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Supplementary Information

for

Amphiphilic Peptide Coated Superparamagnetic Iron Oxide Nanoparticles for *in vivo* MR Tumor Imaging



Figure S1. Images of hydrophobic iron oxide nanoparticles in the absence and presence of magnetic field when they are dispersed in hexane (10 mg/mL, concentration based on the iron content)



Figure S2. Number distribution of hydrodynamic diameter of the SPIONs dispersed in hexane.



Figure S3. Liquid chromatogram of Lauryl-PPPGK-Am and mass spectrum of corresponding peptide amphiphile molecule. Mass data $[M-H]^+$ (calculated) = 676.47, $[M-H]^+$ (observed) = 676.48.



Figure S4. Thermal gravimetric analysis of a) SPIONs before assembly with PA, b) SPION/K-PA and peptide samples.



Figure S5. TEM images of peptide coated SPIONs, scale bars: 10 nm and 20 nm, respectively.



Figure S6. a) Dynamic light scattering (DLS) analysis to determine the dispersion stability of peptide coated SPIONs prepared in two different media, b) photographs taken after a week dispersion in corresponding media.



Figure S7. Calibration curve plotted for iron content.



Figure S8. Dose-dependent cytotoxicity profiles of SPION/KPA and K-PA on different cell lines in comparison with the no-treatment group (NT). Cell viability of HUVEC (**a**, **b**), and MCF-7 cells (**c**, **d**) treated with K-PA (50-200 μ g/mL which is equivalent content of iron assembled with PA) and SPION/K-PA (50-200 μ g/mL) at 24 h (left column) or 48 h (right column). Values were compared to those of the untreated controls and are presented as percentages. Data represent means ± standard error of mean (SEM) of a minimum of 3 independent experiments performed in quadruplicates.



Figure S9. Hematoxylin-eosin staining for the tissue morphology of the kidney, liver and spleen of healthy rats (control) prior to or 4 h and 15 days following the injection of SPION/K-PA (5 mg/kg) (original magnification 200x and scale bar:50 μ m).



Figure S10. H&E staining for the tissue morphology of the kidney, liver and spleen of tumor bearing rats (control) prior to or 4 h and 15 days following the injection of SPION/K-PA (5 mg/kg). Photographs were taken at 200x magnification.



Figure S11. Representative photomicrographs of the liver, kidney and spleen of healthy rats (control) prior to or 4 h and 15 days following the injection of SPION/K-PA (5 mg/kg) stained with Prussian blue for iron accumulation. Positive staining appears in blue dots. (Original magnification 200x. Scale bar: $50 \mu m$).



Figure S12. Prussian blue staining for iron accumulation in the tumor, kidney, liver and spleen of timor baring rats (control) prior to or 4 h and 15 days following the injection of SPION/K-PA (5 mg/kg) (original magnification 200x, scale bar is 50 μ m).

2 θ (degrees)	Calculated d-spacing (nm)	JCPDS data (nm)	hkl
30.02	0.2961	0.2967	220
35.43	0.2524	0.2532	311
43.11	0.2096	0.2099	400
53.31	0.1740	0.1714	422
57.11	0.1610	0.1615	511
62.75	0.1479	0.1484	440

Table S1. The d-spacing values (nm) calculated from peak positions in XRD pattern inserted in Figure 2a and standard atomic spacing for magnetite together with respective Miller indices (*hkl*) from the JCPDS card (19-0629).

Table S2. Physical properties of fatty acid/amine capped SPIONs and PA functionalized SPIONs (a: in water, b: in 5% dextrose, c: in hexane).

Sample	TEM size (nm)	DLS size (nm)	Zeta potential (mV)
SPION	5.5±0.6	7.1 ± 0.3^{c}	N/A
SPION/K-PA ^a	5.3±0.8	58.0±0.7	18.3±1.4
SPION/K-PA ^b	N/A	58.6±0.1	23.7±1.7