

Supporting Information

Identification of new P300 acetyltransferase inhibitors from natural products by a customized virtual screening method

Guo-Bo Li,^a Lu-Yi Huang,^a Hui Li,^a Sen Ji,^a Lin-Li Li,^b and Sheng-Yong Yang^{a,*}

^a State Key Laboratory of Biotherapy/Collaborative Innovation Center for Biotherapy, West China Hospital, West China Medical School, Sichuan University, Sichuan 610041, China

^b Key Laboratory of Drug Targeting and Drug Delivery System of Ministry of Education, West China School of Pharmacy, Sichuan University, Chengdu, Sichuan, 610041, China

* to whom correspondence should be addressed. Tel.: +86-28-85164063; Fax: +86-28-85164060. E-mail: yangsy@scu.edu.cn.

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Experimental Methods.

Customized Virtual Screening Procedures. On the basis of the hypothesis that essential pharmacophore features for potent p300 HAT inhibitor are hydrogen-bonds with residues Arg1410, Thr1411, and Trp1466, we established a target-customized structure-based virtual screening method to screen potential hits from natural product libraries. All the chemical structures of natural products commercially available in Specs Natural Products, AnalytiCon Discovery Natural Products, Indofine Natural Products, and Princeton Natural Products were downloaded from ZINC database because they are well prepared for docking. This first step is to use three rules including (i) molecule weight ≤ 500 , (ii) number of polar atoms ≤ 10 , and (iii) number of rotatable bonds ≤ 10 to filter the collected natural products to exclude the large complicated compounds by an in-house program derived from our previously programs. The second step uses Autodock vina to predict the binding modes and affinities for the filtered compounds from the first step. The crystal structure of p300 HAT in complex with Lys-CoA (PDB entry: 3BIY) was used as the docking template. Gasteiger-Marsili charges were added to the models of protein and compounds, and then non-polar hydrogens were merged onto their respective heavy atoms using AutoDockTools and Raccoon. The grid center was set as the co-crystal ligand Lys-CoA (x, y, z = -17.628, 15.959, 1.475), and the grid size was $26\text{\AA} \times 26\text{\AA} \times 26\text{\AA}$, which encompassed the entire p300 HAT binding site. The other parameters for Vina were set as default. The third step uses hydrogen-bonding

analysis to analyse the docking poses using an in-house program. The hydrogen bonds were defined as the same with that described in ID-Score method¹, and only the compounds likely to form hydrogen-bonds with the three key residues were kept. The final step is manually visualisation to confirm whether or not the predicted binding poses are reasonable. Using this customized virtual screening protocol, we performed a screening campaign against 28,969 natural compounds, and obtained 110 potential hit compounds for p300 HAT (see Supporting Information Table S1).

Enzymatic Inhibitory Assays. Natural compounds **NP-1** to **NP-15** were purchased from Specs company and used without further purification. p300 HAT inhibitory assays were performed using labelled 3H-Ac-CoA and substrate peptide by Shanghai ChemPartner Co., Ltd. The purchased materials include p300 (BPS, Cat. No. 50071), 3H-Ac-CoA (PerkinElmer, Cat. No. NET290), Ac-CoA (Sigma, Cat. No. A2056), C646 (Calbiochem, Cat. No. 382113), and 384-well Flashplate (Perkin Elmer, Cat. No. SMP410A001PK). The assay buffer is modified Tris buffer, and the substrate is H3 peptide. The reaction is initiated by the addition of 10 μ L the substrate solution. After incubation of 60 mins at room temperature, the reaction was stopped by adding 10 μ L stop solution. Volume of 25 μ L reaction solutions are transferred to Flashplate, and incubated for 1 hour, and washed three times using dH₂O and 0.1% Tween-20 prior to reading on Microbeta. The inhibition activity is calculated according to the equation: $\text{Inh\%} = (\text{Max-Signal})/(\text{Max-Min}) \times 100$. All the assays were replicated twice, and the

means of the replicates were calculated. The IC₅₀ values were determined from dose-response curves using Graphpad Prism 5 after carrying out assays at 10 different concentrations for each compound.

The inhibitory activities of **NP-2**, **NP-3**, **NP-9** and **NP-15** against the other three histone acetylases including PCAF, KAT1, and HAT7 were also carried out by ChemPartner Co., Ltd. The purchased materials include PCAF (Cayman, Cat. No. 10009115), HAT1 (Prospec, Cat. No. enz-504), and KAT7 (Active Motif). The screening protocols for PCAF, KAT1, and HAT7 as well as the calculation method for inhibition activity are the same as that described above for p300 HAT assays. All the assays were replicated twice, and the means of the replicates were calculated. The IC₅₀ values of **NP-2**, **NP-3**, **NP-9** and **NP-15** for PCAF were determined from dose-response curves using Graphpad Prism 5 after carrying out assays at 10 different concentrations.

Cell lines. All cell lines used were obtained from American Type Culture Collection (ATCC). These cell lines were cultured in culture medium supplemented with 10% FBS (Caoyuanlvye). All cell lines were maintained at 37 °C in a CO₂ incubator with 5% CO₂.

Cell viability assay. Effects of compounds on the cell viability of tumor cell lines were examined using a cell viability reagent, MTT (Sigma). Briefly, tumor cells were seeded in a 96-well plate in the absence or presence of tested natural compounds for 96 h. MTT (5 mg/ml in sodium chloride) was added for the last

3 h of incubation, and the absorbances at 570 nm were measured after the dissolve of 20% SDS. The IC₅₀ values were calculated by GraphPad Prism 5.01 software (GraphPad Software, USA).

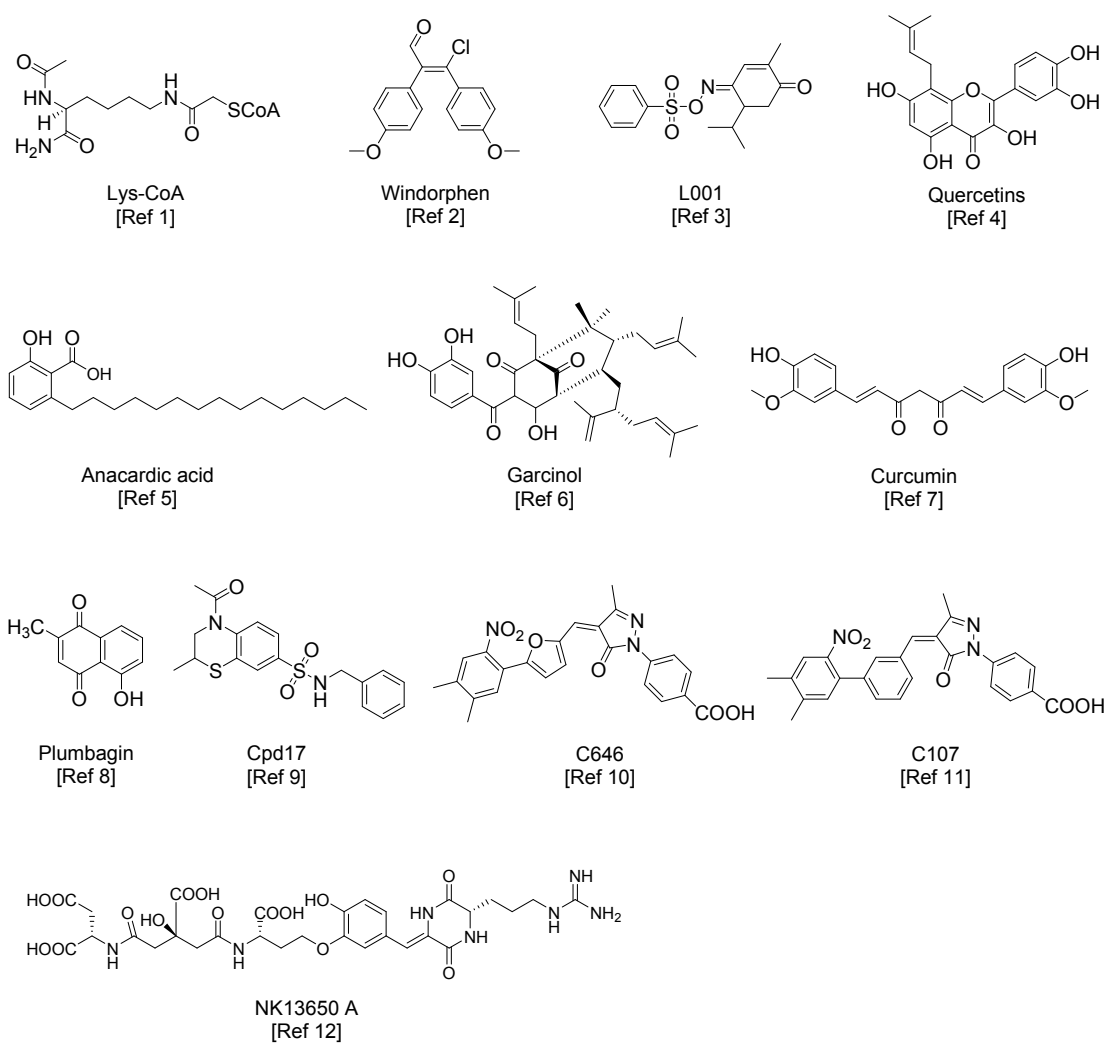


Fig. S1. Chemical structures of known P300 HAT inhibitors²⁻¹³.

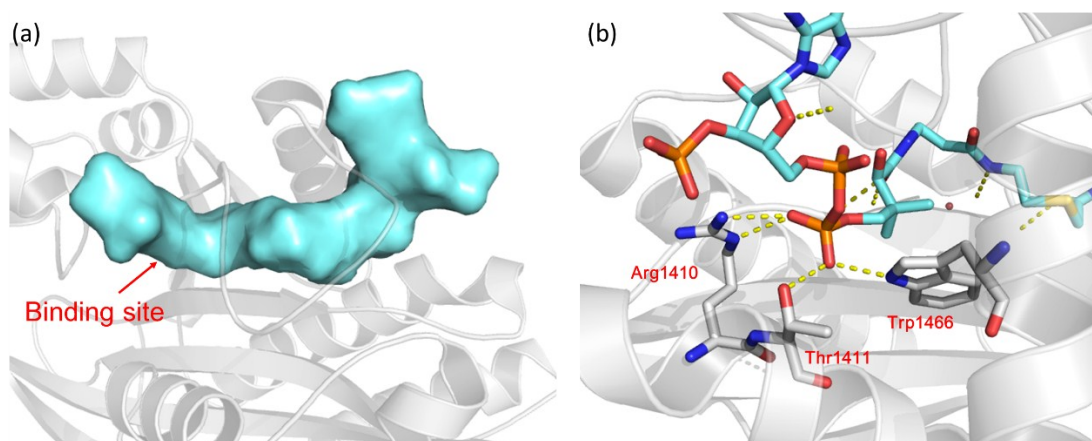


Fig. S2. P300 HAT binding site shape and the key binding feature for substrate Acetyl-CoA. (a) P300 HAT binding site shape, and (b) The key binding feature for substrate Acetyl-CoA which is having hydrogen-bonds with Arg1410, Thr1411, and Trp1466.

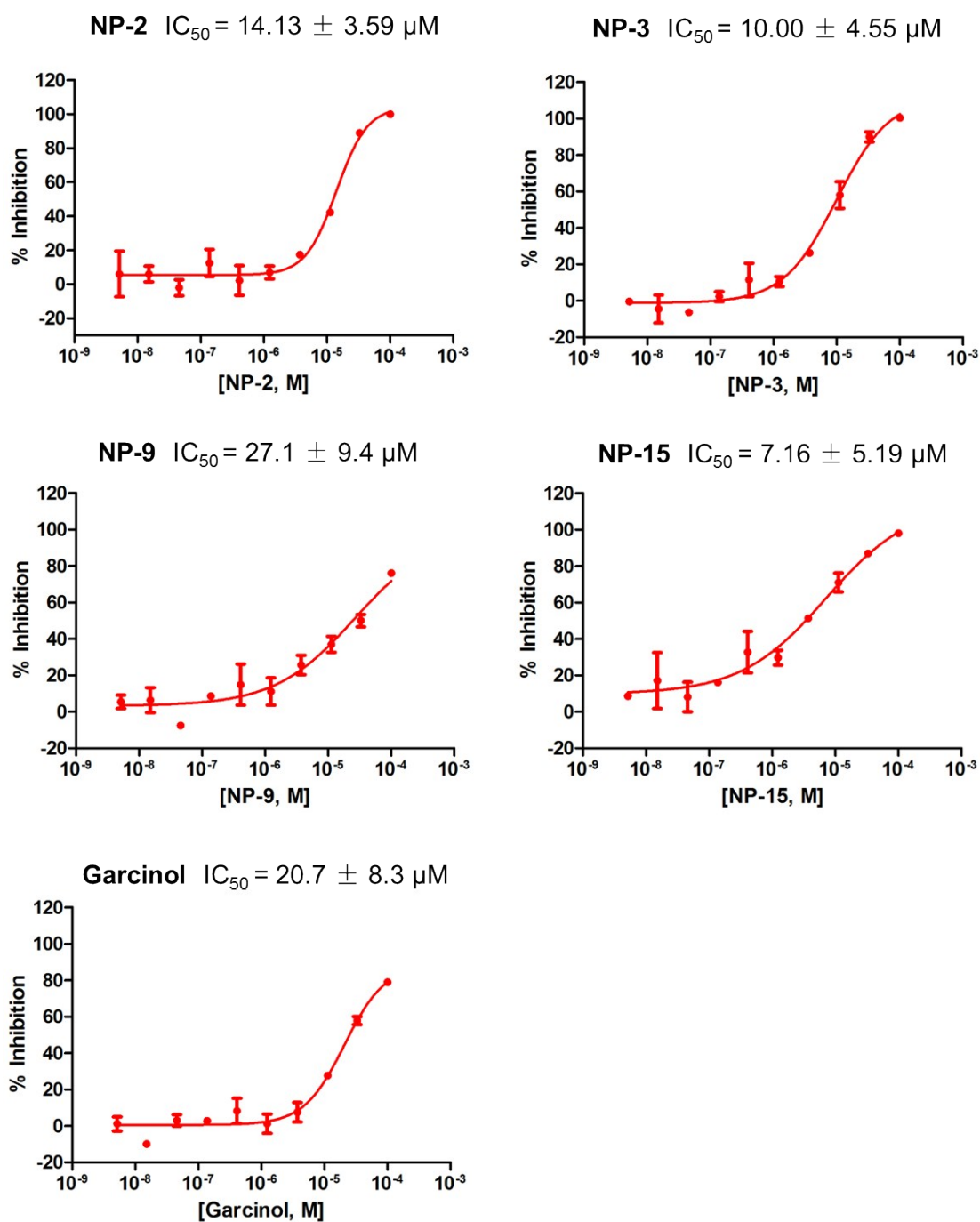


Figure S3. The IC_{50} curves of compounds **NP-2**, **NP-3**, **NP-9**, **NP-15**, and positive control, the known PCAF inhibitor Garcinol (IC_{50} values were reported as *mean* $\pm$ *SD*).

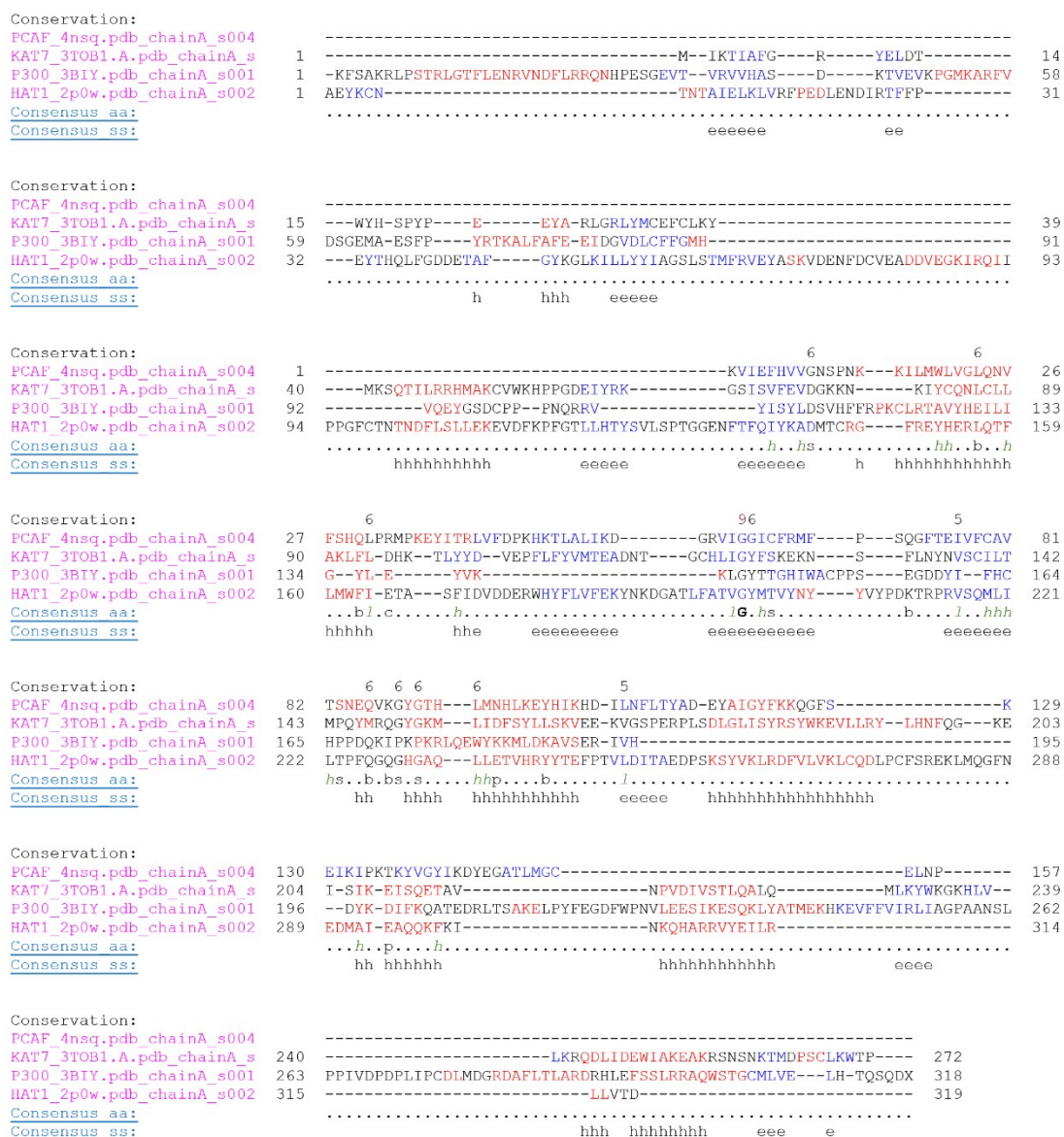


Figure S4. Colored alignment with secondary structure, conservation, and consensus sequence information of p300, PCAF, HAT1, and KAT7 using PROMALS3D¹⁴. The last two lines show consensus amino acid sequence (Consensus_aa) and consensus predicted secondary structures (Consensus_ss). Representative sequences have magenta names and they are colored according to predicted secondary structures (red: alpha-helix, blue: beta-strand). Consensus predicted secondary structure symbols: alpha-helix: h; beta-strand: e. Consensus amino acid symbols are: conserved amino acids are in bold and uppercase letters; aliphatic (I, V, L): l; aromatic (Y, H, W, F): @; hydrophobic (W, F, Y, M, L, I, V, A, C, T, H): h; alcohol (S, T): o; polar residues (D, E, H, K, N, Q, R, S, T): p; tiny (A, G, C, S): t; small (A, G, C, S, V, N, D, T, P): s; bulky residues (E, F, I, K, L, M, Q, R, W, Y): b; positively charged (K, R, H): +; negatively charged (D, E): -; charged (D, E, K, R, H): c.

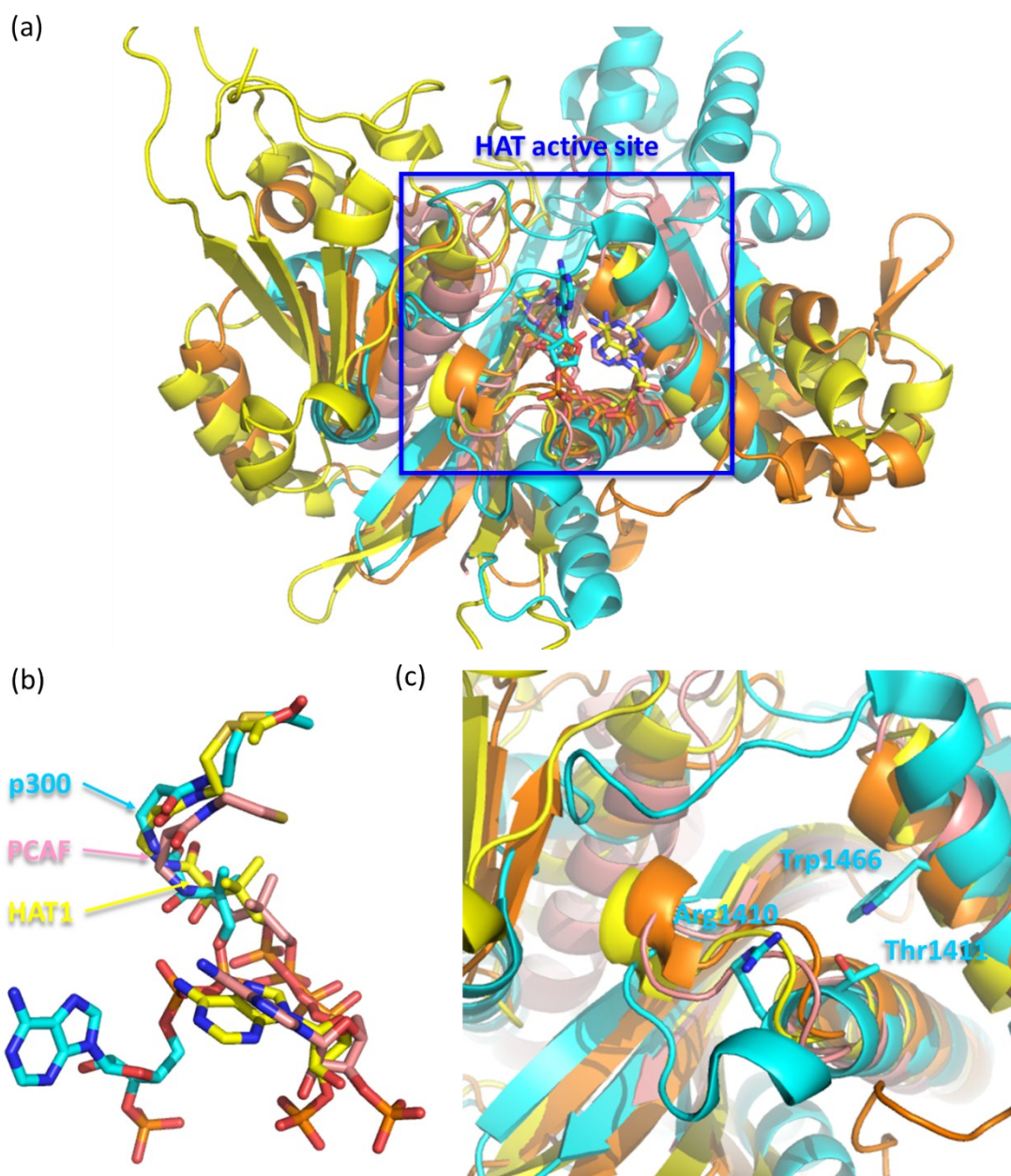
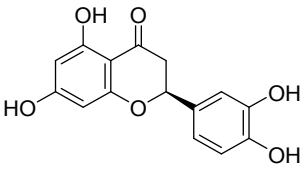
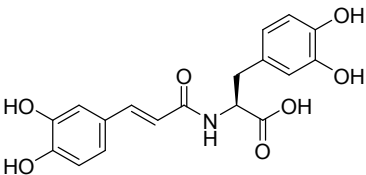
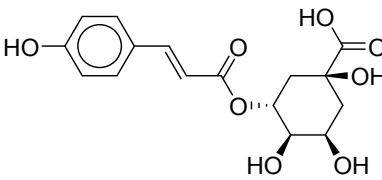
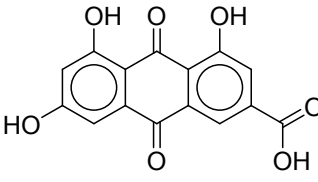
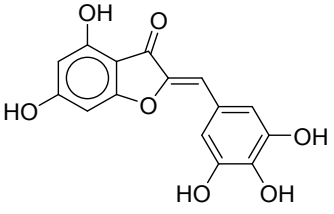
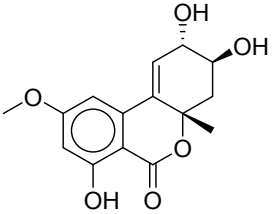
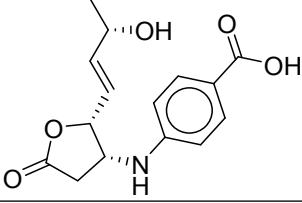
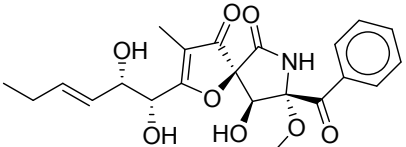
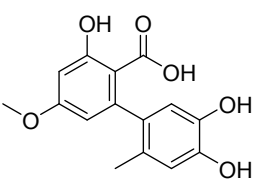
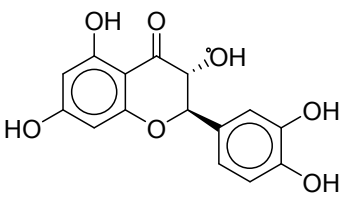
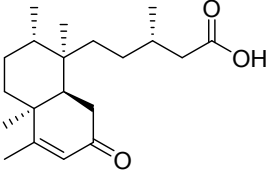
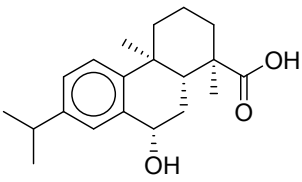
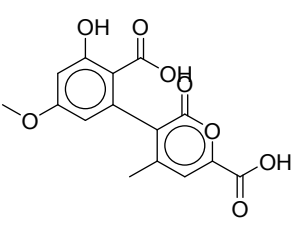
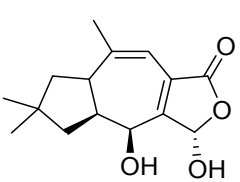
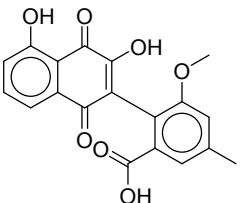
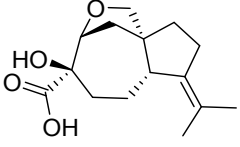
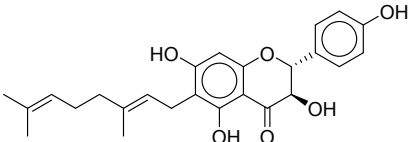
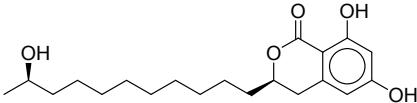
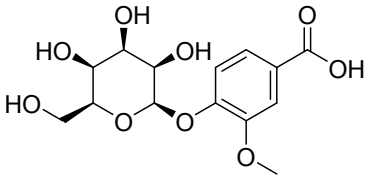
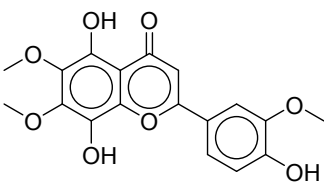
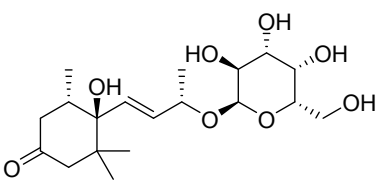
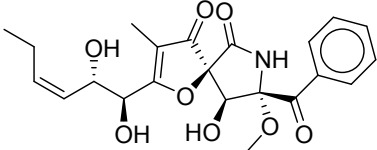
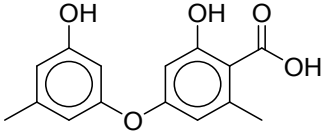
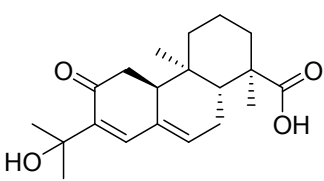
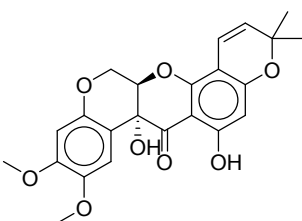


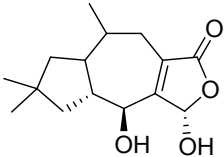
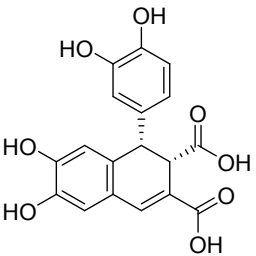
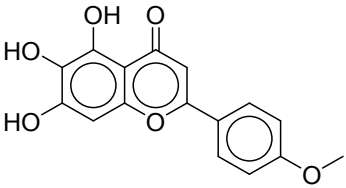
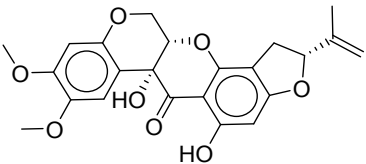
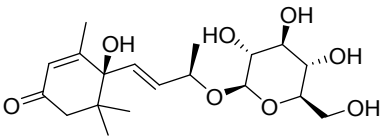
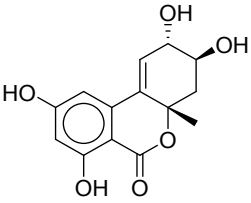
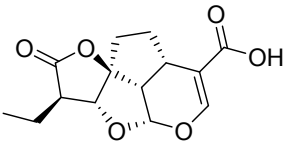
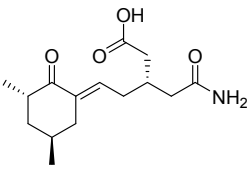
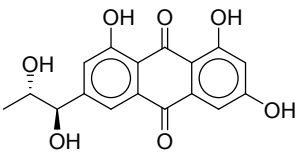
Figure S5. Comparison of tertiary structure, substrate, and key residues of p300, PCAF, HAT1, and KAT7. (a) Alignment of crystal structures of the HAT domain of p300 (PDB code: 4PZS, blue), PCAF (PDB code: 4NSQ, pink), and HAT1 (PDB code: 2P0W, yellow), and the predicted KAT7 HAT domain (orange) using the MICAN program¹⁵, indicating the active site of p300 HAT is significantly different from that of PCAF, HAT1, and KAT7 in protein tertiary structure; (b) The co-crystal substrates (acetyl-CoA or CoA) extracted from (a), the alignment of tertiary structures, showing a significant difference of the binding mode of p300 co-substrate to that of PCAF and HAT1; (c) A comparison of tertiary structures revealed that the key residues Arg1410, Thr1411, and Trp1466 are the specific residues of p300.

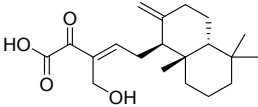
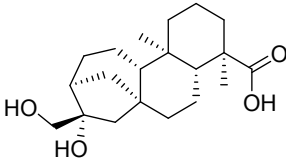
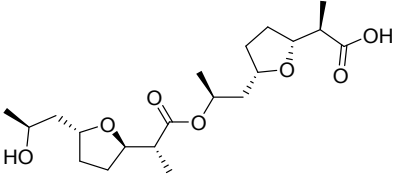
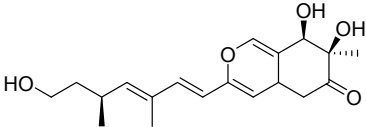
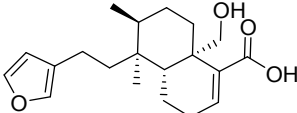
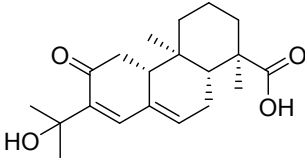
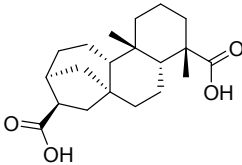
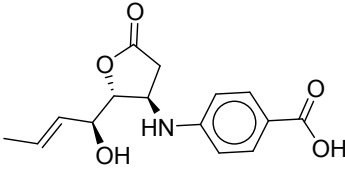
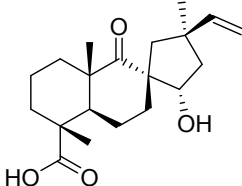
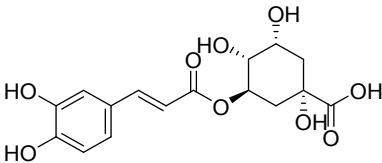
Table S1. Chemical structures of the 110 natural compounds selected as potential P300 inhibitors.

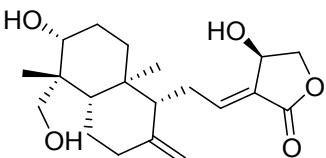
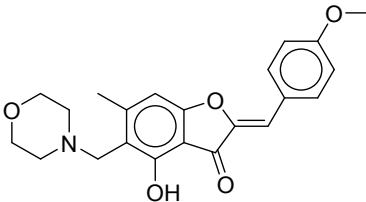
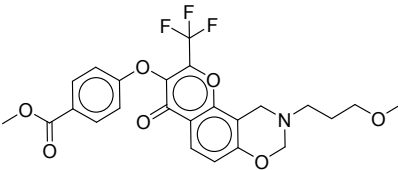
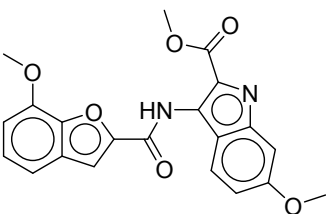
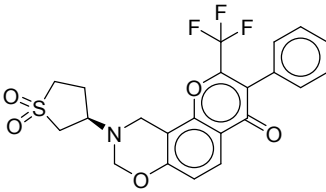
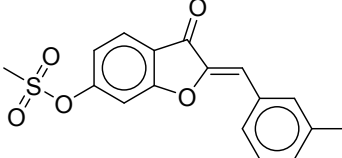
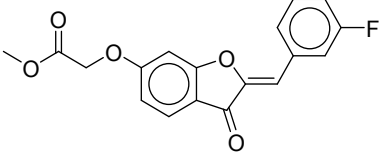
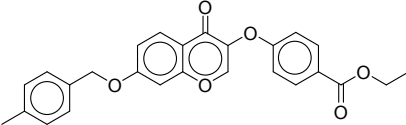
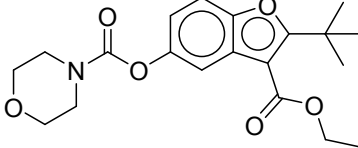
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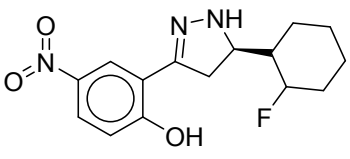
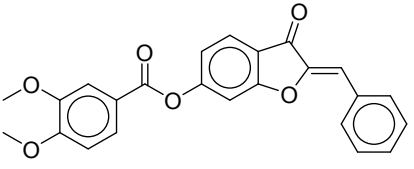
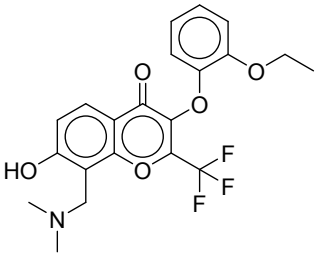
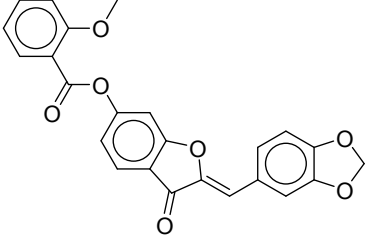
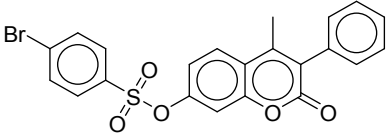
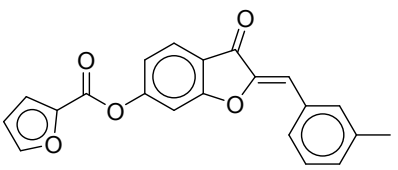
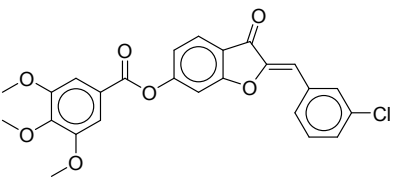
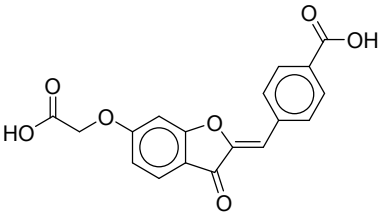
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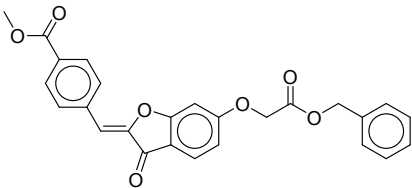
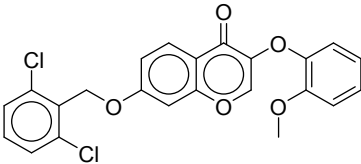
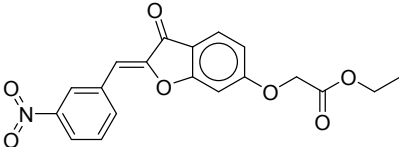
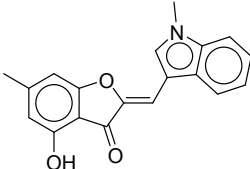
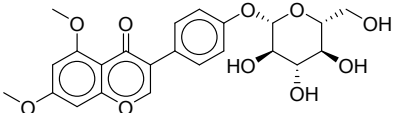
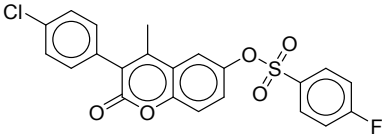
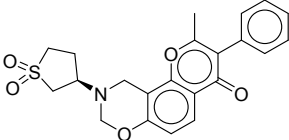
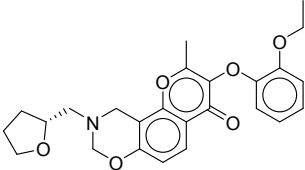
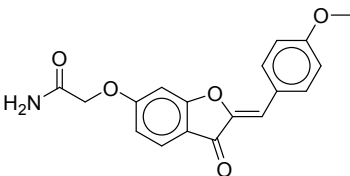
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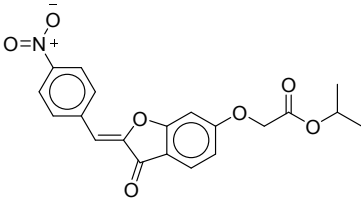
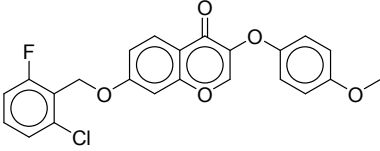
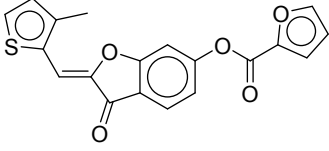
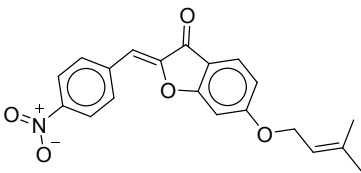
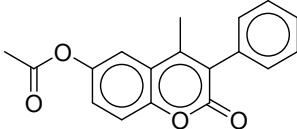
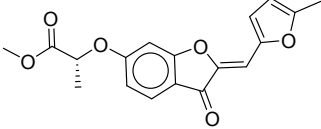
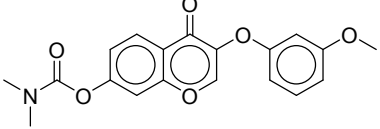
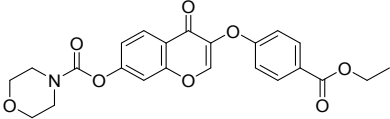
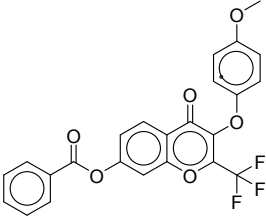
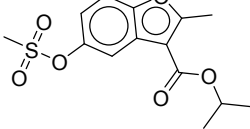
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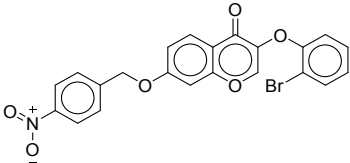
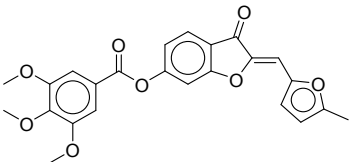
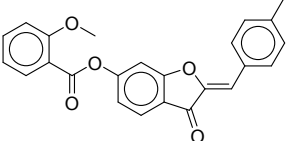
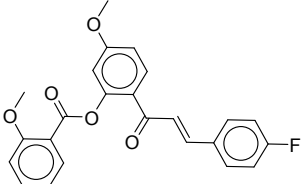
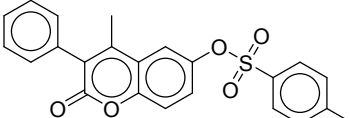
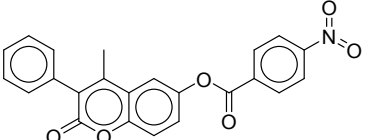
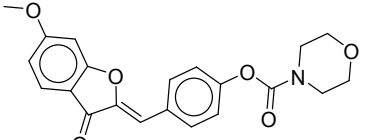
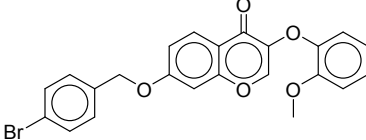
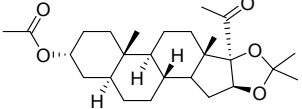
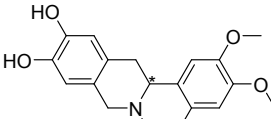
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41	NP-017061	AnalytiCon Discovery NP	
42	NP-006891	AnalytiCon Discovery NP	
43	NP-018849	AnalytiCon Discovery NP	
44	C-002	Indofine NP	

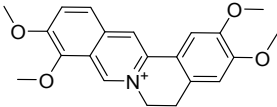
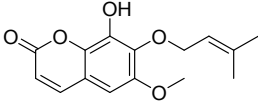
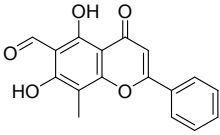
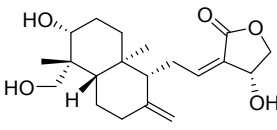
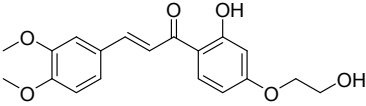
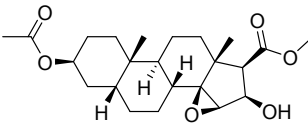
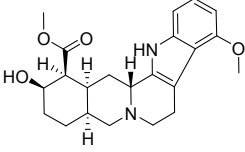
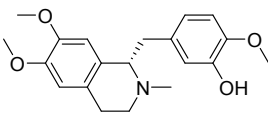
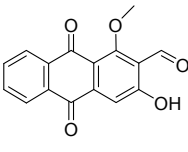
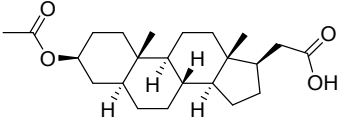
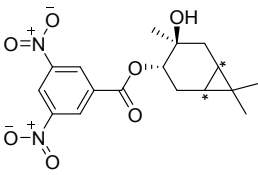
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90 (NP-2)	AP-123/40765218	Specs NP	

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93 (NP-5)	AK-693/43417390	Specs NP	
94 (NP-6)	AM-721/20681006	Specs NP	
95 (NP-7)	AA-504/21163113	Specs NP	
96 (NP-8)	AO-774/41465395	Specs NP	
97 (NP-9)	AK-693/43417378	Specs NP	
98 (NP-10)	AK-198/11618045	Specs NP	
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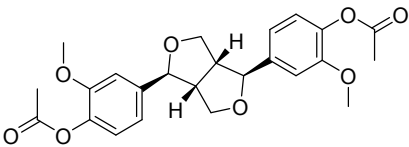
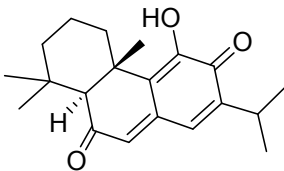
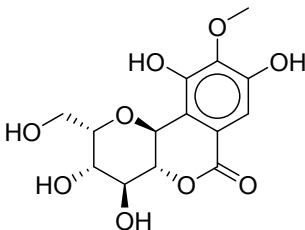
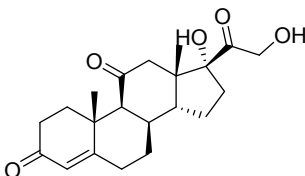
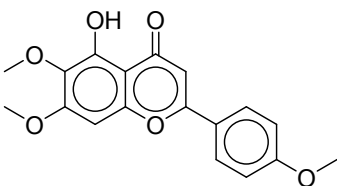
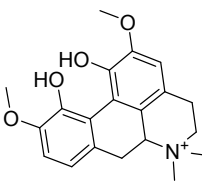
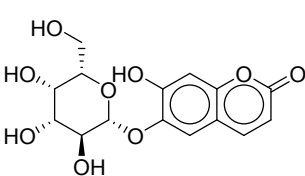
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110	AH-034/11107027	Specs NP	

Table S2. The inhibitory potencies (IC₅₀) of **NP-2**, **NP-3**, **NP-9**, and **NP-15** against eight types of human cancer cell lines.^a

Cancer cell line	Cancer type	IC ₅₀ (μM)			
		NP-2	NP-3	NP-9	NP-15
MDA-MB-231	Triple-negative breast cancer	50-100	>200	>100	11.63
ZR-75-1	Breast cancer	41.09	12.87	>100	2.323
LNCaP	Prostate cancer	50-100	100-200	>100	15.65
DU 145	Prostate cancer	>200	>200	>100	29.98
PANC-1	Pancreatic cancer	100	>200	>100	9.254
HCT 116	Colon cancer	100	100-200	>100	11.81
DLD-1	Colon cancer	>200	>200	>100	6.032
SH-SY5Y	Neuroblastoma	>200	>200	>100	13.21

^a The inhibitory activity was determined by cell viability assays as described in Supporting Information Experimental Methods.

Reference:

1. Li, G.-B.; Yang, L.-L.; Wang, W.-J.; Li, L.-L.; Yang, S.-Y. ID-Score: a new empirical scoring function based on a comprehensive set of descriptors related to protein–ligand interactions. *J. Chem. Inf. Model.* 2013, **53**, 592-600.
2. Lau, O. D.; Kundu, T. K.; Soccio, R. E.; Ait-Si-Ali, S.; Khalil, E. M.; Vassilev, A.; Wolffe, A. P.; Nakatani, Y.; Roeder, R. G.; Cole, P. A. HATs off: selective synthetic inhibitors of the histone acetyltransferases p300 and PCAF. *Mol. Cell* 2000, **5**, 589-595.
3. Hao, J.; Ao, A.; Zhou, L.; Murphy, C. K.; Frist, A. Y.; Keel, J. J.; Thorne, C. A.; Kim, K.; Lee, E.; Hong, C. C. Selective small molecule targeting β -catenin function discovered by in vivo chemical genetic screen. *Cell Rep.* 2013, **4**, 898-904.
4. Yang, H.; Pinello, C. E.; Luo, J.; Li, D.; Wang, Y.; Zhao, L. Y.; Jahn, S. C.; Saldanha, S. A.; Planck, J.; Geary, K. R. Small-molecule inhibitors of acetyltransferase p300 identified by high-throughput screening are potent anticancer agents. *Mol. Cancer Ther.* 2013, **12**, 610-620.
5. Holzhauser, S.; Freiwald, A.; Weise, C.; Multhaup, G.; Han, C. T.; Sauer, S. Discovery and Characterization of Protein - Modifying Natural Products by MALDI Mass Spectrometry Reveal Potent SIRT1 and p300 Inhibitors. *Angew. Chem. Int. Edit.* 2013, **52**, 5171-5174.
6. Sun, Y.; Jiang, X.; Chen, S.; Price, B. D. Inhibition of histone acetyltransferase activity by anacardic acid sensitizes tumor cells to ionizing radiation. *FEBS Lett.* 2006, **580**, 4353-4356.
7. Balasubramanyam, K.; Altaf, M.; Varier, R. A.; Swaminathan, V.; Ravindran, A.;

Sadhale, P. P.; Kundu, T. K. Polyisoprenylated benzophenone, garcinol, a natural histone acetyltransferase inhibitor, represses chromatin transcription and alters global gene expression. *J. Biol. Chem.* 2004, **279**, 33716-33726.

8. Balasubramanyam, K.; Varier, R. A.; Altaf, M.; Swaminathan, V.; Siddappa, N. B.; Ranga, U.; Kundu, T. K. Curcumin, a novel p300/CREB-binding protein-specific inhibitor of acetyltransferase, represses the acetylation of histone/nonhistone proteins and histone acetyltransferase-dependent chromatin transcription. *J. Biol. Chem.* 2004, **279**, 51163-51171.

9. Ravindra, K. C.; Selvi, B. R.; Arif, M.; Reddy, B. A.; Thanuja, G. R.; Agrawal, S.; Pradhan, S. K.; Nagashayana, N.; Dasgupta, D.; Kundu, T. K. Inhibition of lysine acetyltransferase KAT3B/p300 activity by a naturally occurring hydroxynaphthoquinone, plumbagin. *J. Biol. Chem.* 2009, **284**, 24453-24464.

10. Zeng, F.; Peng, S.; Li, L.; Mu, L.; Zhang, Z.; Zhang, Z.; Huang, N. Structure-based identification of drug-like inhibitors of p300 histone acetyltransferase. *Acta Pharm. Sinica* 2013, **48**, 700-708.

11. Bowers, E. M.; Yan, G.; Mukherjee, C.; Orry, A.; Wang, L.; Holbert, M. A.; Crump, N. T.; Hazzalin, C. A.; Liszczak, G.; Yuan, H. Virtual ligand screening of the p300/CBP histone acetyltransferase: identification of a selective small molecule inhibitor. *Chem. Biol.* 2010, **17**, 471-482.

12. Dancy, B. M.; Crump, N. T.; Peterson, D. J.; Mukherjee, C.; Bowers, E. M.; Ahn, Y. H.; Yoshida, M.; Zhang, J.; Mahadevan, L. C.; Meyers, D. J. Live - Cell Studies of p300/CBP Histone Acetyltransferase Activity and Inhibition. *Chembiochem* 2012, **13**,

2113-2121.

13. Tohyama, S.; Tomura, A.; Ikeda, N.; Hatano, M.; Odanaka, J.; Kubota, Y.; Umekita, M.; Igarashi, M.; Sawa, R.; Morino, T. Discovery and characterization of NK13650s, naturally occurring p300-selective histone acetyltransferase inhibitors. *J. Org. Chem.* 2012, **77**, 9044-9052.
14. Pei, J.; Kim, B.-H.; Grishin, N. V. PROMALS3D: a tool for multiple protein sequence and structure alignments. *Nucleic Acids Res.* 2008, **36**, 2295-2300.
15. Minami, S.; Sawada, K.; Chikenji, G. MICAN : a protein structure alignment algorithm that can handle Multiple-chains, Inverse alignments, C α only models, Alternative alignments, and Non-sequential alignments. *BMC Bioinformatics* 2013, **14**, 1-22.