

Supplementary Information

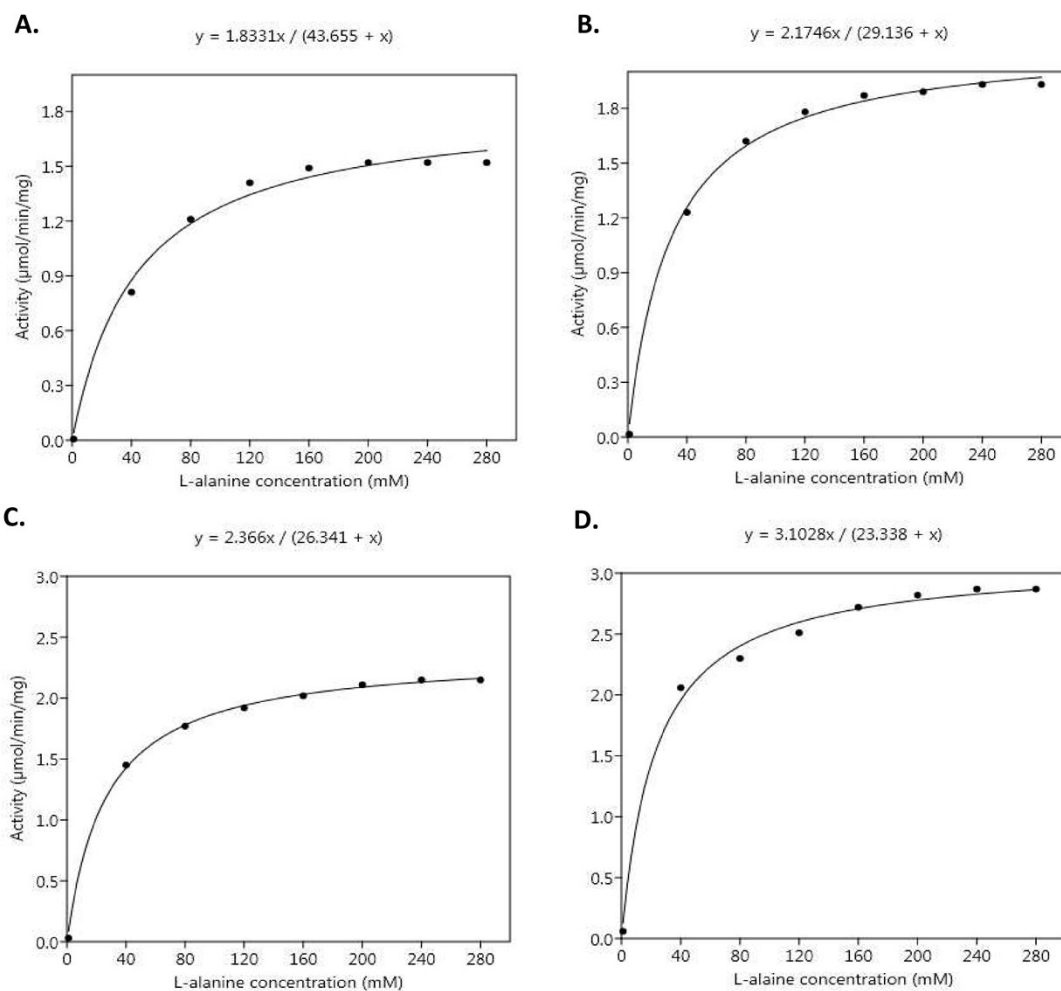


Figure S1: Michaelis-Menten graphs and curve fits using L-alanine as a substrate. **A.** Wild type pm1; **B.** pm1ep1; **C.** pm1ep2; & **D.** pm1ep3.

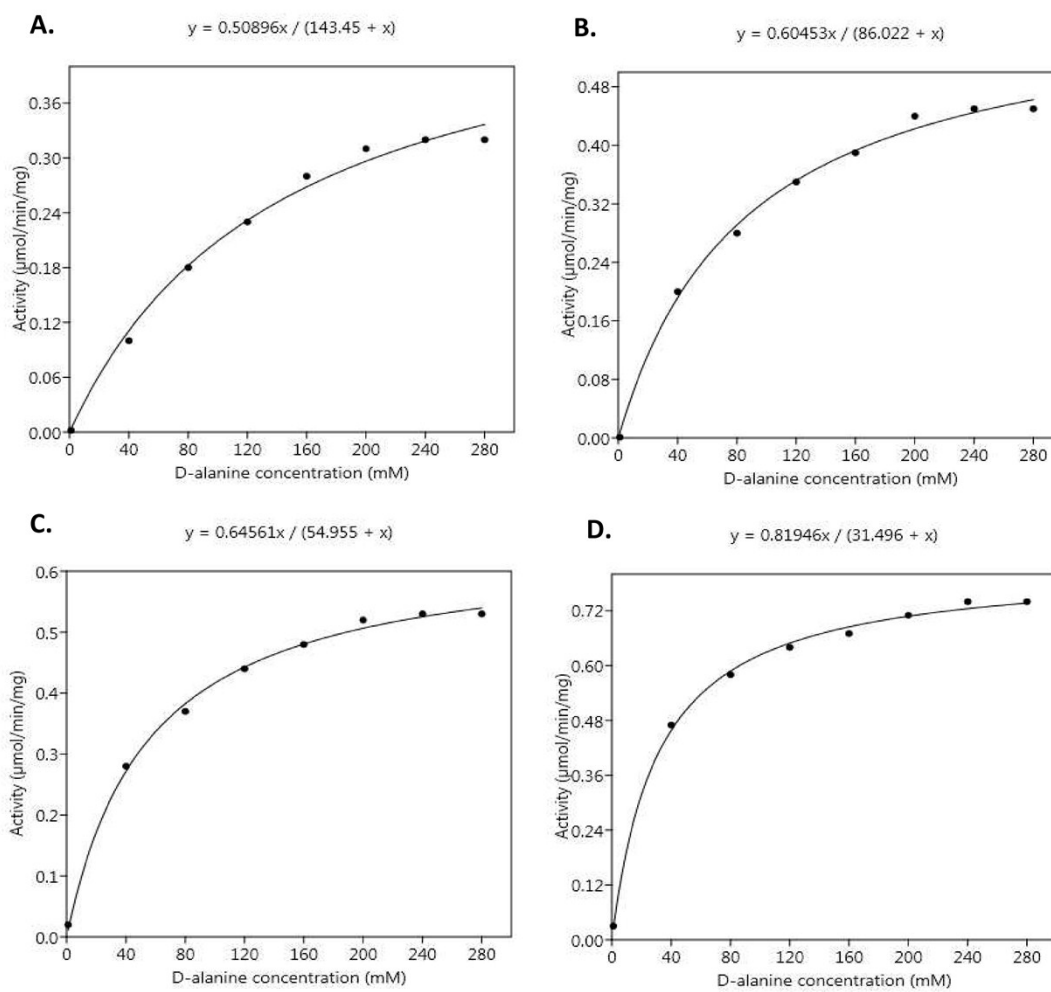


Figure S2: Michaelis-Menten graphs and curve fits using D-alanine as a substrate. **A.** Wild type pm1; **B.** pm1ep1; **C.** pm1ep2; & **D.** pm1ep3.

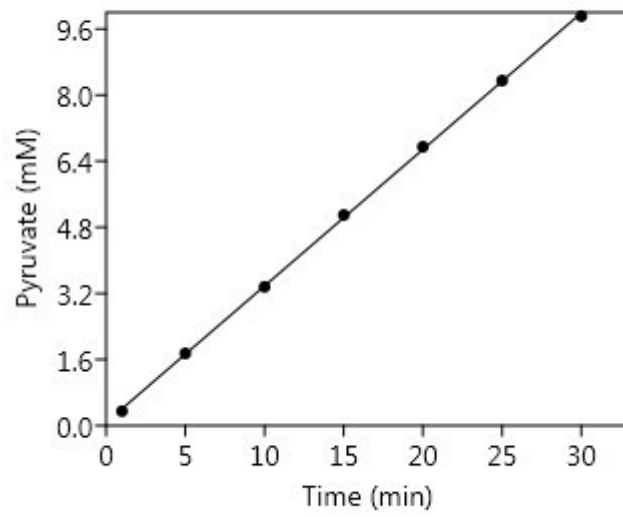


Figure S3: Initial pyruvate production by recombinant strain containing pm1ep3.

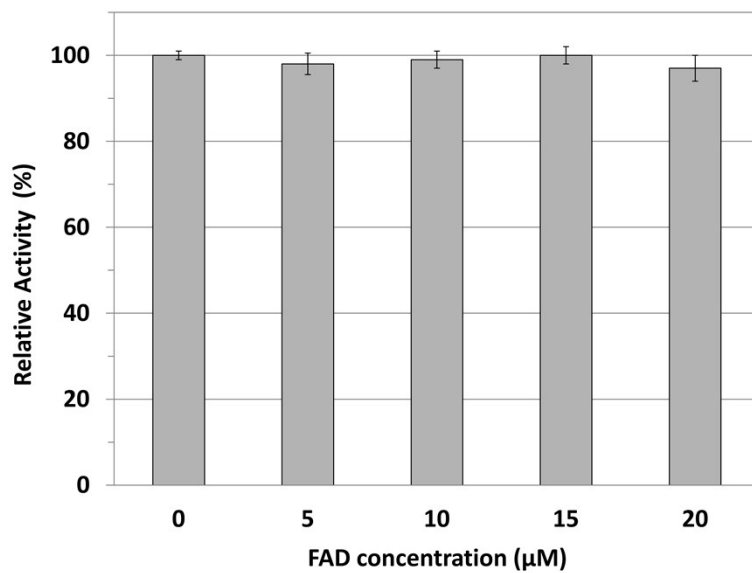


Figure S4: Effect of external concentration of FAD on the enzyme's activity. The highest activity of the pm1 ($1.83 \mu\text{mol}\cdot\text{min}^{-1}\cdot\text{mg}^{-1}$) was defined as 100%.

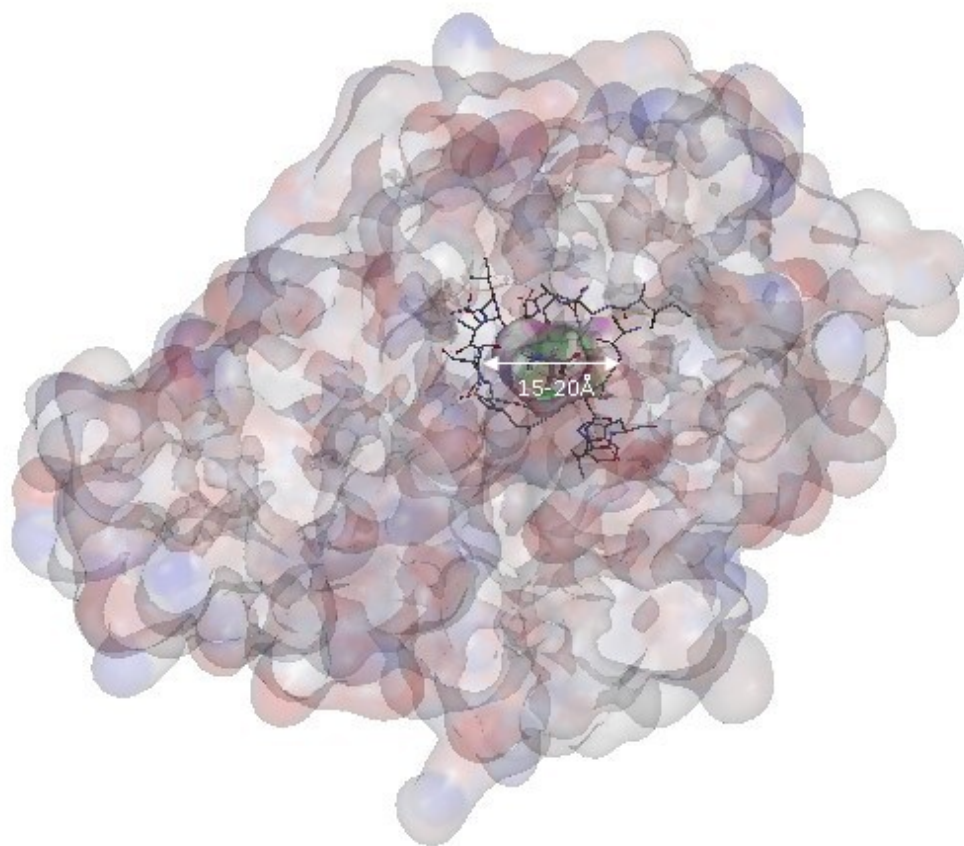


Figure S5: Substrate entrance site in pm1 (surface representation).

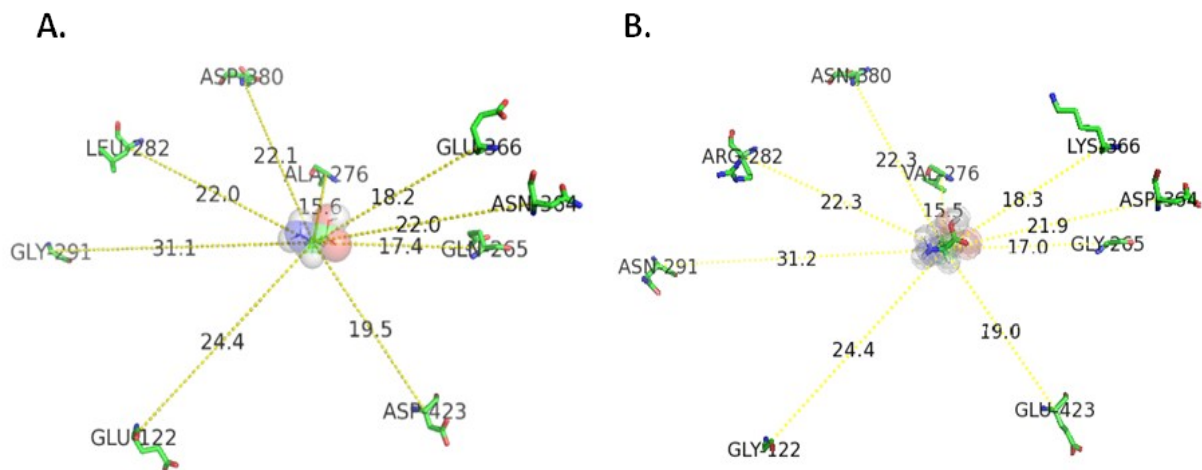


Figure S6: Molecular distances of alanine (substrate) from the mutant sites. **A.** in the wild type pm1; **B.** in the mutant pm1ep3.

Table S1: Primers used in this study

Primer name	Nucleotide sequence (5'-3')
	Primer used for epPCR
Pm1_F1	CGCGGATCCATGGCAATAAGTAGAAGAAAATTTA
Pm1_R1	TCCGAGCTCTTAGAAACGATACAGACTAAATGGT
	Primer used for genes knockout and verification
<i>cycA</i> -knockF	ATGGTAGATCAGGTAAAAGTCGTTGCCGATGATCAGGCTCCGGCTGAACAATTCCGGGGATCCGTCGAC C
<i>cycA</i> -knockR	TTATTTCCGCAGTTCAGCAGCCCGCTTCTTACCAATAAACAACCAGCCCATGTAGGCTGGAGCTGCTTCG
<i>cycA</i> -verF	GCCTGAACAACACAGACAGGTACAGGAAGA
<i>cycA</i> -verR	CTGGATGGCATTGCGCCATCCAGCATGATA
<i>amaP</i> -knockF	ATGATGAACACGGAAGGTAATAACGGTAACAAACCTCTCGGTCTATGGAAATTCCGGGGATCCGTCGAC C
<i>amaP</i> -knockR	TTATACGGTTTTATTGCGCTTCATGACCATTGCCACAATAAGGCTGAGTATGTAGGCTGGAGCTGCTTCG
<i>amaP</i> -verF	TCGGTCGCTAAGCAACTCGGCTATAACGTG
<i>amaP</i> -verR	ACCGCCACCACAATAATACAGGAAGTACTG
<i>lldP</i> -knockF	ATGAATCTCTGGCAACAAAACACTACGATCCC GCCGGGAATATCTGGCTTTCATTCCGGGGATCCGTCGACC
<i>lldP</i> -knockR	TTAAGGAATCATCCACGTTAAGACATAAGCCTGAAGCGTGGTGATCACGCTGTAGGCTGGAGCTGCTTCG
<i>lldP</i> -verF	CATTACACGATGTGCGTGGACTCCAGGAGA
<i>lldP</i> -verR	CGGCAACCTCGTCTGACAGGCGTCTGGGTA

Table S2: Thermodynamic parameter for L-alanine binding to wild type and its mutants.

Enzyme	K_d (mM)	ΔH° (kcal.mol ⁻¹)	ΔG° (kcal.mol ⁻¹)
Wild type (Pm1)	15.21 ± 0.34	-9.53 ± 0.15	-7.08 ± 0.07
Pm1ep1	11.93 ± 0.12	-11.09 ± 0.21	-6.45 ± 0.11
Pm1ep2	9.15 ± 0.11	-12.14 ± 0.08	-5.76 ± 0.08
Pm1ep3	6.76 ± 0.09	-13.27 ± 0.14	-4.97 ± 0.03

K_d and ΔH° were obtained from fit to a binding isotherm, while ΔG° was calculated from $\Delta G^\circ = RT \ln K_d$. Each value represents the mean of three or more independent ITC experiments.

Table S3: K_m values of different amino acid deaminases/oxidases.

Enzyme's name	Origin	Substrate	K_m (mM)	References
L-amino acid deaminase	<i>Proteus mirabilis</i>	L-Phenylalanine	31.55	S5
L-amino acid deaminase	<i>Proteus mirabilis</i>	L-Phenylalanine	26.2	S1
L-amino acid deaminase	<i>Proteus myxofaciens</i>	L-Phenylalanine	2.28	S2
L-Amino Acid Oxidases	<i>Proteus rettgeri</i>	L-Lysine	23.2	S3
L-amino acid deaminase	<i>Proteus vulgaris</i>	L-Methionine	305.0	S4

S1: Y. Hou, G. S. Hossain, J. Li, H. D. Shin, L. Liu, and G. Du, *Appl. Microbiol. Biotechnol.*, 2015, **99**, 8391-8402. S2: D.

P. Pantaleone, A. M. Geller, and P. P. Taylor, *J. Mol. Catal. B: Enzymol.*, 2001, **11**, 795–803. S3: J. A. Duerre, and S.

Chakrabarty, *J. Bacteriol.*, 1975, **121**, 656-663. S4: G. S. Hossain, J. Li, H. D. Shin, G. Du, M. Wang, L. Liu, and J.

Chen, *PLoS One*, 2014, **9**, e114291.