Electronic Supplementary Material (ESI) for Organic & Biomolecular Chemistry. This journal is © The Royal Society of Chemistry 2016

## **Electronic Supplementary Information for:**

## A Radical Addition/Cyclization of Diverse Ethers to 2-Isocyanobiaryls under Mildly Basic Aqueous Conditions

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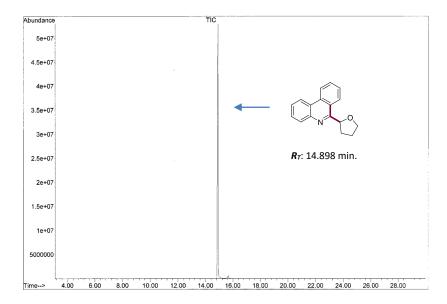
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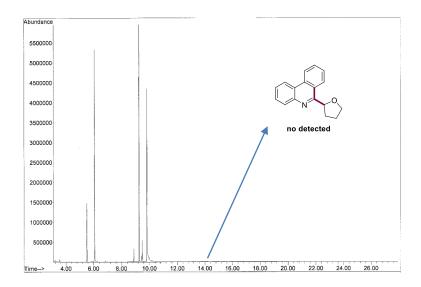
#### **Free Radical Capture Experiments**

#### ✓ Standard procedure

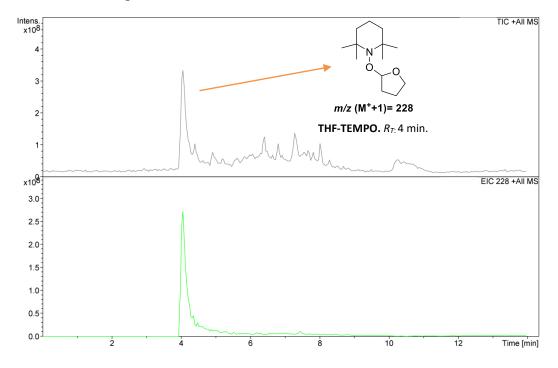
#### GC-MS of compound 3aa

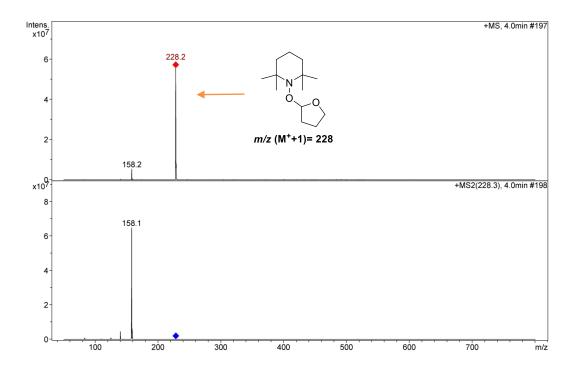


#### ✓ Standard procedure + TEMPO (3 equiv.)



## ✓ ESI-MASS (positive mode).



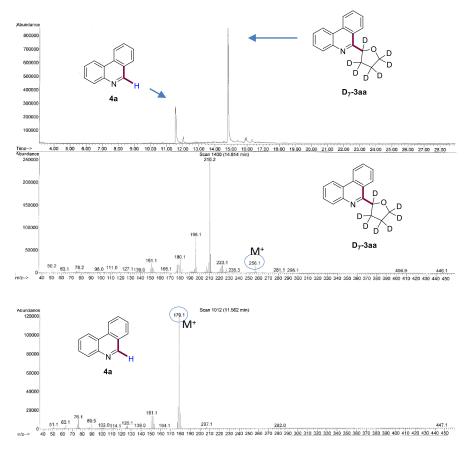


Into a pressure tube were added K<sub>2</sub>CO<sub>3</sub> (54 mg, 0.30 mmol) and D<sub>2</sub>O (0.5 mL). Then, were sequentially added 2-isocyanobiaryl **1a** (54 mg, 0.30 mmol), THF-D<sub>8</sub> (1.5 mL) and *tert*-butyl peroxibenzoate (TBPB) (128 μL, 0.70 mmol). The reaction mixture was stirred under Ar atmosphere for 14 h at 110 °C. After this time, a saturated solution of NaHCO<sub>3</sub> (5 mL) and EtOAc (10 mL) were added and the reaction mixture was extracted with EtOAc (3 x 15 mL). The organic layers were combined and washed with brine (5 mL), dried over MgSO<sub>4</sub>, and concentrated under reduced pressure. The residue was purified by flash chromatography (Hex/EtOAc, 95:5 to 93:7) to afford product **D**7-**3aa** as a white solid (45 mg, 59 %) and compound **4a** as a yellow solid (11 mg, 21 %).

standard procedure
using 
$$D_2O / D_8$$
-THF
$$D_D D$$

$$D_D D$$
1a
$$D_7$$
-3aa, 59%
$$D_7$$
-3aa, 59%
$$D_7$$
-3aa, 59%

Analyzing the GC-MS of the crude reaction mixture, it was possible to confirm that compound **4a** was not deuterated.



# Competition Experiments of 1a with an Equimolar Mixture of THF/D<sub>8</sub>-THF to Examine the Intermolecular Kinetic Isotopic Effect (KIE).

In a pressure tube, the mixture of **1a** (0.30 mmol), **THF/D<sub>8</sub>-THF** (1:1, 1.5 mL) was treated under standard conditions for 1.5 h. The reaction mixture was quenched with NaHCO<sub>3</sub> (5 mL) and was extracted with EtOAc (3 x 15 mL). The organic layers were combined and washed with brine (5 mL), dried over MgSO<sub>4</sub> and concentrated under reduce pressure. The residue was purified by flash column chromatography on silica gel (95:5 to 93:7, hexane/EtOAc) to separate the excess of **1a**, and the mixture of products **3aa/D<sub>7</sub>-3aa** was collected (combined yield: 19%). Using <sup>1</sup>H-NMR, it was found a 76:24 ratio of **3aa/D<sub>7</sub>-3aa**, as shown in **Figure S1**.

$$\begin{array}{c}
\text{standard} \\
\text{procedure} \\
\text{1.5 h} \\
\text{19% yield}
\end{array}$$

$$\begin{array}{c}
\text{3aa / D}_{7}\text{-3aa} = 76:24$$

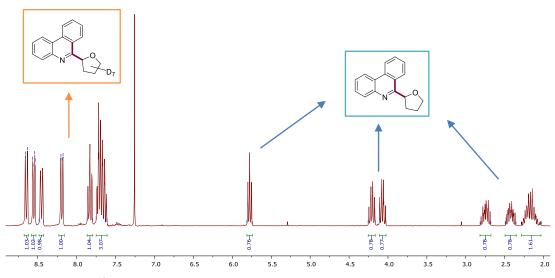
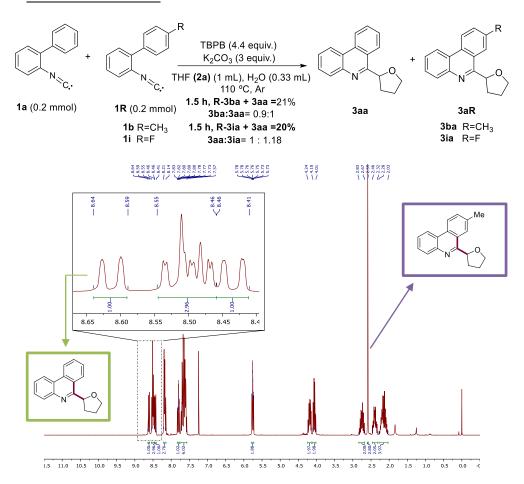


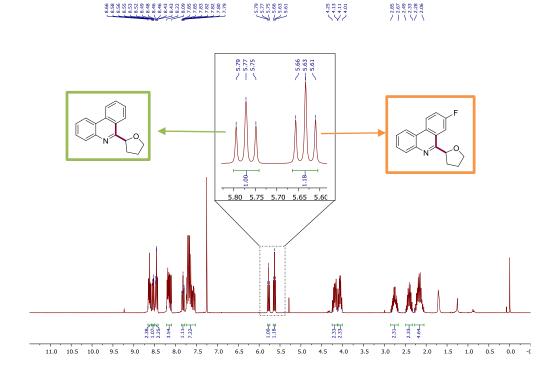
Figure S1. <sup>1</sup>H-NMR (300 MHz, CDCl<sub>3</sub>) of the mixture 3aa/ D<sub>7</sub>-3aa isolated as describe above.

#### Competition Experiments of THF with an Equimolar Mixture of 1a/1b or 1a/1i

#### ✓ Substrates 1a and 1b

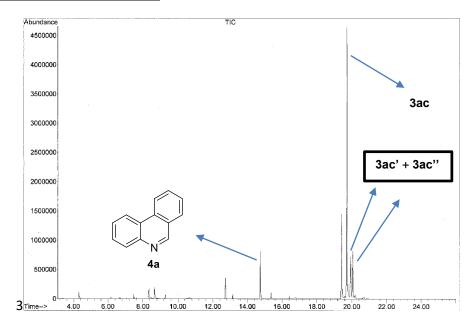


#### ✓ Substrates 1a and 1i

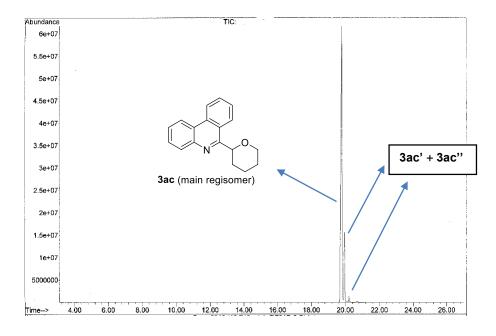


#### Detection of regioisomers upon formation of 3ac by GC-MS

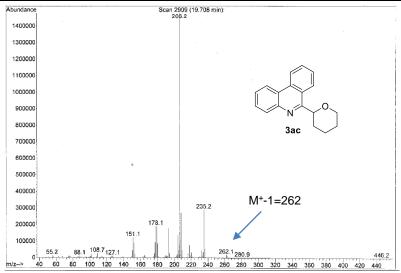
#### GC (MS) of crude reaction mixture:

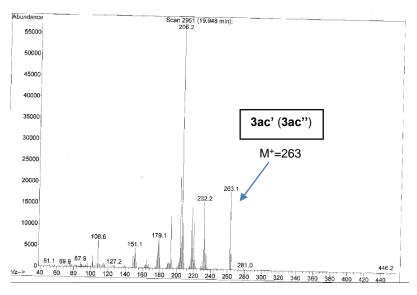


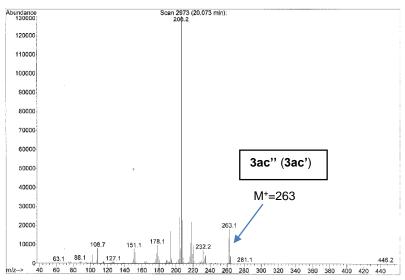
#### GC (MS) of **3ac** after column purification:



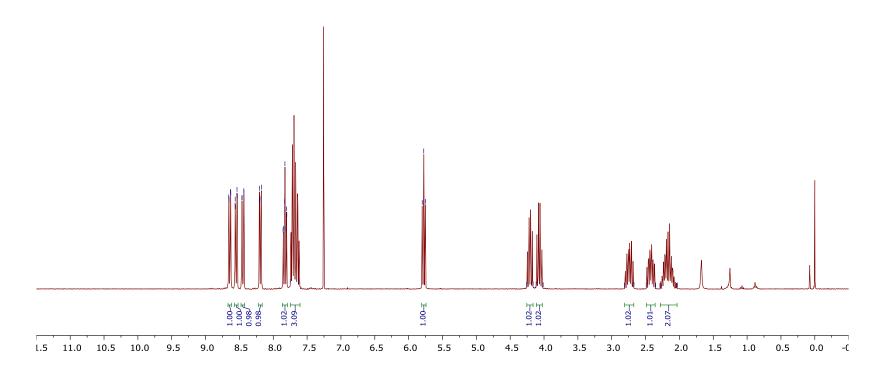
#### MS for selected GC peaks in the formation of **3ac** (**3ac** and regioisomers):



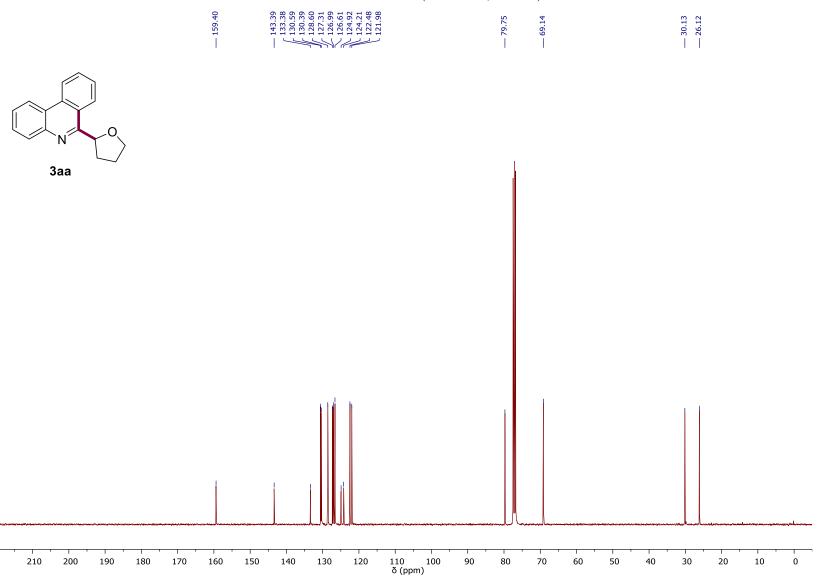


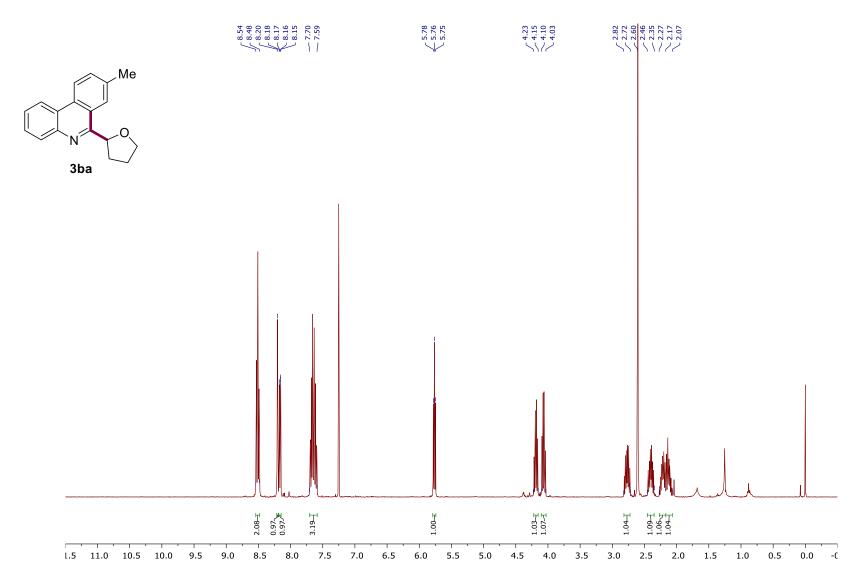


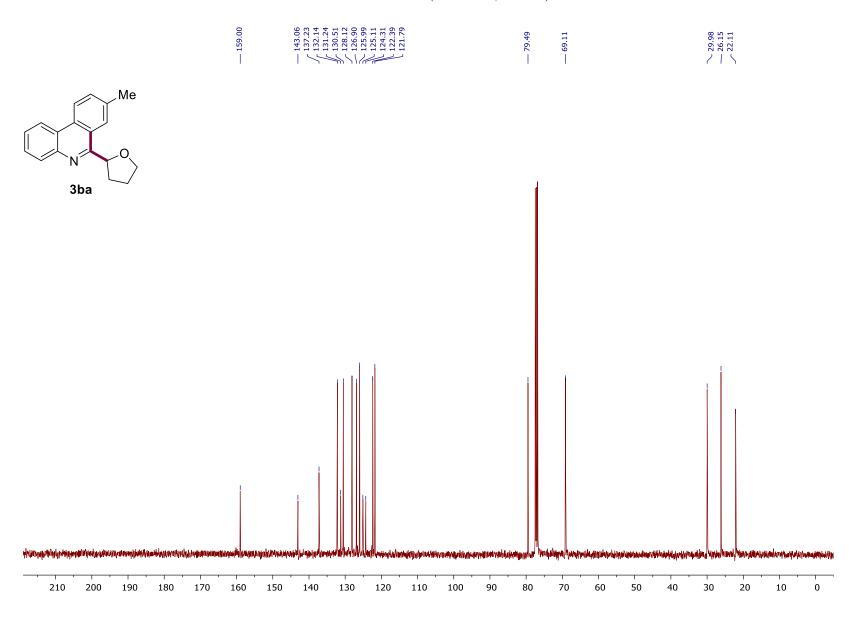


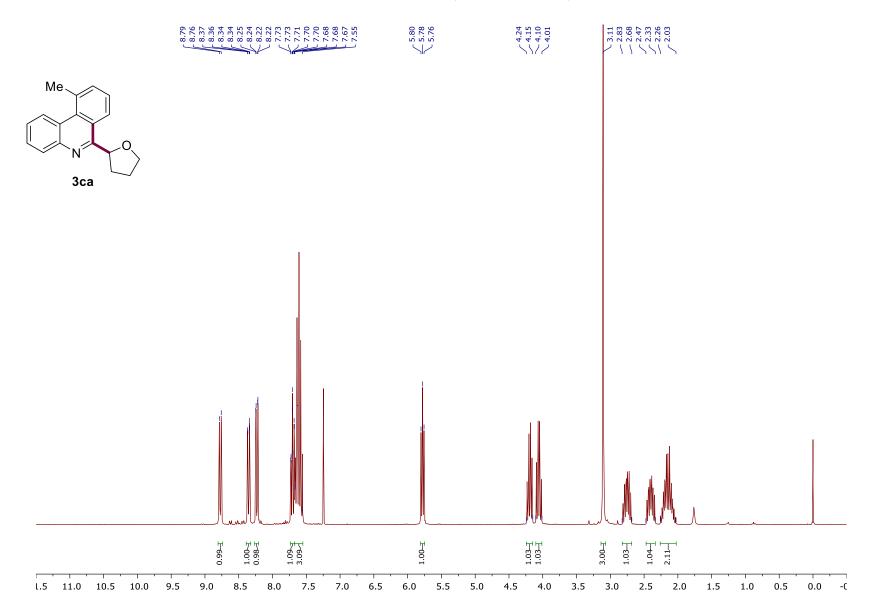




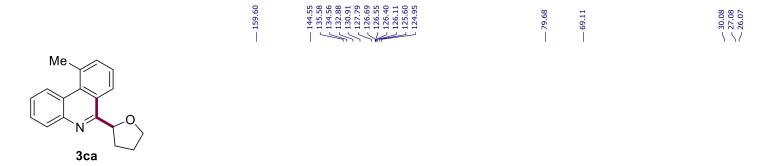


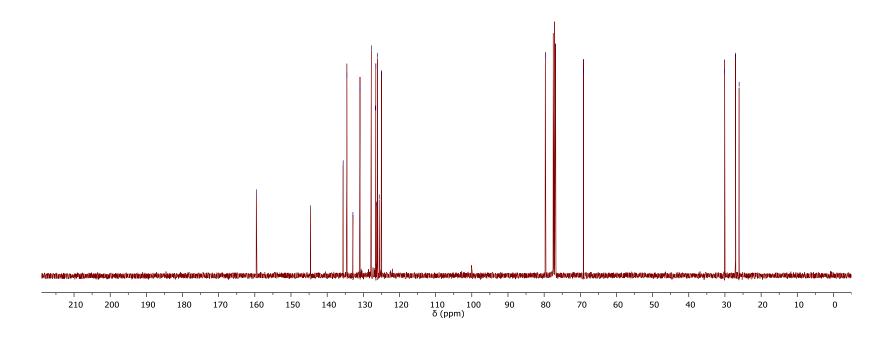


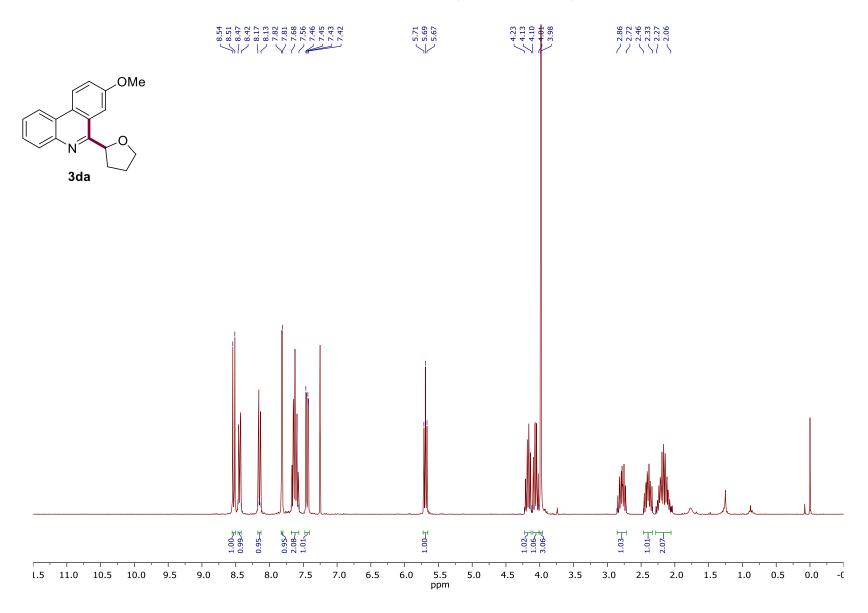




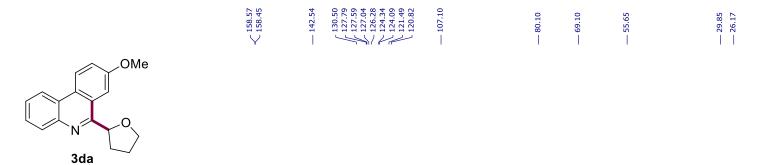
<sup>13</sup>C-NMR (101 MHz, CDCl<sub>3</sub>)

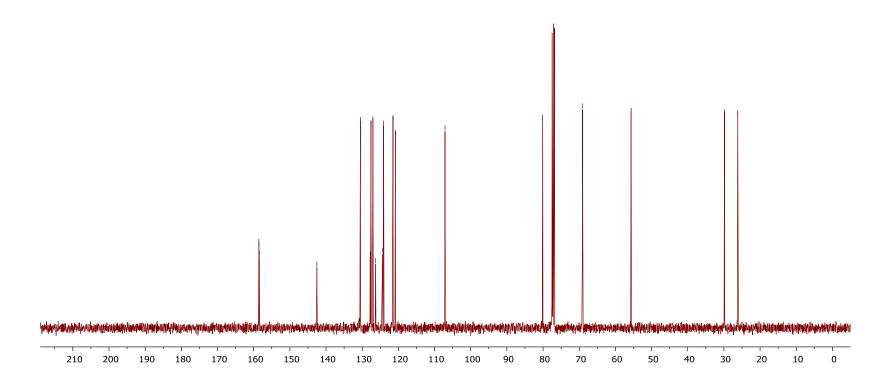


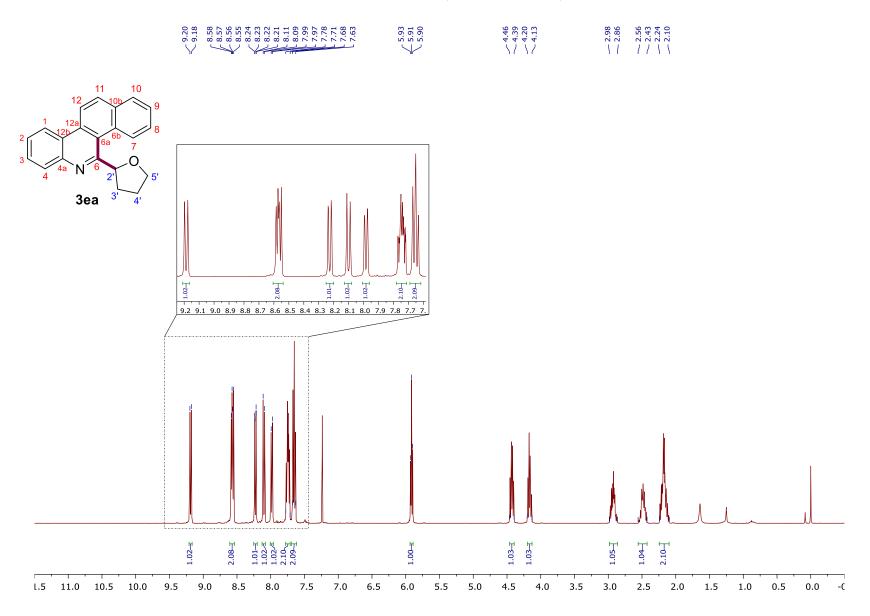




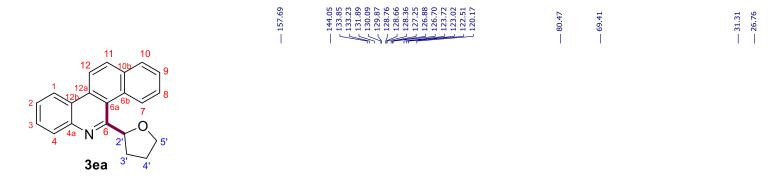
<sup>13</sup>C-NMR (101 MHz, CDCl<sub>3</sub>)

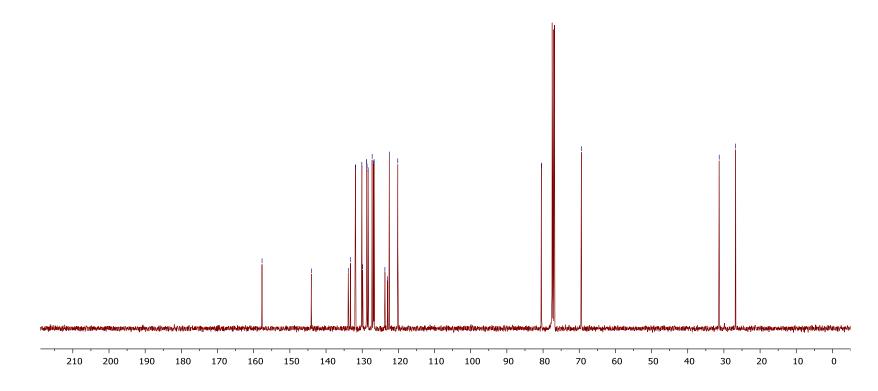


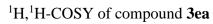


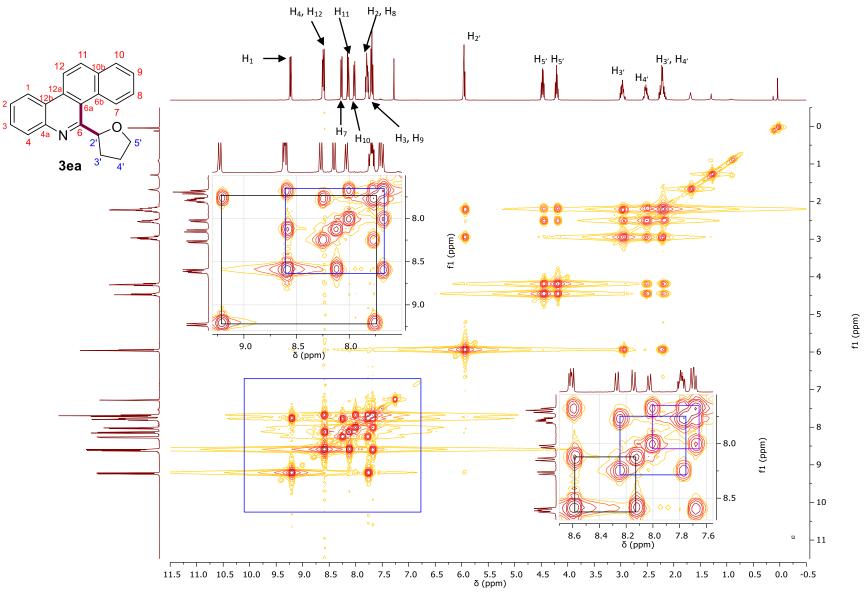


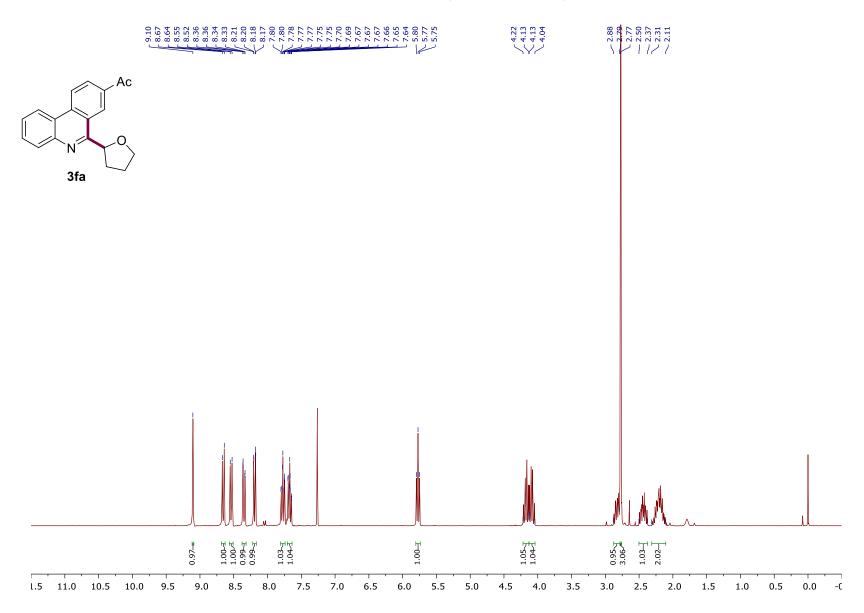
<sup>13</sup>C-NMR (101 MHz, CDCl<sub>3</sub>)



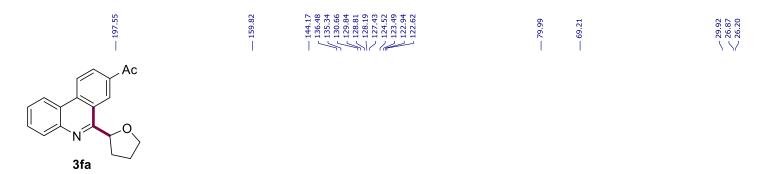


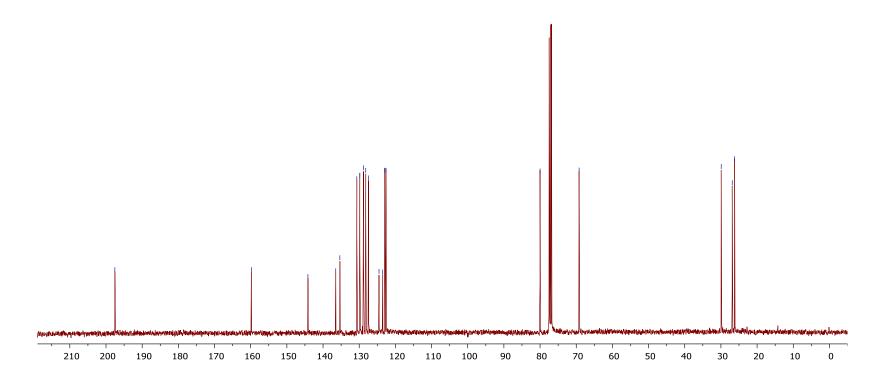






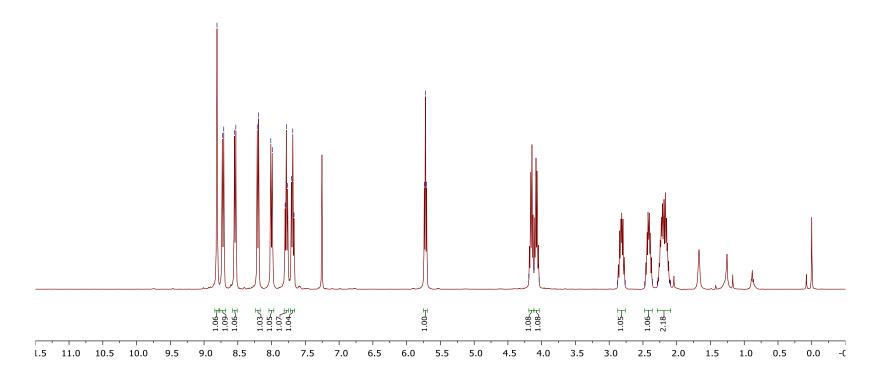
<sup>13</sup>C-NMR (101 MHz, CDCl<sub>3</sub>)

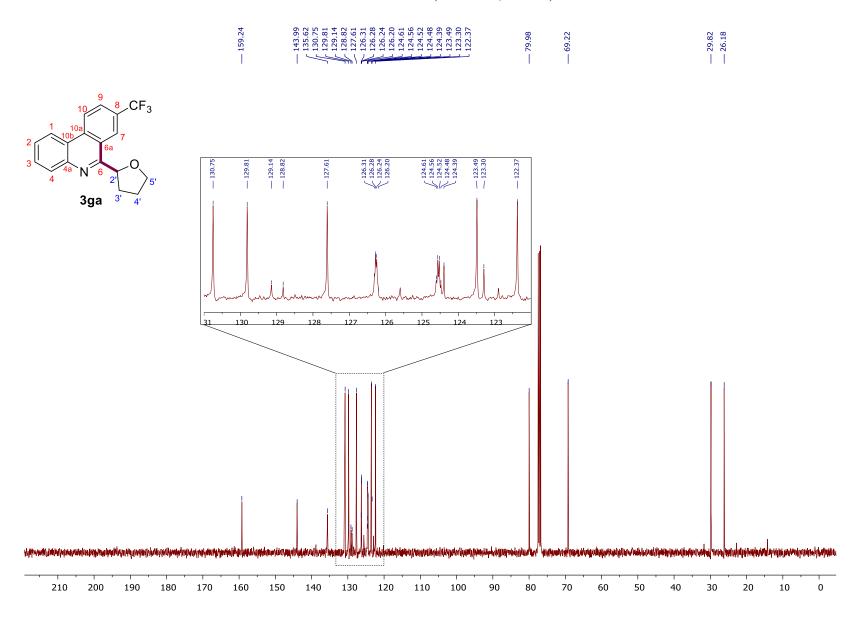


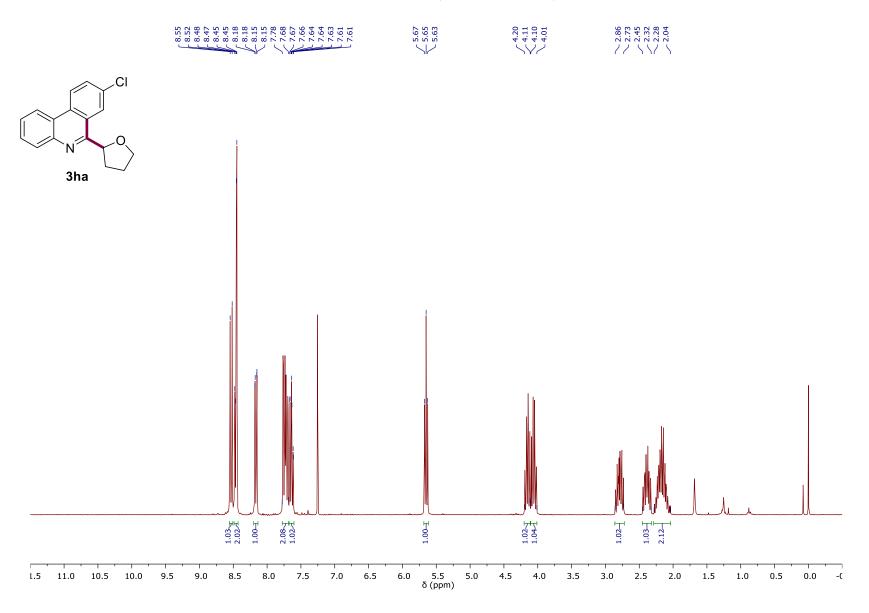




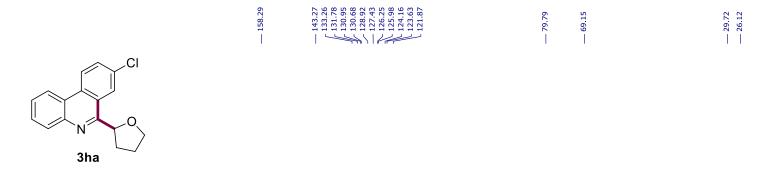


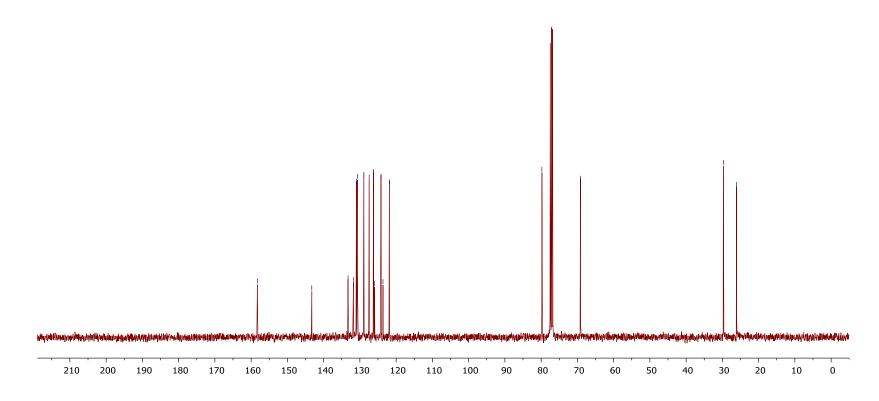






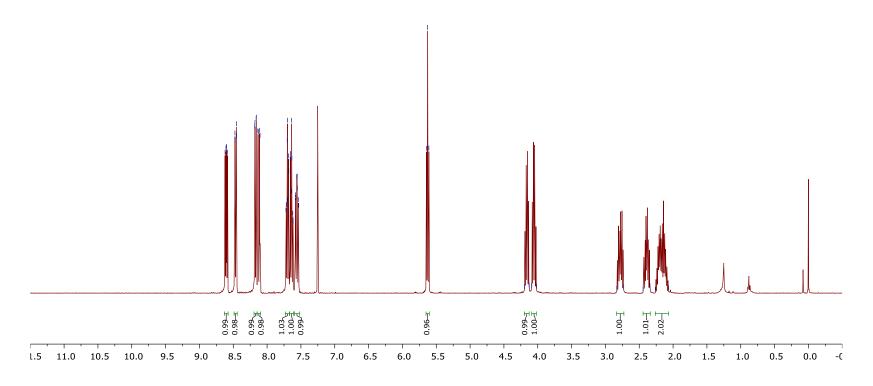
<sup>13</sup>C-NMR (101 MHz, CDCl<sub>3</sub>)

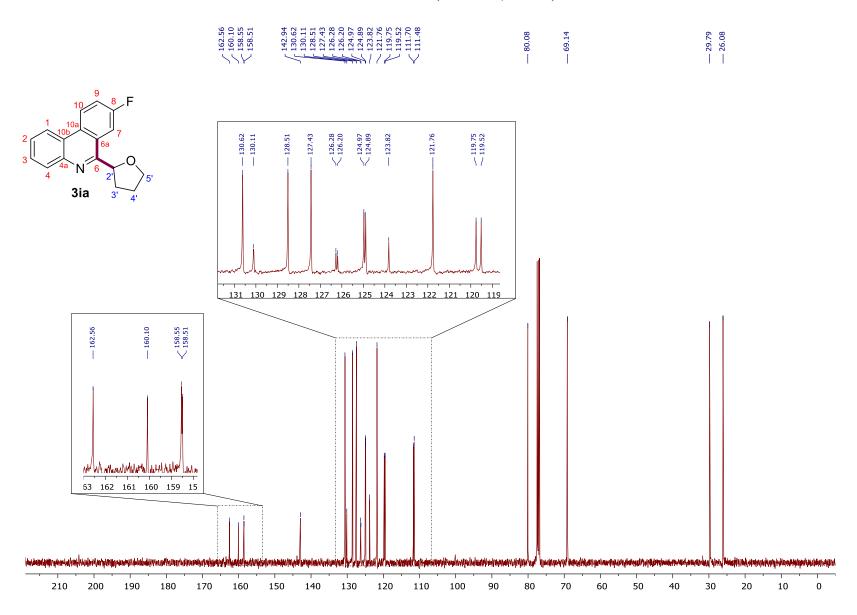


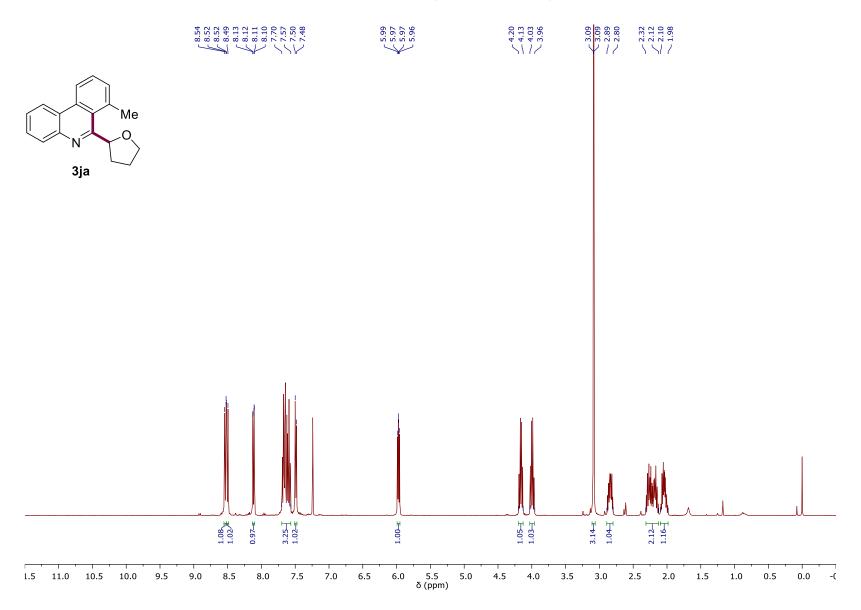




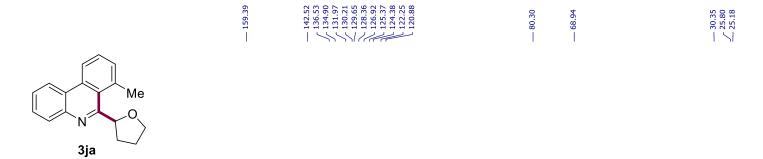


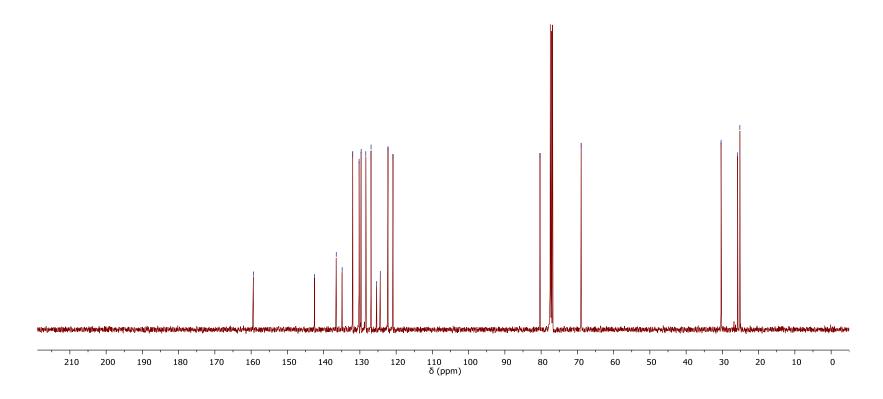


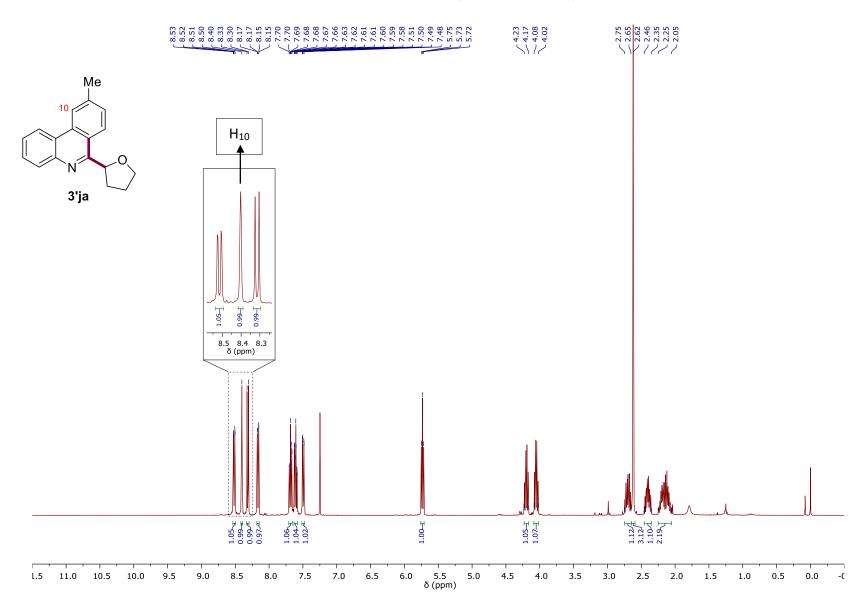




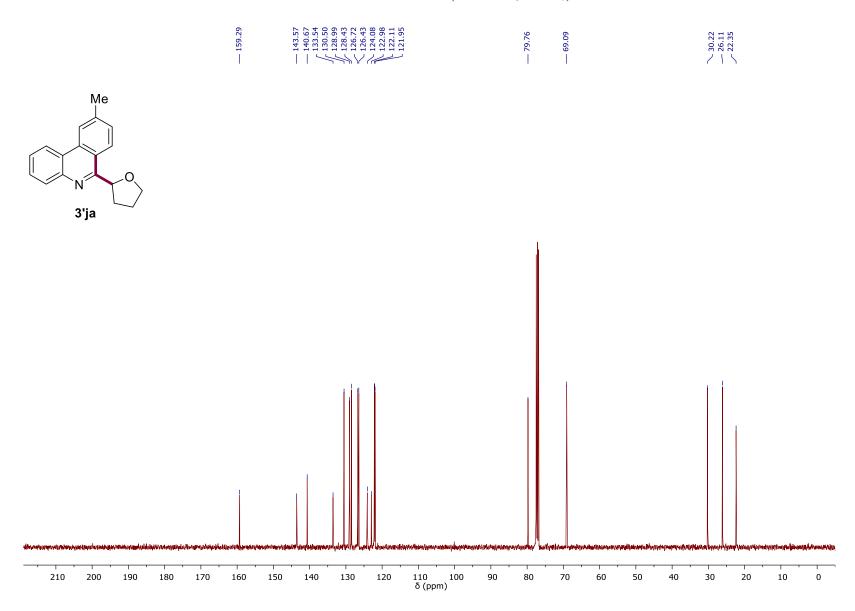
<sup>13</sup>C-NMR (101 MHz, CDCl<sub>3</sub>)

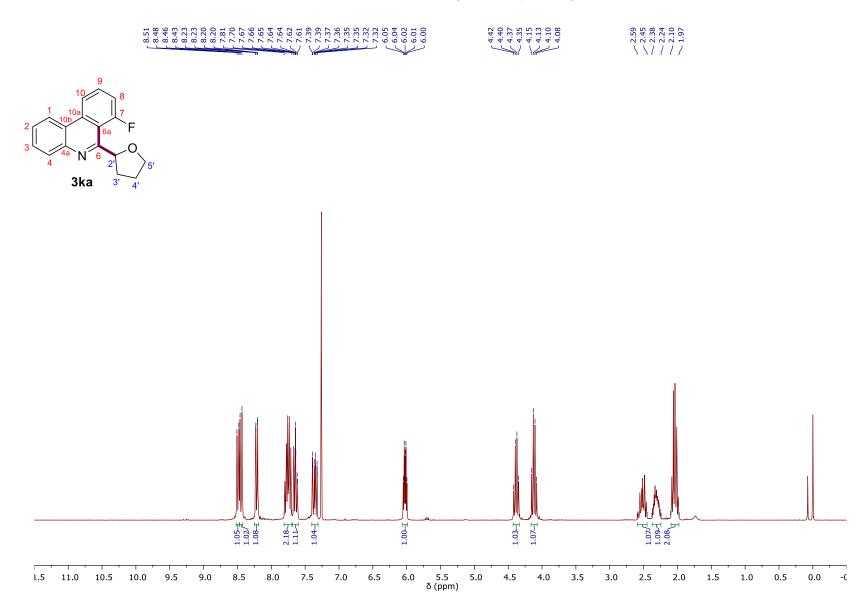


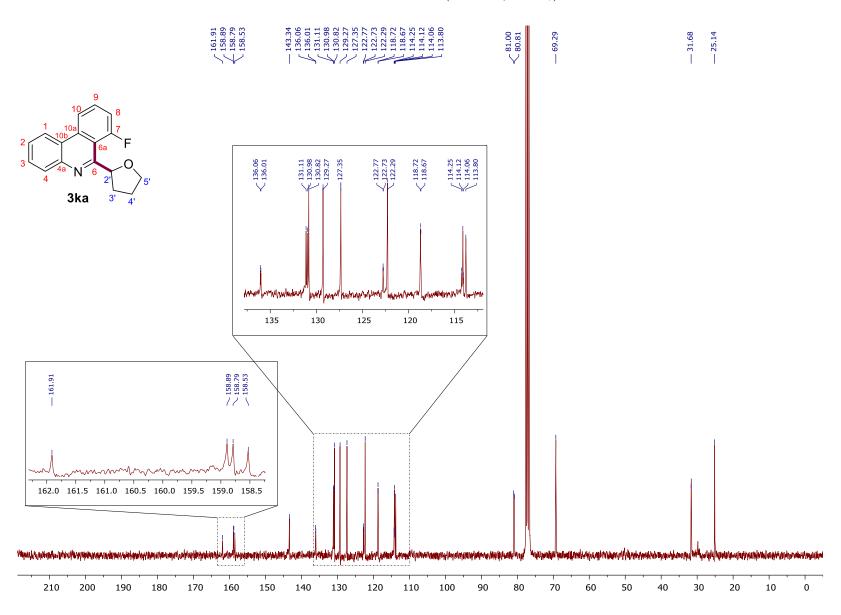


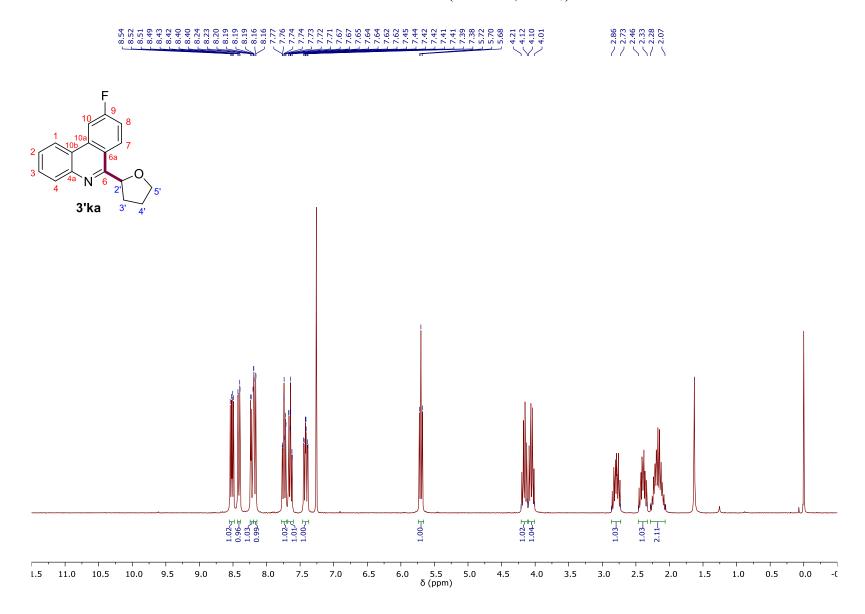


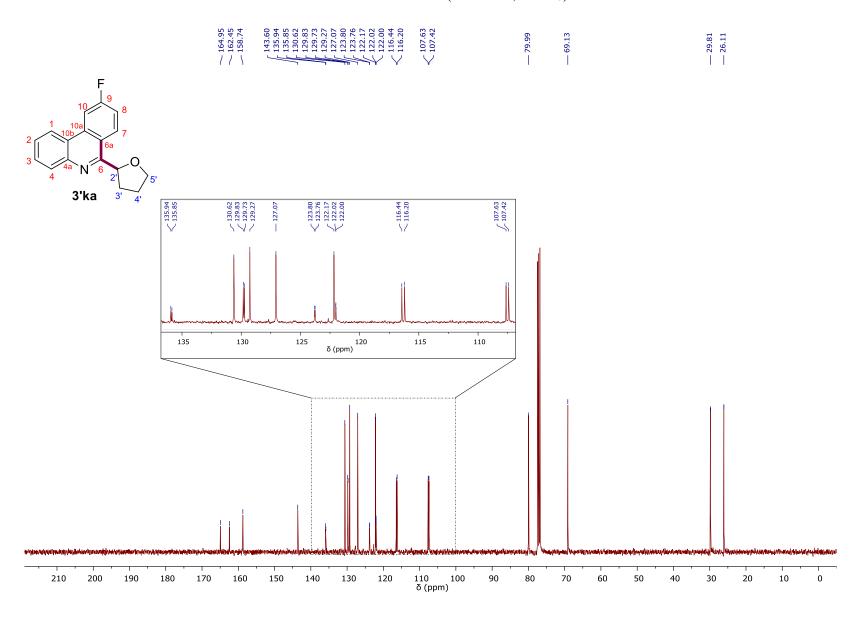
<sup>13</sup>C-NMR (101 MHz, CDCl<sub>3</sub>)



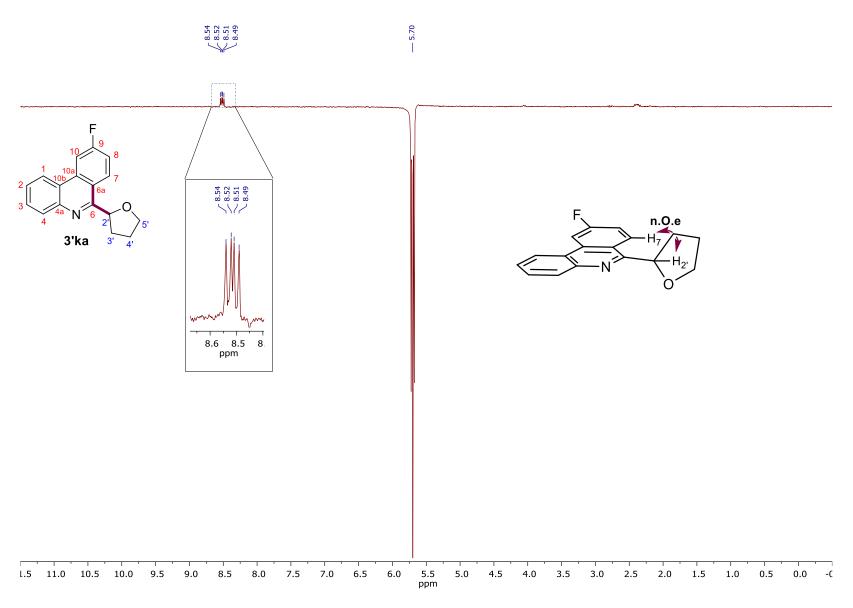


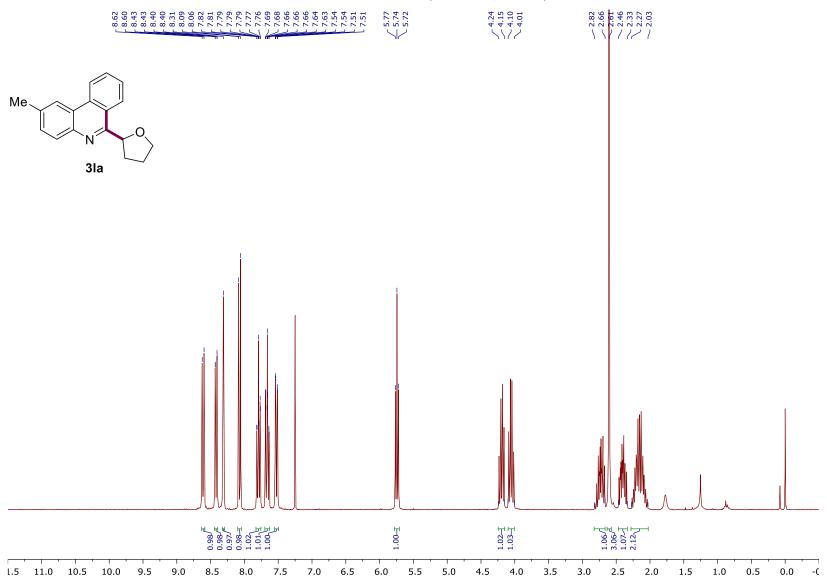






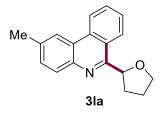
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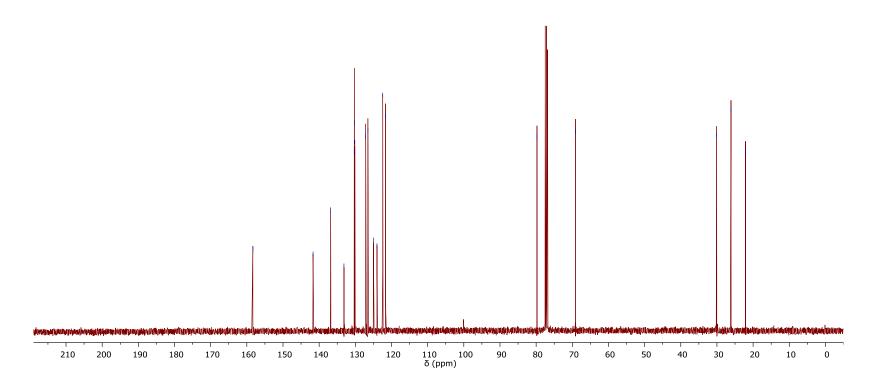


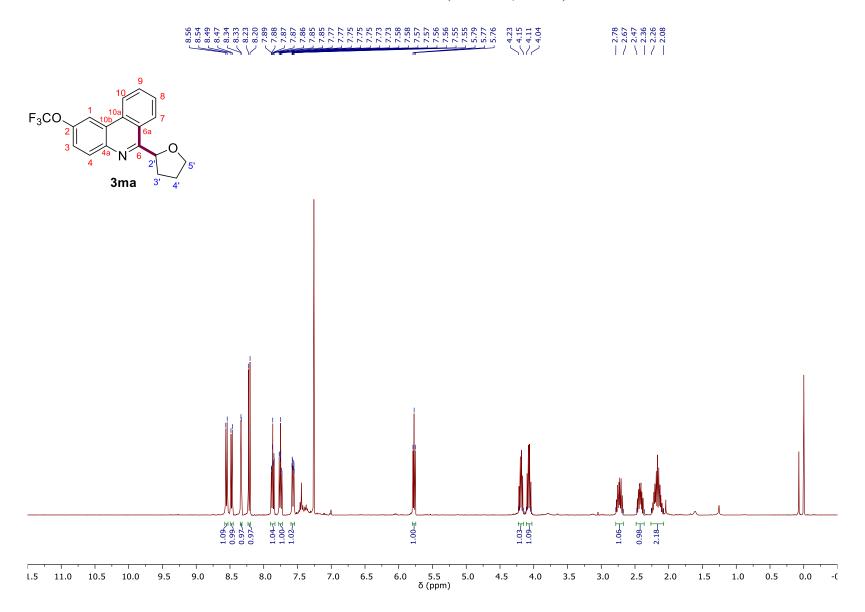


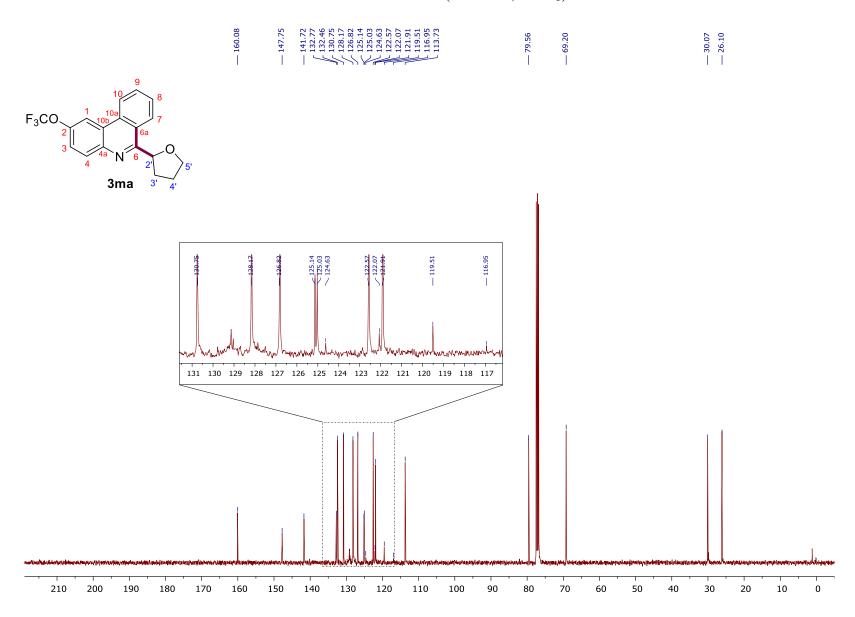
<sup>13</sup>C-NMR (101 MHz, CDCl<sub>3</sub>)





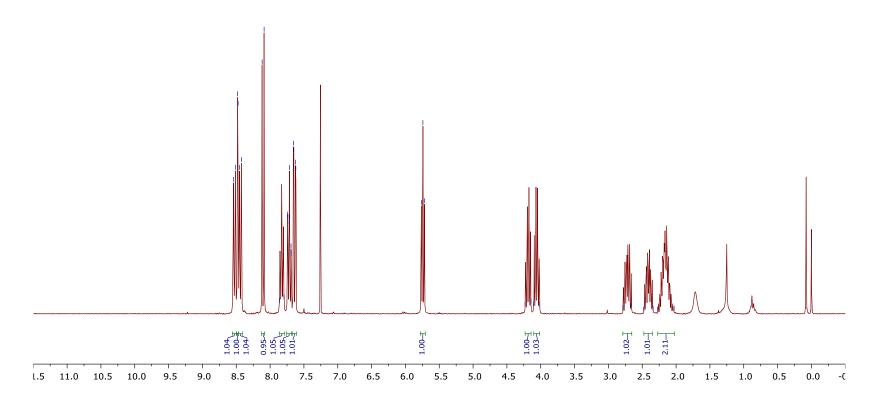




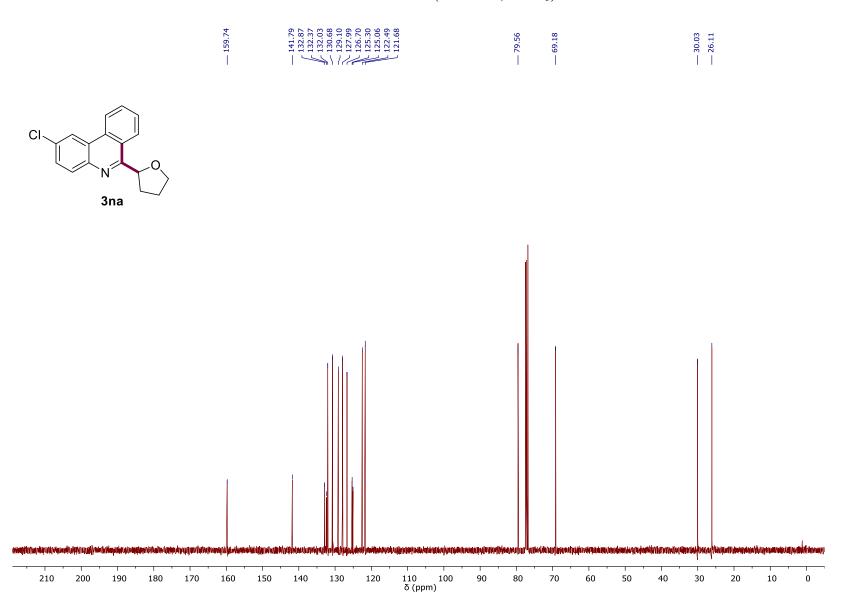








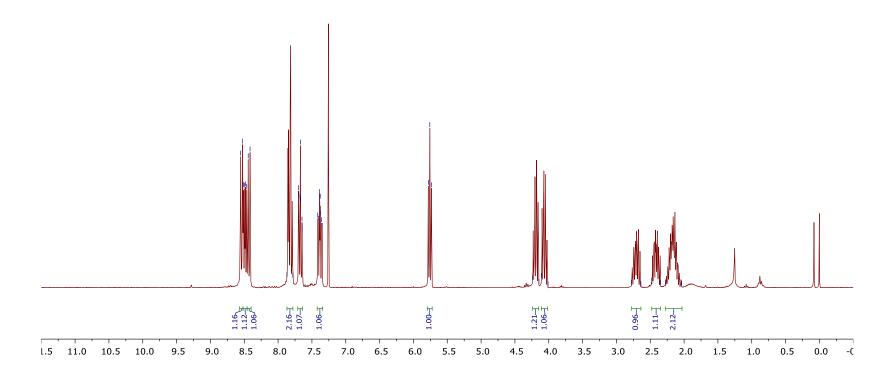
<sup>13</sup>C-NMR (101 MHz, CDCl<sub>3</sub>)



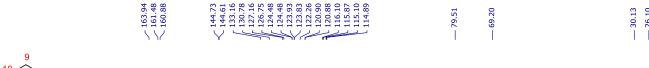
<sup>1</sup>H-NMR (300 MHz, CDCl<sub>3</sub>)



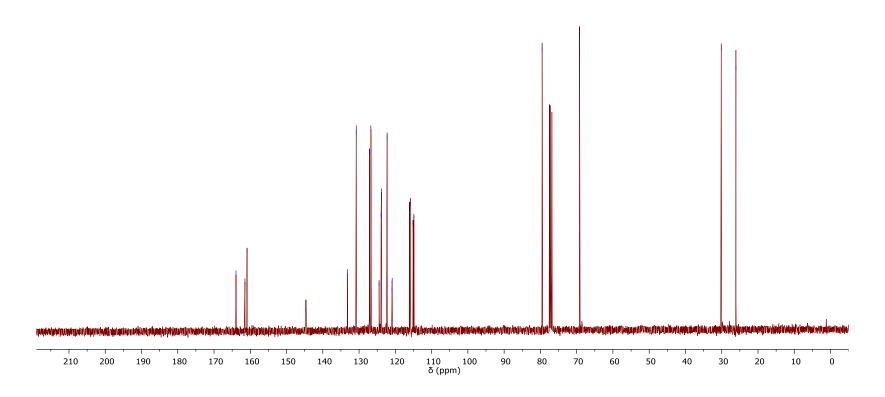


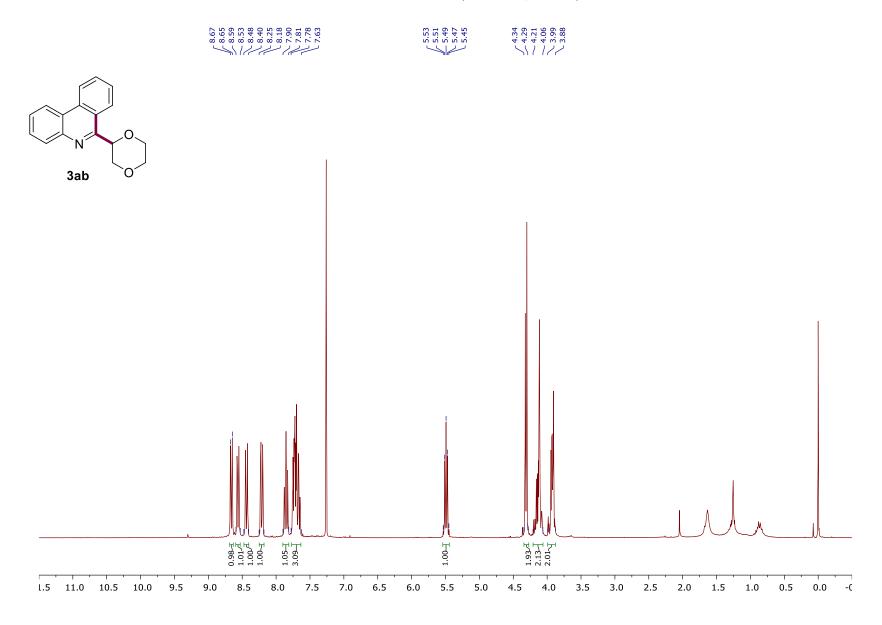


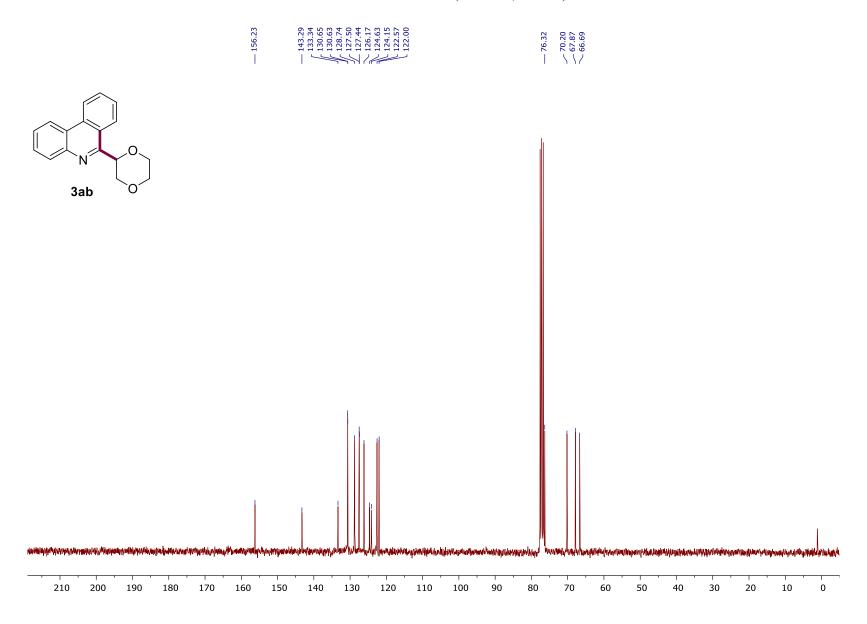
<sup>13</sup>C-NMR (101 MHz, CDCl<sub>3</sub>)



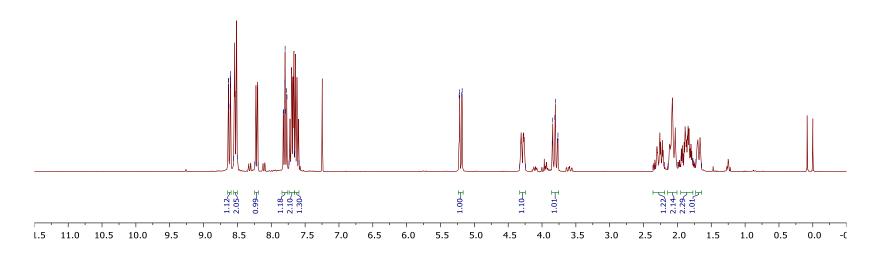






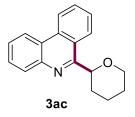


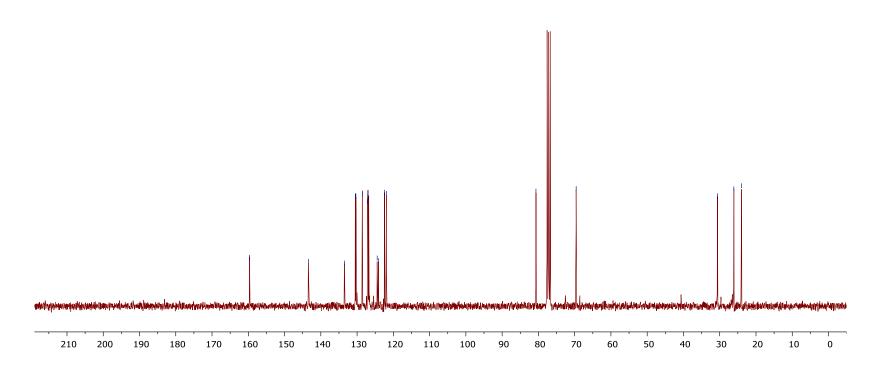


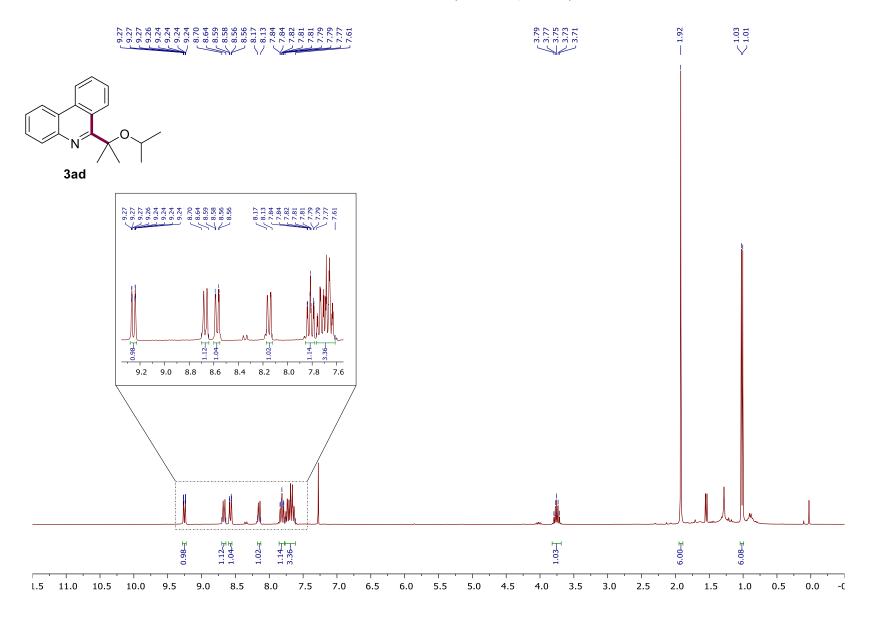


<sup>13</sup>C-NMR (75 MHz, CDCl<sub>3</sub>)

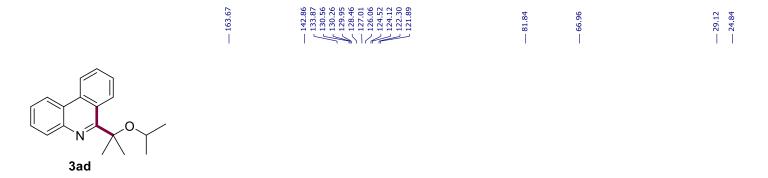


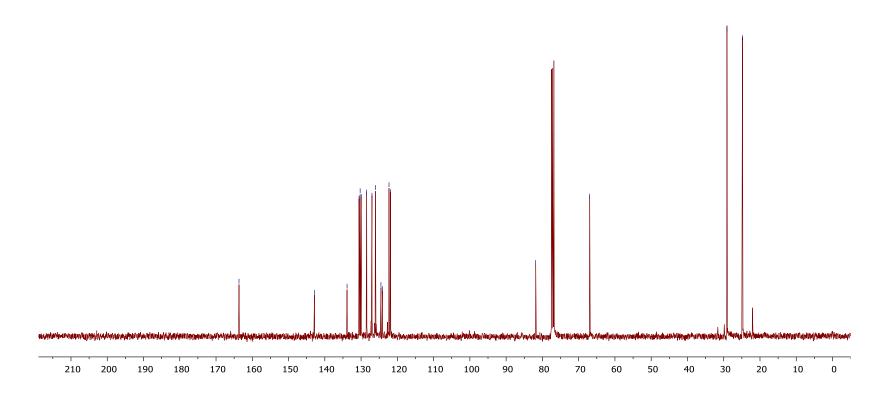


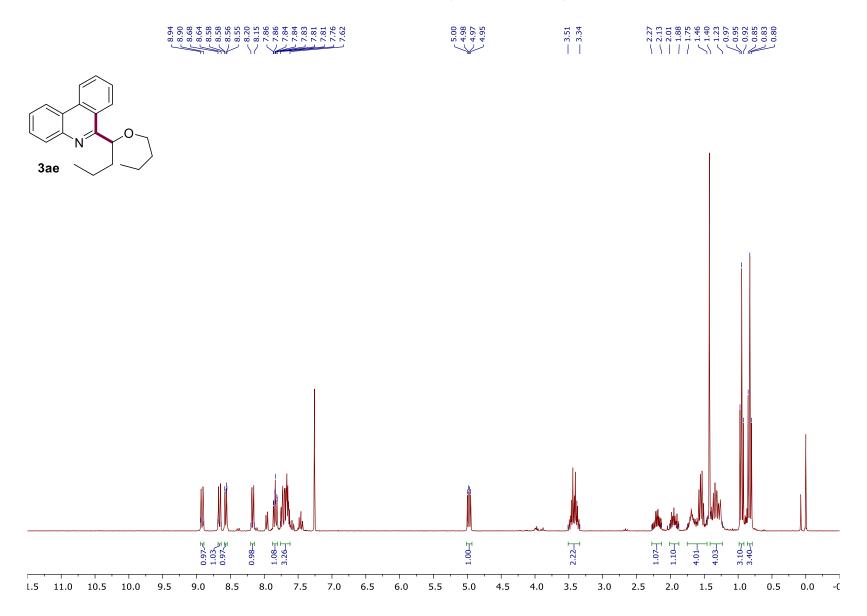


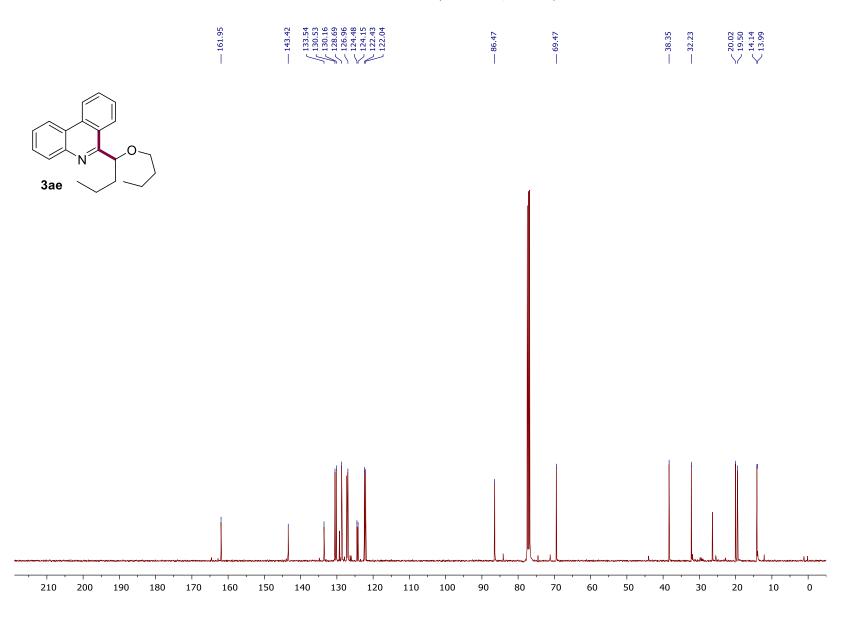


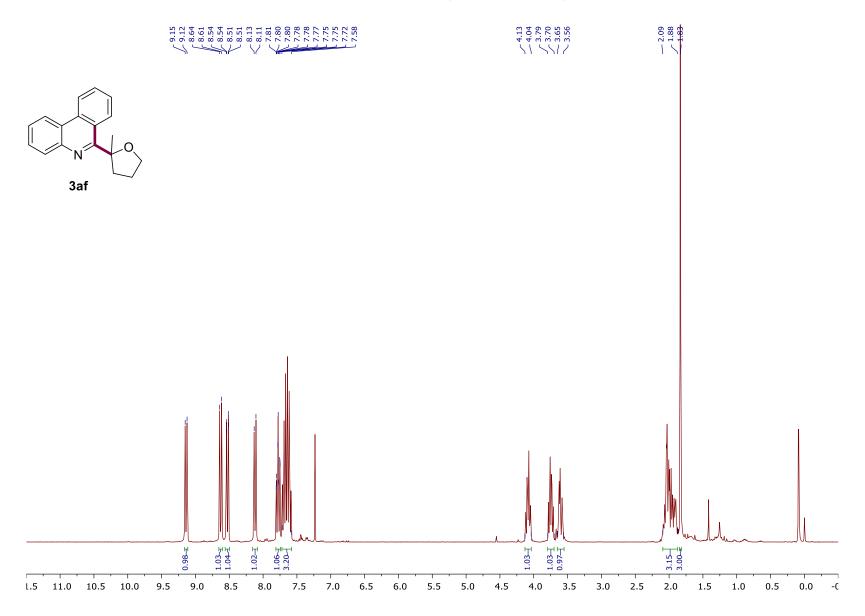
<sup>13</sup>C-NMR (101 MHz, CDCl<sub>3</sub>)



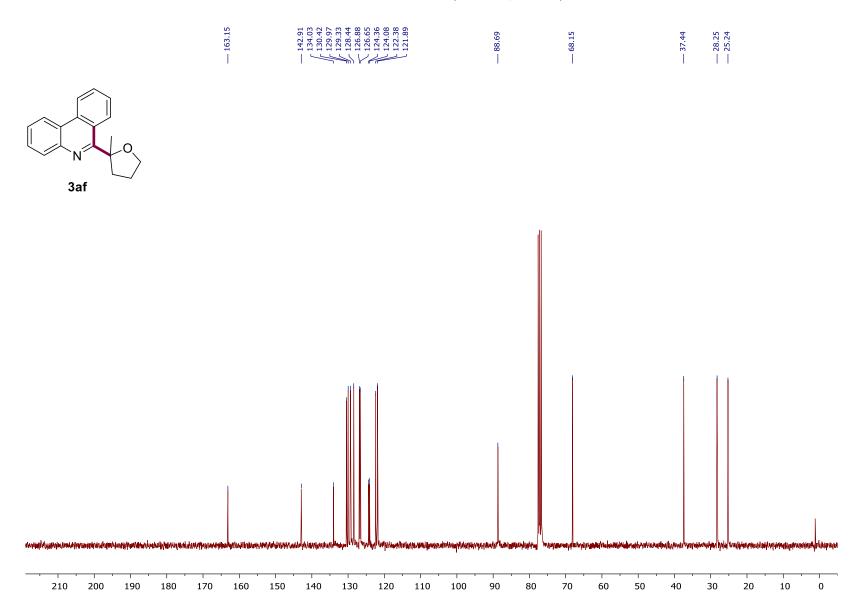


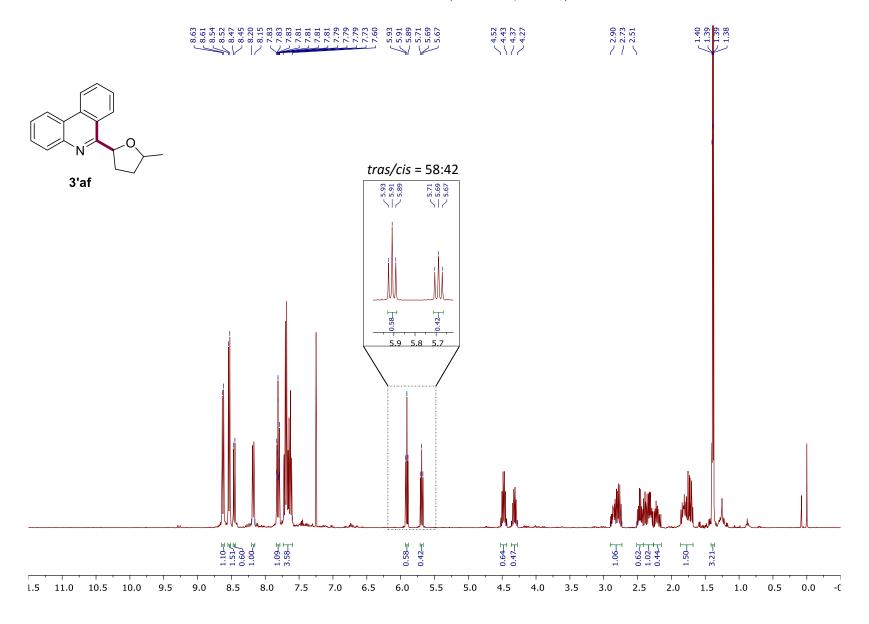




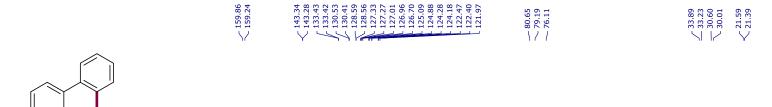


<sup>13</sup>C-NMR (75 MHz, CDCl<sub>3</sub>)

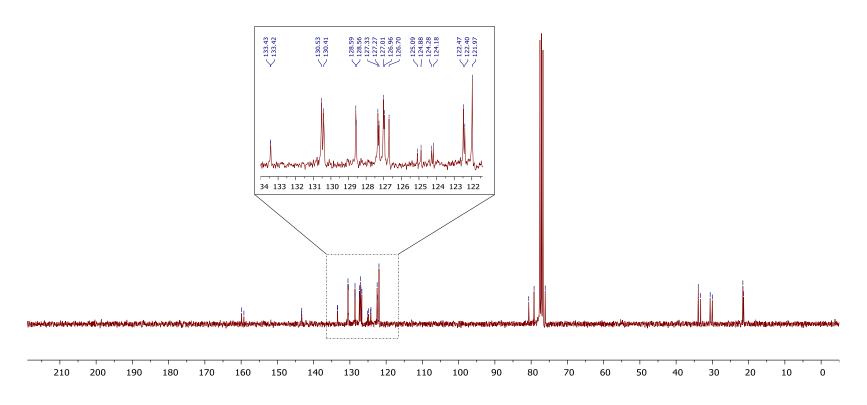


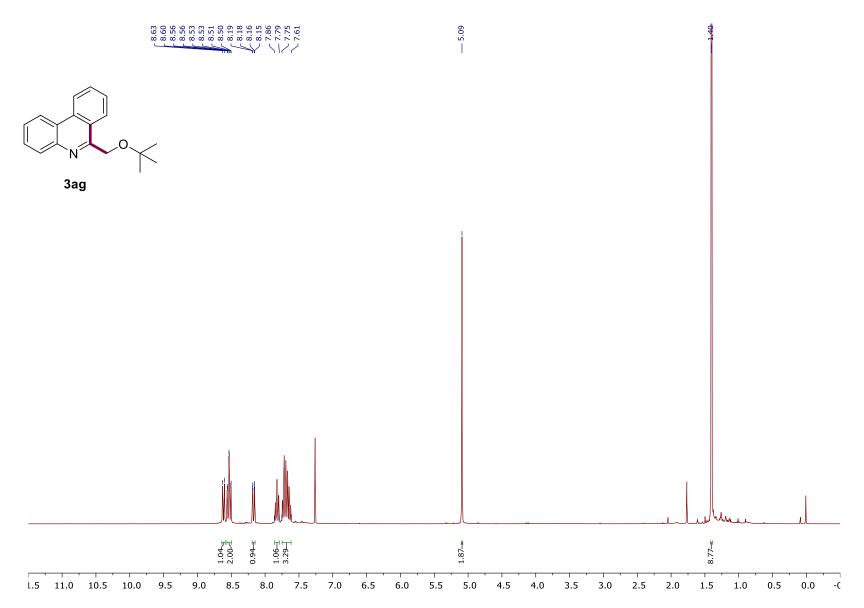


<sup>13</sup>C-NMR (75 MHz, CDCl<sub>3</sub>)

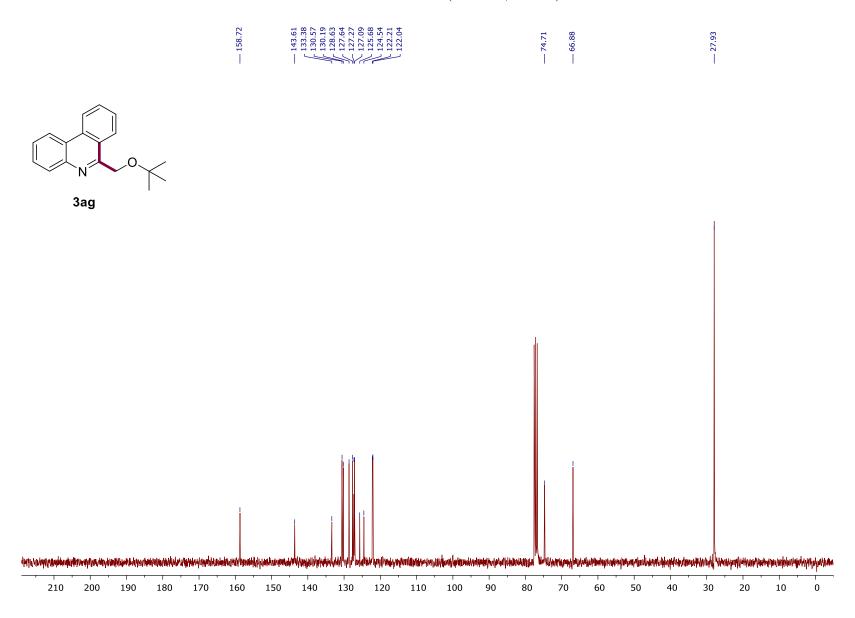


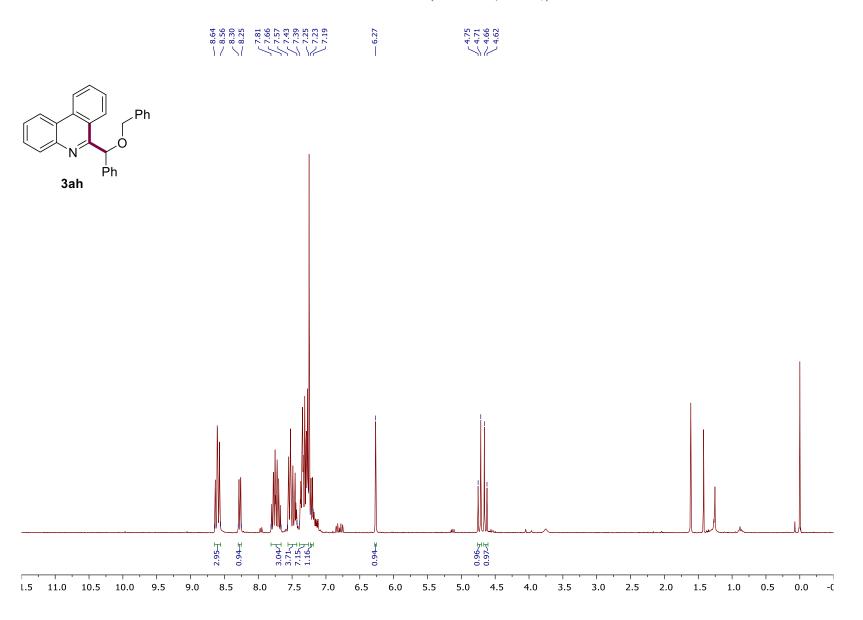




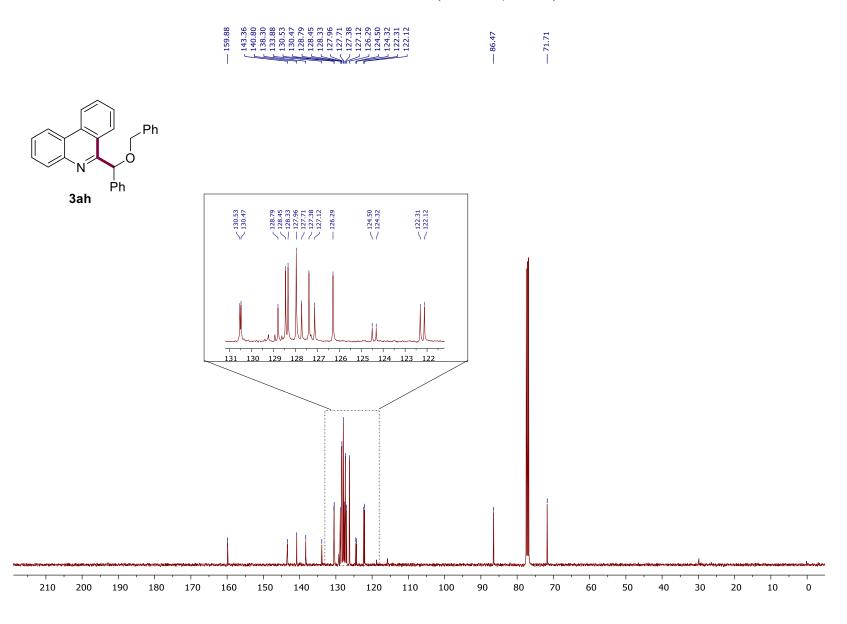


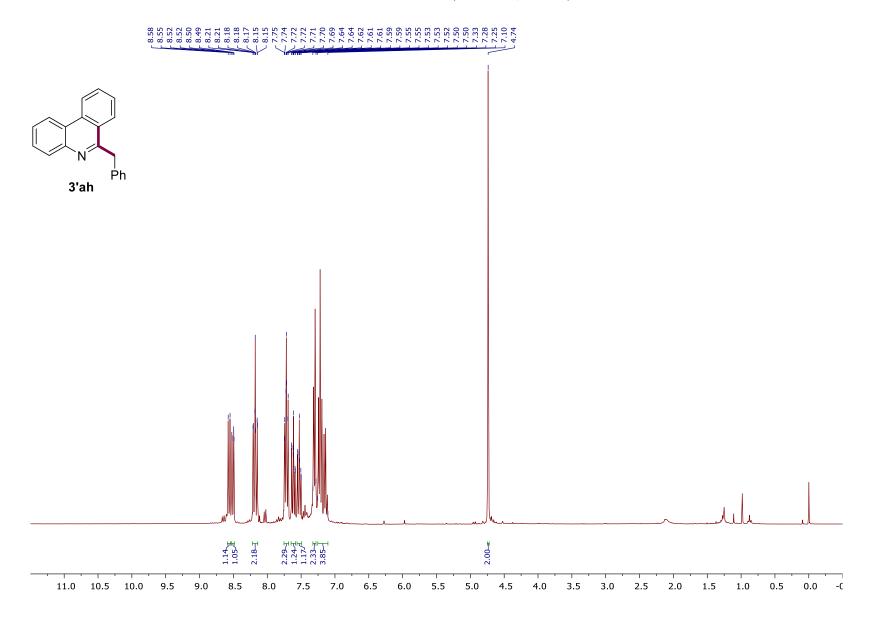
<sup>13</sup>C-NMR (75 MHz, CDCl<sub>3</sub>)



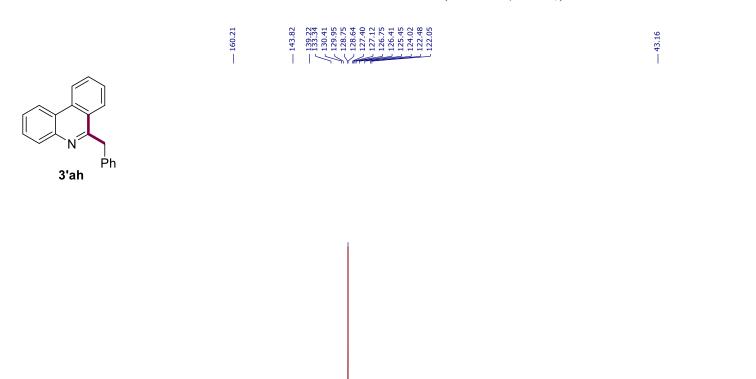


<sup>13</sup>C-NMR (101 MHz, CDCl<sub>3</sub>)





<sup>13</sup>C-NMR (101 MHz, CDCl<sub>3</sub>)



110 100

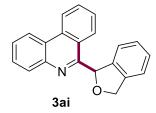
20

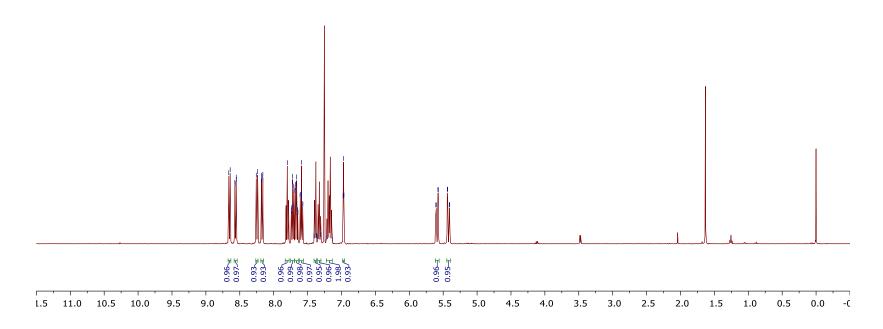
210 200

190 180

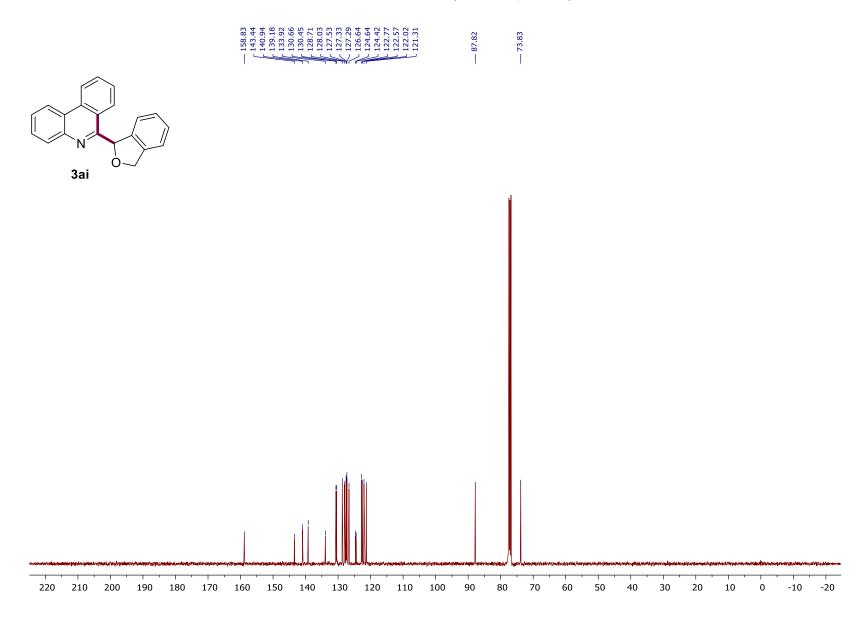
170 160 150 140 130 120

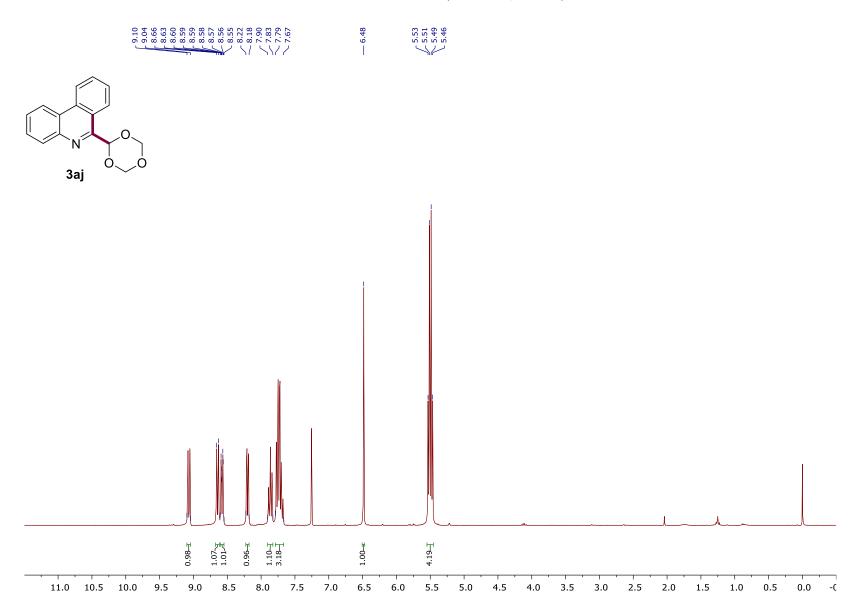
#### 8.65 8.75 8.15





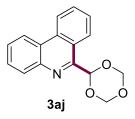
<sup>13</sup>C-NMR (101 MHz, CDCl<sub>3</sub>)

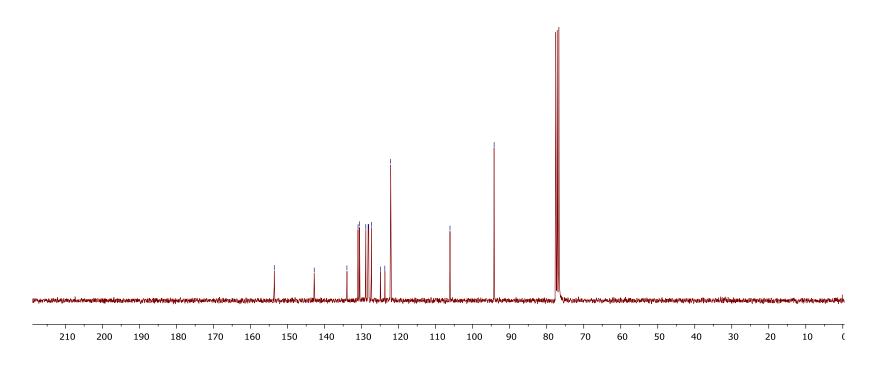


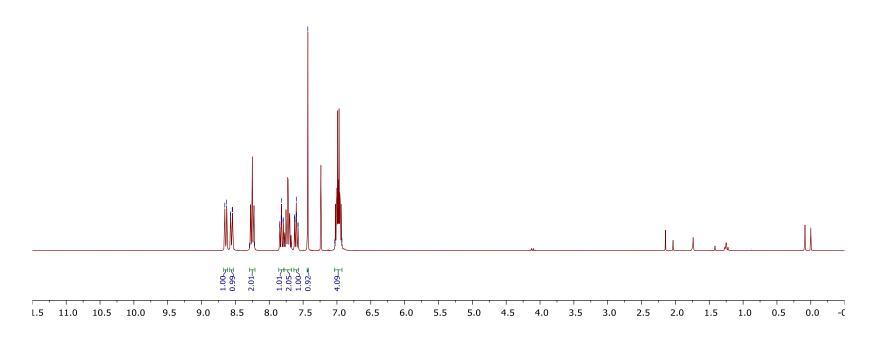


<sup>13</sup>C-NMR (75 MHz, CDCl<sub>3</sub>)

153.63	142.83 133.99 130.98 130.57 128.89 128.26 128.10 127.40 127.40 123.74	106.14	94.21

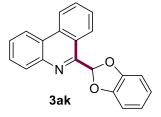


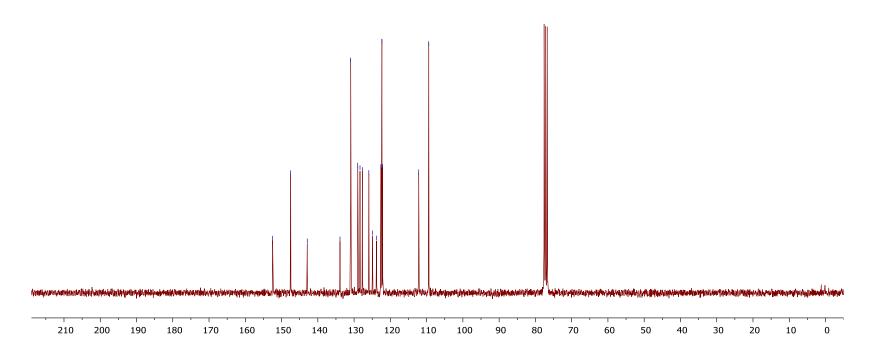




<sup>13</sup>C-NMR (75 MHz, CDCl<sub>3</sub>)



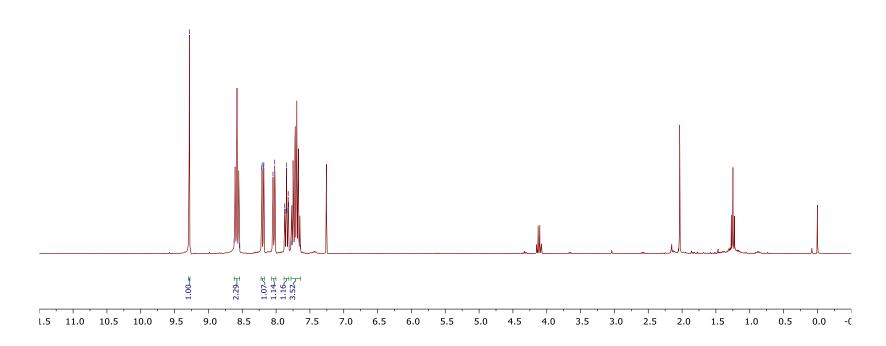








4a



<sup>13</sup>C-NMR (75 MHz, CDCl<sub>3</sub>)

89	.61	67 10 28 28 87 79 79 19 19 22 22 33 33
153,	144	132 131 130 128 128 127 127 127 127 127 127 127
T.	Ï	



4a

