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De novo synthesis of alkyne substituted tryptophans as chemical probes for protein profiling studies

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General experimental: All reagents and solvents were obtained from commercial suppliers and were used as-is without further purification. Anhydrous solvents were obtained from our in-house solvent purification system wherever necessary. NMR spectra were recorded on a 500 MHz ¹H, 125 MHz ¹³C NMR spectrometer at 25 °C. Chemical shifts are reported in ppm (δ) referenced to the NMR solvent residual peak, and coupling constants (*J*) are in hertz. Silica gel flash column chromatography was used to purify all compounds. All reactions were monitored using TLC and LTQ-MS. Reactions were carried out under N₂ atmosphere wherever necessary. Pd(PPh₃)₄ was obtained from Strem Chemicals, and PdCl₂(dppf) was obtained from Acros Organics. For characterization of new compounds, ¹H, ¹³C NMR and LTQ-MS data has been included, whereas for known compounds only ¹H NMR data is reported along with appropriate literature reference.

General procedure for 9-BBN protection of tryptophan:-

Appropriate tryptophan (1eq) was suspended in anhydrous THF (20 mL) and vigorously stirred for 5 min. A solution of 9-BBN in THF (0.5 M, 2.2 eq) was added via syringe and the resulting suspension was stirred under N_2 at ambient temperature until all materials had dissolved (~12 h). The solvents were evaporated to obtain a crude oil from which the product was precipitated by addition of cyclohexane, filtered, and recrystallized from CH₃CN-water to yield corresponding –bicyclononylboronate tryptophan in near quantitative yields.

(1R,5S)-4'-((5-bromo-1H-indol-3-yl)methyl)-9l4-boraspiro[bicyclo[3.3.1]nonane-9,2'-[1,3,2]oxazaborolidin]-5'-one (1): Buff solid. ¹H NMR (d_6 -DMSO, 500 MHz) δ 11.13 (s, 1H), 7.74 (s, 1H), 7.37 (s, 1H), 7.32 (d, J = 8.4 Hz, 1H), 7.17 (d, J = 8.4 Hz, 1H), 6.50-6.46 (br m, 1H), 5.55-5.51 (br m, 1H), 3.84-3.82 (br s, 1H), 3.23-3.19, br m, 1H), 3.10-3.06 (br m, 1H), 1.76-1.39 (br, 12H), 0.40 (d, J = 58.0 Hz, 2H) ppm ; ¹³C NMR (d_6 -DMSO, 125 MHz) δ 173.8, 135.4, 129.5, 126.6, 123.8, 121.1, 113.8, 111.6, 109.6, 55.5, 31.7, 31.6, 26.8, 26.5, 24.6, 24.3, 22.2 ppm; LTQ-MS Calcd for C₁₉H₂₄BBrN₂O₂ – 403.13; found 403.21.

(1R,5S)-4'-((6-bromo-1H-indol-3-yl)methyl)-9l4-boraspiro[bicyclo[3.3.1]nonane-9,2'-[1,3,2]oxazaborolidin]-5'-one (2): Pale yellow solid. ¹H NMR (d_6 -DMSO, 500 MHz) δ 11.11 (s, 1H), 7.54 (s, 1H), 7.50 (d, J = 8.4 Hz, 1H), 7.35 (s, 1H), 7.11 (d, J = 8.4 Hz, 1H), 5.55-5.51 (br m, 1H), 3.84-3.82 (br s, 1H), 3.25-3.21, br m, 1H), 3.12-3.07 (br m, 1H), 1.71-1.32 (br, 12H), 0.40 (d, J = 58.0 Hz, 2H) ppm ; ¹³C NMR (d_6 -DMSO, 125 MHz) δ 173.8, 137.5, 126.6, 125.9, 121.7, 120.5, 114.3, 114.2, 110.0, 55.4, 31.7, 31.6, 31.1, 26.8, 26.5, 24.6, 24.3 ppm; LTQ-MS Calcd for C₁₉H₂₄BBrN₂O₂ – 403.13; found 403.26.

(1R,5S)-4'-((5-hydroxy-1H-indol-3-yl)methyl)-9l4-boraspiro[bicyclo[3.3.1]nonane-9,2'-[1,3,2]oxazaborolidin]-5'-one (5): Off-white crystalline solid. ¹H NMR (d_6 -DMSO, 500 MHz) δ 10.61 (s, 1H), 8.63 (s, 1H), 7.19 (s, 1H), 7.14 (d, J = 8.4 Hz, 1H), 6.84 (s, 1H), 6.60 (d, J = 8.4 Hz, 1H), 6.53-6.49 (br m, 1H), 5.37-5.33 (br m, 1H), 3.81-3.78 (br s, 1H), 3.19-3.15, (br m, 1H), 3.04-2.99 (br m, 1H), 1.73-1.23 (br, 12 H), 0.42 (d, J = 43.5 Hz, 1H) ppm ; ¹³C NMR (d_6 -DMSO, 125 MHz) δ 173.8, 15.8, 131.2, 128.2, 125.2, 112.2, 111.8, 108.4, 102.5, 55.4, 31.7, 31.6, 31.1, 31.0, 26.9, 24.6, 24.3 ppm: LTQ-MS Calcd for C₁₉H₂₅BN₂O₃ – 340.23; found 339.23.

General procedure for BOC protection of bicyclonylboronate tryptophan:-

To a stirred solution of bicyclononylboronate tryptophan (1 eq) in anhydrous DCM (25 mL) was added Boc-anydride (2.5 eq) and cat. dimethylamino pyridine (0.1 eq). The resultant reaction mixture was allowed to stir for 12 h following which the reaction was quenched with sat. NH_4Cl solution and extracted with DCM. The organic layer was washed with water followed by brine, dried over Na_2SO_4 , filtered and evaporated solvents over a rotatory evaporator to give corresponding BOC protected tryptophan moieties.

tert-butyl-5-bromo-3-(((1R,5S)-5'-oxo-9l4-boraspiro[bicyclo[3.3.1]nonane-9,2'-[1,3,2]oxazaborolidin]-4'-yl)methyl)-1H-indole-1-

carboxylate (3): off-white solid. ¹H NMR (d_6 -DMSO, 500 MHz) δ 7.97 (d, J = 8.4 Hz, 1H), 7.84 (s, 1H), 7.74 (s, 1H), 7.47 (d, J = 8.4 Hz, 1H), 6.44-6.40 (Br m, 1H), 5.86-5.82 (br m, 1H), 3.93-3.89 (br s, 1H), 3.26-3.20, br m, 1H), 3.05-3.00 (br m, 1H), 1.74-1.53 (br, 21 H), 0.47 (s, 2H) ppm ; ¹³C NMR (d_6 -DMSO, 125 MHz) δ 173.5, 149.9, 134.1, 132.6, 127.3, 126.6, 122.3, 116.9, 115.8, 115.6, 84.6, 54.6, 31.7, 31.1, 31.0, 28.1, 28.0, 25.9, 24.6, 24.3 ppm; LTQ-MS Calcd for C₂₄H₃₂BBrN₂O₄ – 503.24; found 503.29.

tert-butyl-6-bromo-3-(((1R,5S)-5'-oxo-9l4-boraspiro[bicyclo[3.3.1]nonane-9,2'-[1,3,2]oxazaborolidin]-4'-yl)methyl)-1H-indole-1carboxylate (4): light brown solid. ¹H NMR (d_6 -DMSO, 500 MHz) δ 8.20 (s, 1H), 7.71 (s, 1H), 7.60 (d, J = 8.4 Hz, 1H), 7.42 (d, J = 8.4 Hz, 1H), 6.44-6.41 (br m, 1H), 5.87-5.85 (br m, 1H), 3.89-3.87 (br s, 1H), 3.25-3.21 (br m, 1H), 3.06-3.01 (br m, 1H), 1.76-1.51 (br, 21 H), 0.46 (s, 2H) ppm; ¹³C NMR (d_6 -DMSO, 125 MHz) δ 173.4, 149.2, 129.7, 125.9, 125.8, 121.5, 117.8, 117.5, 116.4, 84.7, 69.5, 54.6, 32.3, 31.7, 31.1, 31.0, 28.1, 28.0, 26.5, 26.0, 24.6, 24.3, 22.2 ppm; LTQ-MS Calcd for C₂₄H₃₂BBrN₂O₄ – 503.24; found 503.19.

NMR data and literature references for intermediate compounds in Scheme 4:

3-benzyl-1H-indole (7)¹: brown oil. ¹H NMR (*CDCl*₃, 500 MHz) δ 8.13 (br s, 1H), 7.51 (d, *J* = 8.0 Hz, 1H), 7.39 (d, *J* = 8.0 Hz, 1H), 7.33-7.26 (br m, 4 H), 7.22-7.16 (obscured t, 2 H), 7.06 (t, *J* = 7.5 Hz, 1H), 7.02 (br s, 1H), 4.14 (s, 2H) ppm.

3-(prop-2-yn-1-yl)-1H-indole (8)²: light yellow oil. ¹H NMR (CDCl₃, 500 MHz) δ 7.95 (br s, 1H), 7.67 (d, *J* = 8.0 Hz, 1H), 7.36 (d, *J* = 8.0 Hz, 1H), 7.24 (t, *J* = 6.5 Hz, 1H), 7.19-7.16 (obscured t, 1 H), 3.73 (s, 2H), 2.17 (s, 1H) ppm.

methyl 2-((diphenylmethylene)amino)-3-hydroxypropanoate (9)³: white crystalline solid. ¹H NMR (d_6 -DMSO, 500 MHz) δ 7.54-7.18 (m, 10H), 4.93 (br, 0.5H), 4.01-3.97 (m, 0.5H), 3.91 (d, J = 16.5 Hz, 1H), 3.86-3.82 (m, 0.5H), 3.65 (s, 3H), 3.56 (d, J = 9.5 Hz, 0.5H) ppm: LTQ-MS Calcd for C₁₇H₁₇NO₃ – 283.33; found 284.14.

Synthesis procedures and NMR data for intermediate compounds in Table 3:

5-ethynyl-1H-indole: brown solid. ¹H NMR (*d*₆-DMSO, 500 MHz) δ 11.28 (s, 1H), 7.69 (s, 1H), 7.38 (d, *J* = 8.4 Hz, 1H), 7.16 (d, *J* = 8.0 Hz, 1H), 6.44 (s, 1H), 3.87 (s, 1H) ppm.

5-(prop-2-yn-1-yloxy)-1H-indole (14)^{4, 5}: light brown solid. ¹H NMR (d_6 -DMSO, 500 MHz) δ 10.94 (s, 1H), 7.28 (s, 1H), 7.11 (s, 1H), 6.76 (d, J = 10.5 Hz, 1H), 6.33 (s, 1H), 4.73 (s, 2H), 3.48 (s, 1H) ppm; LTQ-MS Calcd for C₁₁H₉NO – 171.20; found 172.17 (M + H).

1-(prop-2-yn-1-yl)-5-(prop-2-yn-1-yloxy)-1H-indole (15): During the o-alkylation process of 5-hydroxy indole, 15 was obtained as a minor product (~15% yield) during flash chromatography purification. ¹H NMR (d_6 -DMSO, 500 MHz) δ 7.41 (d, J = 8.4 Hz, 1H), 7.35 (s, 1H), 7.14(s,

1H), 6.87(s, 1H), 5.04 (s, 2H), 4.76 (s, 2H), 3.49 (s, 1H), 3.36 (s, 1H) ppm; ¹³C NMR (*d*₆-DMSO, 125 MHz) δ 152.1, 131.6, 129.5, 129.1, 112.4, 111.1, 104.8, 101.5, 80.3, 79.7, 78.2, 75.8, 56.5, 35.6 ppm; LTQ-MS Calcd for C₁₄H₁₁NO – 209.25; found 208.37 (M – H).
6-iodohex-1-yne⁶: light brown oil. ¹H NMR (*CDC*l₃, 500 MHz) δ 3.21 (br s, 2H), 2.22 (br s, 2H), 1.96 (s, 2H), 1.94 (merged s, 1H), 1.65 (br s, 2H) ppm

5-(hex-5-yn-1-yloxy)-1H-indole (16): To a clean and dry flask with stir bar and acetone (20 mL), 5-hydroxyindole (1 eq), 6-iodohex-1-yne (1.5 eq) and Cs₂CO₃ (3 eq) were added and set for reflux. After 2h, an LTQ MS analysis confirmed the formation of desired product. The reaction mixture was allowed to cool to room temperature, quenched with NH₄Cl and extracted with DCM. The organic layer was washed with water and brine, dried over Na₂SO₄, filtered and solvents were evaporated on a rotatory evaporator. The crude was then purified by flash chromatography using 2:1 Hexanes:Ethyl Acetate system to give **16** as a pure white solid. ¹H NMR (d_6 -DMSO, 500 MHz) δ 10.86 (s, 1H), 7.24 (s, 1H), 7.02 (s, 1H), 6.70 (br s, 1H), 6.30 (s, 1H), 3.93 (br s, 2H), 2.76 (s, 1H), 2.23 (br, 2H), 1.80-1.78 (br m, 2H), 1.62-1.59 (br m, 2H) ppm; ¹³C NMR (d_6 -DMSO, 125 MHz) δ 152.9, 131.5, 128.4, 126.1, 112.3, 112.0, 103.2, 101.2, 84.8, 71.7, 67.8, 28.5, 25.2, 17.9 ppm; LTQ-MS Calcd for C₁₄H₁₆NO – 213.28; found 214.19 + Na (236.09)

1H-indol-5-yl hex-5-ynoate (17): Standard EDC coupling conditions were used to synthesize compound **17**. After purification, white solid was obtained. ¹H NMR (d_6 -DMSO, 500 MHz) δ 11.15 (s, 1H), 7.38 (s, 1H), 7.24 (s, 1H), 6.81 (br s, 1H), 6.41 (s, 1H), 2.85 (s, 1H), 2.67-2.65 (t, J = 6.5 Hz, 2H), 2.29 (br m, 2H), 1.84-1.81 (m, 2H) ppm; ¹³C NMR (d_6 -DMSO, 125 MHz) δ 172.5, 143.9, 134.1, 128.1, 127.1, 115.7, 112.4, 101.6, 84.1, 72.3, 32.9, 23.9, 17.6 ppm; LTQ-MS Calcd for C₁₄H₁₃NO₂ – 227.26; found 226.15 (M – H).

Synthesis of methyl 2-((diphenylmethylene)amino)acrylate (12) using Appel Reaction:

To a clean and dry 500 mL flask with stir bar under a N₂ purge was added **9** (1.0 g, 3.53 mmol, 1 eq) dissolved in DCM (250 mL). This was followed by the addition of CBr₄ (1.76 g, 5.29 mmol, 1.5 eq) and the reaction mixture was allowed to stir for 20 mins. PPh₃ (1.85 g, 7.06 mmol, 2.0 eq) was added slowly over a period of 10 mins making sure that the reaction is under a constant flow of N₂. During the addition of PPh₃, a dark brown coloration is observed which disappears on the addition of additional DCM (100 mL). A TLC at this point (30 mins) confirms the disappearance of **9**, following which the solvents are evaporated over a rotatory evaporator, contents are transferred to a separatory funnel and washed with NaHCO₃, followed by sat. Na₂S₂O₃. The organic layer is finally washed with brine, Na₂SO₄, filtered and solvents were evaporated on a rotatory evaporator. The crude was then purified by flash chromatography using 2:1 Hexanes:Ethyl Acetate system to give **12** as a pale yellow oil (1.23g, 69%). ¹H NMR (*d*₆-DMSO, 500 MHz) δ 7.79 (br, 0.5H), 7.71 (s, 2H), 7.57-755 (br, 2H), 7.45 (s, 4H), 7.23 (s, 1.5H), 5.44 (s, 1H), 4.82 (s, 1H), 3.63 (s, 3H) ppm; LTQ-MS Calcd for C₁₇H₁₅NO₂ – 265.31; found 265.21.

Important Notes for this protocol:

- 1. During the addition of PPh₃, if the brown color persists, add DCM until the color turn bight pale yellow.
- 2. Washing with bicarbonate and thiosulfate is very important.

- 3. Compound 12 has been found to dimerize (compd 13) on standing, so it is safer to store in DCM solution at -80 °C or use immediately.
- 4. The reaction image below shows the course of reaction monitored via TLC.



General synthesis procedure for substituted tryptophan moieties in Table 37:

To a clean and dry flask with stir bar was added corresponding indole compound (1 eq), **12** (1 eq) dissolved in DCM. The reaction mixture was cooled in an ice bath and purged with N_2 for 20 mins. AlCl₃ (2.0 eq) was added and allowed to stir for 2h. After confirming the completion of reaction by TLC, the reaction mixture was quenched with NaHCO₃, filtered over a bed of Celite, washed the organic layer with water and brine, dried over Na₂SO₄, filtered and evaporated the solvents over a rotatory evaporator. The crude obtained from this process was then subjected to flash purification using 2:1 Hexane:Ethyl Acetate.

Methyl-2-((diphenylmethylene)amino)-3-(1H-indol-3-yl)propanoate (11): ¹H NMR (CDCl₃, 500 MHz) δ 7.91 (br s, 1H), 7.59 (br, 2H), 7.32-7.28 (m, 3H), 7.21 (d, J = 7.5 Hz, 1H), 7.15-7.10 (m, 3H), 6.97 (br, 1H), 6.92 (t, J = 7.5 Hz, 1H), 6.59 (br, 2H), 4.42 (br, 1H), 3.72 (s, 3H), 3.47 (obscured br d, 1H), 3.24 (br s, 1H) ppm; LTQ-MS Calcd for C₂₅H₂₂N₂O₂ – 382.46; found 383.22 (M + H).

Methyl-2-((diphenylmethylene)amino)-3-(5-ethynyl-1H-indol-3-yl)propanoate (18): LTQ-MS Calcd for $C_{27}H_{22}N_2O_2 - 406.49$; found 407.21 (M + H).

Methyl-2-((diphenylmethylene)amino)-3-(5-(prop-2-yn-1-yloxy)-1H-indol-3-yl)propanoate (21): ¹H NMR (d_6 -DMSO, 500 MHz) δ 10.66 (s, 1H), 7.47 (d, J = 7.5 Hz, 2H), 7.44-7.41 (obscured t, 1H), 7.35 (t, J = 7.5 Hz, 3H), 7.27 (t, J = 7.5 Hz, 2H), 7.19 (br d, J = 15 Hz, 1H), 6.95 (br, 1H), 6.69 (br, 2H), 6.67 (br, 2H), 4.51 (s, 1H), 4.21 (t, J = 7.0 Hz, 1H), 3.60 (s, 3H), 3.49 (s, 1H), 3.26 (dd, J = 14.0, 5.5 Hz, 1H), 3.03 (dd, J = 14.0, 8.0 Hz, 1H) ppm; ¹³C (d_6 -DMSO, 125 MHz) δ 172.2, 169.6, 151.4, 139.3, 135.7, 132.0, 130.8, 128.8, 128.7, 128.6, 127.6, 125.1, 112.1, 110.1, 102.7, 80.3, 78.0, 66.6, 56.8, 52.2, 29.5 ppm; LTQ-MS Calcd for C₂₈H₂₄N₂O₃ – 436.51; found 437.22 (M + H).

Methyl-2-((diphenylmethylene)amino)-3-(1-(prop-2-yn-1-yl)-5-(prop-2-yn-1-yloxy)-1H-indol-3-yl)propanoate (22): ¹H NMR (d_6 -DMSO, 500 MHz) δ 7.47-7.41(m, 4H), 7.35-7.28 (m, 6H), 7.01 (s, 1H), 6.78 (br, 2H), 6.70 (br s, 1H), 4.94 (s, 2H), 4.53 (s, 2H), 4.20 (br s, 1H), 3.61 (s, 3H), 3.50 (s, 1H), 3.24 (obscured dd, 1H), 3.05-3.01 (obscured dd, 1H) ppm; ¹³C (d_6 -DMSO, 125 MHz) δ 172.2, 169.6, 151.4, 139.3, 135.7, 132.1, 130.8, 128.8, 128.7, 128.6, 128.0, 127.6, 125.1, 112.2, 112.1, 110.1, 102.7, 80.3, 78.0, 66.6, 56.8, 52.3, 29.2 ppm; LTQ-MS Calcd for C₃₁H₂₆N₂O₃ – 474.56; found 475.20 (M + H).

Methyl-2-((diphenylmethylene)amino)-3-(5-(hex-5-yn-1-yloxy)-1H-indol-3-yl)propanoate (23): ¹H NMR (d_6 -DMSO, 500 MHz) δ 10.60 (s, 1H), 7.46 (d, J = 7.5 Hz, 2H), 7.43-7.40 (m, 1H), 7.35 (br s, 3H), 7.29-7.26 (obscured t, 2H), 7.15 (d, J = 9.0 Hz, 1H), 6.92 (s, 1H), 6.67-6.62 (m, 3H), 6.58 (s, 1H), 4.21 (s, 1H), 3.71-3.68 (obscured dd, 2H), 3.59 (s, 3H), 3.25 (obscured dd, 1H), 3.04-3.01 (obscured dd, 1H) 2.79 (s, 1H), 2.23 (br s, 2H), 1.75 (br s, 2H), 1.63-1.58 (br m, 2H) ppm; ¹³C (d_6 -DMSO, 125 MHz) δ 172.2, 169.6, 152.6, 139.3, 135.8, 131.6, 130.8, 128.8, 128.7, 128.6, 128.5, 127.6, 125.1, 112.0, 110.0, 101.5, 84.8, 71.8, 67.7, 66.8, 52.2, 29.6, 28.5, 25.3, 17.9 ppm; LTQ-MS Calcd for C₃₁H₃₀N₂O₃ – 478.59; found 479.27 (M + H).

3-(2-((diphenylmethylene)amino)-3-methoxy-3-oxopropyl)-1H-indol-5-yl hex-5-ynoate (24): ¹H NMR (d_6 -DMSO, 500 MHz) δ 10.87 (s, 1H), 7.48 (d, J = 7.5 Hz, 2H), 7.44-7.41 (m, 1H), 7.36 (br d, 3H), 7.34-7.31 (obscured t, 2H), 7.27 (d, J = 9.0 Hz, 1H), 6.78-6.75 (m, 2H), 6.73 (s, 1H), 4.19 (s, 1H), 3.57 (s, 3H), 3.27-3.24 (obscured dd, 1H), 3.00-2.96 (obscured dd, 1H) 2.88 (s, 1H), 2.67-2.64 (br t, 2H), 2.32 (br s, 2H), 1.84-1.81 (br t, 2H) ppm; ¹³C (d_6 -DMSO, 125 MHz) δ 172.2, 169.8, 143.6, 139.1, 135.8, 134.2, 128.9, 128.8, 128.6, 128.5, 127.6, 125.7, 115.8, 111.9, 110.5, 84.1, 72.3, 66.8, 52.2, 32.8, 29.5, 23.9, 17.6 ppm; LTQ-MS Calcd for C₃₁H₂₈N₂O₄ – 492.58; found 493.27 (M + H).

Deprotection of compound 19 to amine 23 and amino acid 24 (Scheme 4): Protocol as listed in Org. Lett. 2012, 14, 600-603

Compound **19** (1 eq) was dissolved in THF, to it was added HCl (1N, 5 eq) and the reaction mixture was allowed to stir for 6 h. After confirming the completion of reaction by TLC, the contents were diluted with water and extracted with Et_2O . The organic layer, which would now consist of benzophenone protecting group was separated using a separatory funnel. The aqueous layer was neutralized to pH 7.0 using 1N NaOH following which the aqueous content were evaporated via a rotatory evaporator to reduce the contents to minimum. Finally the crude layer was purified

using flash chromatography using 10% MeOH in DCM, collecting fractions corresponding to desired product, evaporated the solvents to give amine 23 as a brown solid. After characterizing by ¹H NMR and LTQ-MS, 23 was subjected to ester deprotection. 23 was added to a flask containing THF:H₂O (1:1) and to it was added LiOH (5 eq). The reaction mixture was allowed to stir overnight and analyzed by LTQ-MS to confirm formation of product. The contents were lyophilized to give 24 as a light brown solid.

Methyl-2-amino-3-(5-(prop-2-yn-1-yloxy)-1H-indol-3-yl)propanoate (23): ¹H NMR (d_6 -DMSO, 500 MHz) δ 10.86 (s, 1H), 7.26 (d, J = 9.0 Hz, 1H), 7.15 (s, 1H), 7.08 (s, 1H), 6.78 (d, J = 8.5 Hz, 1H), 6.02 (br, 2H for NH₂), 4.74 (s, 2H), 4.01 (br, 1H), 3.63 (s, 3H), 3.11-3.08 (br t, 2H) ppm; LTQ-MS Calcd for C₂₈H₂₄N₂O₃ – 272.20; found 273.19 (M + H).

2-amino-3-(5-(prop-2-yn-1-yloxy)-1H-indol-3-yl)propanoic acid (24): LTQ-MS - m/z 258.1

Dimethyl (E)-2,5-bis((diphenylmethylene)amino)hex-2-enedioate (13): During the process of *de novo* tryptophan synthesis from corresponding substituted indoles and compound **12**, a side product is also obtained. After flash chromatography purification, the fractions corresponding to an unknown compound were isolated and evaporated to dryness to give a yellow solid. ¹H NMR (d_6 -DMSO, 500 MHz) δ 7.74 (br m, 2H), 7.58-7.46 (br m, 12H), 7.38-7.29 (br m, 5H), 7.11-7.03 (br d, 4H), 5.78 (br s, 1H), 4.03 (br s, 1H), 3.62 (s, 3H), 3.47 (s, 3H), 2.53 (merged with dmso reference, 2H) ppm; ¹³C (d_6 -DMSO, 125 MHz) δ 171.6, 170.6, 170.5, 163.8, 141.1, 139.2, 138.4, 137.5, 136.6, 135.8, 133.1, 131.7, 130.1, 129.6, 129.3, 129.2, 129.0, 128.8, 128.7, 128.5, 127.9, 127.7, 121.4, 64.0, 52.5, 52.1, 31.5 ppm; LTQ-MS Calcd for C₃₄H₃₀N₂O₄ – 530.22; found 531.23 (M + H). The solid was redissolved in DCM and allowed to crystallize via buffer diffusion using Hexanes to give X-ray quality crystals enabling X-ray crystallography data.

The structure was solved in space group $P^{\overline{1}}$. The two (Ph)₂C=N-C-CO₂Me portions were related through the inversion center, resulting in 2 bridging –CH-CH₂- moieties. Each of these refined to approximately equal occupancy and so were set to 50% for the final refinements. The final R-factor of 6.19% indicated that this is an excellent description of the structure.





Ellipsoid view of crystal structure of **13**. The space group symmetry results in two linking –CH-CH₂- moieties, each with 50% occupancy. One of these has been removed for clarity.



Crystal structure of **13** showing the two half-occupied linking groups colored in green and pink respectively.

Experimental procedure for Protein synthesis studies:

The cells were washed once with PBS (pH=7.4), resuspended in PBS in the presence of EDTA free protease inhibitor (Roche), sonicated (Fisher Scientific, model FB120) using a total of 72, 2-second pulses at 60% amplitude followed by a 21,000 g spin. Protein concentration was measured using the BCA reagent (ThermoFisher), and normalized to 1 μ g/ μ L with PBS for click chemistry. Click chemistry was performed using, 3 μ M azido-tetramethylrhodamine (LumiProbe) in the presence of 500 μ M copper sulfate, 50 μ M TBTA (tris[(1-benzyl-1H-1,2,3-triazol-4-yl)methyl])- amine) in a 4:1 solution of t-butanol/DMSO, and 500 μ M TCEP (tris(2-carboxyethyl)phosphine). After click chemistry, 2× SDS and reducing agent was added to the samples and incubated at 95°C for 5 min. 15 μ g of protein was run on a 10-20% Tris-Glycine Gel (ThermoFisher) with the ECL Ranibow Ladder (Amersham). Rhodamine-labeled proteins were detected using a FluorChemQ fluorescence imager (Protein Simple).





































Indole NH peak at 11.0 ppm.

Indole NH peak @ 10.71 ppm;

OH peak @ 8.55 ppm

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