

## Supporting Information

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### **Ultrasensitive and Unambiguous Bacterial Pathogen Detection through Super Selective interactions between Multivalent Supramolecular Immuno-Nanoparticles (SINs)**

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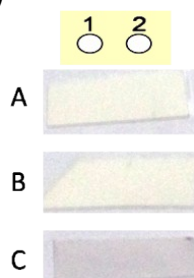
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#### **SUPPORTING FIGURES & TABLES**

#### **Figure S1. Evidence for secondary antibody never binds non-specifically to bacterial cells**

Two different strains were spotted on the membrane ( $10^6$  cells/spot). Blocked with BSA and incubated in different dilutions of direct secondary antibody (A- 1:5000 Secondary antibody, B- 1:2500 Secondary antibody, C- 1:1000 Secondary antibody)



No spots were observed in any of the membrane which implies that the secondary antibody does not bind non-specifically to the bacterial membrane.

**Table S2. Details of strains used**

S.No	Strain code	Strains	S.No	Strain code	Strains
1.	EP1	<i>E.coli</i>	2.	UPEC 129	<i>Uropathogenic E.coli</i>
3.	T56	<i>E.coli</i>	4.	Proteus 487	<i>Proteus</i>
5.	EC1021	<i>E.coli</i>	6.	E.coli 171	<i>E.coli</i>
7.	Pseudo	<i>Pseudomonas aeruginosa</i>	8.	UPEC 475	<i>Uropathogenic E.coli</i>
9.	Acint	<i>Acinetobacter</i>	10.	UPEC 149	<i>Uropathogenic E.coli</i>
11.	Shi.f	<i>Shigella flexneri</i>	12.	Klebsiella 4483	<i>Klebsiella pneumonia</i>
13.	M8	<i>Staphylococcus aureus</i>	14.	S.aureus 3160	<i>Staphylococcus aureus</i>
15.	T113	<i>Klebsiella pneumonia</i>	16.	E.coli 422790	<i>E.coli</i>
17.	M4	<i>Salmonella enterica</i>	18.	S.dysenteriae 1031	<i>Shigella dysenteriae</i>
19.	M11	<i>Proteus mirabilis</i>	20.	UPEC 139	<i>Uropathogenic E.coli</i>
21.	M30	<i>Shigella dysenteriae</i>	22.	S.flexneri 1457	<i>Shigella flexneri</i>
23.	B.C	<i>Bacillus cereus</i>	24.	Proteus 282	<i>Proteus</i>
25.	E.coli 723	<i>E.coli</i>	26.	S.dysenteriae 1021	<i>Shigella dysenteriae</i>
27.	Klebsiella 340053	<i>Klebsiella pneumonia</i>	28.	Klebsiella 3384	<i>Klebsiella pneumonia</i>
29.	Proteus 7002	<i>Proteus mirabilis</i>	30.	E.coli 422790	<i>E.coli</i>
31.	UPEC 478	<i>Uropathogenic E.coli</i>	32.	S.flexneri 9543	<i>Shigella flexneri</i>
33.	S.dys 1311	<i>Shigella dysenteriae</i>	34.	S.aureus 25923	<i>Staphylococcus aureus</i>

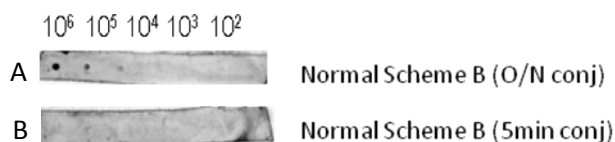
**Table S3. Negative control experiment in ELISA**

ELISA experiment was performed with two different primary S/Ns – specific primary S/Ns (rabbit antibody) and inert or non-specific primary S/Ns (chicken antibody), one that does not bind to our secondary antibody (Anti rabbit antibodies).

<b>Secondary antibody dilution</b>	<b>Scheme B (Full Non-specific)</b>	<b>Scheme D (Full Non-specific)</b>
0.004	0.167	0.057
0.002	0.044	0.024
0.001	0.015	0.018
0.0005	0.023	0.007
0.00025	0.004	0.013
0.000125	0.006	0.019
6.25E-05	0.018	0.015
3.13E-05	0.011	0.012
1.56E-05	0.008	0.017

**Figure S4. Surface density experiment-5min conjugation of primary antibody to GNP**

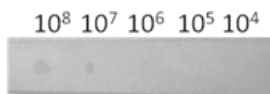
$10^6$  to  $10^2$  CFU of bacterial cells were spotted on the membrane and dot blot assay was carried out with 5min conjugated primary SINS (B) and overnight conjugated primary SINS as control (A).



When the antibodies were conjugated for only 5min to GNPs, no spots were observed on the membrane in none of the dilution of cells spotted. This can be due to the insufficient binding of primary antibodies to the GNPs due to short time incubation.

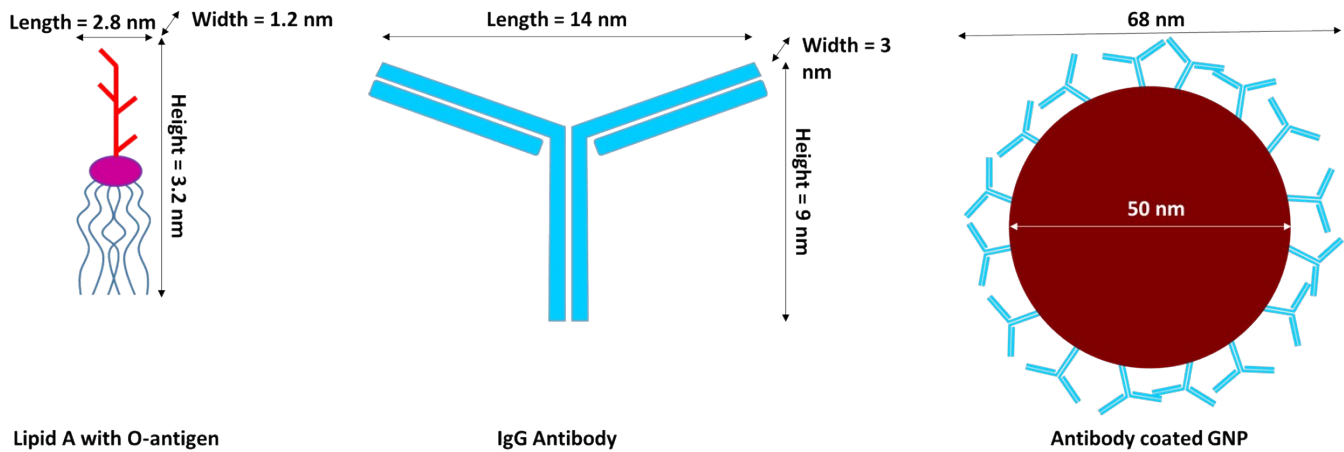
**Figure S5. NBT/BCIP (ALP substrate) direct reaction with bacterial cells**

$10^8$  –  $10^4$  Cells/ $\mu$ l/Spot on nitrocellulose membrane were allowed to react with ALP substrate (NBT/BCIP) directly without primary or secondary antibody



Spot was observed on membrane for  $10^8$  and  $10^7$  CFU cells indicating that bacterial endogenous ALP was reacting directly to the substrate. Hence, cells numbering  $10^6$  CFU or less than  $10^6$  CFU should be spotted on the membrane for the experiment.

**Figure S6. Dimensions of various entities used for calculations**



**Calculations:**

Lipid A with O-antigen area ( $A_O$ ) = Length x Width =  $3.36 \times 10^{-18} \text{ m}^2$

Antibody area ( $A_{Ab}$ ) = Length x Width =  $4.2 \times 10^{-17} \text{ m}^2$

Assuming 90% of the outer membrane is composed of Lipid A,  
 Number of O-antigens buried under a primary antibody =  $SA_{Ab}/SA_O = 11$

Antibody-coated GNP surface area ( $SA_{SIN}$ ) =  $4 \times \pi \times r^2 = 1.45 \times 10^{-14} \text{ m}^2$

Antibody-coated GNP circular area ( $CA_{SIN}$ ) =  $\pi \times r^2 = 3.63 \times 10^{-15} \text{ m}^2$

Maximum number of Antibodies on a GNP =  $SA_{SIN}/SA_{Ab} = 346$

number of O-antigens buried under a SIN =  $CA_{SIN}/A_O = 972$

## SUPPORTING MATERIALS AND METHODS

### M1.Flocculation Assay

In a typical assay, the suitable proportion of protein to be conjugated was identified by mixing different concentrations of antibody to equal amount of gold nanoparticles and kept at room temperature for 30min, followed by addition of 10 $\mu$ l of 10% NaCl. The color change was monitored via absorbance spectra and the gold nanoparticles aggregation can be seen with a peak shift towards 580nm in the samples which has less amount of protein needed to stabilize the fixed amount of nanoparticles. When the antibody molecules are enough to stabilize the gold nanoparticles, then the color change will not take place even after addition of salt.

### M2.Dot blot assay for detection of mixed cell culture

Nitrocellulose membrane were cut into small strips (4 No's) for each scheme and six spots (1-6) were spotted in the following order, each with different combination of cells on each strip.

Spot No.	Combination of cells (Total 10 <sup>6</sup> cells)
1	10 <sup>6</sup> pure positive cells
2	10 <sup>5</sup> positive cells+10 <sup>1</sup> ghost spot negative cells
3	10 <sup>4</sup> positive cells+10 <sup>2</sup> ghost spot negative cells
4	10 <sup>3</sup> positive cells+10 <sup>3</sup> ghost spot negative cells
5	10 <sup>2</sup> positive cells+10 <sup>4</sup> ghost spot negative cells
6	10 <sup>6</sup> Ghost spot negative cells

The experimental procedure was carried out as same as that of pathogen detection for all four schemes

### M3. Simulated Non-specific Surface density dependence of the SInS (Scheme B and Scheme D)

S.No	Scheme B		Scheme D	
1	Fully specific primary SInS	100ng/well	Fully specific primary SInS	100ng/well
2	1(Specific primary SInS) :100 (Non-Specific primary SInS)	100ng/well (1ng+99ng)	1(Specific primary SInS) :100 (Non-Specific primary SInS)	100ng/well (1ng+99ng)
3	1(Specific primary SInS) :1000(Non-Specific primary SInS)	100ng/well (0.1ng+99.9ng)	1(Specific primary SInS) :1000(Non-Specific primary SInS)	100ng/well (0.1ng+99.9ng)

The primary SInS were coated on ELISA plates as mentioned in the table above and kept overnight at room temperature, followed by blocking and subsequent washing steps which are mentioned in the methods for Scheme B and scheme D.

### M4. Dot-blot assay cost analysis

#### Nitrocellulose membrane:

50piece (80mm dia circle) = \$60

1piece = \$1.2

For one reaction (10 samples) we need 1/16<sup>th</sup> of a piece = **\$0.07**

#### Primary Ab:

3ml Serum = \$60

For one reaction we require 3ml of 1:1000 diluted primary antibody = **\$0.06**

#### BSA:

100g = \$90

For one reaction we need 15ml of 0.1% (wt/vol) solution = **\$0.01**

#### Secondary Ab:

1ml = \$225

For one reaction we require 3ml of 1:10000 diluted secondary antibody = **\$0.07**

#### GNP:

1g = \$105

For conjugation we are using only 100µl of 1mM solution (0.04mg of GNP) = **\$0.04**

Total cost per reaction (10 samples) = **\$0.25**