

## New copper(II) complexes of anti-inflammatory drug mefenamic acid: a concerted study including synthesis, physicochemical characterization and their biological evaluation

Raj Pal Sharma,<sup>a,\*</sup> Santosh Kumar,<sup>a</sup> Paloth Venugopalan,<sup>a</sup> Valeria Ferretti,<sup>b,\*</sup> Alketa Tarushi,<sup>c</sup> George Psomas <sup>c,\*</sup> and Maciej Witwicki<sup>d</sup>

<sup>a</sup>*Department of Chemistry, Panjab University, Chandigarh-160014, India.*

<sup>b</sup>*Center for Structural Diffractometry and Department of Chemical and Pharmaceutical Sciences, University of Ferrara, via Fossato di Mortara 17-27, I-44100, Ferrara, Italy.*

<sup>c</sup>*Laboratory of Inorganic Chemistry, Department of General and Inorganic Chemistry, Faculty of Chemistry, Aristotle university of Thessaloniki, GR-54124 Thessaloniki, Greece.*

<sup>d</sup>*Faculty of Chemistry, Wroclaw University, 14 F. Joliot-Curie St., Wroclaw 50-283, Poland.*

## S1. Interaction with CT DNA

The DNA-binding constant complexes **1–3** ( $K_b$ , in  $M^{-1}$ ) may be obtained by monitoring the changes in the absorbance at the corresponding  $\lambda_{max}$  in the UV-Vis spectra with increasing concentrations of CT DNA.  $K_b$  is given by the ratio of slope to the y intercept in plots  $\frac{[DNA]}{(\varepsilon_A - \varepsilon_f)}$  versus [DNA], according to the Wolfe–Shimer equation [1]:

$$\frac{[DNA]}{(\varepsilon_A - \varepsilon_f)} = \frac{[DNA]}{(\varepsilon_b - \varepsilon_f)} + \frac{1}{K_b(\varepsilon_b - \varepsilon_f)} \quad (\text{eq. S1})$$

where [DNA] is the concentration of DNA in base pairs,  $\varepsilon_A = A_{\text{obsd}}/\text{[compound]}$ ,  $\varepsilon_f$  = the extinction coefficient for the free compound and  $\varepsilon_b$  = the extinction coefficient for the compound in the fully bound form.

## S2. Competitive studies with EB

The Stern–Volmer constant  $K_{SV}$  (in  $M^{-1}$ ) is used to evaluate the quenching efficiency for each compound according to the Stern–Volmer equation [2]:

$$\frac{I_0}{I} = 1 + K_{SV}[Q] \quad (\text{eq. S2})$$

where  $I_0$  and  $I$  are the emission intensities in the absence and the presence of the quencher, respectively,  $[Q]$  is the concentration of the quencher (i.e. complexes **1–3**);  $K_{SV}$  is obtained from the Stern–Volmer plots by the slope of the diagram  $\frac{I_0}{I}$  versus  $[Q]$ .

## S3. Interaction with serum albumins

The extent of the inner-filter effect can be roughly estimated with the following formula:

$$I_{\text{corr}} = I_{\text{meas}} \times 10^{\frac{\varepsilon(\lambda_{\text{exc}})cd}{2}} \times 10^{\frac{\varepsilon(\lambda_{\text{em}})cd}{2}} \quad (\text{eq. S3})$$

where  $I_{\text{corr}}$  = corrected intensity,  $I_{\text{meas}}$  = the measured intensity,  $c$  = the concentration of the quencher,  $d$  = the cuvette (1 cm),  $\varepsilon(\lambda_{\text{exc}})$  and  $\varepsilon(\lambda_{\text{em}})$  = the  $\varepsilon$  of the quencher at the excitation and the emission wavelength, respectively, as calculated from the UV-vis spectra of the complexes [3].

The Stern–Volmer and Scatchard graphs are used in order to study the interaction of a quencher with serum albumins. According to Stern–Volmer quenching equation [2]:

$$\frac{I_0}{I} = 1 + k_q \tau_0 [Q] = 1 + K_{SV} [Q] \quad (\text{eq. S4}),$$

where  $I_0$  = the initial tryptophan fluorescence intensity of SA,  $I$  = the tryptophan fluorescence intensity of SA after the addition of the quencher,  $k_q$  = the quenching rate constants of SA,  $K_{SV}$  =

the dynamic quenching constant,  $\tau_0$  = the average lifetime of SA without the quencher,  $[Q]$  = the concentration of the quencher, the dynamic quenching constant ( $K_{SV}$ ,  $M^{-1}$ ) can be obtained by the slope of the diagram  $\frac{I_0}{I}$  versus  $[Q]$ . From the equation:

$$K_{SV} = k_q \tau_0 \quad (\text{eq. S5})$$

and taking  $\tau_0 = 10^{-8}$  s as fluorescence lifetime of tryptophan in SA, the approximate quenching constant ( $k_q$ ,  $M^{-1}s^{-1}$ ) is calculated.

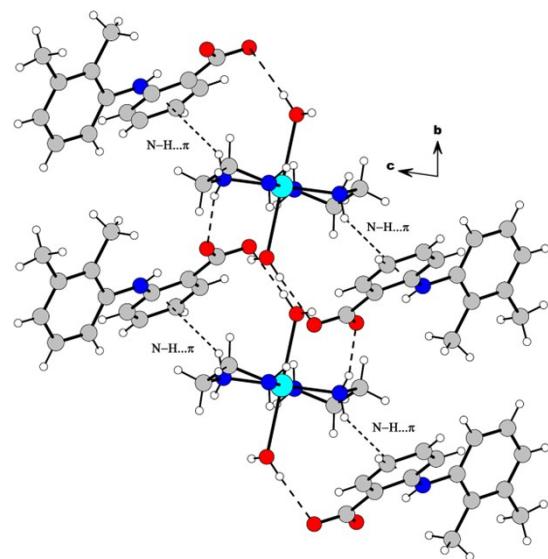
From the Scatchard equation [2]:

$$\frac{\Delta I/I_0}{[Q]} = nK - K \frac{\Delta I}{I_0} \quad (\text{eq. S6})$$

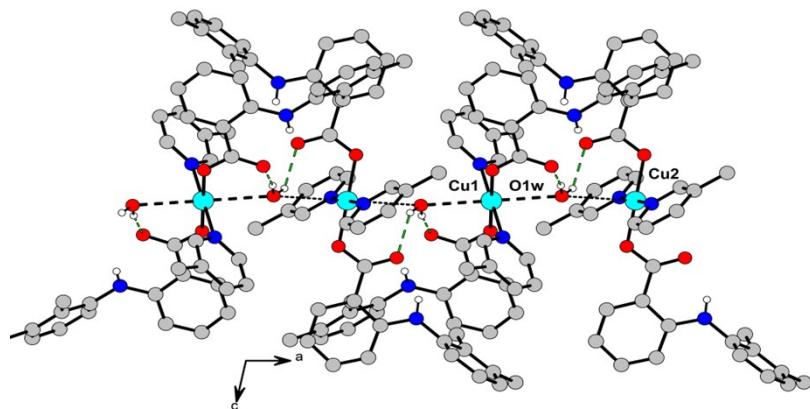
where  $n$  is the number of binding sites per albumin and  $K$  is the association binding constant,  $K$  ( $M^{-1}$ ) is calculated from the slope in plots  $\frac{\Delta I/I_0}{[Q]}$  versus  $\frac{\Delta I}{I_0}$  and  $n$  is given by the ratio of y intercept to the slope [2].

## References

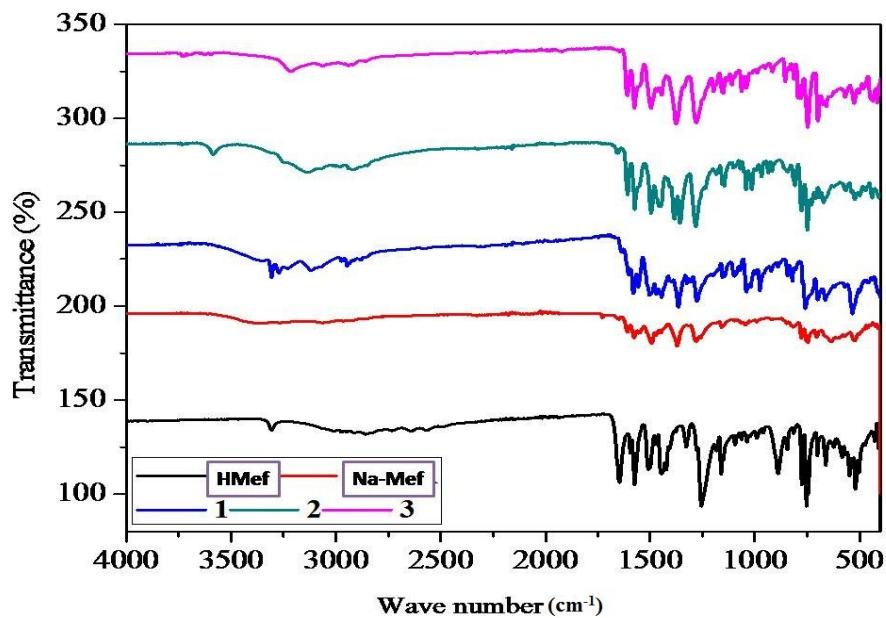
- [1] A. Wolfe, G. Shimer and T. Meehan, *Biochemistry* 1987, **26**, 6392.
- [2] Y. Wang, H. Zhang, G. Zhang, W. Tao and S. Tang, *J. Luminescence* 2007, **126**, 211.
- [3] L. Stella, A.L. Capodilupo and M. Bietti, *Chem. Commun.* 2008, 4744.



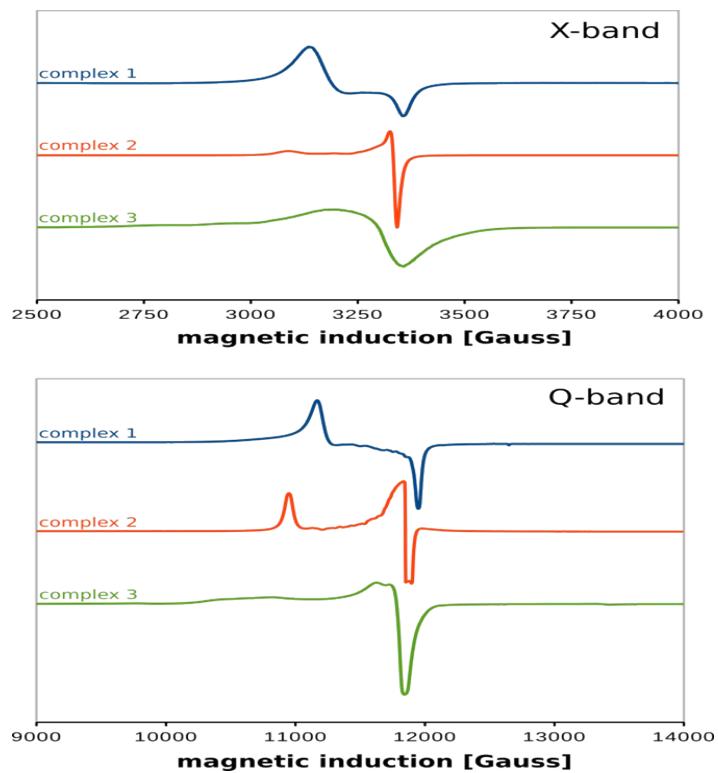
**Figure S1.** Packing diagram of complex **2**, stabilized by various non-covalent interactions such as N-H... $\pi$ , O-H...O and N-H...O hydrogen bonding interactions.



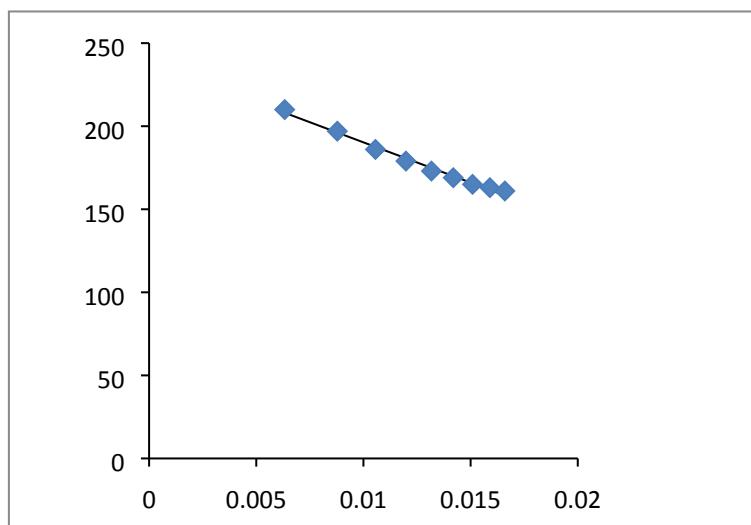
**Figure S2.** Chain of  $[\text{Cu}(\beta\text{-pic})_2(\text{mef})_2]$  in complex **3** running along the *a* axis (Cu..Ow contacts and Ow-H...O hydrogen bonds are shown as black and green broken bonds, respectively)



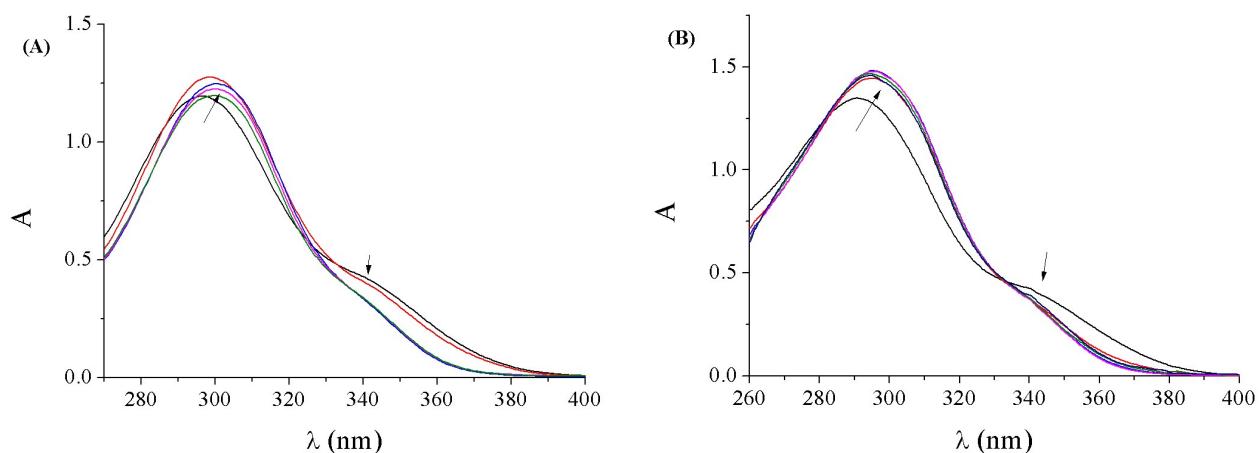
**Figure S3.** FT-IR spectra of complexes **1-3** in comparison with mefenamic acid and sodium salt of mefenamic acid.



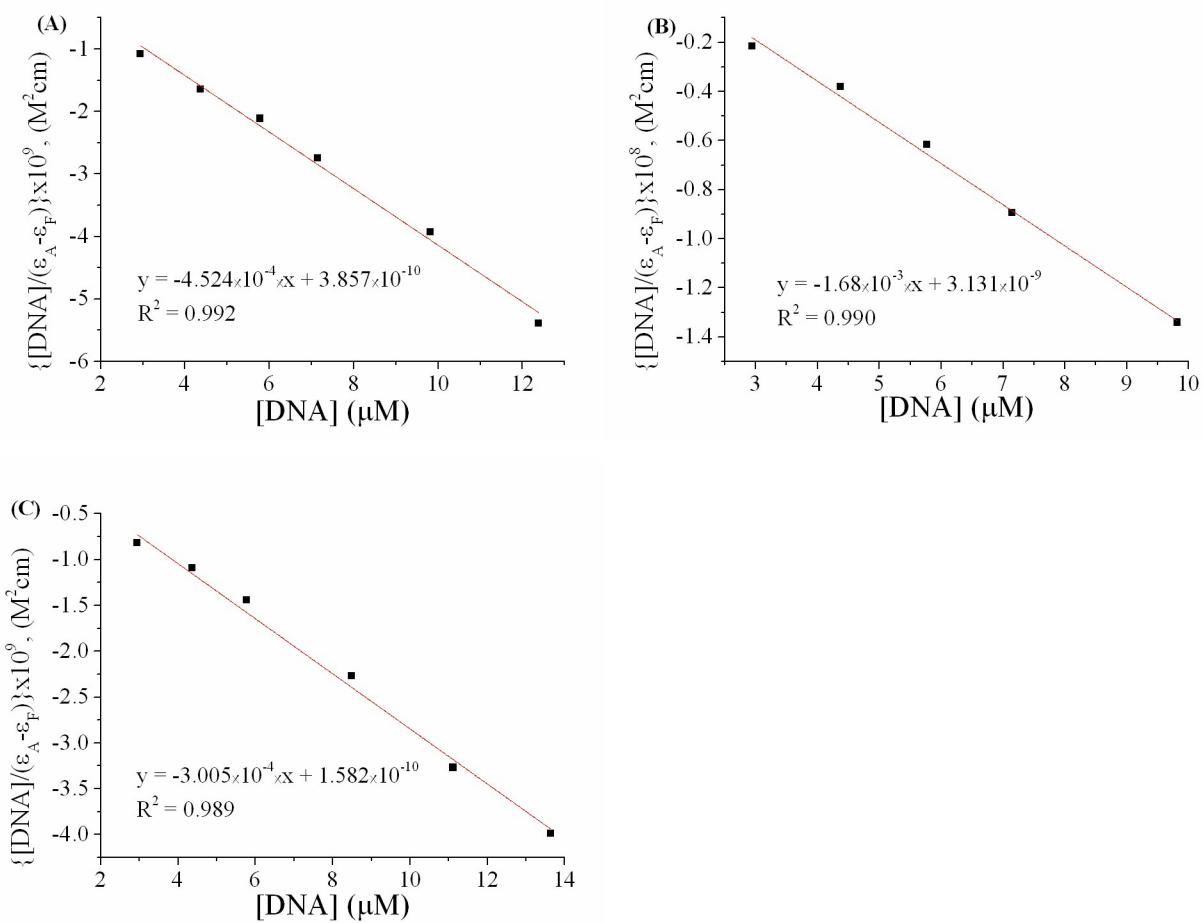
**Figure S4.** X- (9.6 GHz) and Q-band (34 GHz) EPR spectra of powdered complex **1**, **2** and **3** at 77 K (X-band) and 100 K (Q-band). The parameters are given in the text.



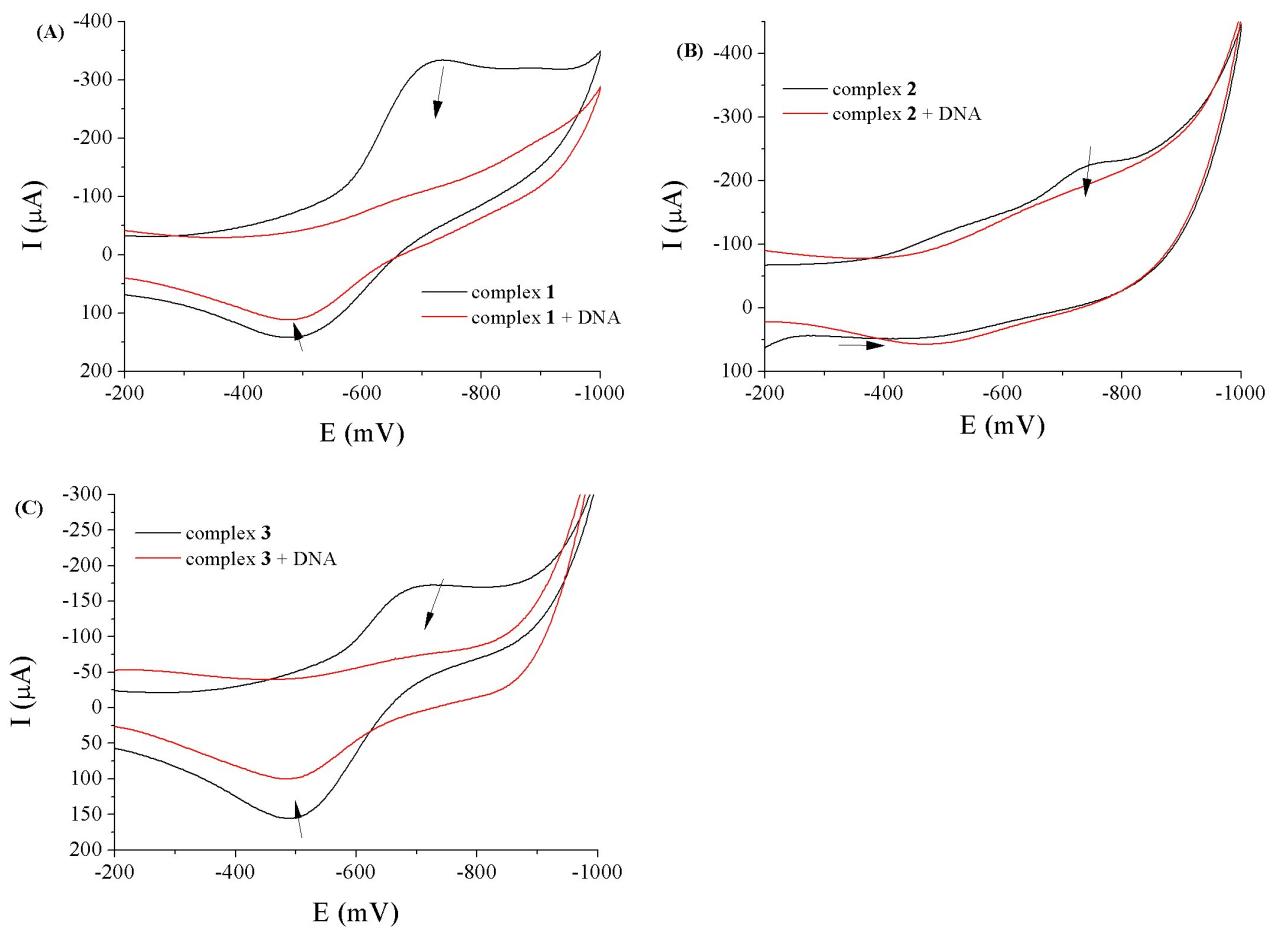
**Figure S5.** Plot of K (molar conductance) versus  $C^{1/2}$  (square root of concentration) of complex **2**.



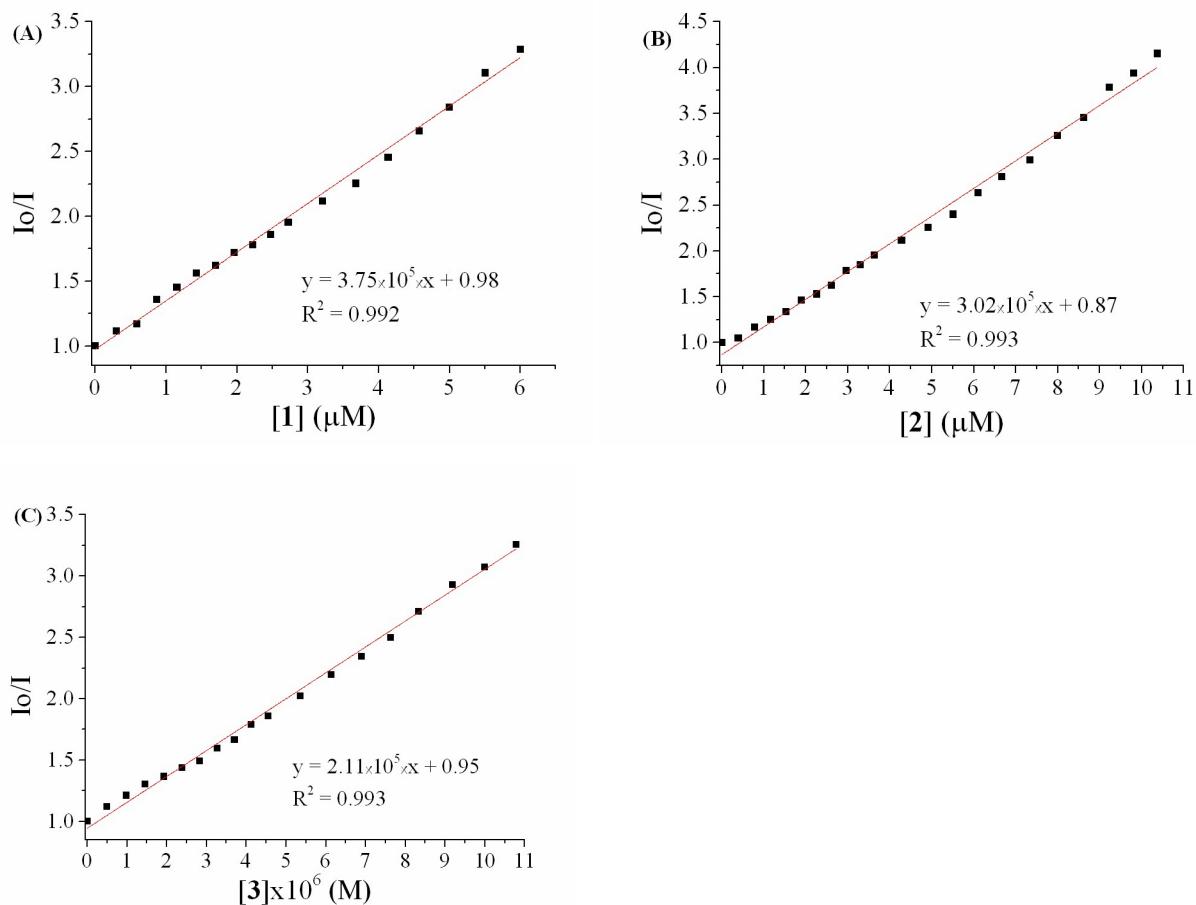
**Figure S6.** UV-Vis spectra of DMSO solution of complex (A) **1** ( $3 \times 10^{-5}$  M) and (B) **2** ( $4 \times 10^{-5}$  M) in the presence of increasing amounts of CT DNA ( $r' = [\text{DNA}]/[\text{compound}] = 0-0.8$ ). The arrows show the changes upon increasing amounts of CT DNA.



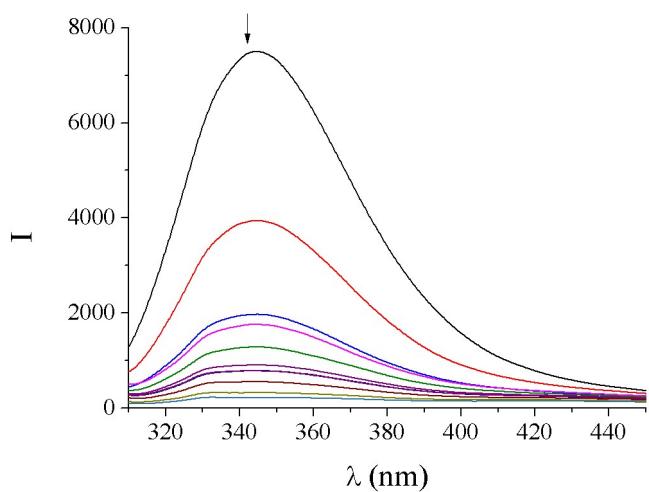
**Figure S7.** (A)–(C) Plot of  $\frac{[DNA]}{(\varepsilon_A - \varepsilon_f)}$  versus [DNA] for complexes **1–3**, respectively.



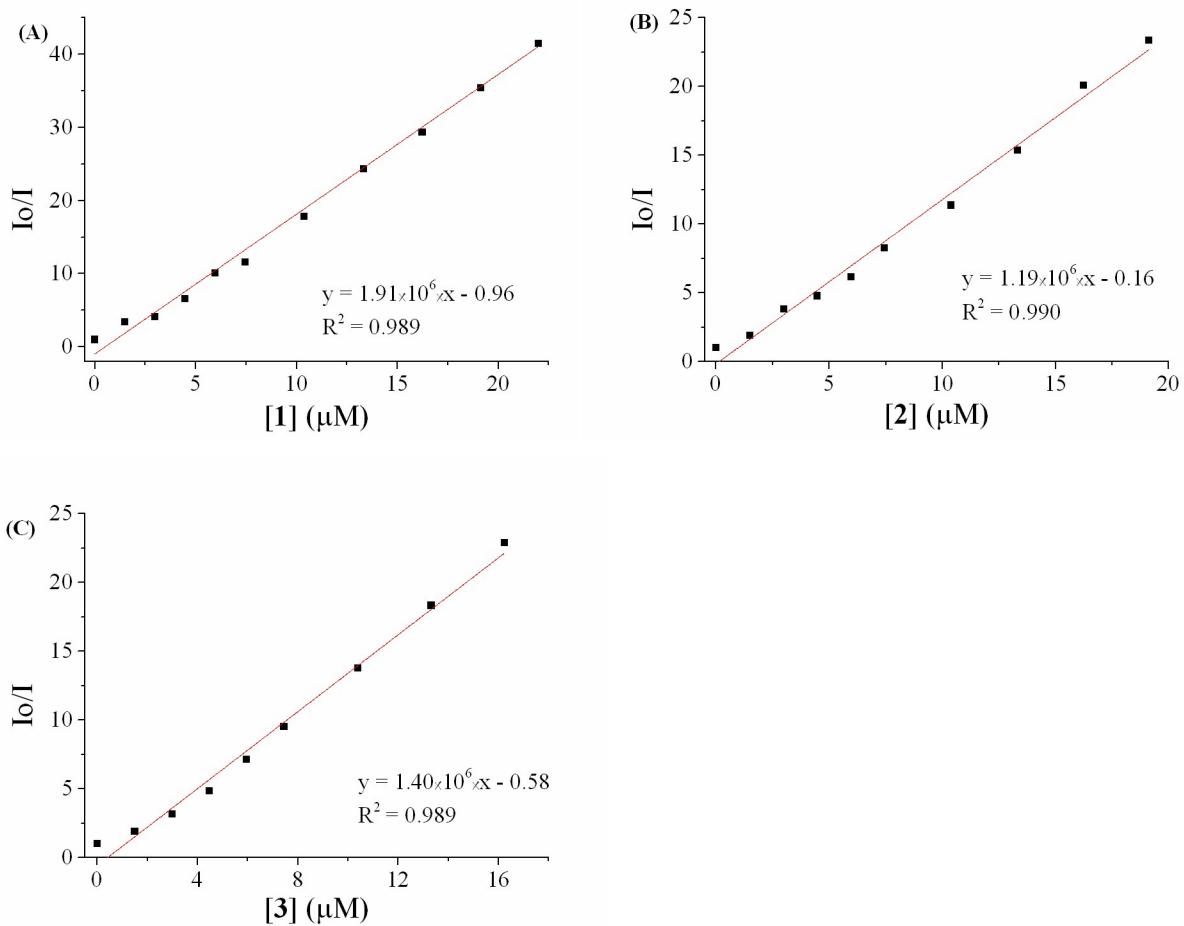
**Figure S8.** Cyclic voltammogram of 0.33 mM 1/2 DMSO/buffer (containing 150 mM NaCl and 15 mM trisodium citrate at pH 7.0) solution of complex (A) **1**, (B) **2** and (C) **3** in the absence or presence of CT DNA. Scan rate = 100 mV  $\text{s}^{-1}$ . Supporting electrolyte = buffer solution.



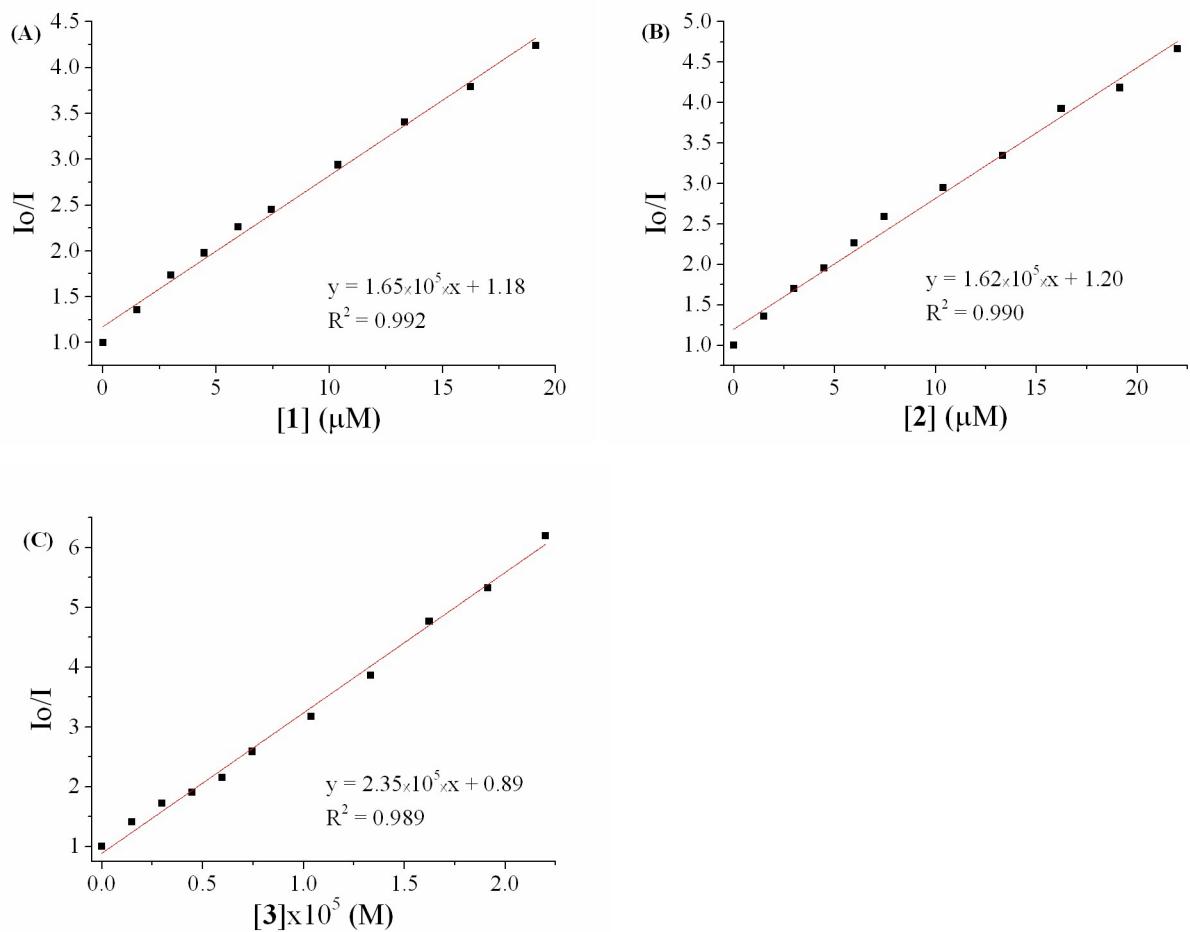
**Figure S9.** (A)–(C) Stern–Volmer quenching plot of EB bound to CT DNA for complexes **1**–**3**, respectively.



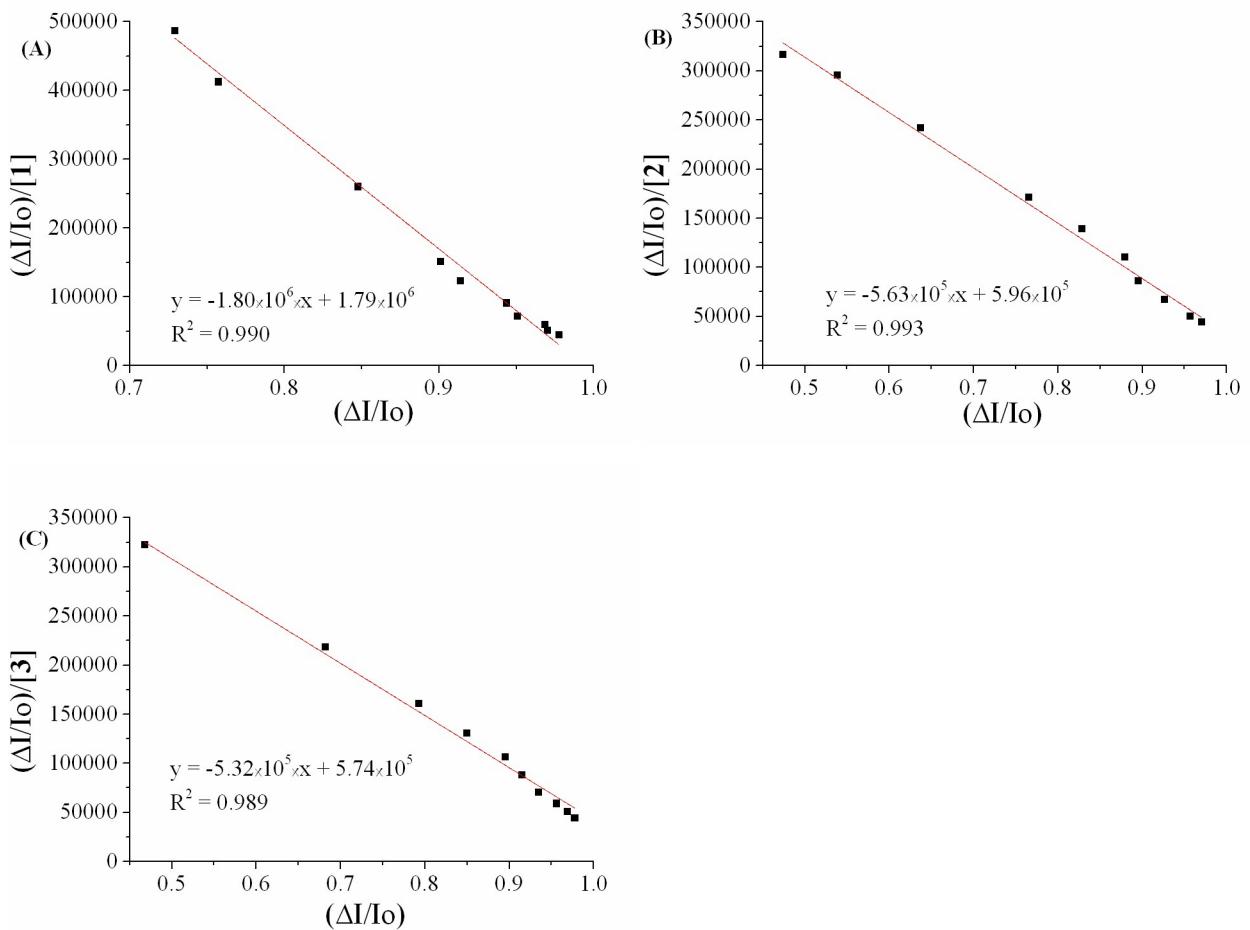
**Figure S10.** Fluorescence emission spectra ( $\lambda_{\text{exc}} = 295 \text{ nm}$ ) for BSA ( $[\text{BSA}] = 3 \mu\text{M}$ ) in buffer solution in the absence and presence of increasing amounts of **2** ( $r = [2]/[\text{BSA}] = 0$ – $7.3$ ). The arrow shows the changes of intensity upon increasing amounts of the complex.



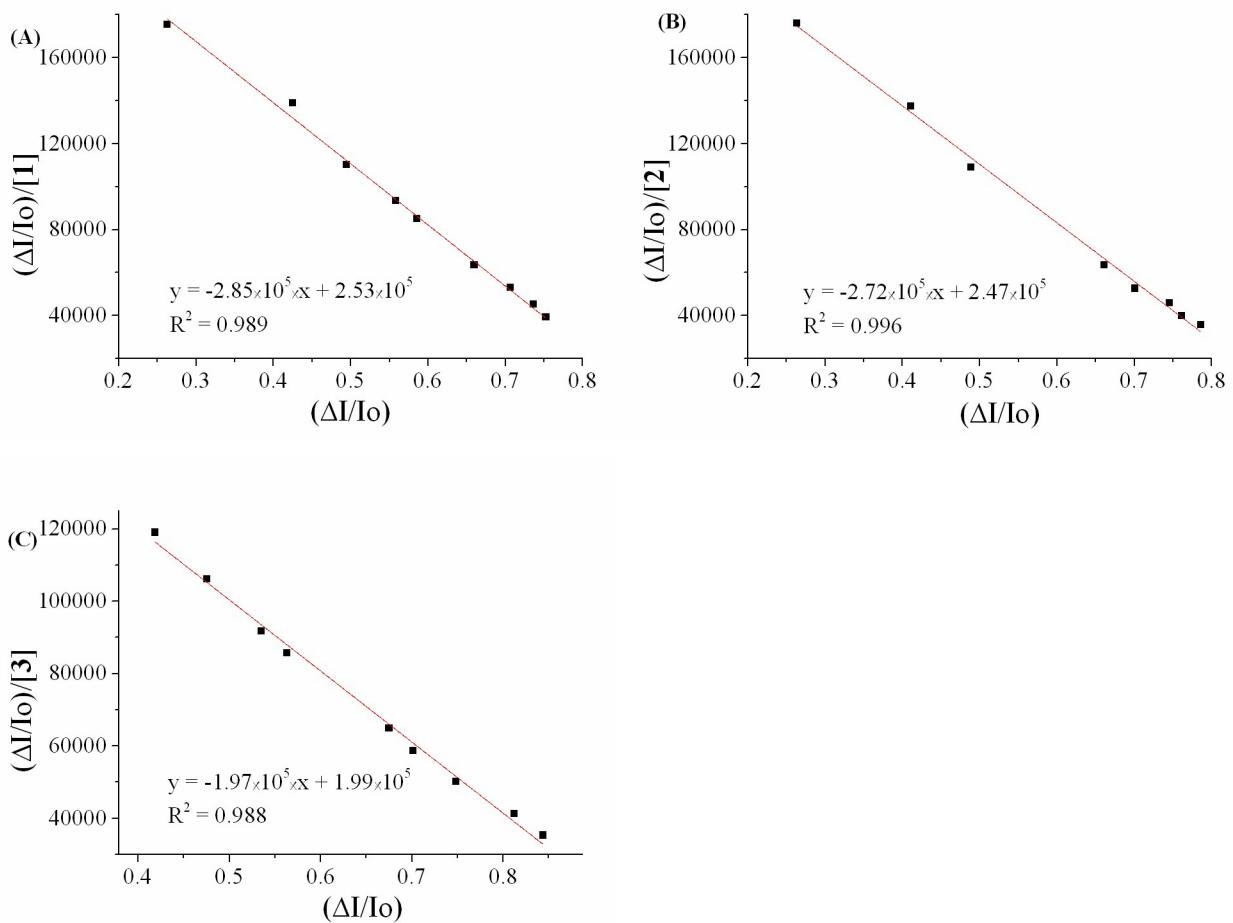
**Figure S11.** (A)–(C) Stern–Volmer quenching plot of BSA for complexes **1**–**3**, respectively.



**Figure S12.** (A)–(C) Stern–Volmer quenching plot of HSA for complexes **1**–**3**, respectively.



**Figure S13.** (A)–(C) Scatchard plot of BSA for complexes **1–3**, respectively.



**Figure S14.** (A)–(C) Scatchard plot of HSA for complexes **1–3**, respectively.

**Table S1.** Hydrogen bonding parameters of complexes **2** and **3**; D= donor, A= acceptor.

D-H...A	D-H (Å)	D...A (Å)	H...A (Å)	<D-H...A (°)	Equivalent positions
<b>Complex 2</b>					
N1-H...O1	0.90	2.968(3)	2.10	161	
O1W-H...O1	0.76(3)	2.832(3)	2.09(3)	164(3)	
N3-H...O1	0.84(3)	2.611(2)	1.89(2)	143(2)	
N2-H...O2 <sup>i</sup>	0.90	2.980(3)	2.10	165	(i) -x,-y,1-z
N2-H...O2 <sup>ii</sup>	0.90	3.098(2)	2.30	146	(ii) x-1,y,z
O1W-H...O2 <sup>iii</sup>	0.80(3)	2.812(3)	2.01(3)	173(3)	(iii)-x,1-y,1-z
<b>Complex 3</b>					
N2-H...O	0.79(4)	2.651(6)	1.99(4)	140(4)	
O1W-H...O2	0.87(7)	2.726(6)	1.86(7)	171(6)	
O1W-H...O4	0.74(7)	2.752(6)	2.12(6)	142(6)	
N4-H...O4	0.80(7)	2.632(5)	1.91(6)	149(7)	