Palladium-Catalyzed γ-C(sp³)–H Carbonylation of Aliphatic Amines: Direct approach to synthesis Pyrrolidones via Oxalyl Amide Assistance

Li Zhang, Changpeng Chen, Jian Han, Zhi-Bin* and Yingsheng Zhao*

College of Chemistry, Chemical Engineering and Materials Science, Soochow University, Suzhou 215123, P. R. China

Supporting information

Table of Contents

1. Reagents			
 Instruments Preparation of various directing groups protected substrates Alkenylation of pivaloyl amide Protected N-heteroarenes Alkynylation of pivaloyl amide Protected N-heteroarenes 			
		6. Gram scale reaction and indole derivatives	
		7. A plausible mechanism to explain 3aa	S17
		8. NMR Spectra	S18

1. Reagents: Unless otherwise noted, all reagents were purchased from commercial suppliers and used without further purification. Column chromatography purifications were performed using 300–400 mesh silica gel.

2. Instruments: NMR spectra were recorded on Varian Inova–400 MHz, Inova–300 MHz, Bruker DRX–400 or Bruker DRX–500 instruments and calibrated using residual solvent peaks as internal reference. Multiplicities are recorded as: s = singlet, d = doublet, t = triplet, dd = doublet of doublets, br = broad singlet, m = multiplet. HRMS analyses were carried out using a Bruker micrOTOF–Q instrument or a TOF–MS instrument.

3. Preparation of various directing groups protected substrates

3.1 Preparation of Ac amide



A solution of Indoline (20 mmol, 1.0 equiv) and triethylamine (2.92 ml, 21 mmol, 1.05 equiv) in CH_2Cl_2 (40 mL) was added dropwise to a solution of Acetic anhydride (25 mmol, 1.25 equiv) in CH_2Cl_2 (50 mL) at 0 °C, then the mixture was stirred for 6 hours at room temperature before quenched by water (50 mL). The organic layer was separated and the aqueous layer was extracted with CH_2Cl_2 (20 mL \times 2). The combined organic phase was washed with brine (30 mL), and then dried over anhydrous Na₂SO₄. Evaporation and column chromatography on silica gelafforded corresponding amide substrates as white solid.



¹H NMR (400 MHz, CDCl₃) δ 8.19 (d, J = 8.0 Hz, 1H), 7.18 – 7.13 (m, 2H), 6.98 (t, J = 7.4 Hz, 1H), 3.98 (t, J = 8.5 Hz, 2H), 3.14 (t, J = 8.5 Hz, 2H), 2.17 (s, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 168.2, 142.4, 130.7, 127.0, 124.1, 123.1, 116.4, 48.2, 27.5, 23.7; HRMS Calcd for C₁₀H₁₂NO [M+H⁺]:162.0919; Found: 162.0931.

3.2 Preparation of N-benzoyl amide.



A solution of Indoline (20 mmol, 1.0 equiv) and triethylamine (2.92 ml, 21 mmol, 1.05 equiv) in CH_2Cl_2 (40 mL) was added dropwise to a solution of Benzoyl Chloride (25 mmol, 1.25 equiv) in CH_2Cl_2 (50 mL) at 0 °C, then the mixture was stirred for 6 hours at room temperature before quenched by water (50 mL). The organic layer was separated and the aqueous layer was extracted with CH_2Cl_2 (20 mL \times 2). The combined organic phase was washed with brine (30 mL), and then dried over anhydrous Na₂SO₄. Evaporation and column chromatography on silica gelafforded corresponding amide substrates as white solid.



¹H NMR (400 MHz, CDCl₃) δ 8.21 (s, 1H), 7.56 – 7.54 m(dd, J = 7.7, 1.5 Hz, 2H), 7.49 – 7.43 (m, 3H), 7.21 (d, J = 7.6 Hz, 1H), 7.13 (s, 1H), 7.02 (s, 1H), 4.07 (s, 2H), 3.11 (t, J = 8.3 Hz, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 169.1, 142.7, 141.1, 137.1, 132.5, 130.4, 128.7, 127.3, 127.2, 125.0, 124.0, 119.5, 117.2, 50.8, 28.2; HRMS Calcd for C₁₅H₁₄NO [M+H⁺]:224.1075; Found: 224.1082. **3.3 Preparation of N- Dimethylcarbamyl amide.**

A solution of Indoline (20 mmol, 1.0 equiv) and triethylamine (2.92 ml, 21 mmol, 1.05 equiv) in CH_2Cl_2 (40 mL) was added dropwise to a solution of Dimethylcarbamyl Chloride (25 mmol, 1.25 equiv) in CH_2Cl_2 (50 mL) at 0 °C, then the mixture was stirred for 6 hours at room temperature before quenched by water (50 mL). The organic layer was separated and the aqueous layer was extracted with CH_2Cl_2 (20 mL × 2). The combined organic phase was washed with brine (30 mL), and then dried over anhydrous Na₂SO₄. Evaporation and column chromatography on silica gelafforded corresponding amide substrates as white solid.



¹H NMR (400 MHz, CDCl₃) δ 7.14 – 7.06 (m, 2H), 6.94 – 6.88 (m, 1H), 6.86 – 6.803 (m, 1H), 3.91 – 3.81 (m, 2H), 2.99 (t, *J* = 8.4 Hz, 2H), 2.90 (d, *J* = 1.2 Hz, 6H); ¹³C NMR (101 MHz, CDCl₃) δ 159.8, 143.8, 130.9, 126.5, 124.3, 120.9, 112.9, 49.9, 37.7, 27.6; HRMS Calcd for C₁₁H₁₅N₂O[M+H⁺]:191.1184; Found: 191.1189.

3.4 Preparation of oxalyl amide



A solution of Indoline (20 mmol, 1.0 equiv) and triethylamine (2.92 ml, 21 mmol, 1.05 equiv) in CH_2Cl_2 (40 mL) was added dropwise to a solution of oxalyl Chloride (25 mmol, 1.25 equiv) in CH_2Cl_2 (50 mL) at 0 °C, then the mixture was stirred for 6 hours at room temperature before quenched by water (50 mL). The organic layer was separated and the aqueous layer was extracted with CH_2Cl_2 (20 mL × 2). The combined organic phase was washed with brine (30 mL), and then dried over anhydrous Na₂SO₄. Evaporation and column chromatography on silica gelafforded corresponding amide substrates as white solid.



¹H NMR (400 MHz, CDCl₃) δ 8.18 (d, *J* = 8.8 Hz, 0.60H), 7.26 – 6.96 (m, 3.30H), 4.17 (s, 0.56H), 4.09 (t, *J* = 8.4 Hz, 1.48H), 3.99 -3.92 (m, 0.72H), 3.79 – 3.68 (m, 1H), 3.56 – 3.49 (m, 1H), 3.20 (t, *J* = 8.4 Hz, 1.45H), 3.14 (t, *J* = 8.4 Hz, 0.52H), 1.57 (d, *J* = 6.8 Hz, 1.54H), 1.51 (d, *J* = 6.8 Hz, 4.46H), 1.25 (d, *J* = 6.6 Hz, 4.51H), 1.14 (s, 1.49H); ¹³C NMR (151 MHz, cdcl₃) δ 164.5, 164.1, 162.6, 162.5, 141.9, 139.9, 133.0, 131.9, 127.8, 127.5, 125.8, 124.9, 124.8, 124.4, 117.4, 113.4, 51.1, 50.7, 48.1, 46.8, 46.1, 46.0, 28.3, 26.9, 21.0, 20.7, 20.3, 20.1; HRMS Calcd for C₁₆H₂₃N₂O₂[M+H⁺]:275.1760; Found: 275.1797.

3.5 Preparation of N-heteroarenes substrates.



General procedures for the preparation of oxalyl amide substrates 1a-1r

A solution of N-heteroarenes (20 mmol, 1.0 equiv) and triethylamine (2.92 ml, 21 mmol, 1.05 equiv) in CH_2Cl_2 (40 mL) was added dropwise to a solution of Pivaloyl Chloride (25 mmol, 1.25 equiv) in CH_2Cl_2 (50 mL) at 0 °C, then the mixture was stirred for 6 hours at room temperature before quenched by water (50 mL). The organic layer was separated and the aqueous layer was extracted with CH_2Cl_2 (20 mL \times 2). The combined organic phase was washed with brine (30 mL), and then dried over anhydrous Na₂SO₄. Evaporation and column chromatography on silica gelafforded corresponding amide substrates as white solid or colour liquid.



¹H NMR (400 MHz, CDCl₃) δ 8.27 – 8.21 (m, 1H), 7.18 (t, *J* = 6.9 Hz, 2H), 7.01 (t, *J* = 7.4 Hz, 1H), 4.24 – 4.19 (m, 2H), 3.12 (t, *J* = 8.1 Hz, 2H), 1.37 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 176.5, 144.8, 130.82, 127.3, 124.3, 123.6, 118.4, 49.5, 40.2, 29.3, 27.8; HRMS Calcd for C₁₃H₁₈NO [M+H⁺]:204.1388; Found: 204.1399.



¹H NMR (400 MHz, CDCl₃) δ 7.95 (d, *J* = 7.2 Hz, 1H), 6.94 (s, 2H), 6.77 (d, *J* = 6.1 Hz, 1H), 4.53 (d, *J* = 5.3 Hz, 1H), 3.09 – 2.88 (m, 1H), 2.28 (d, *J* = 14.9 Hz, 1H), 1.13 (s, 9H), 0.95 (d, *J* = 4.6 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 174.9, 142.5, 129.9, 126.1, 124.0, 122.9, 118.6 55.1, 39.6, 36.0, 27.6, 20.9; HRMS Calcd for C₁₄H₂₀NO [M+H⁺]:218.1545; Found: 218.1558.



¹H NMR (400 MHz, CDCl₃) δ 8.12 (d, *J* = 8.1 Hz, 1H), 7.13 (t, *J* = 8.1 Hz, 1H), 6.99 (d, *J* = 8.0 Hz, 1H), 4.26 (t, *J* = 8.3 Hz, 2H), 3.16 (t, *J* = 8.2 Hz, 2H), 1.36 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 176.8, 146.1, 130.1, 129.2, 128.9, 123.5, 116.6, 49.2, 40.3, 28.7, 27.7; HRMS Calcd for C₁₃H₁₇ClNO [M+H⁺]:238.0999; Found: 238.1009.



¹H NMR (400 MHz, CDCl₃) δ 8.17 (d, *J* = 8.1 Hz, 1H), 7.15 (d, *J* = 7.9 Hz, 1H), 7.06 (t, *J* = 8.1 Hz, 1H), 4.25 (t, *J* = 8.3 Hz, 2H), 3.13 (t, *J* = 8.2 Hz, 2H), 1.36 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 176.9, 145.8, 131.4, 129.1, 126.4, 119.0, 117.1, 48.9, 40.3, 30.8, 27.7; HRMS Calcd for C₁₃H₁₇BrNO [M+H⁺]:282.0494; Found: 282.0503.



¹H NMR (400 MHz, CDCl₃) δ 7.36 (t, *J* = 4.7 Hz, 1H), 7.13 (d, *J* = 7.3 Hz, 2H), 7.10 – 7.04 (m, 1H), 3.81 – 3.76 (m, 2H), 2.76 (t, *J* = 7.1 Hz, 2H), 2.03 – 1.95 (m, 2H), 1.29 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 178.8, 141.3, 132.4, 128.9, 126.2, 126.0, 125.5, 45.6, 40.8, 29.4, 26.5, 24.6; HRMS Calcd for C₁₄H₂₀NO [M+H⁺]:218.1545; Found: 218.1555.



¹H NMR (400 MHz, CDCl₃) δ 7.30 (d, *J* = 8.8 Hz, 1H), 6.98 (d, *J* = 7.1 Hz, 2H), 3.79 (t, *J* = 6.2 Hz, 2H), 2.74 (t, *J* = 7.0 Hz, 2H), 2.33 (s, 3H), 2.03 – 1.96 (m, 2H), 1.32 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 177.6, 137.9, 134.1, 131.4, 128.5, 125.8, 125.1, 44.7, 39.9, 28.5, 25.6, 23.7, 20.5; HRMS Calcd for C₁₅H₂₂NO [M+H⁺]:232.1701; Found: 232.1719..



¹H NMR (400 MHz, CDCl₃) δ 7.65 – 7.58 (m, 1H), 7.01 – 6.97 (m, 1H), 6.87 – 6.81 (m, 2H), 4.31 – 4.26 (m, 2H), 3.97 – 3.92 (m, 2H), 1.35 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 176.4, 146.8, 126.8, 125.6, 125.6, 119.8, 117.0, 66.4, 44.2, 39.8, 28.5; HRMS Calcd for C₁₃H₁₈NO₂ [M+H⁺]:220.1338; Found: 220.1346.



¹H NMR (400 MHz, CDCl₃) δ 7.39 – 7.35 (m, 2H), 7.33 – 7.27 (m, 1H), 7.23 – 7.17 (m, 2H), 3.19 (s, 3H), 1.02 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 178.2, 145.4, 129.3, 128.9, 127.9, 41.5, 40.9, 29.6.; HRMS Calcd for C₁₂H₁₈NO [M+H⁺]:192.1388; Found: 192.1392.



¹H NMR (400 MHz, CDCl₃) δ 7.39 – 7.26 (m, 3H), 7.17 – 7.11 (m, 2H), 3.64 – 3.59 (m, 2H), 1.06 (t, *J* = 9.3 Hz, 3H), 0.97 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 177.2, 143.4, 129.9, 129.0, 127.8, 47.8, 40.9, 29.6, 27.2, 12.7; HRMS Calcd for C₁₃H₂₀NO [M+H⁺]:206.1545; Found: 206.1558.



¹H NMR (400 MHz, CDCl₃) δ 7.39 – 7.34 (m, 3H), 7.14 – 7.12 (m, 2H), 5.01 – 4.91 (m, 1H), 1.00 (s, 3H), 0.98 (s, 3H), 0.98 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 177.3, 139.0, 132.2, 128.3, 128.2, 48.09, 41.5, 29.8, 20.9; HRMS Calcd for C₁₄H₂₂NO [M+H⁺]:220.1701; Found: 220.1710.



¹H NMR (400 MHz, CDCl₃) δ 7.21 – 7.14 (m, 2H), 7.12 (d, *J* = 7.2 Hz, 1H), 7.02 – 6.96 (m, 2H), 3.44 – 3.37 (m, 2H), 1.36 – 1.29 (m, 2H), 1.12 – 1.03 (m, 2H), 0.81 (s, 9H), 0.67 (t, *J* = 7.4 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 176.6, 143.3, 129.3, 128.5, 127.3, 52.4, 40.5, 29.1, 29.0, 19.6, 13.4; HRMS Calcd for C₁₅H₂₄NO [M+H⁺]:234.1858; Found: 134.1861.



¹H NMR (400 MHz, CDCl₃) δ 7.36 – 7.32 (m, 4H), 7.24 (d, *J* = 1.8 Hz, 3H), 7.23 – 7.21 (m, 3H), 1.15 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 179.6, 144.5, 129.3, 129.1, 128.4, 126.9, 117.8, 41.7, 29.7; HRMS Calcd for C₁₇H₂₀NO [M+H⁺]:254.1545; Found: 254.1557.



¹H NMR (400 MHz, CDCl₃) δ 7.32 – 7.27 (m, 3H), 7.25 (t, *J* = 6.7 Hz, 3H), 7.21 – 7.16 (m, 2H), 7.04 – 6.98 (m, 2H), 4.85 (s, 2H), 1.07 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 177.6, 143.3, 137.9, 129.9, 128.9, 128.8, 128.3, 127.9, 127.2, 56.4, 41.1, 29.6; HRMS Calcd for C₁₈H₂₂NO [M+H⁺]:268.1701; Found: 268.1709.



¹H NMR (400 MHz, CDCl₃) δ 7.02 – 6.93 (m, 2H), 6.93 – 6.88 (m, 1H), 6.88 – 6.82 (m, 1H), 2.83 (s, 3H), 1.98 (s, 3H), 0.75 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 177.2, 143.3, 135.2, 130.7, 128.4, 127.6, 126.07, 40.1, 39.0, 28.4, 17.2; HRMS Calcd for C₁₃H₂₀NO [M+H⁺]:206.1545; Found: 206.1553.



¹H NMR (400 MHz, CDCl₃) δ 7.22 – 7.20 (m, 2H), 7.09 (d, J = 1.5 Hz, 2H), 3.14 (s, 3H), 1.01 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 177.7, 158.0 (J_{C-F} =248), 132.3(J_{C-F} =12), 130.4, 129.3 (J_{C-F} =8), 124.0(J_{C-F} = 4), 116.1 (J_{C-F} =20), 40.1, 39.5, 28.3; HRMS Calcd for C₁₂H₁₇FNO [M+H⁺]:210.1294; Found: 210.1299.



¹H NMR (400 MHz, CDCl₃) δ 7.16 (d, *J* = 8.0 Hz, 2H), 7.08 (d, *J* = 8.1 Hz, 2H), 3.17 (s, 3H), 2.36 (s, 3H), 1.02 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 178.2, 142.7, 137.6, 129.8, 128.5, 41.4, 40.8, 29.5, 21.1; HRMS Calcd for C₁₃H₂₀NO [M+H⁺]:206.1545; Found: 206.1557.



¹H NMR (400 MHz, CDCl₃) δ 7.22 – 7.10 (m, 2H), 7.03 (t, *J* = 8.6 Hz, 2H), 3.15 (s, 3H), 1.00 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 177.9, 161.6 (*J*_{C-F} = 246), 141.2 (*J*_{C-F} = 4), 130.1(*J*_{C-F} = 18), 116.0(*J*_{C-F} = 12), 41.3, 40.6, 29.4; HRMS Calcd for C₁₂H₁₇FNO [M+H⁺]:210.1294; Found: 210.1302.



¹H NMR (400 MHz, CDCl₃) δ 7.16 (t, *J* = 7.7 Hz, 1H), 7.03 (d, *J* = 7.6 Hz, 1H), 6.96 – 6.86 (m, 2H), 3.09 (s, 3H), 2.26 (s, 3H), 0.94 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 178.0, 145.2, 139.2, 129.3, 129.0, 128.5, 125.7, 41.3, 40.8, 29.6, 21.3; HRMS Calcd for C₁₃H₂₀NO [M+H⁺]:206.1545; Found: 206.1552.

4. Alkenylation of pivaloyl amide Protected N-heteroarenes

4.1 Substrates Scope of various directing groups protected amides



A mixture of amide (0.2 mmol, 1 equiv), olefin (2 equiv), $[RuCl_2(p-cymene)]_2$ (6.4 mg, 0.05 equiv), AgSbF₆ (13.7 mg, 0.2 equiv), Cu(OAc)₂·H₂O (40 mg, 1 equiv) and THF (1mL) in a 15 mL glass tube (sealed with PTFE cap) was heated at 100 °C for 20 hours. The reaction mixture was cooled to room tempreture, and concentrated in vacuo. The resulting residue was purified by column chromatography on silica gel to give the alkenylated product.

4.11 DG= Ac



¹H NMR (400 MHz, CDCl₃) δ 7.66 (d, *J* = 14.8 Hz, 1H), 7.42 (d, *J* = 7.8 Hz, 1H), 7.23 (d, *J* = 7.2 Hz, 1H), 7.11 (t, *J* = 7.6 Hz, 1H), 6.34 (d, *J* = 15.6 Hz, 1H), 4.27 – 4.22 (m, 2H), 4.17 (d, *J* = 4.8 Hz, 2H), 3.03 (t, *J* = 7.3 Hz, 2H), 2.23 (s, 3H), 1.31 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 170.0, 167.1, 143.0, 141.9, 138.8, 134.4, 125.9, 125.6, 125.4, 117.4, 60.5, 51.0, 29.7, 29.3, 14.3; HRMS Calcd for C₁₅H₁₈NO₃ [M+H⁺]:260.1287; Found: 260.1298.

4.12 DG= benzoyl



¹H NMR (400 MHz, CDCl₃) δ 8.36 (d, *J* = 7.9 Hz, 0.56H), 7.77 (d, *J* = 15.9 Hz, 0.58H), 7.72 – 7.66 (m, 1H), 7.57 – 7.34 (m, 3H), 7.29 (d, *J* = 7.5 Hz, 0.56H), 7.20 (t, *J* = 8.8 Hz, 1H), 7.10 (t, *J* = 7.3 Hz, 0.62H), 6.88 (t, *J* = 7.2 Hz, 0.35H), 6.75 (t, *J* = 7.3 Hz, 0.35H), 6.47 (d, *J* = 15.9 Hz, 0.61H), 6.33 (d, *J* = 15.8 Hz, 0.33H), 5.60 (d, *J* = 8.0 Hz, 0.32H), 4.38 -4.32(m, 0.68H), 4.22 – 4.16 (m, 2H), 3.70 (t, *J* = 8.2 Hz, 1H), 3.13 (dt, *J* = 31.3, 8.1 Hz, 2H), 1.25 (t, *J* = 6.8 Hz, 4H).; ¹³C NMR (101 MHz, CDCl₃) δ 167.1, 166.5, 165.9, 165.8, 141.9 140.6, 140.3, 140.2, 137.6, 136.8, 133.0, 131.4, 130.6, 130.0, 129.8, 129.7, 129.2, 127.2, 127.0, 126.8 126.6, 126.5, 126.2, 125.1, 124.2, 124.1, 123.1 120.8 120.7, 117.2, 113.5, 76.9, 76.8, 76.6, 76.3, 60.2, 49.6, 47.8, 29.2, 27.7, 26.4, 13.7; HRMS Calcd for C₂₀H₂₁NO₃ [M+H⁺]:322.1443; Found: 322.1452.

4.13, DG= Dimethylcarbamyl



¹H NMR (400 MHz, CDCl₃) δ 7.51 (d, *J* = 15.9 Hz, 1H), 7.35 (d, *J* = 7.8 Hz, 1H), 7.18 (dd, *J* = 7.3, 0.9 Hz, 1H), 6.98 (t, *J* = 7.6 Hz, 1H), 6.30 (d, *J* = 15.9 Hz, 1H), 4.23 – 4.18 (m, 2H), 3.93 (t, *J* = 8.1 Hz, 2H), 3.08 (t, *J* = 8.0 Hz, 2H), 2.99 (s, 6H), 1.30 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 166.7, 161.1, 144.3, 141.0, 133.7, 125.5, 125.1, 123.0, 123.0, 116.7, 59.8, 52.2, 37.1, 29.4, 29.2, 13.8.

4.2 Substrates Scope of N-heteroarenes

A mixture of amide (0.2 mmol, 1 equiv), olefin (2 equiv), $[RuCl_2(p-cymene)]_2$ (6.4 mg, 0.05 equiv), AgSbF₆ (13.7 mg, 0.2 equiv), Cu(OAc)₂·H₂O (40 mg, 1 equiv) and THF (1mL) in a 15 mL glass tube (sealed with PTFE cap) was heated at 100 °C for 20 hours. The reaction mixture was cooled to room tempreture, and concentrated in vacuo. The resulting residue was purified by column chromatography on silica gel to give the alkenylated product.



¹H NMR (400 MHz, CDCl₃) δ 7.42 (t, *J* = 12.4 Hz, 2H), 7.23 – 7.21 (m, 1H), 7.08 (t, *J* = 7.6 Hz, 1H), 6.29 (d, *J* = 15.9 Hz, 1H), 4.24 – 4.19 (m, 2H), 4.16 (t, *J* = 7.5 Hz, 2H), 3.05 (t, *J* = 7.5 Hz, 2H), 1.41 (s, 9H), 1.30 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 179.3, 166.9, 143.5, 141.9, 134.5, 125.4, 125.3, 125.0, 124.8, 116.7, 60.0, 50.8, 40.0, 30.6, 28.3, 14.1; HRMS Calcd for C₁₈H₂₄NO₃ [M+H⁺]:302.1756; Found: 302.1762.



¹H NMR (400 MHz, CDCl₃) δ 7.45 (t, *J* = 11.0 Hz, 2H), 7.24 (d, *J* = 7.3 Hz, 1H), 7.11 (t, *J* = 7.6 Hz, 1H), 6.33 (d, *J* = 16.0 Hz, 1H), 4.84 – 4.78 (m, 1H), 4.28 – 4.16 (m, 2H), 3.32 – 3.26 (m, 1H), 2.53 (d, *J* = 15.0 Hz, 1H), 1.41 (s, 9H), 1.31 (t, *J* = 7.1 Hz, 3H), 1.25 (d, *J* = 6.4 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 179.5, 167.7, 143.1, 142.5, 134.8, 127.7, 126.7, 125.8, 125.3, 117.3, 60.7, 57.9, 41.0, 38.4, 29.3, 21.1, 14.7; HRMS Calcd for C₁₉H₂₆NO₃ [M+H⁺]:316.1913; Found: 316.1926.



¹H NMR (400 MHz, CDCl₃) δ 7.36 (d, *J* = 3.3 Hz, 1H), 7.33 (d, *J* = 11.0 Hz, 1H), 7.08 (d, *J* = 8.5 Hz, 1H), 6.27 (d, *J* = 16.0 Hz, 1H), 4.24 – 4.17(m, 4H), 3.10 (t, *J* = 7.6 Hz, 2H), 1.41 (s, 9H), 1.30 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 179.3, 166.5, 144.3, 140.6, 132.7, 131.0, 126.2, 124.6, 123.7, 116.8, 59.89, 50.2, 39.9, 29.8, 28.0, 13.8; HRMS Calcd for C₁₈H₂₃ClNO₃ [M+H⁺]:336.1366 Found: 336.1372.



¹H NMR (400 MHz, CDCl₃) δ 7.32 (d, *J* = 16.0 Hz, 1H), 7.28 (d, *J* = 8.6 Hz, 1H), 7.22 (d, *J* = 8.4 Hz, 1H), 6.28 (d, *J* = 16.0 Hz, 1H), 4.24 - 4.16 (m, 4H), 3.07 (t, *J* = 7.6 Hz, 2H), 1.40 (s, 9H), 1.30 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 179.3, 166.5, 143.9, 140.7, 134.9, 127.5, 126.3, 124.2, 119.8, 116.9, 59.9, 49.8, 39.9, 31.8, 28.0, 13.8; HRMS Calcd for C₁₈H₂₃ClNO₃ [M+H⁺]: 380.0861 Found: 380.0881.



¹H NMR (400 MHz, CDCl₃) δ 7.54 (d, *J* = 15.7 Hz, 1H), 7.48 – 7.46 (m, 1H), 7.17 (t, *J* = 6.3 Hz, 2H), 6.38 (d, *J* = 16.0 Hz, 1H), 4.41 (s, 1H), 4.25 – 4.12 (m, 2H), 3.31 (s, 1H), 2.76 (s, 2H), 2.04 (s, 2H), 1.36 (d, *J* = 23.5 Hz, 9H), 1.30 (d, *J* = 7.1 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 179.6, 167.5, 141.1, 130.9, 130.5, 126.3, 124.5, 118.8, 60.8, 45.8, 29.1, 26.1, 24.9, 14.8; HRMS Calcd for C₁₉H₂₆NO₃ [M+H⁺]:316.1913; Found: 316.1916.



¹H NMR (400 MHz, CDCl₃) δ 7.51 (d, *J* = 14.8 Hz, 1H), 7.28 (s, 1H), 6.99 (s, 1H), 6.37 (d, *J* = 16.0 Hz, 1H), 4.43 (d, *J* = 2.4 Hz, 1H), 4.22 (q, *J* = 7.1 Hz, 2H), 3.28 (s, 1H), 2.72 (s, 2H), 2.32 (s, 3H), 2.00 (s, 2H), 1.39 – 1.29 (m, 12H); ¹³C NMR (101 MHz, CDCl₃) δ 179.1, 167.1, 140.7, 138.8, 135.4, 133.2, 130.8, 130.1, 124.6, 118.0, 60.3, 45.4, 28.7, 21.0, 14.4; HRMS Calcd for C₂₀H₂₈NO₃ [M+H⁺]:330.2069; Found: 330.2071.



¹H NMR (400 MHz, CDCl₃) δ 7.39 (d, *J* = 16.0 Hz, 1H), 7.19 – 7.16 (m, 1H), 7.09 (t, *J* = 7.9 Hz, 1H), 6.91 – 6.89 (m, 1H), 6.36 (d, *J* = 16.0 Hz, 1H), 4.45 (s, 2H), 4.24 – 4.19 (m, 3H), 3.66 (s, 1H), 1.40 (s, 9H), 1.31 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 179.0, 167.1, 148.6, 140.6, 131.1, 126.8, 126.5, 118.7, 118.6, 118.4, 66.9, 60.5, 44.6, 40.3, 28.6, 14.5; HRMS Calcd for C₁₈H₂₄NO₄ [M+H⁺]:318.1705; Found: 318.1720.



¹H NMR (400 MHz, CDCl₃) δ 7.74 -7.68 (m, 2H), 7.43 – 7.34 (m, 2H), 7.23 – 7.17 (m, 1H), 6.46 (d, *J* = 16.1 Hz, 1H), 4.26 (q, *J* = 7.1 Hz, 2H), 3.16 (s, 3H), 1.33 (t, *J* = 7.1 Hz, 3H), 1.04 (s, 9H).; ¹³C NMR (101 MHz, CDCl₃) δ 178.6, 166.6, 144.7, 139.3, 132.6, 130.9,129.9, 128.7, 127.6, 121.1, 60.9, 40.9, 29.3, 14.4; HRMS Calcd for C₁₇H₂₄NO₃ [M+H⁺]: 290.1756; Found: 290.1760.



¹H NMR (400 MHz, CDCl₃) δ 7.75 (d, J = 16.1 Hz, 1H), 7.72 – 7.68 (m, 1H), 7.39 (p, J = 7.7 Hz, 2H), 7.19 – 7.13 (m, 1H), 6.44 (d, J = 16.1 Hz, 1H), 4.26 (m, 2H), 4.20 – 4.12 (m, 1H), 3.01 (d, J = 5.4 Hz,

1H), 1.32 (t, J = 7.1 Hz, 3H), 1.10 (t, J = 7.0 Hz, 3H), 1.01 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 176.8, 165.6, 141.8, 138.6, 132.0, 130.3, 129.4, 127.7, 126.67, 119.9, 59.9, 46.7, 40.2, 28.4, 13.5, 11.4; HRMS Calcd for C₁₈H₂₆NO₃ [M+H⁺]:304.1913; Found: 304.1925.



¹H NMR (400 MHz, CDCl₃) δ 7.86 (d, J = 16.2 Hz, 1H), 7.76 (dd, J = 6.2, 3.2 Hz, 1H), 7.47 – 7.38 (m, 2H), 7.17 (dd, J = 6.2, 3.0 Hz, 1H), 6.46 (d, J = 16.1 Hz, 1H), 4.66 (dt, J = 13.4, 6.7 Hz, 1H), 4.28 (q, J = 7.1 Hz, 2H), 1.35 (t, J = 7.1 Hz, 3H), 1.22 (d, J = 6.6 Hz, 3H), 1.02 (s, 9H), 0.96 (d, J = 6.8 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 177.8, 166.7, 140.7, 134.9, 132.1, 130.2, 128.7, 126.9, 120.0, 60.8, 51.4, 29.92, 29.9, 29.4, 27.4, 22.8, 19.7, 14.4; HRMS Calcd for C₁₉H₂₈NO₃ [M+H⁺]:318.2069; Found: 318.2059.



¹H NMR (400 MHz, CDCl₃) δ 7.78 – 7.67 (m, 2H), 7.42 – 7.35 (m, 2H), 7.19 – 7.14 (m, 1H), 6.44 (d, *J* = 16.1 Hz, 1H), 4.29-4.23 (m, 2H), 4.15 – 4.07 (m, 1H), 2.86 (s, 1H), 1.65-1.57 (m, 1H), 1.47 – 1.39 (m, 1H), 1.32 (t, *J* = 7.1 Hz, 3H), 1.28-1.22 (m, 2H), 1.00 (s, 9H), 0.87 (t, *J* = 7.3 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 177.2, 166.0, 142.4, 138.9, 132.3, 130.6, 129.8, 128.0, 127.0, 120.3, 60.2, 52.2, 40.5, 28.7, 28.6, 19.7, 13.8, 13.4; HRMS Calcd for C₂₀H₂₉NO₃ [M+H⁺]:332.2226; Found: 332.2233.



¹H NMR (400 MHz, CDCl₃) δ 8.00 (d, *J* = 16.0 Hz, 1H), 7.67 – 7.64 (m, 1H), 7.39 – 7.31 (m, 3H), 7.30 (d, *J* = 7.5 Hz, 1H), 7.25 – 7,22 (m, 3H), 7.21 – 7.19 (m, 1H), 6.41 (d, *J* = 16.0 Hz, 1H), 4.29 – 4.23 (m, 2H), 1.33 (t, *J* = 7.1 Hz, 3H), 1.18 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 180.0, 166.7, 144.2, 143.9, 140.1, 132.2, 131.1, 129.64, 129.3, 128.5, 127.8, 127.3, 127.2, 120.4, 60.7, 42.0, 29.7, 14.5; HRMS Calcd for C₂₂H₂₆NO₃ [M+H⁺]:352.1913; Found: 352.1926.



¹H NMR (400 MHz, CDCl₃) δ 7.67 (s, 1H), 7.64 (d, *J* = 7.4 Hz, 1H), 7.33 (t, *J* = 7.3 Hz, 1H), 7.24 – 7.22 (m, 1H), 7.21 – 7.20 (m, 3H), 7.14 – 7.10 (m, 2H), 6.79 – 6.76 (m, 1H), 6.39 (d, *J* = 16.1 Hz, 1H), 5.45 (d, *J* = 13.9 Hz, 1H), 4.29 – 4.23 (m, 2H), 4.06 (d, *J* = 13.9 Hz, 1H), 1.34 (t, *J* = 7.1 Hz, 3H), 1.01 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 177.7, 166.1, 141.9, 139.0, 136.7, 132.9, 131.5, 129.9, 129.4,

128.5, 128.0, 127.2, 127.1, 120.6, 60.5, 55.7, 41.0, 29.1, 14.1; HRMS Calcd for $C_{23}H_{28}NO_3$ [M+H⁺]:366.2069; Found: 366.2074.



¹H NMR (400 MHz, CDCl₃) δ 7.75 (d, *J* = 16.0 Hz, 1H), 7.49 (d, *J* = 6.9 Hz, 1H), 7.22 (t, *J* = 7.6 Hz, 1H), 7.02 (s, 1H), 6.36 (d, *J* = 16.0 Hz, 1H), 4.25 – 4.20 (m, 2H), 2.23 (s, 3H), 1.40 (s, 9H), 1.31 (t, *J* =

7.1 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 176.6, 166.3, 139.9, 135.9, 134.0, 131.8, 127.0, 124.0, 119.4, 60.0, 27.3, 17.7, 13.8; HRMS Calcd for C₁₈H₂₆NO₃ [M+H⁺]:304.1913; Found: 304.1921.



¹H NMR (400 MHz, CDCl₃) δ 7.69 (d, J = 16.1 Hz, 1H), 7.48 (d, J = 7.9 Hz, 1H), 7.38 – 7.32 (m, 1H), 7.16 (t, J = 8.5 Hz, 1H), 6.48 (d, J = 16.1 Hz, 1H), 4.28 – 4.23 (m, 2H), 3.40 (s, 1H), 3.12 (s, 3H), 1.31 (t, J = 7.1 Hz, 3H), 0.99 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 179.0, 166.5, 158.9(J_{C-F} = 247), 138.4(J_{C-F} = 4), 135.5, 132.9(J_{C-F} = 14), 129.9 (J_{C-F} = 8), 123.07 (J_{C-F} = 3), 122.7, 118.1 (J_{C-F} = 21), 61.3, 41.4, 40.1, 29.1, 14.7. HRMS Calcd for C₁₇H₂₃NO₃F [M+H⁺]:308.1662; Found: 308.1673.



¹H NMR (400 MHz, CDCl₃) δ 7.68 (d, *J* = 16.1 Hz, 1H), 7.48 (s, 1H), 7.20 (d, *J* = 7.9 Hz, 1H), 7.07 (d, *J* = 8.0 Hz, 1H), 6.44 (d, *J* = 16.1 Hz, 1H), 4.28 – 4.23 (m, 2H), 3.13 (s, 3H), 2.39 (s, 3H), 1.32 (t, *J* = 7.1 Hz, 3H), 1.02 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 179.0, 166.9, 142.5, 139.7, 138.9, 132.5, 132.0, 130.0, 128.4, 121.1, 61.1, 41.3, 29.6, 21.6, 14.8; HRMS Calcd for C₁₈H₂₆NO₃ [M+H⁺]:304.1913; Found: 304.1924.



¹H NMR (400 MHz, CDCl₃) δ 7.71 – 7.67 (m, 1H), 7.43 – 7.40 (m, 1H), 7.25 – 7.21 (m, 1H), 7.20 – 7.08 (m, 1H), 6.49 (d, *J* = 16.1 Hz, 1H), 4.35 – 4.29 (m, 2H), 3.21 (s, 3H), 1.38 (t, *J* = 7.1 Hz, 3H), 1.11 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 178.0 165.5, 161.4(*J*_{C-F} = 247), 140.2, 137.5 (*J*_{C-F} = 2), 134.1(*J*_{C-F} = 9), 130.9(*J*_{C-F} = 8)), 121.69, 117.2(*J*_{C-F} = 23), 113.28(*J*_{C-F} = 23), 60.4, 40.4, 28.6, 13.8 HRMS Calcd for C₁₇H₂₃NO₃F [M+H⁺]:308.1662; Found: 308.1669.



¹H NMR (400 MHz, CDCl₃) δ 7.68 (d, J = 16.1 Hz, 1H), 7.58 (d, J = 8.0 Hz, 1H), 7.18 (d, J = 8.0 Hz, 1H), 7.00 (s, 1H), 6.41 (d, J = 16.1 Hz, 1H), 4.28 – 4.23 (m, 2H), 3.15 (s, 3H), 2.38 (s, 3H), 1.32 (t, J = 7.1 Hz, 3H), 1.04 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 178.0, 166.2, 144.0, 141.1, 138.7, 129.8, 128.9, 126.9, 119.4, 60.1, 40.3, 28.7, 20.7, 13.8; HRMS Calcd for C₁₈H₂₆NO₃ [M+H⁺]:304.1913; Found: 304.1927.

4.2 Substrates Scope of Olefins



A mixture of amide (0.2 mmol, 1 equiv), olefin (2 equiv), $[RuCl_2(p-cymene)]_2$ (6.4 mg, 0.05 equiv), AgSbF₆ (13.7 mg, 0.2 equiv), Cu(OAc)₂·H₂O (40 mg, 1 equiv) and THF (1mL) in a 15 mL glass tube (sealed with PTFE cap) was heated at 100 °C for 20 hours. The reaction mixture was cooled to room tempreture, and concentrated in vacuo. The resulting residue was purified by column chromatography on silica gel to give the alkenylated product



¹H NMR (400 MHz, CDCl₃) δ 7.50 (d, *J* = 16.0 Hz, 1H), 7.43 – 7.30 (m, 6H), 7.23 (d, *J* = 7.3 Hz, 1H), 7.09 (t, *J* = 7.6 Hz, 1H), 6.36 (d, *J* = 16.0 Hz, 1H), 5.21 (s, 2H), 4.16 (t, *J* = 7.5 Hz, 2H), 3.05 (t, *J* = 7.5 Hz, 2H), 1.38 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 179.8, 167.3, 144.1, 143.2, 136.6, 135.1, 128.9, 128.7, 128.5, 126.1, 126.0, 125.6, 125.5, 116.9, 66.6, 51.5, 40.6, 31.2, 28.9; HRMS Calcd for C₂₃H₂₆NO₃ [M+H⁺]:364.1913; Found: 364.1909.



¹H NMR (400 MHz, CDCl₃) δ 7.39 (d, *J* = 7.8 Hz, 1H), 7.33 (d, *J* = 16.0 Hz, 1H), 7.21 (d, *J* = 6.8 Hz, 1H), 7.07 (t, *J* = 7.6 Hz, 1H), 6.22 (d, *J* = 16.0 Hz, 1H), 4.15 (t, *J* = 7.5 Hz, 2H), 3.04 (t, *J* = 7.5 Hz, 2H), 1.50 (s, 9H), 1.41 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 179.7, 166.8, 144.0, 141.3, 135.1, 126.5, 125.75, 125.7, 125.4, 119.6, 80.5, 51.5, 40.6, 31.3, 29.0, 28.7; HRMS Calcd for C₂₀H₂₈NO₃ [M+H⁺]:330.2069; Found: 330.2069.



¹H NMR (400 MHz, CDCl₃) δ 7.47 (d, *J* = 16.0 Hz, 1H), 7.41 (d, *J* = 7.8 Hz, 1H), 7.23 (d, *J* = 0.9 Hz, 1H), 7.09 (t, *J* = 7.6 Hz, 1H), 6.33 (d, *J* = 16.0 Hz, 1H), 6.02 – 5.92 (m, 1H), 5.39 – 5.33 (m, 1H), 5.26 – 5.22 (m, 1H), 4.68 – 4.66 (m, 2H), 4.17 (t, *J* = 7.5 Hz, 2H), 3.06 (t, *J* = 7.5 Hz, 2H), 1.41 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 179.8, 166.9, 143.9, 142.8, 134.9, 132.6, 125.8, 125.8, 125.4, 125.2, 118.1, 116.6, 65.1, 51.2, 40.4, 30.9, 28.7; HRMS Calcd for C₁₉H₂₃NO₃ [M+H⁺]:314.1756; Found: 314.1787.



¹H NMR (400 MHz, CDCl₃) δ 7.33 – 7.26 (m, 2H), 7.21 (d, *J* = 16.6 Hz, 1H), 7.11 (t, *J* = 7.6 Hz, 1H), 5.71 (d, *J* = 16.6 Hz, 1H), 4.19 (t, *J* = 7.5 Hz, 2H), 3.05 (t, *J* = 7.5 Hz, 2H), 1.40 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 179.4, 149.2, 143.5, 135.5, 126.9, 125.7, 125.5, 124.9, 119.1, 95.3, 5.5, 40.5, 31.0, 28.8; HRMS Calcd for C₁₆H₁₉NO₂ [M+H⁺]:255.1497; Found: 255.1521.



¹H NMR (400 MHz, CDCl₃) δ 7.33 – 7.27 (m, 3H), 7.10 (t, *J* = 7.6 Hz, 1H), 6.71 (d, *J* = 15.5 Hz, 1H), 4.18 (t, *J* = 7.6 Hz, 2H), 3.07 (t, *J* = 7.7 Hz, 2H), 3.05 (s, 3H), 1.38 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 179.5, 143.8, 143.4, 134.7, 126.5, 125.6, 125.3, 124.7, 123.4, 51.1, 43.9, 40.2, 30.7, 28.4; HRMS Calcd for C₁₆H₂₂NO₃S [M+H⁺]:308.1320; Found: 308.1388.



¹H NMR (400 MHz, CDCl₃) δ 7.33 (d, *J* = 7.8 Hz, 1H), 7.18 (d, *J* = 7.4 Hz, 1H), 7.17 – 7.08 (m, 1H), 7.05 (t, *J* = 7.5 Hz, 1H), 6.03 (t, *J* = 18.1 Hz, 1H), 4.17 – 4.07 (m, 6H), 3.00 (t, *J* = 7.4 Hz, 2H), 1.35 (s, 9H), 1.31 (t, *J* = 7.0 Hz, 6H); ¹³C NMR (101 MHz, CDCl₃) δ 179.0, 146.9, 146.8, 143.1, 134.5, 126.3, 126.1, 125.4, 125.0, 112.8, 110.9, 61.8, 61.8, 51.0, 40.0, 30.7, 28.3, 16.4, 16.3; HRMS Calcd for C₁₉H₂₉NO₄P [M+H⁺]:366.1834; Found: 366.1836.



¹H NMR (400 MHz, CDCl₃) δ 8.02 – 7.96 (m, 2H), 7.56 – 7.53 (m, 2H), 7.50 (d, *J* = 11.0 Hz, 1H), 7.47 (t, *J* = 4.9 Hz, 2H), 7.29 (d, *J* = 15.8 Hz, 2H), 7.13 (t, *J* = 7.6 Hz, 1H), 4.18 (t, *J* = 7.5 Hz, 2H), 3.07 (t, *J* = 7.5 Hz, 2H), 1.38 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 192.1, 179.4, 144.3, 143.2, 138.6, 134.9, 132.4, 129.1, 128.8, 128.6, 128.3, 126.4, 126.0, 125.4, 125.2, 121.7, 51.2, 40.3, 30.9, 28.6, 28.4; HRMS Calcd for C₂₂H₂₃NO₂ [M+H⁺]:334.1807; Found: 334.03.



¹H NMR (400 MHz, CDCl₃) δ 7.41 (d, J = 7.8 Hz, 1H), 7.33 (d, J = 16.3 Hz, 1H), 7.24 – 7.19 (m, 1H), 7.08 (t, J = 7.6 Hz, 1H), 6.54 (d, J = 16.3 Hz, 1H), 4.16 (t, J = 7.5 Hz, 2H), 3.04 (t, J = 7.5 Hz, 2H), 2.69 – 2.64 (m, 2H), 1.40 (s, 9H), 1.13 (t, J = 7.3 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 201.8, 179.5,

144.1, 140.8, 135.1, 126.2, 126.1, 125.5, 125.5, 51.5, 40.6, 33.2, 31.1, 28.9, 8.7; HRMS Calcd for C₁₈H₂₄NO₃ [M+H⁺]:286.1807; Found: 286.1816.



¹H NMR (400 MHz, CDCl₃) δ 7.30 (d, *J* = 7.5 Hz, 1H), 7.11 – 6.98 (m, 2H), 6.25 – 6.00 (m, 2H), 4.11 (t, *J* = 7.4 Hz, 2H), 3.00 (t, *J* = 7.4 Hz, 2H), 1.86 (d, *J* = 5.1 Hz, 3H), 1.40 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 178.8, 141.6, 134.5, 129.4, 128.8, 125.4, 125.1, 124.5, 122.6, 51.2, 40.2, 31.2, 28.7, 19.0; HRMS Calcd for C₁₆H₂₂NO [M+H⁺]:244.1701; Found: 244.1700.

5. Alkynylation of pivaloyl amide Protected N-heteroarenes



A mixture of amide (0.2 mmol, 1 equiv), alkyne; (2 equiv), $[RuCl_2(p-cymene)]_2$ (3.2 mg, 0.05 equiv), KPF₆ (7.2 mg, 0.2 equiv), Cu(OAc)₂·H₂O (12.6 mg, 1 equiv) and H₂O (0.5mL) in a 15 mL glass tube (sealed with PTFE cap) was heated at 120 °C for 24 hours. The reaction mixture was cooled to room tempreture, and concentrated in vacuo. The resulting residue was purified by column chromatography on silica gel to give the alkenylated product.



¹H NMR (400 MHz, CDCl₃) δ 7.34 (d, *J* = 7.6 Hz, 1H), 7.16 – 7.14 (m, 1H), 6.97 (t, *J* = 7.6 Hz, 1H), 4.14 (t, *J* = 7.6 Hz, 2H), 3.02 (t, *J* = 7.6 Hz, 2H), 1.37 (s, 9H), 1.13 (d, *J* = 2.3 Hz, 21H); ¹³C NMR (101 MHz, CDCl₃) δ 176.6, 145.3, 134.2, 132.8, 124.2, 123.8, 115.1, 104.5, 96.7, 50.8, 39.7, 30.7, 28.3, 18.8, 11.3; HRMS Calcd for C₂₄H₃₈NOSi [M+H⁺]:384.2723; Found: 384.2732.



¹H NMR (400 MHz, CDCl₃) δ 7.34 (d, J = 7.8 Hz, 1H), 7.18 (d, J = 7.4 Hz, 1H), 7.00 (t, J = 7.6 Hz, 1H), 4.78 – 4.72 (m, 1H), 3.31 – 3.25 (m, 1H), 2.48 (d, J = 14.9 Hz, 1H), 1.37 (s, 9H), 1.24 (d, J = 6.4 Hz, 3H), 1.18 – 1.05 (m, 21H); ¹³C NMR (101 MHz, CDCl₃) δ 176.8, 144.8, 134.0, 132.2, 125.0, 124.5, 117.4, 104.8, 95.6, 57.3, 40.2, 38.1, 28.9, 20.9, 18.9, 18.9, 11.5; HRMS Calcd for C₂₅H₄₀NOSi [M+H⁺]:398.2879; Found: 398.2885.



¹H NMR (400 MHz, CDCl₃) δ 7.11 – 7.09 (m, 1H), 6.99 (t, *J* = 7.9 Hz, 1H), 6.85 – 6.83 (m, 1H), 4.38 (s, 1H), 4.22 (s, 2H), 3.76 (s, 1H), 1.36 (s, 9H), 1.12 (s, 21H); ¹³C NMR (101 MHz, CDCl₃) δ 176.9, 147.8, 128.2, 126.3, 125.4, 121.1, 116.9, 104.2, 95.2, 66.5, 43.9, 39.4, 28.0, 18.3, 10.9; HRMS Calcd for C₂₄H₃₈NO₂Si [M+H⁺]:400.2672; Found: 400.2667.

6. Gram scale reaction and Indole derivatives

6.1 Gram scale reaction



A mixture of amide (5 mmol, 1.02g), olefin (2 equiv), $[RuCl_2(p-cymene)]_2$ (91.5 mg, 0.03 equiv), AgSbF₆ (342.5 mg, 0.2 equiv), Cu(OAc)₂ .H₂O (1g, 1 equiv) and THF (10mL) in a 50 mL glass tube (sealed with PTFE cap) was heated at 100 °C for 24 hours. The reaction mixture was cooled to room tempreture, and concentrated in vacuo. The resulting residue was purified by column chromatography on silica gel to give the alkenylated product.

6.2 Indole derivatives



A mixture of **3a** (0.2 mmol, 1 equiv), DDQ (3 equiv) DCE (1mL) in a 15 mL glass tube (sealed with PTFE cap) was heated at 100 °C for 36 hours. The reaction mixture was cooled to room tempreture, and concentrated in vacuo. The resulting residue was purified by column chromatography on silica gel to give the alkenylated product3y.



¹H NMR (400 MHz, CDCl₃) δ 7.81 (d, *J* = 15.8 Hz, 1H), 7.61 – 7.58 (m, 2H), 7.47 (d, *J* = 7.5 Hz, 1H), 7.28 (d, *J* = 7.6 Hz, 1H), 6.63 (d, *J* = 3.7 Hz, 1H), 6.29 (d, *J* = 15.7 Hz, 1H), 4.2 9 – 4.24 (m, 2H), 1.56 (s, 9H), 1.34 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 179.0, 166.5, 142.5, 134.4, 130.8, 126.0, 123.5, 123.1, 122.8, 121.9, 117.2, 107.0, 59.9, 41.6, 28.5, 13.8; HRMS Calcd for C₁₈H₂₂NO₃ [M+H⁺]:300.1600; Found: 300.1610.

6.3 Removal of Directing Group



A mixture of 3aa (48.6 mg, 0.2 mmol, 100 mol %) in EtOH (2 mL) was added aqueous s-KOH

solution (1 mL) at room temperature. The reaction mixture was stirred for 20 h at 100 °C and cooled to 0 °C. The reaction mixture was neutralized with aqueous $s-NH_4Cl$ solution, and partitioned between DCM and H_2O . The organic layer was dried over MgSO₄ and concentrated in vacuo. The resulting residue was purified by column chromatography on silica gel to give the alkenylated product **3ac.**



¹H NMR (400 MHz, DMSO) δ 7.00 (d, J = 7.7 Hz, 1H), 6.88 (d, J = 7.0 Hz, 1H), 6.49 (t, J = 7.4 Hz, 1H), 6.40 – 6.36 (m, 1H), 6.10 – 6.01 (m, 1H), 5.50 (s, 1H), 3.43 – 3.36 (m, 2H), 2.89 (t, J = 8.6 Hz, 2H), 1.84 –1.82 (m, 3H); ¹³C NMR (101 MHz, DMSO) δ 149.7, 129.8, 127.8, 124.0, 123.9, 123.0, 118.8, 117.7, 46.6, 29.5, 19.0; HRMS Calcd for C₁₁H₁₄N [M+H⁺]:160.1126; Found: 160.1138.

7. A plausible mechanism to explain 3aa



A proposed reaction mechanism of 3aa(3x was coreected to 3aa) was shown as above. Firstly, the $[\operatorname{RuCl}_2(\operatorname{p-cymene})]_2$ catalyst is activated by Ag⁺ in the reaction. The ruthenium species coordinated with the directing group of pivaloyl, followed by ortho metalation, affording a six-membered metallacycle intermediate I, and then Regioselectiv coordinative insertion of olefin into the Ru-C bond provides an eight-membered ruthenium intermediate II. Futher, β -oxygen elimination affords the products $3aa.^1$

[1] J. Park, N. K. Mishra, Sharma, S. Han, Y. Shin, T. Jeong, J. S. Oh, J. H. Kwak, Y. H. Jung and I. S. Kim, J. Org. Chem. 2015, 80, 1818;

8. NMR Spectra























S28































































































S73



S74