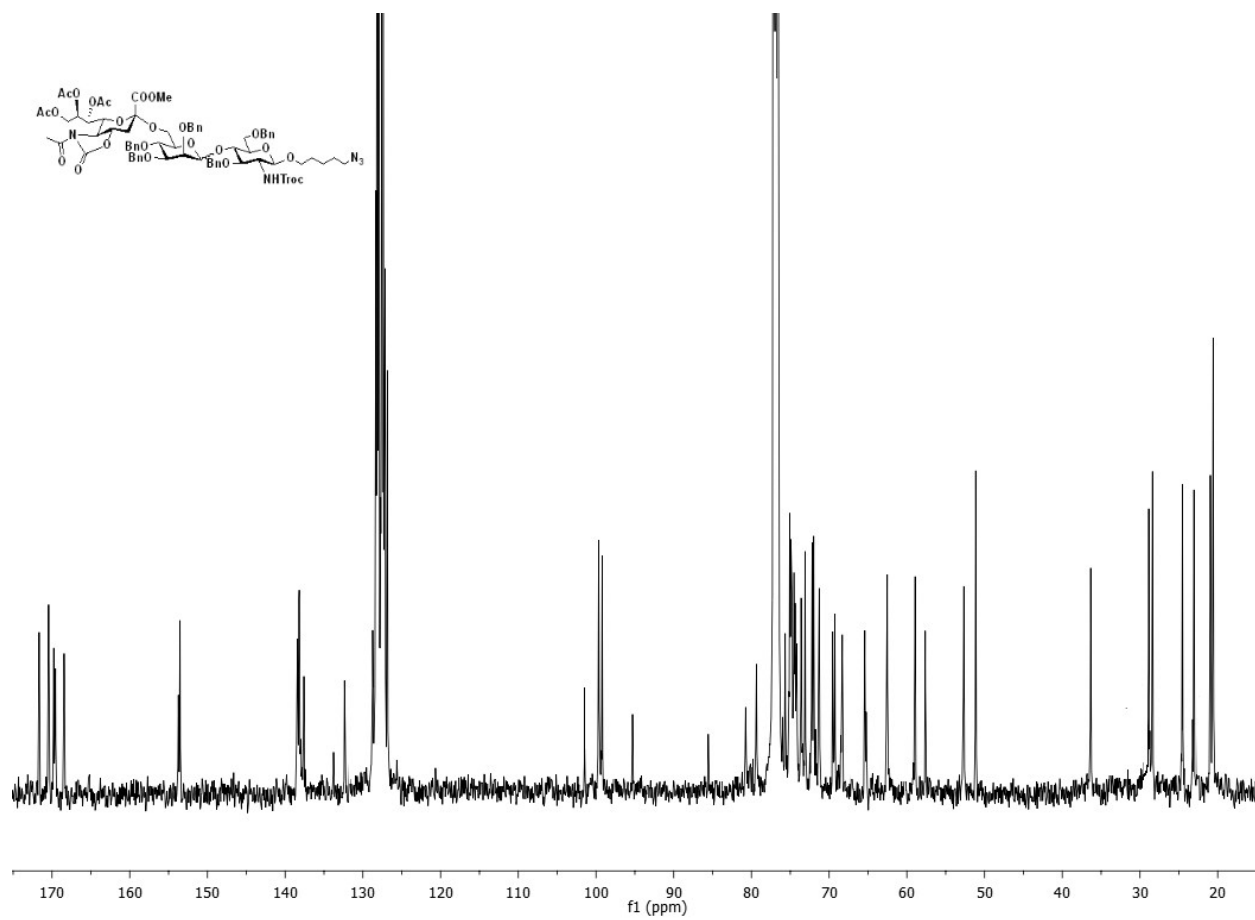


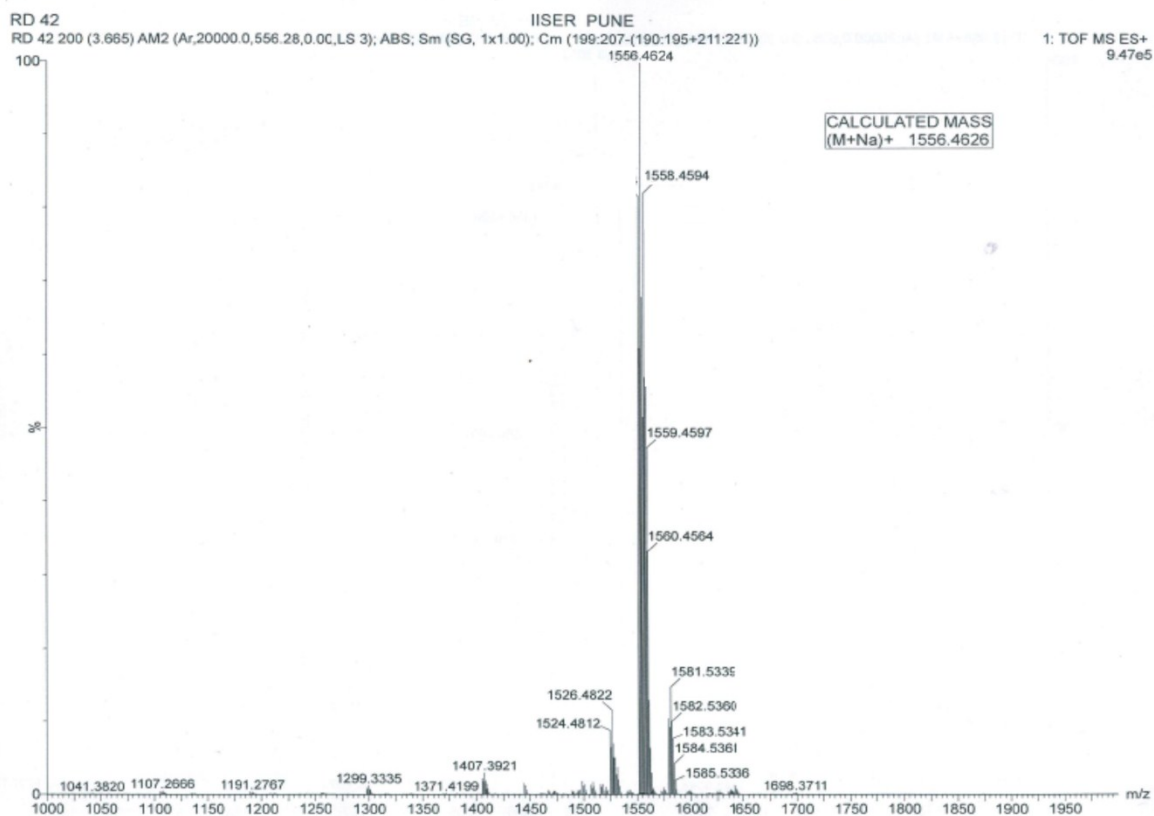


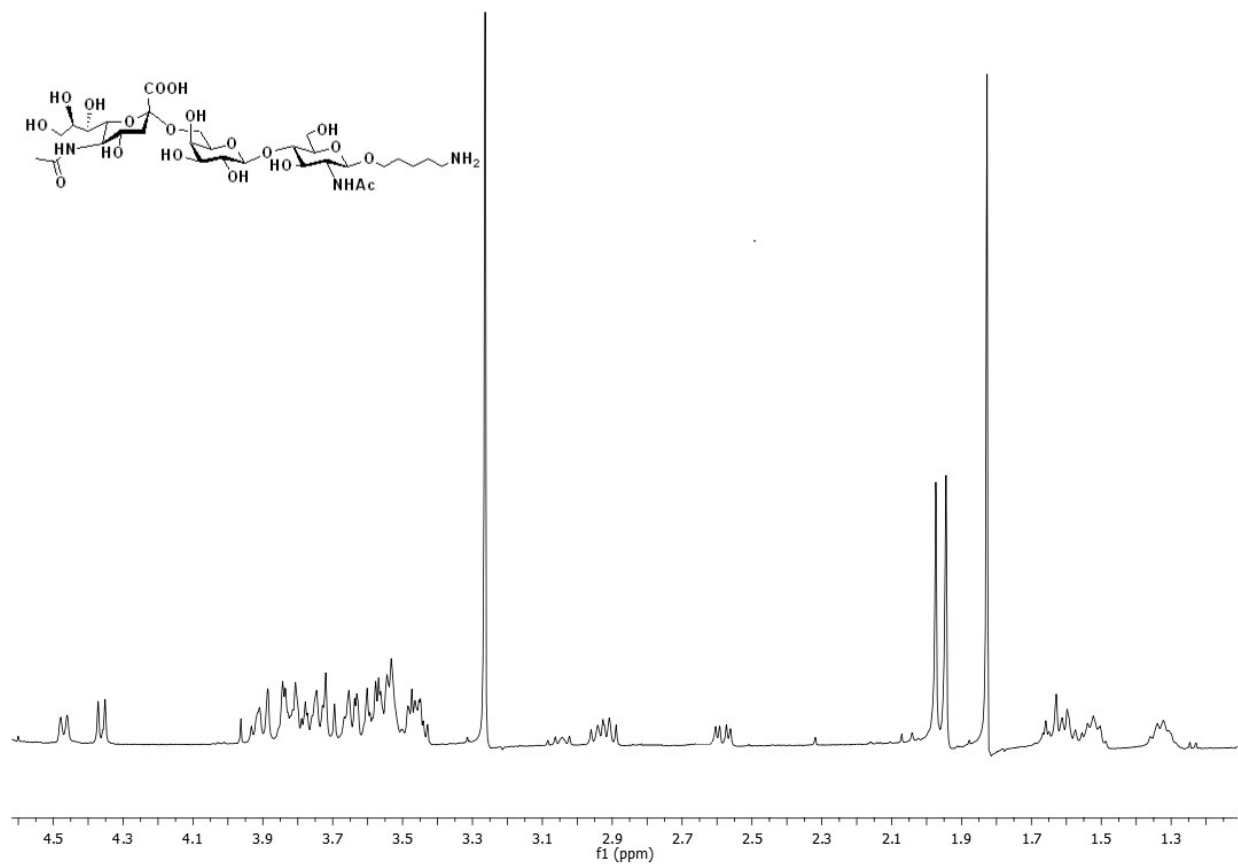


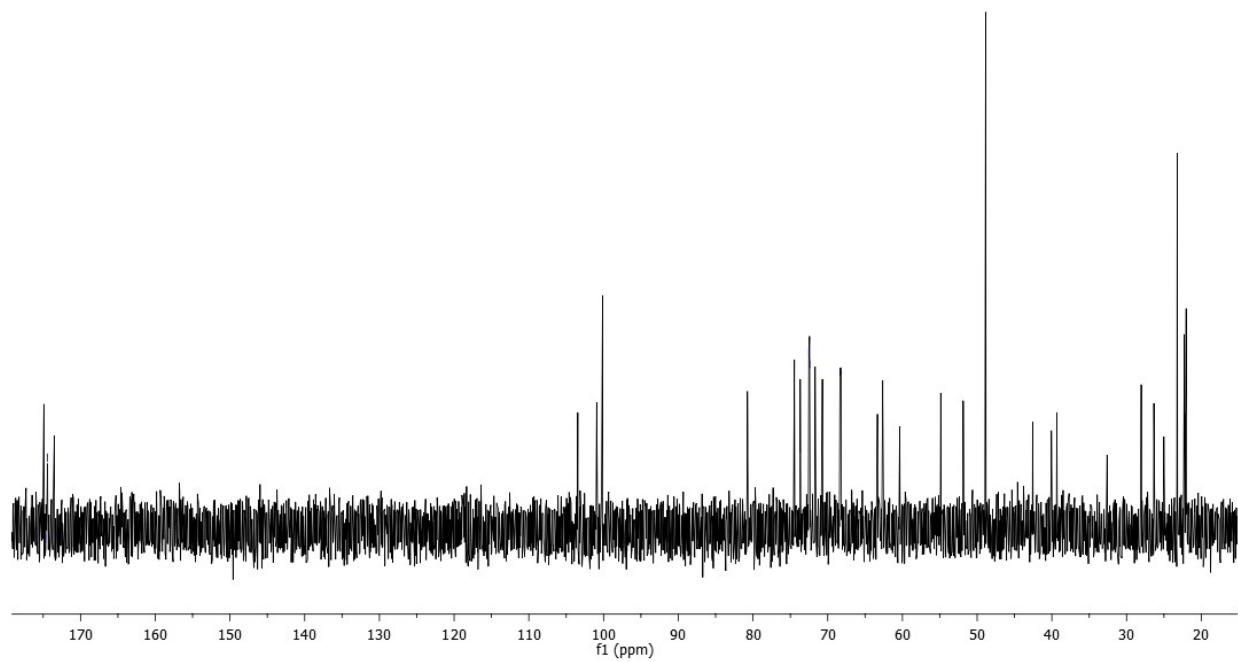
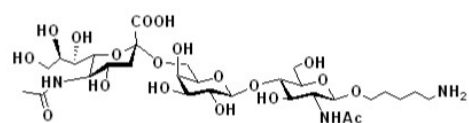










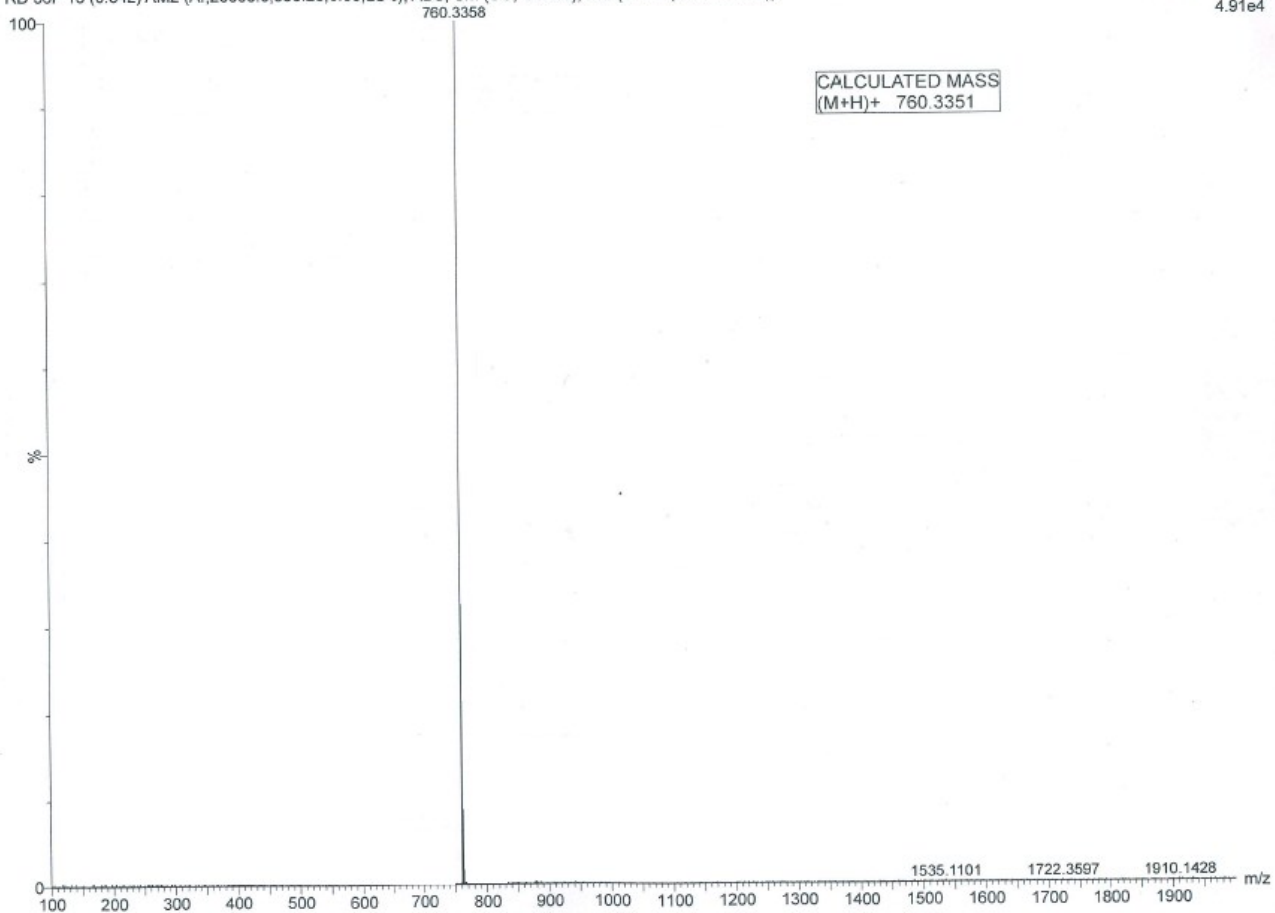


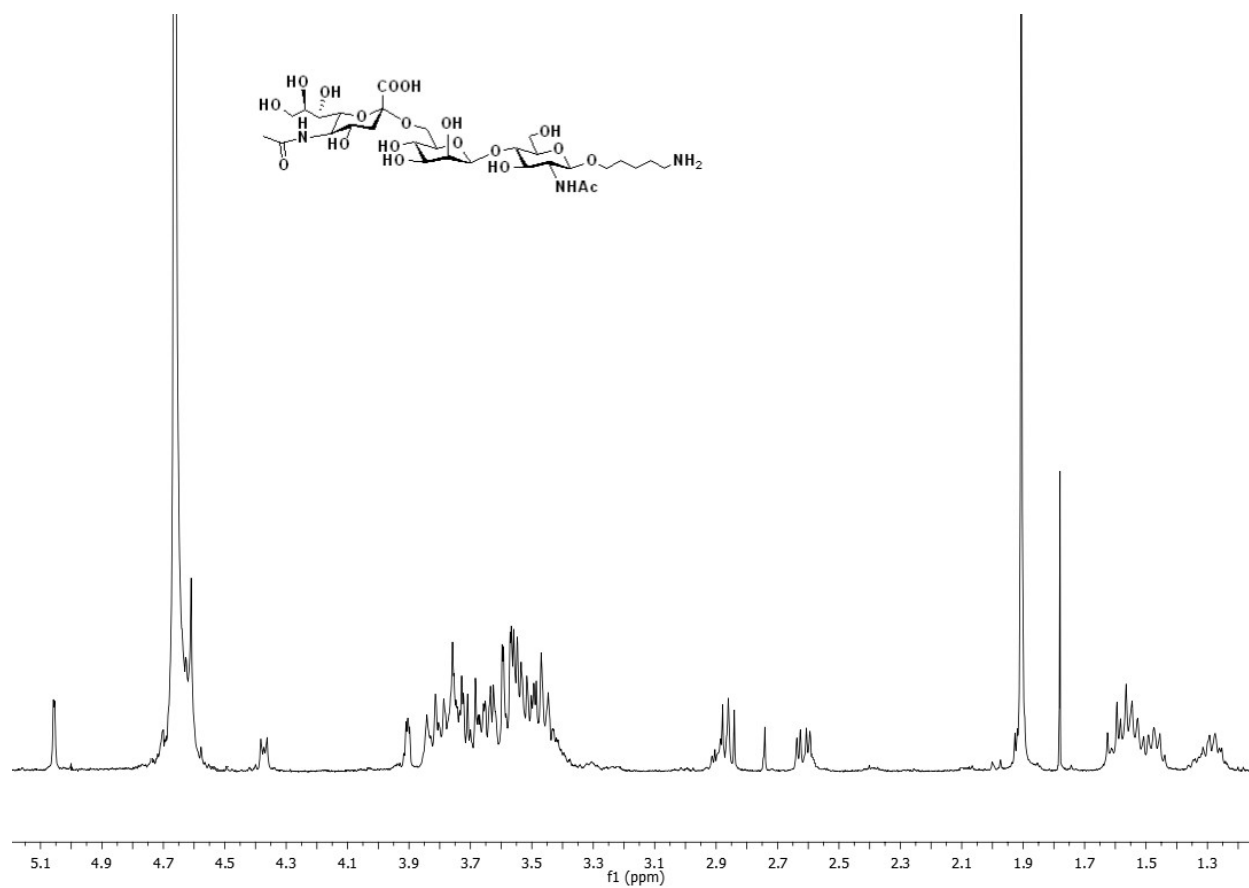
RD 36F

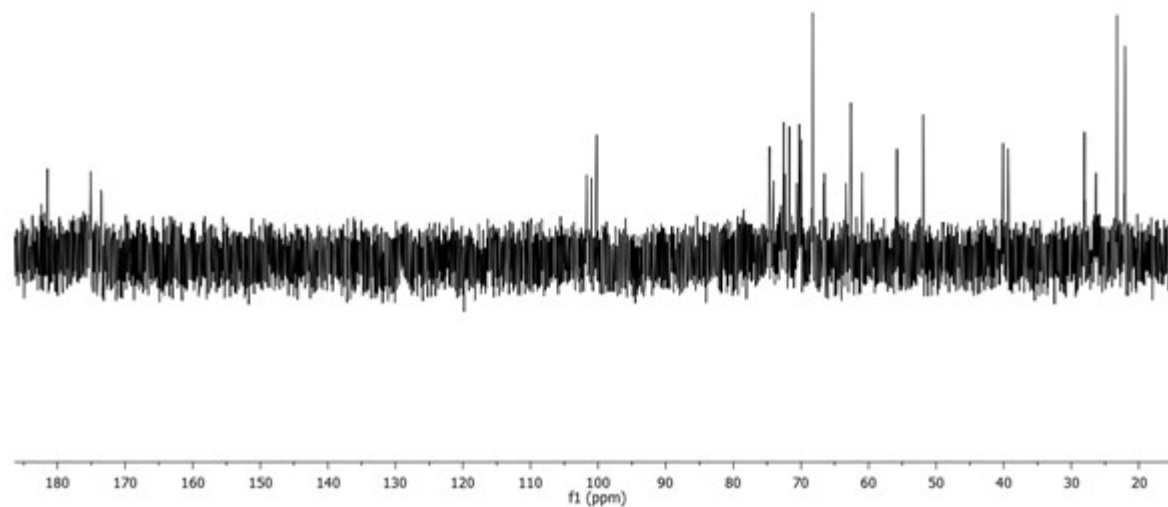
IISER PUNE

RD 36F 18 (0.342) AM2 (Ar,20000.0,556.28,0.00,LS 3); ABS; Sm (SG, 1x1.00); Cm (17:21-(11:17+22:33))

1: TOF MS ES+
4.91e4







RD 44F

IISER PUNE

1: TOF MS ES+

RD 44F 18 (0.342) AM2 (Ar,20000.0,556.28,0.00,LS 3); ABS; Sm (SG, 1x1.00); Cm (17:21-(9:14+23:33))

