

Supplementary Material

Rapid Preparation of Highly Reliable PDMS Double Emulsion Microfluidic Devices

Shunbo Li^{a*}, Xiuqing Gong^a, Ciara S. Mc Nally^a, Muling Zeng^a, Thembaninkosi Gaule^{a,b}, Clara Anduix-Canto^a, Alexander N. Kulak^a, Lukmaan Abbas Bawazer^a, Michael J. McPherson^b and Fiona C. Meldrum^{a*}

S1. Microfluidic chip design for smaller double emulsion droplets generation

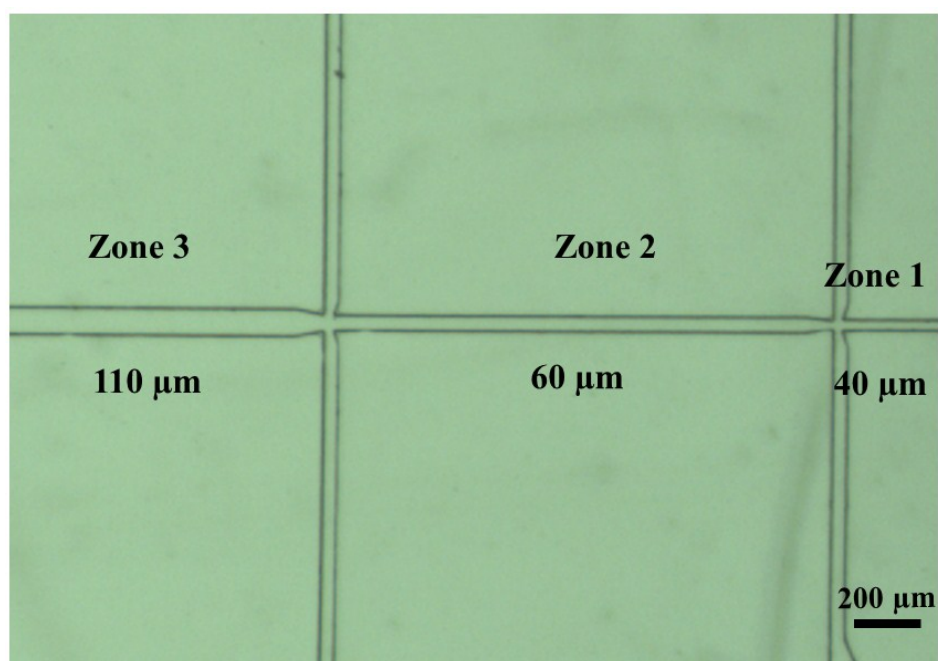


Figure S1. A microfluidic chip that is fabricated to generate smaller double emulsion droplets.

S2. Solvent Extraction of the PDMS

Solvent extraction of the cured PDMS sections was performed using Soxhlet extraction against 2-propanol (Figure S1). The PDMS sections were placed in a Soxhlet thimble, and the solvent raised to its boiling point. Condensation of the vapour causes the solvent to fill the section containing the solid material, washing the PDMS and extracting the uncured oligomers. Once this section is full, the solvent empties into the still pot via a siphon action and the cycle continues. This extraction process was run for 4 days.

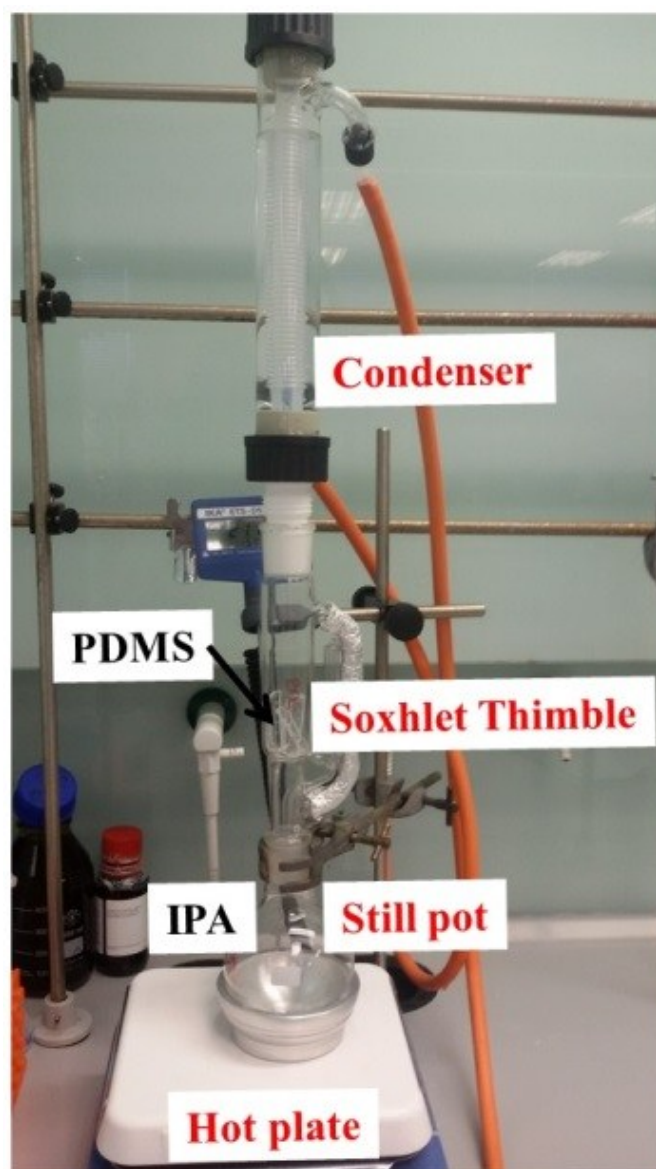


Figure S2. Set-up for PDMS extraction: hot plate, still pot, Soxhlet thimble and condenser. PDMS was put in Soxhlet thimble and 2-propanol (IPA) was used as the solvent.

S3. Statistical Analysis of the Patterning and Coating Results

In order to show the reliability of our chip fabrication method, 12 chips were patterned with epoxy and were then plasma-treated for 2.5 mins. Drops of 0.24 wt% Rhodamine 6G solution were then placed in zones 1 and 3. The region filled with red color is hydrophilic, while the white region is hydrophobic. The results are illustrated in Fig. S3.

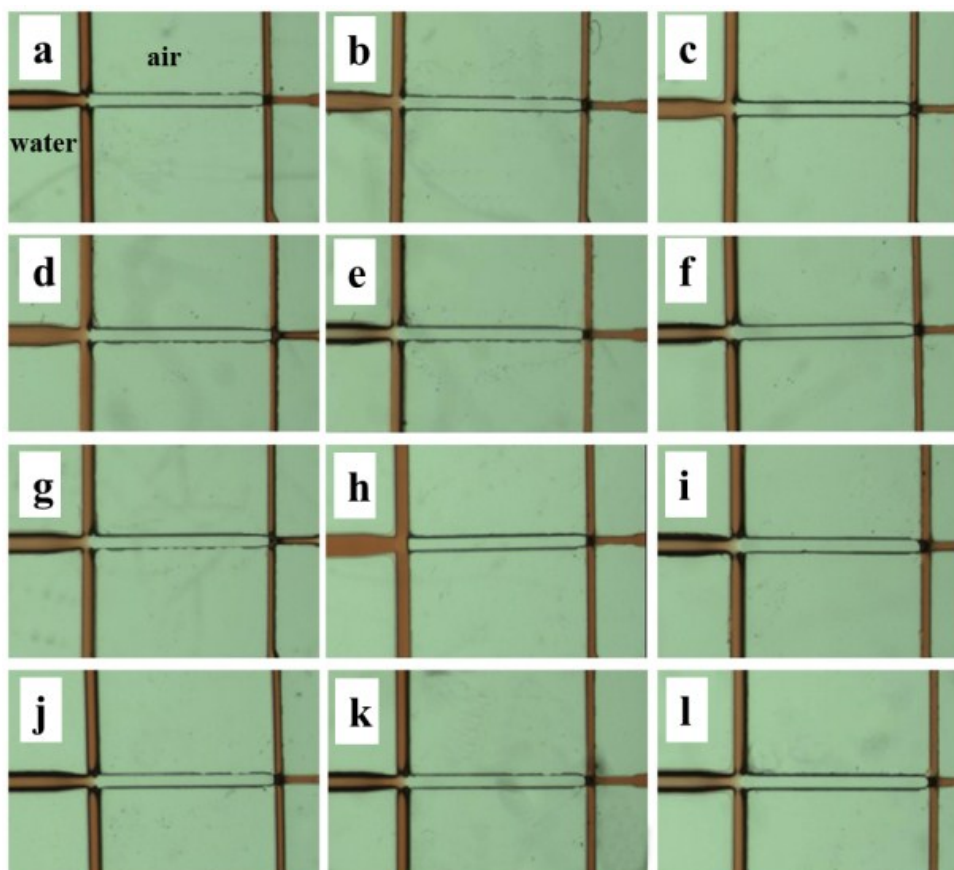


Figure S3. Optical microscope images of 12 chips after patterning, coating and filling with 0.24 wt% Rhodamine 6G solution.

S4. Statistical Analysis of Double Emulsion Formation

The reproducibility of double emulsion generation in the fabricated chips was also investigated. Five chips with the larger channel dimensions (channel depth 110 μm , and widths 85 μm , 150 μm and 250 μm for the first, second and third channel zones) and five with the smaller dimensions (channel depth 23 μm , and widths 40 μm , 60 μm and 110 μm for the first, second and third channel zones) were used to generate double emulsion droplets under flow rates of 40 $\mu\text{l h}^{-1}$ – 50 $\mu\text{l h}^{-1}$ – 500 $\mu\text{l h}^{-1}$ and 30 $\mu\text{l h}^{-1}$ – 50 $\mu\text{l h}^{-1}$ – 700 $\mu\text{l h}^{-1}$ (inner water – oil phase – outer water). The droplets were collected and the size distributions of 1200 droplets are shown below. “Relative frequency” represents the relative number of double emulsion droplets and single phase droplets generated by the prepared devices, as well as the size distribution of cores and shells of double emulsion droplets.

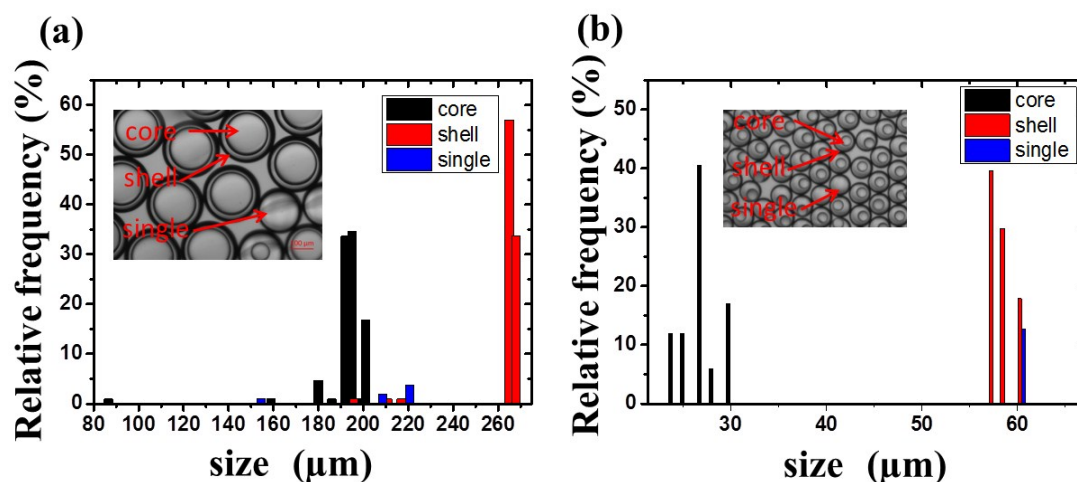


Figure S4. Histograms of 1200 droplets generated by (a) 5 chips with larger channel sizes and (b) 5 chips that generate small double emulsion droplets. “Core” and “shell” represent the core and shell of the double emulsion droplets, and “single” represents single phase droplets (ie double emulsion droplets failed to form). The flow rates were held at 40 $\mu\text{l h}^{-1}$ – 50 $\mu\text{l h}^{-1}$ – 500 $\mu\text{l h}^{-1}$ and 30 $\mu\text{l h}^{-1}$ – 50 $\mu\text{l h}^{-1}$ – 700 $\mu\text{l h}^{-1}$ (inner water – oil phase – outer water) for (a) and (b) respectively.