

## Electronic Supplementary Information

### Mercury<sup>II</sup>-mediated base pairs in DNA: unexpected behavior in metal ion binding and duplex stability induced by 2'-deoxyuridine 5-substituents

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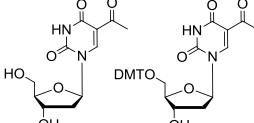
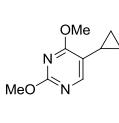
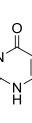
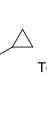
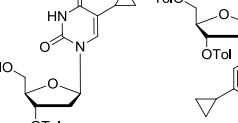
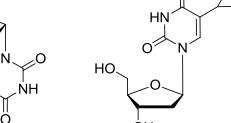
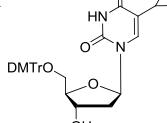
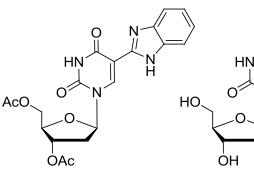
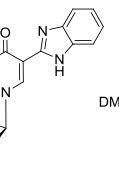
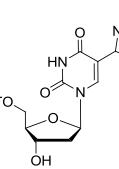
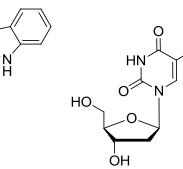
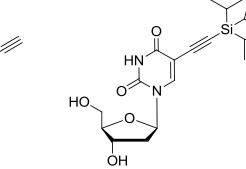
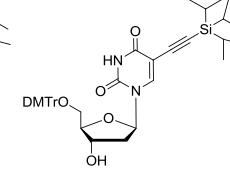
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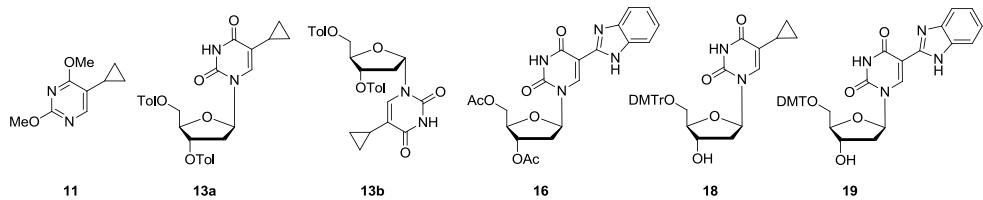
**Table S1**  $^{13}\text{C}$  NMR data of 5-substituted 2'-deoxyuridine nucleosides<sup>a,b</sup>

	C2	C4	C5	C6	5-substituents	C-1'	C-2'	C-3'	C-4'	C-5'					
<b>2<sup>1</sup></b>	149.9	161.6	112.1	146.5	193.4, 30.2	85.5	- <sup>[c]</sup>	70.2	87.9	61.0					
<b>17<sup>d</sup></b>	150.0	161.7	112.3	146.4	193.5, 30.6	86.6	- <sup>[c]</sup>	70.8	86.4	64.0					
<b>11</b>	163.0	169.4	116.0	154.3	6.8, 5.8	-	-	-	-	-					
<b>12</b>	164.4	150.8	113.4	135.7	7.4, 5.1	-	-	-	-	-					
<b>13a</b>	149.9	163.3	115.6	134.2	8.0, 5.3, 5.2	84.9	36.0	74.7	81.2	64.2					
<b>13b</b>	149.7	163.4	114.3	134.1	8.2, 5.2, 4.6	86.1	37.2	74.6	83.4	63.9					
<b>3<sup>2</sup></b>	149.8	163.3	115.1	134.1	7.7, 5.4, 5.3	83.8	- <sup>[c]</sup>	70.1	87.0	60.9					
<b>18</b>	149.8	163.1	115.0	134.0	8.1, 4.9	83.8	39.0	70.3	85.2	63.6					
<b>16</b>	149.5	161.8	104.5	140.3	145.8, 142.4, 134.3, 121.7, 117.7, 112.3,	85.2	37.2	74.1	81.9	63.7					
<b>9<sup>3</sup></b>	149.5	161.8	103.9	140.9	146.0, 121.6, 117.9, 112.2	85.3	- <sup>[c]</sup>	70.5	87.8	61.3					
<b>19</b>	149.4	161.9	103.7	140.6	145.9, 142.5, 134.3, 121.6, 121.5, 117.9, 112.2	86.1	40.7	70.5	85.9	63.6					
<b>4<sup>4</sup></b>	149.5	161.6	97.5	144.5	83.6, 76.4	84.7	- <sup>[c]</sup>	70.9	87.5	61.8					
<b>7<sup>d</sup></b>	149.4	161.4	99.5	144.4	98.3, 93.0, 52.6, 18.5, 10.7	84.9	- <sup>[c]</sup>	69.8	87.6	60.7					
<b>14<sup>1</sup></b>	148.7	160.9	98.3	144.1	98.3, 93.3, 54.4, 17.8, 10.1	84.8	- <sup>[c]</sup>	69.9	85.3	63.1					
	<b>2</b>		<b>17</b>		<b>11</b>		<b>12</b>		<b>13a</b>		<b>13b</b>		<b>3</b>		<b>18</b>
	<b>16</b>		<b>9</b>		<b>19</b>		<b>4</b>		<b>7</b>		<b>14</b>				

<sup>a</sup>Measured in DMSO-*d*<sub>6</sub> at 298 K. <sup>b</sup>Pyrimidine numbering. <sup>c</sup>Superimposed by DMSO. <sup>d</sup>Assigned by 2D spectra (HSQC and HMBC). All the other assignments were done by using <sup>1</sup>H-<sup>13</sup>C gated-decoupled spectra and DEPT-135 spectra.

**Table S2** Selected  $^1\text{H}$ - $^{13}\text{C}$  coupling constants (Hz) of 5-substituted 2'-deoxyuridine nucleosides<sup>a,b</sup>

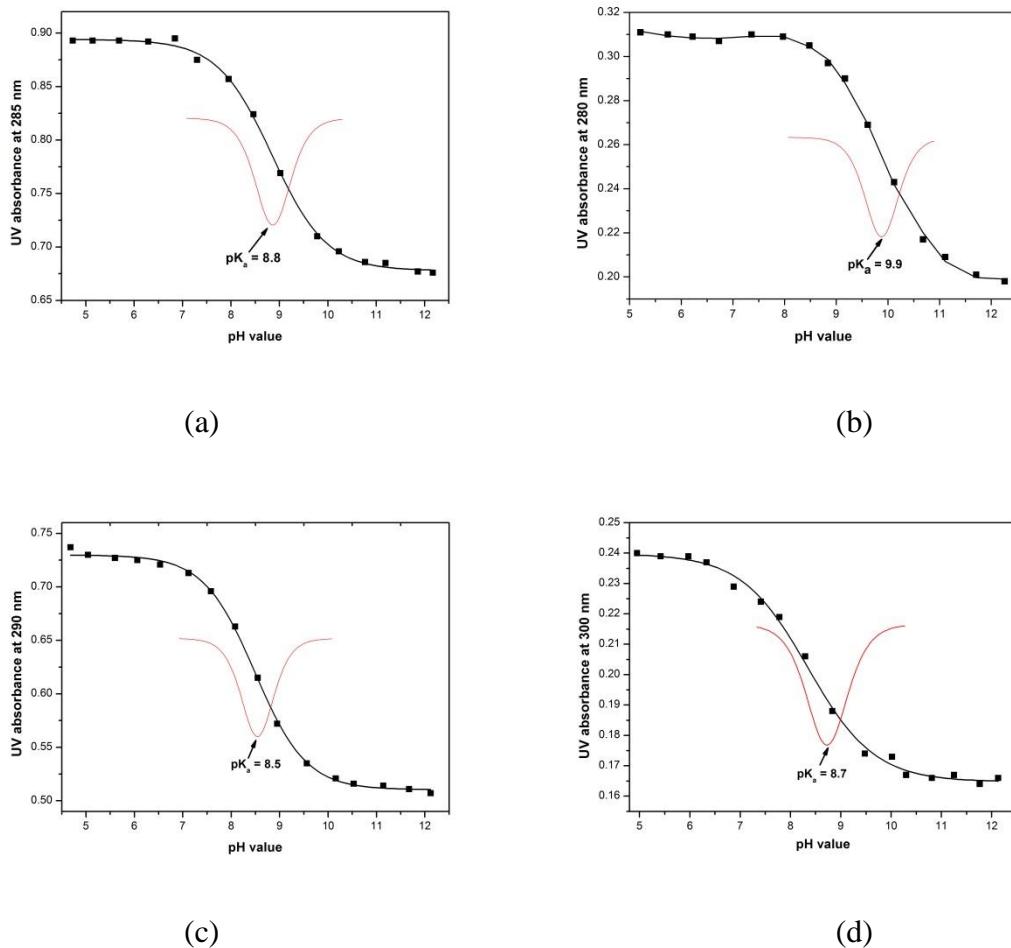
$^1\text{H}$ - $^{13}\text{C}$ coupling constants	<b>11</b>	<b>13a</b>	<b>13b</b>	<b>18</b>	<b>16</b>	<b>19</b>
$^1J(\text{C}6, \text{H-C}6)$	178.2	178.3	177.8	177.3	183.0	184.5
$^3J(\text{C}6, \text{H-C}1')$	4.3	n.d.	n.d.	4.8	3.5	4.3
$^3J(\text{C}2, \text{H-C}6)$ or $^3J(\text{C}2, \text{H-C}1')$	9.5	7.8	8.2	7.9	8.3	8.3
$^3J(\text{C}4, \text{H-C}6)$	9.4	10.0	10.0	10.0	9.2	9.4
$^1J(\text{C}1', \text{H-C}1')$	-	167.4	168.7	169.0	170.5	167.2
$^1J(\text{C}3', \text{H-C}3')$	-	161.4	154.0	147.2	159.2	150.0
$^1J(\text{C}4', \text{H-C}4')$	-	153.6	154.3	149.0	152.3	151.9
$^1J(\text{C}5', \text{H-C}5')$	-	152.6	149.4	142.4	147.9	141.7



<sup>a</sup>Measured in DMSO- $d_6$  at 298 K. <sup>b</sup>Pyrimidine numbering. n.d.: not detected.

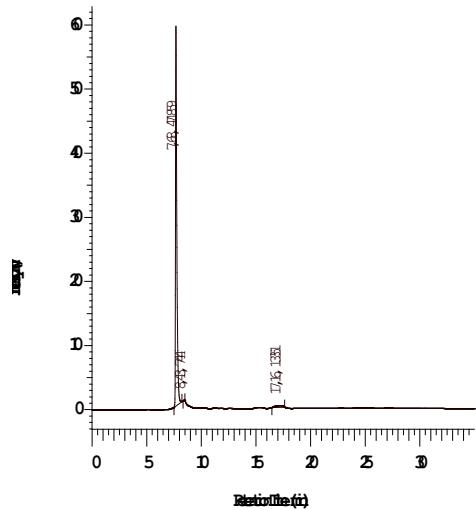
## $pK_a$ Determination by UV

Nucleosides (**2**, **3**, **4**, or **7**) were dissolved in 0.1 M sodium phosphate buffer, pH 4.5. An aqueous NaOH solution (4 M) and concentrated phosphorus acid were used to adjust the pH value of the buffer. At defined pH values, the UV absorbance of nucleosides was measured (Fig. S1).

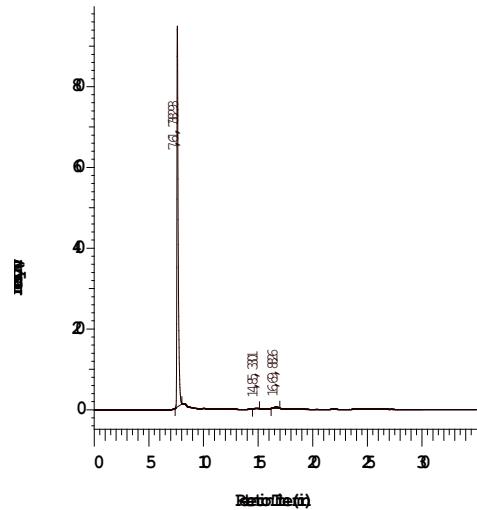


**Fig. S1** UV absorbance *vs* pH. (a) Nucleoside **2** at 285 nm; (b) nucleoside **3** at 280 nm; (c) nucleoside **4** at 290 nm; (d) nucleoside **7** at 300 nm. All measurements were performed in 0.1 M sodium phosphate buffer.

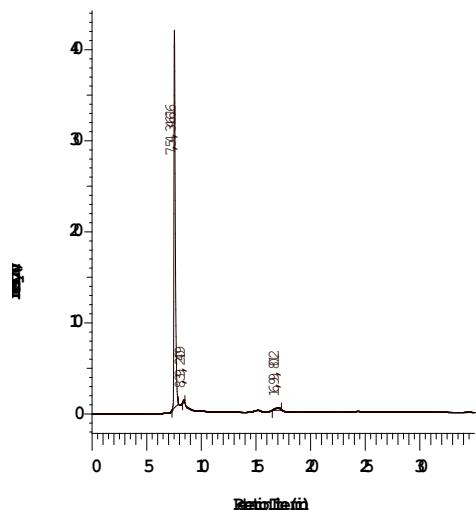
**Reversed-phase HPLC profiles of purified oligonucleotides determined at 260 nm.**



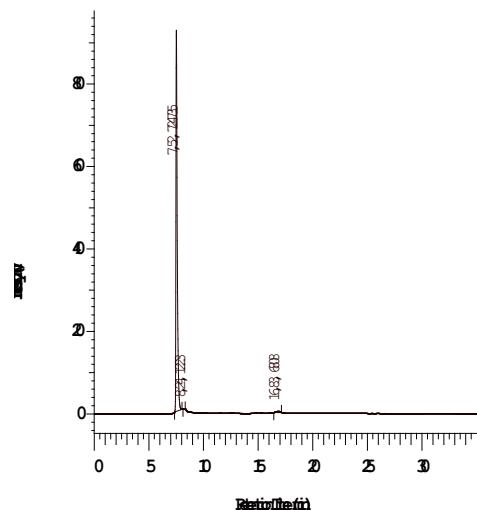
(a) 5'-d( TAG GTC **2**AT ACT) ODN **29**



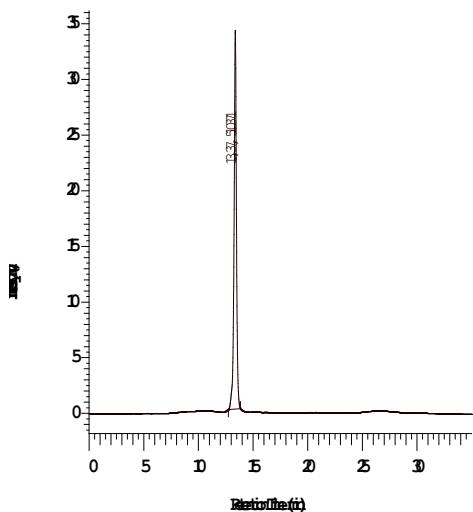
(b) 5'-d( AGT AT**2** GAC CTA) ODN **30**



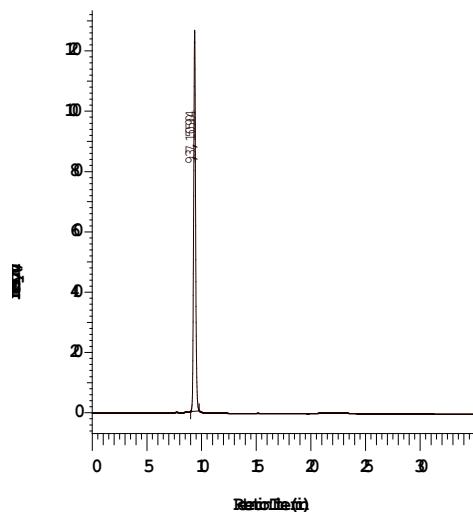
(c) 5'-d( TAG GTC **3**AT ACT) ODN **31**



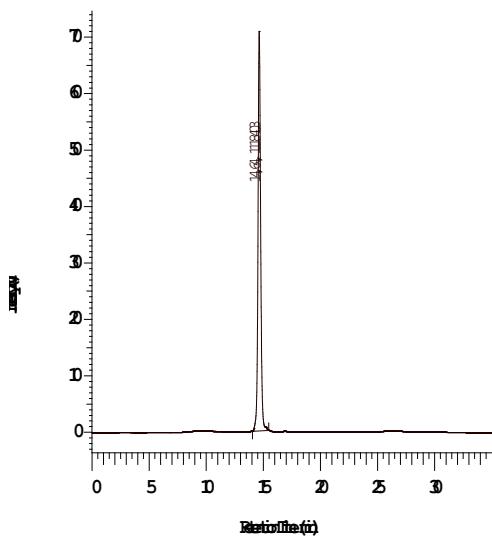
(d) 5'-d( AGT AT**3** GAC CTA) ODN **32**



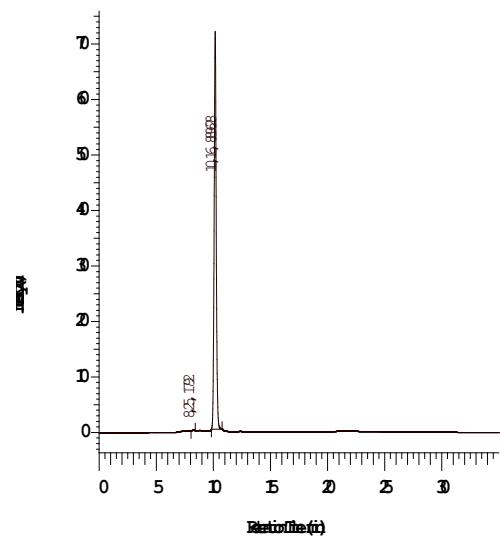
(e) 5'-d(TAG GTC **5**AT ACT) ODN **35**



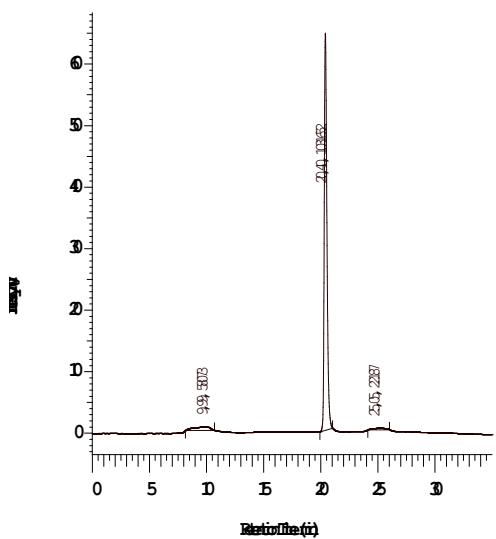
(f) 5'-d(AGT AT**5** GAC CTA) ODN **36**



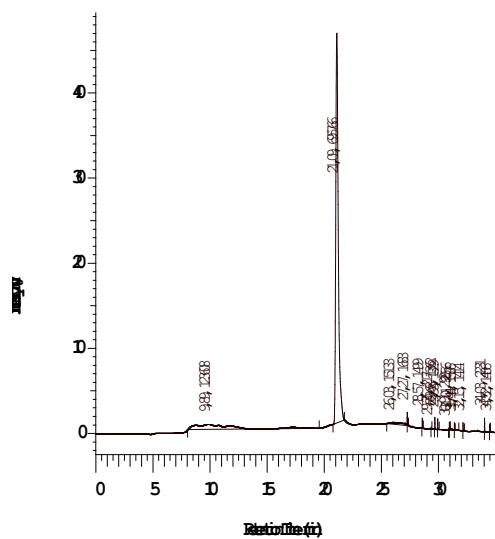
(g) 5'-d(TAG GTC **6**AT ACT) ODN **37**



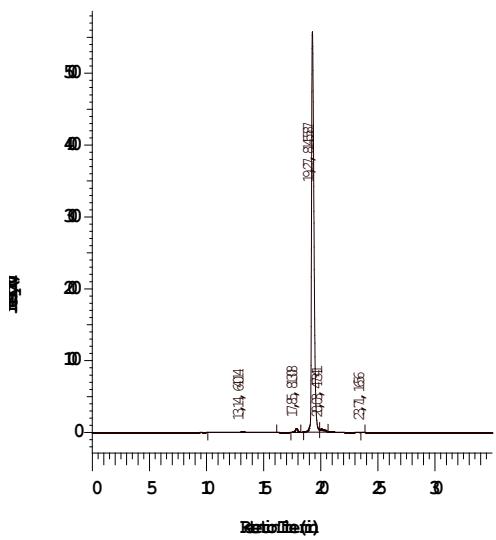
(h) 5'-d(AGT AT**6** GAC CTA) ODN **38**



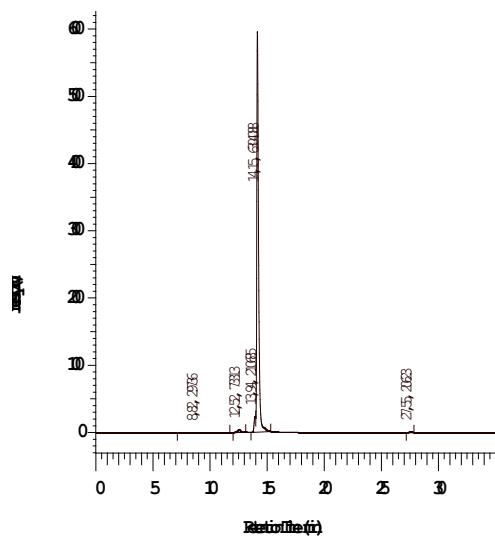
(i) 5'-d(TAG GTC 7AT ACT) ODN **39**



(j) 5'-d(AGT AT7 GAC CTA) ODN **40**

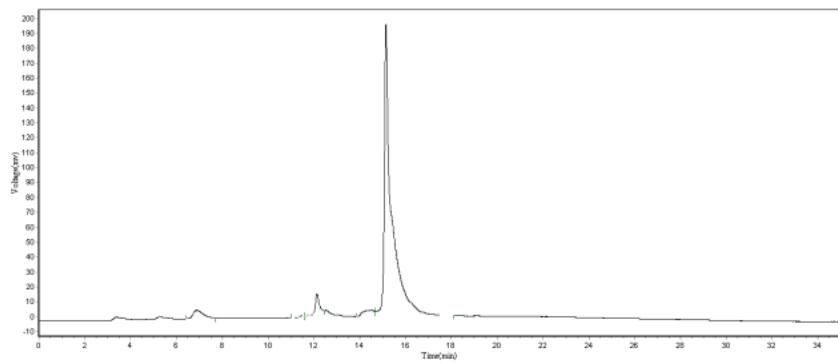


(k) 5'-d(TAG GTC 9AT ACT) ODN **43**

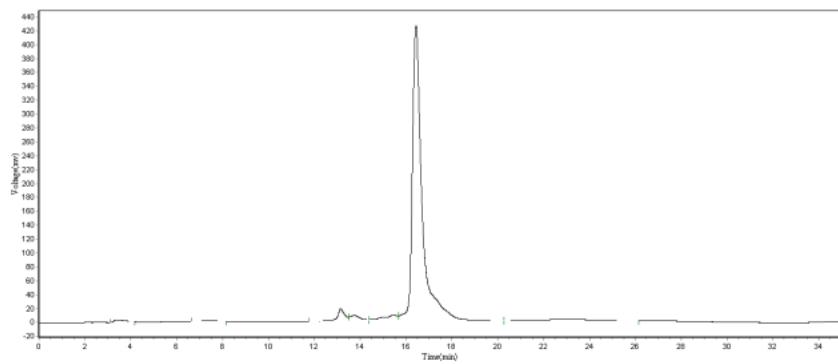


(l) 5'-d(AGT AT9GAC CTA) ODN **44**

**Fig. S2** HPLC purity profiles of oligonucleotides. (a) ODN **29**; (b) ODN **30**; (c) ODN **31**; (d) ODN **32**; (e) ODN **35**; (f) ODN **36**; (g) ODN **37**; (h) ODN **38**; (i) ODN **39**; (j) ODN **40**; (k) ODN **43**; (l) ODN **44**. For elution the following solvent system was used: 0.1 M ( $\text{Et}_3\text{NH}$ )OAc : MeCN (95:5) (pH 7.0) (A) and MeCN (B). Gradient: 0-20 min 0-20% B in A, 20-25 min 20% B in A, 25-30 min 20-0% B in A, flow rate 0.8 mL min<sup>-1</sup>.



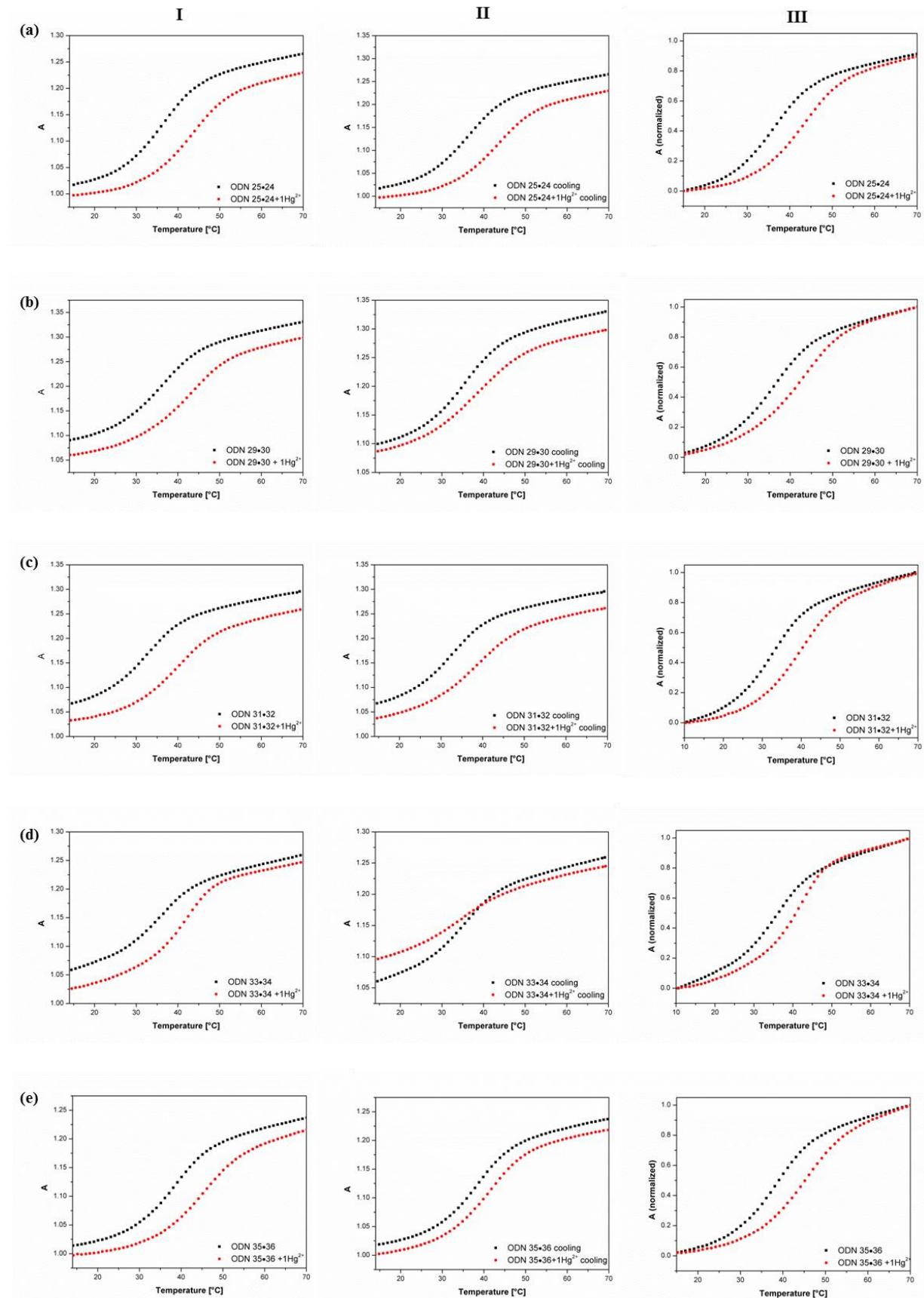
(a) 5'-d(TAG GTC **8**AT ACT) ODN **41**

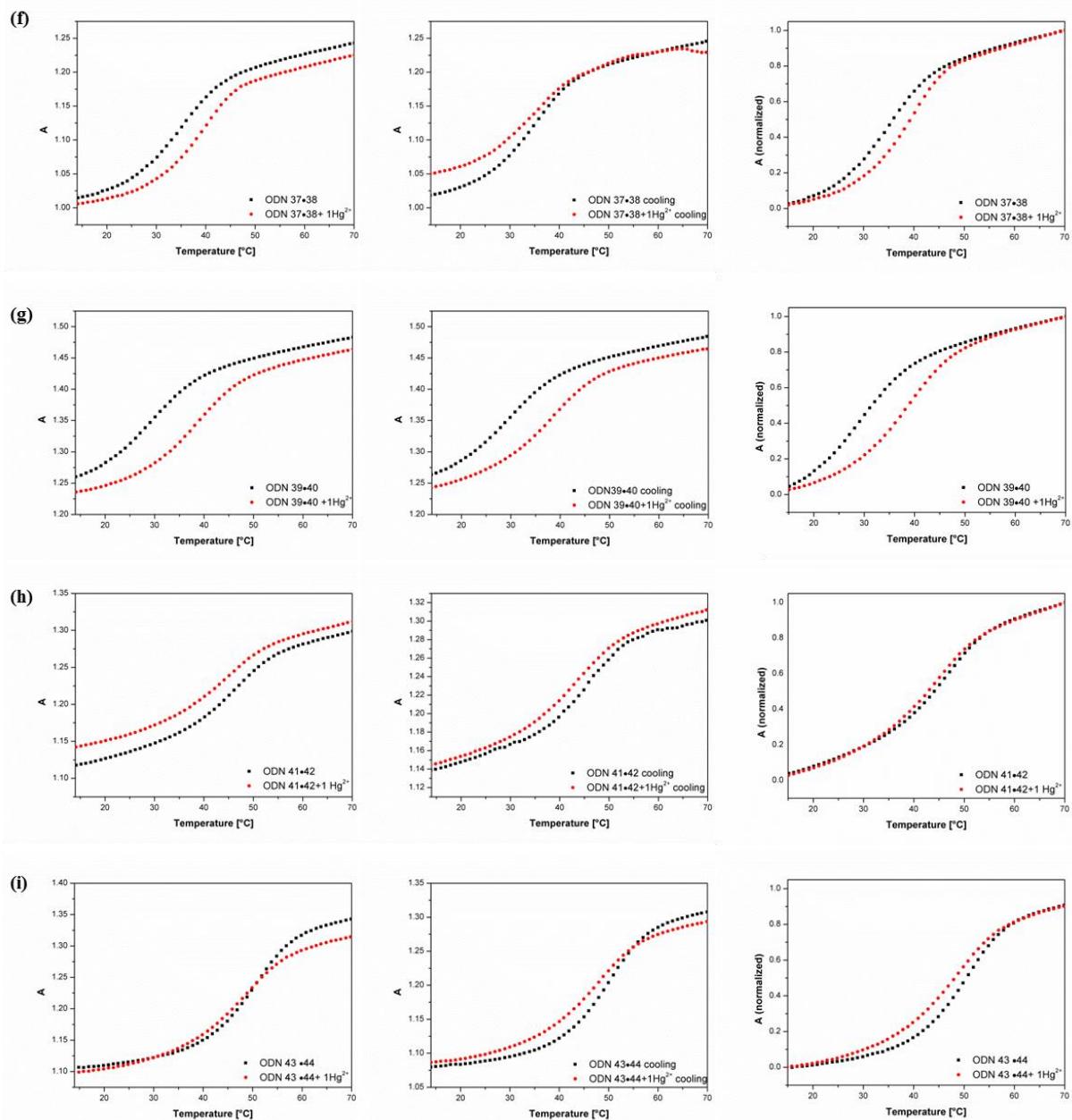


(b) 5'-d(AGT AT**8** GAC CTA) ODN **42**

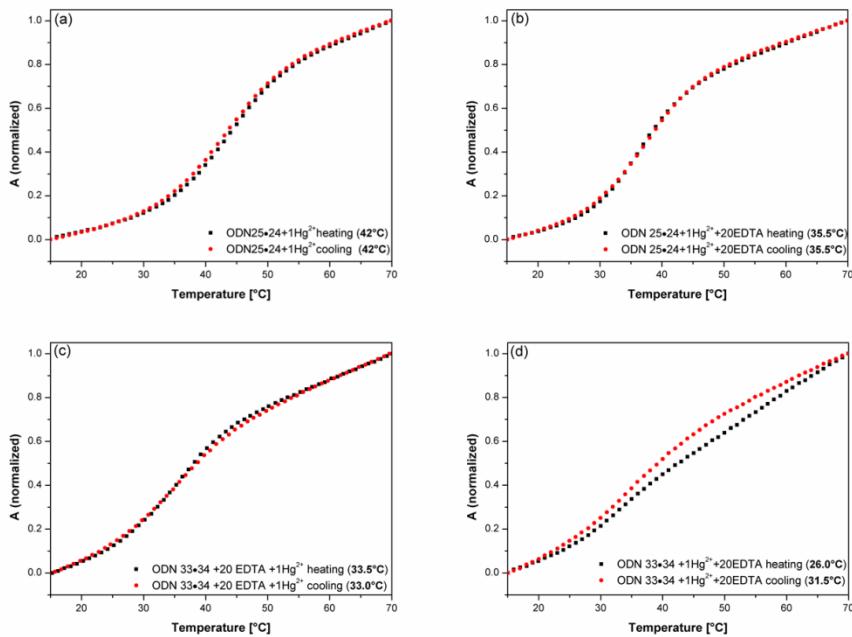
**Fig. S3** HPLC profiles of phenyltriazolyl modified oligonucleotides (crude mixture): (a) ODN **41**; (b) ODN **42**. For elution the following solvent system was used: 0.1 M ( $\text{Et}_3\text{NH}$ )OAc : MeCN (95:5) (pH 7.0) (A) and MeCN (B). Gradient: 0-20 min 0-20% B in A, 20-25 min 20% B in A, 25-30 min 20-0% B in A, flow rate  $0.8 \text{ mL min}^{-1}$ .

## Melting profiles of duplexes



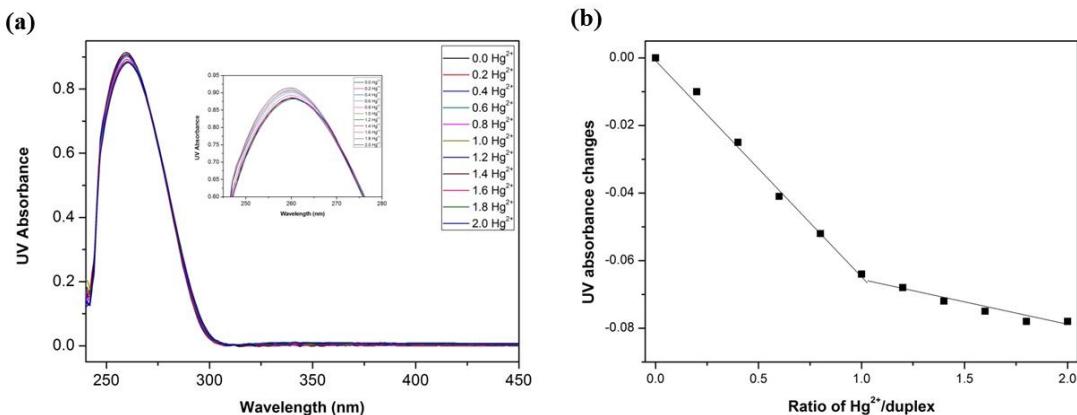


**Fig. S4** The original and normalized melting curves of duplexes obtained from heating and cooling experiments with a single-strand concentration of 5  $\mu\text{M}$  + 5  $\mu\text{M}$  in 10 mM Mops, 100 mM NaNO<sub>3</sub> (pH 7.0) at 260 nm, absence and in presence of 1 equiv. of Hg<sup>2+</sup>. Figures in column I: original melting curves (heating). Figures in column II: original melting curves (cooling). Figures in column III: normalized melting curves (heating). Relative absorbance A (normalized) =  $(A - A_{\min}) / (A_{\max} - A_{\min})$  at 260 nm. : a) ODN 24•25; b) ODN 29•30; c) ODN 31•32; d) ODN 33•34; e) ODN 35•36; f) ODN 37•38; g) ODN 39•40; h) ODN 41•42; i) ODN 43•44.

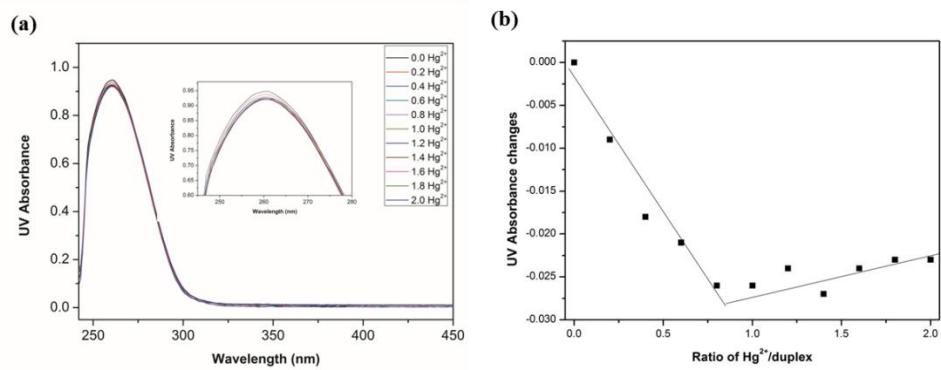


**Fig. S5** The melting curves of duplexes obtained from heating and cooling experiments with a single-strand concentration of 5  $\mu\text{M}$  + 5  $\mu\text{M}$  in 10 mM Mops, 100 mM NaNO<sub>3</sub> (pH 7.0) at 260 nm, in presence of 1 equiv. of Hg<sup>2+</sup>. Relative absorbance A(normalized) =  $(A - A_{\min}) / (A_{\max} - A_{\min})$  at 260 nm: a) ODN 25•24 + 1Hg<sup>2+</sup>; b) ODN 25•24 + 1Hg<sup>2+</sup> + 20 equiv. EDTA; c) ODN 33•34+1 Hg<sup>2+</sup>; d) ODN 33•34 + 1Hg<sup>2+</sup> + 20 equiv. EDTA.

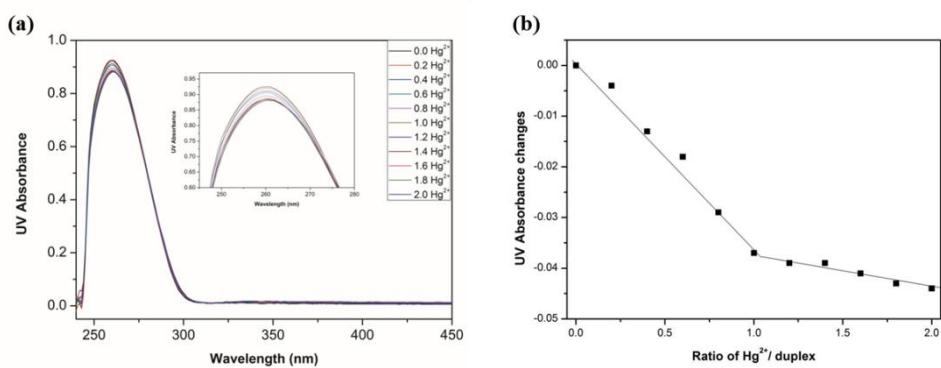
### Stoichiometric titrations of oligonucleotides



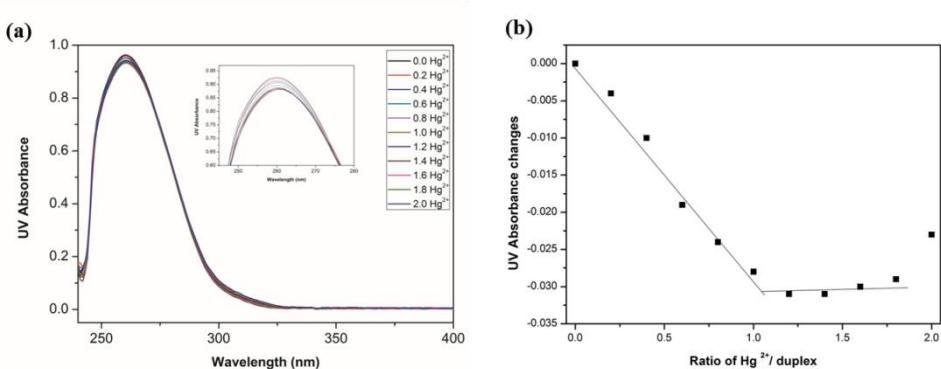
**Fig. S6** (a) UV spectrophotometric titration of 5  $\mu\text{M}$  ODN 25•24 with increasing concentration of Hg<sup>2+</sup> ions (0–2.0 equiv.) in buffer (10 mM Mops, 100 mM NaNO<sub>3</sub>, pH 7.0). (b) Graph of ratio of equivalents of Hg<sup>2+</sup>/duplex vs changes in absorbance at 260 nm from (a).



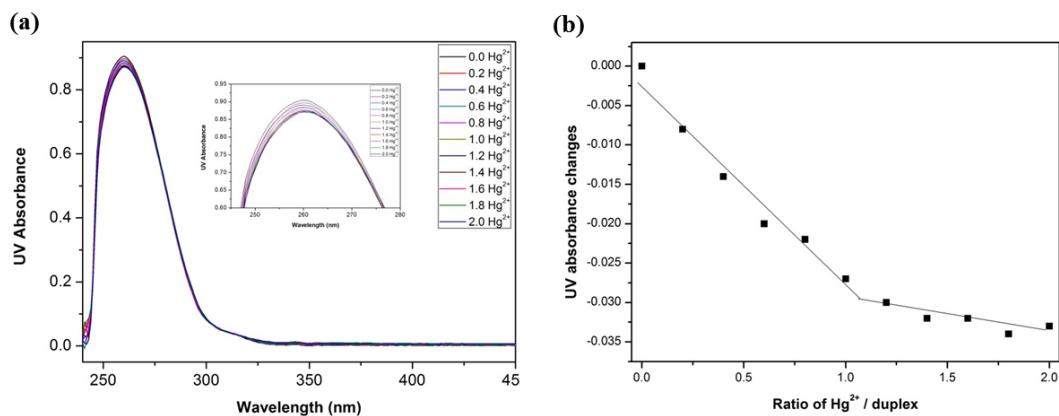
**Fig. S7** (a) UV spectrophotometric titration of 5  $\mu\text{M}$  ODN **29•30** with increasing concentration of  $\text{Hg}^{2+}$  ions (0–2.0 equiv.) in buffer (10 mM Mops, 100 mM  $\text{NaNO}_3$ , pH 7.0). (b) Graph of ratio of equivalents of  $\text{Hg}^{2+}$ /duplex *vs* changes in absorbance at 260 nm from (a).



**Fig. S8** (a) UV spectrophotometric titration of 5  $\mu\text{M}$  ODN **31•32** with increasing concentration of  $\text{Hg}^{2+}$  ions (0–2.0 equiv.) in buffer (10 mM Mops, 100 mM  $\text{NaNO}_3$ , pH 7.0). (b) Graph of ratio of equivalents of  $\text{Hg}^{2+}$ /duplex *vs* changes in absorbance at 260 nm from (a).



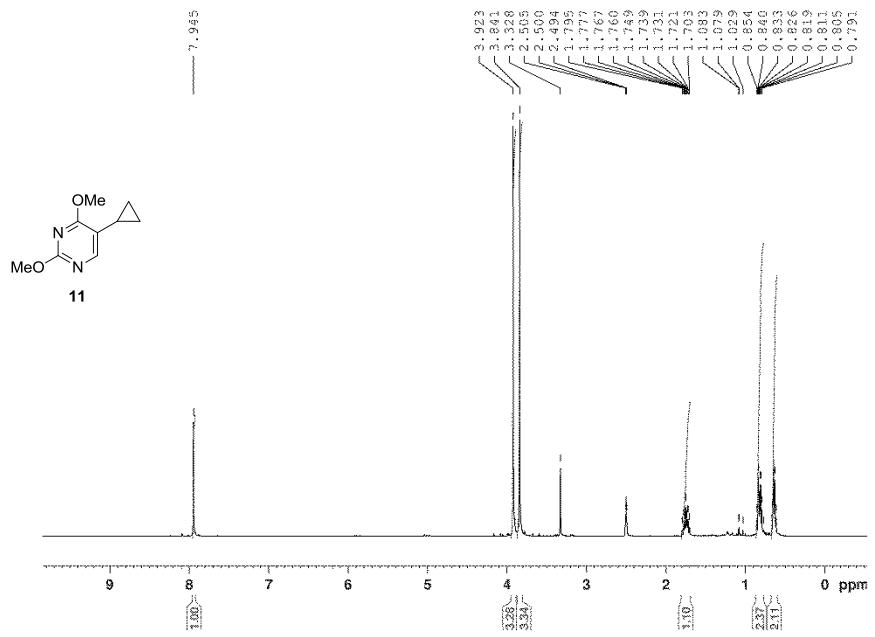
**Fig. S9** (a) UV spectrophotometric titration of 5  $\mu\text{M}$  ODN **33•34** with increasing concentration of  $\text{Hg}^{2+}$  ions (0–2.0 equiv.) in buffer (10 mM Mops, 100 mM  $\text{NaNO}_3$ , pH 7.0). (b) Graph of ratio of equivalents of  $\text{Hg}^{2+}$ /duplex *vs* changes in absorbance at 260 nm from (a).



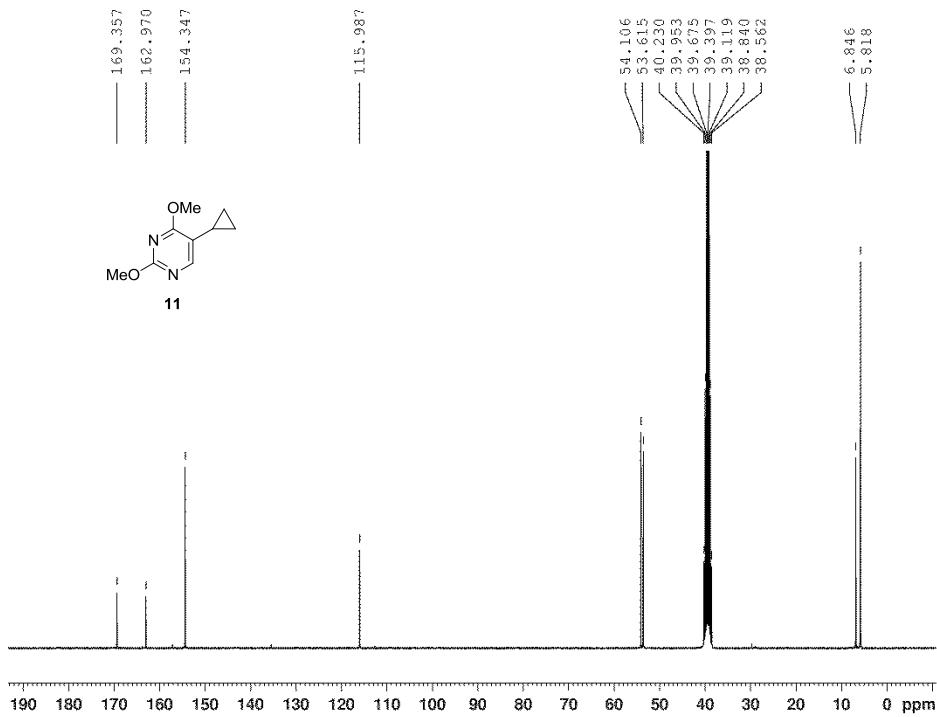
**Fig. S10** (a) UV spectrophotometric titration of 5  $\mu\text{M}$  ODN **35•36** with increasing concentration of  $\text{Hg}^{2+}$  ions (0–2.0 equiv) in buffer (10 mM Mops, 100 mM  $\text{NaNO}_3$ , pH 7.0). (b) Graph of ratio of equivalents of  $\text{Hg}^{2+}$ /duplex *vs* changes in absorbance at 260 nm from (a).

## References

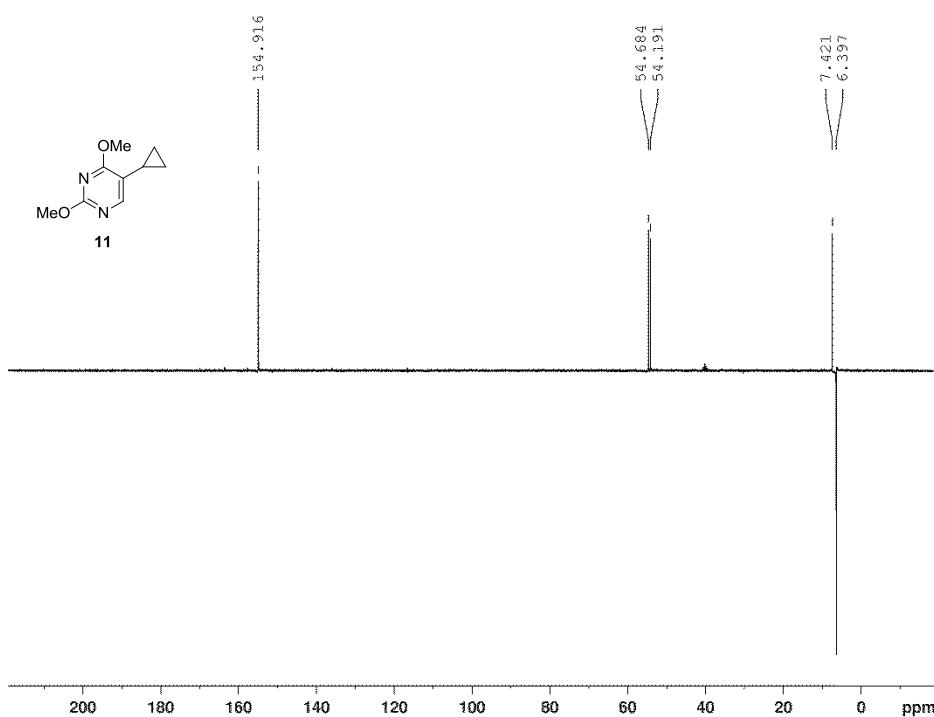
- (1) S. A. Ingale, H. Mei, P. Leonard and F. Seela, *J. Org. Chem.*, 2013, **78**, 11271-11282.
- (2) I. Basnak, A. Balkan, P. L. Coe and R. T. Walker, *Nucleos. Nucleot.*, 1994, **13**, 177-196.
- (3) J. Krim, C. Grünewald, M. Taourirte and J. W. Engels, *Bioorg. Med. Chem.*, 2012, **20**, 480-486.
- (4) V. Borsenberger, M. Kukwikila and S. Howorka, *Org. Biomol. Chem.*, 2009, **7**, 3826-3835.



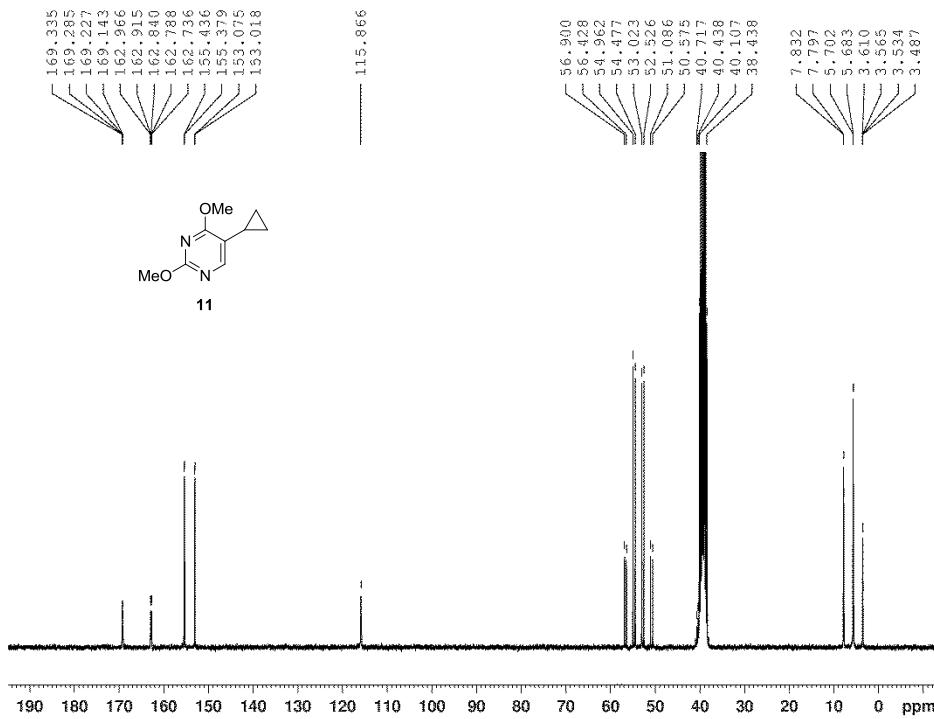
**Fig. S11**  $^1\text{H}$  NMR spectrum of compound **11**.



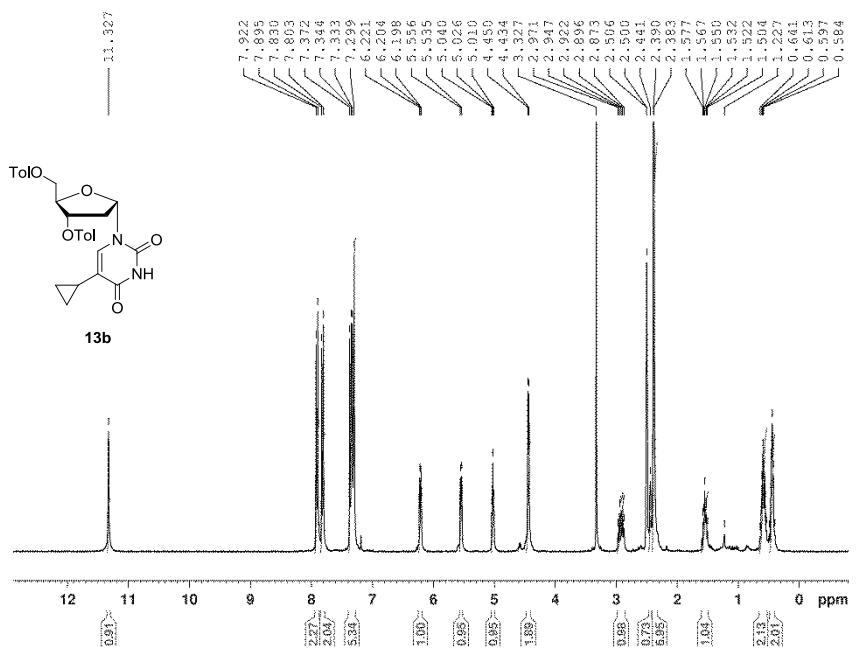
**Fig. S12**  $^{13}\text{C}$  NMR spectrum of compound **11**.



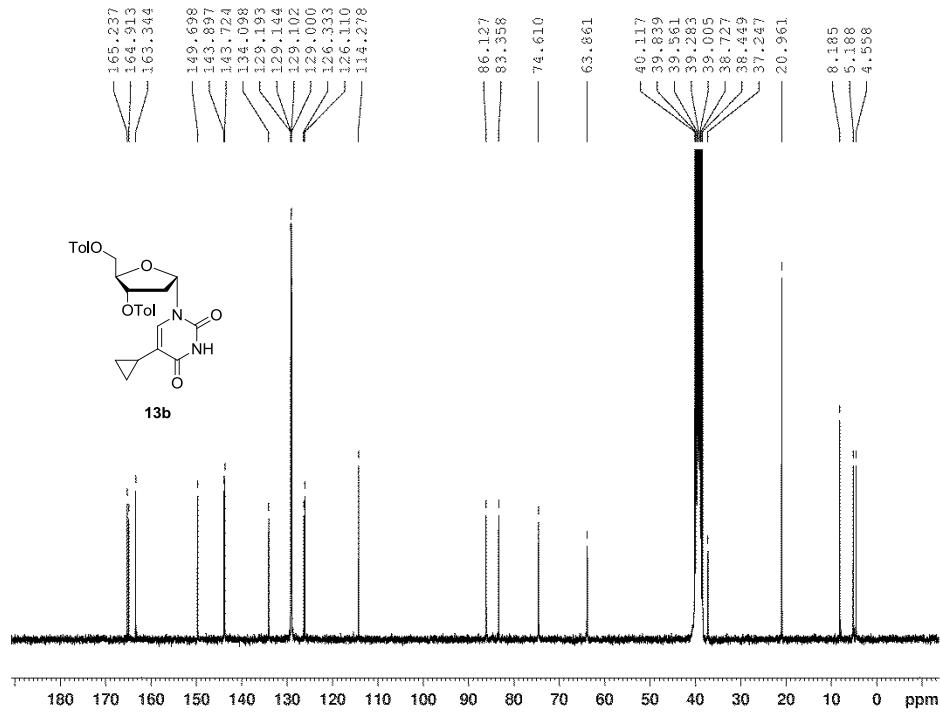
**Fig. S13** DEPT-135 spectrum of compound **11**.



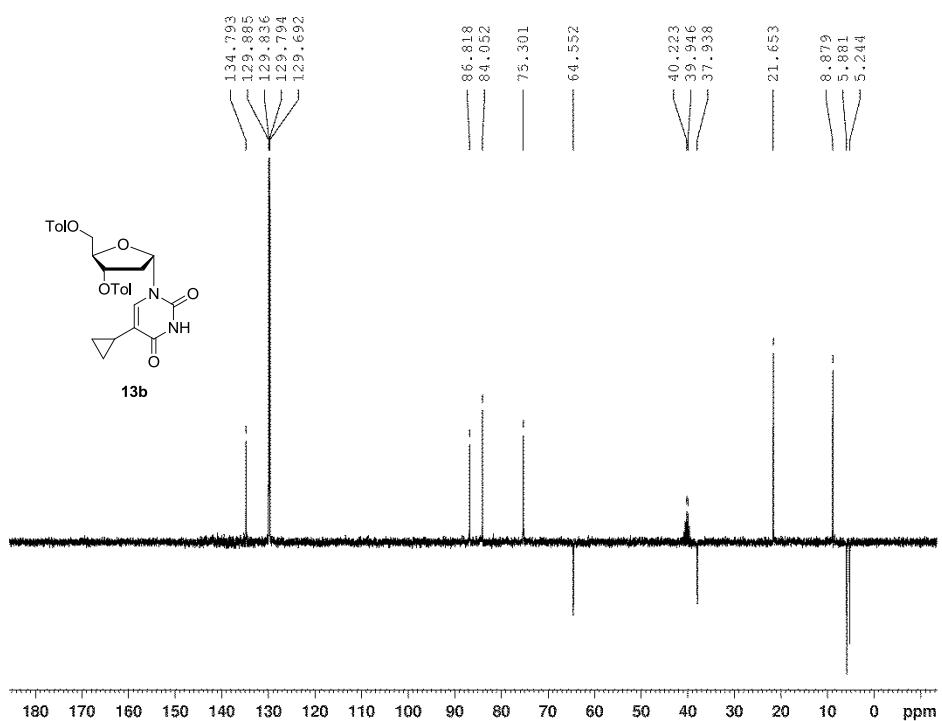
**Fig. S14**  $^1\text{H}$ - $^{13}\text{C}$  gated-decoupled spectrum of compound **11**.



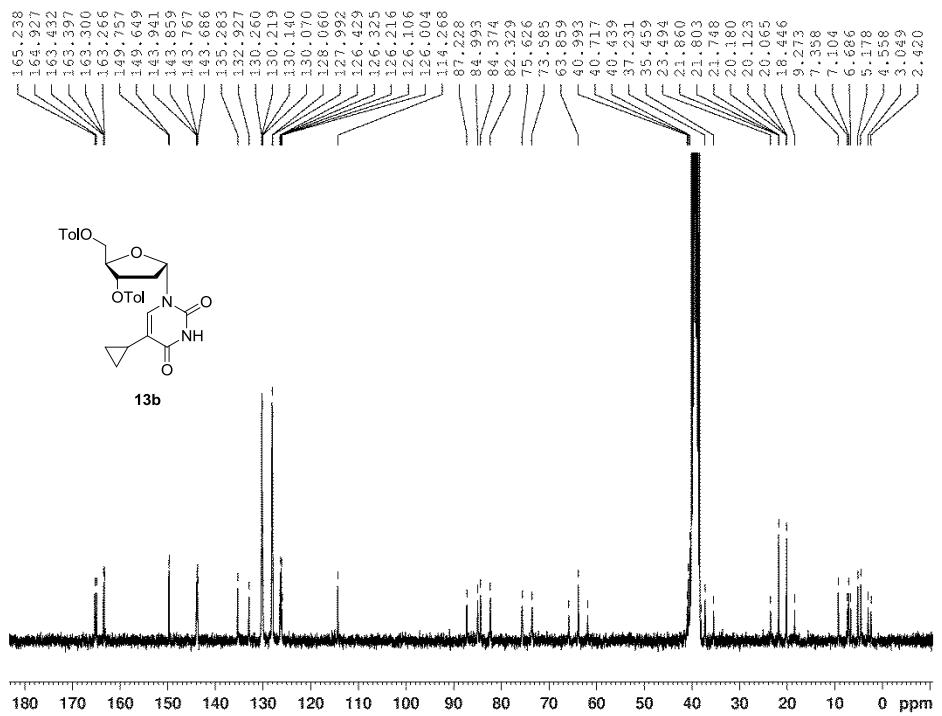
**Fig. S15** <sup>1</sup>H NMR spectrum of compound **13b**.



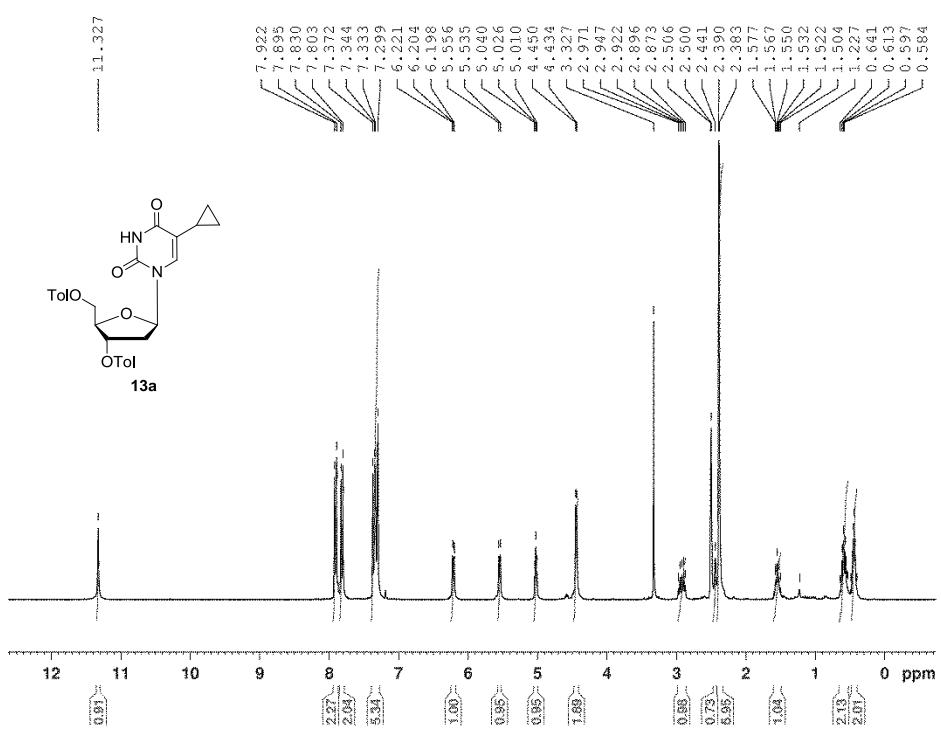
**Fig. S16** <sup>13</sup>C NMR spectrum of compound **13b**.



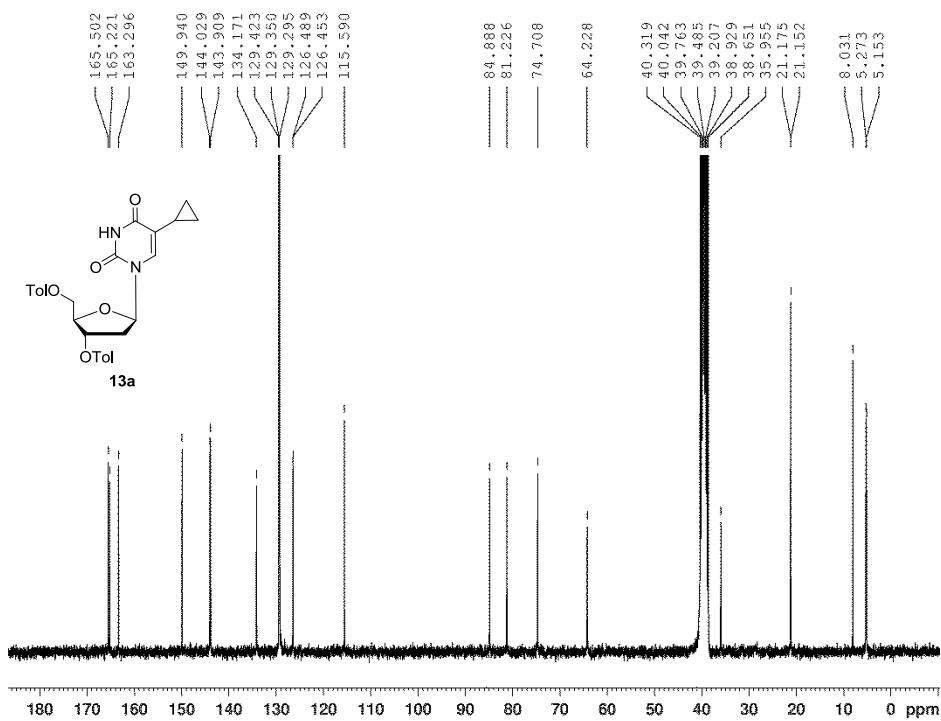
**Fig. S17** DEPT-135 spectrum of compound **13b**.



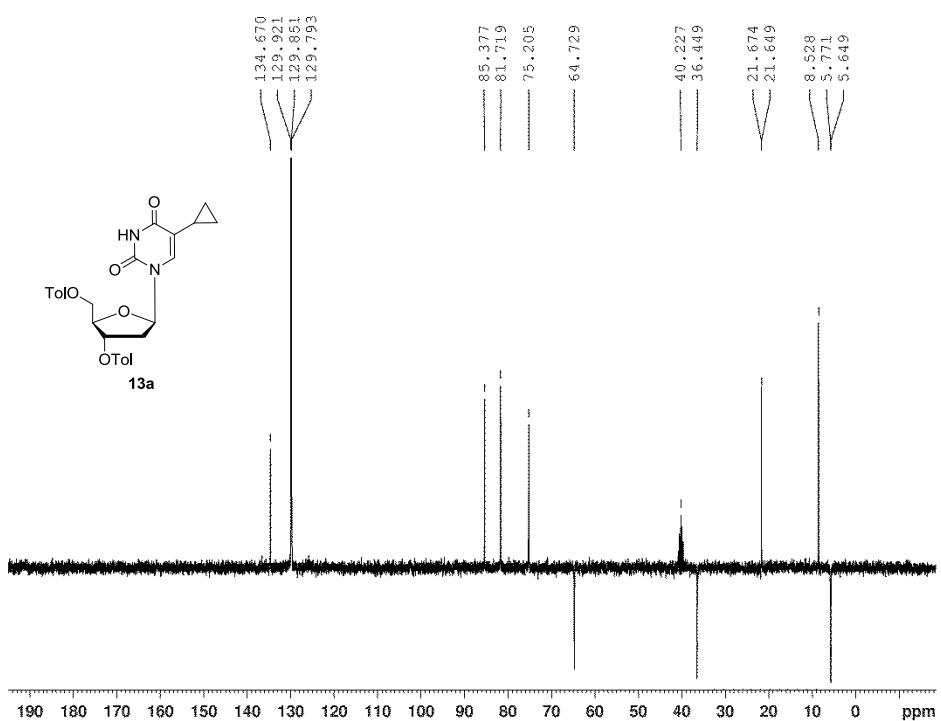
**Fig. S18**  $^1\text{H}$ - $^{13}\text{C}$  gated-decoupled spectrum of compound **13b**.



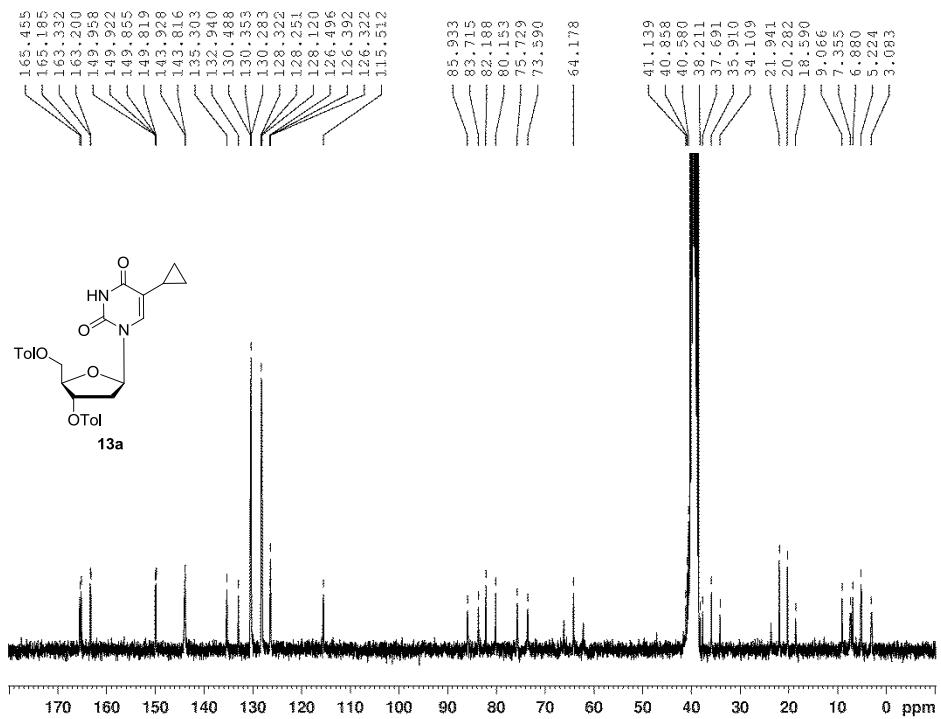
**Fig. S19** <sup>1</sup>H NMR spectrum of compound **13a**



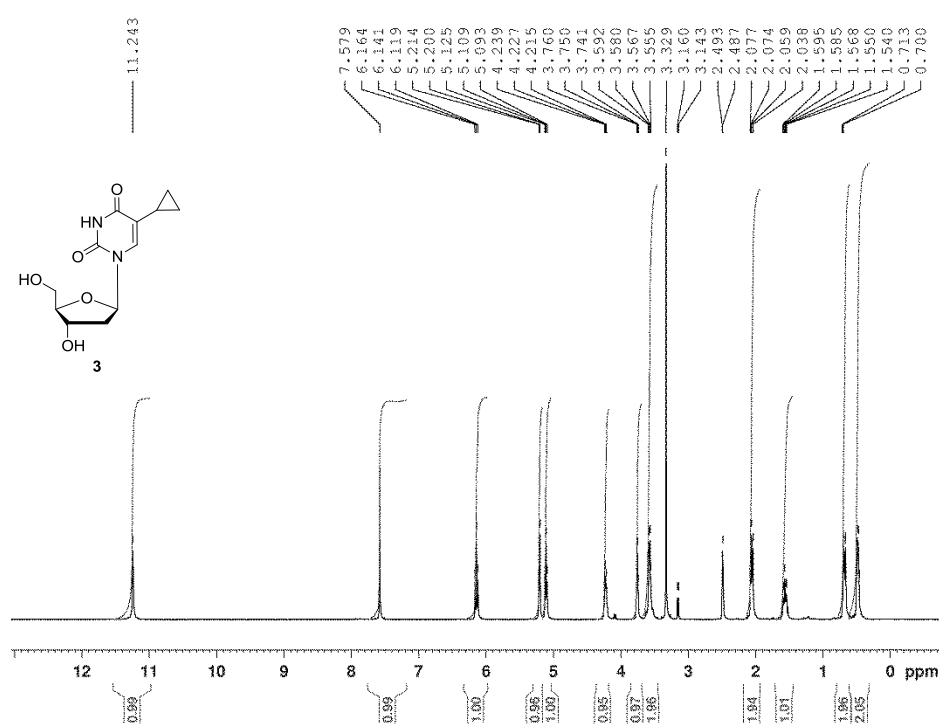
**Fig. S20** <sup>13</sup>C NMR spectrum of compound **13a**.



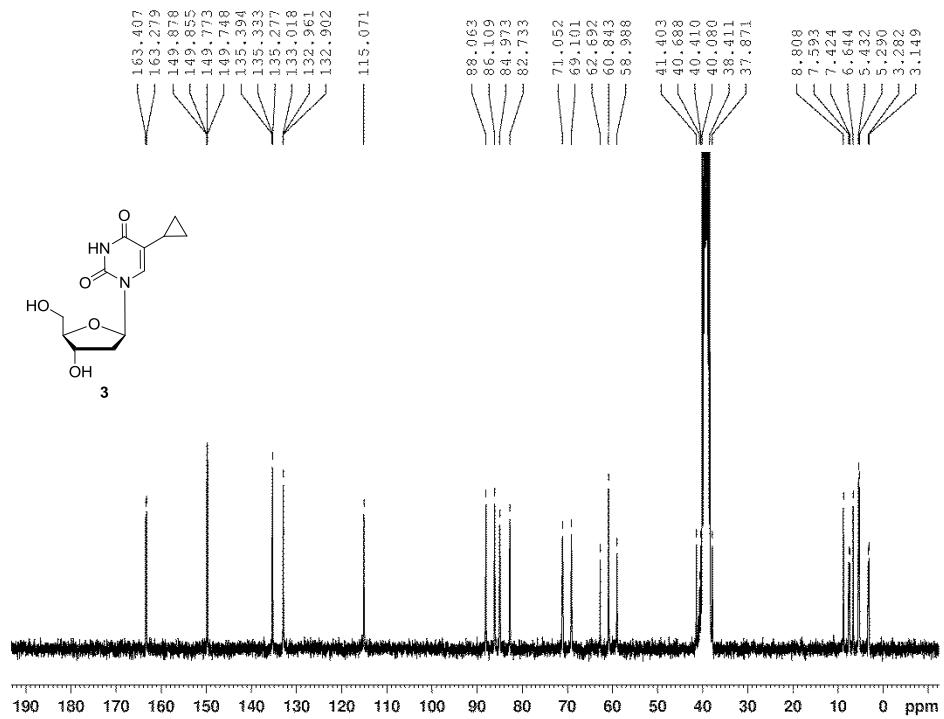
**Fig. S21** DEPT-135 spectrum of compound **13a**



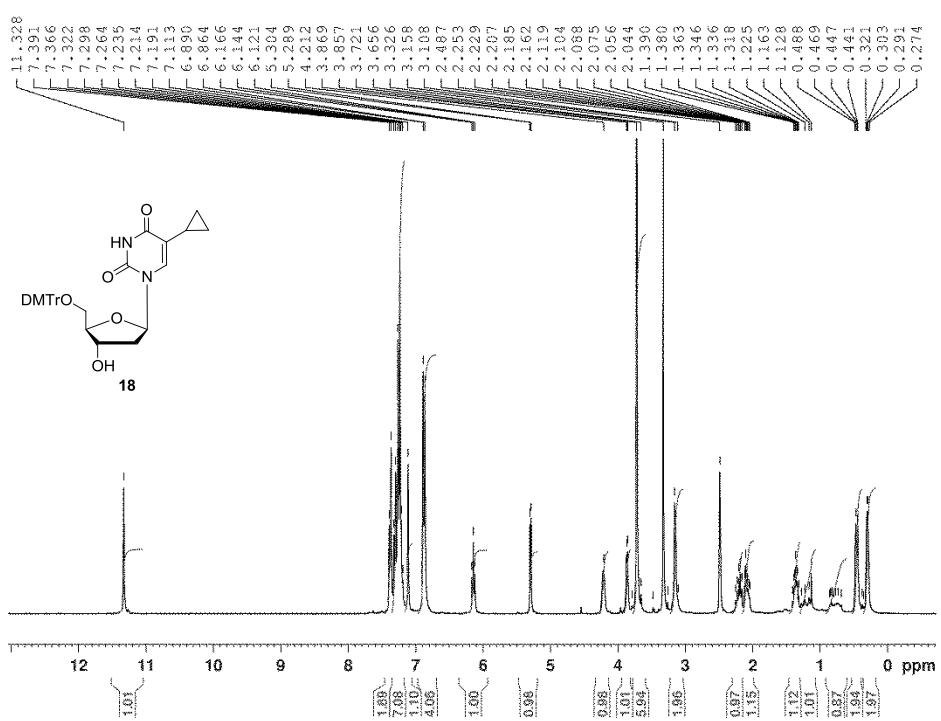
**Fig. S22**  $^1\text{H}$ - $^{13}\text{C}$  gated-decoupled spectrum of compound **13a**.



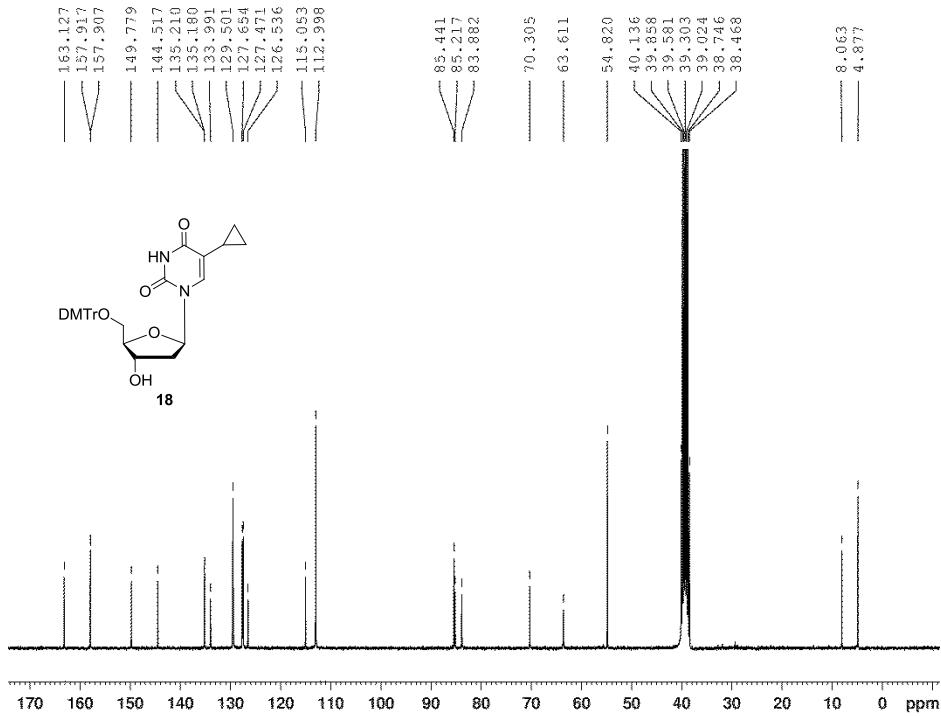
**Fig. S23** <sup>1</sup>H NMR spectrum of compound 3



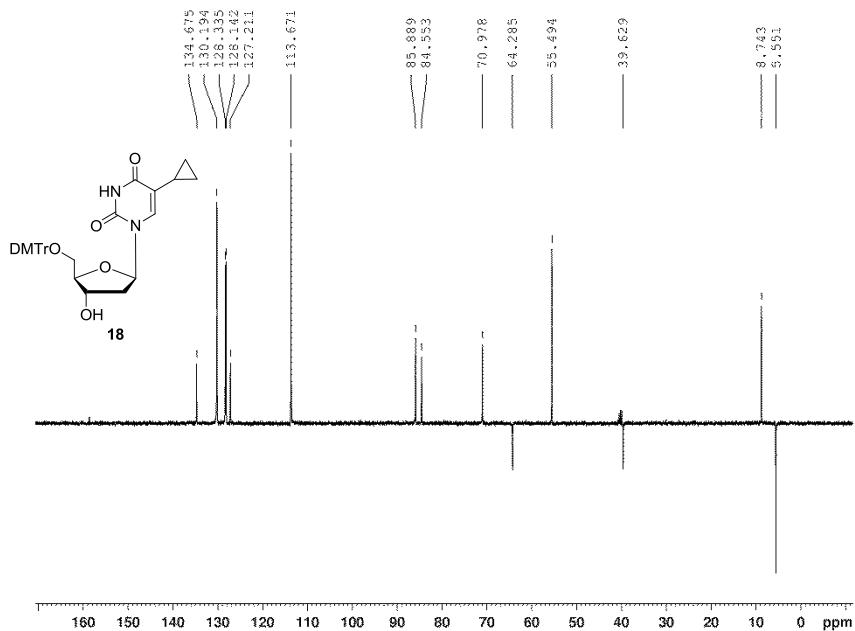
**Fig. S24** <sup>13</sup>C NMR spectrum of compound 3.



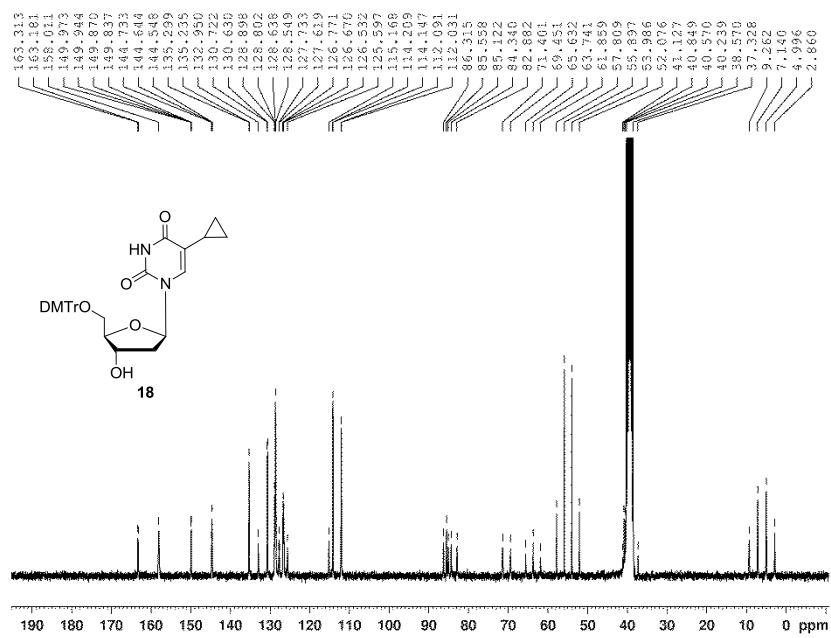
**Fig. S25** <sup>1</sup>H NMR spectrum of compound **18**.



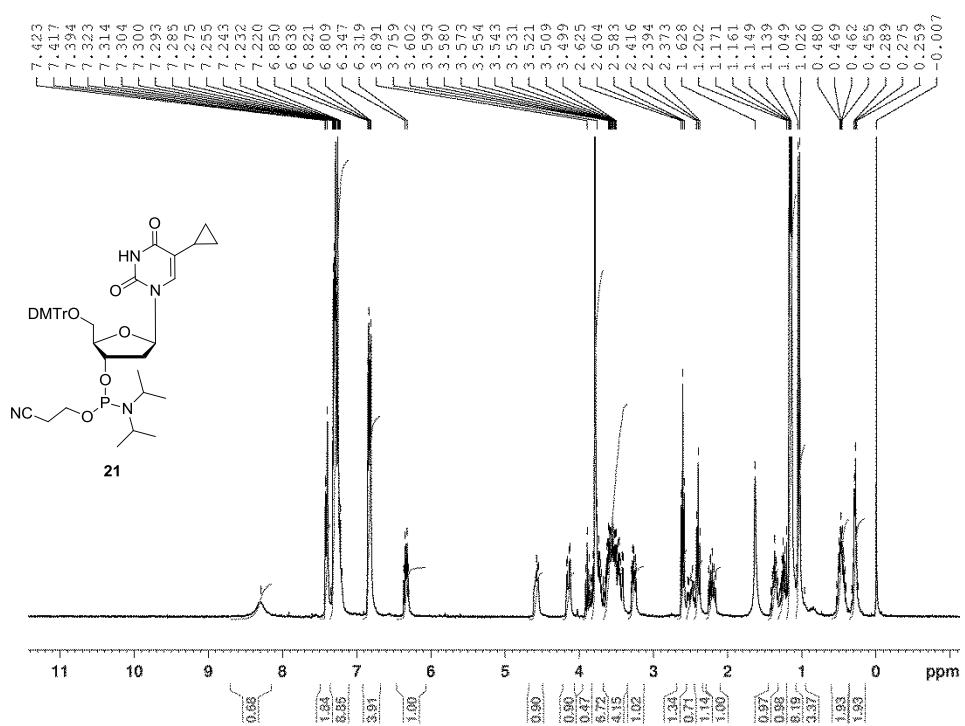
**Fig. S26** <sup>13</sup>C NMR spectrum of compound **18**.



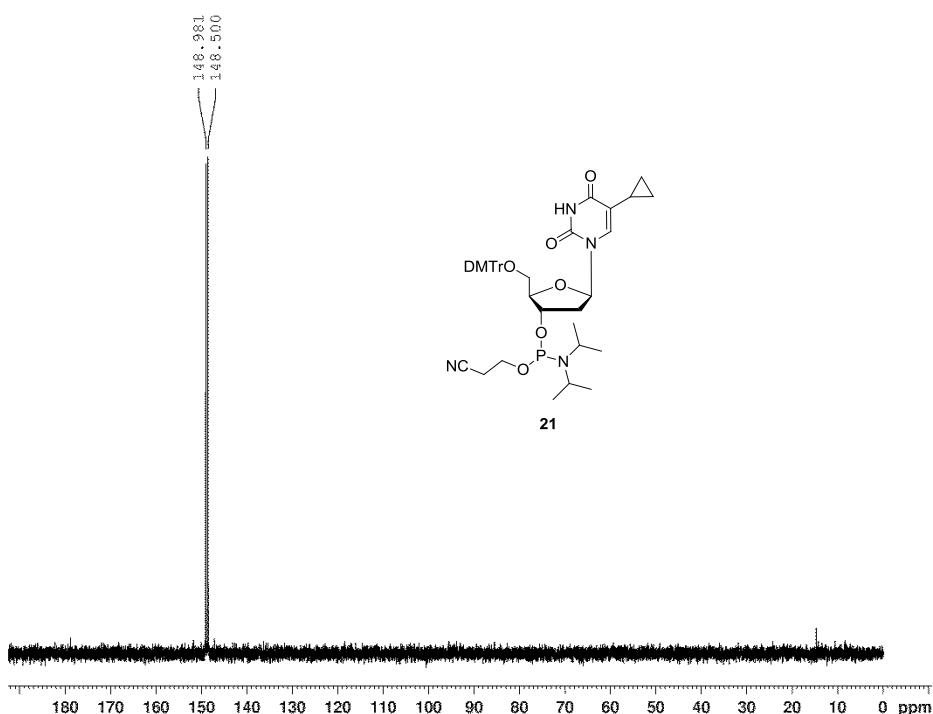
**Fig. S27** DEPT-135 spectrum of compound **18**.



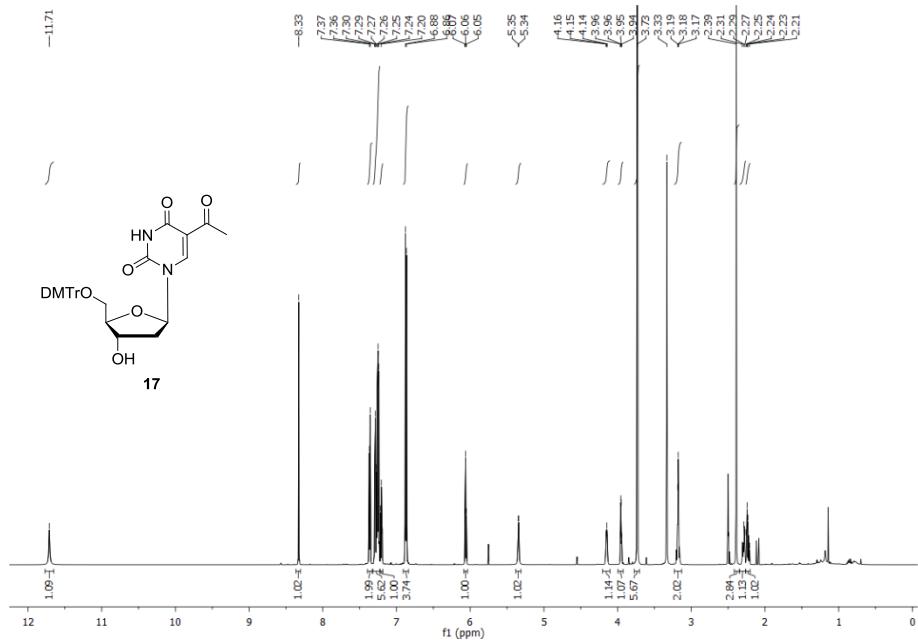
**Fig. S28**  $^1\text{H}$ - $^{13}\text{C}$  gated-decoupled spectrum of compound **18**.



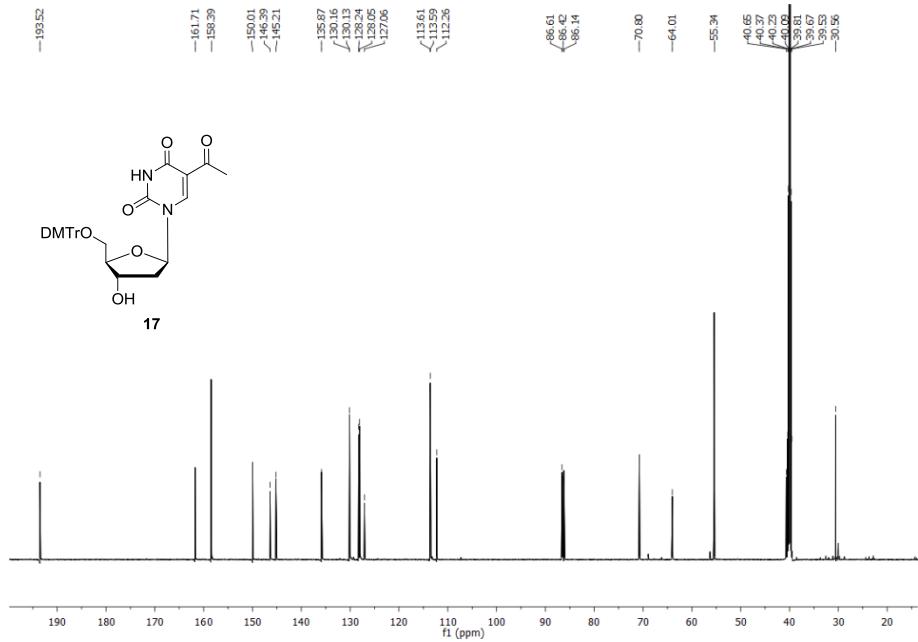
**Fig. S29**  $^1\text{H}$  NMR spectrum of compound **21**.



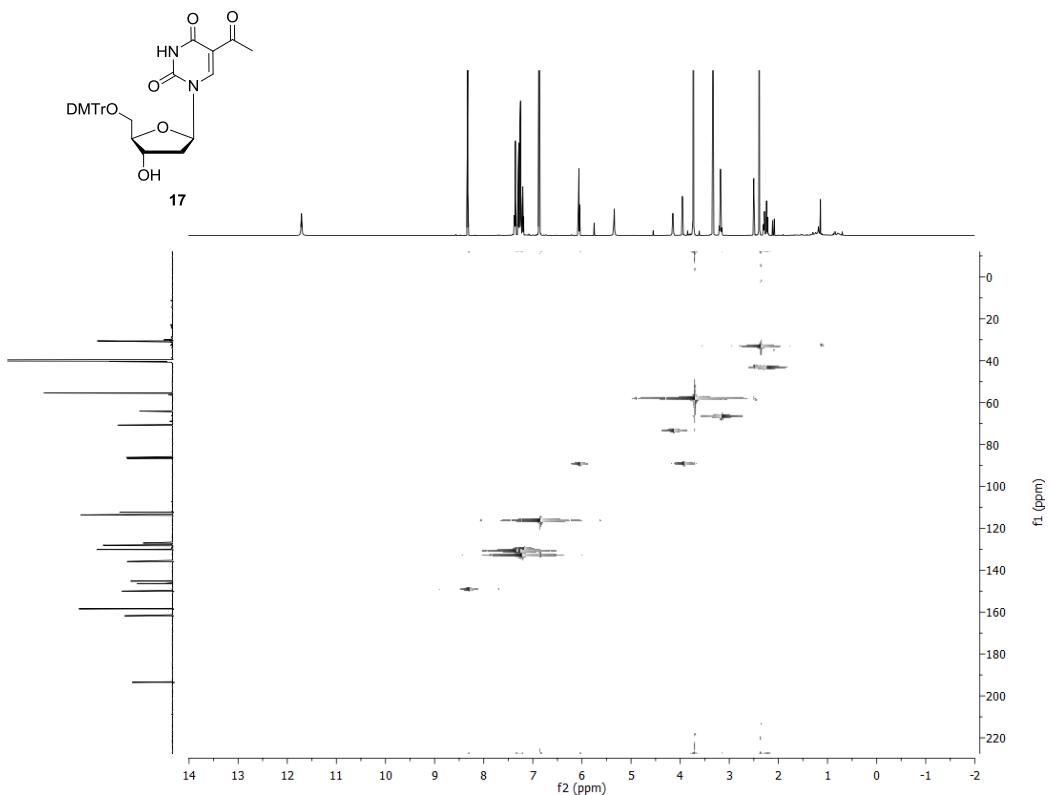
**Fig. S30**  $^{31}\text{P}$  NMR spectrum of compound **21**.



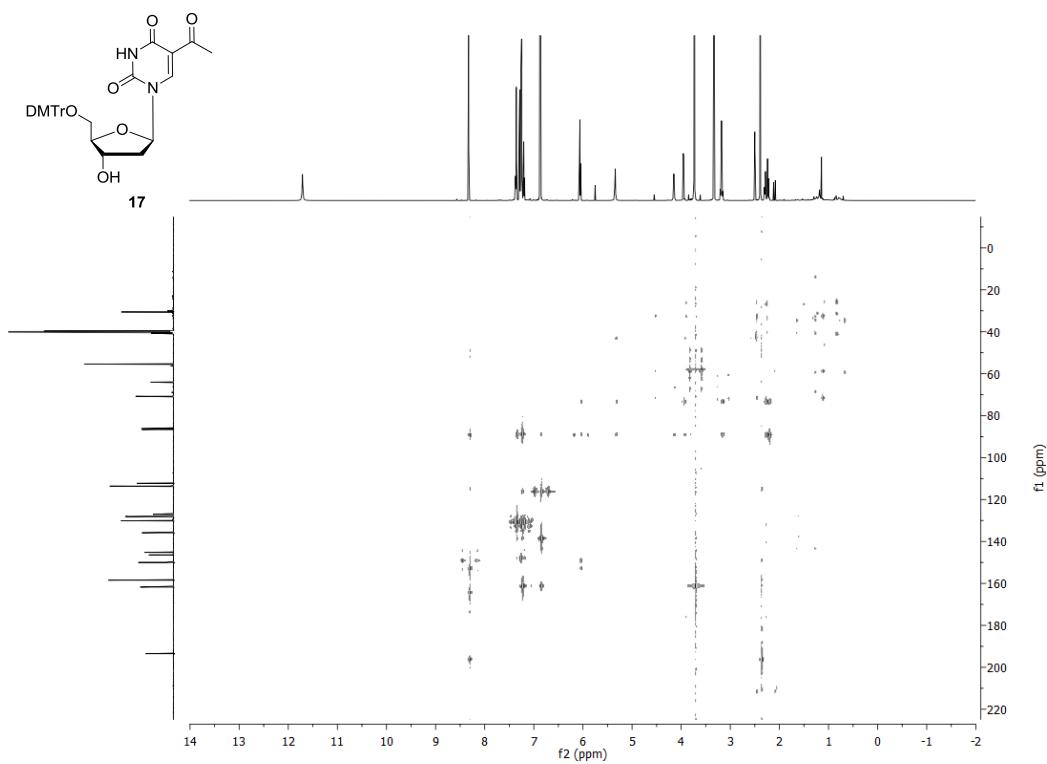
**Fig. S31**  $^1\text{H}$  NMR spectrum of compound **17**.



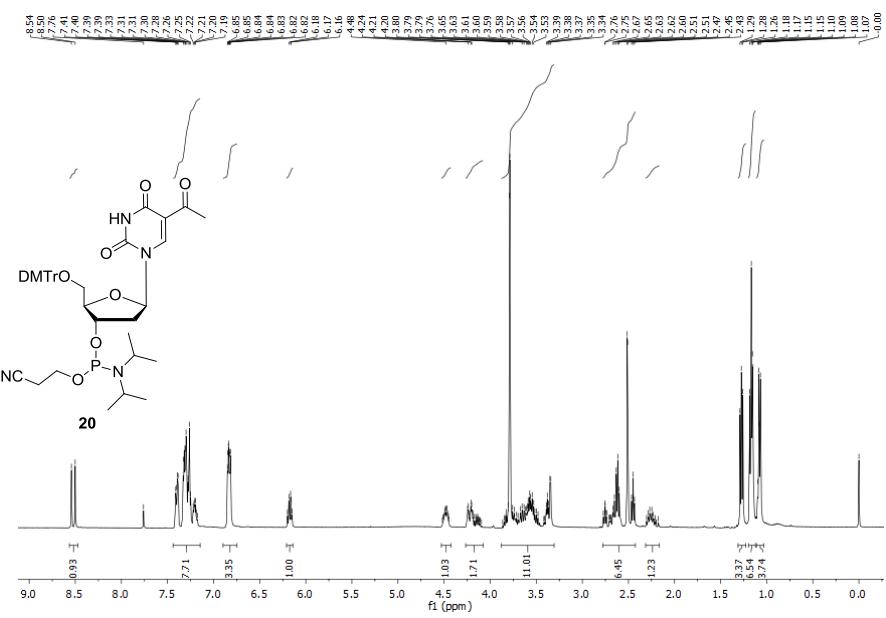
**Fig. S32**  $^{13}\text{C}$  NMR spectrum of compound **17**.



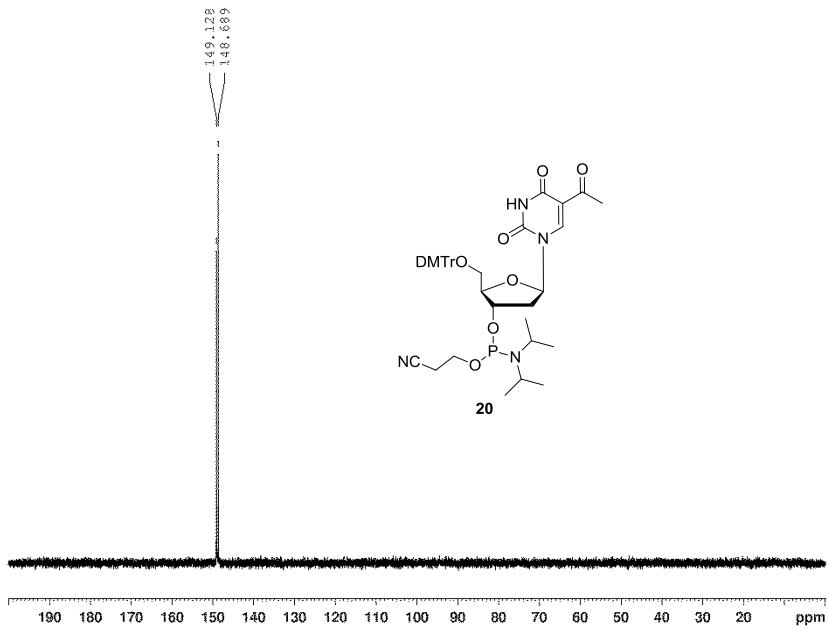
**Fig. S33** HSQC spectrum of compound **17**.



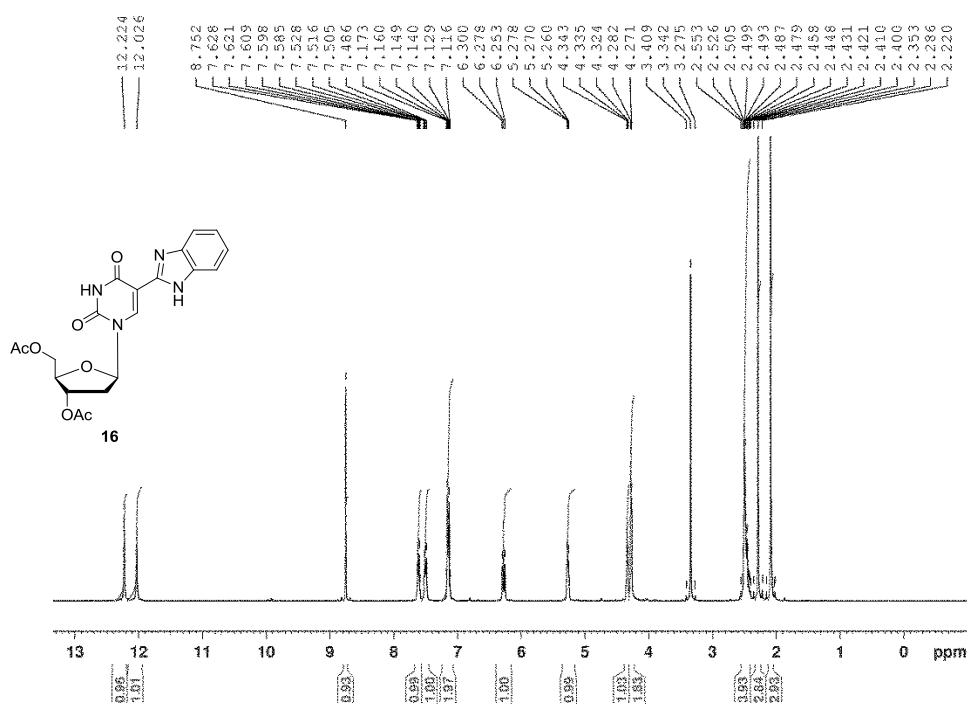
**Fig. S34** HMBC spectrum of compound **17**.



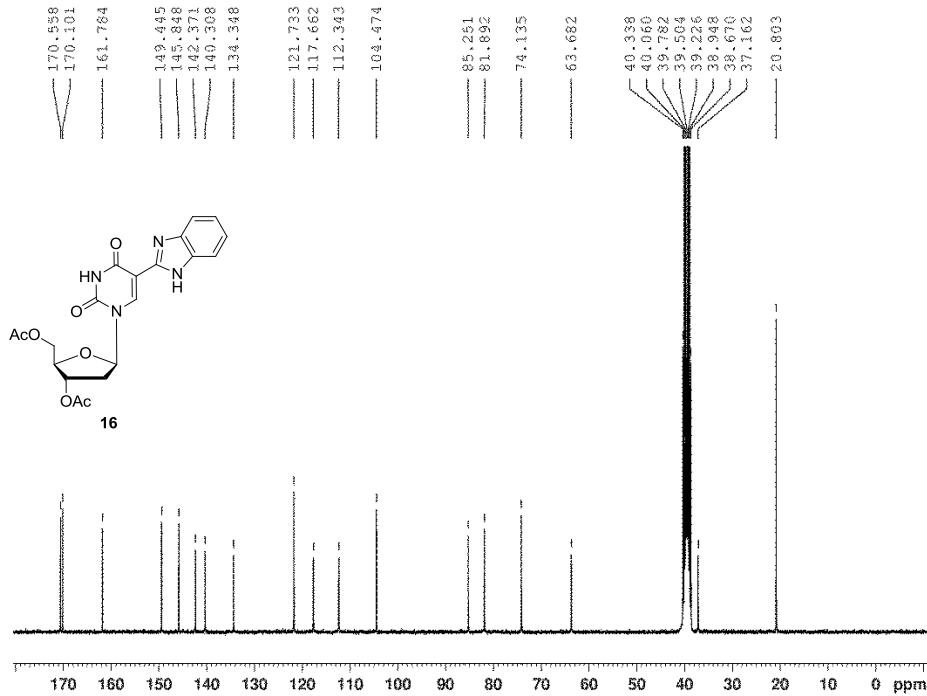
**Fig. S35**  $^1\text{H}$  NMR spectrum of compound **20**.



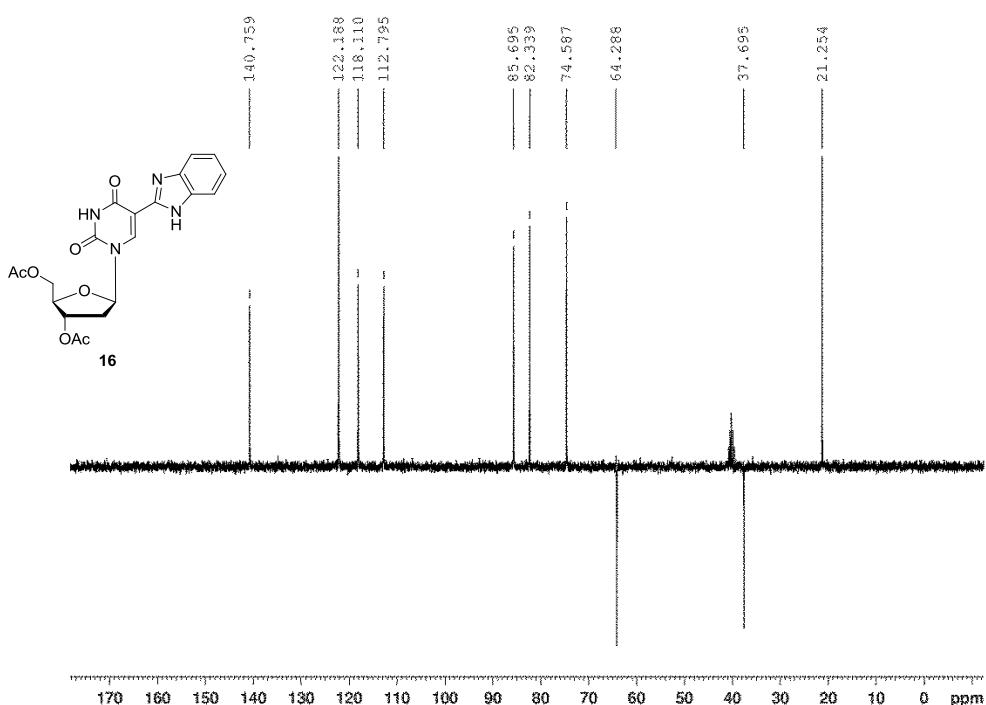
**Fig. S36**  $^{31}\text{P}$  NMR spectrum of compound **20**.



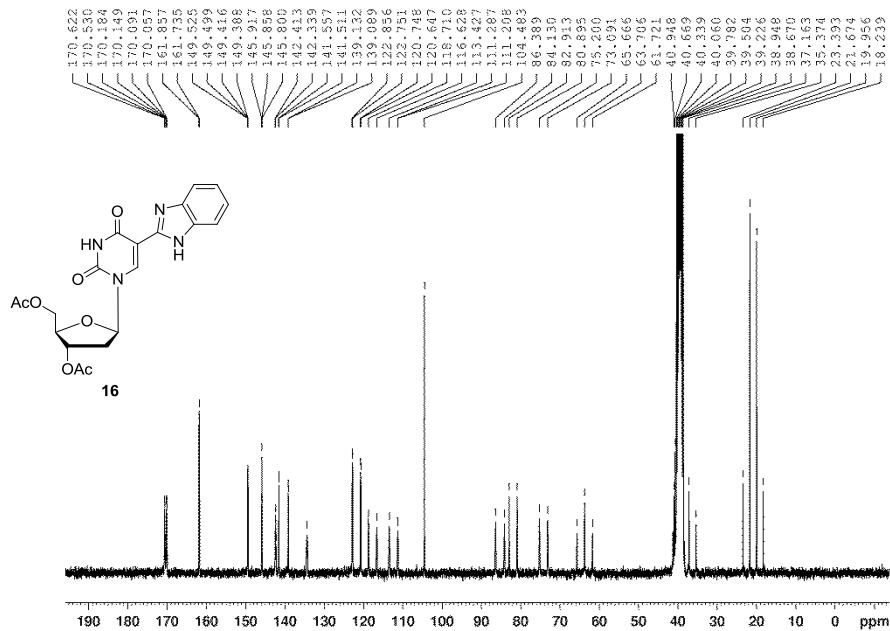
**Fig. S37** <sup>1</sup>H NMR spectrum of compound **16**.



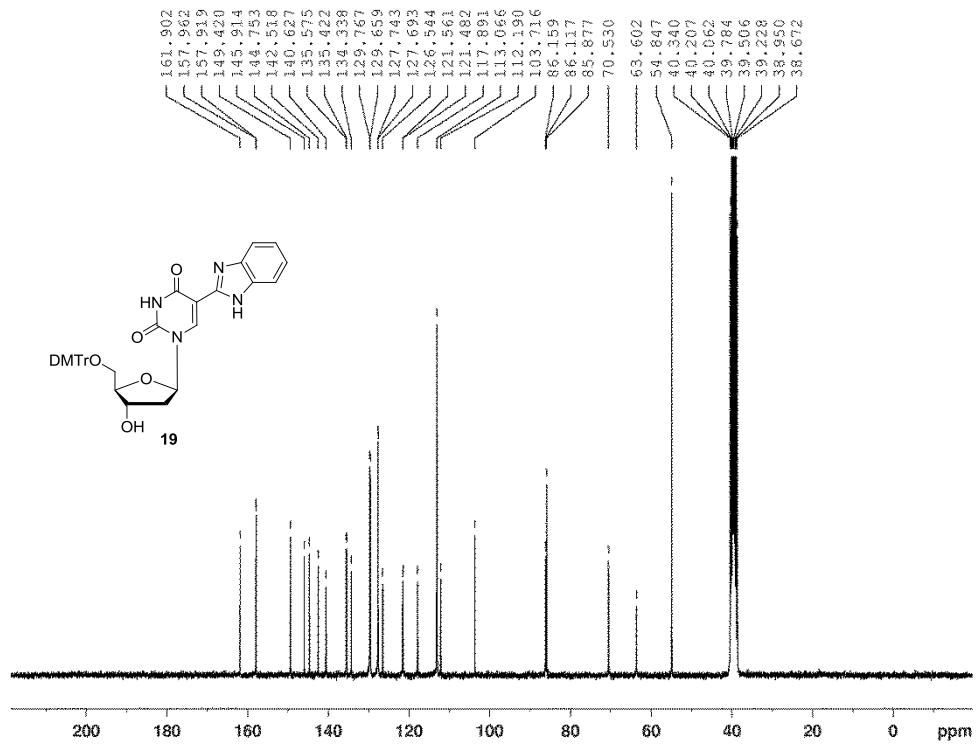
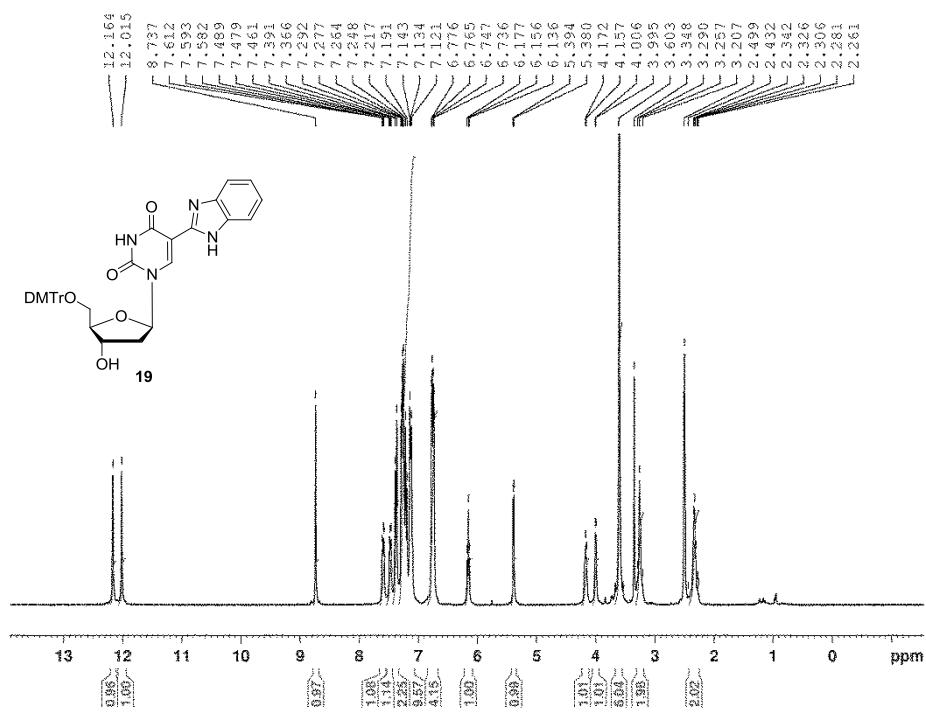
**Fig. S38** <sup>13</sup>C NMR spectrum of compound **16**.

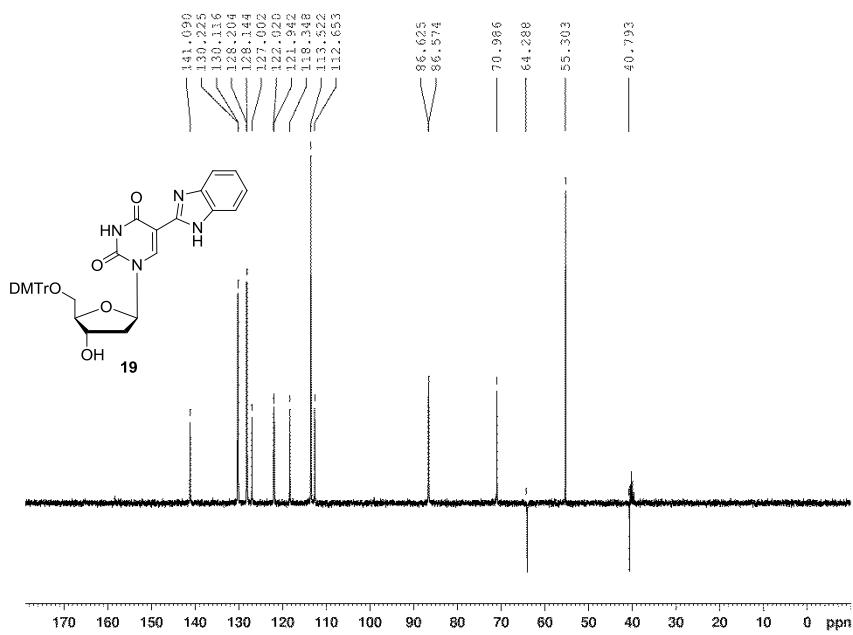


**Fig. S39** DEPT-135 spectrum of compound **16**.

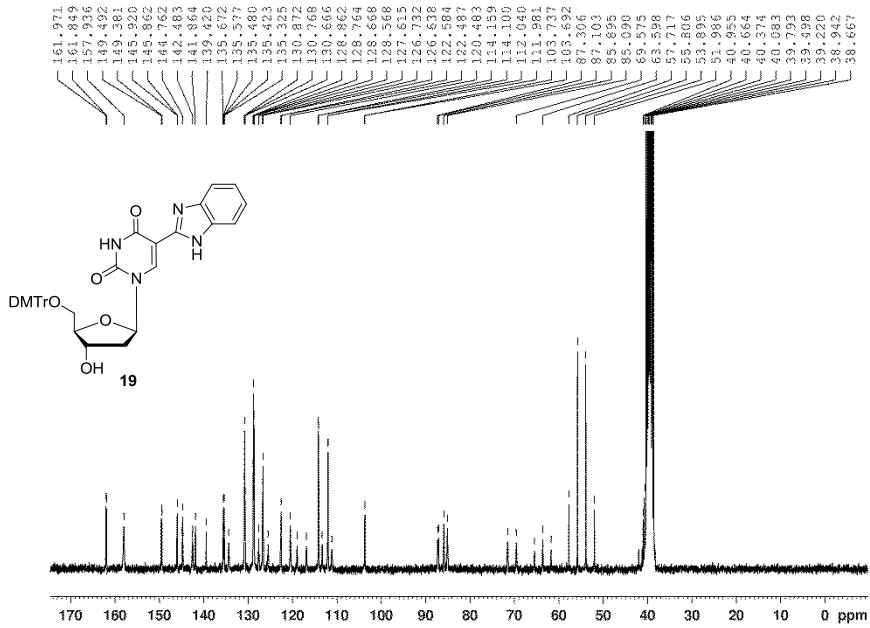


**Fig. S40**  $^1\text{H}$ - $^{13}\text{C}$  gated-decoupled spectrum of compound **16**.

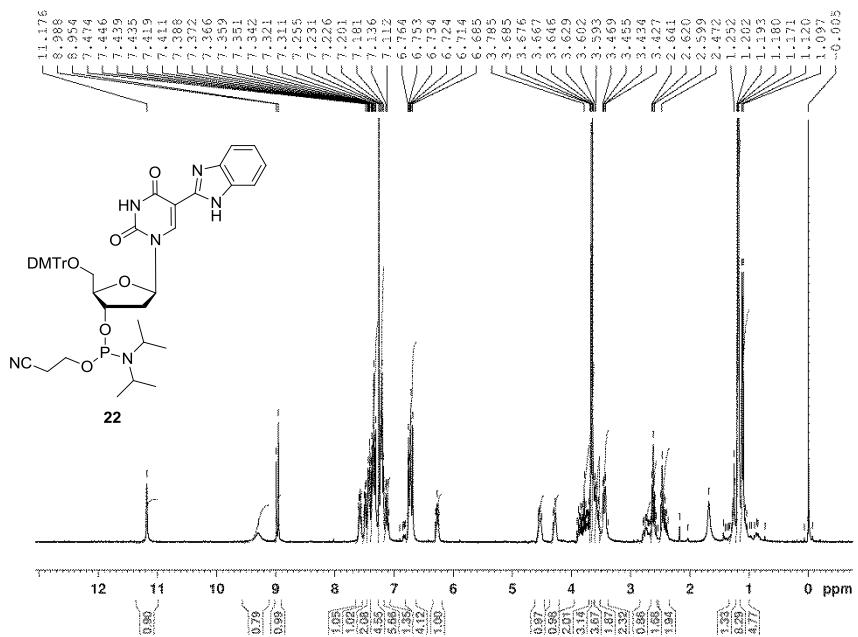




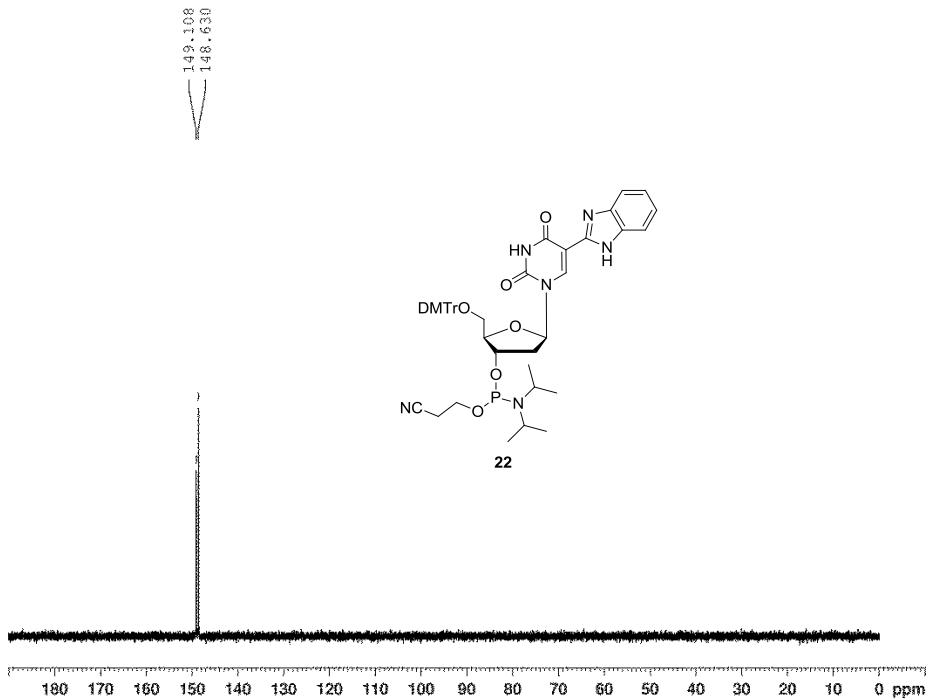
**Fig. S43** DEPT-135 spectrum of compound **19**.



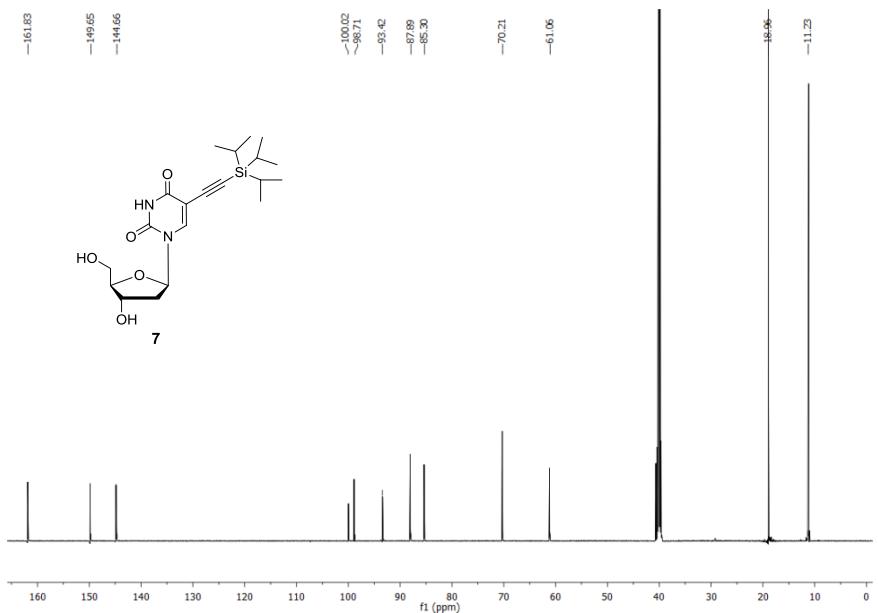
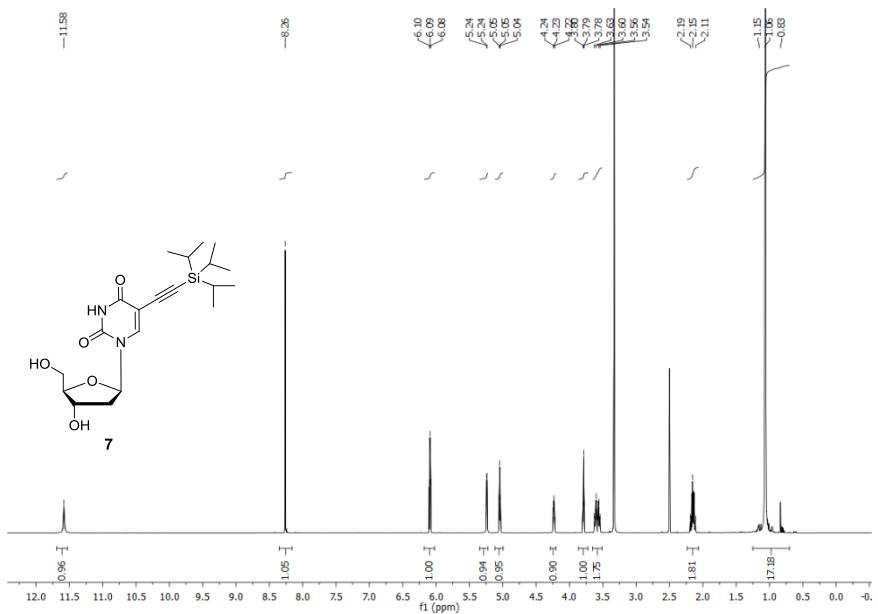
**Fig. S44**  $^1\text{H}$ - $^{13}\text{C}$  gated-decoupled spectrum of compound **19**.



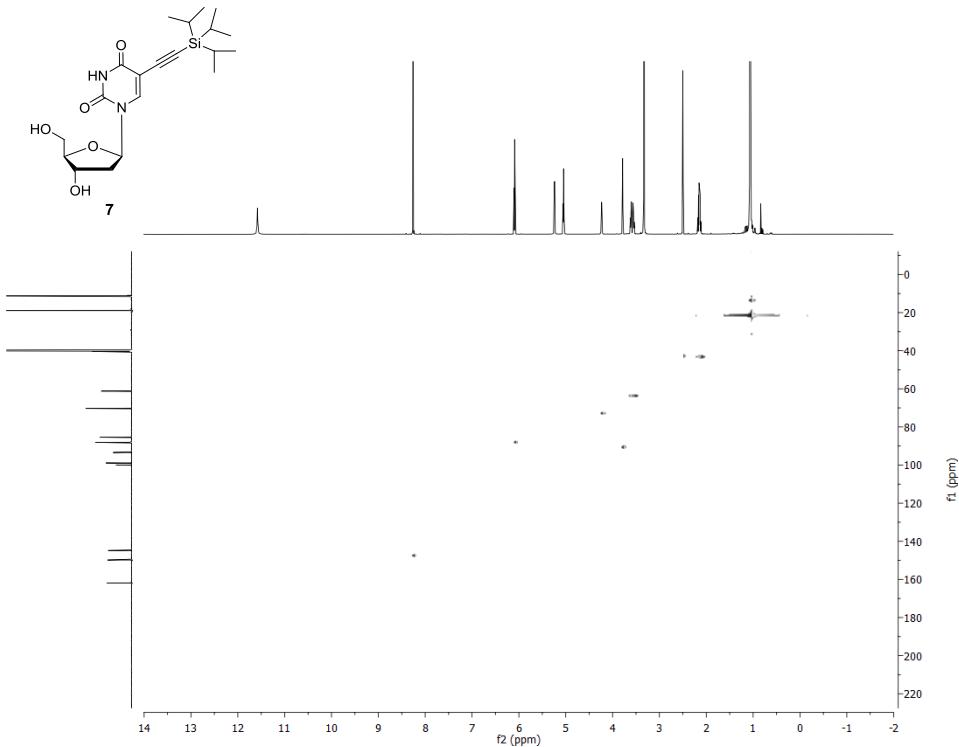
**Fig. S45**  $^1\text{H}$  NMR spectrum of compound **22**.



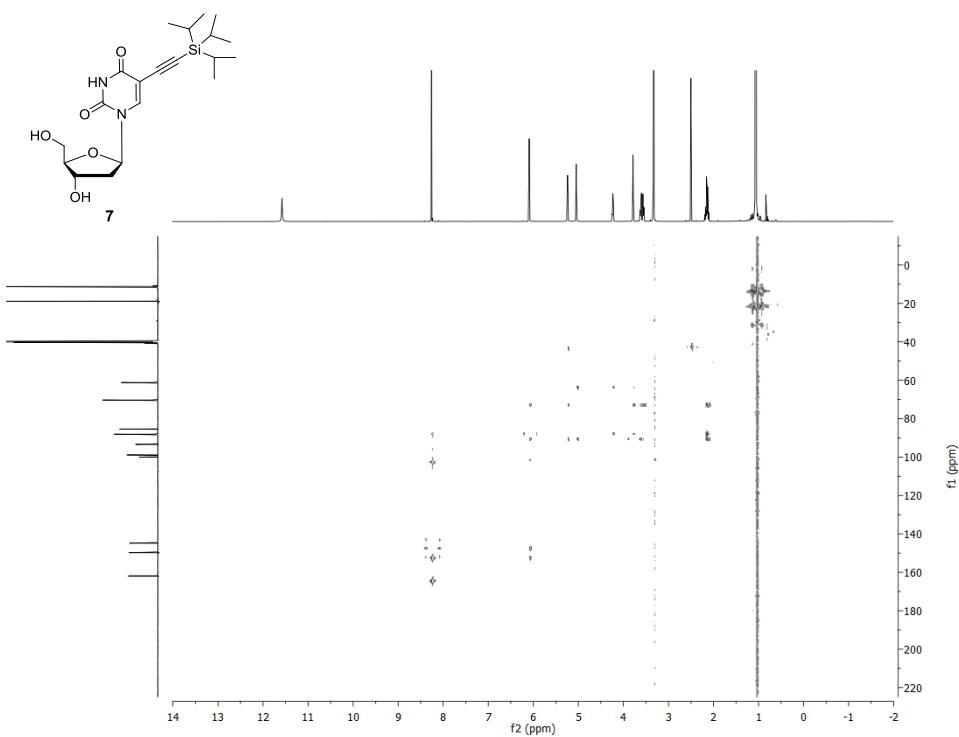
**Fig. S46**  $^{31}\text{P}$  NMR spectrum of compound **22**.



**Fig. S48**  $^{13}\text{C}$  NMR spectrum of compound 7.



**Fig. S49** HSQC spectrum of compound 7.



**Fig. S50** HMBC spectrum of compound 7.