Tweaking of supramoleculargelation properties of a dipeptide based ambidextrous organogelator through a cooperative influence of hydrophobicity, steric bulk and conformational flexibility of side chain residue of a single hydrophobic α-amino acid encrypted on a designed molecular frame

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Scheme 1:



Synthesis of N-Succinimidylpalmitate (1): To stirred suspension of N-Hydroxysuccinimide (5.0 g, 43mmole, 1eqv) in 40 ml of THF:CHCl₃mixture (1:1) and Triethylamine (9 ml, 64.6mmole, 1.5eqv); was added drop wise a solution of Palmitoyl Chloride (12.5 ml, 43mmole, 1eqv) in 20 ml of THF:CHCl₃ mixture (1:1) and stirred for overnight at room temperature. Solid precipitate was filtered and the filtrate was diluted with 25 ml of CHCl₃ and washed sequentially with ice cold 1 (N) HCl (25 ml×2), saturated brine solution (25 ml × 2).Organic layer was collected and dried over anhyd. Na₂SO₄ and evaporated under reduced pressure which was then recrystallized from 25mlofisopropanol to get the desired product (13.8 g;90.8% yield). This compound was used for the next step without any further purification.

Synthesis of N-hexadecanoyl-alanine (2a): To a solution of N-Succinimidylpalmitate (3.0 g, 8.4mmole, 1eqv.), Alanine (0.897 g, 10.08mmole, 1.2eqv.) in 40 ml of 65% Ethanol; Triethylamine (2.5 ml, 17.9mmole, 2.1eqv.) was added and put to reflux ($60-80^{\circ}C$) for 20hrs. Then the reaction mixture was concentrated under reduced pressure and taken up in 25 ml Ethyl acetate and washed sequentially with ice cold 1 (N) HCl (30 ml× 2), saturated brine solution (25 ml×2).Organic layer was collected and dried over anhyd. Na₂SO₄ and evaporated under reduced pressure to give the desired product (2.6 g;94.5% yield; m.p=94^oC). This compound was used for the next step without any further purification.

Synthesis of N-hexadecanoyl-valine (2b): To a solution of N-Succinimidylpalmitate (3.0 g, 8.4mmole, 1eqv.), Valine (1.2 g, 10.08mmole, 1.2eqv.) in 40 ml of 65% Ethanol; Triethylamine (3 ml, 21.56mmole, 2.5eqv.) was added and put to reflux ($60-80^{\circ}$ C) for 20hrs. Then the reaction mixture was concentrated under reduced pressure and taken up in 25 ml Ethyl acetate and washed sequentially with ice cold 1 (N) HCl (30 ml×2), saturated brine solution (25 ml× 2).Organic layer was collected and dried over anhyd. Na₂SO₄ and evaporated under reduced pressure to give the desired product (2.867 g;95.95% yield; m.p=98^oC). This compound was used for the next step without any further purification.

Synthesis of N-hexadecanoyl-isoleucine (2c): To a solution of N-Succinimidylpalmitate (1.6 g, 4.526mmole, 1eqv.), Isoleucine (0.713 g, 5.44 mmole, 1.2eqv.) in 25 ml of 65% Ethanol; Triethylamine (1.3 ml, 9.34mmole, 2.1eqv.) was added and put to reflux ($60-80^{\circ}$ C) for 20hrs. Then the reaction mixture was concentrated under reduced pressure and taken up in 25 ml Ethyl acetate and washed sequentially with ice cold 1 (N) HCl (30 ml×2), saturated brine solution (25 ml× 2).Organic layer was collected and dried over anhyd. Na₂SO₄ and evaporated under reduced pressure to give the desired product (1.572 g;94% yield). This compound was used for the next step without any further purification.

4.63Synthesis of N-hexadecanoyl-leucine (2d): To a solution of N-Succinimidylpalmitate (2.0 g, 5.6mmole, 1eqv.), Leucine (0.885 g, 6.7mmole, 1.2eqv.) in 25 ml of 65% Ethanol; Triethylamine (1.6 ml, 11.5mmole, 2.05eqv.) was added and put to reflux (60-80°C) for 20hrs. Then the reaction mixture was concentrated under reduced pressure and taken up in 25 ml Ethyl acetate and washed sequentially with ice cold 1 (N) HCl (30 ml× 2), saturated brine solution (25 ml×2).Organic layer was collected and dried over anhyd. Na₂SO₄ and evaporated under reduced pressure to give the desired product (1.48 g;71.4% yield). This compound was used for the next step without any further purification.

Synthesis of N-hexadecanoyl-phenylalanine (2e): To a solution of N-Succinimidylpalmitate (2.0 g, 5.5mmole, 1eqv.), Phenyl alanine (1.11 g, 6.7mmole, 1.2eqv.) in 30 ml of 65% Ethanol; Triethylamine (1.6 ml, 11.5mmole, 2.05eqv.) was added and put to reflux (60-80°C) for °20hrs. Then the reaction mixture was concentrated under reduced pressure and taken up in 25 ml Ethyl acetate and washed sequentially with ice cold 1 (N) HCl (30 ml×2), saturated brine solution (25 ml×2).Organic layer was collected and dried over anhyd Na₂SO₄ and evaporated under reduced pressure to give the desired product (1.6 g; 70.35% yield). This compound was used for the next step without any further purification.

Synthesis of N-hexadecanoyl-glycine (2f): To a solution of N-Succinimidylpalmitate (0.6 g, 1.7 mmole, 1 eqv.), glycine (1.0 g, 2.1 mmole, 1.2eqv.) in 30 ml of 65% Ethanol; Triethylamine (0.6 ml, 4.2 mmole, 2.05 eqv.) was added and put to reflux (60-80°C) for °20hrs. Then the reaction mixture was concentrated under reduced pressure and taken up in 25 ml Ethyl acetate and washed sequentially with ice cold 1 (N) HCl (30 ml×2), saturated brine solution (25 ml×2).Organic layer was collected and dried over anhyd Na₂SO₄ and evaporated under reduced pressure to give the desired product (0.5 g; 75% yield). This compound was used for the next step without any further purification.

Synthesis of PTSA-salt of β -Alanine benzyl ester (3): In 100 ml of benzene a mixture of β -Alanine (5 g, 56.12mmol), PTSA.H2O (11.7419 g, 64.36mmol) and benzyl alcohol (11.642ml, 112.29mmol) was taken. After refluxing in Dean-Stark apparatus for 24hrs, 40 ml of diethyl ether was added to it with immediate precipitation of PTSA-salt of β -Alanine benzyl ester. The salt was then filtered, washed with 10 ml of diethyl ether and was dried under reduced pressure. This compound was used for the next step without any further purification.

Synthesis of 4 (a): To a solution of N-hexadecanoyl-alanine (0.500 g, 1.5mmole, 1eqv.) in 5 ml of THF, HOBt (0.304 g, 2.2mmole, 1.5eqv.) was added and kept on stirring in ice water bath at 0°C.After 10mins of stirring, DMAP (0.018 g, 0.15mmole, 0.093eqv.) and solution of DCC (0.618 g, 2.9mmole,1.93eqv.) in 5 ml of CHCl₃ was added to it and kept stirring. Finally, after 15mins, K_2CO_3 (0.311 g, 2.2mmole, 1.47eqv) and PTSA-salt of β -Alanine benzyl ester (0.632 g, 1.8mmole, 1.2eqv.) was added and kept on stirringovernight. The reaction mixture was then concentrated under reduced pressure and taken up in 30 ml CHCl₃ and washed sequentially with ice cold 1 (N) HCl (25 ml×2), saturated brine (25 ml×2). The organic layer was collected and dried over anhyd. Na₂SO₄and then purified by silica column chromatography (60% EtOAc/Petroleum ether) to get the desired product. (0.532 g, 72% yield).

¹HNMR (300MHz, CDCl₃) : δ ::0.85-0.87 (t, -NHCOCH₂CH₂ (CH₂)₁₂CH₃, 3H), 1.24-1.32 (t, -NHCOCH₂CH₂(CH₂)₁₂CH₃, 24H), 1.59-1.65 (s, -NHCOCH(CH₃)NHCOCH₂CH₂ (CH₂)₁₂CH₃, 3H), 1.90 (s, -NHCOCH₂CH₂ (CH₂)₁₂CH₃, 2H), 2.1-2.2 (t, -NHCOCH₂CH₂ (CH₂)₁₂CH₃, 2H), 2.5-2.6 (t, -CONHCH₂CH₂COOCH₂Ph, 2H), 3.50-3.54 (t, -CONHCH₂CH₂COOCH₂Ph, 2H), 3.50-3.54 (t, -CONHCH₂CH₂COOCH₂Ph, 2H), 3.50-3.54 (t, -CONHCH₂CH₂CH₂COOCH₂Ph, 2H), 6.11-6.13 (d, -CONHCH₂CH₂COOCH₂Ph, 1H), 6.59-6.62 (s, -NHCOCH₂CH₂ (CH₂)₁₂CH₃, 1H), 7.26-7.38 (bs, aromatic, 5H). ¹³C NMR(75 MHz, CDCl₃) : $\delta_{\rm C}$: 173.27, 172.93, 171.407, 135.6, 128.62, 128.4, 128.28, 77.46, 77.041, 76.61, 66.59, 49.23, 36.7, 34.976, 33.99, 33.69, 31.9, 31.2, 29.63, 29.49, 29.33, 29.28, 25.75, 25.54, 25.544, 24.865, 24.77, 22.67, 19.13, 18.22, 14.09.IR (KBr) v: 3849.86, 3685.76, 3304.68, 2918.91, 2850.13, 1732.23, 1633.84, 1538.08, 1454.64, 1244.43, 1044.03, 882.92, 695.39.ESI-MS (m/z) $C_{29}H_{48}N_2O_4$ (EXACT MASS=488.36)511.34 (100%, [M + Na]⁺), 527.31(12%, [M + K]⁺).

Synthesis of 4 (b):To a solution of N-hexadecanoyl-valine (0.500 g, 1.41mmole, 1eqv.) in 5 ml of THF, HOBt (0.285 g, 2.11mmole, 1.5 eqv) was added and kept on stirring in ice water bath at 0^{0} C.After 10mins of stirring, DMAP (0.017 g, 0.14mmole, 0.1eqv.) and solution of DCC (0.561 g, 2.72mmole, 1.93 eqv) in 5 ml of CHCl₃ was added to it and kept stirring. Finally, after 15mins, K₂CO₃ (0.286 g, 2.07mmole, 1.47eqv.) and PTSA-salt of β -Alanine benzyl ester (0.595 g, 1.69mmole, 1.2eqv.) was added and kept on stirringovernight. Thereaction mixture was then concentrated under reduced pressure and taken up in 30 ml CHCl₃ and washed sequentially with ice cold 1 (N) HCl (25 ml× 2), saturated brine (25 ml × 2). The organic layer was collected and dried over anhyd. Na₂SO₄and then purified by silica column chromatography (40%EtOAc/Petroleum ether) to get the desired product. (0.525 g, 72% yield).

¹HNMR (300 MHz, CDCl₃) : δ :: 0.875-0.918 (m, -NHCOCH₂CH₂ (CH₂)₁₂C<u>*H*₃ and >CHCH (C<u>*H*</u>₃)₂, 9H), 1.27 (bs, -NHCOCH₂CH₂(C<u>*H*</u>₂)₁₂CH₃, 24H), 1.58-1.65 (m, -NHCOCH₂C<u>*H*</u>₂ (CH₂)₁₂CH₃, >CHC<u>*H*</u> (CH₃)₂, 3H), 2.11-2.24 (t, -NHCOC<u>*H*</u>₂CH₂(CH₂)₁₂CH₃, 2H), 2.59-2.62 (t, -CONHCH₂C<u>*H*</u>₂COOCH₂Ph, 2H), 3.5-3.6 (m, CONHC<u>*H*</u>₂CH₂COOCH₂Ph, 2H), 4.17-4.2 (t, >C<u>*H*</u>CH (CH₃)₂, 1H), 5.09-5.1 (s, -CONHCH₂CH₂COOC<u>*H*</u>₂Ph, 2H), 6.11-6.13 (d, -N<u>*H*</u>COCH₂CH₂ (CH₂)₁₂CH₃), 1H), 6.52 (s, -CON<u>*H*</u>CH₂CH₂COOC<u>*H*</u>2Ph, 1H), 7.26-7.35 (bs, aromatic, 5H). ¹³C NMR(75 MHz, CDCl₃) : δ_{C} : 173.6, 172.4, 171.8, 135.64, 128.6, 128.34, 128.18, 77.47, 77.05, 76.62, 66.53, 57.53, 41.35, 36.53, 35.02, 33.9, 33.47, 31.9, 29.67, 29.64, 29.61, 29.5, 29.33, 29.26, 25.64, 24.81, 22.82, 22.66, 22.20, 14.08. IR (KBr) v: 3295.17, 2918.06, 2849.75, 1737.69, 1637.40, 1557.89, 1461.89, 1271.73, 1187.97, 1098.5, 1007.4, 940.48, 885.17, 697.04.ESI-MS (m/z) C₃₁H₅₂N₂O₄ (EXACT MASS=516.37) 517.379 (100%, [M + H]⁺), 539.37 (20%, [M + Na]⁺).</u>

Synthesis of 4 (c): To a solution of N-hexadecanoyl-isoleucine (1.5 g, 4.06mmole, 1eqv.) in 5 ml of THF,HOBt (0.822 g,6.09 mmole,1.5eqv.) was added and kept on stirring in ice water bath at 0^{0} C.After 10mins of stirring, DMAP (0.045 g,0.373mmole, 0.092eqv.) and solution of DCC (1.7 g 8.241mmole, 2.03 eqv.) in 5 ml of CHCl₃ was added to it and kept stirring. Finally, after 15mins, K₂CO₃ (0.823 g, 5.968mmole, 1.47eqv.) and PTSA-salt of β -Alanine benzyl ester (1.7 g, 4.87mmole, 1.2eqv.) was added and kept on stirringovernight. The reaction mixture was then concentrated under reduced pressure and taken up in 30 ml CHCl₃ and washed sequentially with ice cold 1 (N) HCl (25 ml× 2), saturated brine (25 ml × 2). The organic layer was collected and dried over anhyd. Na₂SO₄and then purified by silica column chromatography (45%EtOAc/Petroleum ether) to get the desired product. (0.532 g, 72% yield).

¹HNMR (300MHz, CDCl₃) : δ ::0.84-0.895 (m, -NHCOCH₂CH₂ (CH₂)₁₂CH₃ and >CHCH (CH₃)CH₂CH₃, 9H), 1.2-1.38 (bs, -NHCOCH₂CH₂ (CH₂)₁₂CH₃) and >CHCH (CH₃)CH₂CH₃, 26H), 1.55-1.61 (s, -NHCOCH₂CH₂ (CH₂)₁₂CH₃ and >CHCH (CH₃)CH₂CH₃, 3H), 2.09-2.3 (m, -NHCOCH₂CH₂ (CH₂)₁₂CH₃, 2H), 2.56-2.60 (t, -CONHCH₂CH₂COOCH₂Ph, 2H), 3.4-3.58 (q, CONHCH₂CH₂COOCH₂Ph, 2H), 4.17-4.22 (t, >CHCH (CH₃)CH₂CH₃, 1H), 5.09-5.13 (bs, -CONHCH₂CH₂COOCH₂Ph, 2H), 5.9-6.15 (d, -NHCOCH₂CH₂ (CH₂)₁₂CH₃, 1H), 6.3-6.35 (s, CONHCH₂CH₂COOCH₂Ph, 1H), 7.3-7.35 (bs, aromatic, 5H). ¹³C NMR(75 MHz, CDCl₃) : $\delta_{\rm C}$: 173.1, 171.9, 171.4, 135.5, 128.6, 128.4, 128.3, 128.2, 128.18, 77.46, 77.05, 76.62, 66.59, 66.5, 57.47, 49.45, 37.37, 36.68, 34.96, 33.9, 33.66, 31.9, 29.67, 29.64, 29.5, 29.33, 29.27, 25.71, 25.02, 24.84, 22.67, 15.33, 14.08, 11.22.IR (KBr) v: 3291, 2918.06,

2849.88, 1732.99, 1633.86, 1556.75, 1463.94, 1170.12, 1018.61, 952.44, 883, 697.02.ESI-MS (m/z) $C_{32}H_{54}N_2O_4$ (EXACT MASS= 530.40)553.4 (100%, [M + Na]⁺).

Synthesis of 4 (d):To a solution of N-hexadecanoyl-leucine (1.38 g, 3.7mmole, 1eqv.) in 5 ml of THF, HOBt (0.749 g, 5.55mmole, 1.5 eqv.) was added and kept on stirring in ice water bath at 0^{0} C.After 10mins of stirring, DMAP (0.042 g, 0.34mmole, 0.092eqv.) and solution ofDCC (1.54 g, 7.511mmole, 2.03 eqv) in 5 ml of CHCl₃ was added to it and kept stirring. Finally, after 15mins, K₂CO₃ (1.751 g, 5.439mmole, 1.47eqv.) and PTSA-salt of β -Alanine benzyl ester (1.57 g, 4.44mmole, 1.2eqv.) was added and kept on stirringovernight. The reaction mixture was then concentrated under reduced pressure and taken up in 30 ml CHCl₃ and washed sequentially with ice cold 1 (N) HCl (25 ml × 2), saturated brine (25 ml × 2).The organic layer was collected and dried over anhyd. Na₂SO₄and then purified by silica column chromatography (55% EtOAc/Petroleum ether) to get the desired product. (1.378 g, 70% yield).

¹HNMR (300MHz, CDCl₃) : δ ::0.85-0.911 (q, -NHCOCH₂CH₂ (CH₂)₁₂CH₃and >CHCH₂CH (CH₃)₂, 9H), 1.15-1.25 (bs, -NHCOCH₂CH₂ (CH₂)₁₂CH₃, 24H), 1.55-1.65 (d, -NHCOCH₂CH₂ (CH₂)₁₂CH₃and >CHCH₂CH (CH₃)₂, 5H), 2.08-2.1 (t, -NHCOCH₂CH₂ (CH₂)₁₂CH₃, 2H), 2.55-2.6 (q, -CONHCH₂CH₂COOCH₂Ph, 2H), 3.49-3.55 (q, -CONHCH₂CH₂CQOCH₂Ph, 2H), 4.37-4.4 (t, >CHCH₂CH (CH₃)₂, 1H), 5.1-5.15 (bs, -CONHCH₂CH₂COOCH₂Ph, 2H), 5.8-5.89 (s, -NHCOCH₂CH₂ (CH₂)₁₂CH₃, 1H), 6.5-6.58 (s, CONHCH₂CH₂COOCH₂Ph, 1H), 7.3-7.73 (bs, aromatic, .5H).¹³C NMR(75 MHz, CDCl₃) : $\delta_{\rm C}$: 173.2, 172.3, 171.8, 135.5, 128.59, 128.38, 128.43, 128.23, 128.18, 77.47, 77.05, 76.62, 66.53, 51.53, 41.35, 36.53, 35.01, 33.9, 33.47, 31.9, 29.67, 29.64, 29.61, 29.5, 29.33, 29.26, 25.64, 24.81, 24.77, 22.82, 22.66, 22.2, 14.08. IR (KBr) v: 3296.52, 2917.83, 2849.96, 1737.71, 1635.24, 1540.04, 1464.26, 1387.95, 1171.89, 1071.86, 960.70, 696.15.ESI-MS (m/z) C₃₂H₅₄N₂O₄ (EXACT MASS= 530.40) 531.4 (100%, [M + H]⁺).

Synthesis of 4 (e): To a solution of N-hexadecanoyl-phenylalanine (1.58 g, 3.9mmole, 1eqv) in 5 ml of THF, HOBt (0.790 g, 5.85mmole, 1.5 eqv.) was added and kept on stirring in ice water bath at 0^{0} C.After 10mins of stirring, DMAP (0.043 g, 0.358mmole, 0.092eqv.) and solution of DCC (1.631 g, 7.917mmole, 1.93 eqv) in 5 ml of CHCl₃ was added to it and kept stirring. Finally, after 15mins, K₂CO₃ (0.792 g, 5.73mmole, 1.47eqv.) and PTSA-salt of β -Alanine benzyl ester (1.64 g, 4.68mmole, 1.2eqv.) was added and kept on stirringovernight. The reaction mixture was then concentrated under reduced pressure and taken up in 30 ml CHCl₃ and washed sequentially with ice cold 1 (N) HCl (25 ml × 2), saturated brine (25 ml × 2). The organic layer was collected and dried over anhyd Na₂SO₄and then purified by silica column chromatography (30%EtOAc/Petroleum ether) to get the desired product. (1.24 g, 56.4% yield).

¹HNMR (300MHz, CDCl₃) : δ ::0.89-.0.95 (t, -NHCOCH₂CH₂ (CH₂)₁₂CH₃, 3H), 1.15-1.35 (bs, -NHCOCH₂CH₂(CH₂)₁₂CH₃, 24H), 1.51-1.6 (d, -NHCOCH₂CH₂ (CH₂)₁₂CH₃, 2H), 2.1-2.28 (t, -NHCOCH₂CH₂ (CH₂)₁₂CH₃, 2H), 2.3-2.55 (m, -CONHCH₂CH₂COOCH₂Ph, 2H), 2.99-3.15 (dd, >CHCH₂Ph, 2H), 3.37-3.49 (m, -CONHCH₂CH₂COOCH₂Ph, 2H), 4.5-4.59 (d, >CHCH₂Ph, 1H), 5.07-5.14 (bs, -CONHCH₂CH₂COOCH₂Ph, 2H), 6.07-6.14 (d and s, -*NH*COCH₂CH₂ (CH₂)₁₂CH₃and -CONHCH₂CH₂COOCH₂Ph, 2H), 7.16-7.35 (m, >CHCH₂Ph and-CONHCH₂CH₂COOCH₂Ph, 1OH).¹³C NMR(75 MHz, CDCl₃) : $\delta_{\rm C}$: 173.095, 171.805, 170.956, 136.626, 135. 56, 129.22, 128.63, 128.60, 128.41, 128.27, 126.942, 77.79, 77.05, 76.63, 66.52, 54.45, 49.51, 38.72, 36.55, 34.76, 33.79, 33.51, 31.92, 29.69, 29.66, 29.48, 29.35, 29.21, 25.57, 25.463, 24.78, 22.69, 14.12.IR (solid)

v: 3583, 3512, 3130, 2989, 2905, 2274, 2239, 1698, 1634, 1574, 1539, 1428, 1369, 1023, 905, 786. ESI-MS (m/z) $C_{35}H_{52}N_2O_4$ (EXACT MASS= 564.4) 565.42 (100%, [M + H]⁺).

Synthesis of 4 (f): To a solution of N-hexadecanoyl-glycine (0.500 g, 1.59mmole, 1eqv.) in 5 ml of THF, HOBt (0.322 g, 2.39mmole, 1.5eqv.) was added and kept on stirring in ice water bath at 0°C.After 10mins of stirring, DMAP (0.019 g, 0.159mmole, 0.1eqv.) and solution of DCC (0.633 g, 3 mmole, 1.93eqv.) in 5 ml of CHCl₃ was added to it and kept stirring. Finally, after 15mins, K_2CO_3 (0.322 g, 2.33mmole, 1.47eqv) and PTSA-salt of β -Alanine benzyl ester(0.65 g, 1.9mmole, 1.2eqv.) was added and kept on stirringovernight. The reaction mixture was then concentrated under reduced pressure and taken up in 30 ml CHCl₃ and washed sequentially with ice cold 1 (N) HCl (25 ml×2), saturated brine (25 ml×2).The organic layer was collected and dried over anhyd. Na₂SO₄and then purified by silica column chromatography (60%EtOAc/Petroleum ether) to get the desired product. (0.62 g, 82% yield).

¹HNMR (300MHz, CDCl₃) : δ ::0.85-0.90 (t, -NHCOCH₂CH₂ (CH₂)₁₂CH₃, 3H), 1.24-1.25 (t, -NHCOCH₂CH₂(CH₂)₁₂CH₃, 24H), 1.5-1.66 (s, - NHCOCH₂CH₂ (CH₂)₁₂CH₃, 2H), 2.0-2.2 (t, -NHCOCH₂CH₂ (CH₂)₁₂CH₃, 2H), 2.5-2.6 (s, -CONHCH₂CH₂COOCH₂Ph, 2H), 3.54-3.6(s, -CONHCH₂CH₂CH₂COOCH₂Ph, 2H), 4.20-4.24 (s, -NHCOC<u>H</u>(CH₃)NHCOCH₂CH₂ (CH₂)₁₂CH₃, 1H), 5.2 (s, -CONHCH₂CH₂COOC<u>H</u>₂Ph, 2H), 6.11-6.13 00-6.1(d, -CON<u>H</u>CH₂CH₂COOCH₂Ph, 1H), 6.60-6.62 (s, -N<u>H</u>COCH₂CH₂ (CH₂)₁₂CH₃, 1H), 7.3-7.38 (bs, aromatic, 5H).¹³C NMR(75 MHz, CDCl₃) : δ_{C} : 173.27, 172.93, 171.407, 135.6, 128.62, 128.4, 128.28, 77.46, 77.041, 76.61, 66.59, 49.23, 36.7, 34.976, 33.99, 33.69, 31.9, 31.2, 29.49, 29.33, 29.28, 25.75, 25.54, 25.54, 24.86, 24.77, 22.67, 19.13, 18.22, 14.09.IR (KBr) v: 3321, 2914, 2849, 1725, 1667, 1520, 1422, 1214, 1077, 898, 719, 647.ESI-MS (m/z) C₂₉H₄₈N₂O₄ (EXACT MASS=474.36) 475.369 (12%, [M + H]⁺), 497.244 (100%, [M + Na]⁺).



¹H-NMR SPECTRA OF 4a.



¹³C-NMR SPECTRA OF **4a.**



IR (in KBr) SPECTREA OF 4a



¹H-NMR of **4b**.



¹³C-NMR SPECTRA OF **4b.**



IR (in KBr) SPECTREA OF 4b



¹H-NMR of **4c**.



¹³C-NMR SPECTRA OF **4c.**



IR (in KBr) SPECTREA OF **4c**



¹H-NMR of **4d**.



¹³C-NMR SPECTRA OF **4d.**



IR (in KBr) SPECTRA OF 4d



¹H-NMR of **4e**.



¹³C-NMR SPECTRA OF **4e.**



IR (solid) SPECTRA OF 4e.



¹H-NMR of **4f**.



¹³C-NMR SPECTRA OF **4f**.



IR SPECTRA OF 4f.

THERMAL STABILITY:



FigureS1. $T_{gel}(^{\circ}C)$ versus concentration (moles/lit) plot of different gelator in 80%(v/v) EtOH in H₂O.



FigureS2. $T_{gel}(^{\circ}C)$ versus concentration (moles/lit) plot of different gelator in Xylene



FigureS3. $T_{gel}(^{\circ}C)$ versus concentration (moles/lit) plot of **4f** gelator in acetonitrile



FigureS4. T_{gel} (°C) versus concentration (moles/lit) plot of **4f** gelator in 80% (v/v) EtOH in H₂O.

RHEOLOGY:



FigureS5. Strain amplitude sweep experiment (at 10rads/sec)andTemperature sweep experiment at constant frequency of 2Hz and heating rate 2°C/min for (**4a**) in acetonitrile.



FigureS6. Strain amplitude sweep experiment (at 10rads/sec) andTemperature sweep experiment at constant frequency of 2Hz and heating rate 2°C/min for (**4c**) in acetonitrile.



FigureS7. Strain amplitude sweep experiment (at 10rads/sec)andTemperature sweep experiment at constant frequency of 2Hz and heating rate 2°C/min for (**4e**) in acetonitrile.

RHEOLOGY:



FigureS8. Strain amplitude sweep experiment (at 10rads/sec)for(i) **4f** in acetonitrile and (ii) **4c** in EtOH.

FT-IR



FigureS9. FT-IR spectra of 4a in acetonitrile



FigureS10. FT-IR spectra of 4b in acetonitrile



FigureS11. FT-IR spectra of 4c in acetonitrile



FigureS12. FT-IR spectra of 4d in acetonitrile





FigureS13. FT-IR spectra of **4e** in acetonitrile.



FigureS14. FT-IR spectra of **4e** in acetonitrile.

SOLVENT- EFFECT:



FigureS15. Plot of GN versus E_TN of the gelators in aromatic solvents.



FigureS16. Plot of GN versus E_TN of the gelators in aromatic solvents.



FigureS17. Plot of GN versus E_TN of the gelators in aprotic polar solvents.



FigureS18. Plot of GN versus E_TN of the gelators in aprotic polar solvents.

SOLVENT-EFFECT



FigureS19. Plot of GN versus E_T 30 of the gelators in protic polar solvents.

SOLVENT-EFFECT



Figure S20. GN versus δ_p plot of gelators in polar protic solvents.



Figure S21. GN versus δ_{p} plot of gelators in polar aprotic solvents.



Figure S22. GN versus δ_a plot of gelators in polar protic solvents.

SOLVENT-EFFECT



Figure S23. Teas plot of Hansen Parameters for 4a in solvents of gelation



Figure S24. Teas plot of Hansen Parameters for 4b in solvents of gelation



Figure S25. Teas plot of Hansen Parameters for 4d in solvents of gelation

Panel–I



Panel-II



Figure S26.Plotsof₁(a) $\delta_h vs\delta_p(b)\delta_a vs\delta_h$, (c) $\delta_d vs\delta_h$ and(d) $\delta_a vs\delta_p(e) \delta_d vs\delta_p(f) \delta_d vs\delta_a$ (HSP)[panel I]and(a) $\beta vs.\alpha$, (b) $\pi^* vs.\alpha$ and(c) $\pi^* vs.\beta$ (KT)[panel II]of gelators**4a-4e**[shaded zones are prevailin g area of gelation].

XRD:



Figure S27XRD profile of xero-gels (intensity vs. 2 θ) for**4b**in acetonitrile [inset represent s respective plots of d⁻¹ vs. $\sqrt{(h^2+l^2+k^2)}$]



 $Figure \ S28 \text{XRD profile of xero-gels (intensity vs. 2\theta) for \ \textbf{4e} \ in \ acetonitrile \ [inset \ represent \ s \ respective \ plots \ of \ d^{-1} \ vs. \ \sqrt{(h^2+l^2+k^2)]}$



Figure S29XRD profile of xero-gels (intensity vs. 2 θ) for **4a** in EtOH [inset represent s respective plots of d⁻¹ vs. $\sqrt{(h^2+l^2+k^2)}$]



Figure S30xRD profile of xero-gels (intensity vs. 2 θ) for **4a** in EtOAc [inset represent s respective plots of d⁻¹ vs. $\sqrt{(h^2+l^2+k^2)}$]



Figure S31XRD profile of xero-gels (intensity vs. 2 θ) for **4b** in EtOAc [inset represent s respective plots of d⁻¹ vs. $\sqrt{(h^2+l^2+k^2)}$]



Figure S32XRD profile of xero-gels (intensity vs. 20) for **4b** in EtOH [inset represent s respective plots of d⁻¹ vs. $\sqrt{(h^2+l^2+k^2)}$]



Figure S33XRD profile of xero-gels (intensity vs. 20) for **4c** in EtOH [inset represent s respective plots of d⁻¹ vs. $\sqrt{(h^2+l^2+k^2)}$]



Figure S34XRD profile of xero-gels (intensity vs. 20) for **4c** in EtOAc [inset represent s respective plots of d^{-1} vs. $\sqrt{(h^2+l^2+k^2)}$]



Figure S35XRD profile of xero-gels (intensity vs. 20) for **4d** in EtOAc [inset represent s respective plots of d⁻¹ vs. $\sqrt{(h^2+l^2+k^2)}$]



Figure S36XRD profile of xero-gels (intensity vs. 20) for **4d** in EtOH [inset represent s respective plots of d⁻¹ vs. $\sqrt{(h^2+l^2+k^2)}$]



Figure S37XRD profile of xero-gels (intensity vs. 20) for **4e**in EtOAc [inset represent s respective plots of d⁻¹ vs. $\sqrt{(h^2+l^2+k^2)}$]



Figure S38XRD profile of xero-gels (intensity vs. 2 θ) for **4e** in EtOH [inset represent s respective plots of d⁻¹ vs. $\sqrt{(h^2+l^2+k^2)}$]



Figure S39XRD profile of xero-gels (intensity vs. 20) for **4f** in acetonitrile [inset represent s respective plots of d⁻¹ vs. $\sqrt{(h^2+l^2+k^2)}$]



Figure S40XRD profile of xero-gels (intensity vs. 20) for **4f** in EtOH [inset represent s respective plots of $d^{-1}vs$. $\sqrt{(h^2+l^2+k^2)}$]



Figure S41XRD profile of xero-gels (intensity vs. 20) for **4f**in EtOAc [inset represent s respective plots of d^{-1} vs. $\sqrt{(h^2+l^2+k^2)}$]



Figure S42XRD profile of xero-gels (intensity vs. 2 θ) for **4a** in 80% EtOH-Water [inset represent s respective plots of d⁻¹ vs. $\sqrt{(h^2+l^2+k^2)}$]



Figure S43XRD profile of xero-gels (intensity vs. 2 θ) for **4b** in 80% EtOH-Water [inset represent s respective plots of d⁻¹ vs. $\sqrt{(h^2+l^2+k^2)}$]



 $\label{eq:Figure S44xRD} Figure \ S44xRD \ profile \ of \ xero-gels \ (intensity \ vs. \ 2\theta) \ for \ \textbf{4c} \ in \ 80\% \ EtOH-Water \ [inset \ represent \ s \ respective \ plots \ of \ d^{-1} \ vs. \ \sqrt{(h^2+l^2+k^2)]}$







 $\label{eq:Figure S46xRD profile of xero-gels (intensity vs. 2\theta) for \mbox{4e in 80\% EtOH-Water [inset represent s respective plots of d^-1 vs. $\sqrt(h^2+l^2+k^2)$]}$



Figure S47XRD profile of xero-gels (intensity vs. 20) for **4f** in 80% EtOH-Water [inset represents respective plots of d^{-1} vs. $\sqrt{(h^2+l^2+k^2)}$]

UV and FLUORESCENCE:



Figure S48U.V spectra of gelator 4a and 4e in acetonitrile at different concentrations.



Figure S49U.V spectra of gelator 4a and 4e in acetonitrile at different concentrations.



Figure S50Fluorescence excitation spectra of gelator **4a** and **4e** in acetonitrile at different concentrations.