

Supporting Information for:

A Ni-PyBisulidine Complex for the Asymmetric Hydrophosphonylation of Aldehydes

Youmao Zeng,^{a‡} Ping Deng,^{a‡} Shixiong Zhang,^a Hong Yan,^a Guojuan Liang,^a Yan Xiong^b and Hui Zhou^{*a}

hzhou@cqmu.edu.cn

^a *School of Pharmaceutical Science, Chongqing Medical University, Chongqing, China*

^b *School of Chemistry and Chemical Engineering, Chongqing University, Chongqing, China*

Contents:

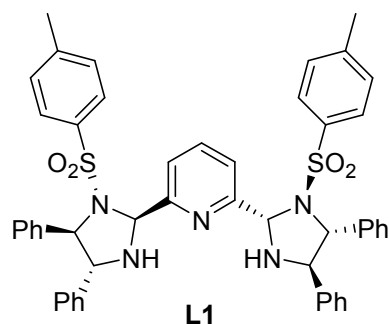
1. General information	S2
2. The preparation and characterization of the ligands	S2
3. General procedures for the preparation of the catalyst.....	S3
4. ESI-HRMS analysis of Ni-L1 complex and the proposed structure of the complex	S3
5. The effects of the catalyst loading and the concentration of benzaldehyde.	S5
6. General Procedure for Catalytic Asymmetric Reaction	S5
7. Characterization of products	S6
8. Copy of NMR spectra.	S19

1. General information

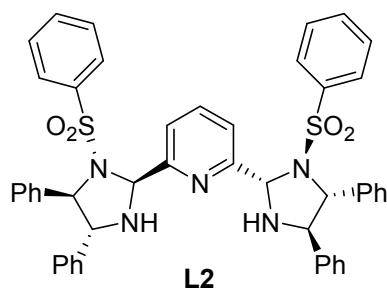
All reagents were obtained from Adamas, Aladin, TCI, or Acros etc. without further purification unless otherwise noted. High resolution mass spectra were measured on commercial instruments. NMR spectra were recorded on commercial instruments and operating at 500 MHz for ^1H NMR and 125 MHz for ^{13}C NMR. Chemical shifts were reported in ppm from tetramethylsilane with the solvent resonance as the internal standard (CDCl_3 , $\delta = 7.26$) in ^1H NMR spectra and Chemical shifts were reported in ppm from the tetramethylsilane with the solvent resonance as internal standard (CDCl_3 , $\delta = 77.0$) in ^{13}C NMR spectra. Spectra are reported as follows: chemical shift (δ ppm), multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet), coupling constants (Hz), integration, and assignment. The enantiomeric excess (*ee*) was determined by HPLC analysis. Analytical HPLC was performed on a Shimadzu liquid chromatography (LC-16), using a chiral DAICEL CHIRALCEL OD-H or CHIRALPAK AS-H or CHIRALPAK AD-H column at 210 nm. Optical rotations were measured on a commercial polarimeter and are reported as follows: $[\alpha]_D^{25}$ ($c = \text{g}/100 \text{ mL}$, solvent).

2. The preparation and characterization of the ligands

Ligands **L1-L4** were prepared according to the literature.¹



^1H NMR (500MHz, CDCl_3): δ 8.02-7.96 (m, 3H), 7.53 (d, $J = 8.2 \text{ Hz}$, 4H), 7.25-7.17 (m, 4H), 7.16-7.10 (m, 16H), 7.00 (d, $J = 7.2 \text{ Hz}$, 4H), 5.83 (s, 2H), 4.71 (d, $J = 5.4 \text{ Hz}$, 2H), 4.26 (d, $J = 5.4 \text{ Hz}$, 2H), 2.39 (s, 6H).

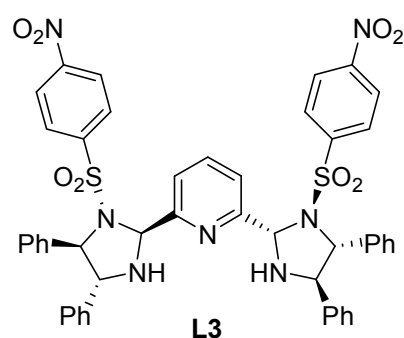


^1H NMR (500 MHz, CDCl_3) δ 7.99-7.98 (m, 2H), 7.65-7.63 (m, 3H), 7.52-7.49 (m, 2H), 7.35 (t, $J = 7.9 \text{ Hz}$, 4H), 7.18-7.09 (m, 16H), 6.98-6.97 (m, 4H), 5.88 (s, 2H), 4.71 (d, $J = 5.8 \text{ Hz}$, 2H), 4.26 (d, $J = 5.8 \text{ Hz}$, 2H). ^{13}C NMR (125 MHz, CDCl_3) δ 158.14, 139.51, 139.01, 138.22, 138.08, 128.99, 128.62, 128.37, 127.85, 127.61, 127.10, 126.79, 123.99,

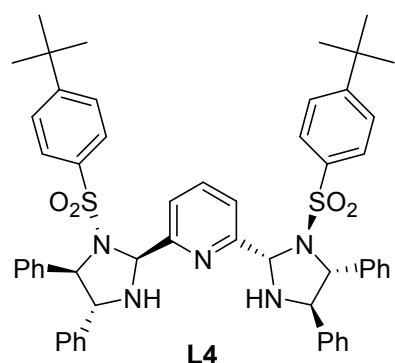
78.30, 71.64, 69.49. HRMS (ESI): m/z calcd for $\text{C}_{47}\text{H}_{41}\text{N}_5\text{NaO}_4\text{S}_2$ [$\text{M} + \text{Na}$] $^+$: 826.24922, found

[1] A. Gonzalez-de-Castro, C. M. Robertson, J. Xiao, *J. Am. Chem. Soc.*, **2014**, *136*, 8350-8360.

826.24686.



^1H NMR (500 MHz, CDCl_3) δ 8.02-7.94 (m, 7H), 7.50 (d, J = 8.2 Hz, 4H), 7.20-7.11 (m, 20H), 5.75 (s, 2H), 4.96 (s, 2H), 4.45 (s, 2H). ^{13}C NMR (125 MHz, CDCl_3) δ 156.32, 156.07, 149.97, 143.28, 139.37, 139.25, 138.34, 128.84, 128.53, 128.33, 128.16, 125.83, 79.32, 70.67, 68.97. HRMS (ESI): m/z calcd for $\text{C}_{47}\text{H}_{39}\text{N}_7\text{NaO}_8\text{S}_2$ [$\text{M} + \text{Na}$] $^+$: 916.21937, found 916.21896.



^1H NMR (500 MHz, CDCl_3) δ 8.01-7.99 (m, 2H), 7.61 (d, J = 8.5 Hz, 4H), 7.37 (d, J = 8.6 Hz, 4H), 7.18-7.13 (m, 7H), 7.10-7.06 (m, 10H), 6.96-6.94 (m, 4H), 5.93 (s, 2H), 4.67 (d, J = 6.1 Hz, 2H), 4.25 (d, J = 6.0 Hz, 2H), 1.34 (s, 18H).

3. General procedures for the preparation of the catalyst

Method A (in situ): The mixture of $\text{Ni}(\text{OAc})_2 \cdot 4\text{H}_2\text{O}$ (0.02 mmol) and ligand (0.02 mmol) was stirred in a certain solvent (0.5 mL) at 35 °C for 1 h.

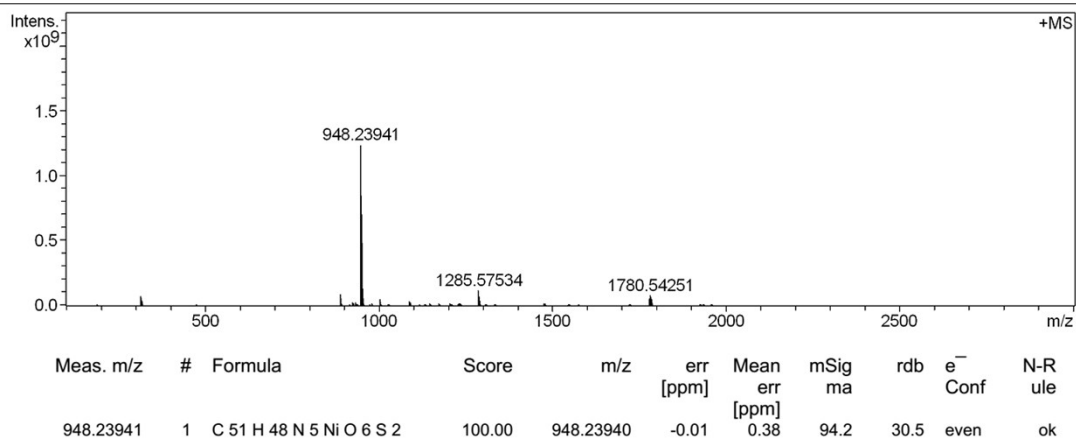
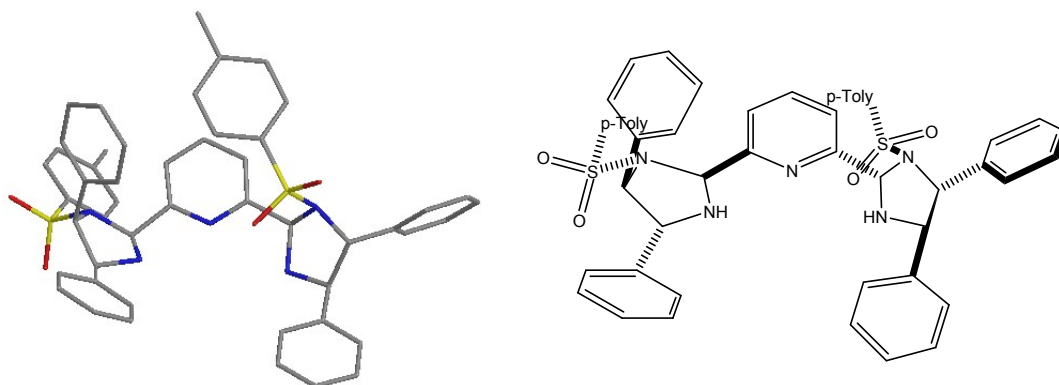
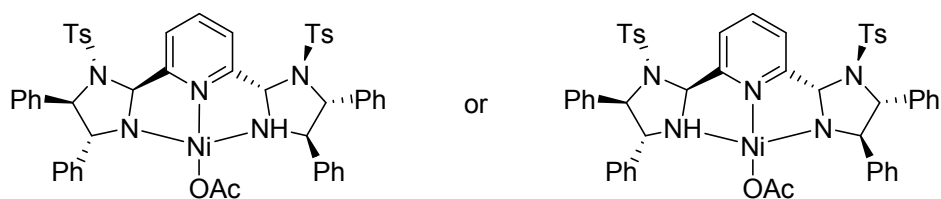
Method B (pre-prepared): The mixture of the **L1** (0.4 mmol) and $\text{Ni}(\text{OAc})_2 \cdot 4\text{H}_2\text{O}$ (0.4 mmol) was stirred in methanol (20 mL) at 80 °C for 4 h. After cooling down to room temperature, the precipitate was collected by filtration. Then, the solid was washed with MeOH and dried under reduced pressure to afford the Ni-**L1** complex as a green solid, 0.36g, 83% yield.

4. ESI-HRMS analysis of Ni-**L1** complex and the proposed structure of the complex

In the ESI-HRMS chart of Ni-**L1** complex (Figure S1), the peak corresponding to Ni/**L1** was detected. The peak of m/z = 948.23941 was assigned to [**L1** + $\text{Ni}(\text{OAc})_2$ - HOAc + H] $^+$ (Calcd. For $\text{C}_{51}\text{H}_{48}\text{N}_5\text{NiO}_6\text{S}_2$: 948.23940). On the basis of the related structure of Fe-PyBisulidine complex (*J. Am. Chem. Soc.*, **2014**, *136*, 8350-8360) and the geometry of **L1** optimized using Chem3D at MM2 level (Figure S2), we speculated that the ligand could coordinate to the Ni via pyridine nitrogen and secondary amine nitrogens. According to the ESI-HRMS analysis of the complex and the balance of total charge, one proton should be removed (Figure S3).

Acquisition Parameter

Polarity	Positive	n/a	n/a	No. of Laser Shots	200
n/a	n/a	No. of Cell Fills	1	Laser Power	20.0 lp
Broadband Low Mass	150.5 m/z	n/a	n/a	n/a	n/a
Broadband High Mass	3000.0 m/z	n/a	n/a	n/a	n/a
Acquisition Mode	Single MS	n/a	n/a	Calibration Date	Mon Jul 25 11:54:52 2016
Pulse Program	basic	n/a	n/a	Data Acquisition Size	1048576
Source Accumulation	0.010 sec	n/a	n/a	Apodization	Sine-Bell Multiplication
Ion Accumulation Time	0.020 sec	n/a	n/a		
Flight Time to Acq. Cell	0.002 sec	n/a	n/a		

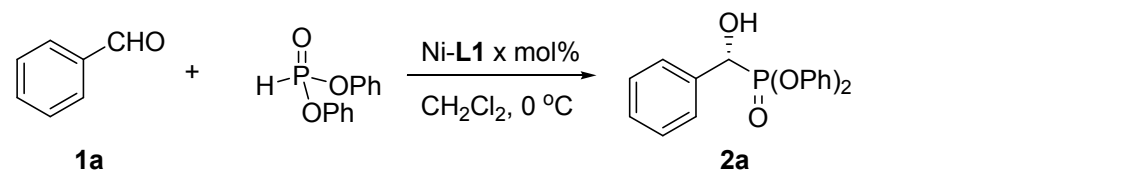
**Figure S1.** The ESI-HRMS chart of Ni-L1**Figure S2.** The geometry of L1 optimized using Chen3D (8.0) at MM2 level

Calcd. For C₅₁H₄₈N₅NiO₆S₂ [M + H]⁺: 948.23940, Found: 948.23941

Figure S3. The proposed structure of Ni-L1 complex

5. The effects of the catalyst loading and the concentration of benzaldehyde.

Table S1 The effects of the catalyst loading and the concentration of aldehyde. ^a



1a + $\text{H}-\text{P}(\text{OPh})_2$ $\xrightarrow[\text{CH}_2\text{Cl}_2, 0\text{ }^\circ\text{C}]{\text{Ni-L1 x mol\%}}$ **2a**

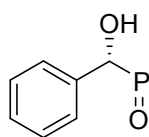
Entry	Time (h)	The loading of Ni-L1 (mol %)	The concentration of benzaldehyde (M)	Yield (%) ^h	ee (%) ⁱ
1 ^b	17	10	0.1	44	93
2 ^c	15	10	0.2	90	93
3 ^d	15	10	0.4	98	93
4 ^d	17	5	0.4	74	93
5 ^e	15	2	1	2	5
6 ^f	15	1	2	5	5
7 ^g	39	5	0.5	24	85

^a The catalyst was prepared *in situ*. ^b The reaction was carried out: aldehyde (0.1 mmol), diphenylphosphite (0.1 mmol), 1.0 mL CH₂Cl₂. ^c The reaction was carried out: aldehyde (0.2 mmol), diphenylphosphite (0.2 mmol), 1.0 mL CH₂Cl₂. ^d The reaction was carried out: aldehyde (0.2 mmol), diphenylphosphite (0.2 mmol), 0.5 mL CH₂Cl₂. ^e The reaction was carried out: aldehyde (1.0 mmol), diphenylphosphite (1.0 mmol), 1.0 mL CH₂Cl₂. ^f The reaction was carried out: aldehyde (2.0 mmol), diphenylphosphite (2.0 mmol), 1.0 mL CH₂Cl₂. ^g The reaction was carried out: aldehyde (0.5 mmol), diphenylphosphite (0.5 mmol), 1.0 mL CH₂Cl₂. ^h Isolated yield. ⁱ Enantiomeric excesses were determined by HPLC analysis.

6. General Procedure for Catalytic Asymmetric Reaction

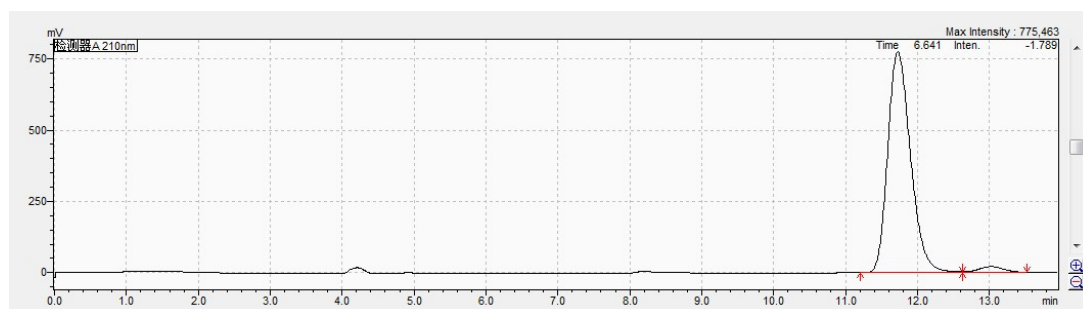
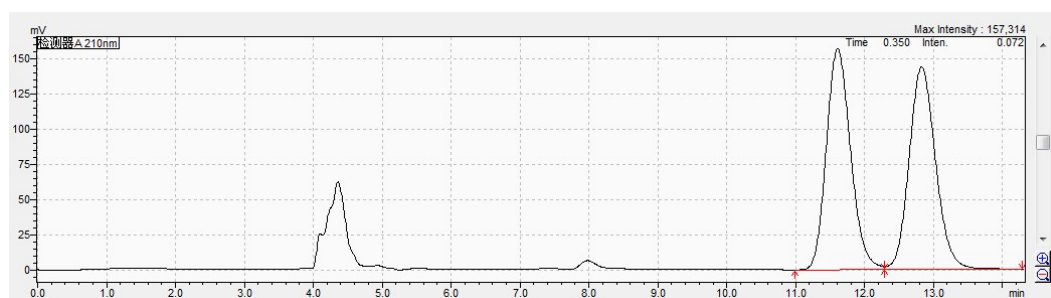
The mixture of diphenylphosphite (0.2 mmol) and the chiral complex (0.02 mmol, 10 mol%) was stirred in CH₂Cl₂ (0.5 mL) at -10 °C for 30 min followed by the addition of the aldehyde (0.2 mmol). The stirring was continued for the time indicated in Table 3 at -10 °C. The residue was purified by silica gel flash column chromatography (petroleum ether / AcOEt, 2:1) to afford the products.

7. Characterization of products

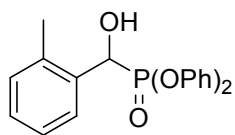


2a

White solid, 62.6 mg, 92% yield, 95% *ee*, $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.57-7.59 (m, 2H), 7.39-7.23 (m, 8H), 7.16-7.13 (m, 2H), 7.06-7.0 (m, 4H), 5.34 (d, $J = 8.5$ Hz, 1H), 3.54 (brs, 1H). $[\alpha]_{\text{D}}^{25} = +38.9$ (c 0.80, CHCl_3) [lit.² $[\alpha]_{\text{D}}^{24} = -36.5$ (c 1.0, CHCl_3) for *S* enantiomer in 91% *ee*]. HPLC (CHIRALCEL OD-H column, hexane / 2-propanol = 90 / 10, flow 0.7 mL/min, detection at 210 nm) t_{r} (major) = 11.7 min and t_{r} (minor) = 13.0 min.



Peak#	Retention Time	Area	Area%
1	11.728	17050904	97.334
2	13.022	467056	2.666
Total		17517960	100.000

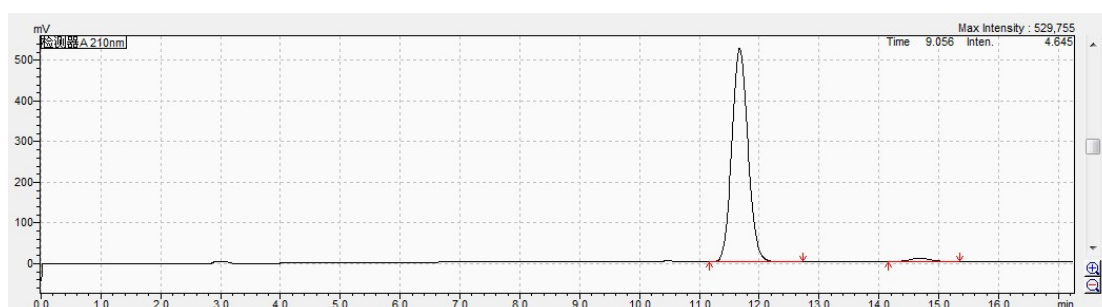
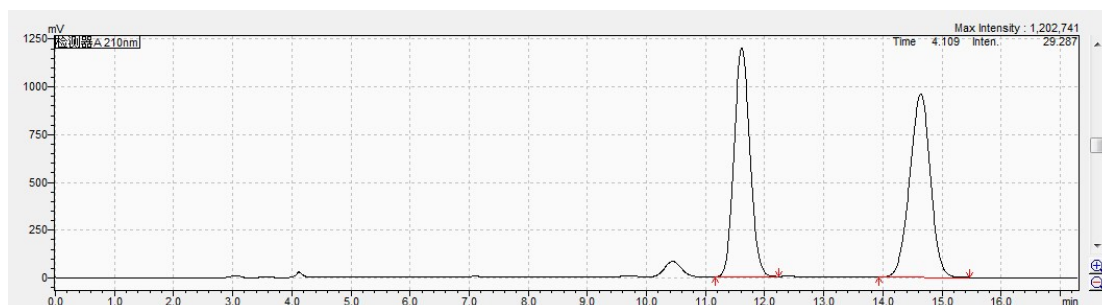


2b

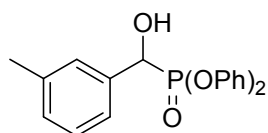
White solid, 65.8 mg, 93% yield, 96% *ee*, $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.78 (brs, 1H), 7.29-7.22 (m, 8H), 7.18-7.13 (m, 2H), 7.09-7.07 (m, 2H), 7.00-6.99 (m, 2H), 5.63 (d, $J = 9.4$ Hz, 1H), 3.77 (brs, 1H), 2.45 (s, 3H). $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 150.3 (d, $J = 10.1$ Hz), 136.2 (d, $J = 8.0$ Hz), 133.9, 130.5, 129.7 (d, $J = 5.9$ Hz), 128.6 (d, $J = 2.1$ Hz), 127.6 (d, $J = 4.2$ Hz), 126.4 (d, $J = 1.8$ Hz), 125.2 (d, $J = 9.3$ Hz), 120.6 (d, $J = 3.7$ Hz), 120.5 (d, $J = 3.6$ Hz), 67.2 (d, $J = 161.8$ Hz), 19.6. HRMS (ESI): m/z calcd for $\text{C}_{20}\text{H}_{19}\text{NaO}_4\text{P}$ $[\text{M} + \text{Na}]^+$: 377.09132, found 377.09042. $[\alpha]_{\text{D}}^{25} =$

[2] S. Hirashima, R. Arai, K. Nakashima, N. Kawai, J. Kondo, Y. Koseki, T. Miura, *Adv. Synth. Catal.*, **2015**, 357, 3863-3867.

+63.3 (c 0.41, CHCl₃). HPLC (CHIRALPAK AD-H column, hexane / 2-propanol = 80/20, flow 1 mL/min, detection at 210 nm) t_r (major) = 11.7 min and t_r (minor) = 14.7 min.

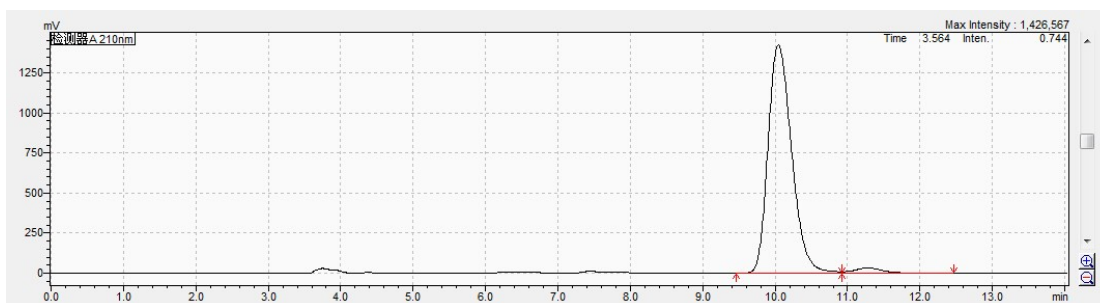
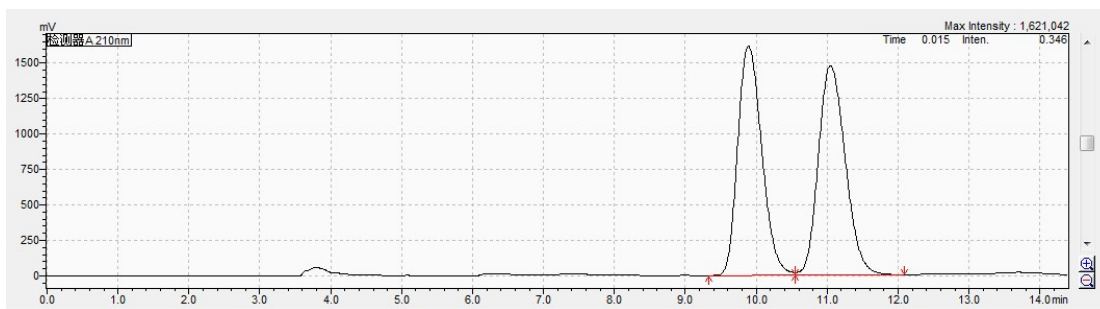


Peak#	Retention Time	Area	Area%
1	11.673	9810696	98.024
2	14.681	197744	1.976
Total		10008440	100.000

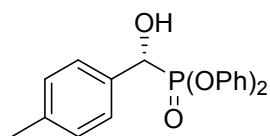


2c

White solid, 71mg, 99% yield, 95% ee, ¹H NMR (500 MHz, CDCl₃) δ 7.30-7.24 (m, 2H), 7.20-7.11 (m, 5H), 7.08-7.00 (m, 3H), 7.00-6.95 (m, 2H), 6.94-6.89 (m, 2H), 5.14 (d, *J* = 9.2 Hz, 1H), 4.64 (brs, 1H), 2.24 (s, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 149.3 (d, *J* = 5.6 Hz), 149.2 (d, *J* = 6.2 Hz), 137.0 (d, *J* = 1.7 Hz), 134.4, 128.6 (d, *J* = 10.0 Hz), 128.2 (d, *J* = 2.8 Hz), 127.3 (d, *J* = 0.9 Hz), 127.1 (d, *J* = 6.2 Hz), 124.1 (d, *J* = 11.3 Hz), 123.6 (d, *J* = 6.2 Hz), 119.6 (d, *J* = 4.0 Hz), 119.5 (d, *J* = 4.0 Hz), 69.6 (d, *J* = 160.2 Hz), 20.4. HRMS (ESI): *m/z* calcd for C₂₀H₁₉NaO₄P [M + Na]⁺: 377.09132, found 377.09030. [α]_D²⁵ = +26.5 (c 0.32, CHCl₃). HPLC (CHIRALCEL OD-H column, hexane / 2-propanol = 90/10, flow 0.8 mL/min, detection at 210 nm) t_r (major) = 10.0 min and t_r (minor) = 11.2 min.



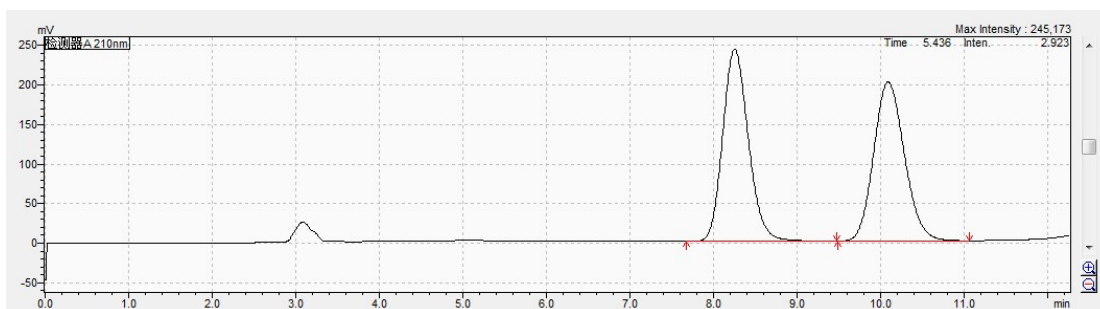
Peak#	Retention Time	Area	Area%
1	10.044	32196171	97.464
2	11.280	837720	2.536
Total		33033891	100.000

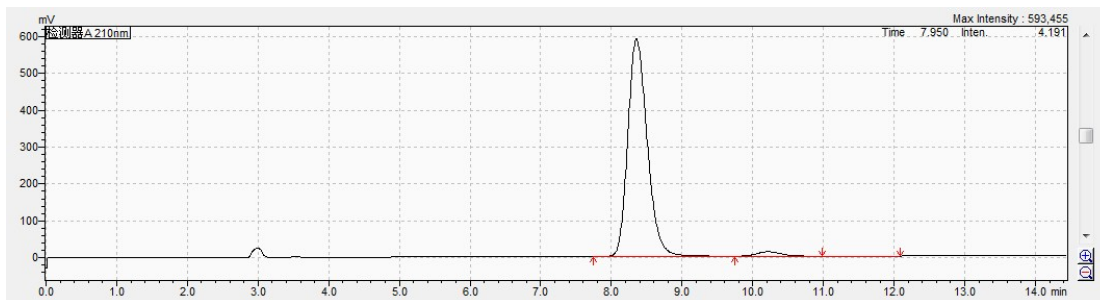


2d

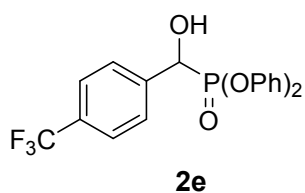
White solid, 71 mg, 99% yield, 95% ee, $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.55-7.45 (m, 2H), 7.35-7.24 (m, 4H), 7.24-7.13 (m, 4H), 7.12-7.09 (m, 2H), 7.07-7.01 (m, 2H), 5.28 (d, $J = 8.0$ Hz, 1H), 4.16 (brs, 1H), 2.39 (s, 3H). $[\alpha]_{\text{D}}^{25} = +40.9$ (c 0.57, CHCl_3) [lit.² $[\alpha]_{\text{D}}^{25} = -38.0$ (c 1.0, CHCl_3)

for *S* enantiomer in 91% ee]. HPLC (CHIRALCEL OD-H column, hexane / 2-propanol = 90/10, flow 1 mL/min, detection at 210nm) t_{r} (major) = 8.4 min and t_{r} (minor) = 10.2 min.

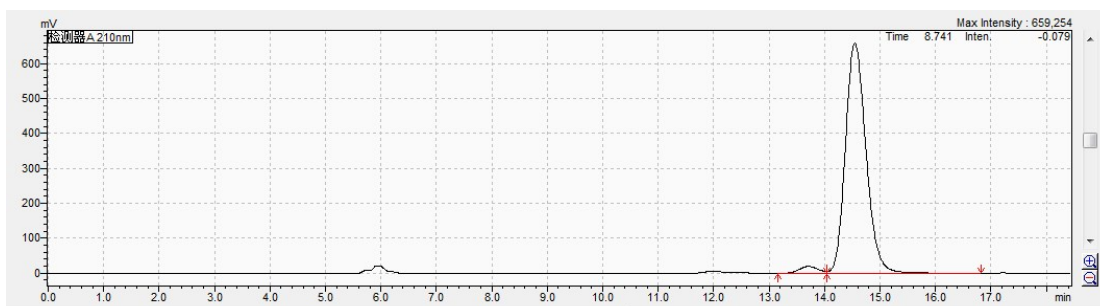




Peak#	Retention Time	Area	Area%
1	8.355	11355306	97.590
2	10.218	280468	2.410
Total		11635774	100.000

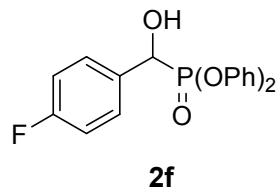


White solid, 77.5 mg, 95% yield, 95% ee, ^1H NMR (500 MHz, CDCl_3) δ 7.69-7.55 (m, 4H), 7.32-7.22 (m, 4H), 7.20-7.12 (m, 2H), 7.10-7.05 (m, 2H), 7.03-6.95 (m, 2H), 5.27 (d, $J = 10.4$ Hz, 1H), 5.21 (brs, 1H). ^{13}C NMR (125 MHz, CDCl_3) δ 150.2 (d, $J = 6.2$ Hz), 150.1 (d, $J = 6.6$ Hz), 139.8, 129.8 (d, $J = 10.1$ Hz), 127.7 (d, $J = 5.9$ Hz), 125.6, 125.5, 125.3 (qui, $J = 3.3$ Hz), 120.7 (d, $J = 4.1$ Hz), 120.5 (d, $J = 4.1$ Hz), 70.1 (d, $J = 159.6$ Hz). HRMS (ESI): m/z calcd for $\text{C}_{20}\text{H}_{16}\text{F}_3\text{NaO}_4\text{P}$ [$\text{M} + \text{Na}$] $^+$: 431.06305, found 431.06167. $[\alpha]_{\text{D}}^{25} = +32.6$ (c 0.41, CHCl_3). HPLC (CHIRALCEL OD-H column, hexane / 2-propanol = 90/10, flow 0.5 mL/min, detection at 210 nm) t_{r} (minor) = 13.7 min and t_{r} (major) = 14.5 min.

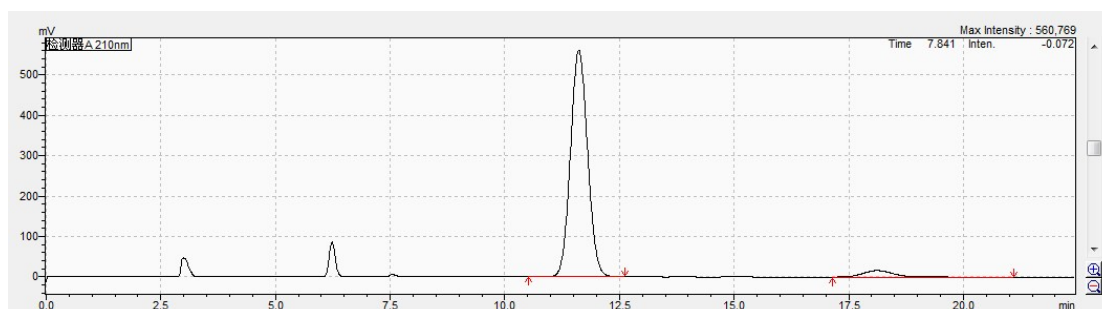
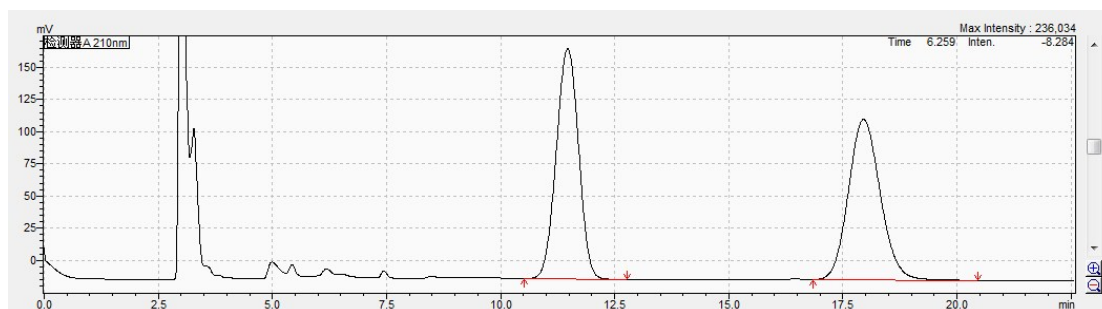


Peak#	Retention Time	Area	Area%
-------	----------------	------	-------

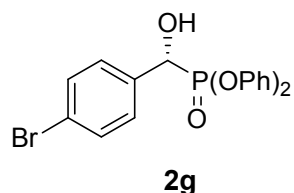
1	13.710	453161	2.601
2	14.547	16971678	97.399
Total		17424839	100.000



White solid, 66.6 mg, 93% yield, 90% ee, ^1H NMR (500 MHz, CDCl_3) δ 7.56-7.43 (m, 2H), 7.30-7.20 (m, 4H), 7.19-7.11 (m, 2H), 7.11-7.05 (m, 2H), 7.05-6.96 (m, 4H), 5.19 (d, $J = 9.0$ Hz, 1H), 5.02 (brs, 1H). ^{13}C NMR (125 MHz, CDCl_3) δ 161.5 (d, $J = 3.5$ Hz), 159.5 (d, $J = 3.7$ Hz), 148.0 (d, $J = 1.9$ Hz), 147.9 (d, $J = 2.3$ Hz), 129.2 (m), 127.4, (d, $J = 10.0$ Hz), 127.1 (d, $J = 6.4$ Hz), 127.0 (d, $J = 6.4$ Hz), 123.1 (d, $J = 12.8$ Hz), 118.4 (d, $J = 4.0$ Hz), 118.3 (d, $J = 4.0$ Hz), 113.2 (d, $J = 2.4$ Hz), 113.0 (d, $J = 2.4$ Hz), 67.7 (d, $J = 161.3$ Hz). HRMS (ESI): m/z calcd for $\text{C}_{19}\text{H}_{16}\text{FNaO}_4\text{P} [\text{M} + \text{Na}]^+$: 381.06624, found 381.06607. $[\alpha]_{\text{D}}^{25} = +30.3$ (c 0.35, CHCl_3). HPLC (CHIRALPAK AS-H column, hexane / 2-propanol = 80/20, flow 1 mL/min, detection at 210 nm) t_{r} (major) = 11.6 min and t_{r} (minor) = 18.1 min.

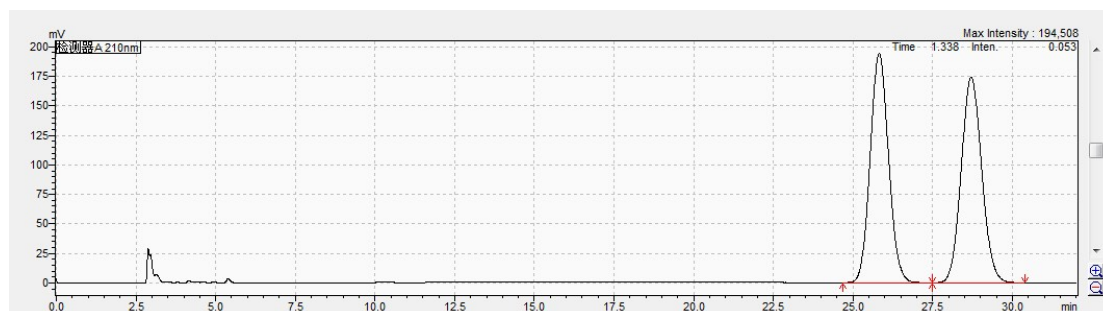


Peak#	Retention Time	Area	Area%
1	11.613	14589556	95.025
2	18.118	763829	4.975
Total		15353385	100.000

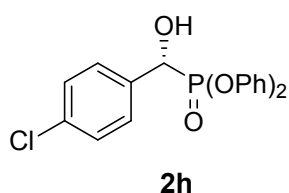


White solid, 78.6 mg, 94% yield, 95% ee, ^1H NMR (500 MHz, CDCl_3) δ 7.50-7.43 (m, 2H), 7.42-7.36 (m, 2H), 7.30-7.22 (m, 4H), 7.20-7.11 (m, 2H), 7.09-7.04 (m, 2H), 7.02-6.96 (m, 2H), 5.15 (d, $J =$

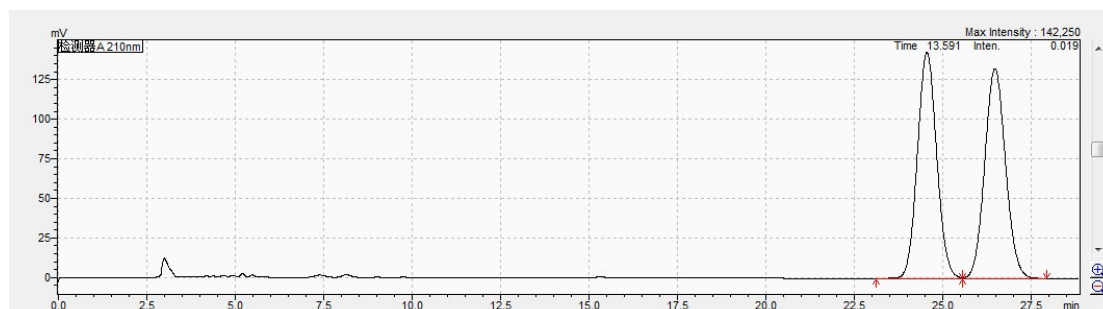
9.5 Hz, 1H), 5.08 (brs, 1H). $[\alpha]_D^{25} = +35.9$ (c 0.46, CHCl₃) [lit.² $[\alpha]_D^{24} = -45.8$ (c 1.0, CHCl₃) for *S* enantiomer in 97% *ee*]. HPLC (CHIRALPAK AD-H column, hexane / 2-propanol = 90/10, flow 1 mL/min, detection at 210nm) t_r (minor) = 25.9 min and t_r (major) = 28.9 min.

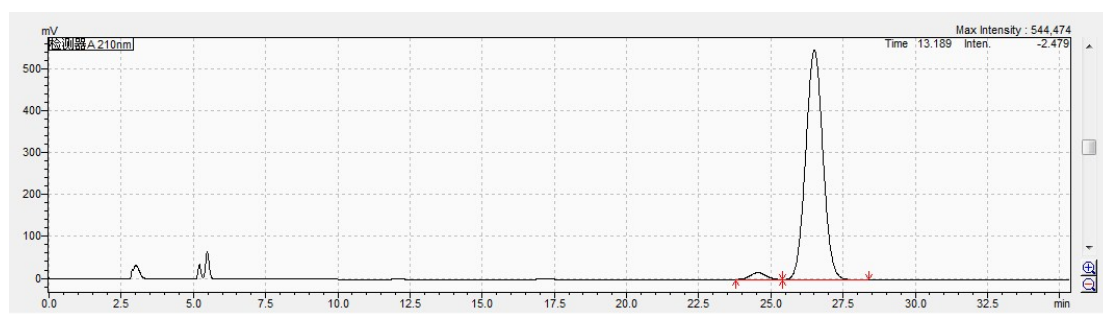


Peak#	Retention Time	Area	Area%
1	25.881	1751599	2.643
2	28.895	64533802	97.357
Total		66285401	100.000

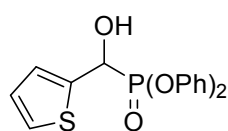


White solid, 73.3 mg, 98% yield, 95% *ee*, ¹H NMR (500 MHz, CDCl₃) δ 7.55-7.43 (m, 2H), 7.41-7.24 (m, 7H), 7.24-7.15 (m, 2H), 7.13-7.00 (m, 4H), 5.34-5.26 (m, 1H), 5.25-5.19 (m, 1H). $[\alpha]_D^{25} = +39.8$ (c 0.42, CHCl₃) [lit.² $[\alpha]_D^{20} = -48.3$ (c 1.0, CHCl₃) for *S* enantiomer in 93% *ee*]. HPLC (CHIRALPAK AD-H column, hexane / 2-propanol = 90/10, flow 1 mL/min, detection at 210 nm) t_r (minor) = 24.5 min and t_r (major) = 26.5 min.



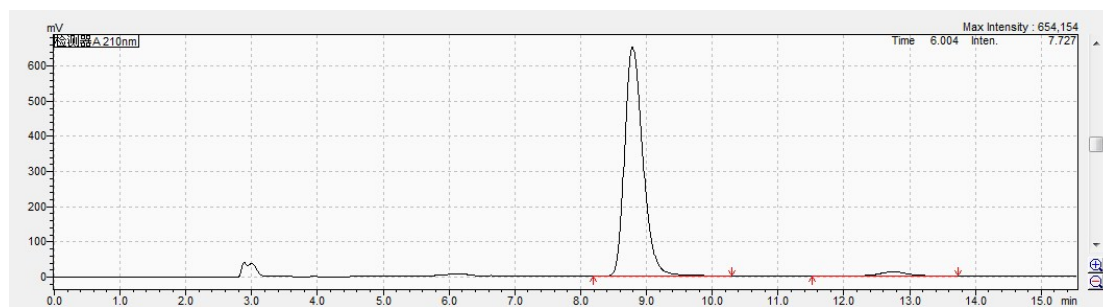
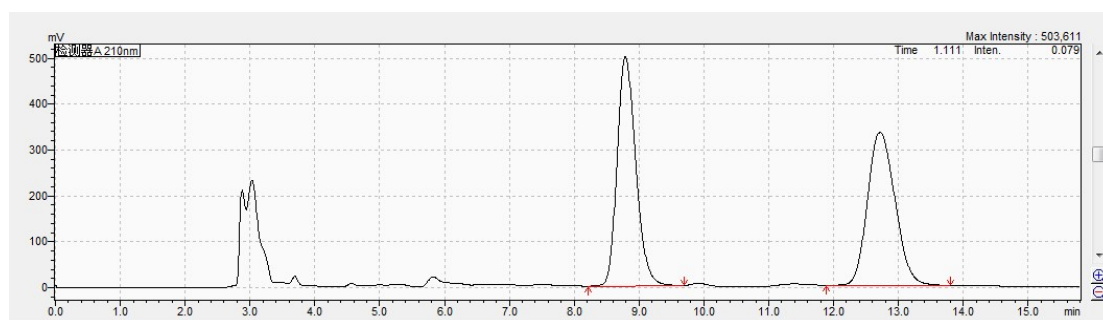


Peak#	Retention Time	Area	Area%
1	24.549	606217	2.602
2	26.508	22696022	97.398
Total		23302239	100.000



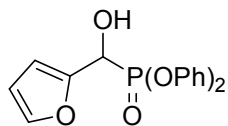
2i

White solid, 69.2 mg, 99% yield, 95% ee, ^1H NMR (500 MHz, CDCl_3) δ 7.36-7.34 (m, 1H), 7.33-7.23 (m, 7H), 7.19-7.15 (m, 2H), 7.15-7.11 (m, 2H), 7.09-7.04 (m, 2H), 7.03-7.00 (m, 1H), 5.51 (d, $J = 9.7$ Hz, 1H), 3.95 (brs, 1H). ^{13}C NMR (125 MHz, CDCl_3) δ 150.8 (d, $J = 11.6$ Hz), 138.4, 130.3 (d, $J = 7.6$ Hz), 127.7, 127.1 (d, $J = 3.2$ Hz), 126.0 (d, $J = 10.6$ Hz), 121.2 (d, $J = 3.9$ Hz), 121.1 (d, $J = 3.7$ Hz), 116.5, 67.3 (d, $J = 169.0$ Hz). HRMS (ESI): m/z calcd for $\text{C}_{17}\text{H}_{15}\text{NaO}_4\text{PS}$ [$\text{M} + \text{Na}$] $^+$: 369.03209, found 369.03203. $[\alpha]_{\text{D}}^{25} = +19.9$ (c 0.41, CHCl_3). HPLC (CHIRALCEL OD-H column, hexane / 2-propanol = 90/10, flow 1 mL/min, detection at 210 nm) t_{r} (major) = 8.8 min and t_{r} (minor) = 12.8 min.



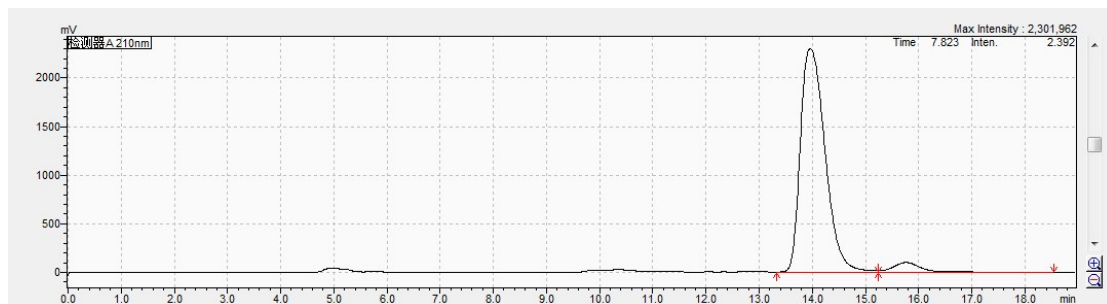
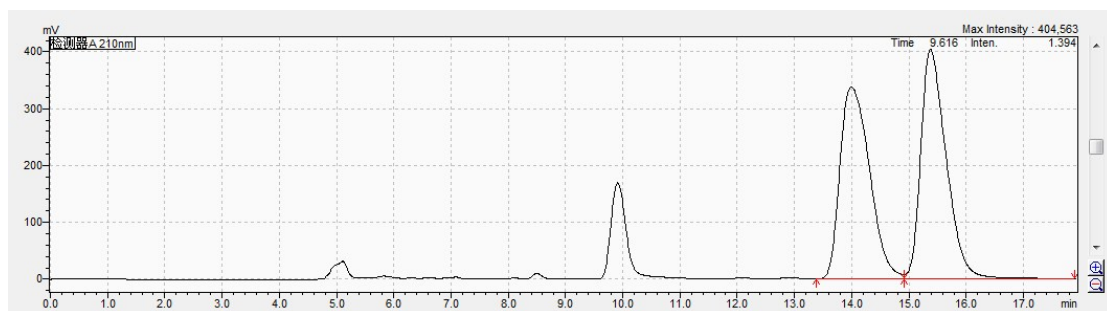
Peak#	Retention Time	Area	Area%
-------	----------------	------	-------

1	8.789	12548503	97.560
2	12.753	313840	2.440
Total		12862343	100.000

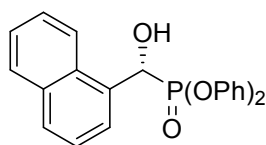


2j

White solid, 64.7 mg, 91% ee, 98% yield, ^1H NMR (500 MHz, CDCl_3) δ 7.46 (s, 1H), 7.38-7.26 (m, 4H), 7.25-7.13 (m, 4H), 7.13-7.05 (m, 2H), 6.65-6.55 (s, 1H), 6.46-6.34 (s, 1H), 5.33 (d, $J = 12.3$ Hz, 1H), 4.16 (brs, 1H). ^{13}C NMR (125 MHz, CDCl_3) δ 150.2 (d, $J = 9.3$ Hz), 148.7, 143.3, 129.7 (d, $J = 10.5$ Hz), 125.4 (d, $J = 14.3$ Hz), 120.7 (d, $J = 3.6$ Hz), 120.6 (d, $J = 3.7$ Hz), 110.9, 110.4, (d, $J = 6.7$ Hz), 64.5 (d, $J = 168.9$ Hz). HRMS (ESI): m/z calcd for $\text{C}_{17}\text{H}_{15}\text{NaO}_5\text{P}$ [$\text{M} + \text{Na}$] $^+$: 353.05493, found 353.05419. $[\alpha]_{\text{D}}^{25} = +13.8$ (c 0.33, CHCl_3). HPLC (CHIRALCEL OD-H column, hexane / 2-propanol = 90/10, flow 0.6 mL/min, detection at 210nm) t_{r} (major) = 14.0 min and t_{r} (minor) = 15.8 min.



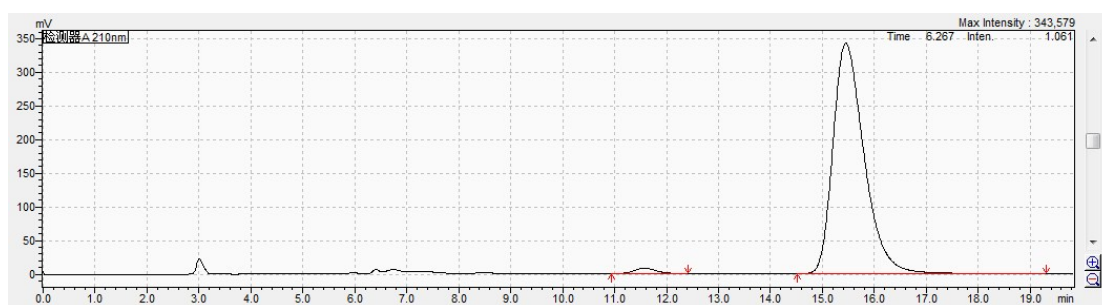
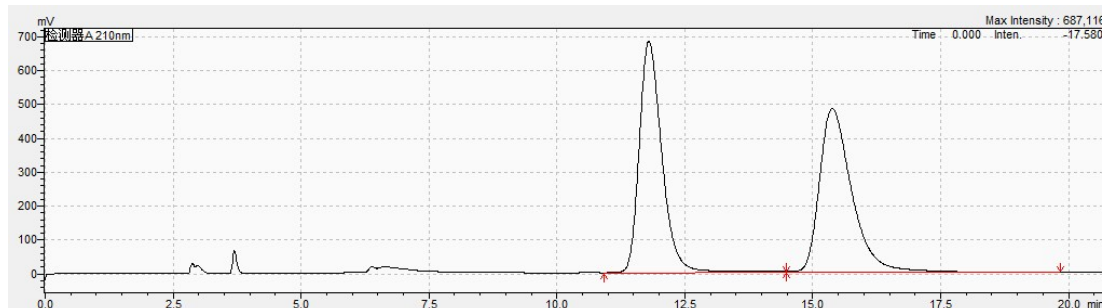
Peak#	Retention Time	Area	Area%
1	13.963	74005617	95.608
2	15.762	3399664	4.392
Total		77405281	100.000



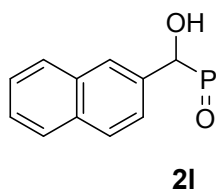
2k

White solid, 70.2 mg, 90% yield, 97% ee, ^1H NMR (500 MHz, CDCl_3) δ 8.18 (d, $J = 7.7$, 1H), 8.00 (brs, 1H), 7.96-7.84 (m, 2H), 7.62-7.47 (m, 3H), 7.33-7.24 (m, 3H), 7.22-7.13 (m, 3H), 7.11-7.04 (m, 3H), 6.92 (d, J

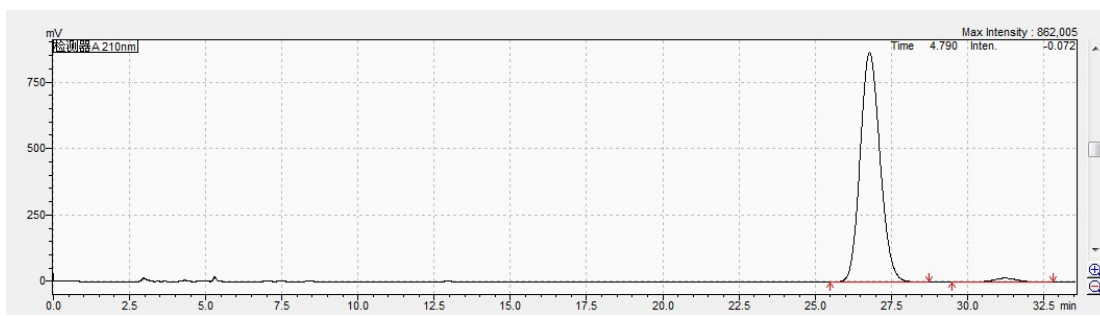
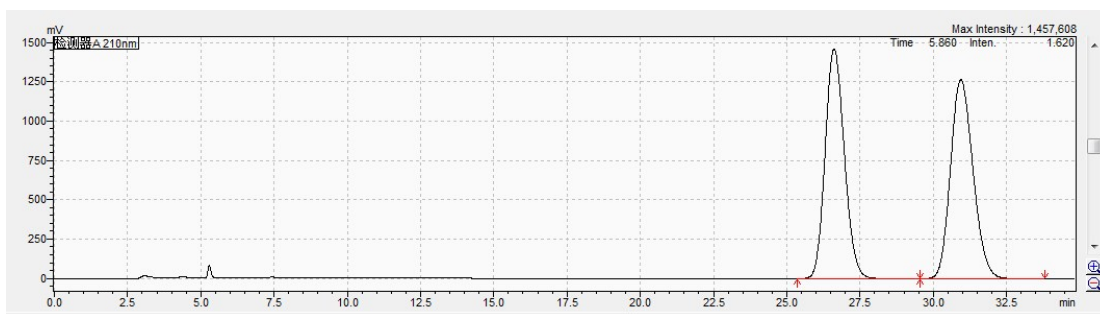
= 7.0 Hz, 2H), 6.22 (d, $J = 9.7$ Hz, 1H), 3.95 (brs, 1H). $[\alpha]_{\text{D}}^{25} = +98.9$ (c 0.39, CHCl_3) [lit.² $[\alpha]_{\text{D}}^{22} = -131.2$ (c 1.0, CHCl_3) for *S* enantiomer in 93% *ee*]. HPLC (CHIRALCEL OD-H column, hexane / 2-propanol = 90/10, flow 1 mL/min, detection at 210 nm) t_{r} (minor) = 11.6 min and t_{r} (major) = 15.5 min.



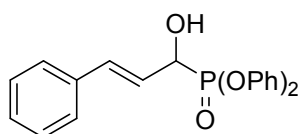
Peak#	Retention Time	Area	Area%
1	11.575	227560	1.572
2	15.452	14247005	98.428
Total		14474565	100.000



White solid, 78 mg, 99% yield, 96% *ee*, ^1H NMR (500 MHz, CDCl_3) δ 8.03 (s, 1H), 7.90-7.78 (m, 3H), 7.73-7.65 (m, 1H), 7.55-7.44 (m, 2H), 7.30-7.17 (m, 5H), 7.17-6.98 (m, 6H), 5.49 (d, $J = 8.7$ Hz, 1H), 3.89 (brs, 1H). ^{13}C NMR (125 MHz, CDCl_3) δ 150.3 (d, $J = 7.7$ Hz), 133.4, 133.1, 132.8, 129.7 (d, $J = 8.6$ Hz), 128.3 (d, $J = 8.1$ Hz), 127.7, 126.8 (d, $J = 8.2$ Hz), 126.4 (d, $J = 13.2$ Hz), 125.3 (d, $J = 10.8$ Hz), 124.9 (d, $J = 4.2$ Hz), 120.6 (d, $J = 3.9$ Hz), 120.5 (d, $J = 3.7$ Hz), 70.9 (d, $J = 160.0$ Hz). HRMS (ESI): m/z calcd for $\text{C}_{23}\text{H}_{19}\text{NaO}_4\text{P}$ $[\text{M} + \text{Na}]^+$: 413.09132, found 413.09020. $[\alpha]_{\text{D}}^{25} = +45.2$ (c 0.38, CHCl_3). HPLC (CHIRALPAK AD-H column, hexane / 2-propanol = 80/20, flow 1mL/min, detection at 210 nm) t_{r} (major) = 26.8 min and t_{r} (minor) = 31.2 min.

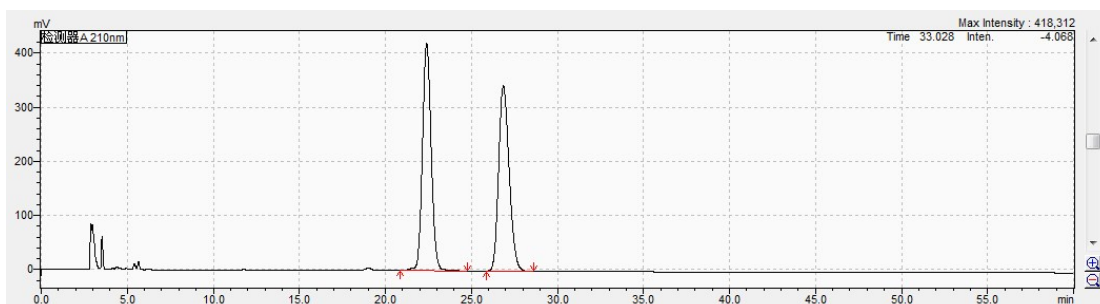


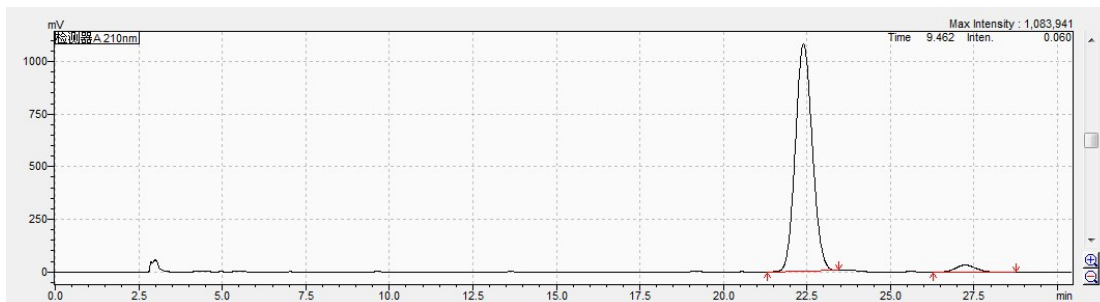
Peak#	Retention Time	Area	Area%
1	26.789	39011588	98.125
2	31.235	745444	1.875
Total		39757032	100.000



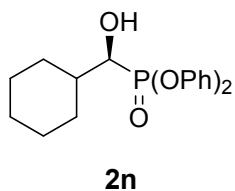
2m

White solid, 66.6 mg, 91% yield, 93% ee, ^1H NMR (500 MHz, CDCl_3) δ 7.36-7.22 (m, 11H), 7.22-7.11 (m, 7H), 6.88-6.79 (m, 1H), 6.45-6.35 (m, 1H), 4.97-4.86 (m, 1H), 4.54 (t, $J = 6.2$ Hz, 1H). ^{13}C NMR (125 MHz, CDCl_3) δ 150.4 (d, $J = 5.0$ Hz), 136.2 (d, $J = 2.9$ Hz), 133.9 (d, $J = 14.1$ Hz), 129.8 (d, $J = 5.7$ Hz), 128.7, 128.2, 126.9 (d, $J = 1.8$ Hz), 125.4 (d, $J = 10.9$ Hz), 120.9 (d, $J = 4.1$ Hz), 120.8 (d, $J = 4.1$ Hz), 69.5 (d, $J = 161.5$ Hz). HRMS (ESI): m/z calcd for $\text{C}_{21}\text{H}_{19}\text{NaO}_4\text{P}$ [$\text{M} + \text{Na}$] $^+$: 389.09132, found 389.09106. $[\alpha]_{\text{D}}^{25} = +29.3$ (c 0.35, CHCl_3). HPLC (CHIRALPAK AD-H column, hexane / 2-propanol = 90/10, flow 1 mL/min, detection at 210 nm) t_r (major) = 22.4 min and t_r (minor) = 27.2 min.

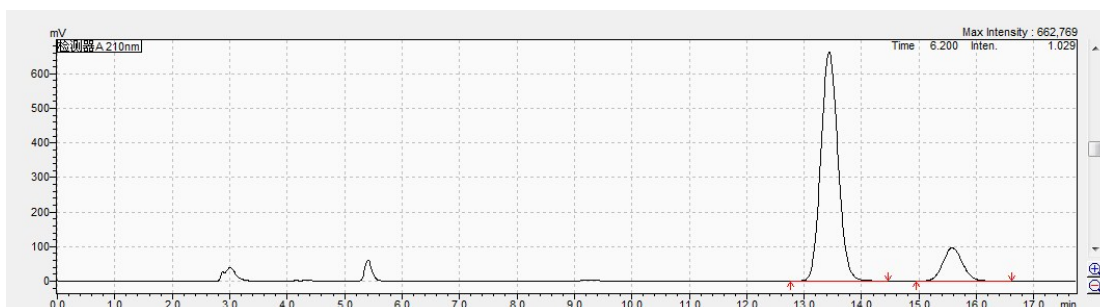
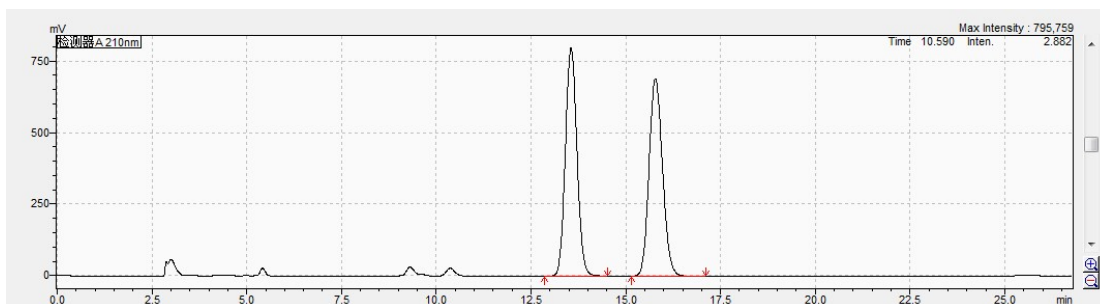




Peak#	Retention Time	Area	Area%
1	22.399	38213753	96.427
2	27.233	1415901	3.573
Total		39629654	100.000

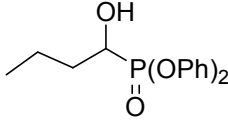


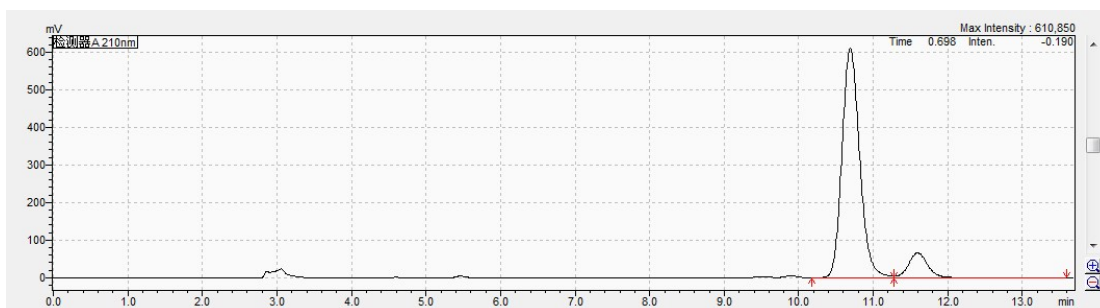
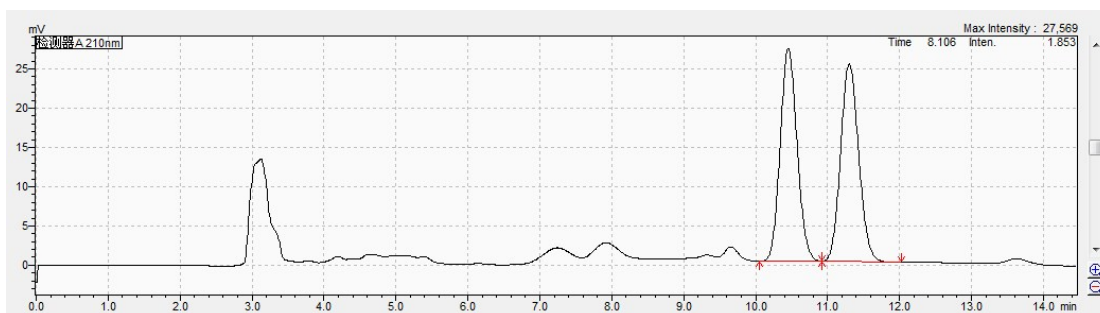
White solid, 69.2 mg, 99% yield, 72% ee, $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.34-7.26 (m, 4H), 7.22-7.11 (m, 6H), 3.95 (t, $J = 5.6$, 1H), 3.49 (brs, 1H), 2.17-2.05 (m, 1H), 2.02-1.93 (m, 1H), 1.90-1.82 (m, 1H), 1.82-1.72 (m, 2H), 1.71-1.60 (m, 1H), 1.43-1.11 (m, 6H). $[\alpha]_{\text{D}}^{25} = +1.4$ (c 0.42, CHCl_3) [lit.² $[\alpha]_{\text{D}}^{21} = +0.7$ (c 1.0, CHCl_3) for *S* enantiomer in 66% ee]. HPLC (CHIRALPAK AD-H column, hexane / 2-propanol = 90/10, flow 1 mL/min, detection at 210 nm) t_{r} (major) = 13.4 min and t_{r} (minor) = 15.6 min.



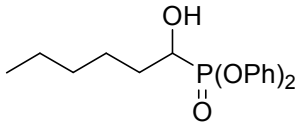
Peak#	Retention Time	Area	Area%
-------	----------------	------	-------

1	13.435	13703285	85.891
2	15.578	2250921	14.109
Total		15954206	100.000

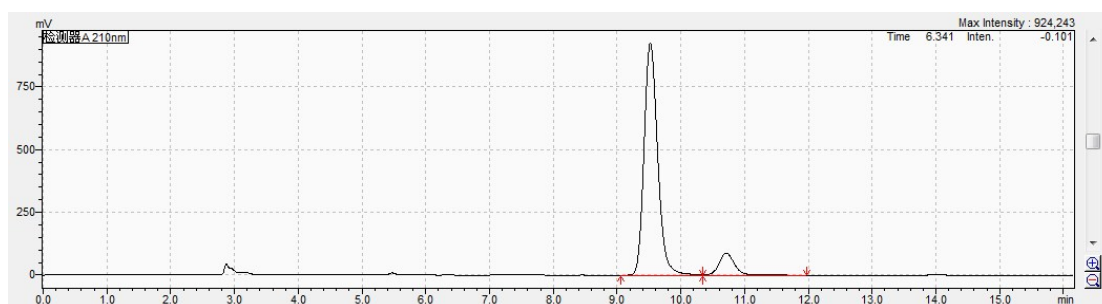
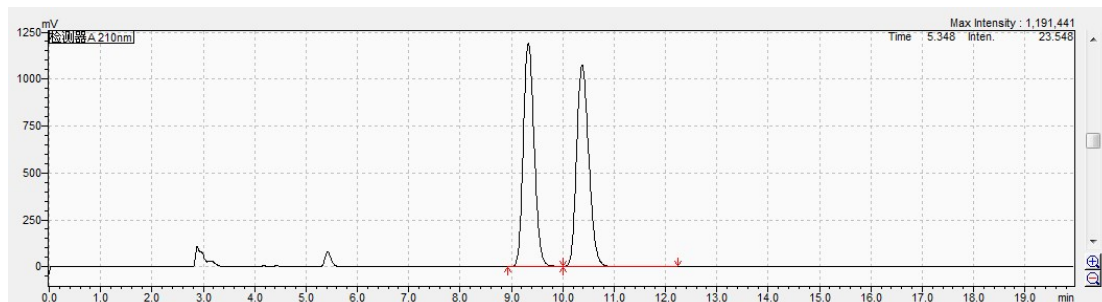

 Colorless oil, 53.2 mg, 87% yield, 79% ee, ¹H NMR (500 MHz, CDCl₃) δ 7.38-7.28 (m, 4H), 7.23-7.12 (m, 6H), 4.24-4.17 (m, 1H), 2.87 (brs, 1H), 2.02-1.84 (m, 2H), 1.57-1.35 (m, 2H), 0.98 (t, *J* = 7.4 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 150.3 (d, *J* = 9.9 Hz), 129.8, 125.3 (d, *J* = 3.8 Hz), 120.7 (t, *J* = 4.1 Hz), 67.6 (d, *J* = 158.9 Hz), 33.3, 18.9 (d, *J* = 14.2 Hz), 13.7. HRMS (ESI): *m/z* calcd for C₁₆H₁₉NaO₄P [M + Na]⁺: 329.09132, found 329.09133. [α]_D²⁵ = -7.0 (c 1.48, CHCl₃). HPLC (CHIRALPAK AD-H column, hexane / 2-propanol = 90/10, flow 1 mL/min, detection at 210nm) *t*_r (major) = 10.692 min and *t*_r (minor) = 11.594 min.



Peak#	Retention Time	Area	Area%
1	10.692	10041464	89.385
2	11.594	1192463	10.615
Total		11233927	100.000


 White solid, 66.8 mg, 99% yield, 81% ee, ¹H NMR (500 MHz, CDCl₃) δ 7.33-7.26 (m, 4H), 7.21-7.12 (m, 6H), 4.16-4.10 (m, 1H), 3.88 (brs, 1H), 1.94-1.87 (m, 2H), 1.42-1.26 (m, 6H), 0.88 (t, *J* = 6.9 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 150.4 (d, *J* = 10.6 Hz),

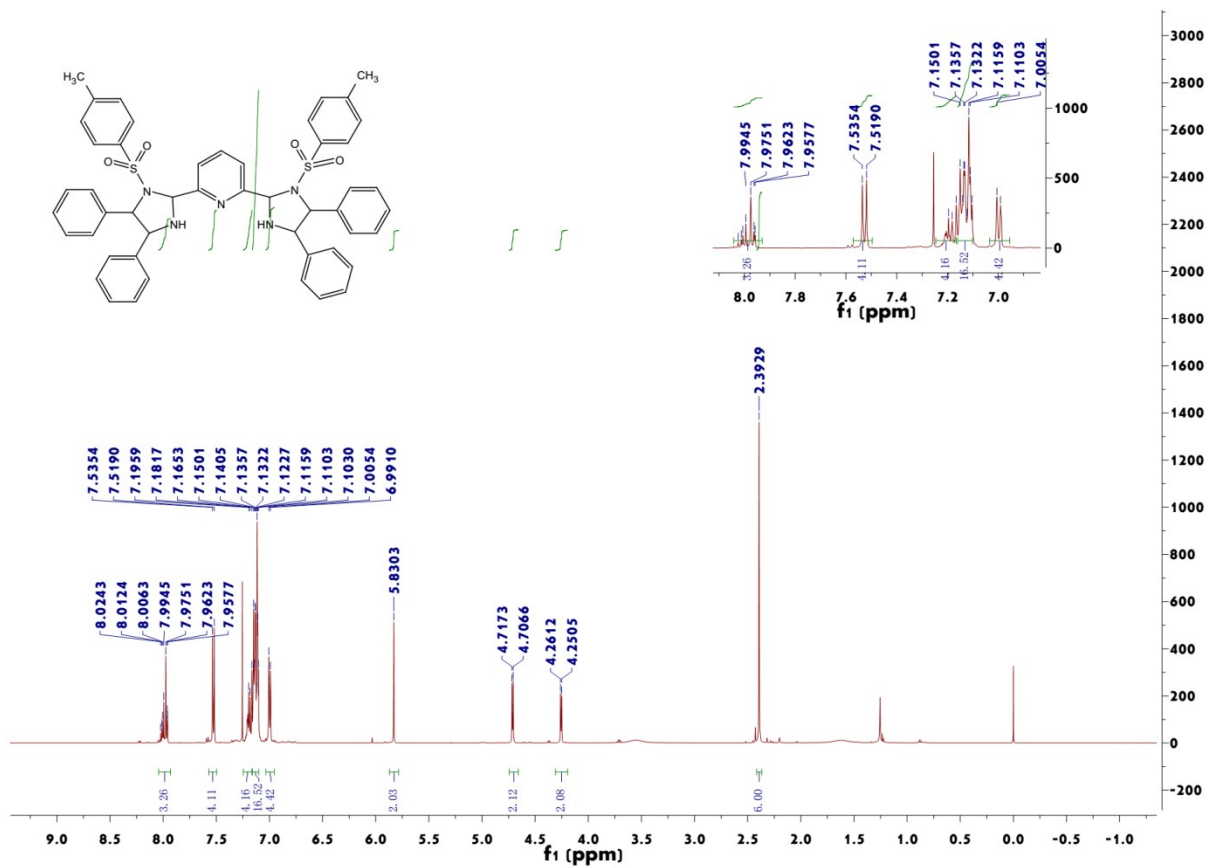
129.8, 125.2 (d, $J = 5.8$ Hz), 120.8 (d, $J = 4.0$ Hz), 120.7 (d, $J = 4.1$ Hz), 67.8 (d, $J = 158.7$ Hz), 31.5, 31.3 (d, $J = 0.8$ Hz), 25.4 (d, $J = 13.9$ Hz), 22.5, 14.1. HRMS (ESI): m/z calcd for $C_{18}H_{23}NaO_4P [M + Na]^+$: 357.12262, found 357.12250. $[\alpha]_D^{25} = -10.1$ (c 0.40, $CHCl_3$). HPLC (CHIRALPAK AD-H column, hexane / 2-propanol = 90/10, flow 1 mL/min, detection at 210nm) t_r (major) = 9.5 min and t_r (minor) = 10.7 min.



Peak#	Retention Time	Area	Area%
1	9.523	13857672	90.325
2	10.711	1484417	9.675
Total		15342089	100.000

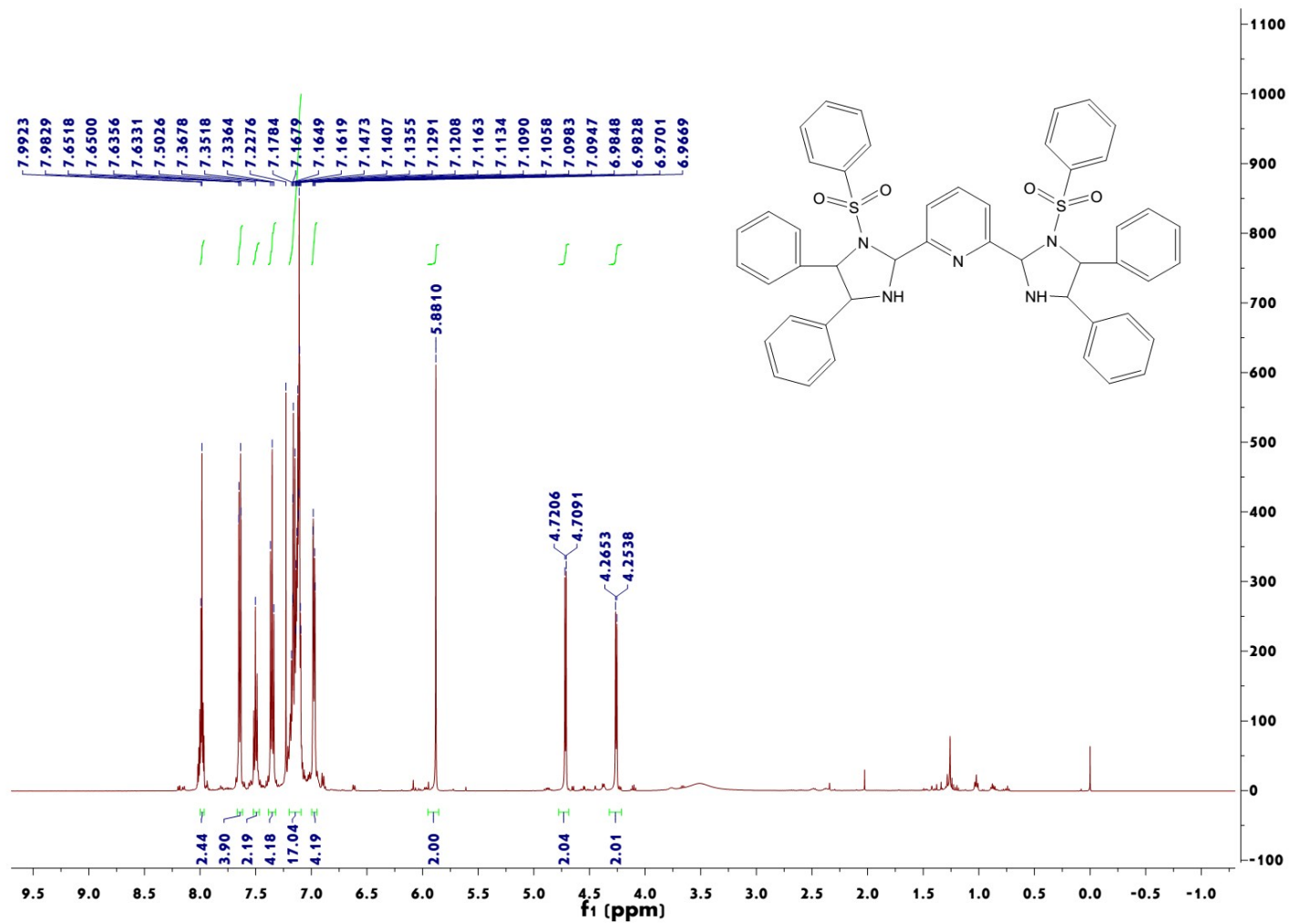
8. Copy of NMR spectra.

L1



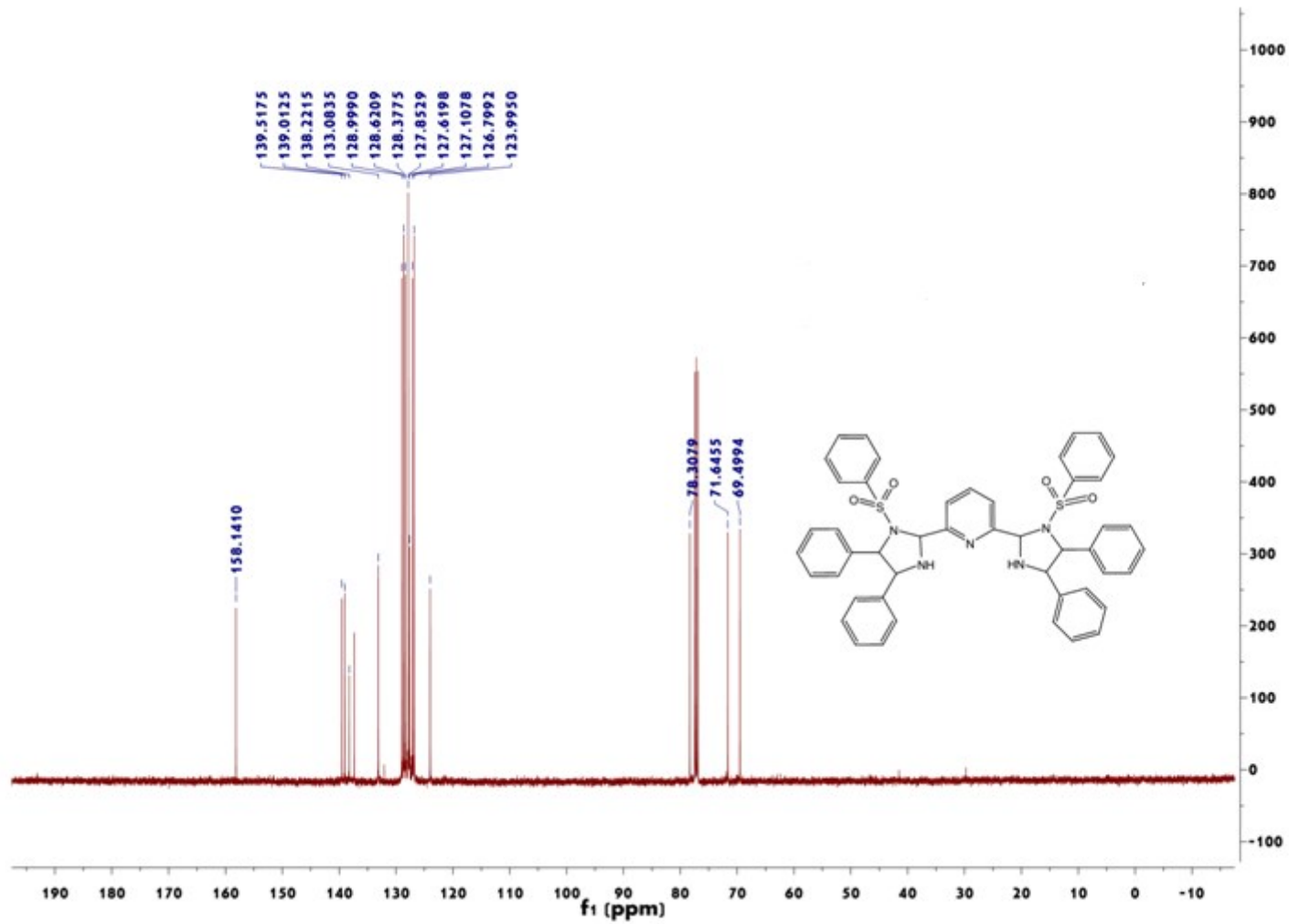
S19

L2



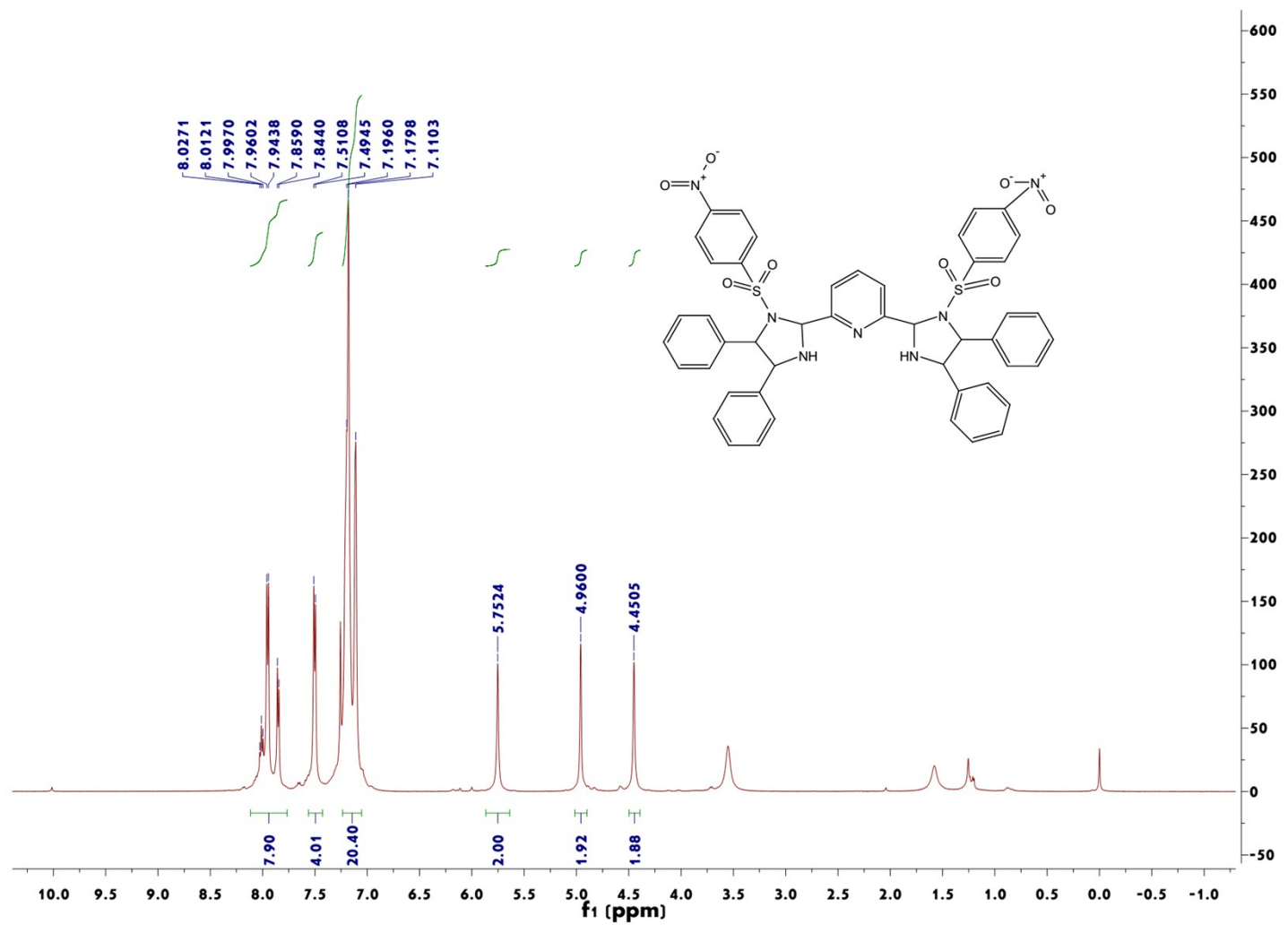
S20

L2



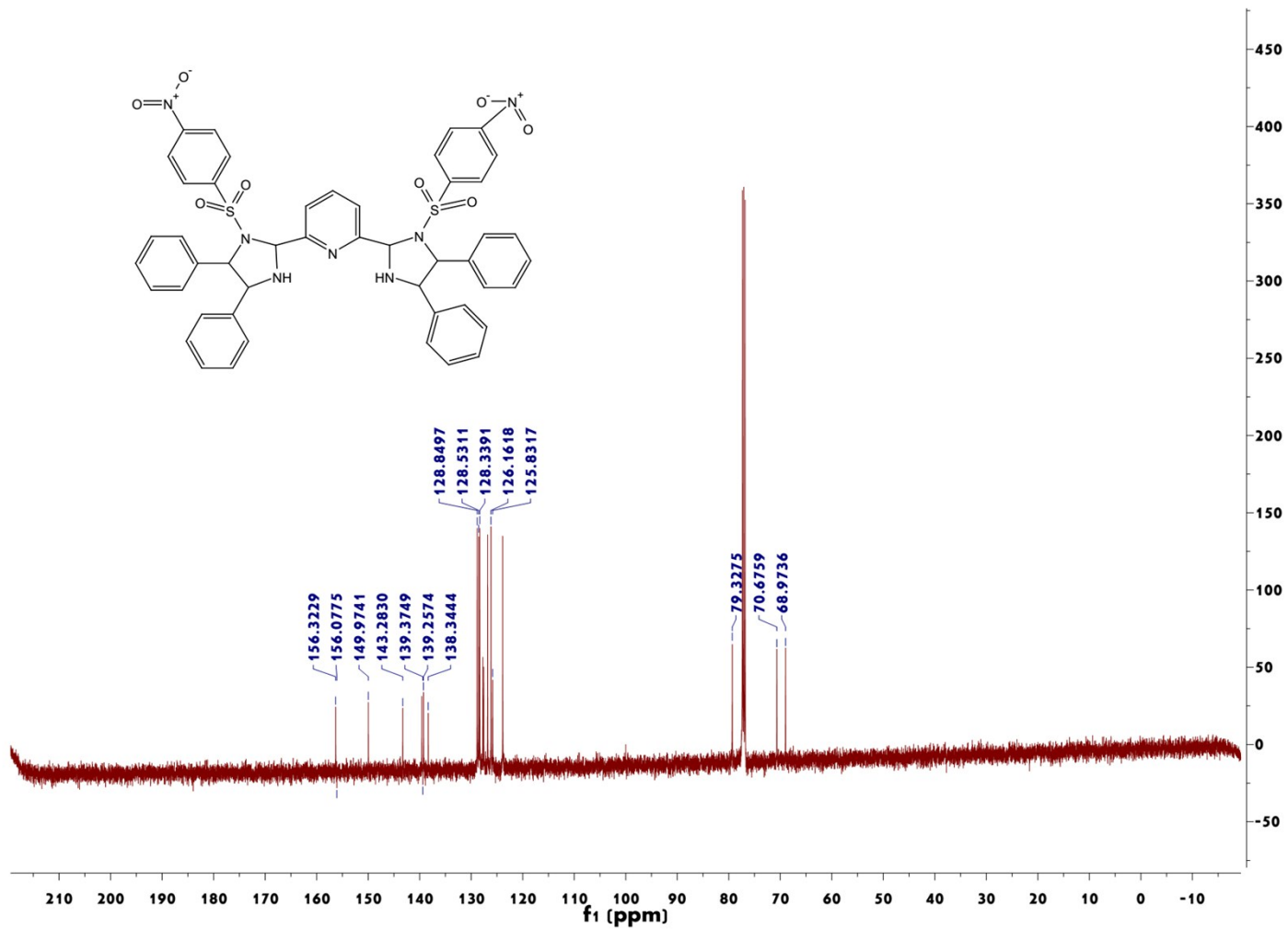
S21

L3



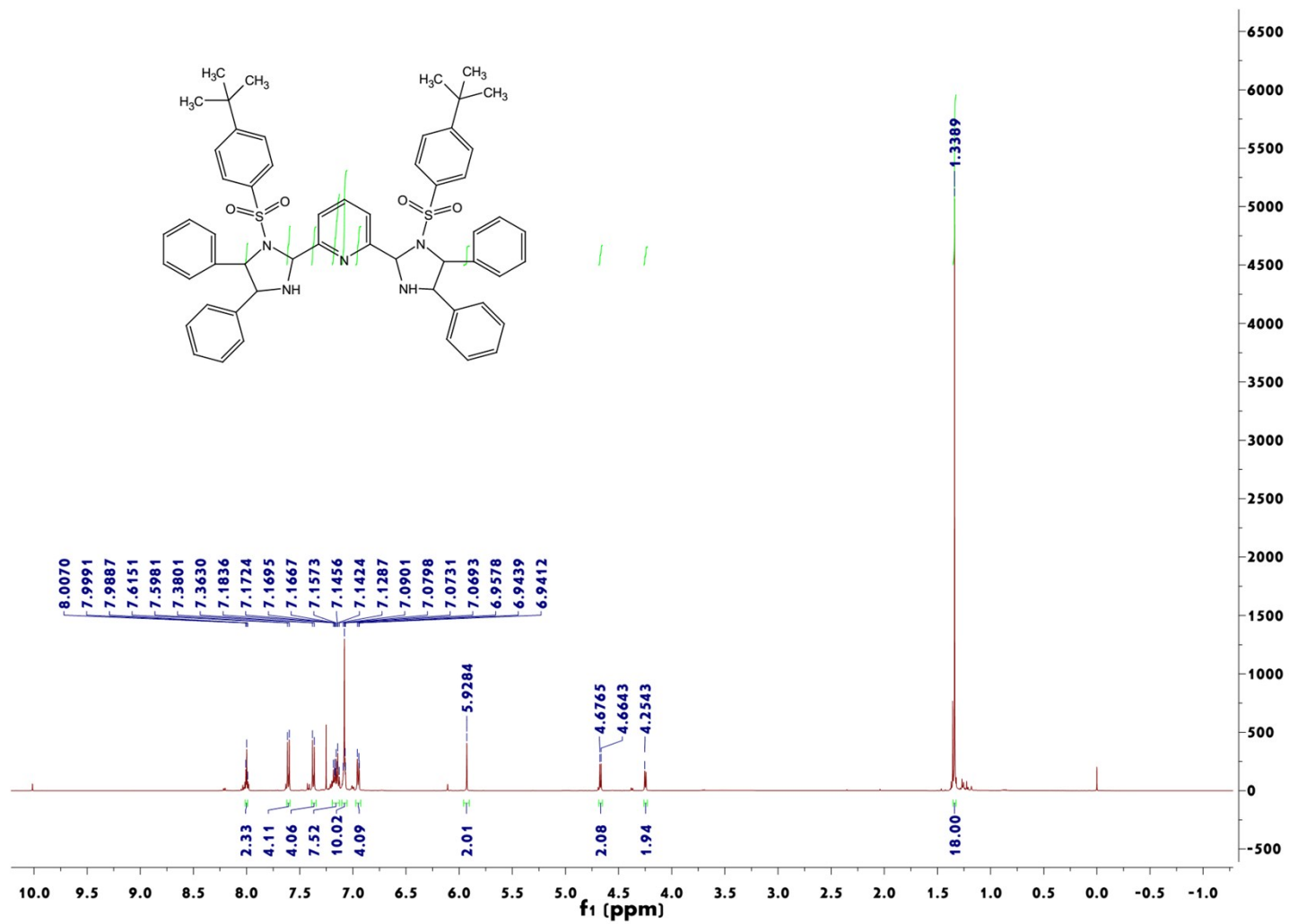
S22

L3



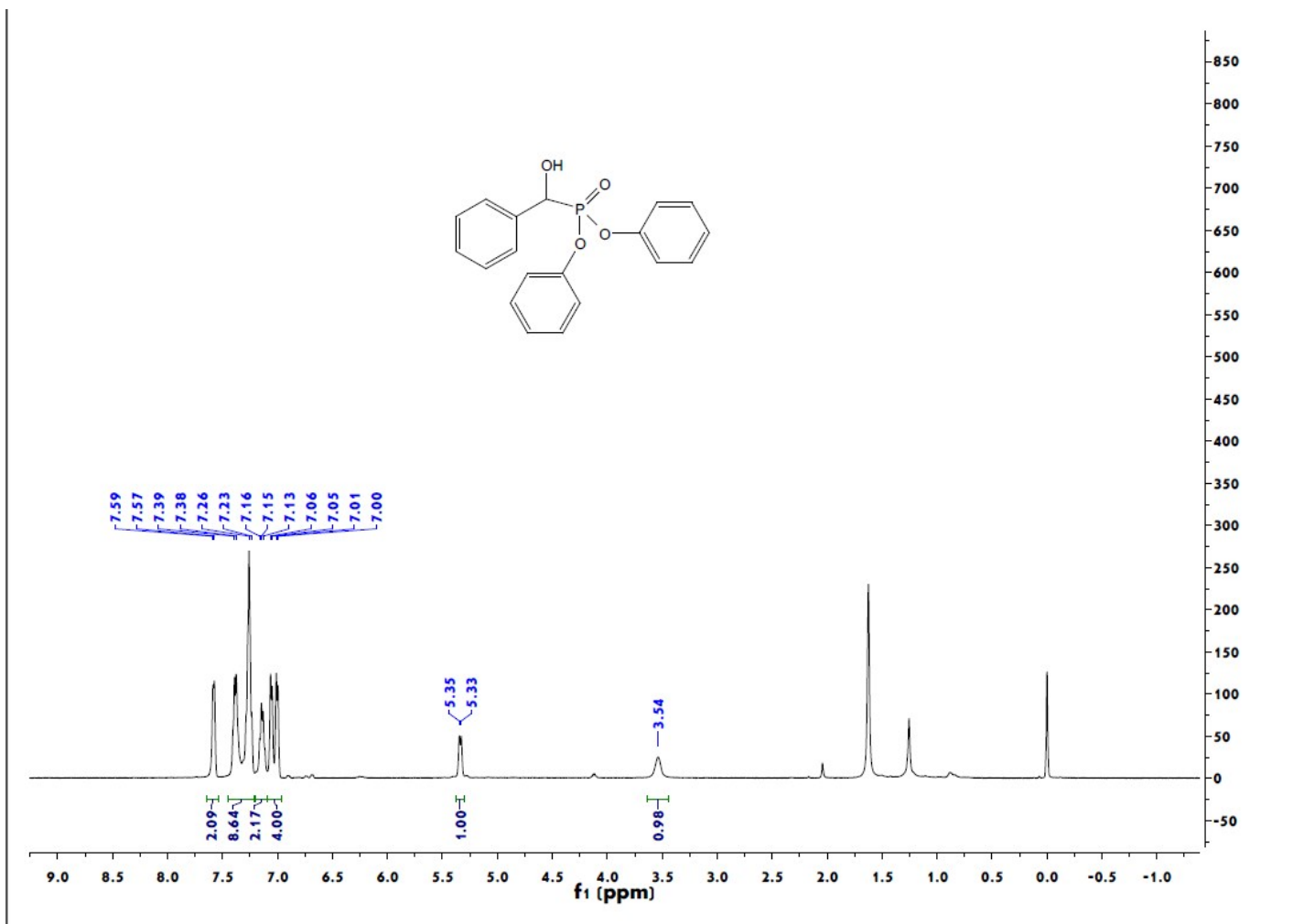
S23

L4



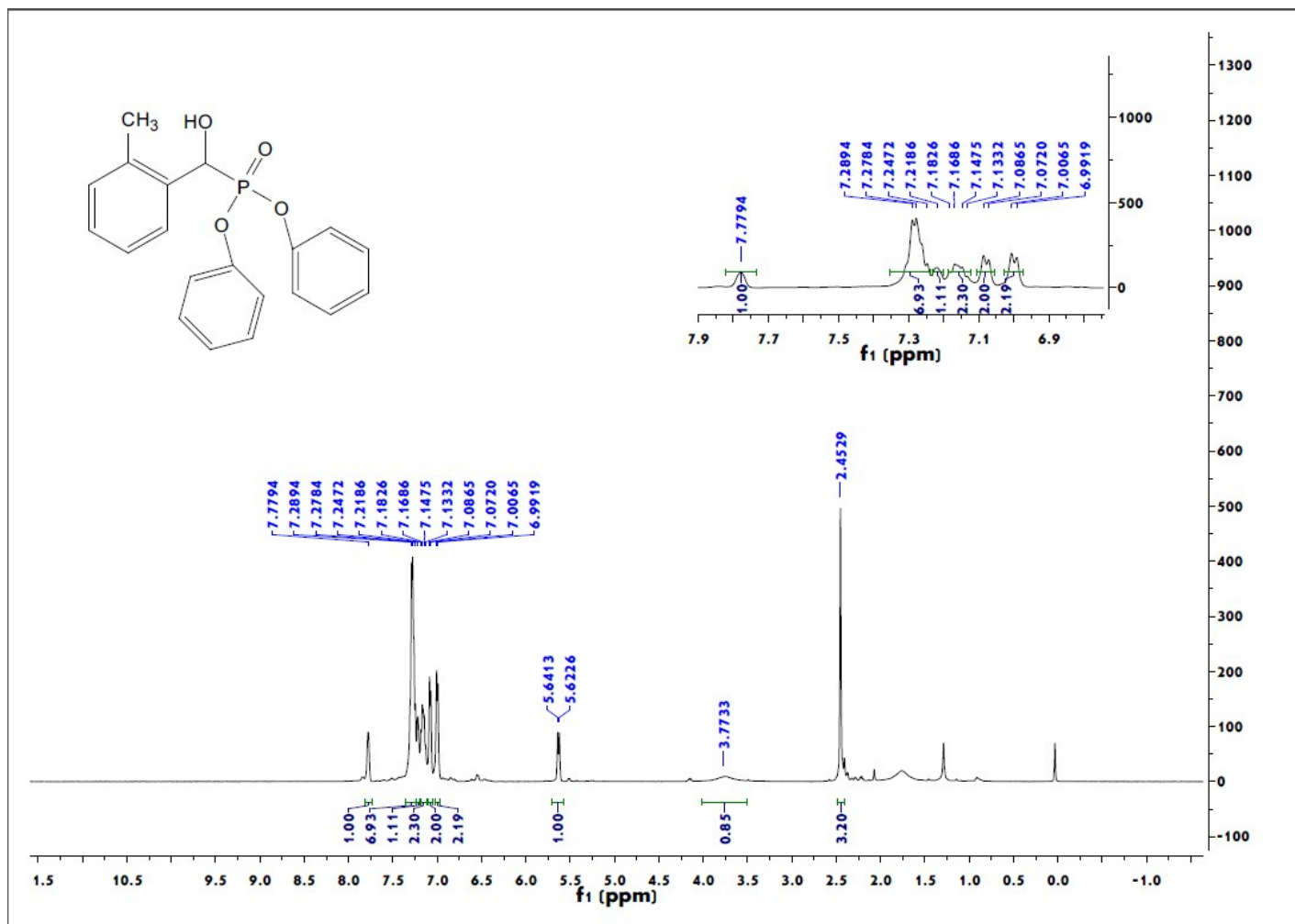
S24

2a

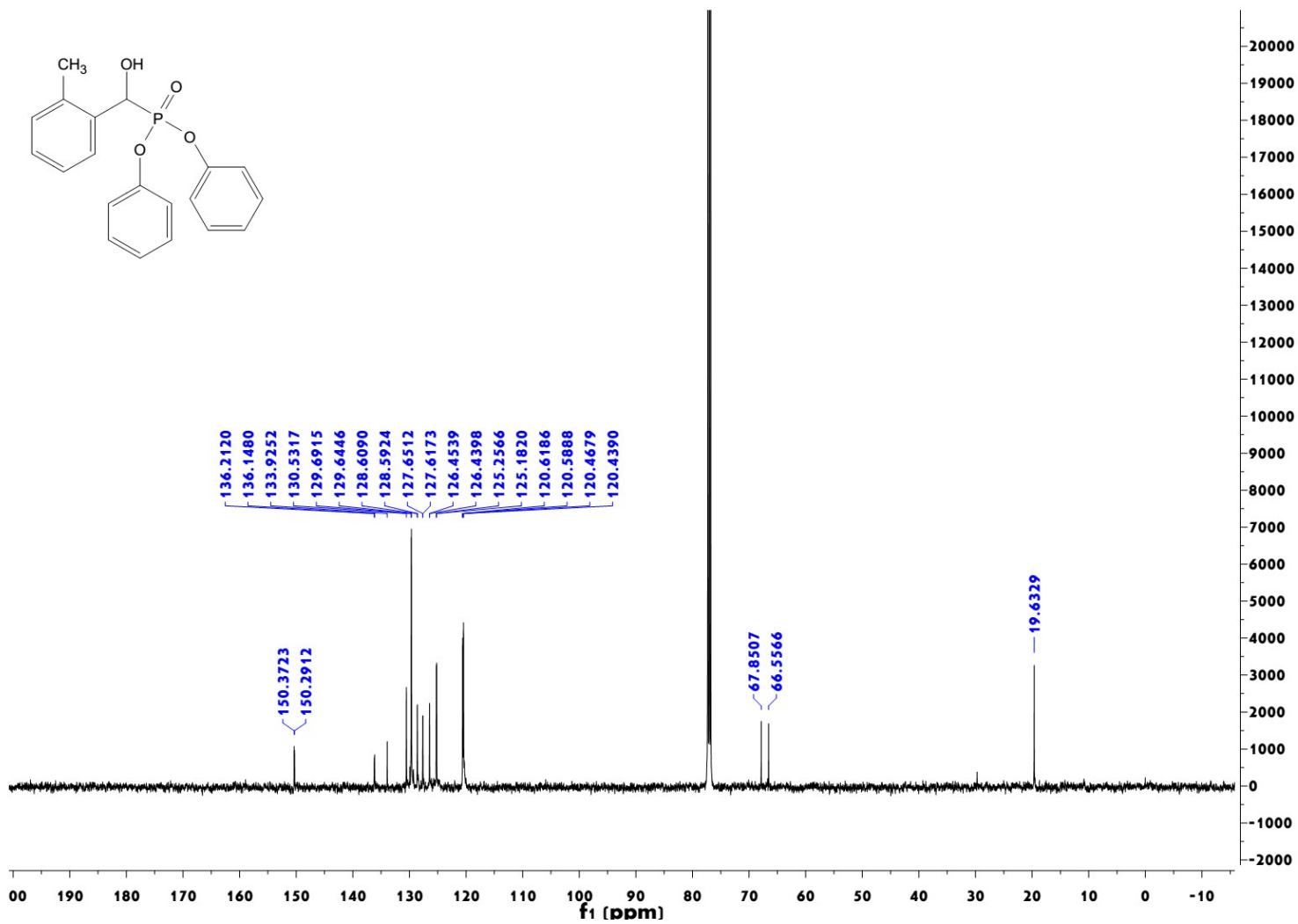


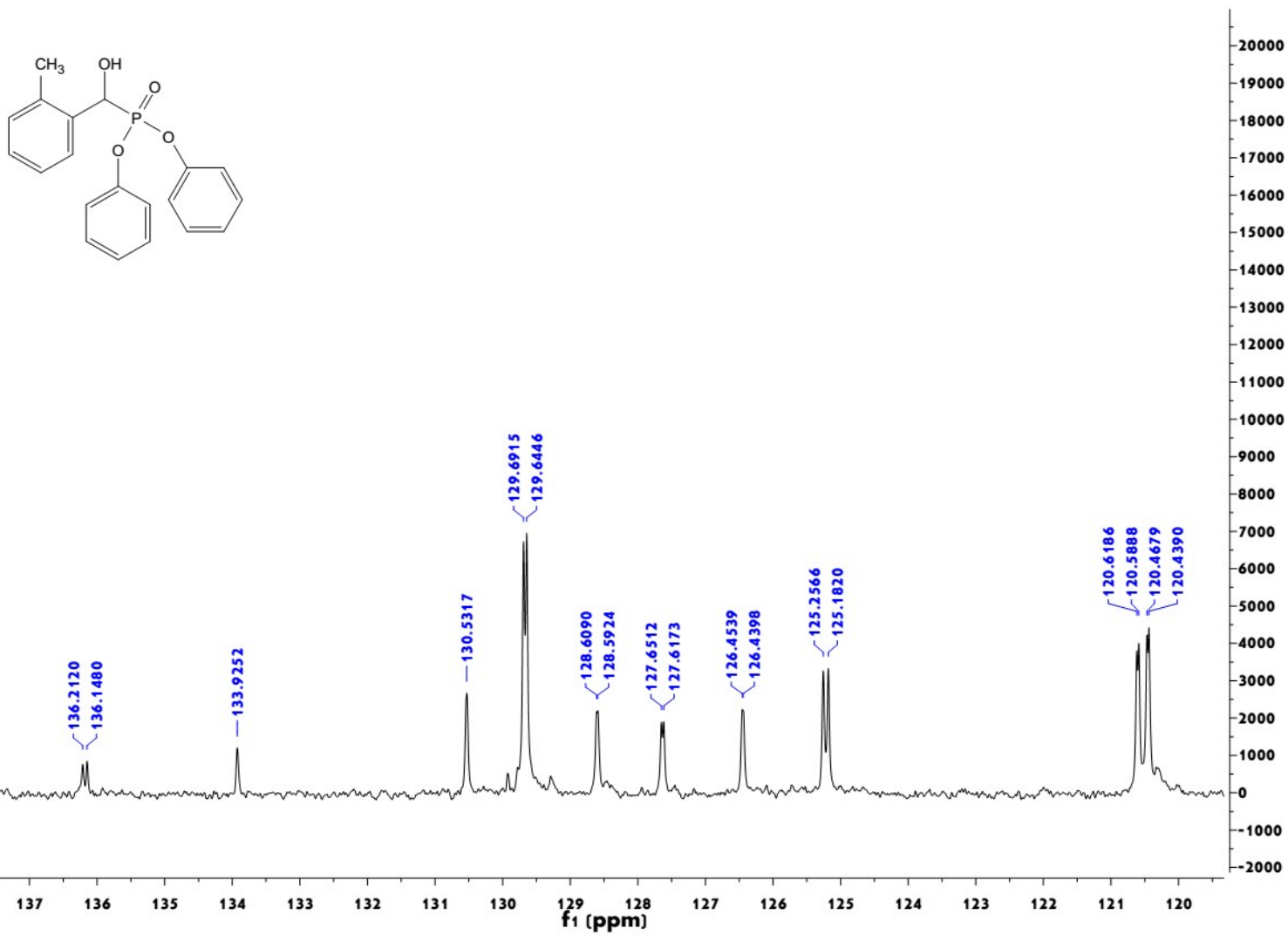
S25

2b

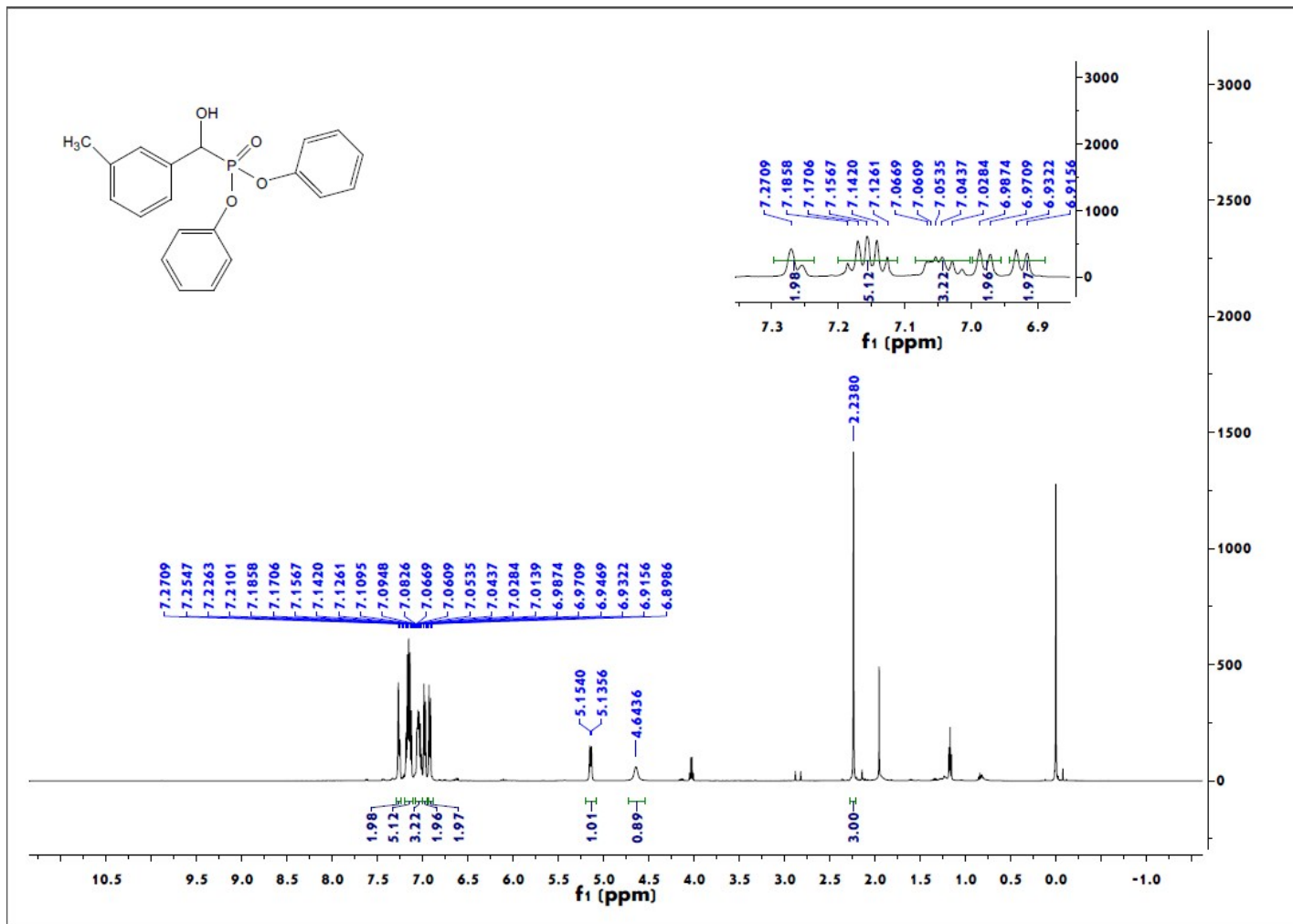


2b

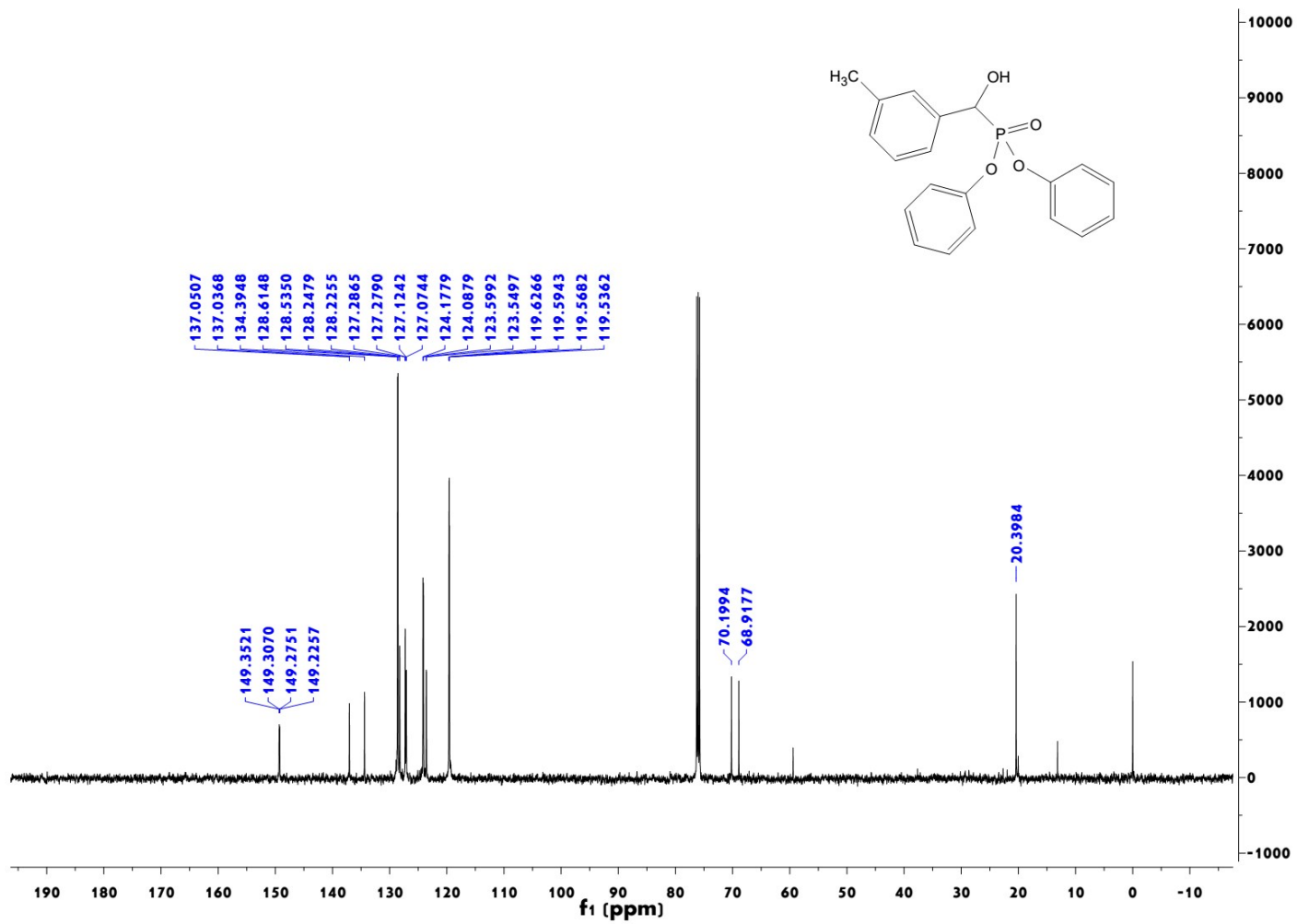


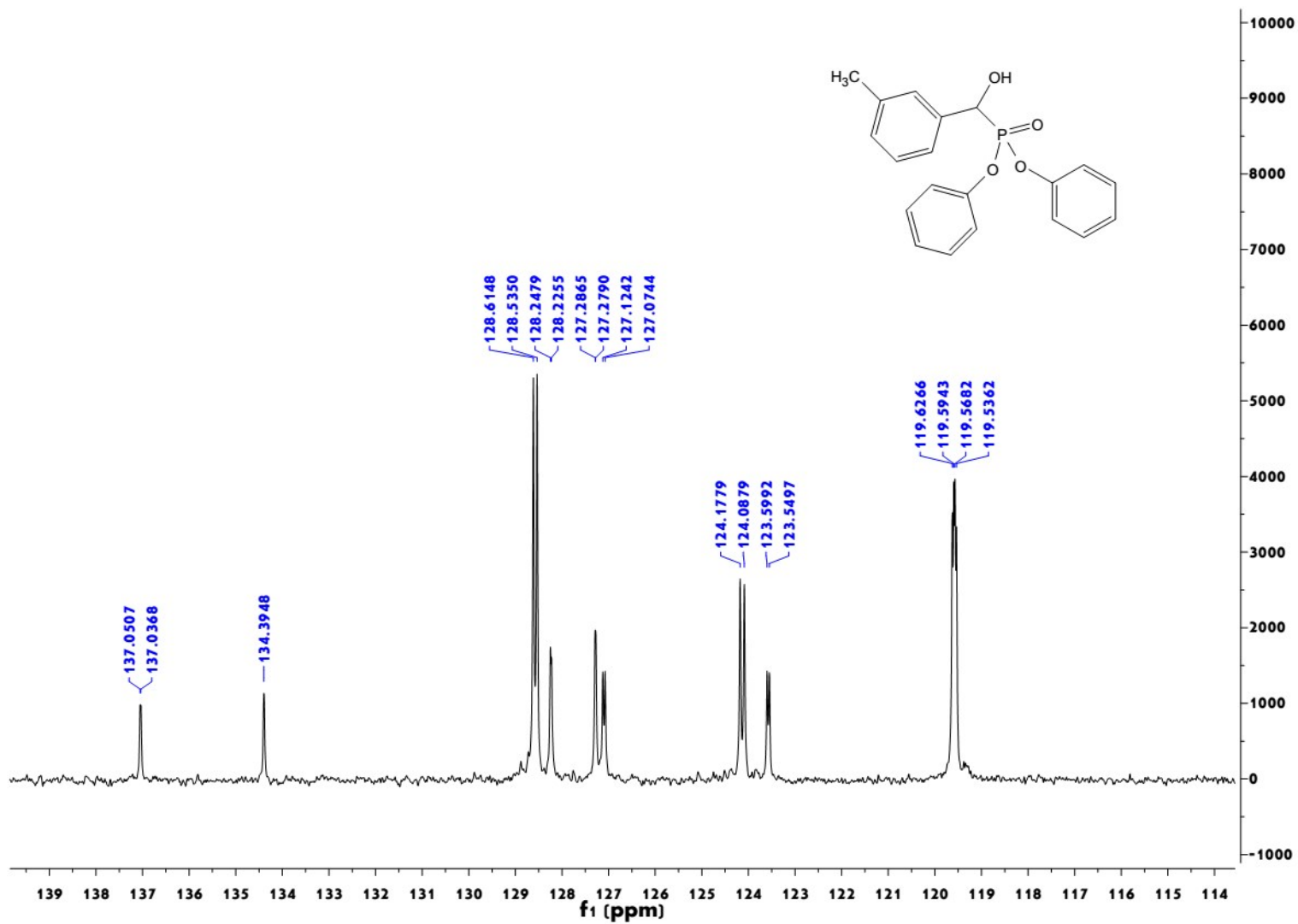


2c

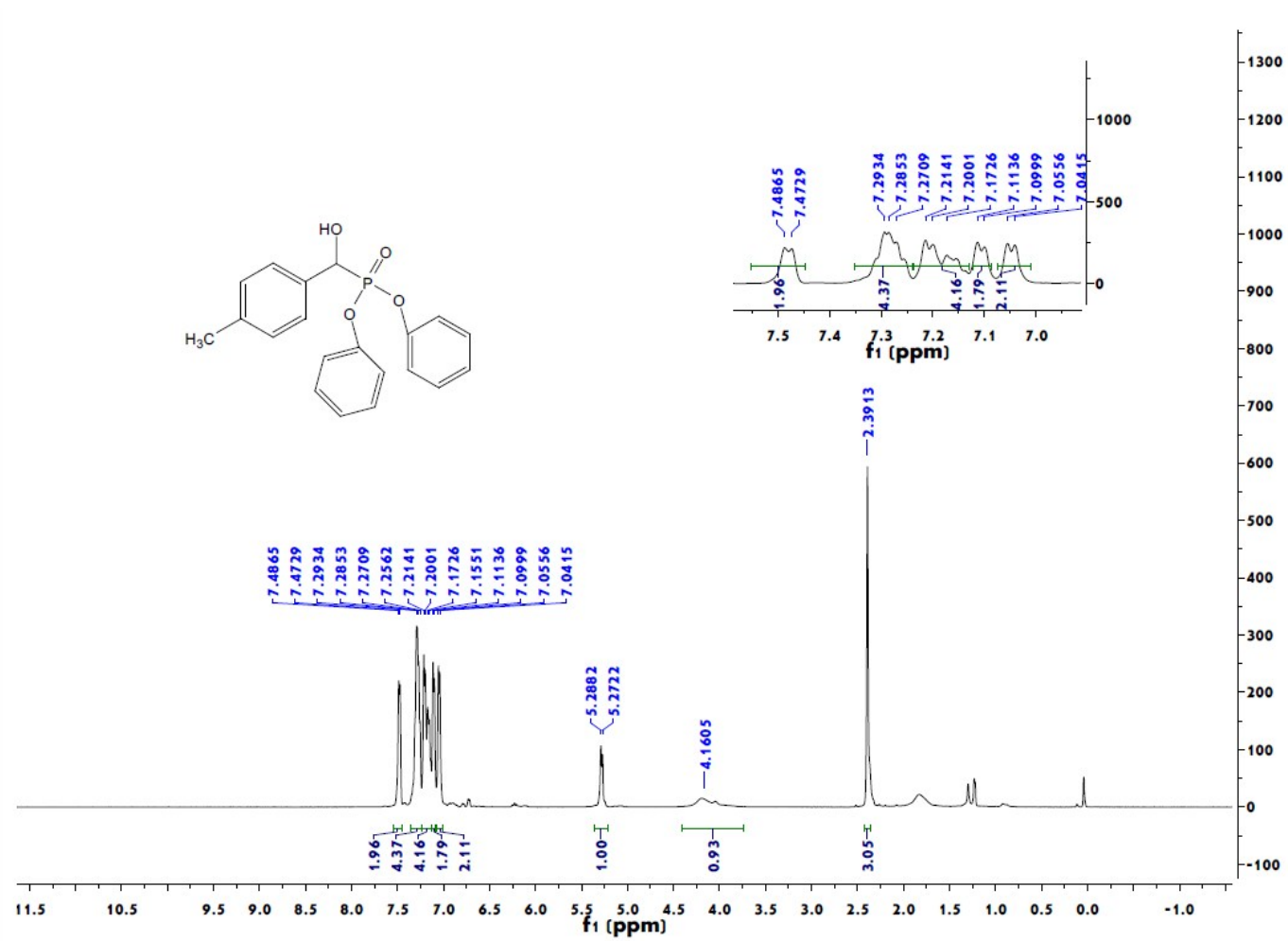


2c



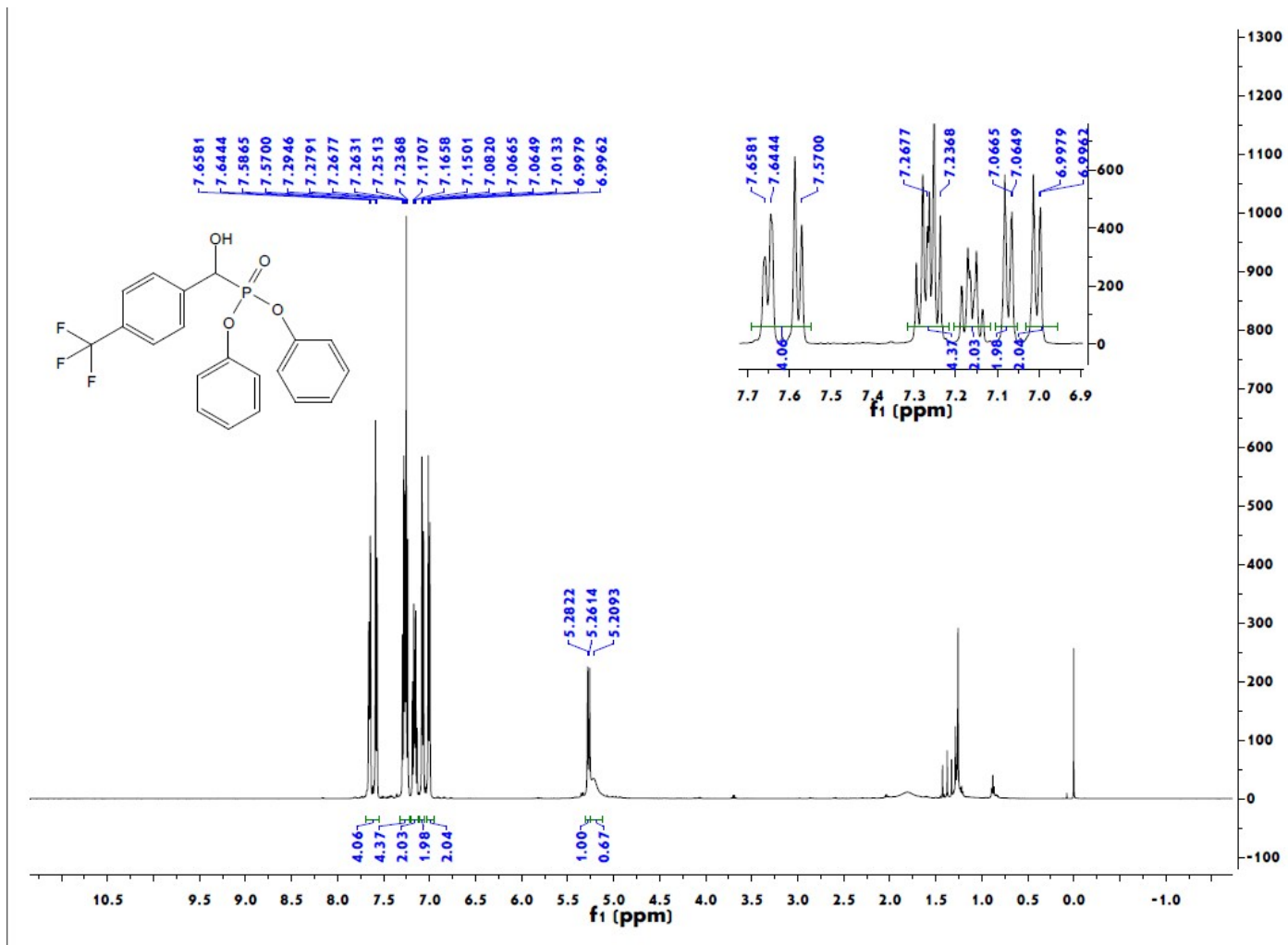


2d

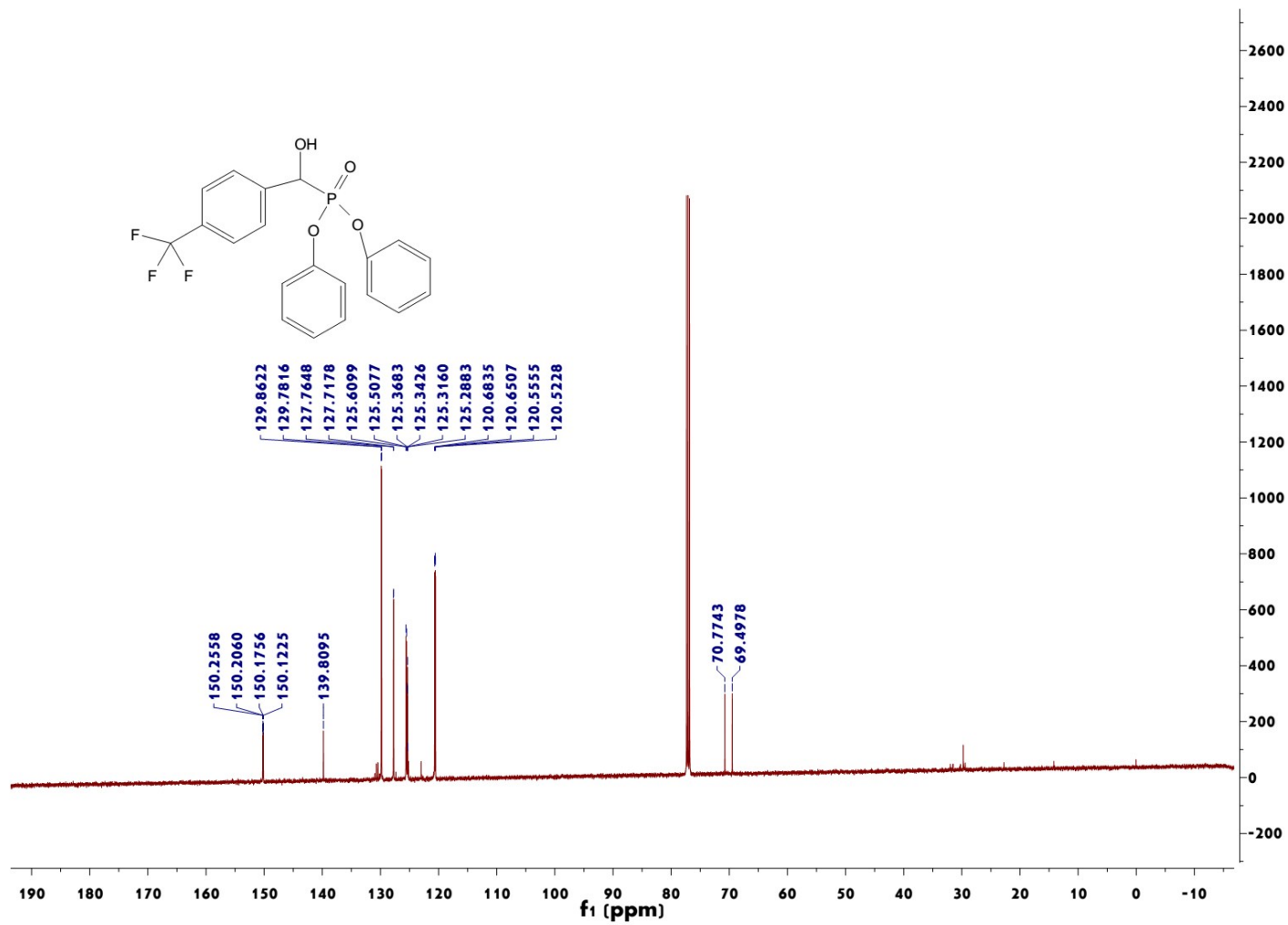


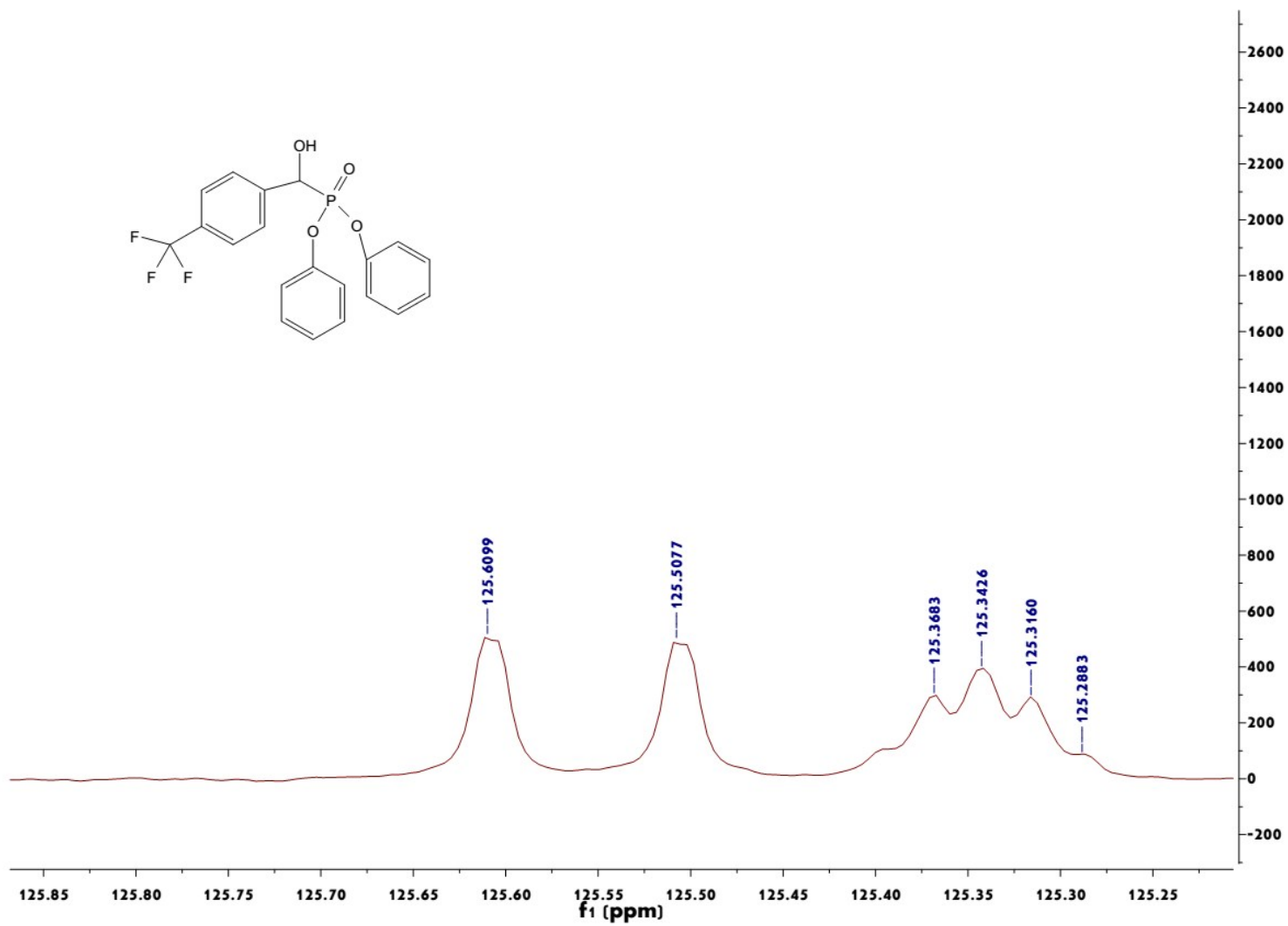
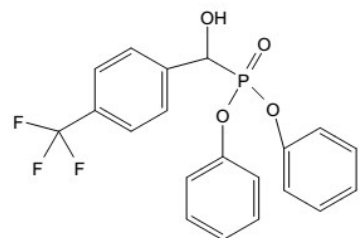
S32

2e

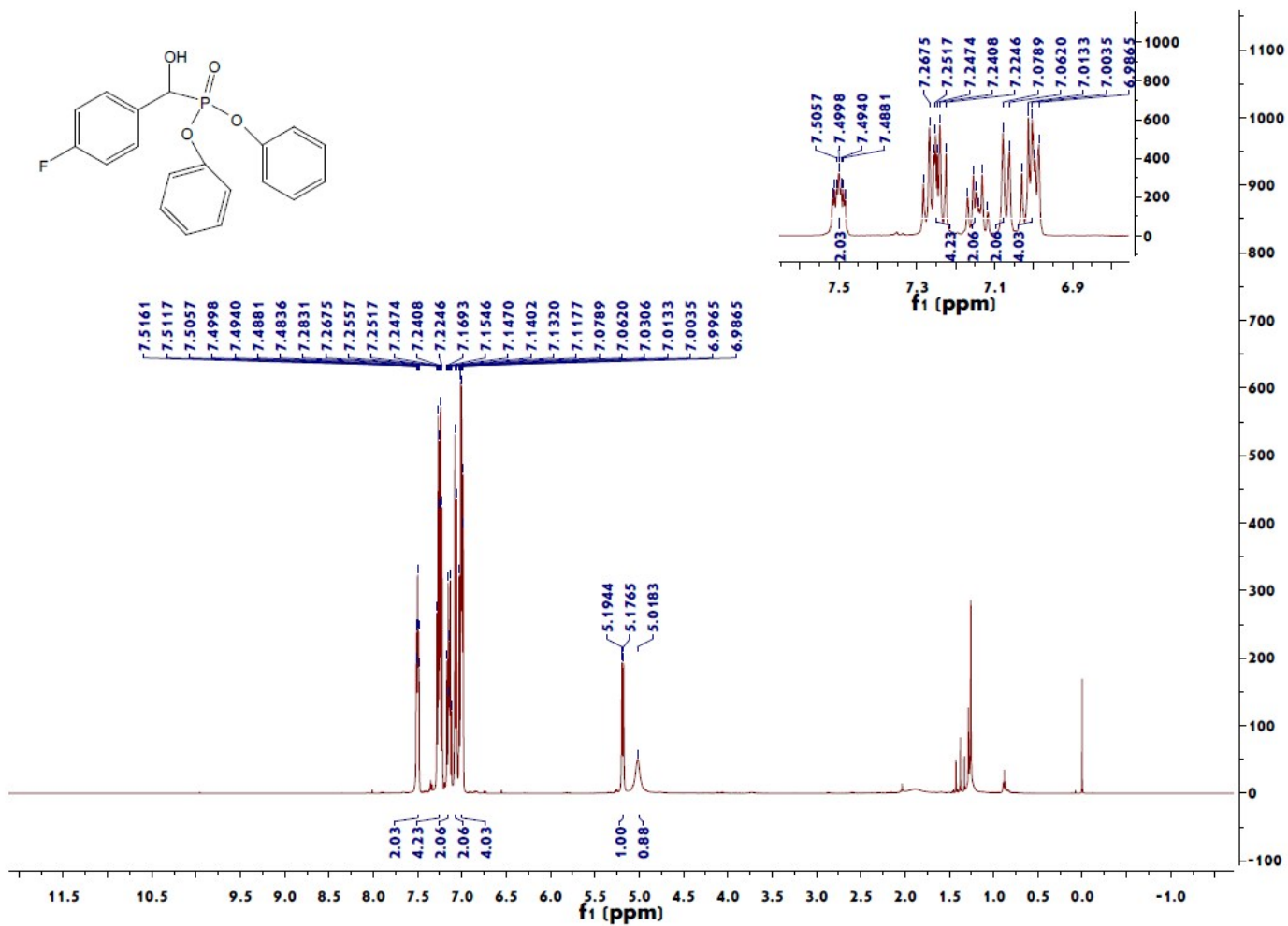


2e

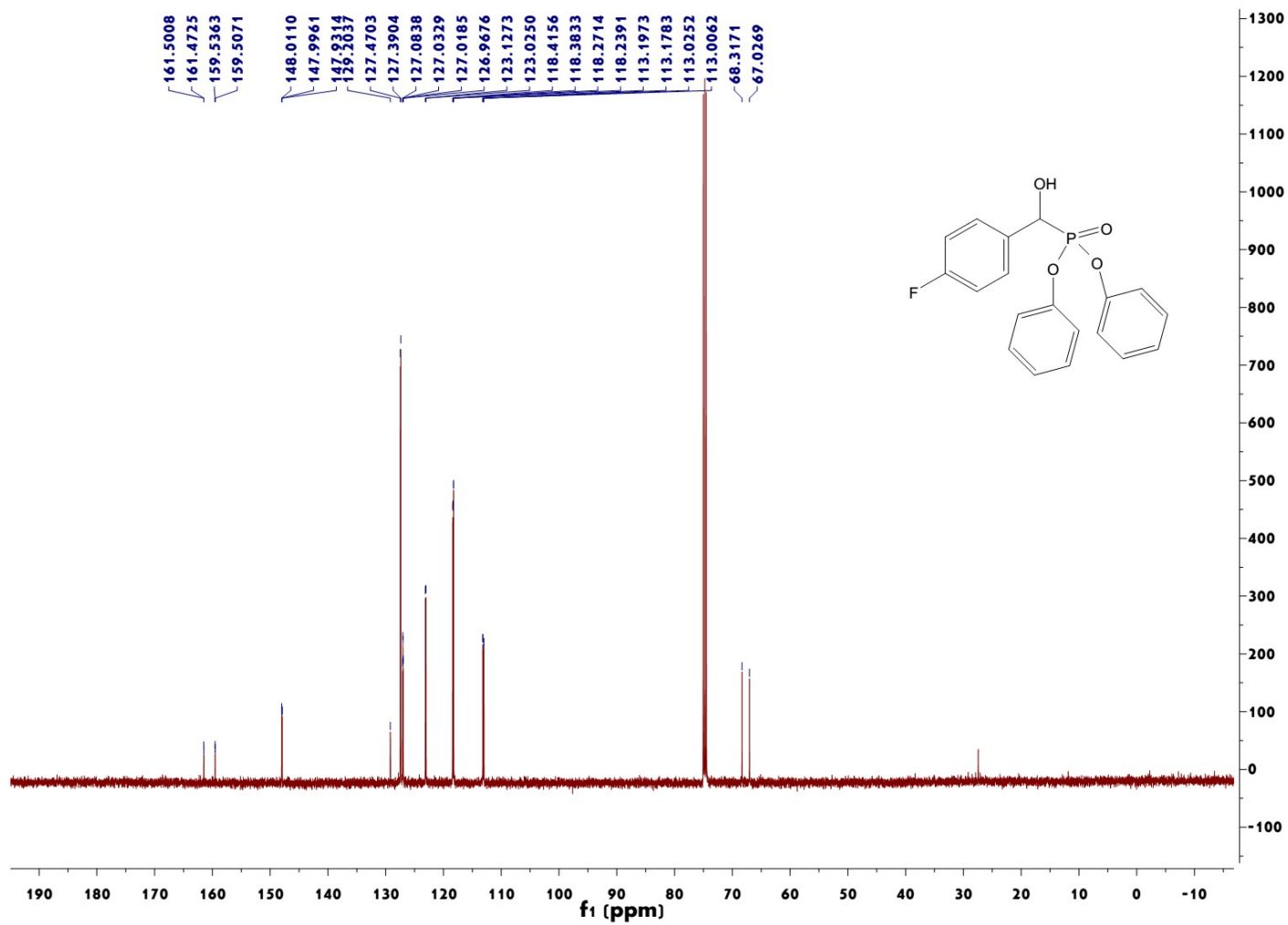


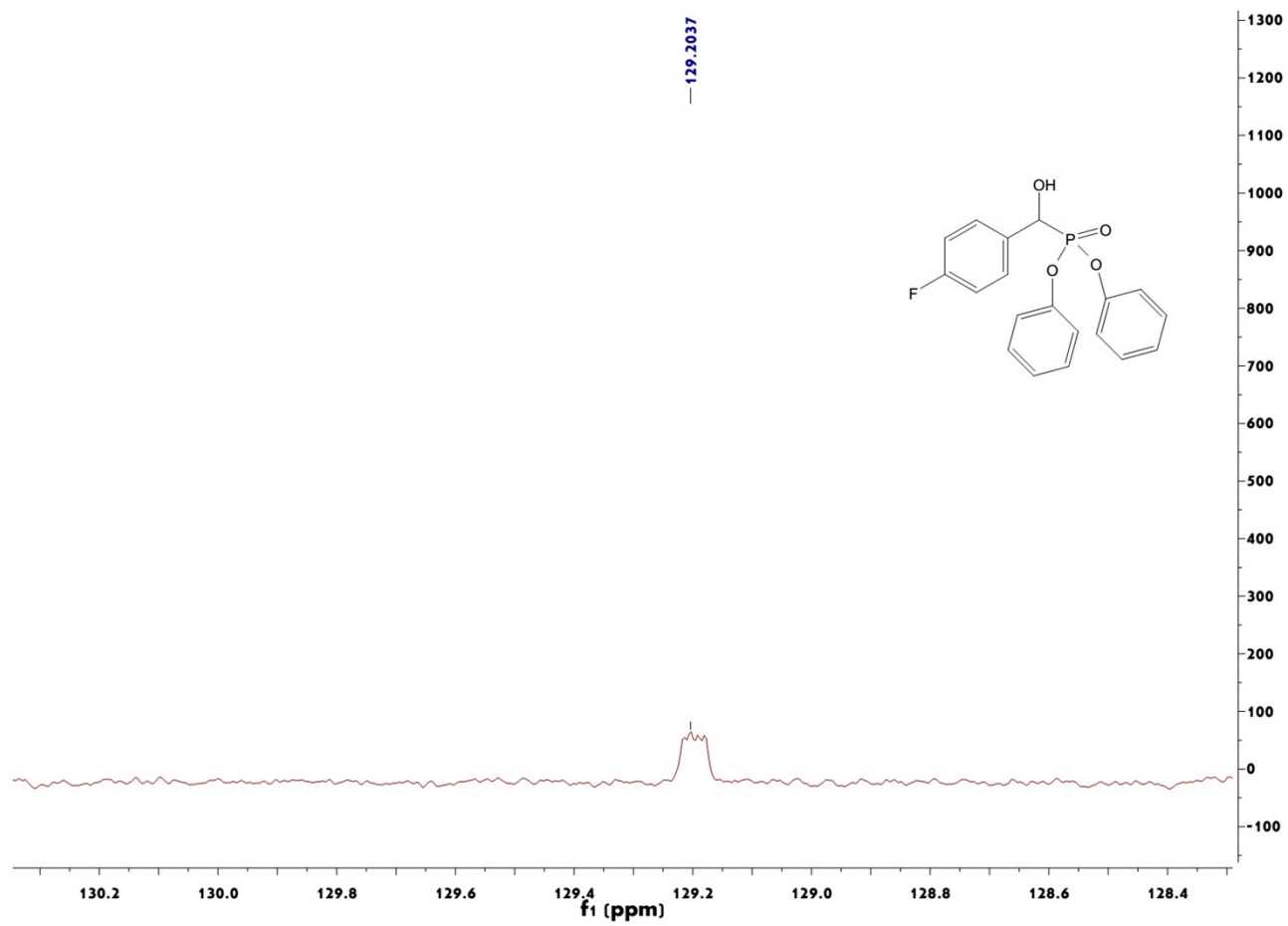


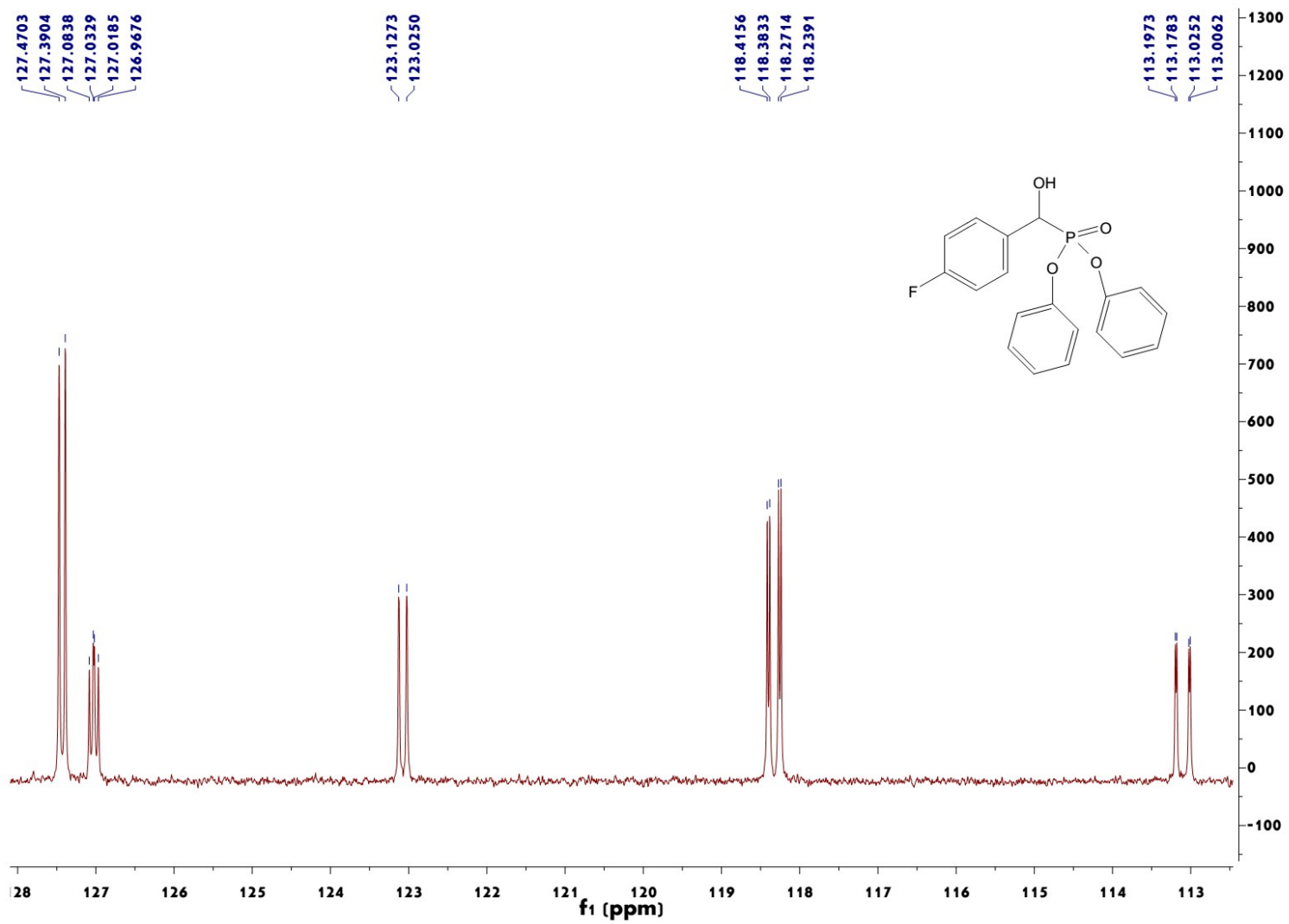
2f



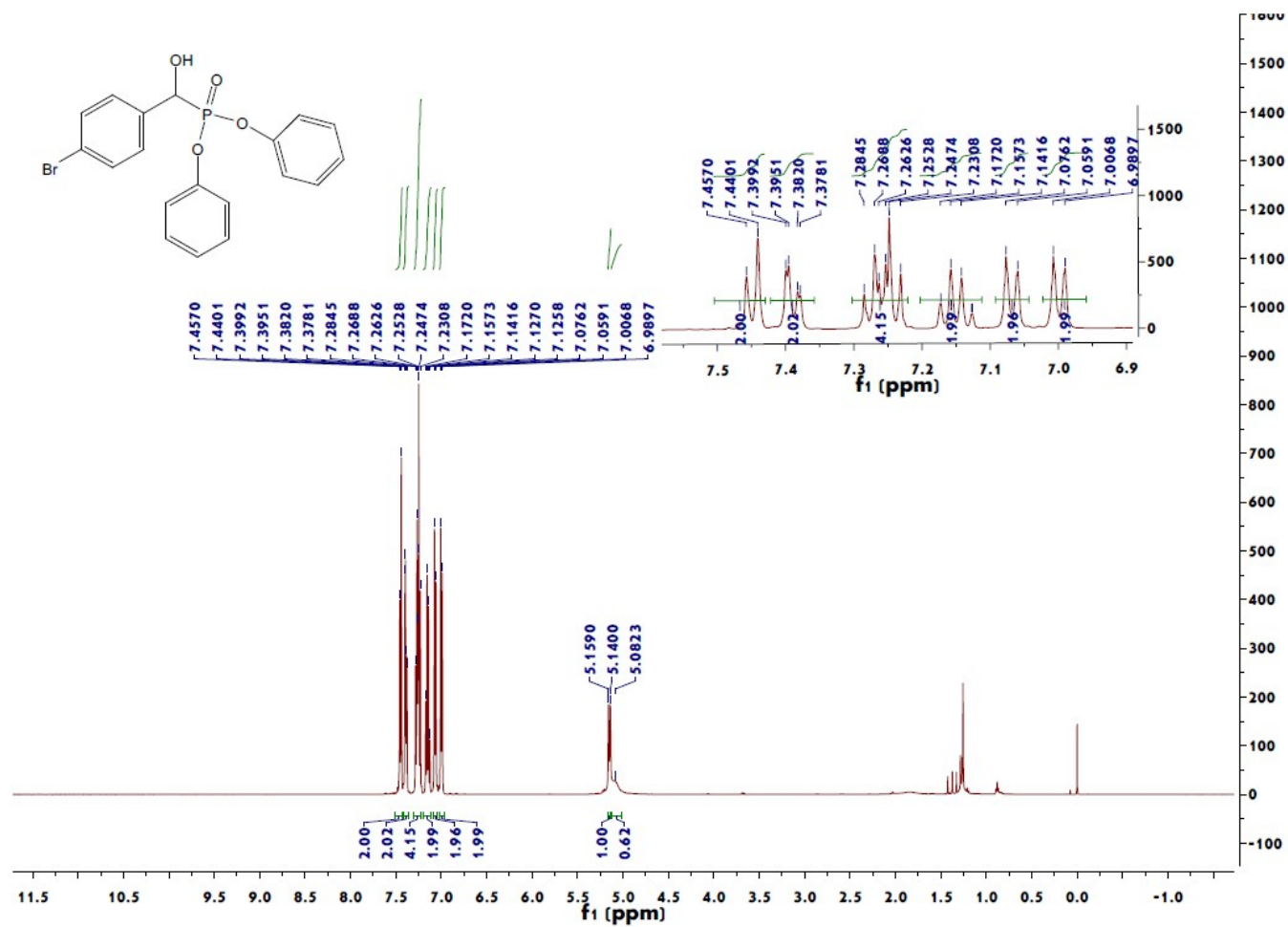
2f



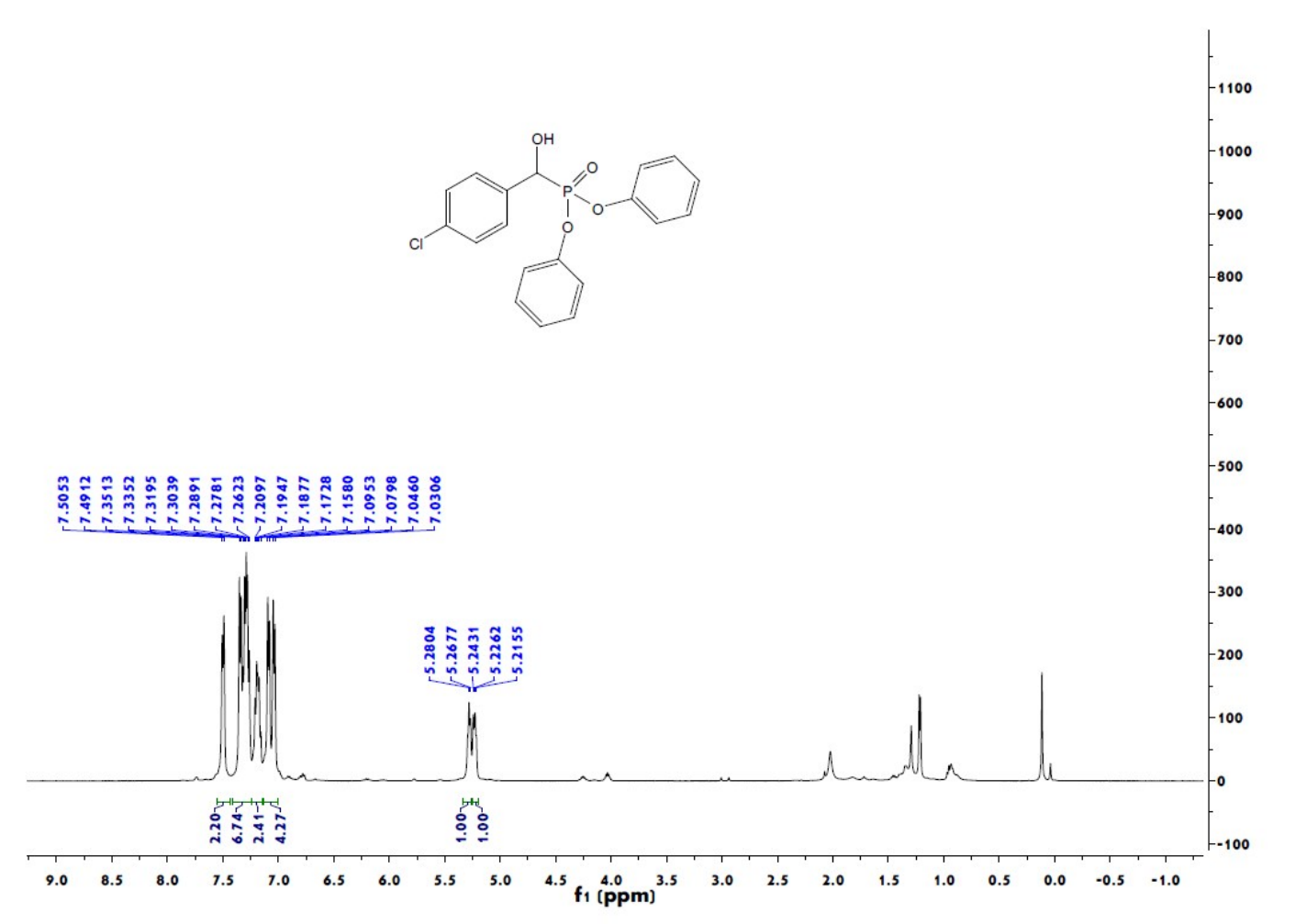




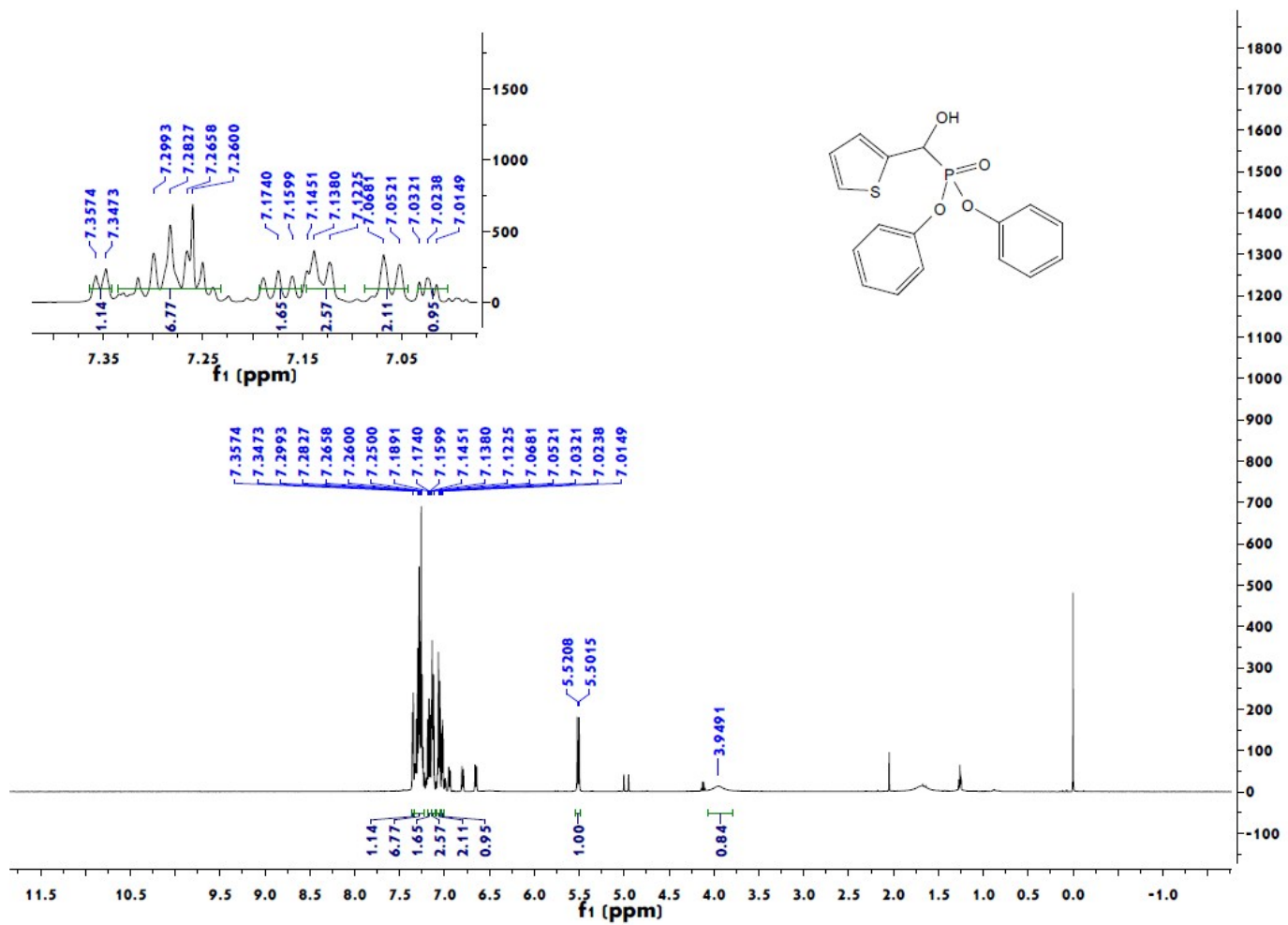
2g



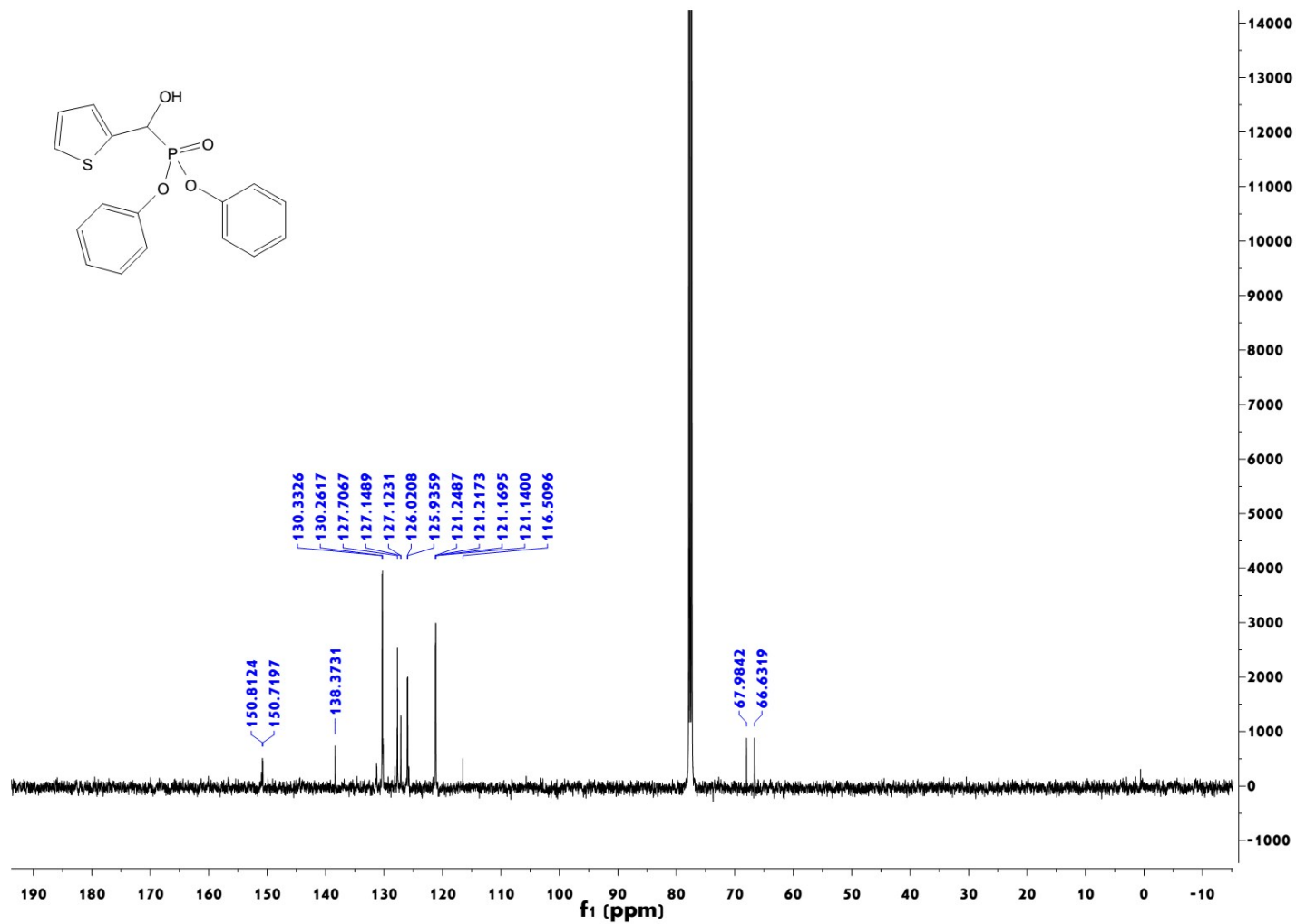
2h

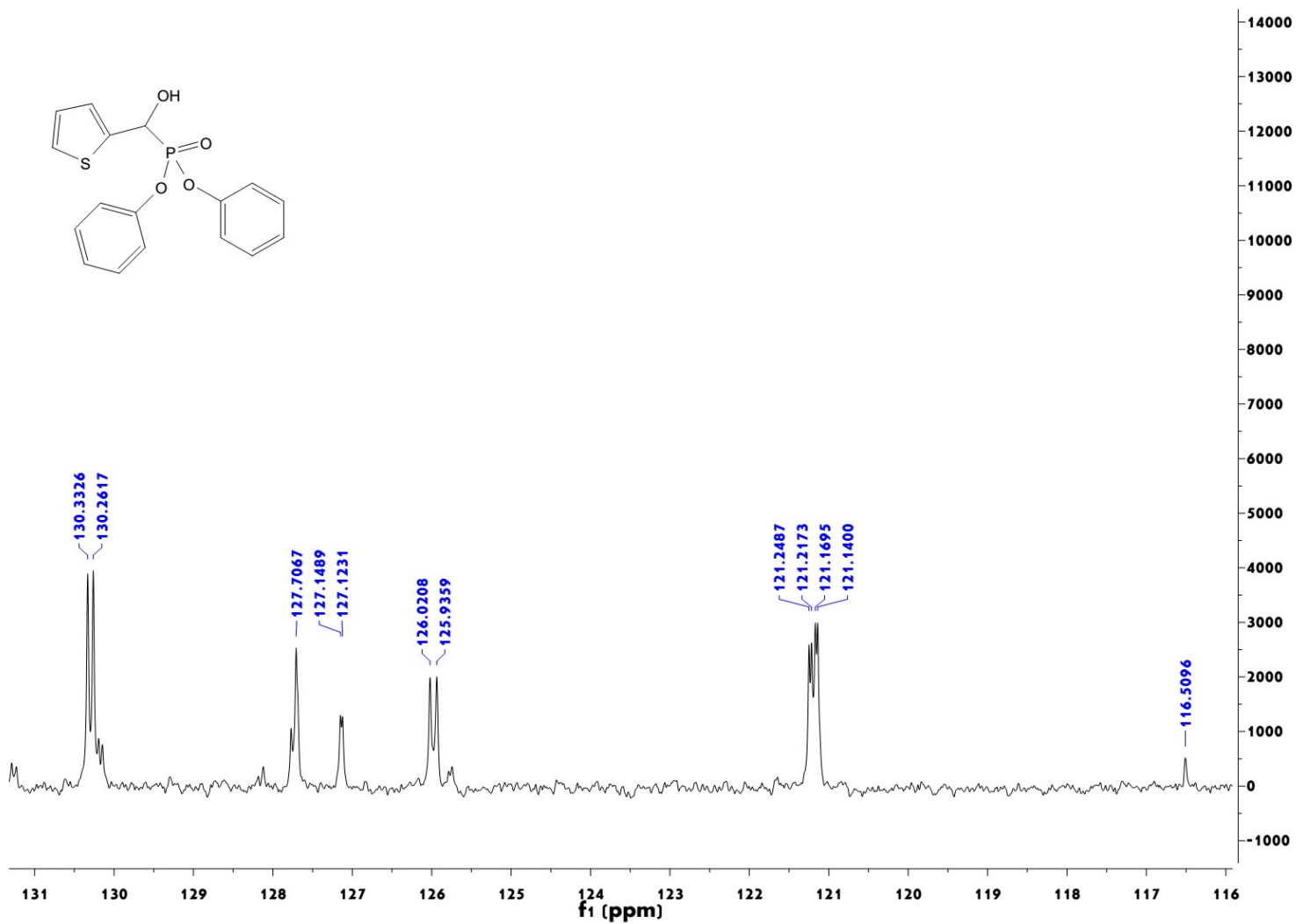
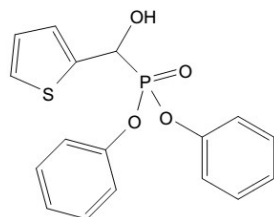


2i

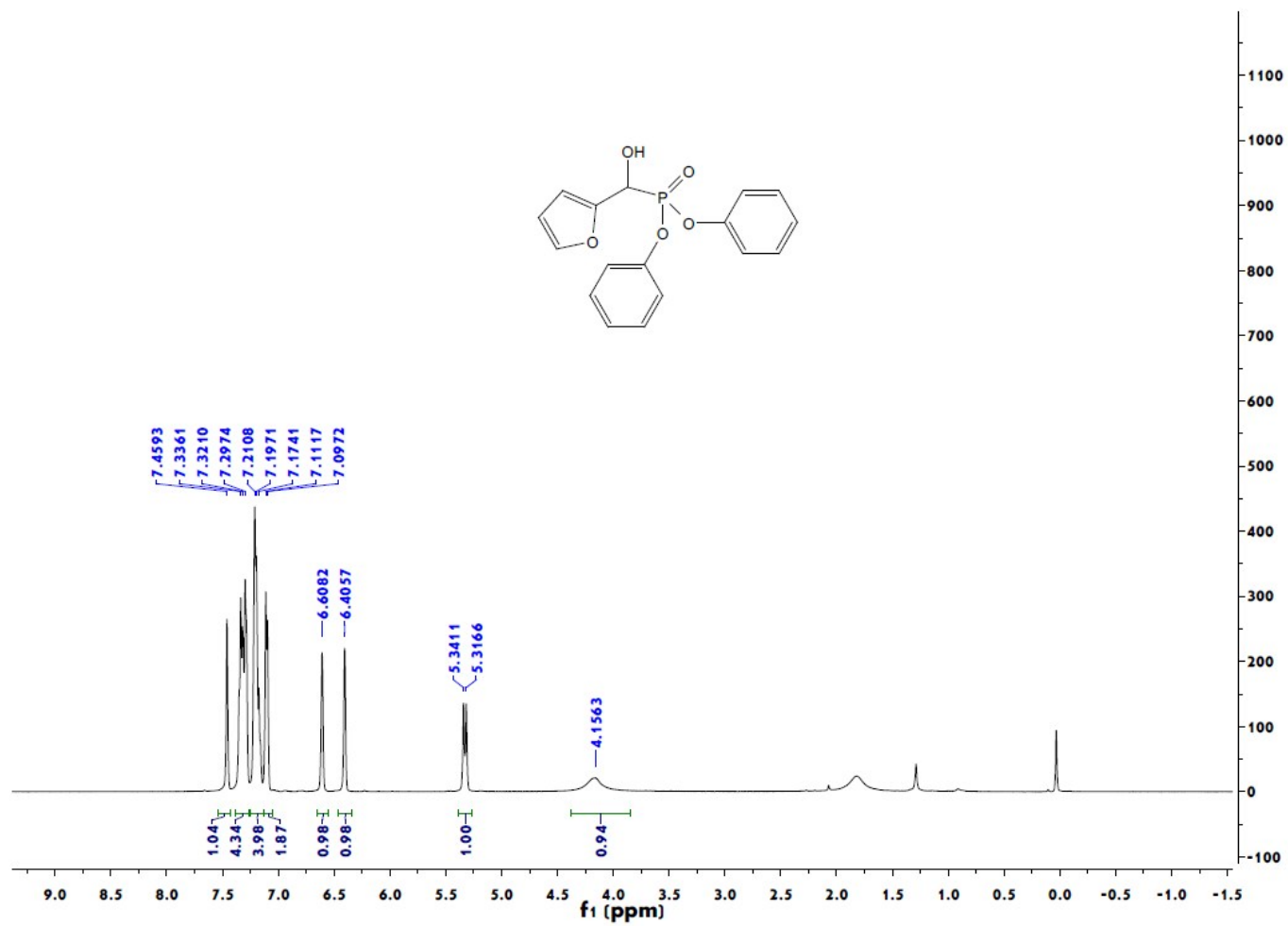


2i

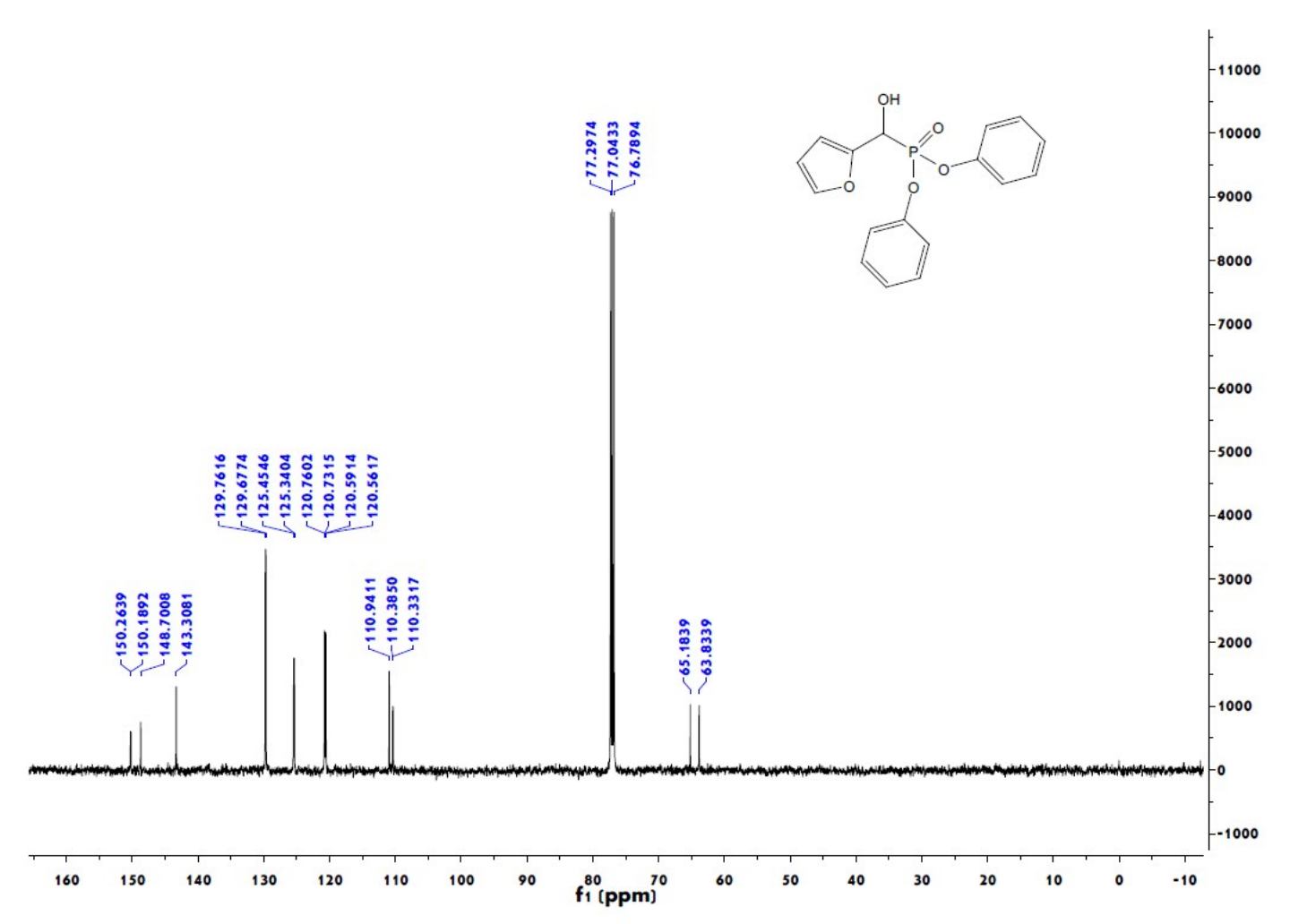




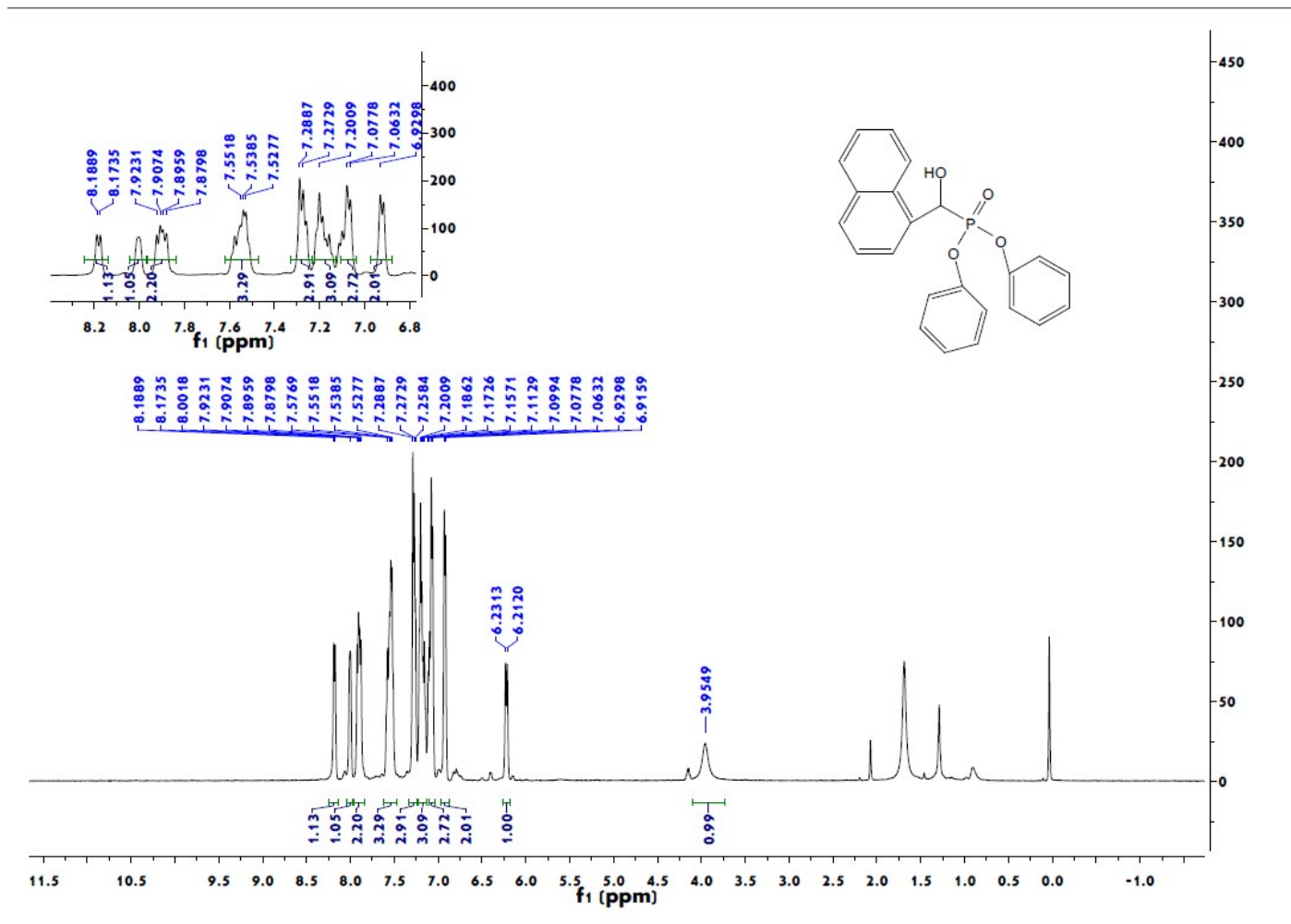
2j



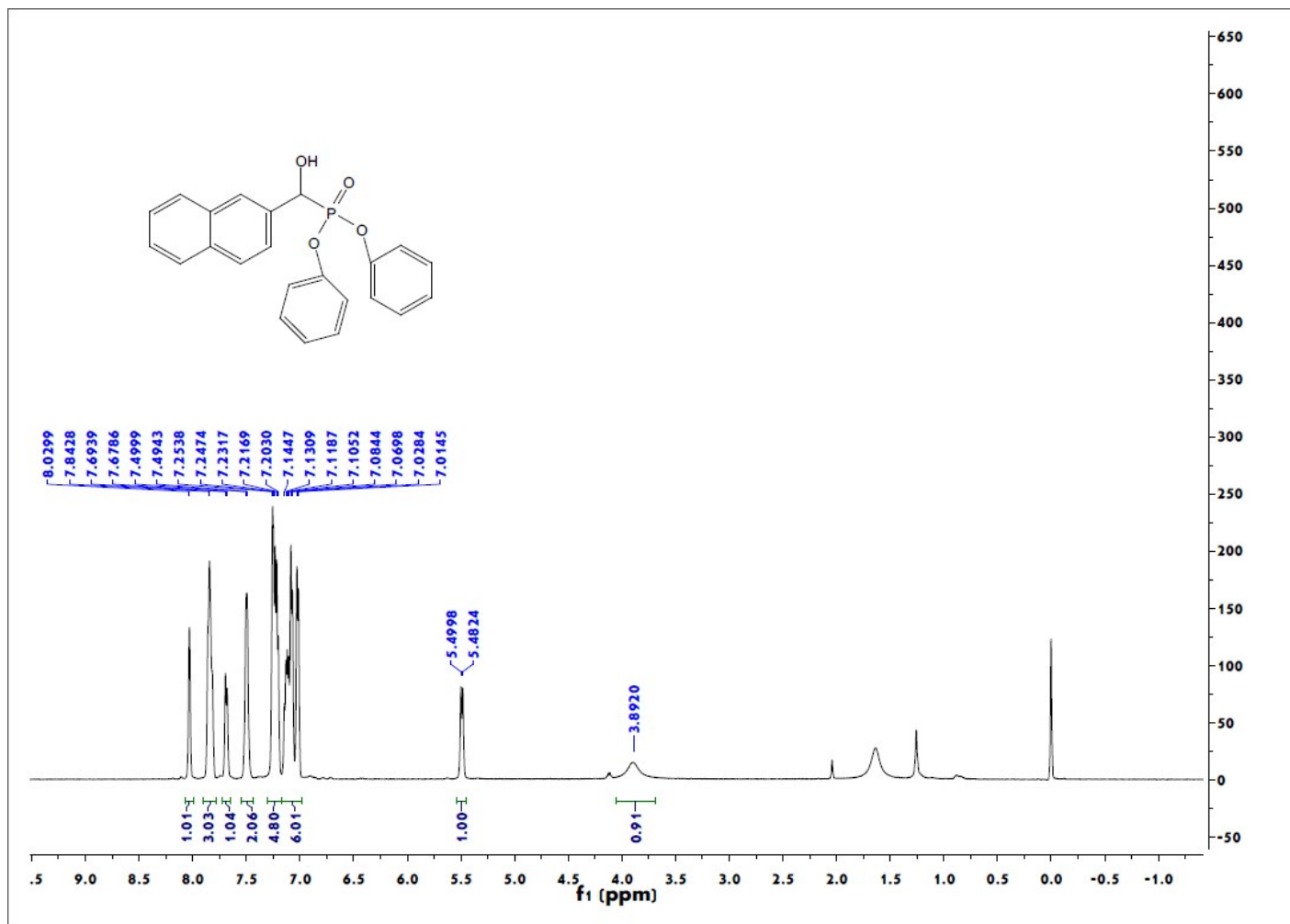
2j



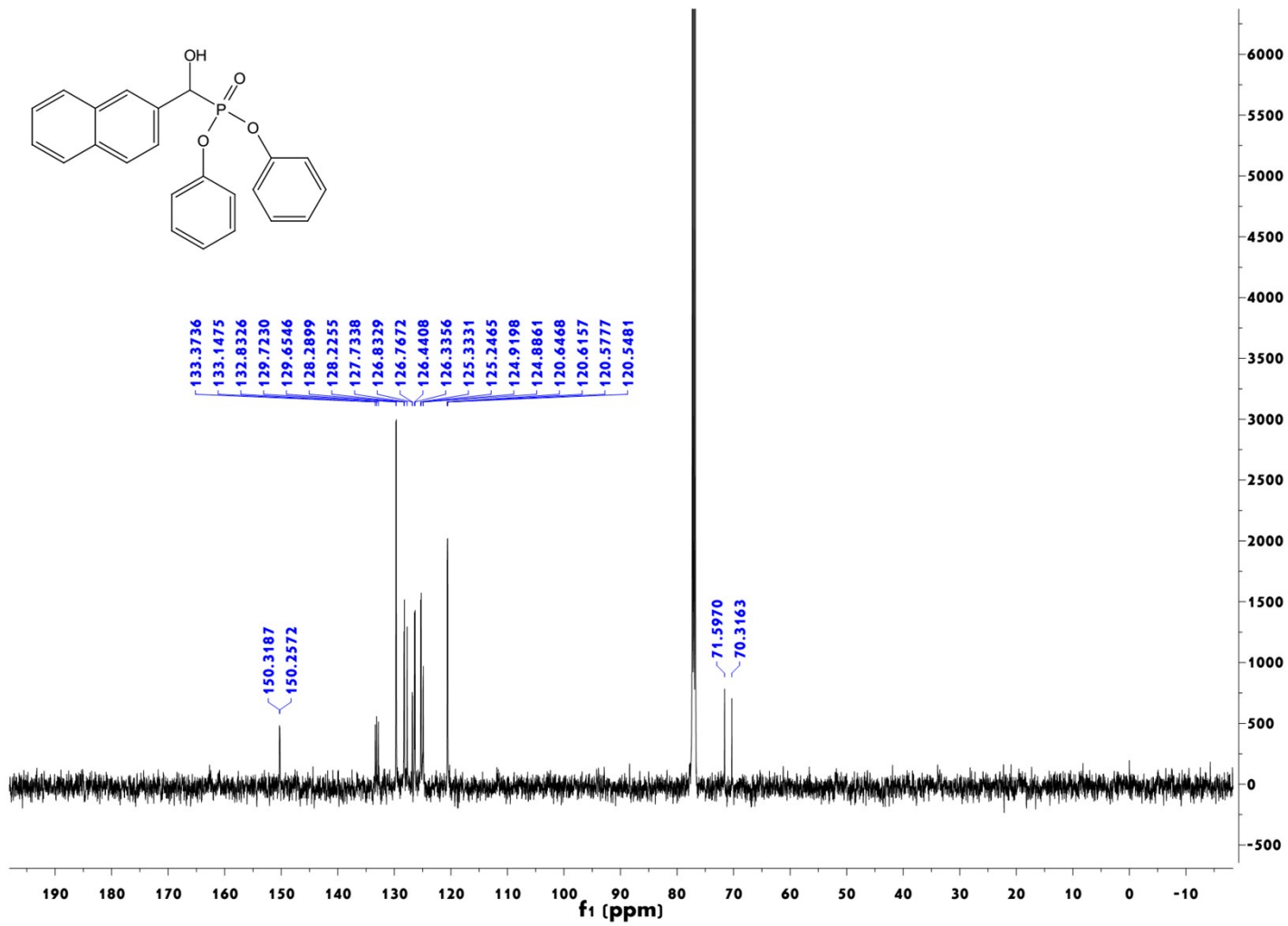
2k

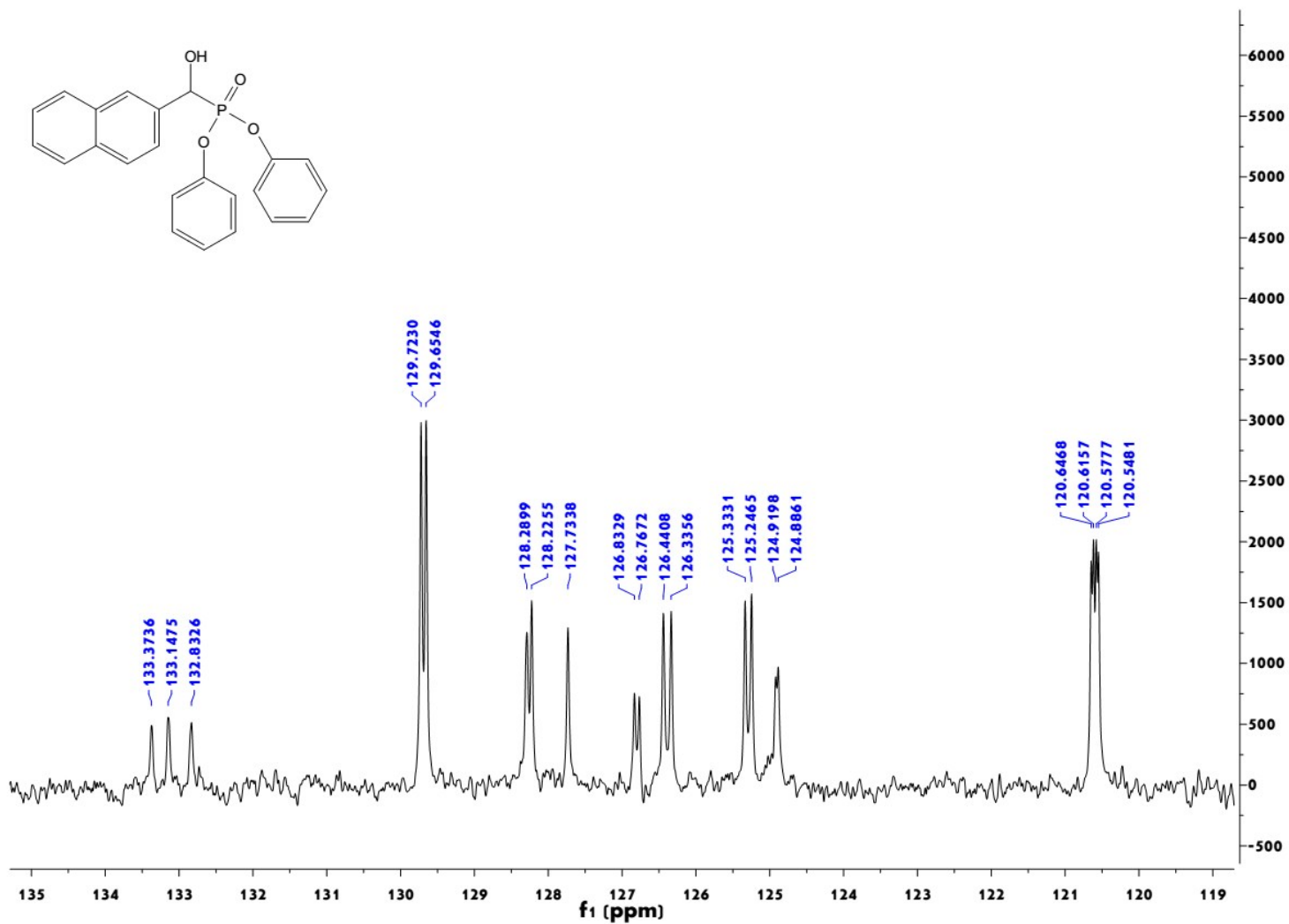
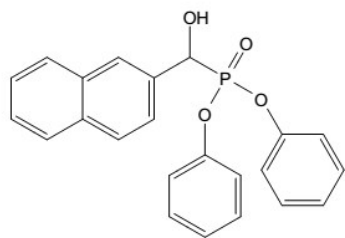


S47

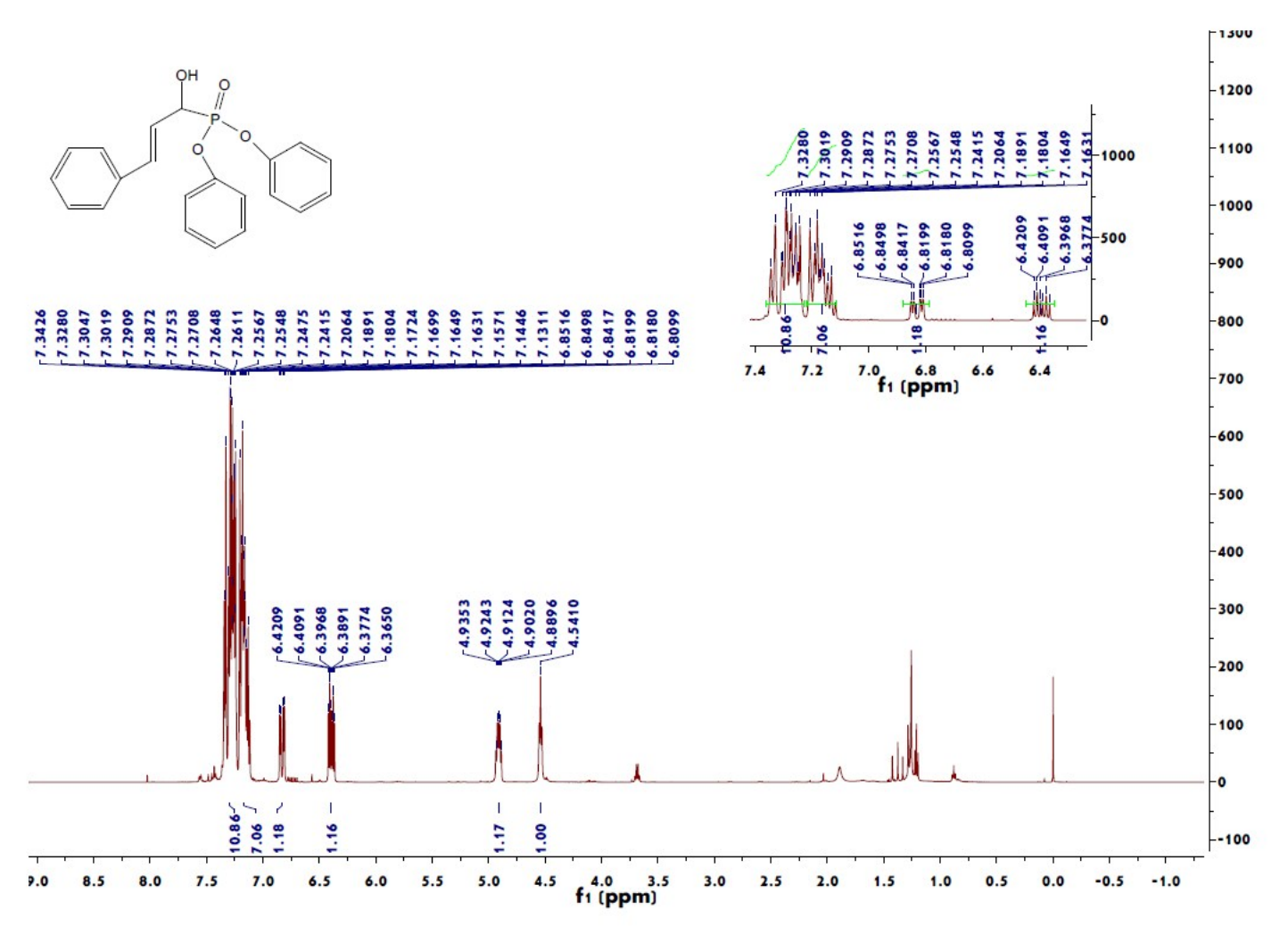


21

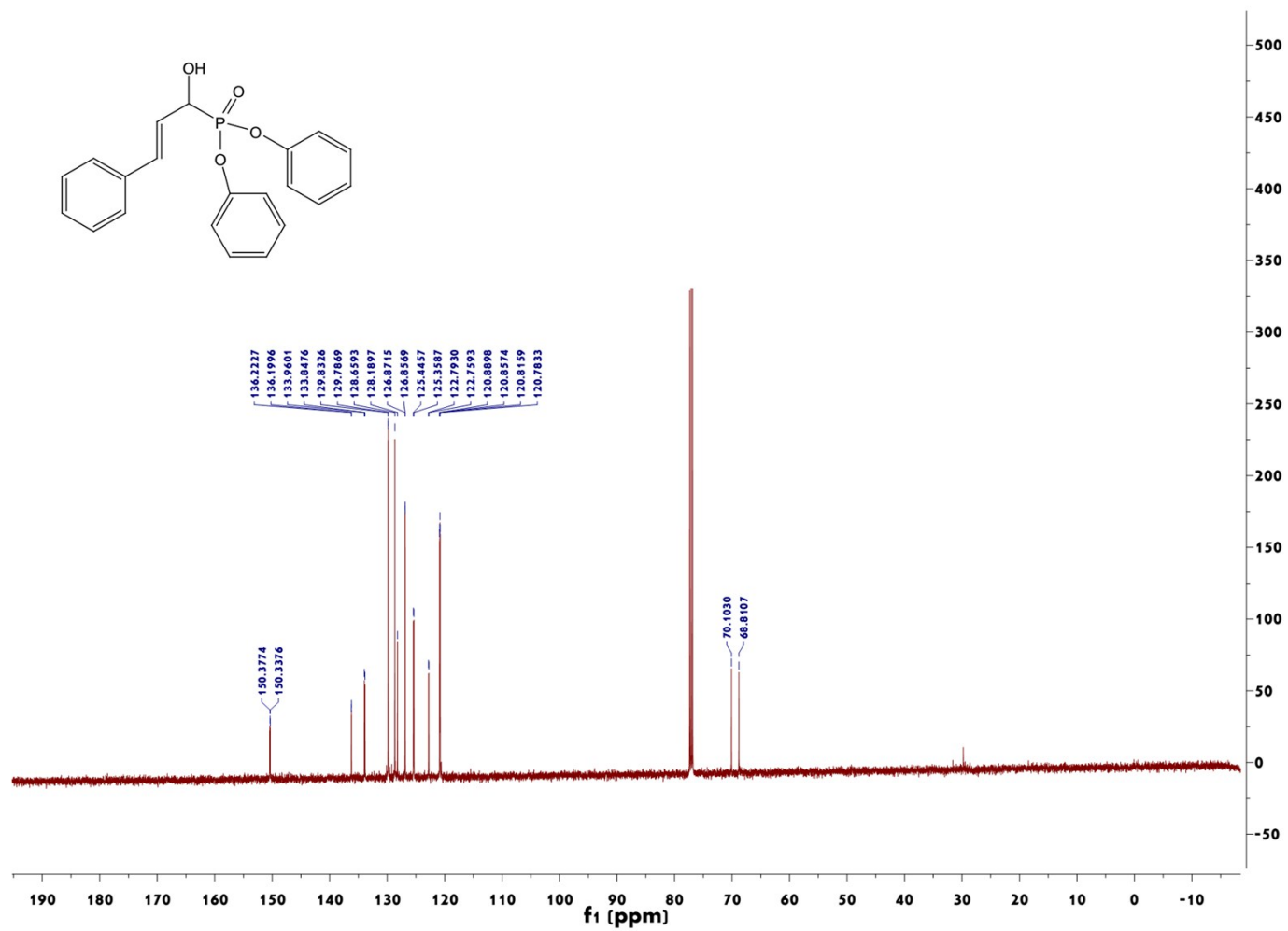




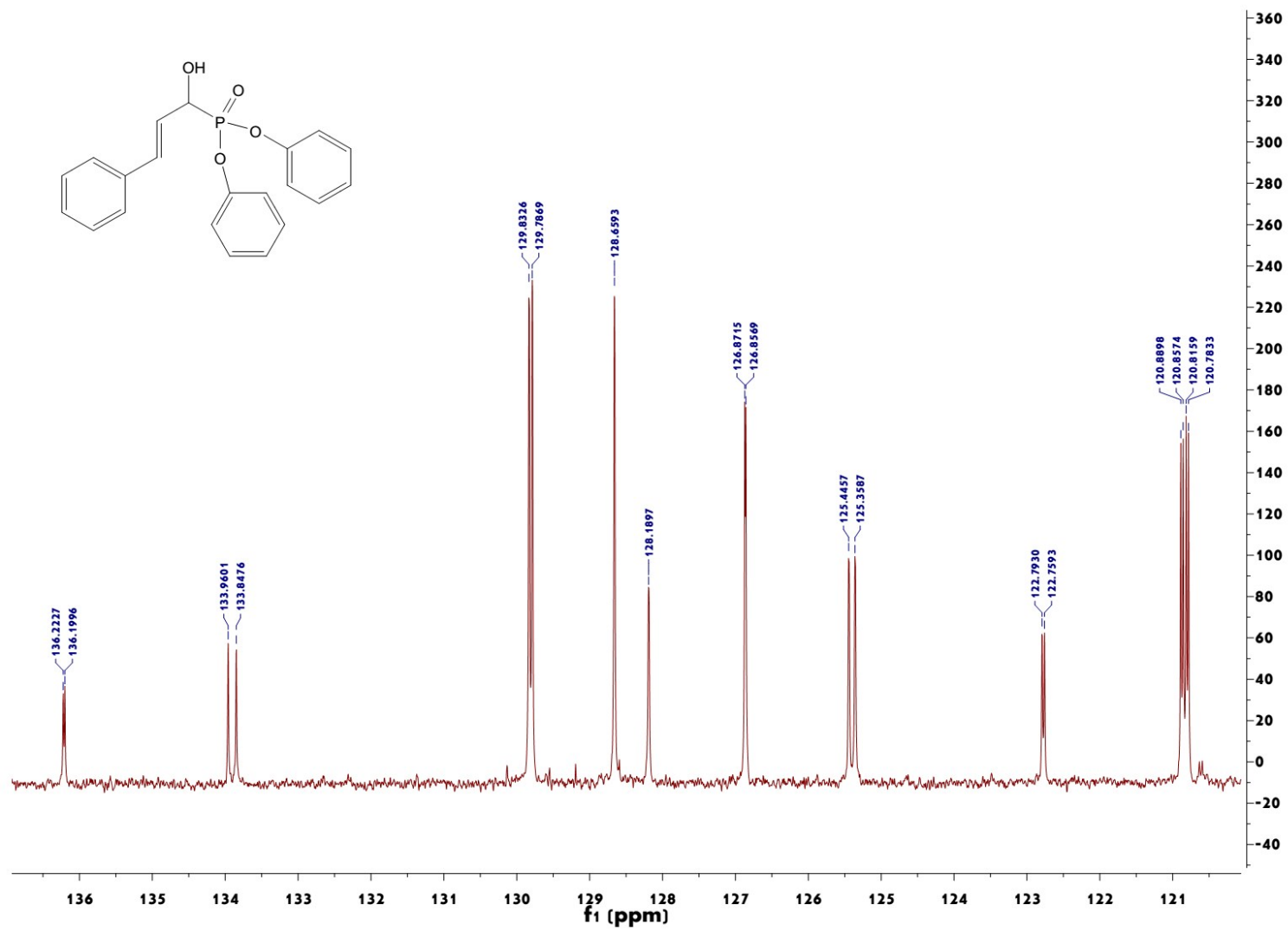
2m



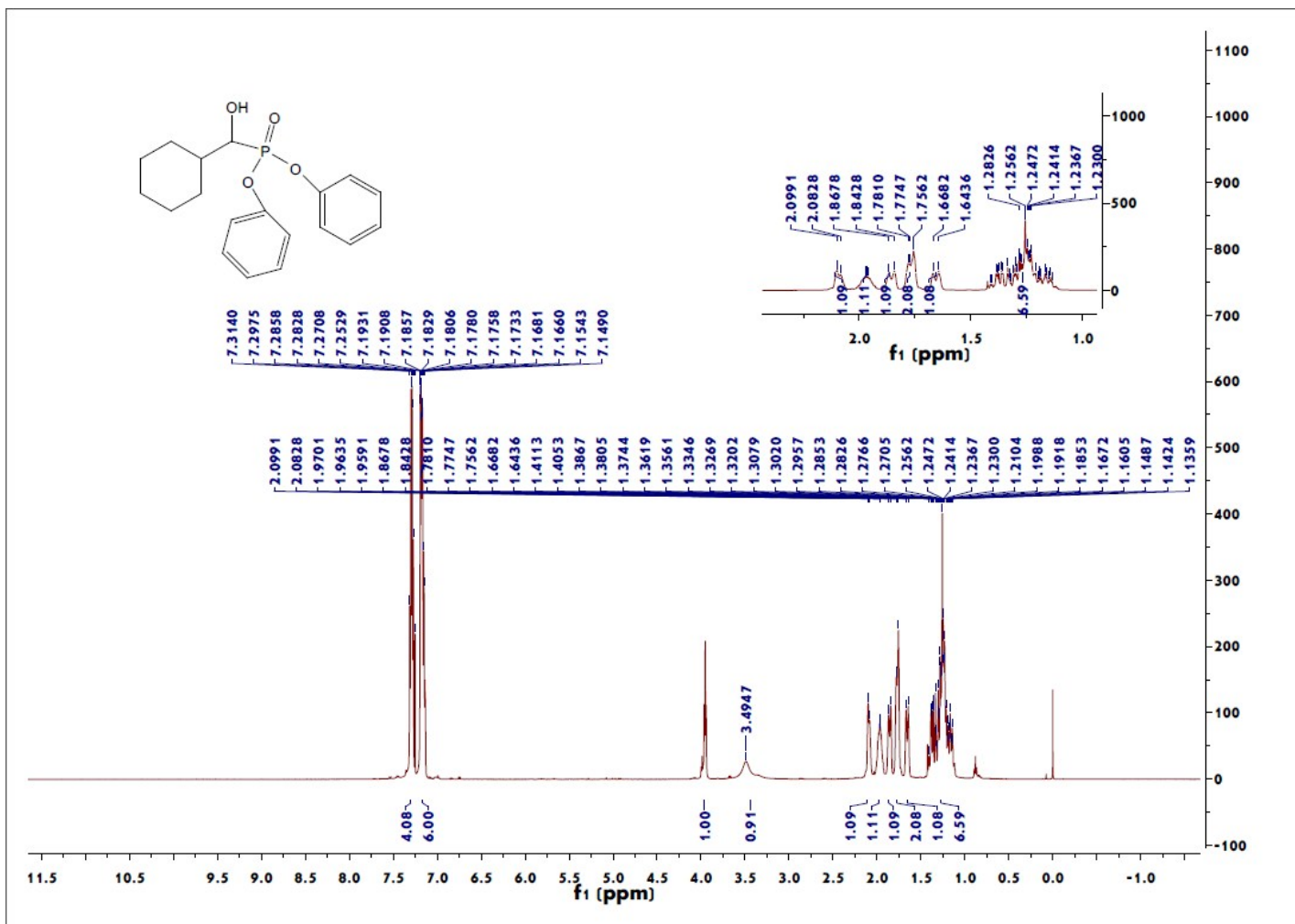
2m



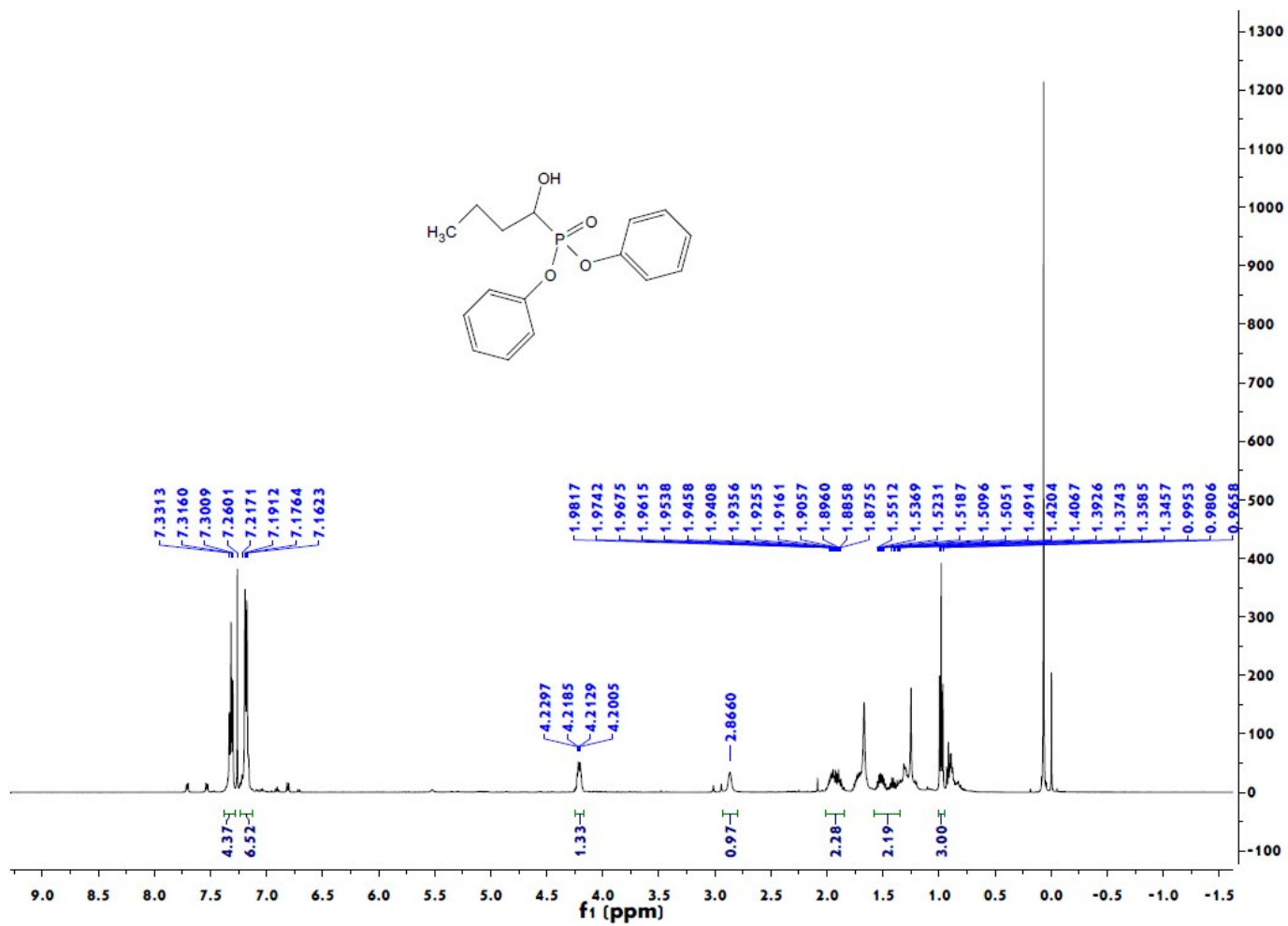
S52



2n

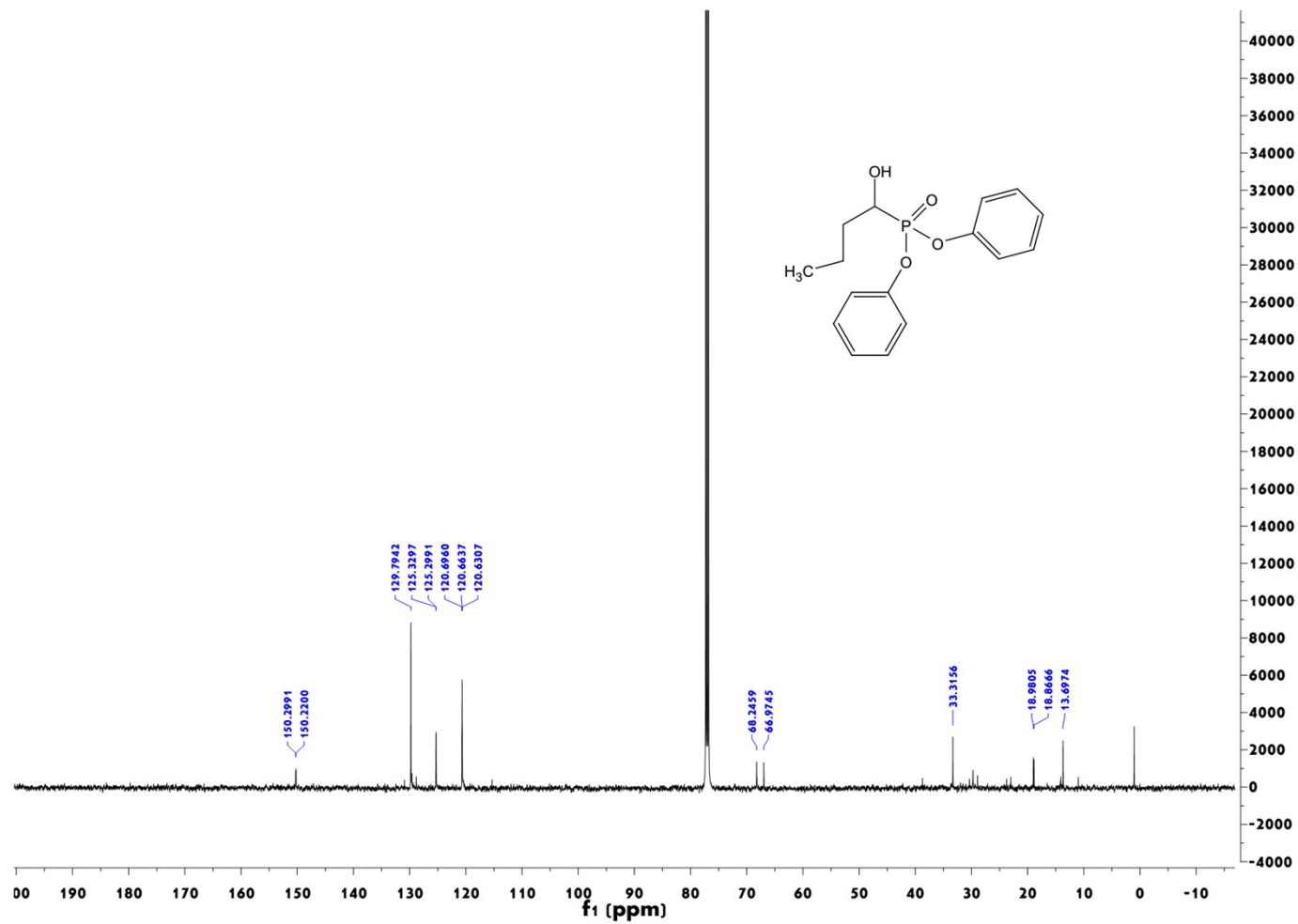


2o

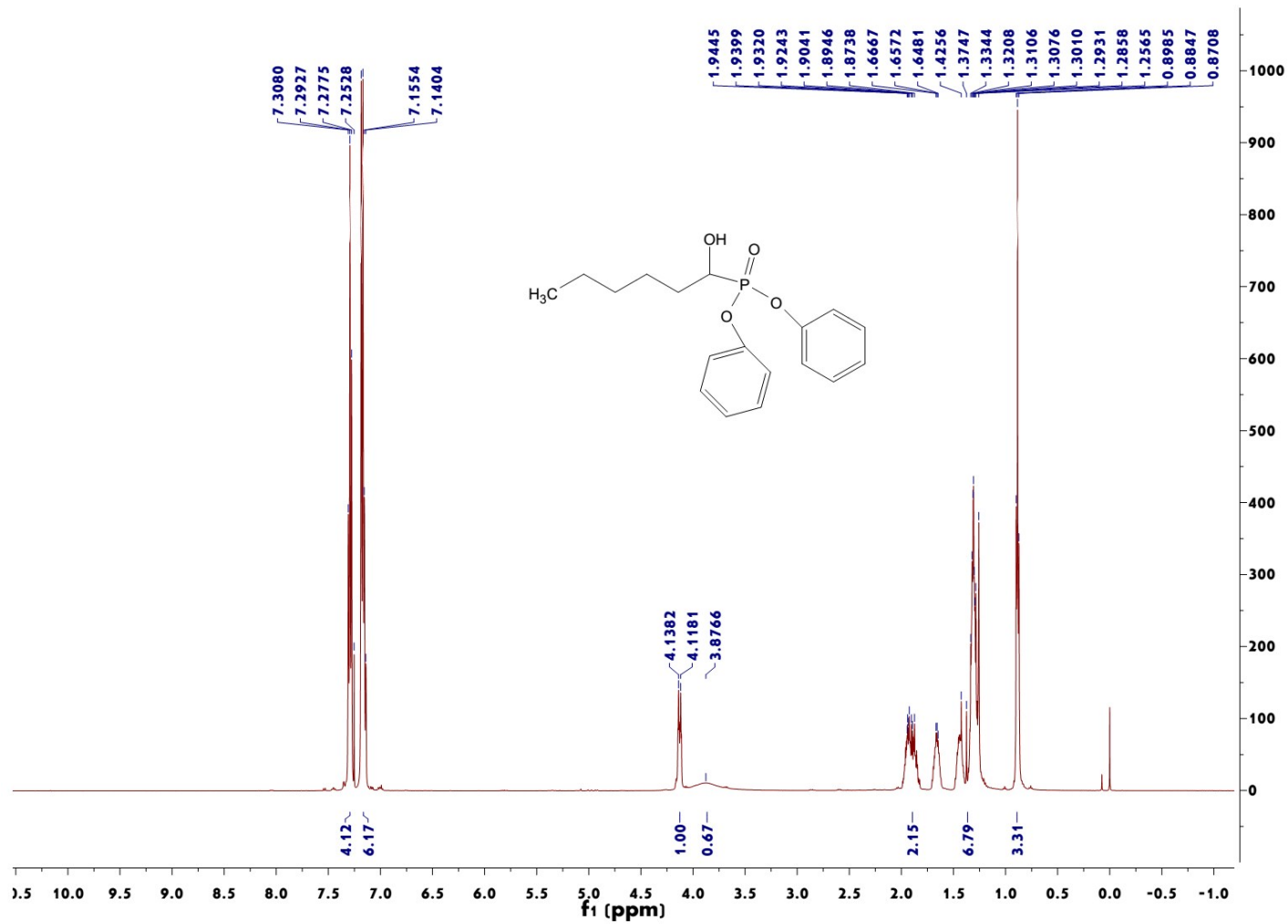


S55

2o



2p



2p

