

Supplementary information for:

Tuning the properties of pH responsive Nanoparticles to control cellular interactions in vitro and ex vivo

S. K. Mann^{ab}, A. Dufour^a, J. J. Glass^{cd}, R. De Rose^{cd}, S. J. Kent^{cd}, G. K. Such^b and A. P. R. Johnston^{*ae}

^aDrug Delivery, Disposition and Dynamics, Monash Institute of Pharmaceutical Sciences, Monash University, Parkville, Victoria 3052, Australia. Email: angus.johnston@monash.edu

^bDepartment of Chemistry, The University of Melbourne, Parkville, Victoria 3010, Australia.

^cDepartment of Microbiology and Immunology, The Peter Doherty Institute for Infection and Immunity, The University of Melbourne, VIC 3000, Australia

^dARC Centre of Excellence in Convergent Bio-Nano Science and Technology, The University of Melbourne, Melbourne, VIC 3010, Australia.

^eARC Centre of Excellence in Convergent Bio-Nano Science and Technology, Monash University, Parkville, Australia

Synthesis of pentafluorophenyl methacrylate

Pentafluorophenol (5.0 g, 2.7 mmol), triethylamine (4.1 g, 41 mmol) and 4-dimethylaminopyridine (0.67 g, 27 mmol) were dissolved in dry DCM (30 ml). Nitrogen was bubbled through the solution for 5 min at 0°C. Methacrylic anhydride (6.1 ml, 41 mmol) was added slowly via a syringe with vigorous stirring. The resulting solution was stirred at room temperature for 12 h under a nitrogen atmosphere. The solution was diluted with DCM (20 ml) and washed with 0.1 M HCl (50 mL), water (50 mL) and brine (50 mL). The organic layer was dried over MgSO₄, filtered and concentrated *in vacuo*. The crude product was purified by automated column chromatography with a gradient of *n*-hexane and ethyl acetate to give pentafluorophenyl methacrylate as a colourless liquid (4.2 g, 61%). ¹H NMR (400 MHz; CDCl₃): δ/ppm: 6.45 (quintet, J = 1 Hz), 5.91 (qd, J = 1.6, 1.3 Hz), 2.09 (dd, J = 1.6, 1.0 Hz); ¹⁹F NMR (376 MHz; CDCl₃): δ/ppm: -152.77 (d, 2F, J = 16 Hz, *ortho*), -158.17 (1F, t, J = 22 Hz, *para*), -162.46 (2F, dd, J = 16, 22 Hz, *meta*).

Table S1. Polymer synthesis

Polymer	Monomer:CTA:AIBN	Time (h)
PEGMA ₈	28:1:0.1	16
PEGMA ₃₇	176:1:0.1	16
PEG ₄₅ -b-PDEAEMA ₉₇	100:1:0.1	15
PEGMA ₈ -b-PDEAEMA ₁₀₀	110:1:0.1	18
PEGMA ₃₇ -b-PDEAEMA ₉₀	110:1:0.1	16
PDEAEMA	500:10:1	16
PDEAEMA-r-PPFPMA	500:10:1	16

*All polymers were synthesized in 1,4-Dioxane at 60°C.

Table S2. Polymer characterization

Polymer	PEG/PEGMA Mn (NMR)	PDEAEMA Mn (NMR)	Mn (GPC)	Mw (GPC)	PDI
PEG ₄₅ -b-PDEAEMA ₉₇	2,000	17,700	16,962	19,086	1.13
PEGMA ₈ -b-PDEAEMA ₁₀₀	2,400	18,200	16,818	21,847	1.30
PEGMA ₃₇ -b-PDEAEMA ₉₀	11,100	16,700	55,151	90,038	1.63
PDEAEMA	-	36,000	-	-	-
PDEAEMA-r-PPFPMA	-	49,000	27,940	42,540	1.52

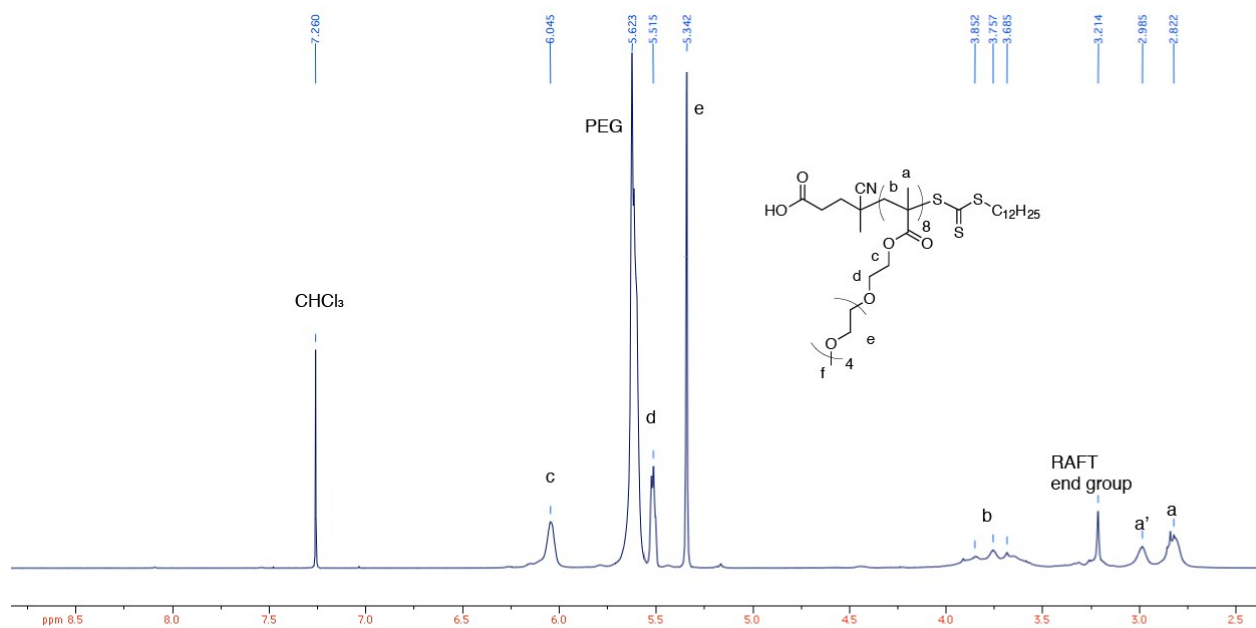


Figure S1. ^1H NMR of PEGMA₈ macro RAFT agent (CDCl_3).

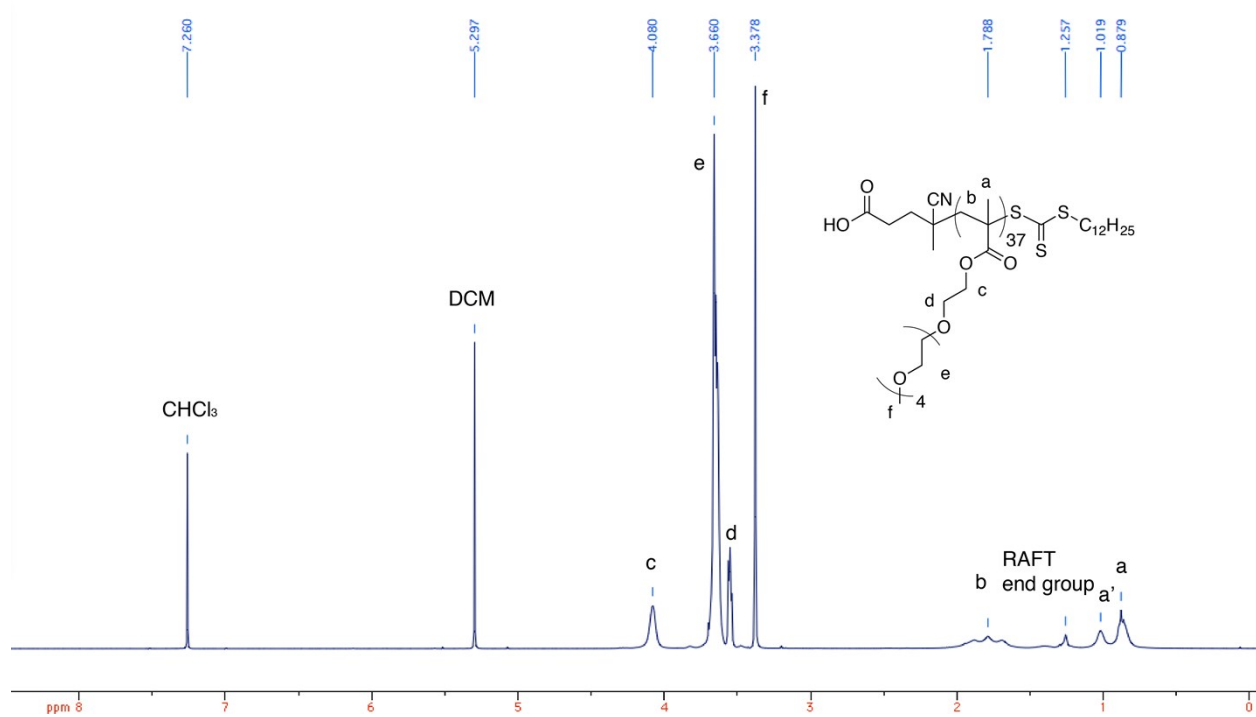


Figure S2. ^1H NMR of PEGMA₃₇ macro RAFT agent (CDCl_3).

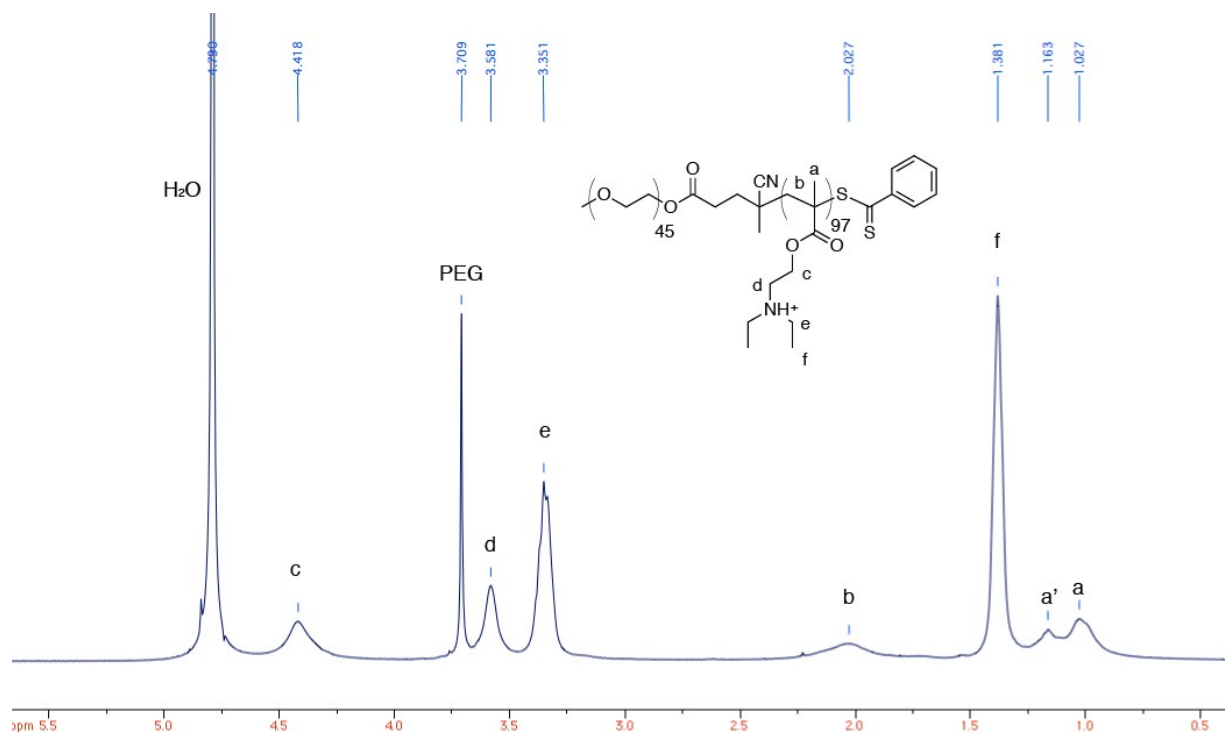


Figure S3. ^1H NMR of $\text{PEG}_{45}\text{-b-PDEAEMA}_{97}$ (D_2O).

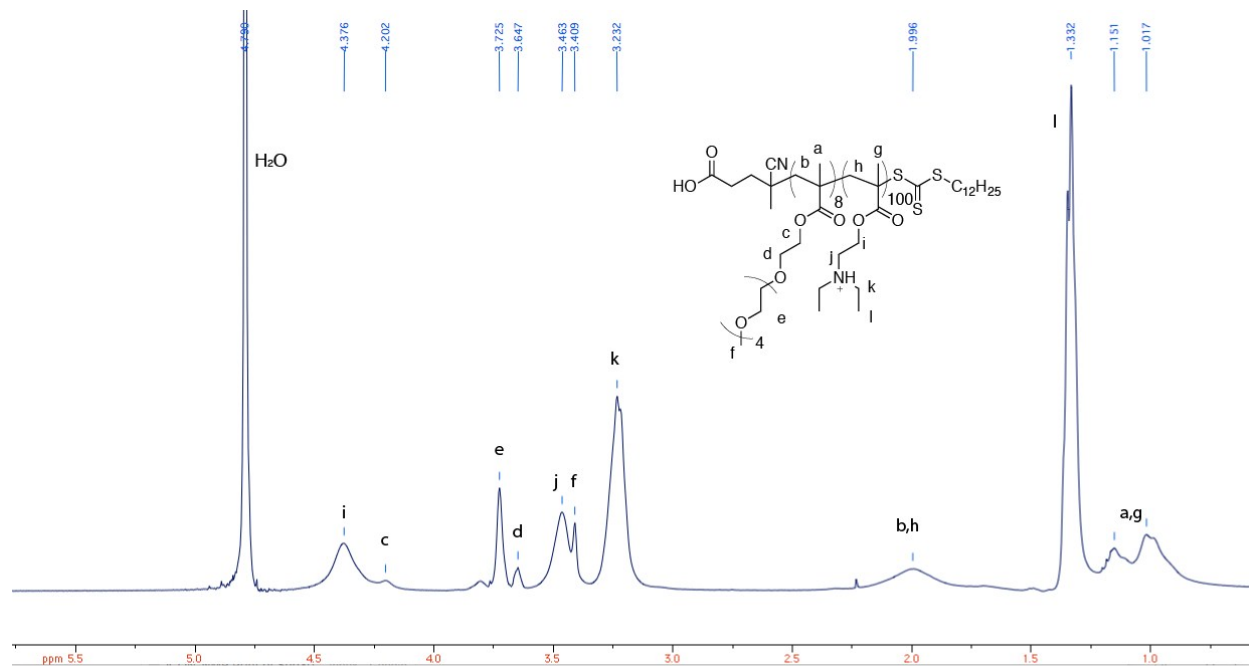


Figure S4. ^1H NMR of $\text{PEGMA}_8\text{-b-PDEAEMA}_{100}$ (D_2O).

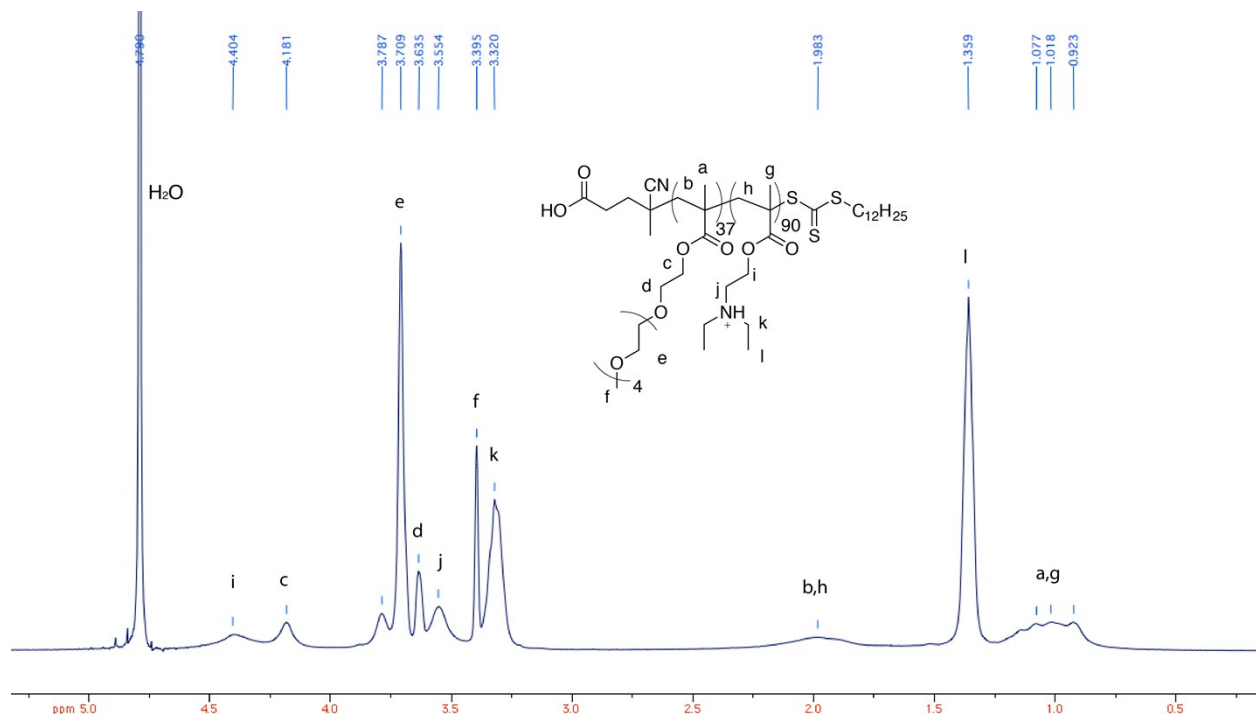


Figure S5. ^1H NMR of PEGMA₃₇-b-PDEAEMA₉₀ (D₂O)

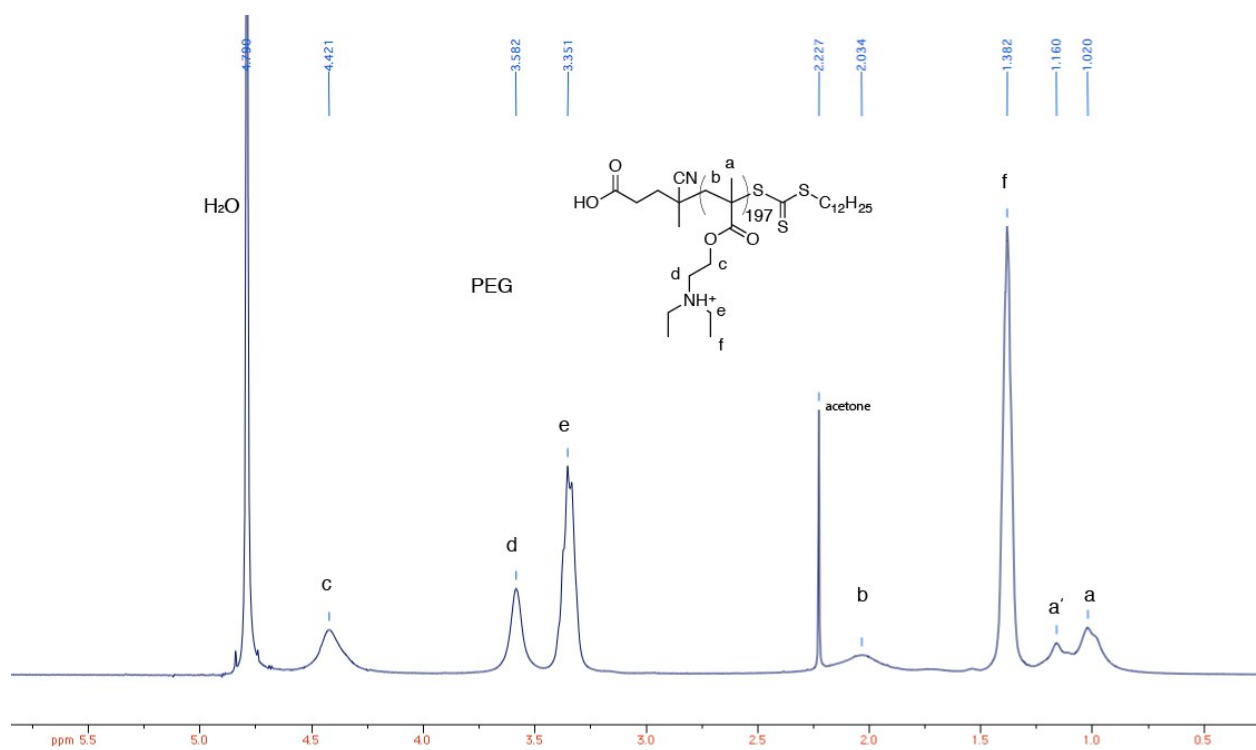


Figure S6. ^1H NMR of PDEAEMA (D₂O).

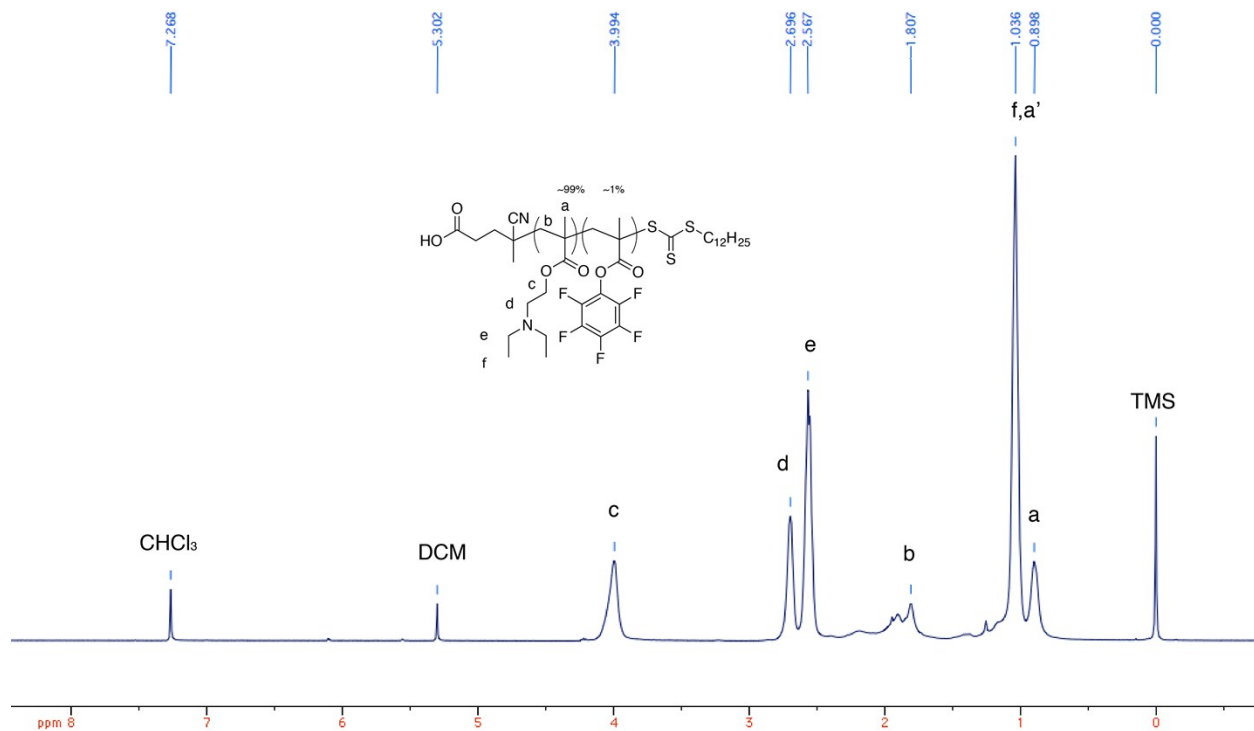


Figure S7. ^1H NMR of P(PDEAEMA-*r*-PFPMA) (CDCl_3).

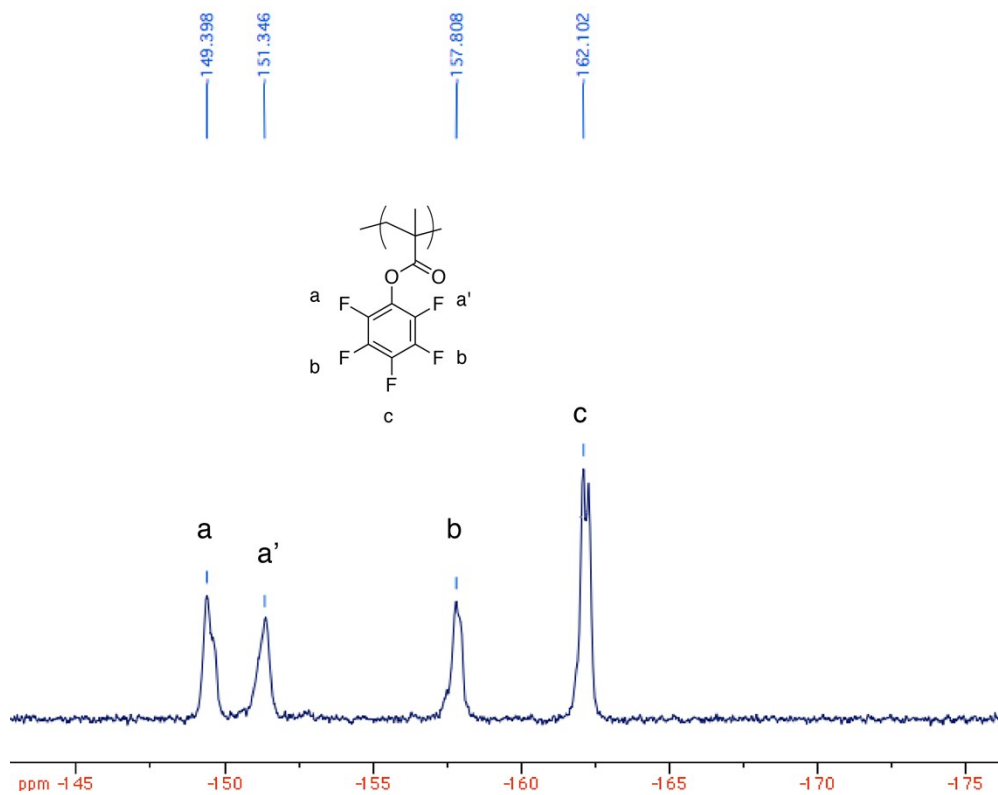


Figure S8. ^{19}F NMR of P(PDEAEMA-*r*-PFPMA).

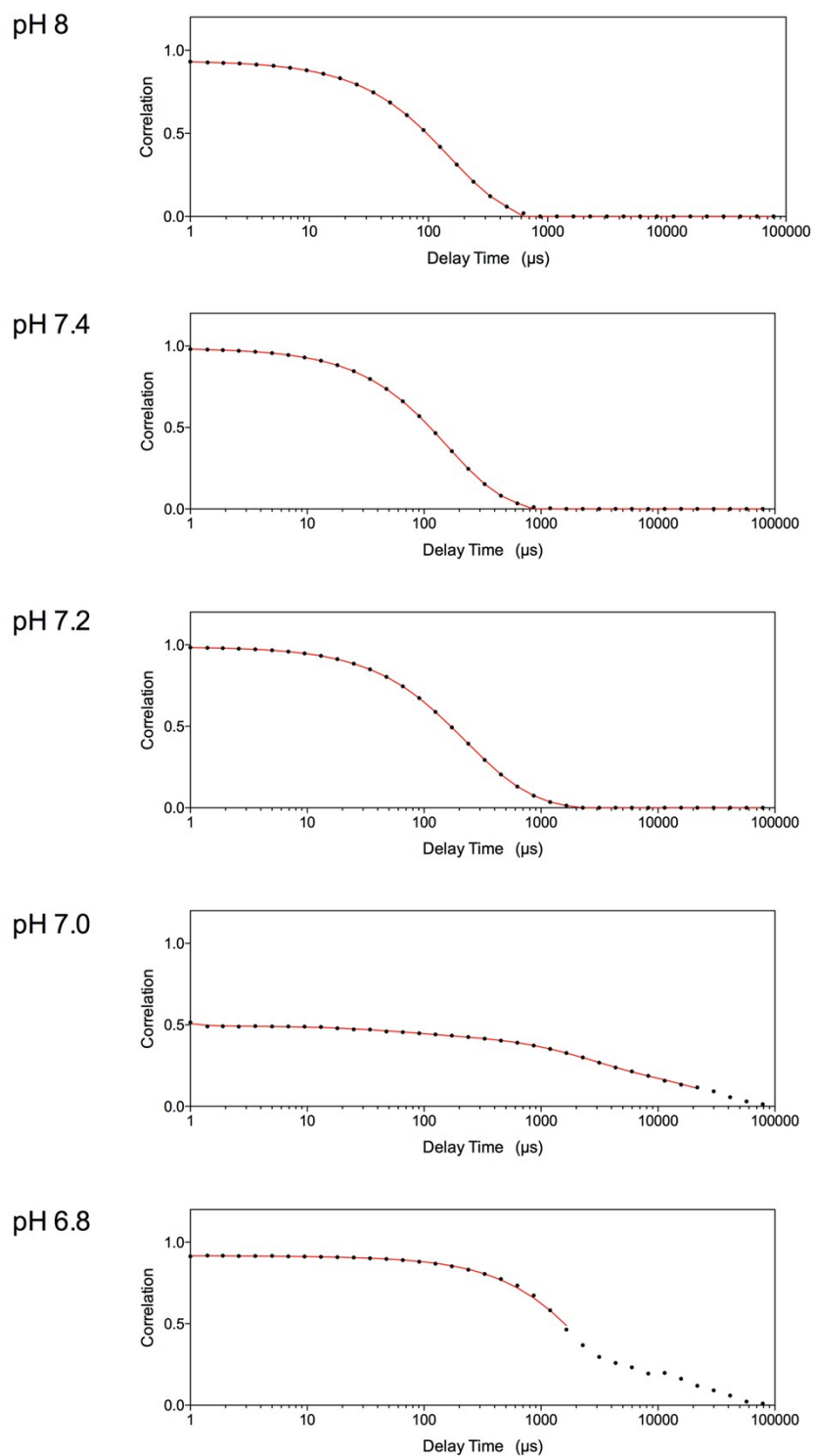


Figure S9. pH-dependent disassembly of nanoparticles analysis by DLS. Correlation function (black circles) and fitting function (red line) for PEGMA₈ NPs adjusted to selected pH values at 37°C.

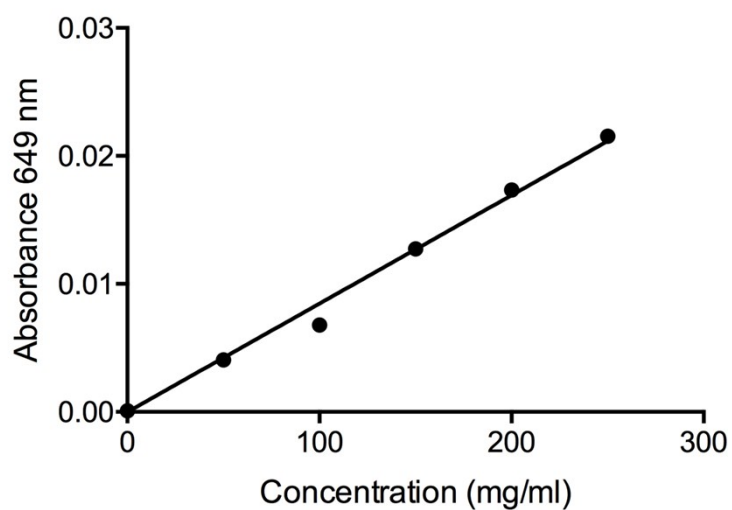
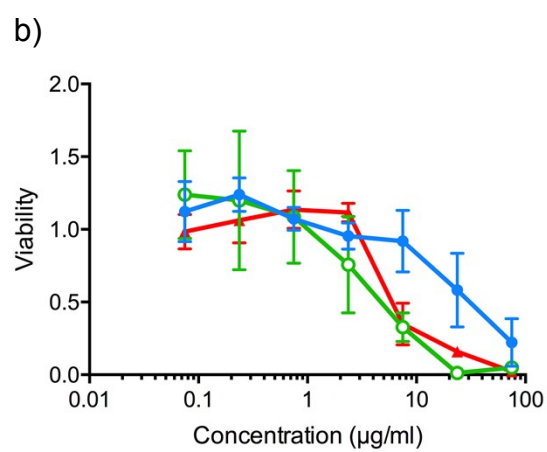
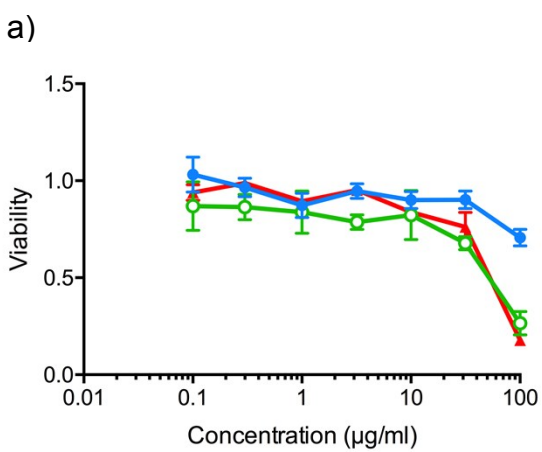


Figure S10. Cy5-labelled PDEAEMA standard curve.



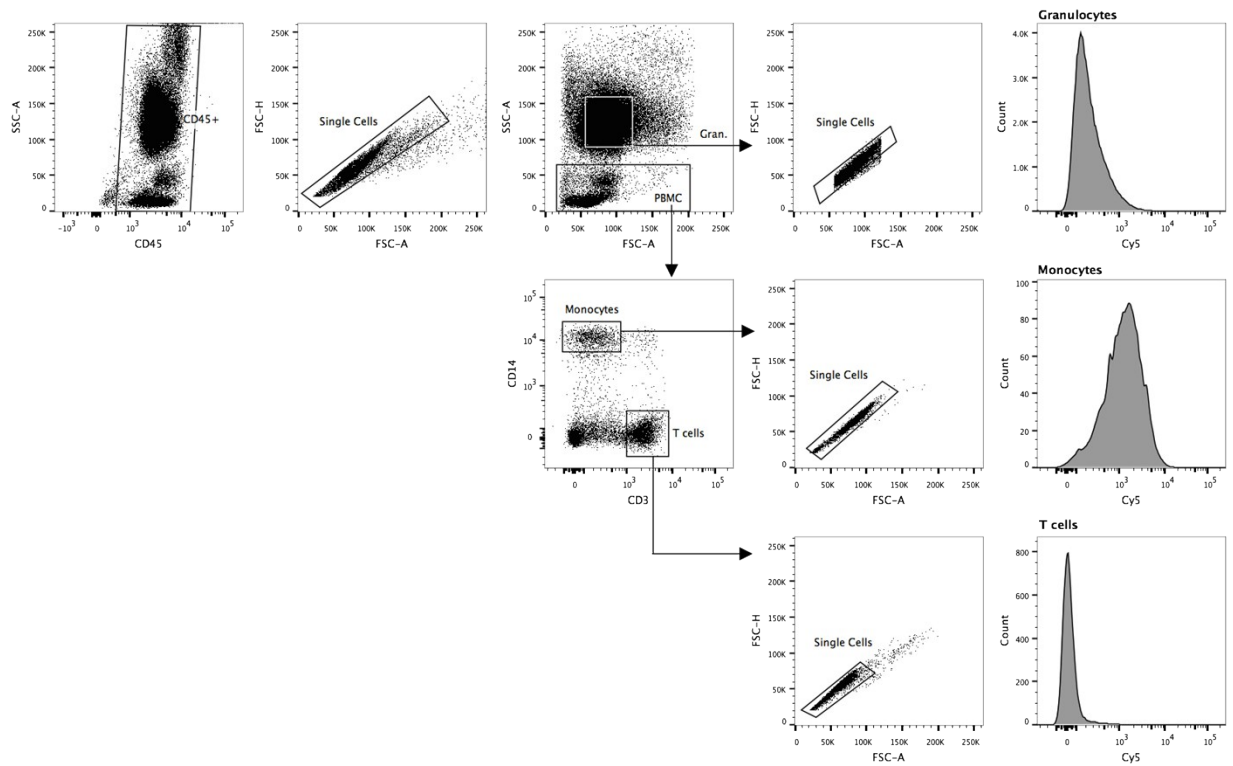
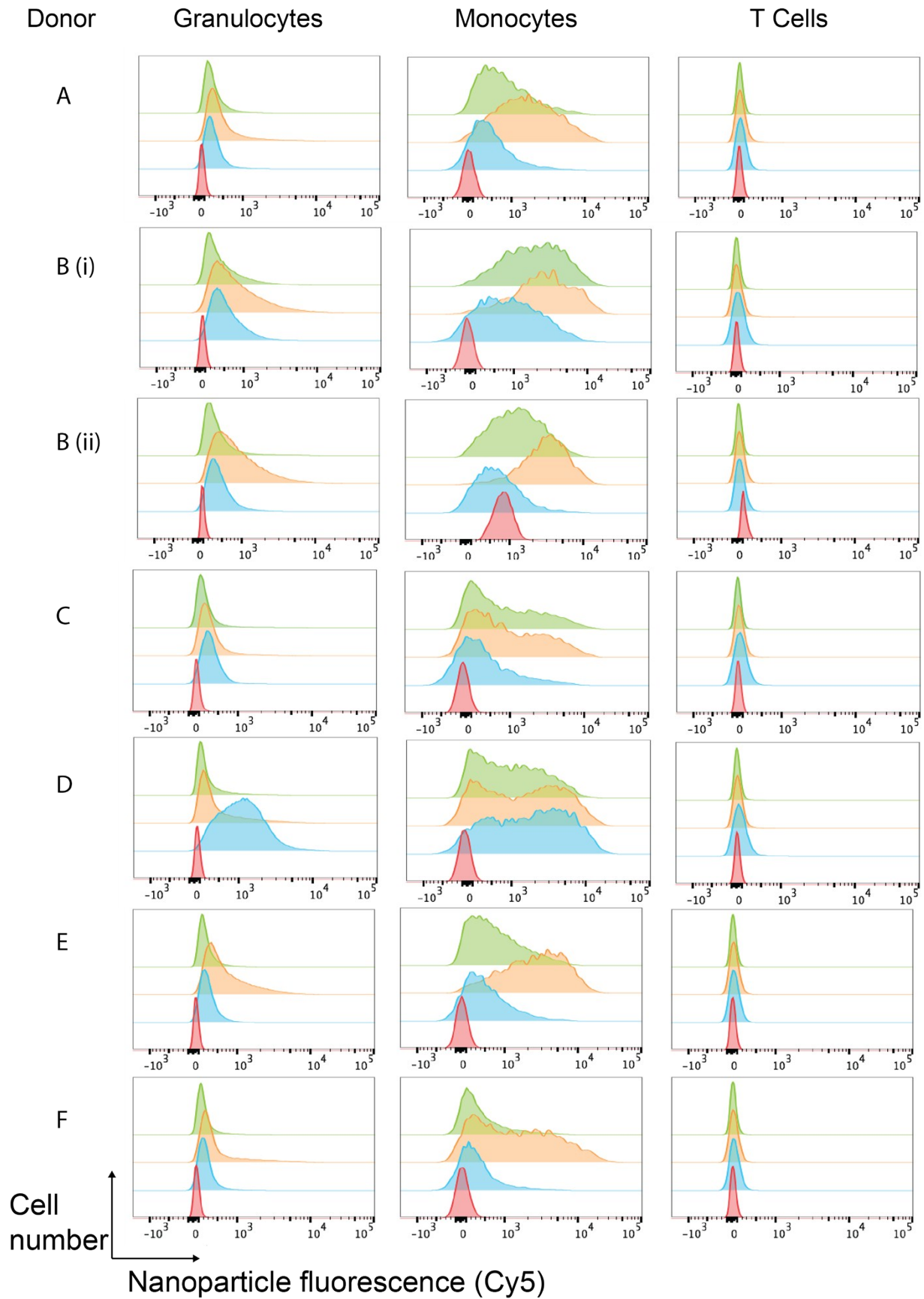


Figure S12. Gating strategy for identification of monocyte, granulocyte and T cell association with nanoparticles. Representative flow cytometry plots from a single donor after association with PEGMA₈. Leukocytes are identified as CD45⁺ before selecting single cells, Granulocytes and peripheral blood mononuclear cells (PBMC) identified by forward (FSC) and side scatter (SSC) properties. PBMC are identified as monocytes (CD14⁺ CD3⁻) and T cells (CD3⁺ CD14⁻). After again gating on single cells, Cy5 signals were analyzed to identify nanoparticle association.



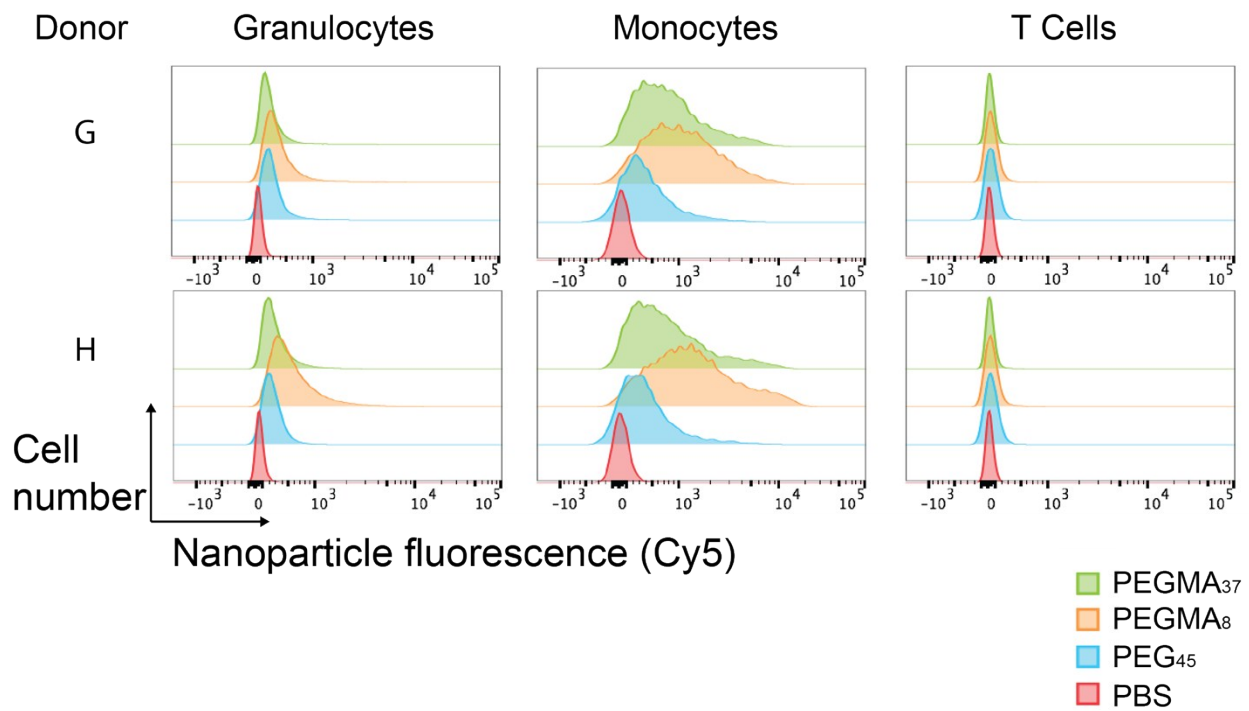


Figure S13. Flow cytometry histograms for monocyte, granulocyte and T-cell association of nanoparticles in 8 donors. The fluorescence was normalized to account for the different concentration of fluorescent labeling of the particles.