Supplementary Information

Discrimination of Enantiomers of Dipeptide Derivatives with Two Chiral Centers by Tetraaza Macrocyclic Chiral Solvating Agents Using ¹H NMR Spectroscopy

^a College of Chemistry, Beijing Normal University, Beijing 100875, P. R. China.

Email: linai@bnu.edu.cn. jxzhang@bnu.edu.cn

^b Department of Chemistry, Capital Normal University, Beijing 10048, P. R. China.

^c Department of Chemistry, Missouri University of Science and Technology, Rolla, Missouri 65409, USA

Contents

Crystallographic data for TAMCSAs 1a·2CH ₃ COCH ₃ , 1b and 1c·CDCl ₃	S2
Theoretically proposed model of G1-1 and G1-2 with TAMCSA 1c	S3
Synthesis of TAMCSAs 1a-1c and chiral diimines 3a-3c	S4
Synthesis of dipeptide derivatives with two chiral centers G1-1-G8-2	S6
Determination of enantiomeric excesses of G1 in the presence of TAMCSA 1c	S13
Determination of the stoichiometry of (±)-G1 with TAMCSA 1c (Job plots)	S13
Determination of the association constants of G5-1 and G5-2 with TAMCSA 1a	S13
Partial ¹ H NMR spectra of discriminations of enantiomers of (±)-G1-G8	S14
¹ H NMR, ¹³ C NMR, ¹ H- ¹ H COSY, ¹ H- ¹³ C HSQC and HRMS Spectra	S21
¹ H NMR spectra of discrimination of enantiomers of (\pm) -G1-G8	S55
¹ H NMR spectra of determination of enantiomeric excesses of G1	S67
¹ H NMR spectra of (±)-G1 with TAMCSA 1c (Job plots)	S72
ESI mass spectrum of complex of (±)-G1 with TAMCSA 1c	S76

Lixia Fang,^a Caixia Lv,^a Guo Wang,^b Lei Feng,^a Pericles Stavropoulos,^c Guangpeng Gao,^a Lin Ai^{*,a} and Jiaxin Zhang^{*,a}

References

Crystallographic data for TAMCSAs 1a·2CH₃COCH₃, 1b and 1c·CDCl₃

<i>, , , , , , , , , , , , , , , , , , , </i>		5 57 .	
	1a·2CH ₃ COCH ₃	1b	1c·CDCl ₃
Formula	$C_{42}H_{50}N_4O_6$	$C_{36}H_{36}Cl_{2}N_{4}O_{4}\\$	$C_{37}H_{36}DBr_2Cl_3N_4O_4$
M	706.86	659.59	868.88
Temperature (K)	110(2)	110(2)	100(2)
Crystal system	Monoclinic	Monoclinic	Tetragonal
Space group	<i>P</i> 2(1)	<i>C</i> 2	P43
a/Å	9.0188(8)	37.839(4)	13.811(3)
b/Å	21.1444(19)	25.190(2)	13.811(3)
$c/\text{\AA}$	10.7833(9)	13.6300(14)	20.076(4)
a/deg	90	90	90
β /deg	111.097(2)	101.307(2)	90
γ/deg	90	90	90
$V/\text{\AA}^3$	1918.5(3)	12739(2)	3829.5(16)
Ζ	2	12	4
$D_C/Mg m^{-3}$	1.224	1.032	1.507
F_{000}	756	4152	1760
μ/mm^{-1}	0.082	0.188	2.370
θ (range)/deg	2.02 to 25.25	1.90 to 25.25	1.790 to 27.514
Reflections collected/unique	9662/3569 [$R_{int} = 0.0362$]	17687/17687 [$\mathbf{R}_{int} = 0.0000$]	25246/7907 [$R_{int} = 0.0384$]
Parameters	480	1280	465
$R_1 \left[I > 2\sigma(I) \right]$	0.0357	0.0668	0.0317
$wR_2 [I > 2\sigma(I)]$	0.0756	0.1739	0.0679
Goodness of fit on F ⁻²	1.025	1.011	1.027
CCDC deposition numbers	1007044	1007045	1007047

Table S1. Crystallographic data for TAMCSAs 1a · 2CH₃COCH₃, 1b and 1c · CDCl₃.





Figure S1. Theoretically proposed model of G1-1 and G1-2 with TAMCSA 1c.



Synthesis of TAMCSAs 1a-1c and chiral diimines 3a-3c.¹

Procedure of synthesis of chiral diimines 3a-3c. Salicylaldehyde or its derivatives (2 mmol) was added to a solution of diamine (0.76 g, 2 mmol) in dried MeOH (10 mL) and the mixture was refluxed under nitrogen atmosphere. After yellow precipitate was formed, the reaction mixture continued to be stirred for two hours. The chiral diimines **3a-3c** was obtained as yellow solid and used in the next step without further purification.

Procedure of synthesis of TAMCSAs 1a-1c. To a solution of chiral diimines **3** (1.2 mmol) in dried THF (60 mL) was added zinc powder (0.78 g, 12 mmol) and MsOH (1.15g, 12 mmol) in dried THF (20 mL) under nitrogen atmosphere at -18°C. The mixture was stirred for 24h. The reaction mixture was basified to pH = 9-10 with saturated NaHCO₃ solution. The precipitate formed was filtered off and washed with CHCl₃. The organic layer was separated from filtrate. The water layer was extracted with CHCl₃ (15 mL × 3). The combined organic layer was dried over anhydrous Na₂SO₄. The solvent was removed under reduce pressure and the residue was

purified by column chromatography on silica gel to afford TAMCSAs **1a-1c**. Meanwhile, the chiral compounds **2a-2c** were also obtained as known chiral compounds.²

TAMCSA **1a**: $R_f = 0.4$ (petroleum ether / ethyl acetate = 2/1); 31% yield; mp. 191-193 °C; [α]_D²⁰-9.3 (*c* 0.03, THF); ¹H NMR (400 MHz, CDCl₃) δ : 1.11-1.18 (m, 2H), 1.25-1.31 (m, 2H), 1.62-1.64 (m, 2H), 1.76 (d, *J* = 11.8 Hz, 2H), 4.05 (br, 2H), 4.42 (s, 2H), 4.71 (s, 2H), 6.13 (d, *J* = 7.6 Hz, 2H), 6.50-6.54 (m, 2H), 6.87(d, *J* = 7.2 Hz, 2H), 6.94 (d, *J* = 8.0 Hz, 2H), 7.03-7.07 (m, 2H), 7.27-7.33 (m, 6H), 7.53-7.54 (m, 4H), 10.14 (br, 2H); ¹³C NMR (100 MHz, CDCl₃) δ : 24.5, 31.7, 53.2, 62.1, 69.4, 117.2, 119.2, 125.9, 127.4, 127.5, 128.3, 128.4, 128.6, 138.2, 155.0, 173.2; IR (KBr): 3300, 2932, 1663, 1527, 1493, 1454, 753, 698 cm⁻¹; ESI-HRMS: calcd for C₃₆H₃₉N₄O₄ 591.2971, found 591.2970 ([M+H]⁺).

TAMCSA **1b**: $R_f = 0.3$ (petroleum ether / ethyl acetate = 2/1); 29% yield; mp. 188-190 °C; $[\alpha]_D^{20}$ -27.4 (*c* 0.03, THF); ¹H NMR (400 MHz, CDCl₃) δ : 1.08-1.16 (m, 2H), 1.21-1.29 (m, 2H), 1.62 (d, *J* = 7.6 Hz, 2H), 1.73 (d, *J* = 12.8 Hz, 2H), 4.04-4.13 (m, 2H), 4.49 (s, 2H), 4.66(s, 2H), 6.06 (d, *J* = 1.7 Hz, 2H), 6.81 (d, *J* = 8.7 Hz, 2H), 6.96-7.02 (m, 4H), 7.33-7.37 (m, 6H), 7.54-7.56 (m, 4H), 10.09 (br, 2H); ¹³C NMR (100 MHz, DMSO-d₆) δ : 24.5, 31.7, 52.7, 62.1, 66.0, 116.5, 121.6, 126.5, 127.3, 127.4, 128.1, 129.0, 139.2, 142.3, 155.0, 172.2; IR (KBr): 3314, 2931, 1662, 1547, 1486, 700 cm⁻¹; ESI-HRMS: calcd for C₃₆H₃₇Cl₂N₄O₄ 659.2192, found 659.2198 ([M+H]⁺).

TAMCSA **1c**: $R_f = 0.3$ (petroleum ether / ethyl acetate = 2/1); 33% yield; mp. 198-200 °C; $[\alpha]_D^{20}$ -34.4 (*c* 0.03, THF); ¹H NMR (400 MHz, CDCl₃) δ : 1.09-1.14 (m, 2H), 1.24-1.27 (m, 2H), 1.63 (d, *J* = 8.2 Hz, 2H), 1.75 (d, *J* = 13.3 Hz, 2H), 1.86 (br, 2H), 4.04 (br, 2H), 4.45 (s, 2H), 4.64(s, 2H), 6.17 (s, 2H), 6.79-6.81 (m, 4H), 7.17 (dd, *J* = 8.6Hz, *J* = 2.2Hz, 2H), 7.34-7.35 (m, 6H), 7.52-7.54 (m, 4H), 10.01 (br, 2H); ¹³CNMR (100 MHz, DMSO-d₆) δ : 25.0, 32.2, 53.2, 62.5,

S5

66.5, 109.8, 117.5, 127.0, 127.8, 128.6, 130.8, 132.4, 139.7, 156.0, 156.1, 172.6; IR (KBr): 3387, 2933, 1655, 1528, 1483, 698 cm⁻¹; ESI-HRMS: calcd for $C_{36}H_{37}Br_2N_4O_4$ 747.1182, found 747.1184 ([M+H]⁺).

Synthesis of dipeptide derivatives with two chiral centers G1-1-G8-2.³



For example, synthetic procedure of G1-1: *D*-PheOCH₂Ph (0.26 g, 1 mmol) was added to a solution of *N*-Ts-*D*-phenylglycine (0.30 g, 1 mmol) in dry ethyl acetate (8 mL) at room temperature. A solution of DCC (0.25 g, 1.2 mmol) in dry ethyl acetate (5 mL) was added dropwise into the above solution while stirring under nitrogen atmosphere at ice-bath. And then the reaction mixture was allowed to room temperature and stirred for over night. The precipitate formed was filtered off and filtrate was concentrated under reduce pressure. The crude product was purified by column chromatography on silica gel (petroleum ether / ethyl acetate = 3/1) to afford dipeptide derivative with two chiral centers **G1-1**.

G1-1. 85% yield; mp. 158-160 °C; $R_f = 0.3$ (petroleum ether / ethyl acetate = 3/1); $[\alpha]_D^{20}$ -49.1 (*c* 0.02, THF). ¹H NMR (400 MHz, CDCl₃) δ : 2.36 (s, 3H), 2.96 (dd, J = 13.9 Hz, J = 5.8 Hz, 1H), 3.03 (dd, J = 13.8 Hz, J = 5.8 Hz, 1H), 4.71 (d, J = 5.0 Hz, 1H), 4.70-4.75 (m, 1H), 5.01 (d, J = 12.0 Hz, 1H), 5.08 (d, J = 12.1 Hz, 1H), 5.83 (d, J = 4.8 Hz, 1H), 6.11 (d, J = 7.4 Hz, 1H), 6.83 (d, J = 7.0 Hz, 2H), 7.02 (d, J = 7.3 Hz, 1H), 7.15 7.34 (m, 10H), 7.35 (d, J = 2.9 Hz, 1H), 7.59 (d, J = 8.1 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ : 21.5, 37.5, 53.6, 60.5, 67.4, 127.2, 127.3, 127.5, 128.6, 128.7, 129.0, 129.2, 129.5, 134.8, 135.2, 135.9, 136.6, 143.5, 168.5, 170.3; IR

(KBr): 3273, 1739, 1658, 1548, 1344, 1164, 1084, 707 cm⁻¹; MALDI-HRMS: calcd for $C_{31}H_{30}N_2NaO_5S$ 565.1773, found 565.1768 ([M+Na]⁺).

G1-2. 80% yield; mp. 155-157 °C; $R_f = 0.3$ (petroleum ether / ethyl acetate = 3/1); $[\alpha]_D^{20} + 50.0$ (*c* 0.02, THF). ¹H NMR (400 MHz, CDCl₃) & 2.36 (s, 3H), 2.98 (dd, *J* = 13.9 Hz, *J* = 5.8 Hz, 1H), 3.04 (d, *J* = 13.9 Hz, *J* = 5.8 Hz, 1H), 4.71 (d, *J* = 4.8 Hz, 1H), 4.71-4.75 (m, 1H), 5.01 (d, *J* = 12.1 Hz, 1H), 5.08 (d, *J* = 12.1 Hz, 1H), 5.84 (d, *J* = 4.9 Hz, 1H), 6.12 (d, *J* = 7.5 Hz, 1H), 6.83 (d, *J* = 6.8 Hz, 2H), 7.02 (d, *J* = 7.0 Hz, 2H), 7.15-7.23 (m, 10H), 7.34-7.35 (m, 3H), 9.59 (d, *J* = 8.28 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃) & 21.5, 37.5, 53.6, 60.5, 67.4, 127.2, 127.3, 127.5, 128.5, 128.6, 128.7, 129.0, 129.2, 129.5, 134.8, 135.2, 136.0, 136.7, 143.5, 168.5, 170.4; IR (KBr): 3269, 2361, 1734, 1655, 1495, 1150, 696 cm⁻¹; ESI-HRMS: calcd for C₃₁H₃₀N₂NaO₅S 565.1768, found 565.1738 ([M+Na]⁺).

G2-1. 52% yield; mp. 160-162 °C; $R_f = 0.3$ (petroleum ether / ethyl acetate = 3/1); $[\alpha]_D^{20}$ -39.0 (*c* 0.02, THF). ¹H NMR (400 MHz, CDCl₃) δ : 2.36 (s, 3H), 2.89 (dd, *J* = 13.8 Hz, *J* = 5.4 Hz, 1H), 2.97 (dd, *J* = 13.8 Hz, *J* = 5.3 Hz, 1H), 4.73 (d, *J* = 4.8 Hz, 1H), 4.79-4.84 (m, 1H), 5.08 (d, *J* = 12.0 Hz, 1H), 5.14 (d, *J* = 12.0 Hz, 1H), 5.87 (d, *J* = 4.8 Hz, 1H), 6.09 (d, *J* = 8.2 Hz, 1H), 6.54 (d, *J* = 7.1 Hz, 2H), 6.98-7.02 (m, 2H), 7.06-7.16 (m, 5H), 7.21-7.30 (m, 5H), 7.37-7.38 (m, 3H), 7.56 (d, *J* = 8.3 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ : 21.5, 37.5, 53.2, 60.5, 67.4, 127.0, 127.2, 127.6, 128.5, 128.6, 128.7, 129.0, 129.2, 129.5, 134.8, 134.9, 136.4, 136.7, 143.4, 168.3, 170.6; IR (KBr): 3258, 1734, 1658, 1345, 1161, 1085, 685 cm⁻¹; ESI-HRMS: calcd for C₃₁H₃₀N₂NaO₅S 565.1768, found 565.1734 ([M+Na]⁺).

G2-2. 70% yield; mp. 159-161 °C; $R_f = 0.3$ (petroleum ether / ethyl acetate = 3/1); $[\alpha]_D^{20}$ +40.0 (*c* 0.02, THF). ¹H NMR (400 MHz, CDCl₃) δ : 2.36 (s, 3H), 2.90 (dd, *J* = 13.9 Hz, *J* = 5.4 Hz, 1H), 2.97 (dd, *J* = 13.9 Hz, *J* = 5.3 Hz, 1H), 4.73 (d, *J* = 4.9 Hz, 1H), 4.79-4.84 (m, 1H), 5.08 (d,

S7

J = 12.0 Hz, 1H), 5.14 (d, J = 12.0 Hz, 1H), 5.87 (d, J = 4.8 Hz, 1H), 6.09 (d, J = 8.1 Hz, 1H), 6.54 (d, J = 7.0 Hz, 1H), 6.98-7.02 (m, 2H), 7.06-7.16 (m, 5H), 7.22-7.30 (m, 5H), 7.37-7.38 (m, 3H), 7.56 (d, J = 8.3 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃) δ : 21.5, 37.5, 53.1, 60.5, 67.5, 127.0, 127.2, 127.6, 128.5, 128.7, 129.0, 129.1, 129.5, 134.7, 134.8, 136.4, 136.6, 143.4, 168.2, 170.1; IR (KBr): 3269, 1745, 1648, 1344, 1150, 696 cm⁻¹; ESI-HRMS: calcd for C₃₁H₃₀N₂NaO₅S 565.1768, found 565.1743 ([M+Na]⁺).

G3-1. 50% yield; mp. 156-158 °C; $R_f = 0.3$ (petroleum ether / ethyl acetate = 3/1); $[\alpha]_D^{20}$ -48.0 (*c* 0.02, THF). ¹H NMR (400 MHz, CDCl₃) &: 2.40 (s, 3H), 2.79 (dd, *J* = 13.9 Hz, *J* = 6.5 Hz, 1H), 3.07 (dd, *J* = 13.9 Hz, *J* = 6.0 Hz, 1H), 3.95-4.00 (m, 1H), 4.91 (d, *J* = 7.5 Hz, 1H), 5.10 (d, *J* = 12.4 Hz, 1H), 5.16 (d, *J* = 12.4 Hz, 1H), 5.44 (d, *J* = 6.8 Hz, 1H), 6.86-6.88 (m, 2H), 7.10-7.13 (m, 2H), 7.16-7.21 (m, 8H), 7.28-7.30 (m, 5H), 7.58 (d, *J* = 8.3 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃) &: 21.6, 38.3, 56.8, 57.4, 67.4, 127.1, 127.2, 127.9, 128.3, 128.5, 128.8, 128.9, 129.3, 129.8, 135.0, 135.1, 135.9, 136.0, 143.8, 169.6, 169.8; IR (KBr): 3247, 1734, 1648, 1161, 1096, 696 cm⁻¹; ESI-HRMS: calcd for C₃₁H₃₀N₂NaO₅S 565.1768, found 565.1746 ([M+Na]⁺).

G3-2. 62% yield; mp. 158-160 °C; $R_f = 0.3$ (petroleum ether / ethyl acetate = 3/1); $[\alpha]_D^{20}$ +46.0 (*c* 0.02, THF). ¹H NMR (400 MHz, CDCl₃) & 2.40 (s, 3H), 2.79 (dd, *J* = 14.0 Hz, *J* = 6.5 Hz, 1H), 3.07 (dd, *J* = 14.0 Hz, *J* = 6.0 Hz, 1H), 3.95-4.00 (m, 1H), 4.92 (d, *J* = 7.5 Hz, 1H), 5.10 (d, *J* = 12.4 Hz, 1H), 5.16 (d, *J* = 12.4 Hz, 1H), 5.44 (d, *J* = 6.8 Hz, 1H), 6.86-6.88 (m, 2H), 7.10-7.13 (m, 2H), 7.16-7.21 (m, 8H), 7.28-7.32 (m, 5H), 7.58 (d, *J* = 8.3 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃) & 21.6, 38.4, 56.8, 57.5, 67.4, 127.1, 127.2, 127.3, 127.9, 128.3, 128.5, 128.8, 128.9, 129.3, 129.8, 135.0, 135.1, 135.9, 136.1, 143.7, 169.6, 169.8; IR (KBr): 3247, 1745, 1658, 1150, 1096, 696 cm⁻¹; ESI-HRMS: calcd for C₃₁H₃₀N₂NaO₅S 565.1768, found 565.1736 ([M+Na]⁺).

G4-1. 49% yield; mp. 148-150 °C; $R_f = 0.3$ (petroleum ether / ethyl acetate = 3/1); $[\alpha]_D^{20}$ -7.5 (*c* 0.02, THF). ¹H NMR (400 MHz, CDCl₃) δ : 2.38 (s, 3H), 2.92 (dd, *J* = 13.0 Hz, *J* = 5.6 Hz, 1H), 2.97 (dd, *J* = 13.0 Hz, *J* = 5.9 Hz, 1H), 3.89-3.94 (m, 1H), 4.93 (d, *J* = 7.0 Hz, 1H), 5.11 (d, *J* = 12.4 Hz, 1H), 5.17 (d, *J* = 12.3 Hz, 1H), 5.40 (d, *J* = 7.0 Hz, 1H), 6.88 (d, *J* = 7.0 Hz, 2H), 7.10-7.20 (m, 10H), 7.28-7.31 (m, 5H), 7.52 (d, *J* = 8.3 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ : 21.5, 38.5, 56.7, 57.7, 67.4, 127.1, 127.2, 127.9, 128.4, 128.5, 128.8, 128.9, 129.2, 129.7, 135.0, 135.6, 135.8, 143.7, 169.5, 170.0; IR (KBr): 3291, 1734, 1648, 1507, 1161, 1452, 1085, 739 cm⁻¹; ESI-HRMS: calcd for C₃₁H₃₀N₂NaO₅S 565.1768, found 565.1744 ([M+Na]⁺).

G4-2. 50% yield; mp. 149-151 °C; $R_f = 0.3$ (petroleum ether / ethyl acetate = 3/1); $[\alpha]_D^{20}$ +7.0 (*c* 0.02, THF). ¹H NMR (400 MHz, CDCl₃) δ : 2.38 (s, 3H), 2.92 (dd, *J* = 14.0 Hz, *J* = 6.6 Hz, 1H), 2.97 (dd, *J* = 13.9 Hz, *J* = 6.8 Hz, 1H), 3.89-3.94 (m, 1H), 4.93 (d, *J* = 7.0 Hz, 1H), 5.11 (d, *J* = 12.4 Hz, 1H), 5.17 (d, *J* = 12.4 Hz, 1H), 5.40 (d, *J* = 7.0 Hz, 1H), 6.87- 6.89 (m, 2H), 7.10-7.20 (m, 10H), 7.28-7.31 (m, 5H), 7.52 (d, *J* = 8.3 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃) δ : 21.5, 38.5, 56.7, 57.7, 67.4, 127.1, 127.2, 127.9, 128.4, 128.5, 128.9, 129.2, 129.7, 135.0, 135.6, 135.8, 143.7, 169.5, 170.0; IR (KBr): 3291, 1745, 1637, 1507, 1161, 1085, 696 cm⁻¹; ESI-HRMS calcd for C₃₁H₃₀N₂NaO₅S 565.1768, found 565.1744 ([M+Na]⁺).

G5-1: 62% yield; mp. 109-110 °C; $R_f = 0.3$ (petroleum ether / ethyl acetate = 5/2); $[\alpha]_D^{20} = -2.2$ (*c* 0.04, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ : 1.22 (d, *J* = 7.1 Hz, 3H), 2.39 (s, 3H), 3.00 (d, *J* = 5.8 Hz, 2H), 3.71-3.78 (m, 1H), 4.75 (dt, *J* = 7.6 Hz, *J* = 5.8 Hz, 1H), 5.09 (d, *J* = 12.1 Hz, 1H), 5.10 (d, *J* = 5.2 Hz, 1H), 5.16 (d, *J* = 12.1 Hz, 1H), 6.38 (d, *J* = 7.6 Hz, 1H), 6.87-6.89 (m, 2H), 7.19-7.22 (m, 3H), 7.27-7.29 (m, 4H), 7.35-7.37 (m, 3H), 7.73 (d, *J* = 8.2 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃) δ : 19.5, 21.5, 37.7, 52.3, 53.4, 67.4, 127.1, 127.3, 128.5, 128.6,

129.3, 129.8, 134.9, 135.4, 136.7, 143.8, 170.7, 170.8; IR (KBr): 3285, 2917, 1747, 1669, 1156, 1089, 699 cm⁻¹; TOF-HRMS: calcd for $C_{26}H_{29}N_2O_5S$ 481.1791, found 481.1794 ([M+H]⁺).

G5-2: 72% yield; mp. 110-111 °C; $R_f = 0.3$ (petroleum ether / ethyl acetate = 5/2); $[\alpha]_D^{20} = +2.2$ (*c* 0.04, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ : 1.22 (d, *J* = 7.1 Hz, 3H), 2.40 (s, 3H), 3.01 (d, *J* = 5.8 Hz, 2H), 3.74 (dq, *J* = 7.2 Hz, *J* = 7.1 Hz, 1H), 4.75 (dt, *J* = 7.8 Hz, *J* = 5.8 Hz, 1H), 5.07 (d, *J* = 5.9 Hz, 1H), 5.09 (d, *J* = 11.9 Hz, 1H), 5.16 (d, *J* = 12.1 Hz, 1H), 6.36 (d, *J* = 7.6 Hz, 1H), 6.87-6.89 (m, 2H), 7.16-7.22 (m, 3H), 7.28-7.30 (m, 4H), 7.36-7.38 (m, 3H), 7.73 (d, *J* = 8.2 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃) δ : 19.6, 21.5, 37.7, 52.2, 53.4, 67.4, 127.1, 127.3, 128.5, 128.6, 129.3, 129.8, 134.9, 145.4, 136.7, 143.8, 170.7, 170.8; IR (KBr): 3289, 2976, 1746, 1641, 1155, 1097, 695 cm⁻¹; TOF-HRMS: calcd for C₂₆H₂₉N₂O₅S 481.1791, found 481.1794 ([M+H]⁺).

G6-1: 72% yield; mp. 173-174 °C; $R_f = 0.3$ (petroleum ether / ethyl acetate = 7/2); $[\alpha]_D^{20} = -37.5$ (*c* 0.02, THF); ¹H NMR (400 MHz, CDCl₃) δ : 0.75 (d, *J* = 6.8 Hz, 3H), 0.84 (d, *J* = 6.8 Hz, 3H), 1.92-2.00 (m, 1H), 2.34 (s, 3H), 2.80 (dd, *J* = 13.8 Hz, *J* = 6.1 Hz, 1H), 2.95 (dd, *J* = 13.8 Hz, *J* = 5.3 Hz, 1H), 3.48 (dd, *J* = 8.4 Hz, *J* = 4.8 Hz, 1H), 4.71-4.75 (m, 1H), 5.05 (d, *J* = 12.1 Hz, 1H), 5.12 (d, *J* = 12.1 Hz, 1H), 5.27 (d, *J* = 8.4 Hz, 1H), 6.05 (d, *J* = 7.7 Hz, 1H), 6.81 (d, *J* = 7.0 Hz, 2H), 7.17-7.22 (m, 7H), 7.27-7.36 (m, 3H), 7.73 (d, *J* = 8.1 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃) δ : 17.1, 19.0, 21.5, 31.7, 37.9, 53.2, 61.7, 67.4, 127.2, 127.4, 128.5, 128.6, 129.2, 134.8, 135.3, 136.8, 143.7, 169.8, 170.7; IR (KBr): 3260, 2916, 2362, 1735, 1653, 1179, 706 cm⁻¹; TOF-HRMS: calcd for C₂₈H₃₃N₂O₅S 509.2104, found 509.2104 ([M+H]⁺).

G6-2: 80% yield; mp. 168-169 °C; $R_f = 0.3$ (petroleum ether / ethyl acetate = 7/2); $[\alpha]_D^{20} = +37.1$ (*c* 0.02, THF); ¹H NMR (400 MHz, CDCl₃) δ : 0.75 (d, *J* = 6.8 Hz, 3H), 0.84 (d, *J* = 6.8

Hz, 3H), 1.92-2.00 (m, 1H), 2.34 (s, 3H), 2.80 (dd, J = 13.8 Hz, J = 6.1 Hz, 1H), 2.95 (dd, J = 13.9 Hz, J = 5.3 Hz, 1H), 3.48 (dd, J = 8.4 Hz, J = 4.8 Hz, 1H), 4.71-4.75 (m, 1H), 5.06 (d, J = 12.0 Hz, 1H), 5.12 (d, J = 12.0 Hz, 1H), 5.26 (d, J = 8.4 Hz, 1H), 6.05 (d, J = 7.8 Hz, 1H), 6.80-6.82 (m, 2H), 7.17-7.25 (m, 7H), 7.28-7.36 (m, 3H), 7.73 (d, J = 8.2 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃) δ : 17.1, 19.0, 21.5, 31.7, 37.8, 53.2, 61.7, 67.4, 127.2, 127.4, 128.5, 128.6, 129.2, 129.6, 134.8, 135.3, 136.8, 143.7, 169.8, 170.7; IR (KBr): 3272, 2953, 2362, 1746, 1641, 1191, 683 cm⁻¹; TOF-HRMS: calcd for C₂₈H₃₃N₂O₅S 509.2104, found 509.2104 ([M+H]⁺).

G7-1: 37% yield; mp. 109-110 °C; $R_f = 0.3$ (petroleum ether / ethyl acetate = 1/3); $[\alpha]_D^{20} = -24.4$ (*c* 0.02, THF); ¹H NMR (400 MHz, CDCl₃) δ : 1.82-1.88 (m, 2H), 2.18-2.24 (m, 2H), 2.35 (s, 3H), 2.91 (dd, *J* = 14.0 Hz, *J* = 7.1 Hz, 1H), 2.97 (dd, *J* = 13.9 Hz, *J* = 6.0 Hz, 1H), 3.74-3.79 (m, 1H), 4.64-4.69 (m, 1H), 5.06 (d, *J* = 12.1 Hz, 1H), 5.13 (d, *J* = 12.1 Hz, 1H), 5.47 (s, 1H), 5.60 (s, 1H), 6.45 (d, *J* = 6.6 Hz, 1H), 6.96-6.98 (m, 2H), 7.20-7.22 (m, 4H), 7.25-7.27 (m, 3H), 7.33-7.35 (m, 3H), 7.69 (d, *J* = 8.2 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃) δ : 21.5, 28.7, 31.0, 37.7, 53.7, 55.9, 67.2, 127.0, 127.3, 128.4, 128.5, 128.6, 129.2, 129.7, 135.0, 135.9, 136.6, 143.6, 170.6, 171.1, 175.6; IR (KBr): 3341, 2356, 1724, 1662, 1152, 799 666 cm⁻¹; TOF-HRMS: calcd for C₂₈H₃₂N₃O₆S 538.2006, found 538.2005 ([M+H]⁺).

G7-2: 41% yield; mp. 176-177 °C; $R_f = 0.3$ (petroleum ether / ethyl acetate = 1/3); $[\alpha]_D^{20} = +24.1$ (*c* 0.02, THF); ¹H NMR (400 MHz, CDCl₃) δ : 1.84-1.88 (m, 2H), 2.18-2.26 (m, 2H), 2.35 (s, 3H), 2.91 (dd, J = 13.9 Hz, J = 7.1 Hz, 1H), 2.97 (dd, J = 13.9 Hz, J = 5.9 Hz, 1H), 3.74-3.79 (m, 1H), 4.64-4.69 (m, 1H), 5.06 (d, J = 12.2 Hz, 1H), 5.13 (d, J = 12.2 Hz, 1H), 5.47 (s, 1H), 5.60 (s, 1H), 6.46 (d, J = 7.2 Hz, 1H), 6.96-6.98 (m, 2H), 7.20-7.22 (m, 4H), 7.25-7.27 (m, 3H), 7.33-7.35 (m, 3H), 7.69 (d, J = 8.2 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃) δ : 21.5, 28.7, 31.0, 37.7, 53.7, 55.9, 67.2, 127.0, 127.3, 128.4, 128.5, 128.6, 129.2, 129.7, 135.0, 135.9, 136.6, 143.6,

170.6, 171.1, 175.6; IR (KBr): 3329, 2344, 1736, 1662, 1152, 812, 702 cm⁻¹; TOF-HRMS: calcd for $C_{28}H_{32}N_3O_6S$ 538.2006, found 538.2009 ([M+H]⁺).

G8-1: 33% yield; mp. 152-153 °C; $R_f = 0.3$ (petroleum ether / ethyl acetate = 5/2); $[\alpha]_D^{20} = -37.0$ (*c* 0.02, THF); ¹H NMR (400 MHz, CDCl₃) δ : 1.29 (d, *J* = 7.2 Hz, 3H), 2.37 (s, 3H), 4.42-4.49 (m, 1H), 4.80 (d, *J* = 5.4 Hz, 1H), 5.05 (d, *J* = 12.2 Hz, 1H), 5.12 (d, *J* = 12.2 Hz, 1H), 5.89 (d, *J* = 5.4 Hz, 1H), 6.20 (d, *J* = 7.0 Hz, 1H), 7.15-7.18 (m, 4H), 7.20-7.24 (m, 5H), 7.33-7.35 (m, 3H), 7.59 (d, *J* = 8.3 Hz, 2H) ; IR (KBr): 3260, 2349, 1723, 1641, 1344, 1155, 683 cm⁻¹; TOF-HRMS: calcd for C₂₅H₂₇N₂O₅S 467.1635, found 467.1632 ([M+H]⁺).

G8-2 : 36% yield; mp. 153-154 °C; $R_f = 0.3$ (petroleum ether / ethyl acetate = 5/2); $[\alpha]_D^{20} = +37.6$ (*c* 0.02, THF); ¹H NMR (400 MHz, CDCl₃) δ : 1.29 (d, *J* = 7.1 Hz, 3H), 2.37 (s, 3H), 4.42-4.49 (m, 1H), 4.80 (d, *J* = 5.4 Hz, 1H), 5.05 (d, *J* = 12.2 Hz, 1H), 5.12 (d, *J* = 12.2 Hz, 1H), 5.88 (d, *J* = 5.4 Hz, 1H), 6.19 (d, *J* = 7.0 Hz, 1H), 7.15-7.18 (m, 4H), 7.20-7.25 (m, 5H), 7.32-7.35 (m, 3H), 7.59 (d, *J* = 8.2 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃) δ : 18.0, 21.4, 48.6, 60.3, 67.3, 127.3, 127.5, 128.1, 128.5, 128.6, 128.9, 129.4, 135.0, 136.1, 136.8, 143.4, 168.5, 171.8; IR (KBr): 3244, 1736, 1650, 1346, 1164, 678 cm⁻¹; TOF-HRMS: calcd for C₂₅H₂₇N₂O₅S 467.1635, found 467.1633 ([M+H]⁺).

Determination of enantiomeric excesses of G1 in the presence of TAMCSA 1c.

To evaluate the accuracy of determination of enantiomeric excess (ee) by ¹H NMR spectroscopy, the following nine samples were prepared containing **G1-1** with 85, 65, 45, 25, 0, -25, -45, -65 and -85% *ee*, respectively. And then all the samples were obtained by adding 1 equiv of TAMCSA **1c** to the above solutions with a concentration of 5 mM in CDCl₃, respectively. Their ¹H NMR spectra were recorded on a 400 MHz spectrometer and their enantiomeric purities were calculated based on the integration of ¹H NMR signals of *NH* protons of Ts*NH* group of **G1-1** and **G1-2**.

Determination of the stoichiometry of (±)-G1 with TAMCSA 1c (Job plots).³

The samples of dipeptide derivative (±)-G1 with TAMCSA 1c were dissolved in CDCl₃ (0.5 mL) with a concentration of 10 mM, respectively. The solutions were distributed among the nine NMR tubes with the molar fractions X of (±)-G1 from 0.1 to 0.9. The ¹H NMR spectra of all the samples were recorded on a 400 MHz spectrometer. The Job plots of G1-1 and G1-2 with TAMCSA 1c exhibited a maximum value ($X^*\Delta\delta = 0.042$ ppm) at molar fraction of X = 0.5. The results indicate that TAMCSA 1c can form a 1:1 complex with (±)-G1.

Determination of the association constants of G5-1 and G5-2 with TAMCSA 1a.

To determine the association constants, the samples of **G5-1** and **G5-2** with TAMCSA **1a** were prepared with a constant concentration (2 mM) of TAMCSA **1a** and varying concentrations of **G5-1** or **G5-2** with 0.0, 1.0, 2.0, 3.0, 4.0, 5.0, 6.0, 7.0, 8.0, 9.0 and 10.0 mM in CDCl₃, respectively and their ¹H NMR spectra were measured. Based on the nonlinear curve-fitting method, the association constants were obtained for their complexes.

Partial ¹H NMR spectra of discriminations of enantiomers of (±)-G1-G8.

TAMCSA /Guest	Proton	Spectra ^{b,d}	Spectra ^{c,d}	TAMCSA /Guest	Proton	Spectra ^{b,d}	Spectra ^{c,d}
1a/(±)-G1	PhCH ₂		M	,	ArH	7.75 ppm	7.75 ppm
		0 3.05 3.00 2.95		1b/(±)-G5	CH ₃	1.24 ppm	1.25 ppm
	PhCH ₂ O	5.10 5.00	5.1 5.0		PhCH ₃	ppm	2.40 ppm
	CONH	6.30 ppm			PhCH ₂	3.00 ppm	
	ArH	5 7.60 ppm			CH ₃ CH	3.75 ppm	 3.75 ppm
1b/(±)-G1	PhCH ₂	3.05 3.00 2.95	 3.10 3.05300 2.95		PhCH ₂ O	5.13 5.04	5.10 ppm
		3.05 3.00 2.95				5.13 5.04	£111 £111
	PhCH ₂ O	5.10 5.00 ppm	<u></u> 5.1 ppm		CONH	6.56 ppm	6.48 ppm
		5.10 5.00 ppm	برالی_ برالی_ 5.1 ppm		ArH	7.75 ppm	7.75 ppm

Table S2. Partial ¹H NMR spectra of (±)-**G1-G8** (10×10^{-3} M) in the presence of TAMCSAs **1a-1c** (10×10^{-3} M) by ¹H NMR spectroscopy in CDCl₃ at room temperature, respectively.

	CONH	6.30 ppm		1c/(±)-G5	CH ₃	1.25 ppm
	ArH	7.64 ppm	 		PhCH ₃	2.40 ppm
1c/(±)-G1	PhCH ₂	3.02 ppm 2.96			PhCH ₂	3.00 ppm
	PhCH ₂ O	2.96 ppm 2.96			CH₃CH	5 3.75 ppm
		5.10 ppm 5.00	15 5.05 ppm		PhCH ₂ O	5.10 ppm
		•• .10				5.10 ppm
	CONH	6.30 ppm	6.25 ppm		CONH	65 6.55 ppm
	Ar <i>H</i>	7,60 ppm	7.65 ppm		ArH	7.75 ppm
1a/(±)-G2	PhCH ₂	0 2.94 ppm 2.88	 	1a/(±)-G6	(<i>CH</i> ₃) ₂ CH	0.76 ppm
		0 2.94 ppm 2.88	###### ##### 00 2.95 2.90 2.1			0.84 ppm

Â 1.24 ppm ▲ Ą 2.40 ppm Â 3.00 ppm ▲ Â. 3.75 ppm 14 A 5.15 ppm £Ĵ 5.15 ppm A 6.50 ppm Δ_{i} 7.75 ppm ★ Ŵ 0.76 ppm Â 0.84 ppm



						*	.×€ L
	CONH	ppm	6.12 ppm		PhCH ₂ O	5.10 ppm	5.10 ppm
	Ar <i>H</i>	7.58 ppm	 7.60 ppm		CONH	6.20 ppm	
		M	<u> </u>				
1a/(±)-G3	PhCH ₃	2.40 ppm	2.40 ppm		ArH	7.75 ppm	7.75 ppm
						XW.	
	PhCH ₂	2.84 ppm	2.80 ppm	1c/(±)-G6	(<i>CH</i> ₃) ₂ CH	0.76 ppm	0.76 ppm
		3.08 ppm	3.10 ppm			0.84 ppm	0.84 ppm
		The second secon	xM. xM.				_ ^.^.
	PhCH ₂ O	.20 5.10 ppm	5.15 ppm		PhCH ₂	2.80 ppm	2.80 ppm ★
	PhCH	5.44 ppm	5.45 5.4 ppm		(CH ₃) ₂ CH <i>CH</i>	3.50 ppm	3.50 ppm
		7.58				*	×ÛL ≏ŨL
	Ar <i>H</i>	-	ppm		PhCH ₂ O	5.10 ppm	5.10 ppm
		240 236) ki
$1b/(\pm)$ -G3	PhCH ₃	2.10 2.50 ppm	2.40 ppm		CONH	6.2 6. ppm	6.16 ppm
		285 - 280	 XX				
	PhCH ₂	ppm	2.85 ppm		ArH	ppm	7.75 ppm

		3.10 3.05 3.00	3.10 3.05 ppm	1a/(±)-G7	Ph <i>CH</i> .	2.35	2.35
	DFCH O	5.15 5.10		IW(2) (37	1 110/1/3	2 90	
	PnC <i>H</i> ₂ O	5.44 	20 5.15 ppm 3.10		PhCH ₂		2.95 ppm • •
	Ph <i>CH</i>	7.60 7.56				00 2.95 ppm	2.95 ppm
	Ar <i>H</i>	ppm	7.60 ppm		PhCH ₂ O	5.10 ppm	5.10 ppm
1c/(±)-G3	PhCH ₃	2.40 ppm	2.40 ppm			5.10 ppm	5.10 ppm
	PhCH ₂	2.84 ppm	2.85 2.80 ppm		Ar <i>H</i>	7.70 ppm	7.70 ppm
			3.10 3.05 ppm	1b/(±)-G7	PhCH ₃	2.35 ppm	2.36 ppm
	PhCH ₂ O	5.15 5.10 ppm) 5.15 5.10		PhCH ₂ O	5.10 ppm	5.10 ppm
	Ph <i>CH</i>	5.45 ppm	5.46 ppm	1c/(±)-G7	PhCH ₃	2.35 2.30 ppm	2.36 ppm
	ArH	7.58 ppm			PhCH ₂ O	5.10 ppm	5.10 ppm

. .





^{*a*} $\Delta\Delta\delta = \Delta\delta_1 - \Delta\delta_2$; $\Delta\delta_1 = \delta_1 - \delta_{free}$; $\Delta\delta_2 = \delta_2 - \delta_{free}$, the numbers "1 and 2" present a pair of enantiomers of dipeptide derivatives, respectively.

^b Partial ¹H NMR spectra of the corresponding protons of (±)-**G1-G8** (10×10^{-3} M), H:G = 1:1. ^c Overlaid ¹H NMR spectra of the corresponding protons of one of the two enantiomers of (±)-**G1-G8** (10×10^{-3} M).

^{*d*} The following different signs stand for the corresponding stereoisomers: G1-1 (\bigcirc), G1-2 (\bullet); G2-1 (\square), G2-2 (\blacksquare); G3-1 (\bigtriangledown), G3-2 (\blacktriangledown); G4-1 (\diamondsuit), G4-2 (\blacklozenge); G5-1 (\triangle), G5-2 (\blacktriangle); G6-1 (\diamondsuit), G6-2 (\bigstar); G7-1 (\diamondsuit), G7-2 (\bigstar); G8-1 (\heartsuit), G8-2 (\blacktriangledown).

¹H NMR, ¹³C NMR, ¹H-¹H COSY, ¹H-¹³C HSQC and HRMS Spectra



Figure S3. ¹³C NMR spectrum of TAMCSA 1a·2·CH₃COCH₃ in CDCl₃ (100 MHz).

Elemental Composition Report

Single Mass Analysis Tolerance = 3.0 PPM / DBE: min = -1.5, max = 50.0 Element prediction: Off Number of isotope peaks used for i-FIT = 3









Page 1



Figure S6. ¹³C NMR spectrum of TAMCSA 1b in DMSO-d₆ (100 MHz).

Elemental Composition Report

Single Mass Analysis Tolerance = 3.0 PPM / DBE: min = -1.5, max = 50.0 Element prediction: Off Number of isotope peaks used for i-FIT = 3





Page 1



Figure S9. 13 C NMR spectrum of TAMCSA 1c in DMSO-d₆ (100 MHz).

Single Mass Analysis

Tolerance = 2.0 PPM / DBE: min = -1.5, max = 50.0 Element prediction: Off Number of isotope peaks used for i-FIT = 3

Monoisotopic Mass, Even Electron Ions 676 formula(e) evaluated with 2 results within limits (up to 50 best isotopic matches for each mass) Н C: 0-40 H: 0-40 N: 0-10 O: 0-10 Br: 0-2 Phò 0 LCX-20130508-1 9 (0.154) AM (Top, 12, Ht, 5000.0, 0.00, 1.00); Cm (9) NH HÌN TOF MS ES+ Br 1.01e+003 'NH HN 749.1184 100-0 PhQ н 747.1184 751.1212 % 750.1261 748.1379 752.1302 750.9486 0 m/z 747.00 748.00 749.00 750.00 751.00 752.00









Figure S12. ¹³C NMR spectrum of dipeptide derivative G1-1 in CDCl₃ (100 MHz).



SmartFormula



Figure S13. HRMS spectrum of dipeptide derivative G1-1.



Figure S15. ¹H-¹³C HSQC of dipeptide derivative G1-1.



Figure S17. ¹³C NMR spectrum of dipeptide derivative G1-2 in CDCl₃ (100 MHz).



Figure S18. HRMS spectrum of dipeptide derivative G1-2.







Figure S20. ¹³C NMR spectrum of dipeptide derivative G2-1 in CDCl₃ (100 MHz).



Figure S21. HRMS spectrum of dipeptide derivative G2-1.







Figure S25. ¹³C NMR spectrum of dipeptide derivative G2-2 in CDCl₃ (100 MHz).



Figure S26. HRMS spectrum of dipeptide derivative G2-2.



Figure S27. ¹H NMR spectrum of dipeptide derivative G3-1 in CDCl₃ (400 MHz).



Figure S29. HRMS spectrum of dipeptide derivative G3-1.



Figure S31. ¹³C NMR spectrum of dipeptide derivative G3-2 in CDCl₃ (100 MHz).



Figure S32. HRMS spectrum of dipeptide derivative G3-2.






Figure S35. ¹H NMR spectrum of dipeptide derivative G4-1 in CDCl₃ (400 MHz).



Figure S36. ¹³C NMR spectrum of dipeptide derivative G4-1 in CDCl₃ (100 MHz).



Figure S37. HRMS spectrum of dipeptide derivative G4-1.



Figure S38. ¹H-¹H COSY of dipeptide derivative G4-1.



Figure S39. HSQC of dipeptide derivative G4-1.



Figure S41. ¹³C NMR spectrum of dipeptide derivative G4-2 in CDCl₃ (100 MHz).



Figure S43. ¹H NMR spectrum of dipeptide derivative G5-1 in CDCl₃ (400 MHz).



Figure S45. HRMS spectrum of dipeptide derivative G5-1.

7,77395 7,77895 7,77895 7,72009 7,72009 7,23091 6,68742 6,69746 6,69746 6,69746 6,69746 6,69746 6,69746 6,69766 6,69746 6,69766 6,69746 6,69746 6,69746 6,69746 6,69746 6,69746 6,69746 6,69746 6,697666 6,69766 6,697





Figure S48. HRMS spectrum of dipeptide derivative G5-2.





Figure S51. ¹H NMR spectrum of dipeptide derivative G6-1 in CDCl₃ (400 MHz).



Figure S53. HRMS spectrum of dipeptide derivative G6-1.

7,7,7,00 7,7,340 7,7,340 7,7,340 7,7,340 7,7,340 7,7,340 7,7,340 6,60055 6,50005 6,



Figure S55. ¹³C NMR spectrum of dipeptide derivative G6-2 in CDCl₃ (100 MHz).





Figure S59. HRMS spectrum of dipeptide derivative G7-1.

7,7010 7,73841 7,73841 7,73294 7,72194 7,72194 7,72194 6,448366,44836 6,448366,44836 6,448366,44836 6,448366,448366,





Figure S62. HRMS spectrum of dipeptide derivative G7-2.











Figure S67. HRMS spectrum of dipeptide derivative G8-1.

7,15998 7,75791 7,75791 7,75802 7,75802 7,72402 7,711681 7,711683 7,7116335 7,7116335 7,7116335 7,7116





Figure S70. HRMS spectrum of dipeptide derivative G8-2.





Figure S71. ¹H NMR spectrum of (\pm) -G1 with TAMCSA 1a (1:1) in CDCl₃ (400 MHz), [1a] = 10 mM.



Figure S72. ¹H NMR spectrum of (\pm) -G1 with TAMCSA 1b (1:1) in CDCl₃ (400 MHz), [1b] = 10 mM.



Figure S73. ¹H NMR spectrum of (\pm) -G1 with TAMCSA 1c (1:1) in CDCl₃ (400 MHz), [1c] = 10 mM.





Figure S75. ¹H NMR spectrum of (\pm)-G2 with TAMCSA 1b (1:1) in CDCl₃ (400 MHz), [1b] = 10 mM.







Figure S77. ¹H NMR spectrum of (\pm) -G3 with TAMCSA 1a (1:1) in CDCl₃ (400 MHz), [1a] = 10 mM.



Figure S79. ¹H NMR spectrum of (\pm) -G3 with TAMCSA 1c (1:1) in CDCl₃ (400 MHz), [1c] = 10 mM.



Figure S80.¹H NMR spectrum of (\pm) -G4 with TAMCSA 1a (1:1) in CDCl₃ (400 MHz), [1a] = 10 mM.





Figure S81.¹H NMR spectrum of (\pm)-G4 with TAMCSA 1b (1:1) in CDCl₃ (400 MHz), [1b] = 10 mM.





Figure S82. ¹H NMR spectrum of 1:1 mixture of 1c with (\pm)-G4 (1:1) in CDCl₃ (400 MHz), [1c] = 10 mM.

















Figure S89.¹H NMR spectrum of (\pm) -G7 with TAMCSA 1a (1:1) in CDCl₃ (400 MHz), [1a] = 10 mM.



Figure S90.¹H NMR spectrum of (\pm)-G7 with TAMCSA 1b (1:1) in CDCl₃ (400 MHz), [1b] = 10 mM.









Figure S94.¹H NMR spectrum of (\pm) -G8 with TAMCSA 1c (1:1) in CDCl₃ (400 MHz), [1c] = 10 mM.

¹H NMR spectra of determination of enantiomeric excesses of G1 in the presence of TAMCSA 1c.



Figure S95.¹H NMR spectrum of G1-1 and G1-2 (*ee* $_{G1-1}$ % = 85 %) with TAMCSA 1c in CDCl₃ (400 MHz), [1c] = 5 mM.



Figure S96. H NMR spectrum of GI-1 and GI-2 ($ee_{GI-1}\% = 65\%$) with TAMCSA Ic in CDCl₃ (400 MHz), [Ic] = 5 mM.



Figure S97.¹H NMR spectrum of **G1-1** and **G1-2** (*ee* $_{G1-1}$ % = 45 %) with TAMCSA 1c in CDCl₃ (400 MHz), [1c] = 5 mM.



1. 7. 5. 6. 11. 7. 5. 6. 11. 7. 5. 6. 11. 7. 5. 6. 11. 7. 5. 6. 11. 7. 5. 6. 11. 7. 5. 6. 11. 7. 5. 6. 11. 7. 5. 6. 11. 7. 5. 6. 11. 7. 5. 6. 11. 7. 5. 6. 11. 7. 5. 6. 11. 5. 11. 5. 6. 11. 5. 6. 11. 5. 6. 11. 5. 6. 11. 5. 6. 11. 5. 6. 11. 5. 6. 11. 5. 6. 11. 5. 6. 11. 5. 6. 11. 5. 6. 11. 5. 6. 11. 5. 6. 11. 5. 6. 11. 5. 6. 11. 5. 6. 11. 5.



Figure S99.¹H NMR spectrum of G1-1 and G1-2 (*ee* $_{G1-1}\% = 0\%$) with TAMCSA 1c in CDCl₃ (400 MHz), [1c] = 5 mM.





Figure S101.¹H NMR spectrum of G1-1 and G1-2 (*ee* $_{G1-1}$ % = -45 %) with TAMCSA 1c in CDCl₃ (400 MHz), [1c] = 5 mM.



Figure S102.¹H NMR spectrum of G1-1 and G1-2 (*ee* G1-1% = -65%) TAMCSA 1c with in CDCl₃ (400 MHz), [1c] = 5 mM. 7.6126 1.2586 1.2407 1.2033 1.1783

5 Ę. 3112



Figure S103.¹H NMR spectrum of G1-1 and G1-2 (*ee* G1-1% = -85%) TAMCSA 1c with in CDCl₃ (400 MHz), [1c] = 5 mM.





3.5

0.53

0.49

4.5

5.0

2.29

7.0

4.0 0.52 0.52 0.48

6.5

0.97

6.0

5.5

8 2 8 8 8.97

7.5

8.0

2.78-

2.0

2.5

6

3.0

0.77 0.68 0.82 0.58 0.58

1.5

1.0

0.5

0.0
25,080 27,280 27,280 27,280 27,280 27,280 27,280 27,280 27,280 27,280 27,280 27,280 27,280 27,280 27,280 26,887 26,897 26,997



Figure S106. ¹H NMR spectrum of (±)-G1 with TAMCSA 1c (0.7:0.3) in CDCl₃ (400 MHz), $[1c] + [(\pm)-G1] = 10$ mM.





Figure S107. ¹H NMR spectrum of (±)-G1 with TAMCSA 1c (0.6:0.4) in CDCl₃ (400 MHz), $[1c] + [(\pm)-G1] = 10$ mM.



Figure S108. ¹H NMR spectrum of (±)-G1 with TAMCSA 1c (0.5:0.5) in CDCl₃ (400 MHz), $[1c] + [(\pm)-G1] = 10$ mM.



Figure S109. ¹H NMR spectrum of (±)-G1 with TAMCSA 1c (0.4:0.6) in CDCl₃ (400 MHz), $[1c] + [(\pm)-G1] = 10$ mM.



Figure S110. ¹H NMR spectrum of (±)-G1 with TAMCSA 1c (0.3:0.7) in CDCl₃ (400 MHz), $[1c] + [(\pm)-G1] = 10$ mM.



3528 3394 9810 7762 7626 7352 7352 04530 0462 Ď j 357 871(.113

Figure S111. ¹H NMR spectrum of (±)-G1 with TAMCSA 1c (0.8:0.2) in CDCl₃ (400 MHz), $[1c] + [(\pm)-G1] = 10$ mM.



Figure S112. ¹H NMR spectrum of (±)-G1 with TAMCSA 1c (0.1:0.9) in CDCl₃ (400 MHz), $[1c] + [(\pm)-G1] = 10$ mM.

ESI mass spectrum of complex of (±)-G1 with TAMCSA 1c.



Figure S113. ESI mass spectrum of complex of dipeptide derivative (±)-G1 with TAMCSA 1c.

References

- (a) T. Shono, N. Kise, H. Oike, M. Yoshimoto, E. Okazaki, *Tetrahedron Lett.*, 1992, 38, 5559-5562;
 (b) N. Kise, H. Oike, E. Okazaki, M. Yoshimoto and T. Shono, *J. Org. Chem.*, 1995, 60, 3980-3992;
 (c) T. J. Wenzel and J. E. Thurston, *J. Org. Chem.*, 2000, 65, 1243-1248;
 (d) N. Kise, T. Iwasaki, Y. Yasuda and T. Sakurai, *Tetrahedron Lett.*, 2008, 49, 7074-7077.
- G. P. Gao, C. X. Lv, Q. J. Li, L. Ai and J. X. Zhang, *Tetrahedron Lett.*, 2015, 56, 6742-6746.
- 3 M. R. Davis, E. K. Singh, H. Wahyudi, L. D. Alexander, J. B. Kunicki, L. A. Nazarova, K. A. Fairweather, A. M. Giltrap, K. A. Jolliffe and S. R. McAlpine, *Tetrahedron*, 2012, 68, 1029-1051.