

## Electronic Supplementary Information (ESI)

### Environmentally sensitive nanohydrogels decorated with three-strand oligonucleotide helix for controlled loading and prolonged release of intercalators

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### Formulas used in Electrochemical Impedance Spectroscopy (EIS) analysis

The impedance of the CPE parameter in the Ershler – Randles model applied in fitting calculated data to experimental EIS points is described by Eq.(1):

$$Z_{CPE} = T^{-1}(j\omega)^{-\phi} \quad (1)$$

where,  $\omega$  is angular frequency,  $j = (-1)^{1/2}$ ,  $T$  is capacitive coefficient, and  $\phi$  is the exponent value.

The average double layer capacitance,  $C_{dl}$  is combined with the capacitive coefficient  $T$  and can be calculated according to Eq. (2):

$$T = C_{dl}^{\phi}(R_s^{-1}R_{CT}^{-1})^{1-\phi} \quad (2)$$

where  $R_s$  and  $R_{CT}$  are the values of solution resistance and the resistance of the charge transfer.

As the semi-infinite diffusion of 1-electron simple redox species ( $[\text{Fe}(\text{CN})_6]^{3-/4-}$ ) takes place, the mass transfer resistance (Warburg impedance,  $W$ ), visible in EIS plots as a linear part (see Fig.5 of ms.), can be estimated from Eq.(3):

$$W = \sigma \omega^{1/2} (1 - j) \quad (3)$$

where the Warburg parameter,  $\sigma$ , is described as (Eq.(4)):

$$\sigma = \frac{RT}{n^2 F^2 \sqrt{2}} \left[ \frac{1}{\sqrt{D_o}} \frac{1}{C_o} + \frac{1}{\sqrt{D_R}} \frac{1}{C_R} \right] \quad (4)$$

and  $D_o$  and  $C_o$  are diffusion coefficient and concentration of oxidized form, respectively, and  $D_R$  and  $C_R$  are diffusion coefficient and concentration of the reduced form of the redox species.

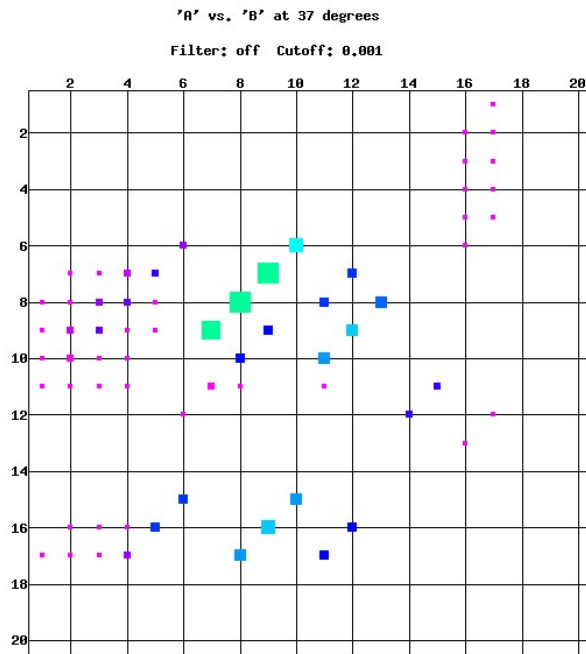
The electron transfer-rate constant,  $k_0$ , can be determined using Eq. (5):

$$k_0 = \left( \frac{\sigma}{R_{CT}} \right) / \frac{2\xi^\alpha}{2D_{ox}} \quad (5)$$

where  $k_0$  is electron-transfer rate constant,  $\xi = (\sqrt{D_{Ox}/D_{Red}})$  (for 1-electron, fast and reversible electrode process of  $[\text{Fe}(\text{CN})_6]^{3-/4-} \approx 1$ ),  $\alpha$  is transfer coefficient and is assumed to be equal to 0.5,  $D_{ox}$  is the diffusion coefficient of  $[\text{Fe}(\text{CN})_6]^{3-/4-}$  that was taken from <sup>1</sup> as  $0.896 \times 10^{-5} \text{ cm}^2\text{s}^{-1}$  and corrected to the value of  $0.726 \times 10^{-5} \text{ cm}^2\text{s}^{-1}$ , as the diffusion coefficient of  $[\text{Fe}(\text{CN})_6]^{3-/4-}$  has 19% lower values in the PNIPA gel environment compared to aqueous conditions <sup>2</sup>.

# 1. Figures

- Oligo 1 5' Acryd-GGGGG-GCTCTTGGAACT 3'
- Oligo 2 5' Acryd GGGGG-TGAGTAGACT 3'



Range	Color
$0.000 \leq P \leq 0.010$	#FF00FF
$0.010 < P < 0.012$	#CC00FF
$0.012 \leq P < 0.017$	#9900FF
$0.017 \leq P < 0.023$	#6600FF
$0.023 \leq P < 0.033$	#3300FF
$0.033 \leq P < 0.046$	#0000FF
$0.046 \leq P < 0.065$	#0033FF
$0.065 \leq P < 0.091$	#0066FF
$0.091 \leq P < 0.128$	#0099FF
$0.128 \leq P < 0.180$	#00CCFF
$0.180 \leq P < 0.253$	#00FFFF
$0.253 \leq P < 0.356$	#00FFCC
$0.356 \leq P < 0.5$	#00FF99
$0.500 \leq P < 0.644$	#00FF66
$0.644 \leq P < 0.747$	#00FF33
$0.747 \leq P < 0.820$	#00FF00
$0.820 \leq P < 0.872$	#33FF00
$0.872 \leq P < 0.909$	#66FF00
$0.909 \leq P < 0.935$	#99FF00
$0.935 \leq P < 0.954$	#CCFF00
$0.954 \leq P < 0.967$	#FFFF00
$0.967 \leq P < 0.977$	#FFCC00
$0.977 \leq P < 0.983$	#FF9900
$0.983 \leq P < 0.988$	#FF6600
$0.988 \leq P < 0.990$	#FF3300
$0.990 \leq P \leq 1.000$	#FF0000

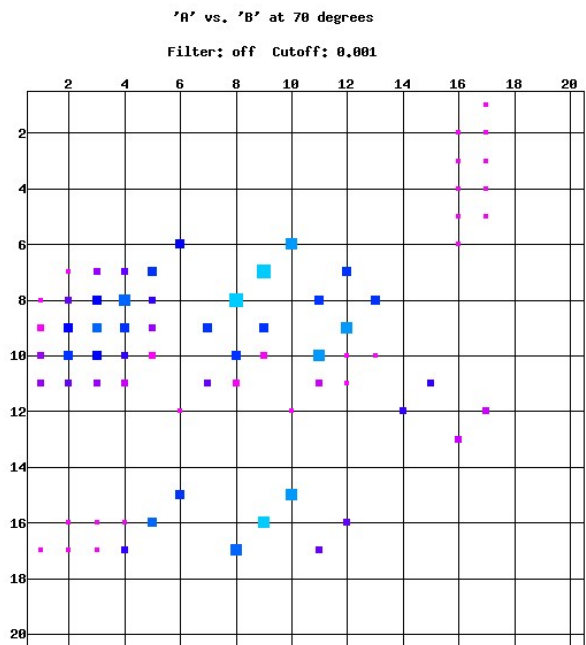


Fig. 1S Simulation of the ability of oligo1 and oligo2 strands for additional selfhybridization.

- Oligo 1-2 5' Acryd GGGGG-TGAGTAGACACTGCTCTTGGAACCT-GGGGG Acryd-3'
- Oligo 3 3' ACTCATCTGTGACGAGAACCTTGA 5'

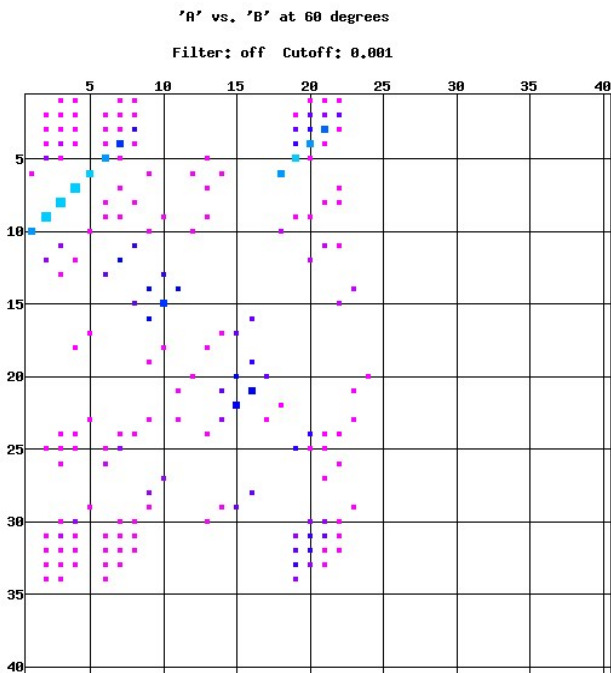
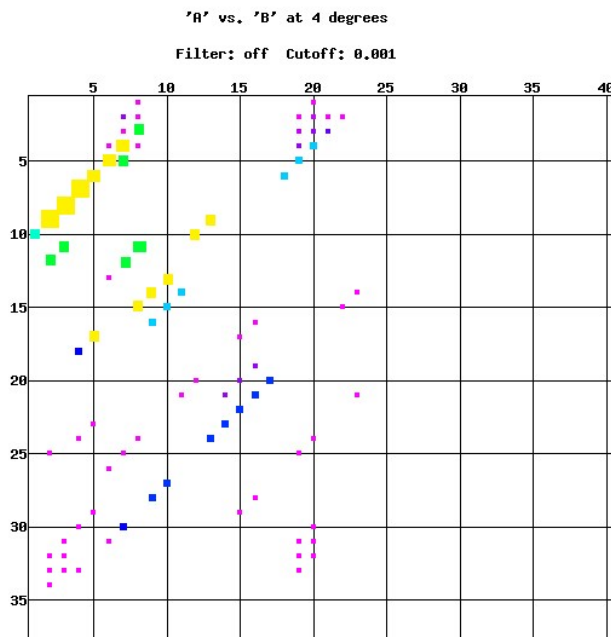
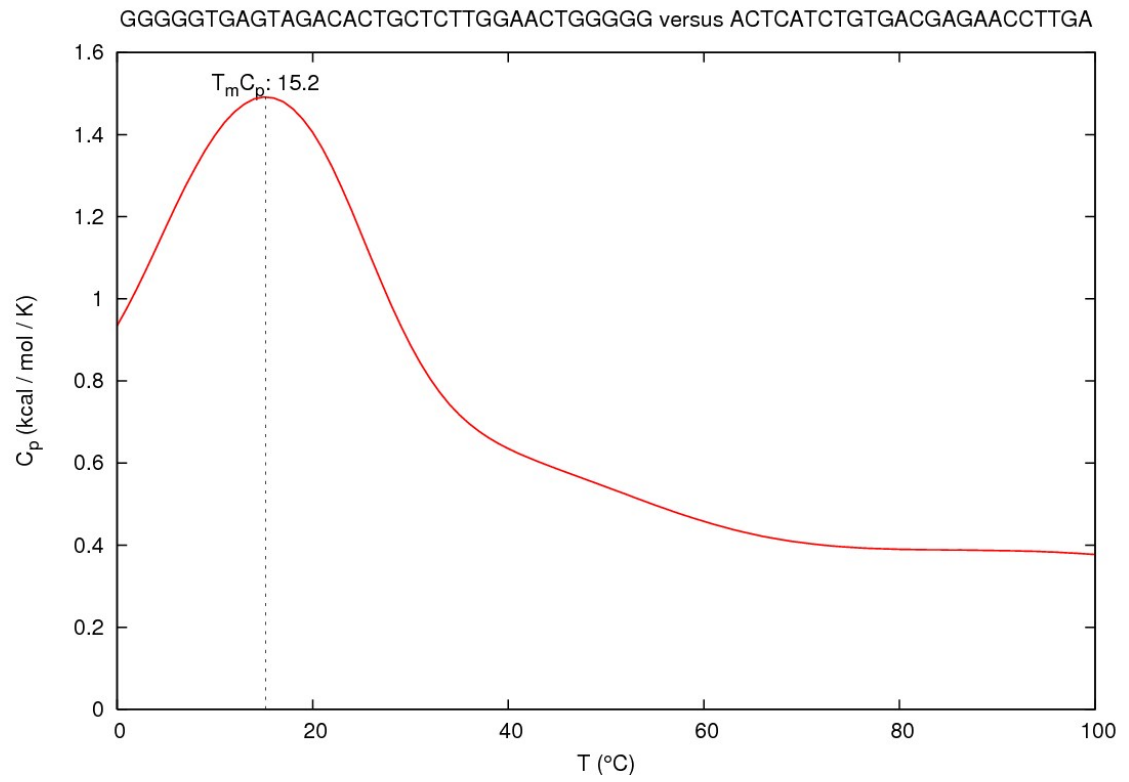


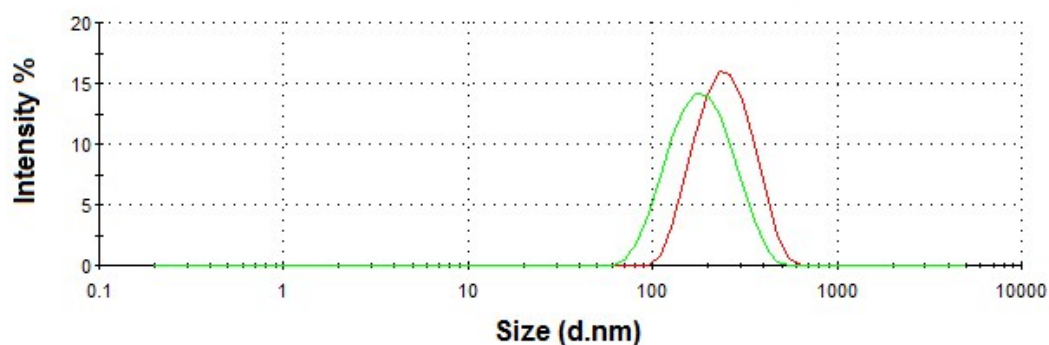
Fig. 2S Simulation of the ability of oligo1-2 and oligo3 strands for selfhybridization.



**Fig. 3S** Simulation of the  $T_m$  and  $C_p$  of oligo1-2-3 tri-segment hybrid.

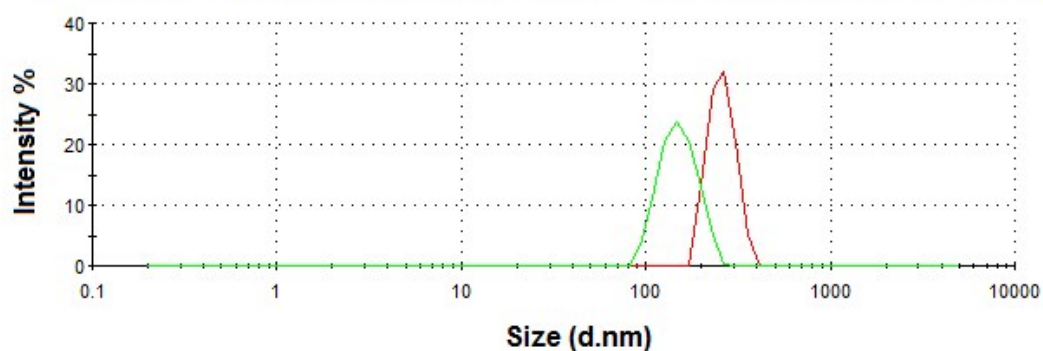
A)

Size distribution of PNIPA-co-AAc NPs by Intensity at 37 and 45°C



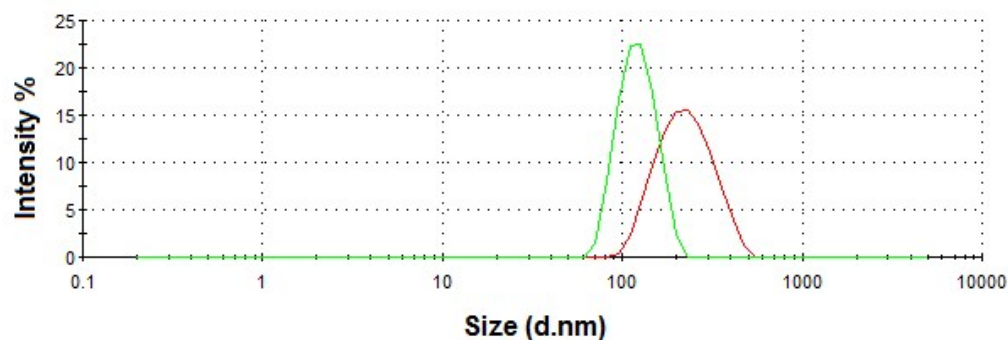
B)

Size distribution of PNIPA-co-AAc-oligo1-2 NPs by Intensity at 37 and 45°C



C)

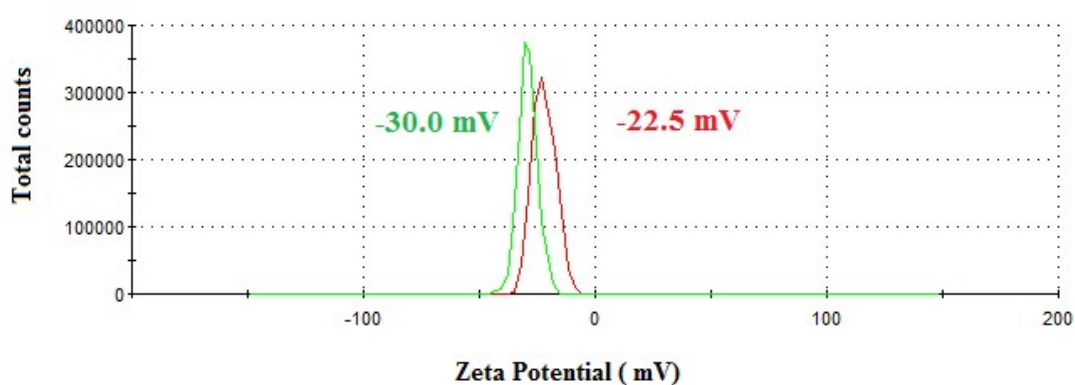
Size distribution of PNIPA-co-AAc-oligo1-2-3 NPs by Intensity at 37 and 45°C



**Fig. 4S** Sizes of PNIPA-co-AAc- (A), PNIPA-co-AAc-oligo1-2- (B) and PNIPA-AAc-oligo1-2-3 nanogels (C) obtained by DLS at 37 and 45 °C, respectively.

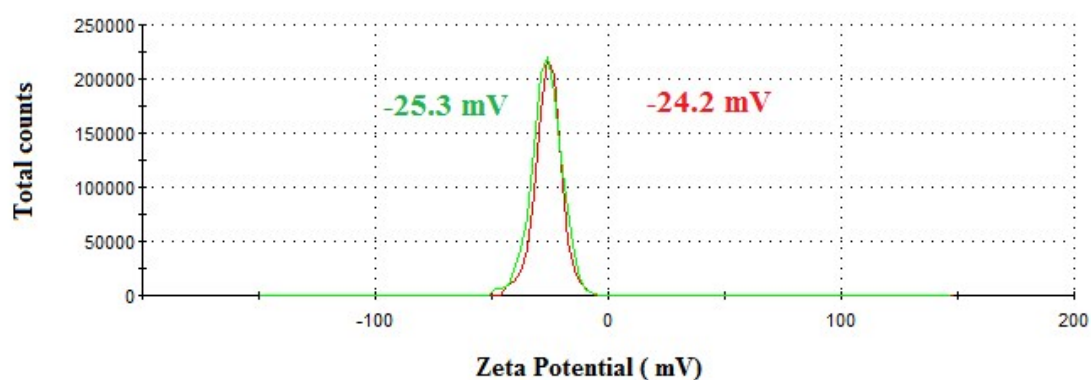
A)

Zeta Potential distribution of PNIPA-co-AAc NPs at 37 and 45°C



B)

Zeta Potential distribution of PNIPA-co-AAc-oligo1-2 NPs at 37 and 45°C



C)

Zeta Potential distribution of PNIPA-co-AAc-oligo1-2-3 NPs at 37 and 45°C

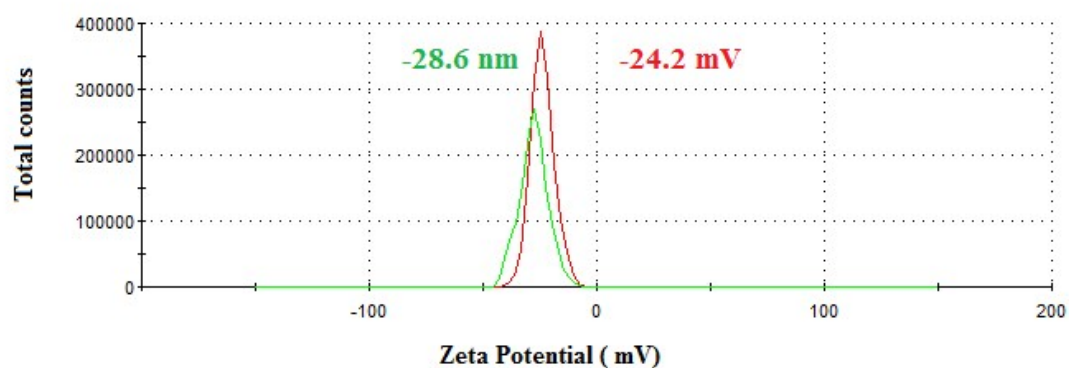
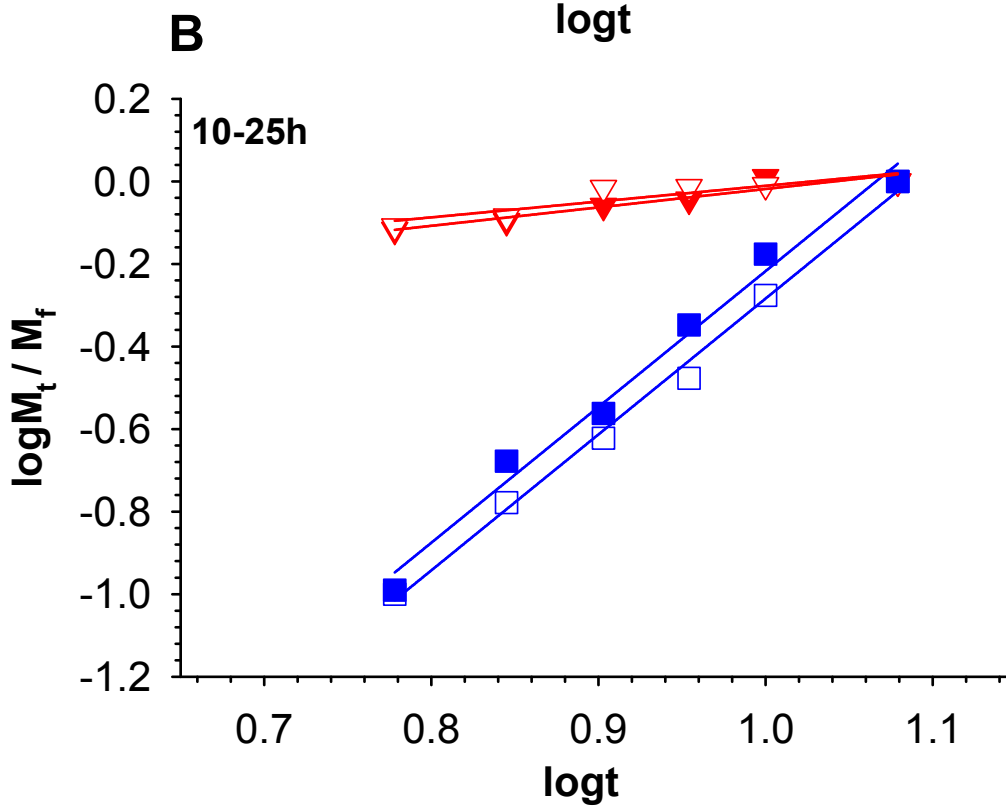
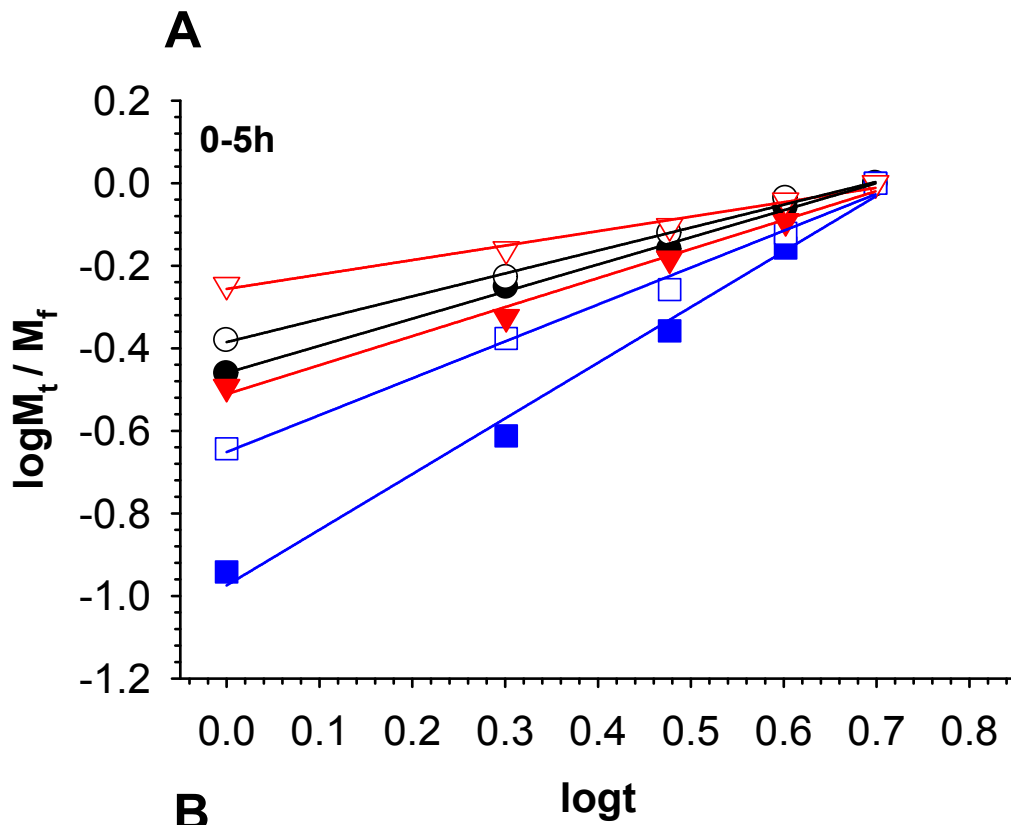
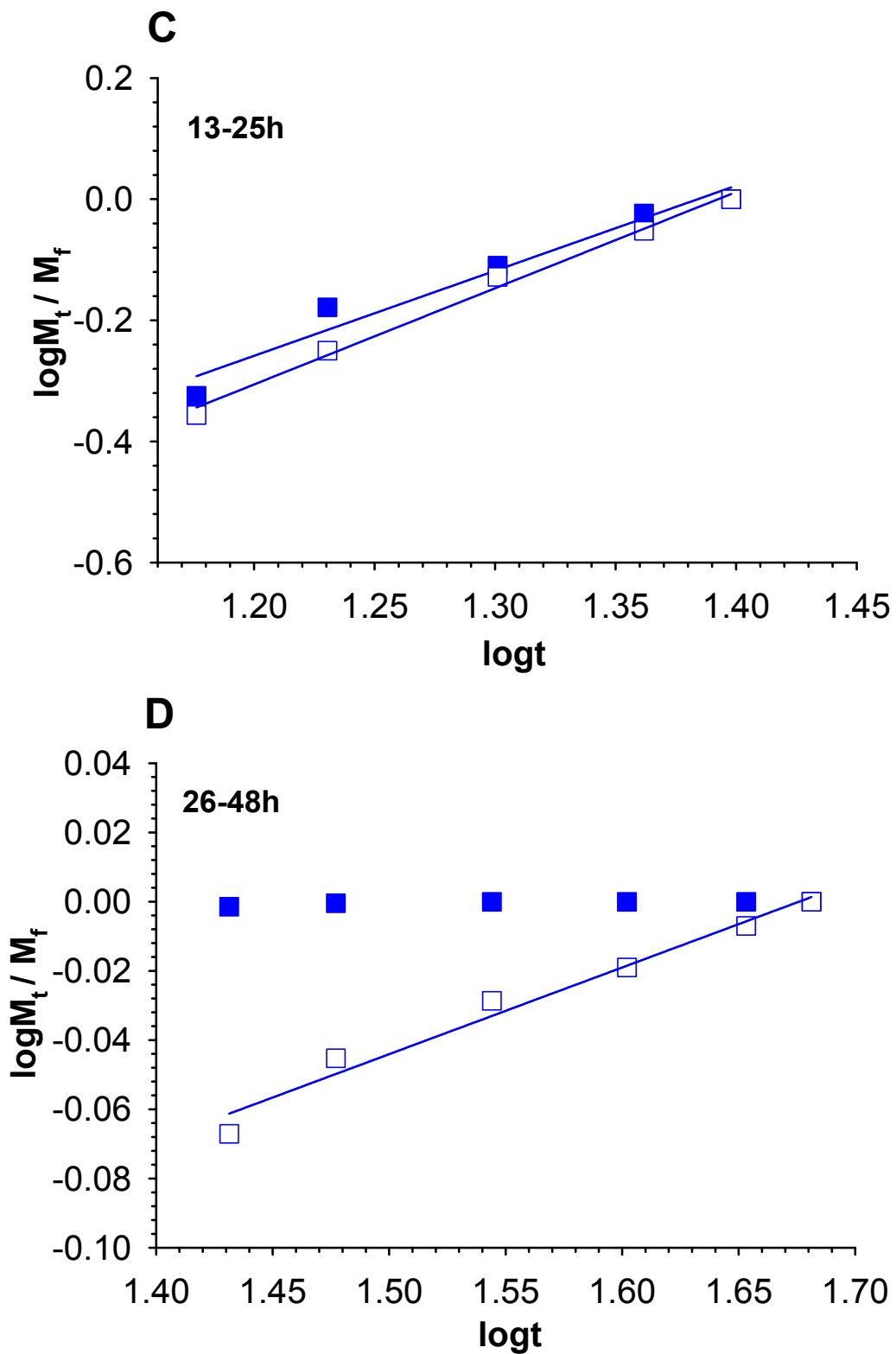


Fig. 5S Zeta potentials of PNIPA-co-AAc- (A), PNIPA-co-AAc-oligo1-2- (B) and PNIPA-AAc-oligo1-2-3 nanogels (C) recorded by DLS at 37 and 45°C, respectively.







**Fig. 6S** Plots of  $\log (M_f/M_t)$  vs.  $\log t$  constructed according to Peppas model in selected ranges of time (A-D) for the release of doxorubicin from: PNIPA-co-AAc- (black circles), PNIPA-co-AAc-oligo1-2- (red triangles) and PNIPA-AAc-oligo1-2-3 nanogels (blue squares). Temperature: 37 °C (filled symbols), 45°C (empty symbols).

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<sup>1</sup> D. Lide, H. Frederikse, Handbook of Chemistry and Physics, CRC Press, New York, 2007.

<sup>2</sup> M. Karbarz, M. Gniadek, Z. Stojek, One dimensional volume-phase transition of N-isopropylacrylamide gels on the surface of gold electrodes, *Electroanal.*, 2005, **17**, 1396-1400.