# Design, synthesis and *in-vitro* anti-leukemic evaluation of ferulic acid analogues as Bcr-Abl inhibitors

Narendran Rajendran, Shankar Subramaniam , Mamilla R. Charan Raja, Himesh Makala Venkata Subbarao, Ulaganathan Venkatasubramanian, Brindha Pemaiah , Santanu Kar Mahapatra\* and Aravind Sivasubramanian\*

Electronic supplementary information (ESI)

# Materials and methods

#### Plant collection and extraction

The aerial parts of *S. brachiata* were collected from the place called killai, near Pichavaram, Tamil Nadu, and India. A voucher specimen was matched with the authentic specimen (herbarium no 5555, regional plant resource center, Puri, India) and was deposited in the herbarium. The aerial parts (~1 kg) were shade dried and powdered. 100 g of the powder was percolated with Millipore water and after 24 h, the extract were filtered through a Whatman No. 41 filter paper. The extraction was repeated thrice and the combined extracts were concentrated in vacuo and further lyophilized to obtain a residue (35 g).

# **Isolation of ferulic acid**

10g of water extract of *S.brachiata* was subjected to column chromatographic separation. Initially the crude extract was separated by using Sephadex LH 20 (Sigma Aldrich, sweden). Sephadex was prepared after the swelling time of 24hrs by using chloroform-methanol (30:70) mixture. There are 18 fractions were collected by increasing polarity of the mobile phase from 30% chloroform: methanol to 100% methanol. The TLC pattern of SB 9 and SB 13 shows similar in spot nature and they were mixed up accordingly. This fraction reveals two spots in major (Rf = 0.488, 0.418) and one more with minor intensity. Further purification was done by using normal silica column (100-200 mesh). Dirty white precipitate was formed along the sides of the test tube. Further purification of the compound yielded 27mg of ferulic acid. The structures of the compound have been identified by using NMR Studies.

#### **Procedure for the synthesis of** (*E*)-**3**-(**4**-acetoxy-**3**-methoxyphenyl) acrylic acid (2)

Ferulic acid (19.4g, 0.1 mol) was added to the solution of NaOH (12.7g, 0.26 mol) in 100 mL water. Then the mixture was stirred and cooled below  $10^{\circ}$ C. To the cold solution acetic anhydride (12.7 mL, 0.125 mol) was added in drop wise manner. The reaction mixture was stirred at  $20^{\circ}$ C for 10min and room temperature for 30min. Then dilute sulfuric acid was added in dropwise till the white precipitate form. Maintain the pH of the solution 4-5 by using dilute

sulfuric acid (5N). Filter the white precipitate and washed several times with distilled water. The product gives white precipitate (15.719 g) with yield of 81%.

# General procedure for the synthesis of amide analogues of ferulic acid (2a-2g)

Add 1.2 equivalent Thionyl chloride (86.34  $\mu$ L, 1.2mmol) to the synthesized (*E*)-3-(4-acetoxy-3-methoxyphenyl) acrylic acid (236 mg, 1mmol) and heat the reaction mixture at 80<sup>o</sup>C for 3 hrs. Remove the free SOCl<sub>2</sub> by using vacuum pump and dissolved in dry CH<sub>2</sub>Cl<sub>2</sub>.

Amine (1.5 mmol) was dissolved in dry  $CH_2Cl_2$  followed by adding Trimethylamine (721 µL, 5 mmol) and cool the solution below  $10^{0}$ C. To this, acid chloride solution was added dropwise by using addition funnel. Maintain the temperature of reaction mixture below  $10^{0}$ C for 10min and 3hrs in room temperature. The reaction mixture was quenched with water and extracted by using  $CH_2Cl_2$  (2 x 20 mL). The organic solution was dried over anhydrous sodium sulfate and solvents were evaporated using roto vacuum evaporator. The products further purified by subjected to column chromatography using chloroform/methanol (99:1) as eluent.

# Esterification and Thio-esterification of ferulic acid (2h-2k)

Ferulic acid chlorides have been prepared as mentioned in the above procedure (SOCl<sub>2</sub> method) and dissolved this acid chloride in dry CH<sub>2</sub>Cl<sub>2</sub>. Thiol (1.5 mmol) was dissolved in dry CH<sub>2</sub>Cl<sub>2</sub> followed by adding Trimethylamine (721  $\mu$ L, 5 mmol) and cool the solution below 10<sup>0</sup>C. To this, acid chloride solution was added dropwise by using addition funnel. Maintain the temperature of reaction mixture below 10<sup>0</sup>C for 10min and 3hrs in room temperature. The reaction mixture was quenched with water and extracted by using CH<sub>2</sub>Cl<sub>2</sub> (2 x 20 mL). The organic solution was dried over anhydrous sodium sulfate and solvents were evaporated using roto vacuum evaporator. The products further purified by subjected to column chromatography using Hexane - ethyl acetate (80:20) as eluent. The product formed as a white precipitate.

# **Procedure for the synthesis of methyl-** (*E*)-**3-**(**4-hydroxy-3-methoxyphenyl**) acrylate (**3**)

Ferulic acid (194mg, 1mmol) was dissolved in 3mL of methanol followed by adding drops of Con.  $H_2SO_4$ . Then the mixture was refluxed at 80<sup>o</sup>C for 3hrs. After 3hrs, the mixture was quenched with water and extracted by using ethyl acetate (2 x 20mL). The organic extract was separated, washed with bicarbonate, dried over anhydrous sodium sulfate and solvents were evaporated under reduced pressure. As such of the obtained compound was used for the synthesis of its respective analogues.

# General procedure for O- alkylation of ferulic acid (3a-3l)

Amine (2.5 mmol) was dissolved in dry  $CH_2Cl_2$  and adds  $K_2CO_3$  (552 mg, 4 mmol). Stir and cool the solution below  $10^{0}C$ . To this add chloroacetyl chloride (315 µL, 4 mmol) dissolved in dry  $CH_2Cl_2$  dropwise by using addition funnel. Maintain the temperature below  $10^{0}C$  for 20min and 3hrs for room temperature. The reaction mixture was quenched with cold water and

extracted with ethyl acetate (2 x 20 mL). The ethyl acetate layer was dried over anhydrous sodium sulfate and solvents removed under reduced pressure. The acetamide formed as precipitate and it was subjected to O-alkylation with FA ester.

Take methyl- (*E*)-3-(4-hydroxy-3-methoxyphenyl) acrylate (208 mg, 1mmol) and dissolved acetonitrile (2mL). Add the acetamide (1.2 mmol) and  $K_2CO_3$  (165.6 mg, 1.2mmol) to the reaction mixture. Reflux the reaction mixture at  $120^{0}C$  for 3hrs. After completion of the reaction, it was quenched with water and extracted with CH<sub>2</sub>Cl<sub>2</sub> (2 x 20 mL). The organic layer then dried with anhydrous sodium sulfate and solvents were removed under reduced pressure. Product was purified by column chromatography by using methanol/chloroform (3:97) as eluent.

# General procedure for the synthesis of ferulic acid ethers (3m-3w)

Take methyl- (*E*)-3-(4-hydroxy-3-methoxyphenyl) acrylate (208 mg, 1mmol) and dissolved Dimethyl formamide (2mL). To this add  $K_2CO_3$  (414 mg, 3mmol) and alkyl bromide (1.5mmol) and heat the reaction mixture at 80<sup>o</sup>C for 3hrs. After the reaction completes, it was quenched with water and extracted by using ethyl acetate (3 x 20 mL). The ethyl acetate layer then dried over with anhydrous sodium sulfate and solvents were removed under reduced pressure. Product was purified by column chromatography using Hexane – ethyl acetate (95:5) as eluent.

# General procedure for the synthesis of 3H-benzo[f]chromen-3-one derivatives from Ferulic acid (4a-4c)

Ferulic acid (194mg, 1mmol) was taken in 5mL RB flask and mixed with powdered 2-naphthol (144mg, 1mmol). Add anhydrous  $AlCl_3$  (133mg, 1mmol) to the reaction mixture and mixed thoroughly. Neat reaction was performed at  $120^{\circ}C$  for 3hrs. After completion of the reaction, the reaction mixture was quenched with water and extracted with ethyl acetate. Then the organic layer was dried over with anhydrous sodium sulfate and solvents were removed under reduced pressure. Product was purified by column chromatography using Hexane – ethyl acetate (80:20) as eluent.

# In silico studies

# **Ligand Preparation**

All the 39 compounds including (E)-4-(3-((4-fluorophenyl)thio)-3-oxoprop-1-en-1-yl)-2methoxyphenyl acetate (**2i**), methyl (E)-3-(3-methoxy-4-(2-oxo-2thiomorpholinoethoxy)phenyl)acrylate (**3j**) along with ferulic acid (**1**) were drawn by using ChemSketch and known BCR-ABL tyrosine kinase inhibitors (Imatinib) was retrieved from Pubchem database. The ligands were subjected to Energy minimization using Avagadro and subsequently the charges were added to them through Autodock 4.0.

# **Protein preparation**

The crystal structure of the target protein BCR-ABL tyrosine kinase<sup>1</sup> (PDB ID: - 3OXZ) was retrieved from PDB database (<u>http://www.rcsb.org/pdb</u>) having resolution 2.2 Å. All the heteroatoms and water molecules were removed by Discovery Analyzer software. Polar hydrogens and Gasteiger charges were assigned after merging of non-polar hydrogen atoms in ADT.

# Molecular docking

To validate drug-target association, the molecular docking stimulation was performed on ligands with BCR-ABL tyrosine kinase by Autodock software 4.0 by employing Lamarckian genetic algorithm<sup>2</sup>. The receptor was kept rigid, while ligands were set flexible to rotate and explore most probable binding poses. The torsional bonds of ligands were set free by Ligand module in AutoDock Tool (ADT). As per earlier reports, the Grid was set by covering active site binding pocket of the protein (chain A) with  $92 \times 92 \times 92$  points in x, y, z direction and followed by Autogrid run. The docking protocol was then implemented on the ligands in the dataset. The study was carried for 20 GA runs, which was found to be optimum to reproduce the pose in its crystal. The other GA parameters like the population size and the genetic operators were kept at their default values. The results were interpreted by observing the least binding energy (kcal/mol) and hydrogen bonds; stacking interactions involving between the active site residues of the target and ligand molecules were analyzed using PyMol molecular visualization tool and LigPlot. The RMSD was calculated to ensure that the selected ligands have conformational similarity.

# Anti-leukemic activity of FA derivatives

# In vitro cell viability assay

All cell lines were procured from the cell repository of the National Centre for Cell Science, Pune, India. The dose and duration dependent cytotoxicity of Ferulic acid and its derivatives on PBML, K562, U937 and HepG2 cell lines were quantitatively estimated by a non-radioactive, colorimetric assay system using tetrazolium salt, 3-[4,5- dimethylthiazol- 2-yl]-2,5-diphenil-tetrazolium bromide (MTT). The percentage of proliferation was calculated by using the following equation:

% Proliferation = [OD sample - OD control] X 100/OD control

The concentration required for a 50% inhibition of viability (IC50) was determined graphically. Multiple linear regressions were used to compare data using GraphPad prism 6 software. Among all the derivatives of FA, 2i and 3j were found as most active compounds and subjected into further biochemical studies.

# Intracellular measurement of reactive oxygen species (ROS)

The production of intracellular ROS was measured using 2,7- dichlorofluorescein diacetate (DCFH2-DA) [43]. To the stock solution (in methanol) of 10 mM DCFH2-DAwas diluted in

culture medium without serum or other additives to yield a 100  $\mu$ M working solution. At the end of exposure with Ferulic acid, 2i and 3j and the cells were washed twice with PBS. Then, cells were incubated in 1.5 mL working solution of DCFH2-DA at 37°C for 30 min. Cells were lysed in alkaline solution and centrifuged at 3000 rpm. One milliliter of supernatant was transferred to a cuvette, and fluorescence was measured at 520 nm with a fluorescence spectrophotometer (Jasco-FP8200) using 485-nm excitation. As a positive control, those cells were incubated with H<sub>2</sub>O<sub>2</sub> (100 mM) for 30 min prior to the analysis. The values were expressed as percent fluorescence microscopy (Carl Zeiss Axio Scope A1). To determine the crucial cues of ROS generation in shikonin derivatives-induced cell death, K562 cells (2×104) were seeded in a 96-well plate with 0.2 mL per well. A stock solution of *N*-acetyl-L-cysteine (NAC) was made with sterile water and added to cells at 10 mM for 1 h [42]. NAC pretreated or untreated K562 cells were cultured with 100 nM FA derivatives for 24 h and cell viability was determined by the MTT method.

# **Determination of reduced glutathione (GSH)**

Reduced glutathione estimation in the cell lysate was performed by the method of Moron et al. The required amount of the cell lysate was mixed with 25% of trichloroacetic acid and centrifuged at 2,000 xg for 15 min to settle the precipitated proteins. The supernatant was aspirated and diluted to 1 mL with 0.2 M sodium phosphate buffer (pH 8.0). Later, 2 mL of 0.6 mM DTNB was added. After 10 minutes the optical density of the yellow-colored complex formed by the reaction of GSH and DTNB (Ellman's reagent) was measured at 405 nm. A standard curve was obtained with standard reduced glutathione. The levels of GSH were expressed as  $\mu$ g of GSH/mg protein.

# Determination of oxidized glutathione (GSSG)

The oxidized glutathione level was measured after derivatization of GSH with 2-vinylpyidine according to the method of Griffith et al. In brief, with 0.5 mL cell lysate, 2  $\mu$ l of 2-vinylpyidine was added and incubates for 1 hr at 37°C. Then the mixture was deprotenized with 4% sulfosalicylic acid and centrifuged at 1,000 xg for 10 min to settle the precipitated proteins. The supernatant was aspirated and GSSG level was estimated with the reaction of DTNB at 412 nm in spectrophotometer and calculated with standard GSSG curve.

#### **Redox ratio (GSH/GSSG)**

Redox ratio was determined for FA, 2i and 3j by taking the ratio of reduced glutathione/oxidized glutathione.

#### **Determination of lipid peroxidation (MDA)**

Lipid peroxidation was estimated by the method of Ohkawa et al. in cell lysate. The reaction mixture contains Tris-HCl buffer (50 mM, pH 7.4), tert-butyl hydroperoxide (BHP) (500  $\mu$ M in

ethanol) and 1 mM FeSO4. After incubating the samples at 37°C for 90 min, the reaction was stopped by adding 0.2 mL of 8% sodium dodecyl sulfate (SDS) followed by 1.5 mL of 20% acetic acid (pH 3.5). The amount of malondialdehyde (MDA) formed during incubation was estimated by adding 1.5 mL of 0.8% thiobarbituric acid (TBA) and further heating the mixture at 95°C for 45 min. After cooling, samples were centrifuged, and the thiobarbituric acid reactive substances (TBARS) were measured in supernatants at 532 nm by using 1.53 x 105 M-1 cm-1 as extinction coefficient. The levels of lipid peroxidation were expressed in terms of n mol/mg protein.

# Quantitative estimation of DNA fragmentation by diphenylamine (DPA) assay

After the treatment schedule, cells were lysed with hypotonic lysis buffer followed by centrifugation. Then Perchloric acid (0.5 M) was added to the pellets containing uncut DNA (resuspended with 200 mL of hypotonic lysis buffer), and to the other half of the supernatants containing DNA fragments. Then 2 volumes of a solution containing 0.088 M DPA, 98% (v/v) glacial acetic acid, 1.5% (v/v) sulfuric acid, and 0.5% (v/v) of 1.6% acetaldehyde solution were added. The samples were stored at 4 °C for 48 h. The colorimetric reaction was quantitated spectrophotometrically at 575 nm. The percentage of fragmentation was calculated as the ratio of DNA in the supernatants to the total DNA confirm the data obtained from the spectrophotometric method, we have analyzed the DNA fragmentation by DNA laddering in 1.2% agarose gel electrophoresis.

# Apoptotic morphological changes by AO/Et-Br staining

Two DNA-binding dyes AO and Et-Br were used for the morphological apoptotic cells. After treatment with FA derivatives for 24 h, K562 cells were collected, washed with cold PBS and then stained with a mixture of AO (100  $\mu$ g/mL) and Et-Br (100  $\mu$ g/mL) at room temperature for 5 min in dark. After proper washing with PBS, the stained cells were observed by a fluorescence microscope (Carl Zeiss Axio Scope A1). Untreated and FA derivatives treated K562 cells were collected and were fixed with 2.5% glutaraldehyde for 15 min, followed by permeabilized with 0.1% Triton X-100 and stained with 1  $\mu$ g/mL DAPI for 5 min at 37°C. The cells were then washed with PBS and examined by fluorescence microscopy (Carl Zeiss Axio Scope A1).

#### Bcr-Abl kinase inhibitory assay - ADP-Glo<sup>TM</sup> Kinase assay kit

The Bcr–Abl inhibitory activity assay was performed using ADP-Glo<sup>TM</sup> Kinase assay kit (Promega, USA) and Abl Kinase Enzyme System (Promega, USA) according to the manufacturer's instructions. General procedures are as the following: Kinases (4ng/µL) were incubated with substrates ( $0.2\mu g/\mu L$ ), compounds ( $3 \times 10^{-5} - 3 \times 10^{-10}$ M) and ATP ( $25\mu$ M) in a final buffer of tris 40mM, MgCl2 10mM, BSA 0.1mg/mL, DTT 1mM in 384-well plate with the total volume of 5µL. The assay plate was incubated at 30°C for 1h. After the plate cooled for 5min at room temperature, 5µl of ADP-Glo reagent was added into each well to stop the reaction and consume the remaining ADP within 40min. At the end, 10µL of kinase detection reagent was

added into the well and incubated for 30min to produce a luminescence signal and the luminescence was measured with the help of a plate-reading luminometer. The signal was correlated with the amount of ATP present in the reaction and was inversely correlated with the kinase activity.

## Assessment of apoptotic cell population by annexin V-FITC/PI assay

Confluent culture of K562 cells  $(2 \times 10^6)$  were seeded in 6 well cell culture plates and treated with FA, 2i, 3j for 24 h. Then the K562 cells were collected, washed with PBS, and re-suspended in PBS. Apoptotic cell death was identified by double staining with annexin V-FITC and PI, using the annexin V-FITC apoptosis detection kit (BD Bioscience) according to the manufacturer's instructions. Data acquisition and analysis were performed in a Becton-Dickinson FACS verse flow cytometer using CellQuest software.

# BCR-Abl mRNA expression study:

Total RNA extracted from untreated and treated K562 cells after 6 hrs (TRIZOL; Invitrogen). 1 µg of total RNA were reverse transcribed using Revert Aid M-MuLV Reverse Transcriptase (Fermentas) to synthesis the cDNA. cDNA was amplified with green TaqDNA polymerase (Fermentas) in 25 µL reaction mixture using specific primers of bcr-abl and gapdh using Veriti Thermal Cycler (Applied Biosystems). Sequences of the PCR primers were as follows: bcr-abl product representing the BCR-ABL junction and kinase, forward: 5'-GAA GCT TCT CCC TGG CAT CCC GT-3' and reverse 5'-GCC AGG CTC TCG GGT GCA GTC C-3'; gapdh, forward 5'-GAG CCA AAC GGG TCA TCA TC-3' and reverse, 5'-CCT GCT TCA CCA CCT TCT TG-3'. The reaction conditions consisted of an initial activation step (5 min at 95°C) and a cycling step (denaturation for 30 s at 94°C, annealing for 30 s at 58°C and extension for 1 min at 72°C for 35 cycles). Glyceraldehyde-3-phosphate dehydrogenase (GAPDH) amplification was also done to ensure equal cDNA input. PCR-amplified product was subsequently run on 1.2% agarose gel, stained with ethidium bromide and visualized under ultraviolet light and analyzed in BioRad gel documentation system. In real time PCR, (Eppendorf RealPlex master cycler, using 2X SYBR premix Ex Taq II (TAKARA Bio), the mRNA expression levels of the target genes were normalized against those of GAPDH levels and expressed as relative fold change compared with untreated controls and quantified by the  $2^{-ddCT}$  method.





Fig. S1 High Performance Thin Layer Chromatography (HPTLC) profile of Standard Ferulic acid (Lane 1) compared with aqueous extract of *Salicornia brachiata* (Lane 2 and 3).



Fig. S2 Compounds synthesized by using –OH Protected ester of ferulic acid



Fig. S3 Compounds synthesized by using Methyl ferulate



Fig. S4 Crvstal structure of the product (a) Structure of 4a, (b) Crystal structure of 4a [CCDC 1449366], (c) Structure of 3g and (d) Crystal structure of 3g [CCDC 1454627].







Fig. S5 LC Chromatogram and mass spectrum of compound 2h



MassPeaks:495 BasePeak:497(5638) Spectrum Mode:Averaged 1.533-2.267(81-137) B3 Mode:Averaged 4.333-8.633(261-5/9) Polanty:Positive Segment 1 - Event 1



Fig. S6 LC Chromatogram and mass spectrum of compound 2i





Fig. S7 LC Chromatogram and mass spectrum of compound 2j



Fig. S8 LC Chromatogram and mass spectrum of compound 3j



Fig S9. DNA Fragmentation studies of FA, 2i, 3j and imatinib

# **Characterization of the compounds**

# (E)-3-(4-hydroxy-3-methoxyphenyl) acrylic acid (1)

Mp: 169-171°C; MF:  $C_{10}H_{10}O_4$ ; Appearance: white solid; <sup>1</sup>H NMR (300 MHz, d<sub>6</sub>-DMSO): 12.13 (s, 1H), 9.56 (s, 1H), 7.48 (d, J=15.9Hz, 1H), 7.28 (d, J=1.8Hz, 1H), 7.08 (dd, J=1.8Hz, 6.3Hz, 1H), 6.78 (d, J=8.1Hz, 1H), 6.36 (d, J=15.9Hz, 1H), 3.81 (s, 3H); <sup>13</sup>C NMR (300 MHz, d<sub>6</sub>-DMSO): 167.98, 149.02, 147.86, 144.49, 125.73, 122.79, 115.57, 115.47, 111.06, 55.61; LC MS (m/z): [M + H<sup>+</sup>]- 194.19

# Methyl (E)-3-(4-acetoxy-3-methoxyphenyl) acrylate (2)

Yield : 89%; Mp: 197-199°C (Reference  $197^{\circ}$ C); MF: C<sub>13</sub>H<sub>14</sub>O<sub>5</sub>; Appearance: white solid; <sup>1</sup>H NMR (300 MHz, d<sub>6</sub>-DMSO): 12.46 (s, 1H), 7.57 (d, J=15.9Hz, 1H), 7.48 (d, J=1.8Hz, 1H), 7.26 (dd, J=1.8Hz, 6.6Hz, 1H), 7.11 (d, J=8.1Hz, 1H), 6.59 (d, J=15.9Hz, 1H), 3.82 (s, 3H), 2.264 (s, 3H); ; LC MS (m/z): [M + H<sup>+</sup>] - 250.25

# (E)-3-(4-hydroxy-3-methoxyphenyl)-N, N-dimethyl acrylamide (2a)

Yield : 45%; Mp: 116-118°C; MF:  $C_{12}H_{15}NO_3$ ; Appearance: white solid; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): 7.60 (d, J=15.3Hz, 1H), 7.10 (dd, J=1.8Hz, 6.3Hz, 1H), 6.99 (d, J=1.8Hz, 1H), 6.91 (d, J=8.1Hz, 1H), 6.73 (d, J=15.6Hz, 1H), 5.91 (s, 1H), 3.93(s, 3H), 3.17(s, 3H), 3.07(s, 3H); <sup>13</sup>C NMR (300 MHz, d<sub>6</sub>-DMSO): 167.04, 147.35, 146.74, 142.60, 127.87, 121.81, 114.78, 110.05, 55.98, 37.45, 35.99; ; LC MS (m/z): [M + H<sup>+</sup>] - 221.26

#### (E)-4-(3-(diethylamino)-3-oxoprop-1-en-1-yl)-2-methoxyphenyl acetate (2b)

Yield : 53%; Mp: 110-112°C; MF:  $C_{16}H_{21}NO_4$ ; Appearance: white solid; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): 7.66 (d, J=15.3Hz, 1H), 7.14 (d, J=8.1Hz, 1H), 7.05 (d, J=3.9Hz, 1H), 7.02 (s, 1H), 6.75 (d, J=15.3Hz, 1H), 3.8 (s, 3H), 3.53-3.44 (m, 4H), 2.32 (s, 3H), 1.28-1.17 (m, 6H); <sup>13</sup>C NMR (300 MHz, d<sub>6</sub>-DMSO): 168.88, 165.49, 151.21, 141.61, 140.64, 134.56, 123.06, 120.05,118.15, 111.79, 55.91, 42.28,41.06, 20.63, 15.05, 13.18; ; LC MS (m/z): [M + H<sup>+</sup>]- 291.15

#### (E)-3-(4-hydroxy-3-methoxyphenyl)-1-(pyrrolidin-1-yl) prop-2-en-1-one (2c)

Yield : 45%; Mp: 174-175°C; MF:  $C_{14}H_{17}NO_3$ ; Appearance: white solid; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): 7.66 (d, J=15.3Hz, 1H), 7.11 (dd, J=1.8Hz, 6.6Hz,1H), 7.0 (d, J=1.8Hz, 1H), 6.91 (d, J=8.1Hz, 1H), 6.57 (d, J=15.6Hz, 1H), 5.88 (s, 1H), 3.93 (s, 3H), 3.65-3.57 (m, 4H), 2.05-1.85 (m, 4H); <sup>13</sup>C NMR (300 MHz, CDCl<sub>3</sub>)165.06, 147.41, 146.74, 141.91, 127.80, 121.82, 116.19, 114.78, 110.11, 55.94, 46.58, 46.04, 26.11, 24.34; LC MS (m/z): [M + H<sup>+</sup>] - 247.12

#### (E)-2-methoxy-4-(3-oxo-3-(piperidin-1-yl) prop-1-en-1-yl) phenyl acetate (2d)

Yield : 40%; Mp: 124-125°C; MF:  $C_{17}H_{21}NO_4$ ; Appearance: white solid; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): 7.52 (d, J=15.6Hz, 1H), 7.05 (d, J=8.4Hz,1H), 6.97 (t, J=8.1Hz,8.4Hz, 2H), 6.76 (d, J=15.3Hz, 1H), 3.79 (s, 3H), 3.59 (s, 2H), 3.51 (s, 2H), 2.25 (s, 3H), 2.1 (s, 1H), 1.63-1.54 (m, 5H); <sup>13</sup>C NMR (300 MHz, CDCl<sub>3</sub>) 168.88, 165.16, 151.26, 141.49, 140.67, 134.56, 123.08,

120.23, 118.08, 111.53, 55.96, 47.06, 43.36, 26.78, 25.6, 24.63, 20.66;; LC MS (m/z):  $[M + H^+]$ - 303.15

## (E)-3-(4-hydroxy-3-methoxyphenyl)-1-morpholinoprop-2-en-1-one (2e)

Yield : 30%; Mp: 116-118°C; MF:  $C_{14}H_{17}NO_4$ ; Appearance: white solid; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): 7.56 (d, J=15.3Hz, 1H), 7.04-7,01 (m,1H), 6.91 (d, J=1.8Hz,1H), 6.84 (d, J=8.4Hz, 1H), 6.61 (d, J=15.3Hz,1H), 5.90 (s, 1H), 3.86 (d, J=3.6Hz, 3H), 3.65 (s, 8H); <sup>13</sup>C NMR (300 MHz, CDCl<sub>3</sub>) 165.93, 147.58, 146.81, 143.52, 127.60, 121.99, 114.85, 113.79, 109.97, 66.88, 55.99, 46.26, 42.46, 29.69; ; LC MS (m/z): [M + H<sup>+</sup>] - 263.12

#### (E)-4-(3-(benzylamino)-3-oxoprop-1-en-1-yl)-2-methoxyphenyl acetate (2f)

Yield : 35%; Mp: 126-129°C; MF:  $C_{19}H_{19}NO_4$ ; Appearance: white solid; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): 7.55 (d, J=15.6Hz, 1H), 7.30-7.18 (m, 6H), 7.03-6.33(m, 3H), 6.27 (d, J=15.6Hz, 1H), 5.88 (s, 1H), 4.50 (d, J=5.7Hz, 1H), 3.7 (s, 3H), 2.2 (s, 3H); <sup>13</sup>C NMR (300 MHz, CDCl<sub>3</sub>) 168.95, 165.59, 151.27, 140.89, 140.64, 138.12, 133.84, 128.76, 127.94, 127.85, 127.61, 123.15, 120.76, 120.56, 111.40, 55.87, 43.89, 20.66; ; LC MS (m/z): [M + H<sup>+</sup>] - 325.13

# (E)-2-methoxy-4-(3-oxo-3-(phenylthio) prop-1-en-1-yl) phenyl acetate (2h)

Yield : 65%; Mp: 119-121°C; MF:  $C_{18}H_{16}O_4S$ ; Appearance: white solid; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): 7.63 (d, J=15.9Hz, 1H), 7.51-7.43 (m, 5H), 7.15 (td, J=1.8Hz, 2H), 7.07 (d, J=8.1Hz, 1H), 6.73 (d, J=15.6Hz, 1H), 3.88 (s, 3H), 2.33 (s, 3H); <sup>13</sup>C NMR (300 MHz, CDCl<sub>3</sub>) 187.76, 168.70, 151.50, 141.83, 140.77, 134.62, 132.95, 129.22, 127.52, 124.30, 123.38, 121.69, 111.59, 55.97, 20.64; ; LC MS (m/z): [M + H<sup>+</sup>] - 328.08.

#### (E)-4-(3-((4-fluorophenyl) thio)-3-oxoprop-1-en-1-yl)-2-methoxyphenyl acetate (2i)

Yield : 70%; Mp: 108-110°C; MF:  $C_{18}H_{15}FO_4S$ ; Appearance: white solid; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): 7.63 (d, J=15.6Hz, 1H), 7.48-7.43 (m, 2H), 7.19-7.11 (m, 4H), 7.08 (d, J=8.4Hz, 1H), 6.72 (d, J=15.9Hz, 1H), 3.89 (s, 3H), 2.33 (s, 3H); <sup>13</sup>C NMR (300 MHz, CDCl<sub>3</sub>) 187.76, 168.73, 151.53, 141.07, 136.78, 136.66, 132.86, 124.04, 123.43, 122.80, 122.75, 121.75, 116.68, 116.38, 111.59, 55.99, 20.66; ; LC MS (m/z): [M + H<sup>+</sup>] - 346.07

#### (E)-2-methoxy-4-(3-oxo-3-(p-tolylthio) prop-1-en-1-yl) phenyl acetate (2j)

Yield : 40%; Mp: 153-155°C; MF:  $C_{19}H_{18}O_4S$ ; Appearance: white solid; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): 7.64 (d, J=15.9Hz, 1H), 7.61 (d, J=1.8Hz, 1H), 7.39-7.36 (m, 3H), 7.30 (d, J=8.4Hz, 2H), 7.21 (s, 1H), 7.16 (t, J=9.3,1H), 3.84 (s, 3H), 2.36 (s, 3H), 2.27 (s, 3H); <sup>13</sup>C NMR (300 MHz, CDCl<sub>3</sub>) 187.32, 168.32, 151.20, 141.39, 140.73, 139.40, 134.46, 132.67, 129.96, 124.48, 123.70, 123.30, 122.35, 112.27, 56.04, 20.81, 20.37; ; LC MS (m/z): [M + H<sup>+</sup>] - 342.09

# Phenyl- (E)-3-(4-acetoxy-3-methoxyphenyl) acrylate (2k)

Yield : 57%; Mp: 162-163°C; MF:  $C_{18}H_{16}O_5$ ; Appearance: white solid; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): 7.83 (d, J=16.2Hz, 1H), 7.44-7.38 (m, 2H), 7.287-7.16 (m, 5H), 7.09 (d, J=8.1, 1H), 6.58 (d, J=16.2Hz, 1H), 3.84 (s, 3H), 2.54 (s, 3H); <sup>13</sup>C NMR (300 MHz, CDCl<sub>3</sub>) 168.72, 165.21, 151.45, 150.72, 145.77, 141.73, 133.08, 129.43, 125.80, 123.23, 121.57, 121.45, 117.49, 111.38, 55.90, 20.62; ; LC MS (m/z): [M + H<sup>+</sup>] - 312.10

# Methyl (E)-3-(4-hydroxy-3-methoxyphenyl) acrylate (3)

Yield: 88%; MF:  $C_{11}H_{12}O_4$ ; Appearance: Brown viscus liquid; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): 7.62 (d, J=15.9Hz, 1H), 7.04 (td, J=1.8Hz, 2H), 6.91 (d, J=8.1Hz, 1H), 6.29 (d, J=15.9Hz, 1H), 3.91 (s, 3H), 3.79 (s, 3H); ; LC MS (m/z): [M + H<sup>+</sup>] - 208.07

#### Methyl (E)-3-(3-methoxy-4-(2-oxo-2-(phenylamino)ethoxy)phenyl)acrylate (3a)

Yield: 51%; Mp: 148-150°C; MF:  $C_{19}H_{19}NO_5$ ; Appearance: White solid; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): 8.76 (s, 1H), 7.66-7.57 (m, 3H), 7.38-7.33 (m, 2H), 7.18-7.10 (m, 3H), 6.96 (d, J=8.1Hz, 1H), 6.35 (d, J=15.9Hz, 1H), 4.67 (s, 2H), 3.97 (s, 3H), 3.81 (s, 3H); <sup>13</sup>C NMR (300 MHz, CDCl<sub>3</sub>) 167.30. 166.18, 149.84, 148.79, 144.05, 137.00, 129.76, 129.03, 124.69, 122.22, 119.86, 116.88, 115.60, 110.57, 69.68, 55.91, 51.66; ; LC MS (m/z): [M + H<sup>+</sup>] - 341.13

#### Methyl (E)-3-(4-(2-((4-chlorophenyl) amino)-2-oxoethoxy)-3-methoxyphenyl)acrylate (3b)

Yield: 48%; Mp: 188-190°C; MF:  $C_{19}H_{18}CINO_5$ ; Appearance: White solid; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): 8.80 (s, 1H), 7.63 (d, J=15.9Hz, 1H), 7.58-7.53 (m, 2H), 7.34-7.28 (m, 2H), 7.15-7.10 (m, 2H), 6.96 (d, J=8.1Hz, 1H), 6.36 (d, J=15.9Hz, 1H), 4.66 (s, 2H), 3.97 (s, 3H), 3.81 (s, 3H); <sup>13</sup>C NMR (300 MHz, CDCl<sub>3</sub>) 167.33. 166.28, 149.89, 148.72, 144.02, 135.64, 130.00, 129.74, 124.12, 122.29, 121.08, 117.08, 115.84, 110.68, 69.77, 56.01, 51.74; ; LC MS (m/z): [M + H<sup>+</sup>] - 375.09

# Methyl (E)-3-(3-methoxy-4-(2-((4-nitrophenyl)amino)-2-oxoethoxy)phenyl)acrylate (3d)

Yield: 53%; Mp: 178-180°C; MF:  $C_{19}H_{18}N_2O_7$ ; Appearance: pale yellow solid; <sup>1</sup>H NMR (300 MHz, d<sub>6</sub>-DMSO): 10.66 (s, 1H), 8.65 (t, J=2.1Hz, 1H), 7.98-7.93 (m, 2H), 7.66-7.57 (m, 2H), 7.42 (d, J=1.8Hz, 1H), 7.26 (dd, J=1.8Hz, 6.6Hz, 1H), 6.96 (d, J=8.4Hz, 1H), 6.61 (d, J=15.9Hz, 1H), 4.81 (s, 2H), 3.81 (s, 3H), 3.71 (s, 3H); <sup>13</sup>C NMR (300 MHz, d<sub>6</sub>-DMSO) 167.16. 166.87, 149.37, 149.23, 147.92, 144.45, 139.49, 130.24, 127.96, 125.43, 122.48, 118.17, 115.88, 113.62, 113.57, 111.15, 67.82, 55.94, 51.30; ; LC MS (m/z): [M + H<sup>+</sup>] - 386.11

# Methyl (E)-3-(3-methoxy-4-(2-((3-methoxyphenyl) amino)-2-oxoethoxy) phenyl)acrylate (3e)

Yield: 40%; Mp: 168-170°C; MF:  $C_{20}H_{21}NO_6$ ; Appearance: White solid; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): 8.75 (s, 1H), 7.61 (d, J=15.9Hz, 1H), 7.34 (t, J=2.1Hz, 2.4Hz, 1H), 7.27-7.22 (m, 1H), 7.14 (d, J=1.8Hz, 2H), 7.11-7.04 (m, 1H), 6.93 (d, J=8.4Hz, 1H), 6.72-6.68 (m, 1H), 6.36 (d, J=15.9Hz, 1H), 4.66 (s, 2H), 3.97 (s, 3H), 3.81 (d, J=3.6, 6H); <sup>13</sup>C NMR (300 MHz, CDCl<sub>3</sub>)

167.37. 166.26, 160.21, 149.93, 148.86, 144.11, 138.28, 129.89, 129.78, 122.29, 116.99, 115.78, 112.07, 110.43, 105.79, 69.83, 56.01, 55.34, 51.74, 29.70; ; LC MS (m/z): [M + H<sup>+</sup>]- 371.14

# Methyl (E)-3-(3-methoxy-4-(2-oxo-2-(o-tolylamino)ethoxy) phenyl)acrylate (3f)

Yield: 42%; Mp: 157-160°C; MF:  $C_{20}H_{21}NO_5$ ; Appearance: Grey solid; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): 10.03 (s, 1H), 7.62 (d, J=15.9Hz, 1H), 7.45-7.37 (m, 3H), 7.26-7.17 (m, 2H), 6.94 (d, J=8.4Hz, 1H), 6.89 (d, J=7.8Hz, 1H), 6.52 (d, J=15.9Hz, 1H), 4.74 (s, 2H), 3.85 (s, 3H), 3.71 (s, 3H), 2.27 (s, 3H); <sup>13</sup>C NMR (300 MHz, CDCl<sub>3</sub>) 166.88. 166.13, 149.51, 144.48, 138.25, 137.98, 128.60, 127.82, 124.33, 122.50, 119.88, 116.55, 115.80, 113.48, 110.11, 67.93, 55.74, 51.28, 21.12; ; LC MS (m/z):  $[M + H^+]$  - 355.19

# Methyl (E)-3-(4-(2-(benzylamino)-2-oxoethoxy)-3-methoxyphenyl) acrylate (3g)

Yield: 30%; Mp: 160-162°C; MF:  $C_{20}H_{21}NO_5$ ; Appearance: White solid; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): 8.50 (t, J=6Hz, 5.7Hz, 1H), 7.60 (d, J=15.9Hz, 1H), 7.40 (, 1H), 7.33-7.22 (m, 6H), 6.93 (d, J=8.4Hz, 1H), 6.60 (d, J=15.9Hz, 1H), 4.613 (s, 2H), 4.33 (d, J=6Hz, 2H), 3.817 (s, 3H), 3.717 (s, 3H); ; LC MS (m/z): [M + H<sup>+</sup>] - 355.14

# Methyl (E)-3-(4-(2-(diphenylamino)-2-oxoethoxy)-3-methoxyphenyl) acrylate (3h)

Yield: 50%; Mp: 166-167°C; MF:  $C_{25}H_{23}NO_5$ ; Appearance: White solid; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): 7.61 (d, J=15.9Hz, 1H), 7.38-7.26 (m, 10H), 7.06(t, J=1.8Hz, 8.4Hz, 2H), 6.78 (d, J=7.8Hz, 1H), 6.31 (d, J=15.9Hz, 1H), 4.66 (s, 2H), 3.86 (s, 3H), 3.79 (s, 3H); <sup>3</sup>C NMR (300 MHz, CDCl<sub>3</sub>) 167.62. 167.03, 149.64, 149.58, 144.66, 129.64, 128.42, 122.14, 115.90, 113.44, 110.56, 67.54, 55.93, 51.64; LC MS (m/z):  $[M + H^+]$  417.16

#### Methyl (E)-3-(3-methoxy-4-(2-morpholino-2-oxoethoxy) phenyl) acrylate (3i)

Yield: 45%; Mp: 110-120°C; MF:  $C_{17}H_{21}NO_6$ ; Appearance: White solid; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): 7.62 (d, J=16.2Hz, 1H), 7.09 (t, J=1.8Hz, 8.1Hz, 2H), 6.93 (d, J=8.1Hz, 1H), 6.32 (d, J=8.1Hz, 1H), 4.79 (s, 2H), 3.90 (s, 3H), 3.80 (s, 3H); 3.64 (d, J=3.9Hz, 8H), <sup>13</sup>C NMR (300 MHz, CDCl<sub>3</sub>) 167.47. 166.10, 149.51, 148.98, 144.39, 128.70, 122.22, 116.21, 113.53, 110.32, 68.42, 66.76, 55.83, 51.63, 45.91, 42.51; ; LC MS (m/z): [M + H<sup>+</sup>] - 335.14

# Methyl (E)-3-(3-methoxy-4-(2-oxo-2-thiomorpholinoethoxy) phenyl) acrylate (3j)

Yield: 52%; Mp: 110-112°C; MF:  $C_{17}H_{21}SNO_5$ ; Appearance: White solid; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): 7.62 (d, J=15.9Hz, 1H), 7.09 (t, J=1.8Hz, 7.8Hz, 2H), 6.93 (d, J=8.1Hz, 1H), 6.32 (d, J=15.9Hz, 1H), 4.77 (s, 2H), 3.92-3.825 (m, 10H), 2.67-2.58 (m, 4H), <sup>13</sup>C NMR (300 MHz, CDCl<sub>3</sub>) 167.48. 166.10, 149.44, 149.02, 144.40, 128.68, 122.23, 116.21, 113.34, 110.29, 68.59, 55.82, 51.63, 48.34, 44.78, 27.94, 27.34; ; LC MS (m/z): [M + H<sup>+</sup>] - 351.13.

# Methyl (E)-3-(3-methoxy-4-(2-oxo-2-(piperidin-1-yl) ethoxy) phenyl) acrylate (3k)

Yield: 38%; Mp: 170-172°C; MF:  $C_{18}H_{23}NO_5$ ; Appearance: White solid; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): 7.62 (d, J=15.9Hz, 1H), 7.08 (t, J=1.8Hz, 7.5Hz, 2H), 6.93 (d, J=8.4Hz, 1H), 6.31 (d, J=15.9Hz, 1H), 4.78 (s, 2H), 3.92 (s, 3H), 3.80 (s, 3H), 3.54-3.49 (m, 4H), 1.66-1.60 (m, 5H), 1.25 (s, 1H), <sup>13</sup>C NMR (300 MHz, CDCl<sub>3</sub>) 167.56. 165.65, 149.53, 144.60, 128.31, 122.33, 115.91, 113.37, 110.34, 68.29, 55.90, 51.60, 46.33, 43.26, 29.66, 26.41, 25.52, 24.41; ; LC MS (m/z): [M + H<sup>+</sup>] - 333.16

#### Methyl (E)-3-(3-methoxy-4-(2-oxo-2-(pyrrolidin-1-yl) ethoxy)phenyl)acrylate (3l)

Yield: 40%; Mp: 123-127°C; MF:  $C_{17}H_{21}NO_5$ ; Appearance: White solid; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): 7.63 (d, J=15.9Hz, 1H), 7.08 (t, J=2.1Hz, 7.2Hz, 2H), 6.91 (t, J=1.2Hz, 7.5Hz, 1H), 6.30 (d, J=15.9Hz, 1H), 4.72 (s, 2H), 3.90 (s, 3H), 3.80 (s, 3H), 3.56-3.49 (m, 4H), 2.01-1.80 (m, 4H); LC MS (m/z): [M + H<sup>+</sup>] - 319.20

#### Methyl (E)-3-(4-ethoxy-3-methoxyphenyl) acrylate (3m)

Yield: 61%; Mp: 94-96°C; MF:  $C_{13}H_{16}O_4$ ; Appearance: White solid; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): 7.64 (d, J=15.9Hz, 1H), 7.10-7.04 (m, 2H), 6.85 (d, J=8.1Hz, 1H), 6.32 (d, J=15.9Hz, 1H), 4.13 (q, J=7.4Hz, 6.9Hz, 2H), 3.90 (s, 3H), 3.80 (s, 3H), 1.48 (t, J=6.9Hz, 7.2Hz, 3H); <sup>13</sup>C NMR (300 MHz, CDCl<sub>3</sub>) 167.70, 150.52, 149.39, 144.85, 127.18, 122.57, 115.33, 112.13, 109.92, 64.35, 55.92, 51.59, 14.68; ; LC MS (m/z): [M + H<sup>+</sup>] - 236.10

#### Methyl (E)-3-(3-methoxy-4-propoxyphenyl) acrylate (3n)

Yield: 70%; Mp: 116-118°C; MF:  $C_{14}H_{18}O_4$ ; Appearance: White solid; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): 7.63 (d, J=16.2Hz, 1H), 7.101-7.047 (m, 2H), 6.81 (d, J=8.1Hz, 1H), 6.30 (d, J=15.9Hz, 1H), 4.01 (t, J=5.4Hz, 6.9Hz, 2H), 3.89 (s, 3H), 3.80 (s, 3H), 1.94-1.82 (m, 2H),1.045 (t, J=7.5Hz, 7.2Hz, 3H); <sup>13</sup>C NMR (300 MHz, CDCl<sub>3</sub>) 167.67, 150.76, 149.48, 144.84, 127.12, 122.54, 115.26, 112.32, 110.11, 70.41, 55.96, 51.55, 22.33, 10.35; ; LC MS (m/z): [M + H<sup>+</sup>]-250.12

#### Methyl (E)-3-(4-butoxy-3-methoxyphenyl) acrylate (30)

Yield: 52%; Mp: 104-106°C; MF:  $C_{15}H_{20}O_4$ ; Appearance: White solid; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): 7.62 (d, J=15.9Hz, 1H), 7.101-7.045 (m, 2H), 6.81 (d, J=8.1Hz, 1H), 6.31 (d, J=15.9Hz, 1H), 4.05 (t, J=6.9Hz, 2H), 3.89 (s, 3H), 3.80 (s, 3H), 1.89-1.79 (m, 2H), 1.56-1.43 (m, 2H), 1.045 (t, J=7.5Hz, 7.2Hz, 3H); <sup>13</sup>C NMR (300 MHz, CDCl<sub>3</sub>) 167.66, 150.79, 149.47, 144.83, 127.09, 122.53, 115.23, 112.28, 110.09, 68.64, 55.95, 51.53, 31.04,19.13,13.91; ; LC MS (m/z): [M + H<sup>+</sup>] - 264.14

### Methyl (E)-3-(3-methoxy-4-(pentyloxy) phenyl) acrylate (3p)

Yield: 55%; Mp: 82-84°C; MF:  $C_{16}H_{22}O_4$ ; Appearance: White solid; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): 7.63 (d, J=15.9Hz, 1H), 7.101-7.045 (m, 2H), 6.81 (d, J=8.1Hz, 1H), 6.31 (d, J=15.9Hz, 1H), 4.04 (t, J=6.9Hz, 2H), 3.89 (s, 3H), 3.79 (s, 3H), 1.90-1.816 (m, 2H), 1.49-1.32 (m, 4H), 0.98 (t, J=7.2Hz, 7.5Hz, 3H); <sup>13</sup>C NMR (300 MHz, CDCl<sub>3</sub>) 167.62, 150.75, 149.43, 144.80,

127.06, 122.50, 115.20, 112.24, 110.05, 68.91, 55.91, 51.49, 28.17, 27.99, 22.37, 13.91; ; LC MS (m/z):  $[M + H^+]$  - 278.15

#### Methyl (E)-3-(4-(hexyloxy)-3-methoxyphenyl) acrylate (3q)

Yield: 41%; Mp: 74-76°C; MF:  $C_{17}H_{24}O_4$ ; Appearance: White solid; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): 7.62 (d, J=15.9Hz, 1H), 7.10-7.04 (m, 2H), 6.80 (d, J=8.1Hz, 1H), 6.31 (d, J=15.9Hz, 1H), 4.04 (t, J=6.9Hz, 2H), 3.89 (s, 3H), 3.79 (s, 3H), 1.90-1.80 (m, 2H), 1.50-1.41 (m, 2H), 1.36-1.28 (m, 4H), 0.90 (t, J=7.5Hz, 7.2Hz, 3H); <sup>13</sup>C NMR (300 MHz, CDCl<sub>3</sub>) 167.68, 150.78, 149.46, 144.85, 127.09, 122.55, 115.23, 112.28, 110.07, 68.98, 55.95, 51.55, 31.52, 28.96, 25.56, 22.53, 13.97; ; LC MS (m/z): [M + H<sup>+</sup>] - 292.17

#### Methyl (E)-3-(4-(heptyloxy)-3-methoxyphenyl) acrylate (3r)

Yield: 52%; Mp: 66-68°C; MF:  $C_{18}H_{26}O_4$ ; Appearance: White solid; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): 7.63 (d, J=15.9Hz, 1H), 7.07 (dd, J=1.8Hz, 2H), 6.85 (d, J=8.1Hz, 1H), 6.31 (d, J=15.9Hz, 1H), 4.04 (t, J=6.9Hz, 6.6Hz, 2H), 3.89 (s, 3H), 3.79 (s, 3H), 1.90-1.80 (m, 2H), 1.49-1.25 (m, 8H), 0.88 (q, J=2.1Hz, 4.5Hz, 3H); <sup>13</sup>C NMR (300 MHz, CDCl<sub>3</sub>) 167.72, 150.82, 149.50, 144.89, 127.13, 122.58, 115.26, 112.58, 110.10, 69.02, 55.98, 51.59, 31.74, 29.03, 25.89, 22.59, 14.09; ; LC MS (m/z): [M + H<sup>+</sup>] - 306.18

#### Methyl (E)-3-(3-methoxy-4-(octyloxy)phenyl)acrylate (3s)

Yield: 55%; Mp: 68-70°C; MF:  $C_{19}H_{28}O_4$ ; Appearance: White solid; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): 7.63 (d, J=15.9Hz, 1H), 7.09 (d, J=1.8Hz, 1H), 7.07-7.043 (m, 1H), 6.85 (d, J=8.4Hz, 1H), 6.30 (d, J=15.9Hz, 1H), 4.04 (t, J=6.9Hz, 2H), 3.89 (s, 3H), 3.79 (s, 3H), 1.90-1.80 (m, 2H), 1.47-1.28 (m, 10H), 0.90-0.86 (m, 3H); <sup>13</sup>C NMR (300 MHz, CDCl<sub>3</sub>) 167.71, 150.82, 149.50, 144.88, 127.12, 122.58, 115.26, 112.31, 110.10, 69.01, 55.97, 51.58, 31.79, 29.33, 29.20, 29.03, 25.92, 22.64, 14.09; ; LC MS (m/z): [M + H<sup>+</sup>] - 320.30

#### Methyl (E)-3-(4-(allyloxy)-3-methoxyphenyl) acrylate (3t)

Yield: 46%; Mp: 84-86°C; MF:  $C_{14}H_{16}O_4$ ; Appearance: White solid; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): 7.62 (d, J=15.9Hz, 1H), 7.10-7.04 (m, 2H), 6.80 (d, J=8.1Hz, 1H), 6.31 (d, J=15.9Hz, 1H), 5.45-5.44(m, 1H), 5.38-5.29 (m, 1H), 4.66-4.64 (m, 2H), 3.90 (s, 3H), 3.80 (s, 3H); <sup>13</sup>C NMR (300 MHz, CDCl<sub>3</sub>) 167.61, 150.06, 149.49, 144.72, 132.70, 127.50, 122.31, 118.34, 115.49, 112.80, 110.01, 69.68, 55.87, 51.56; ; LC MS (m/z): [M + H<sup>+</sup>] - 248.10

#### Methyl (E)-3-(4-((5-cyanopentyl) oxy)-3-methoxyphenyl) acrylate (3u)

Yield: 78%; Mp: 92-94°C; MF:  $C_{17}H_{21}NO_4$ ; Appearance: White solid; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): 7.62 (d, J=15.9Hz, 1H), 7.10-7.04 (m, 2H), 6.80 (d, J=8.1Hz, 1H), 6.31 (d, J=15.9Hz, 1H), 4.06 (t, J=5.4Hz, 6.9Hz, 2H), 3.90 (s, 3H), 3.80 (s, 3H), 2.42-2.37 (m, 2H), 1.92-1.59 (m, 6H); <sup>13</sup>C NMR (300 MHz, CDCl<sub>3</sub>) 167.64, 150.48, 149.55, 144.75, 127.48, 122.48, 119.60, 115.52, 112.48, 110.16, 68.43, 55.95, 51.60, 31.79, 28.24, 25.36, 25.16, 17.07; ; LC MS (m/z): [M + H<sup>+</sup>]- 303.15

# Methyl (E)-3-(4-(benzyloxy)-3-methoxyphenyl) acrylate (3v)

Yield: 42%; Mp: 76-78°C; MF:  $C_{18}H_{18}O_4$ ; Appearance: White solid; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): 7.61 (d, J=15.9Hz, 1H), 7.44-7.30 (m, 5H), 7.06-7.01 (m, 2H), 6.81 (d, J=8.1Hz, 1H), 6.30 (d, J=15.9Hz, 1H), 5.19 (s, 2H), 3.91 (s, 3H), 3.79 (s, 3H); <sup>13</sup>C NMR (300 MHz, CDCl<sub>3</sub>) 167.66, 150.27, 149.76, 144.76, 136.58, 128.65, 128.28, 128.03, 127.73, 127.21,122.37, 115.62, 113.43, 110.26, 70.86, 66.27, 56.00, 51.52; ; LC MS (m/z):  $[M + H^+]$ - 298.12

#### Methyl (E)-3-(4-(hexadecyloxy)-3-methoxyphenyl) acrylate (3w)

Yield: 57%; Mp: 68-70°C; MF:  $C_{27}H_{44}O_4$ ; Appearance: White solid; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): 7.61 (d, J=15.9Hz, 1H), 7.06-7.04 (m, 2H), 6.81 (d, J=8.1Hz, 1H), 6.30 (d, J=15.9Hz, 1H), 4.04 (t,J= 6.9, 2H), 3.89 (s, 3H), 3.80 (s, 3H), 1.90-1.80 (m, 2H), 1.30 (d, J=13.5Hz, 26H), 0.89 (t, J=6.6Hz, 6.3Hz, 3H); ); <sup>13</sup>C NMR (300 MHz, CDCl<sub>3</sub>) 167.69, 150.81, 149.49, 144.86, 127.11, 122.55, 115.24, 112.30, 110.10, 69.01, 63.06, 55.96, 51.55, 32.80, 31.90, 29.67, 29.63, 29.56, 29.51, 29.42, 29.34, 29.01, 25.89, 25.72, 22.67, 14.09; ; LC MS (m/z):  $[M + H^+]$ - 432.32

# (E)-3-(3-methoxy-4-(((4-nitrophenyl) carbamoyl) oxy) phenyl) acrylic acid (3x)

Yield: 40%; Mp: 172-174°C; MF:  $C_{17}H_{14}N_2O_7$ ; Appearance: pale yellow solid; <sup>1</sup>H NMR (300 MHz, d6-DMSO): 12.4 (s, 1H), 11.03 (s, 1H), 8.25 (d, J=9.3Hz, 2H), 7.73 (d, J=9Hz, 2H), 7.63-7.53 (m, 3H), 7.29 (s, 2H), 6.62 (d, J=16.2Hz, 1H), 3.85 (s, 3H); <sup>13</sup>C NMR (300 MHz, d6-DMSO) 167.98, 155.65, 149.01, 147.86, 144.46, 135.59, 126.36, 125.72, 125.10, 122.76, 118, 117.97, 115.58, 115.47, 112.35, 111.08, 55.63, ; LC MS (m/z): [M + H<sup>+</sup>] - 358.08

#### 1-(4-hydroxy-3-methoxyphenyl)-1,2-dihydro-3H-benzo[f]chromen-3-one (4a)

Yield : 35%; Mp: 160-162; MF:  $C_{20}H_{16}O_4$ ; Appearance: White solid; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): 7.83 (q, J=8.7Hz, 7.2Hz, 3H), 7.46 (pd, J=1.5Hz, 2H), 7.34 (d, J=8.7Hz, 1H), 6.78 (d, J=8.4Hz, 1H), 6.61 (q, J=1.8Hz, 3.9Hz, 2H), 5.53(s, 1H), 4.88(q, J=2.4Hz, 3.6Hz, 1H) 3.75 (s, 3H), 3.15 (t, J=6.6Hz, 3.3Hz, 2H); C=74.11%, H=4.99%, O=19.79%; ; LC MS (m/z):  $[M + H^+]$ -320.10

#### 8-bromo-1-(4-hydroxy-3-methoxyphenyl)-1,2-dihydro-3H-benzo[f]chromen-3-one (4b)

Yield : 61%; Mp: 174-176; MF:  $C_{20}H_{15}BrO_4$ ; Appearance: White solid; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): 7.94 (d, J=1.8Hz, 1H), 7.70 (d, J=8.7Hz, 1H), 7.60 (d, J=9Hz, 1H), 7.47 (dd, J=2.1Hz, 1H), 7.29 (d, J=9Hz, 1H), 6.718 (t, J=4.2Hz,4.5Hz,1H), 6.51-6.47(m, 2H), 5.47(s, 1H), 4.71(q, J=2.7Hz,3.6Hz, 1H), 3.69 (s, 3H), 3.16-3.02 (m, 2H); <sup>13</sup>C NMR (300 MHz, CDCl<sub>3</sub>):166.89, 149.87, 147.08, 145.17, 132.23, 132.11, 130.78, 130.71, 129.59, 129.0, 124.87, 119.8, 119.26, 118.73, 118.21, 114.95, 108.99, 55.86, 37.64, 37.43; C=60.1%, H=3.699%, O=15.99%; ; LC MS (m/z):  $[M + H^+]$ - 398.02

#### 7-hydroxy-4-(4-hydroxy-3-methoxyphenyl) chroman-2-one (4c)

Yield : 30%; Mp: 168-170; MF:  $C_{16}H_{14}O_5$ ; Appearance: White solid; <sup>1</sup>H NMR (300 MHz, d6-DMSO): 9.72 (s, 1H), 8.93 (s, 1H), 6.83 (d, J=8.1Hz, 1H), 6.78(d, J=1.8Hz, 1H), 6.69 (d, J=8.1Hz, 1H), 6.55-6.50 (m, 2H), 6.43 (dd, J=1.8Hz, 1H), 4.24 (t, J=6Hz, 6.3Hz, 1H), 3.71 (s, 3H), 3.02 (d, J=6.3Hz, 2H); <sup>13</sup>C NMR (300 MHz, d6-DMSO): 167.86, 157.37, 151.79, 147.63, 145.45, 132.37, 128.83, 119.27, 116.65, 115.37, 111.55, 111.48, 103.12, 55.50, 36.77, 30.59; C=67.15%, H=5.01%, O=28.02%; ; LC MS (m/z): [M + H<sup>+</sup>] - 286.08















	ers	Hz Hz sec usec K sec sec	usec dB MHz	NHZ Hz
Data Parameters 0852N2 1	uisition Paramet 20151208 12:03 39=01 59=01 59=01 59=01 5533 5533 5533 5533 5533 5533 5533 55	6188.115 0.094423 5.2954423 5.2954423 456 80.480 6.00 6.00 6.00 1.0000000	CHANNEL F1	cessing paramete 32768 300.1300849 200 0,30 0,30 1.00
Current NAME EXPNO PROCNO	F2 - Acq Date_ Time INSTRUM PROBHD PULPROG TD SOLVENT SOLVENT SOLVENT SOLVENT	SWH FIDRES AO RG DW DE DE DI TE D1 TD0	NUC1 P1 PL1 SF01	F2 - Pro S1 SF WMDW S35B LLB CGB PC



































































































































Compound code	IC <sub>50</sub> value against K562 cells (μg/mL)	IC <sub>50</sub> value against U937 cells (μg/mL)	IC <sub>50</sub> value against HepG- 2 cells ( µg/mL)	IC <sub>50</sub> value against Normal PBMC cells (ug/mL)
1	>100	54.68	35.07	76.54
2	>100	32.21	>100	>100
2a	>100	>100	>100	>100
2b	>100	>100	>100	>100
2c	>100	>100	>100	>100
2d	>100	>100	>100	>100
2e	>100	>100	>100	>100
2f	>100	16.89	>100	>100
2h	35	21.11	42.18	68.14
2i	4.02	33.14	32.94	51.32
2j	60	>100	51.01	74.18
2k	>100	>100	>100	>100
3	>100	>100	>100	>100
3a	>100	>100	>100	>100
3b	>100	>100	>100	>100
3d	>100	>100	>100	>100
3e	>100	10.12	22.64	>100
3f	>100	>100	>100	>100
3g	>100	>100	>100	>100
3h	>100	>100	>100	>100
3i	>100	>100	>100	>100
Зј	6.30	40.21	38.54	56.98
3k	>100	>100	>100	>100
31	>100	>100	>100	>100
3m	>100	>100	>100	>100
3n	>100	>100	>100	>100
30	>100	>100	>100	>100
3p	>100	>100	>100	>100
3q	>100	>100	>100	>100
3r	>100	>100	>100	>100
<u>3s</u>	>100	>100	>100	>100
3t	>100	>100	>100	>100
<u>3u</u>	>100	19.23	34.56	79.34
3v	>100	>100	>100	>100
3w	>100	>100	>100	>100
3x	>100	>100	>100	>100
4a	>100	>100	>100	>100
4b	>100	>100	>100	>100
4c	>100	>100	>100	>100
Imatinib	0.1		n.d	-

Table S1.	In vitro	anti-proliferative	activity	of Ferulic	acid o	derivatives
	110 10010	and promotative	~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~	or i er ane	acra (	