

SUPPORTING INFORMATION

Visible light-mediated decarboxylative amination of indoline-2-carboxylic acids catalyzed by Rose Bengal

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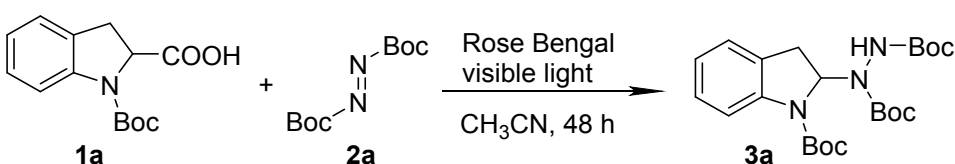
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1. General Information

N-protected indoline-2-carboxylate acids and *N*-Boc-1,2,3,4-tetrahydroisoquinoline-1-carboxylic acid (**1**) were prepared according to the literature¹. Rose Bengal was purchased from Aladdin, Shanghai, China (R104993-1 G, Lot#L15040, 95%). Unless otherwise noted, all reagents were purchased from commercial suppliers and used without further purification. Reactions were monitored by thin-layer chromatography (TLC) with Haiyang GF254 silica gel plates (Qingdao Haiyang chemical industry Co Ltd. Qingdao, China) using UV light and vanillic aldehyde as visualizing agents. Flash column chromatography was performed using 200-300 mesh silica gel at increased pressure. ¹H NMR and ¹³C NMR spectra were recorded on Bruker-AM 600 (600 MHz) (Bruker BioSpin AG Ltd., Beijing, China). Chemical shifts were reported in ppm from TMS with the solvent resonance as the internal standard. Data were reported as follows: chemical shifts (δ) in ppm, coupling constants (*J*) in Hz, and solvent (CDCl₃). High-resolution mass spectra were obtained by using ESI ionization sources (Varian 7.0T FTICR-MS). Melting points were taken on a WPX-4 apparatus and were uncorrected (Yice instrument equipment Co Ltd, Shanghai).

2. Extra information for optimization of the reaction conditions

Table S1 Effect of molar ratio of substrates on the model reaction ^a



1a + **2a** $\xrightarrow[\text{CH}_3\text{CN, 48 h}]{\text{Rose Bengal, visible light}}$ **3a**

Entry	Molar ratio (1a : 2a)	Yield (%) ^b
1	3 : 1	55
2	2.5 : 1	54
3	2 : 1	52
4	1.5 : 1	46
5	1 : 1	41
6	1 : 1.5	49
7	1 : 2	50

^a Reaction conditions: **1a** (0.2-0.6 mmol), **2a** (0.2-0.4 mmol), CH₃CN (1.0 mL), and Rose Bengal (2 mol%) under irradiation of 32 W fluorescent bulb at rt for 48 h.

^b Isolated yield.

Table S2 Effect of solvent volume on the model reaction ^a

Entry	CH ₃ CN (mL)	Yield (%) ^b
1	0.5	54
2	1	52
3	1.5	47
4	2	50
5	2.5	48

^a Reaction conditions: **1a** (0.4 mmol), **2a** (0.2 mmol), CH₃CN, and Rose Bengal (2 mol%) under irradiation of 32 W fluorescent bulb at rt for 48 h.

^b Isolated yield.

Table S3 Effect of visible light on the model reaction ^a

Entry	Visible light (W)	Yield (%) ^b
1	5	25
2	12	34
3	23	52
4	32	54
5	45	54

^a Reaction conditions: **1a** (0.4 mmol), **2a** (0.2 mmol), CH₃CN (0.5 mL), and Rose Bengal (2 mol%) under irradiation of fluorescent bulb at rt for 48 h.

^b Isolated yield.

Table S4 Effect of photocatalyst loading on the model reaction ^a

Entry	Rose Bengal (x mol%)	Yield (%) ^b
1	0.5	58
2	1	66

3	2	53
4	4	48
5	7.5	39
6	10	32

^a Reaction conditions: **1a** (0.4 mmol), **2a** (0.2 mmol), CH₃CN (0.5 mL), and Rose Bengal (x mol%) under irradiation of 23 W fluorescent bulb at rt for 48 h.

^b Isolated yield.

3. Fluorescence of **2a**

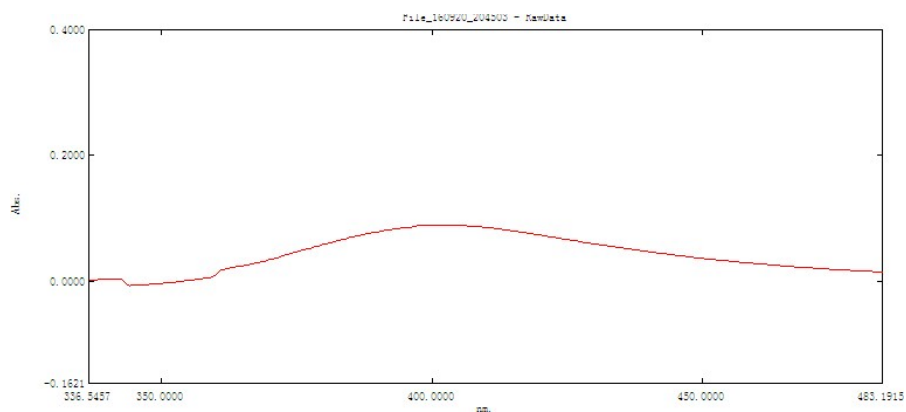
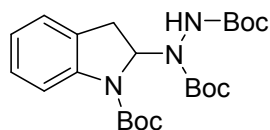


Figure 1. Fluorescence response of **2a** (2.5 mM in CH₃CN).

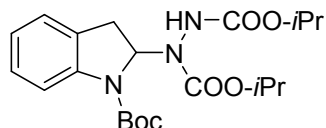
4. Characterization data of the products

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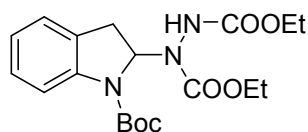
White solid; mp 174-175 °C; ^1H NMR (600 MHz, CDCl_3) δ = 7.73 (m, 1H), 7.11 (m, 2H), 6.92 (t, J =7.3, 1H), 6.51 (br, 1H), 5.85 (d, J =179.1, 1H), 3.30 (m, 2H), 1.50 (m, 27H); ^{13}C NMR (151 MHz, CDCl_3) δ = 155.3, 153.0, 152.1, 142.2, 129.5, 127.1, 124.2, 123.9, 122.5, 114.3, 81.6, 81.1, 70.7, 34.6, 32.9, 28.5, 28.2, 28.2, 28.1. HRMS(ESI) m/z 472.2422 ($\text{M}+\text{Na}^+$), Cal. $\text{C}_{23}\text{H}_{35}\text{N}_3\text{O}_6\text{Na}$, 472.2418.

Di-*iso*-propyl 1-(1-(*tert*-butoxycarbonyl)indolin-2-yl)hydrazine-1,2-dicarboxylate (3b)



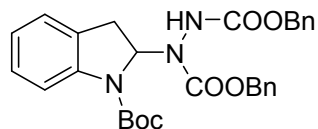
White solid; mp 139-140 °C; ^1H NMR (600 MHz, CDCl_3) δ = 7.60 (m, 1H), 7.12 (m, 2H), 6.92 (t, J =7.2, 1H), 6.54 (br, 1H), 6.04 (d, J =94.7, 1H), 4.89 (m, 2H), 3.32 (m, 2H), 1.54 (s, 9H), 1.17 (m, 12H); ^{13}C NMR (151 MHz, CDCl_3) δ = 155.9, 151.9, 142.0, 127.1, 123.9, 122.5, 114.2, 81.8, 70.7, 70.0, 69.7, 28.4, 21.8. HRMS(ESI) m/z 444.2113 ($\text{M}+\text{Na}^+$), Cal. $\text{C}_{21}\text{H}_{31}\text{N}_3\text{O}_6\text{Na}$, 444.2105.

Diethyl 1-(1-(*tert*-butoxycarbonyl)indolin-2-yl)hydrazine-1,2-dicarboxylate (3c)



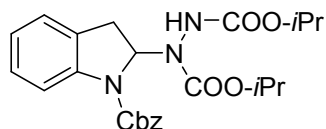
Colorless oil; ^1H NMR (600 MHz, CDCl_3) δ = 7.71 (m, 1H), 7.12 (m, 2H), 6.93 (t, J =7.1, 1H), 6.60 (br, 1H), 6.23 (d, J =89.0, 1H), 4.12 (m, 4H), 3.33 (m, 2H), 1.54 (s, 9H), 1.21 (dd, J =41.5, 35.3, 6H); ^{13}C NMR (151 MHz, CDCl_3) δ = 156.4, 154.5, 151.9, 141.9, 129.0, 127.2, 123.9, 122.6, 114.3, 81.9, 62.6, 62.0, 29.6, 28.4, 14.3. HRMS(ESI) m/z 416.1800 ($\text{M}+\text{Na}^+$), Cal. $\text{C}_{19}\text{H}_{27}\text{N}_3\text{O}_6\text{Na}$, 416.1792.

Dibenzyl 1-(1-(*tert*-butoxycarbonyl)indolin-2-yl)hydrazine-1,2-dicarboxylate (3d)



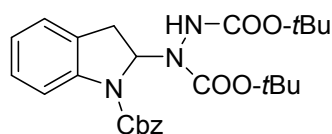
White solid; mp 123-124 °C; ^1H NMR (600 MHz, CDCl_3) δ = 7.66 (m, 1H), 7.31 (dd, J =33.0, 9.9, 8H), 7.08 (dd, J =96.7, 59.9, 5H), 6.62 (br, 1H), 6.34 (d, J =124.1, 1H), 5.06 (m, 4H), 3.19 (m, 2H), 1.46 (d, J =37.9, 9H); ^{13}C NMR (151 MHz, CDCl_3) δ = 156.2, 153.9, 151.9, 141.9, 135.7, 128.6, 128.5, 128.2, 127.9, 127.3, 126.9, 124.0, 122.7, 114.4, 82.0, 68.2, 67.6, 28.4. HRMS(ESI) m/z 540.2112 ($\text{M}+\text{Na}^+$), Cal. $\text{C}_{29}\text{H}_{31}\text{N}_3\text{O}_6\text{Na}$, 540.2105.

Di-*iso*-propyl 1-(1-((benzyloxy)carbonyl)indolin-2-yl)hydrazine-1,2-dicarboxylate (3e)



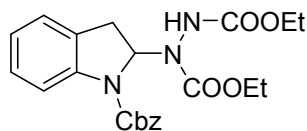
White solid; mp 120-122 °C; ^1H NMR (600 MHz, CDCl_3) δ = 7.72 (m, 1H), 7.37 (m, 5H), 7.13 (m, 2H), 6.96 (d, J =6.7, 1H), 6.67 (br, 1H), 6.19 (d, J =115.2, 1H), 5.29 (m, 2H), 4.86 (m, 2H), 3.38 (m, 2H), 1.21 (m, 12H); ^{13}C NMR (151 MHz, CDCl_3) δ = 156.2, 154.1, 153.9, 152.7, 141.6, 135.9, 128.6, 128.3, 127.3, 123.9, 122.9, 114.4, 69.8, 67.6, 29.7, 21.8. HRMS(ESI) m/z 478.1956 ($\text{M}+\text{Na}^+$), Cal. $\text{C}_{24}\text{H}_{29}\text{N}_3\text{O}_6\text{Na}$, 478.1949.

Di-*tert*-butyl 1-(1-((benzyloxy)carbonyl)indolin-2-yl)hydrazine-1,2-dicarboxylate (3f)



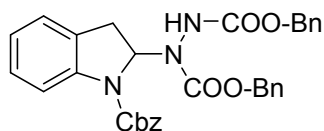
White solid; mp 153-154 °C; ^1H NMR (600 MHz, CDCl_3) δ = 7.75 (m, 1H), 7.37 (m, 5H), 7.12 (dd, J =17.0, 6.9, 2H), 6.95 (t, J =7.3, 1H), 6.61 (br, 1H), 5.82 (d, J =192.4, 1H), 5.30 (d, J =51.7, 2H), 3.30 (m, 2H), 1.35 (dd, J =73.0, 28.7, 18H); ^{13}C NMR (151 MHz, CDCl_3) δ = 155.3, 153.1, 152.7, 141.8, 136.1, 128.7, 128.6, 128.1, 127.2, 124.0, 122.9, 114.4, 81.6, 81.0, 70.8, 67.3, 33.4, 28.0. HRMS(ESI) m/z 506.2266 ($\text{M}+\text{Na}^+$), Cal. $\text{C}_{26}\text{H}_{23}\text{N}_3\text{O}_6\text{Na}$, 506.2262.

Diethyl 1-(1-((benzyloxy)carbonyl)indolin-2-yl)hydrazine-1,2-dicarboxylate (3g)



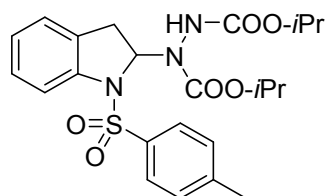
White solid; mp 93-94 °C; ^1H NMR (600 MHz, CDCl_3) δ = 7.81 (m, 1H), 7.37 (dd, J =18.5, 11.6, 5H), 7.13 (m, 2H), 6.96 (t, J =7.2, 1H), 6.66 (br, 1H), 6.29 (d, J =113.6, 1H), 5.26 (s, 2H), 4.07 (s, 4H), 3.35 (m, 2H), 1.19 (m, 6H); ^{13}C NMR (151 MHz, CDCl_3) δ = 156.6, 154.5, 152.8, 141.6, 135.9, 128.6, 128.4, 127.4, 124.0, 123.0, 114.4, 67.8, 62.9, 62.7, 62.0, 29.7, 14.3. HRMS(ESI) m/z 450.1643 ($\text{M}+\text{Na}^+$), Cal. $\text{C}_{22}\text{H}_{25}\text{N}_3\text{O}_6\text{Na}$, 450.1636.

Dibenzyl 1-(1-((benzyloxy)carbonyl)indolin-2-yl)hydrazine-1,2-dicarboxylate (3h)



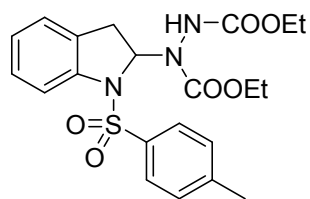
White solid; mp 143-144°C; ^1H NMR (600 MHz, CDCl_3) δ = 7.76 (m, 1H), 7.13 (m, 18H), 6.72 (m, 1H), 6.36 (d, J =139.9, 1H), 5.06 (m, 6H), 3.41 (m, 2H); ^{13}C NMR (151 MHz, CDCl_3) δ = 156.4, 152.7, 135.8, 135.6, 128.6, 128.5, 128.2, 127.8, 128.5, 124.1, 123.1, 114.4, 67.6, 29.7. HRMS(ESI) m/z 574.1957 ($\text{M}+\text{Na}^+$), Cal. $\text{C}_{32}\text{H}_{29}\text{N}_3\text{O}_6\text{Na}$, 574.1949.

Di-*iso*-propyl 1-(1-tosylindolin-2-yl)hydrazine-1,2-dicarboxylate (3i)



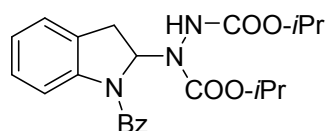
Colorless oil; ^1H NMR (600 MHz, CDCl_3) δ = 7.73 (m, 3H), 7.25 (m, 2H), 7.19 (t, J =7.7, 1H), 7.00 (m, 2H), 6.42 (d, J =25.1, 1H), 4.97 (m, 2H), 3.10 (m, 2H), 2.37 (s, 3H), 1.26 (m, 12H); ^{13}C NMR (151 MHz, CDCl_3) δ = 156.2, 144.4, 141.0, 129.8, 127.6, 124.6, 123.8, 114.4, 70.9, 69.9, 26.9, 21.9, 21.8, 21.7, 21.5. HRMS(ESI) m/z 498.1677 ($\text{M}+\text{Na}^+$), Cal. $\text{C}_{23}\text{H}_{29}\text{N}_3\text{O}_6\text{SNa}$, 498.1669.

Diethyl 1-(1-tosylindolin-2-yl)hydrazine-1,2-dicarboxylate (3j)



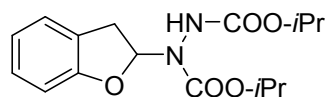
White solid; mp 141-142 °C; ^1H NMR (600 MHz, CDCl_3) δ = 7.73 (d, J =40.7, 3H), 7.22 (dt, J =15.5, 8.6, 3H), 7.00 (m, 2H), 6.42 (m, 2H), 4.19 (m, 4H), 3.21 (s, 2H), 2.37 (s, 3H), 1.26 (m, 6H); ^{13}C NMR (151 MHz, CDCl_3) δ = 156.5, 144.5, 141.0, 129.8, 127.7, 124.6, 123.9, 114.4, 63.1, 62.1, 26.9, 21.5, 14.3. HRMS(ESI) m/z 470.1361 ($\text{M}+\text{Na}^+$), Cal. $\text{C}_{21}\text{H}_{25}\text{N}_3\text{O}_6\text{SNa}$, 470.1356.

Di-iso-propyl 1-(1-benzoylindolin-2-yl)hydrazine-1,2-dicarboxylate (3k)



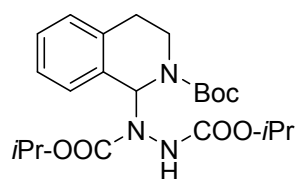
White solid; mp 175-176 °C; ^1H NMR (600 MHz, CDCl_3) δ = 7.30 (m, 9H), 6.29 (m, 2H), 4.92 (m, 2H), 3.40 (m, 2H), 1.26 (d, J =6.2, 12H); ^{13}C NMR (151 MHz, CDCl_3) δ = 169.9, 155.9, 142.3, 130.6, 128.8, 127.0, 124.4, 123.9, 115.6, 69.9, 64.3, 29.7, 21.9, 21.8. HRMS(ESI) m/z 448.1850 ($\text{M}+\text{Na}^+$), Cal. $\text{C}_{23}\text{H}_{27}\text{N}_3\text{O}_5\text{Na}$, 448.1843.

Di-iso-propyl 1-(2,3-dihydrobenzofuran-2-yl)hydrazine-1,2-dicarboxylate (3l)



Colorless oil; ^1H NMR (600 MHz, CDCl_3) δ = 7.12 (m, 2H), 6.82 (m, 2H), 6.11 (d, J =129.1, 1H), 4.99 (dd, J =33.4, 27.4, 2H), 3.37 (m, 2H), 1.25 (m, 12H); ^{13}C NMR (151 MHz, CDCl_3) δ = 157.3, 155.2, 153.5, 126.9, 123.5, 119.8, 107.8, 99.0, 69.1, 28.6, 20.7, 21.0, 20.9, 20.7. HRMS(ESI) m/z 345.1429 ($\text{M}+\text{Na}^+$), Cal. $\text{C}_{16}\text{H}_{22}\text{N}_2\text{O}_5\text{Na}$, 345.1421.

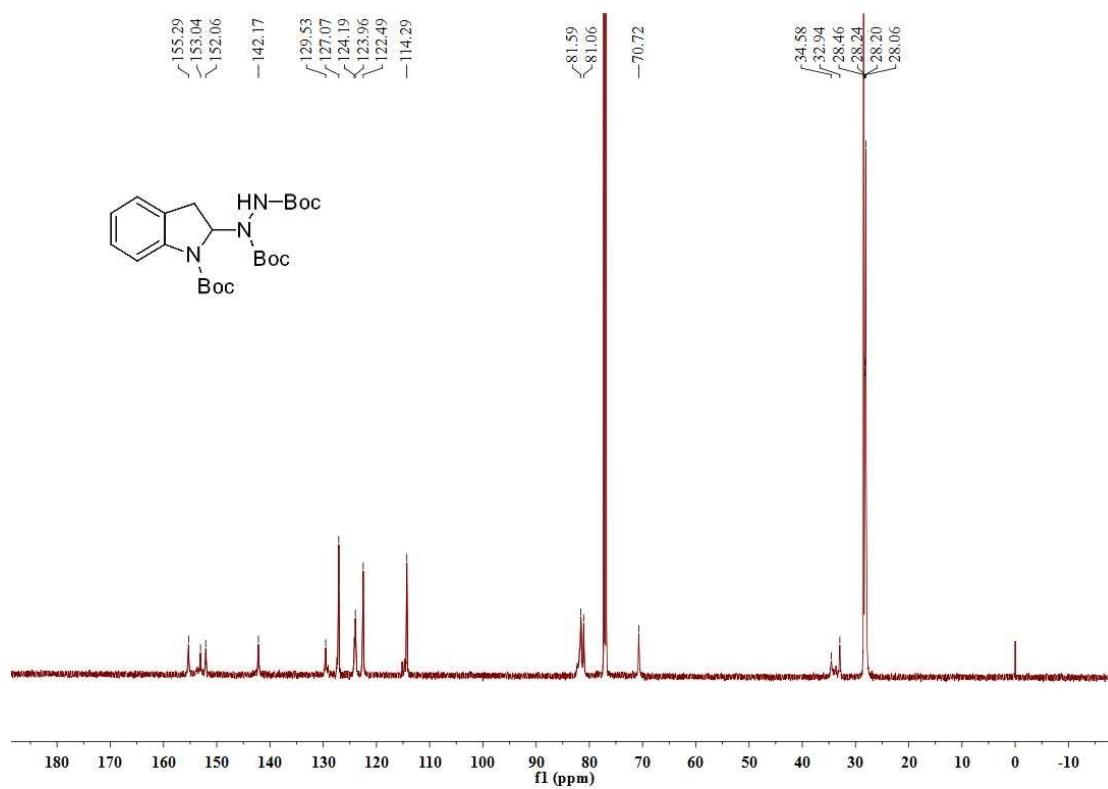
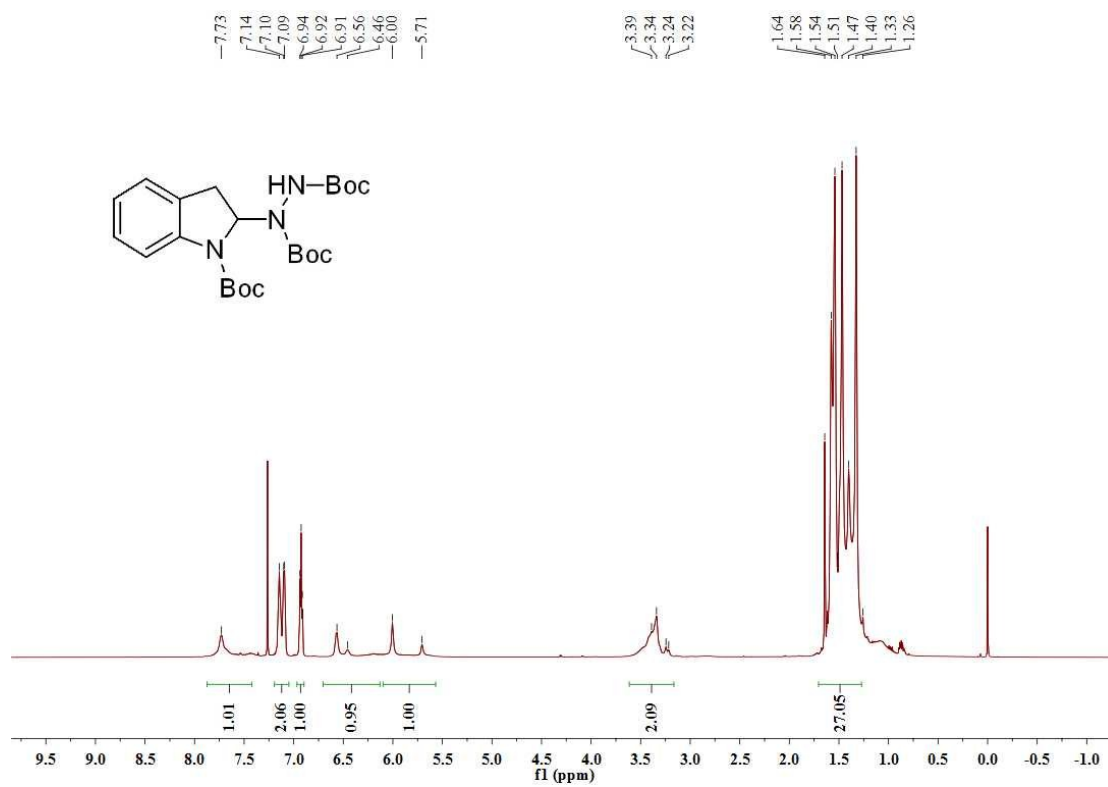
Di-iso-propyl 1-(2-(tert-butoxycarbonyl)-1,2,3,4-tetrahydroisoquinolin-1-yl)hydrazine-1,2-dicarboxylate (3m)



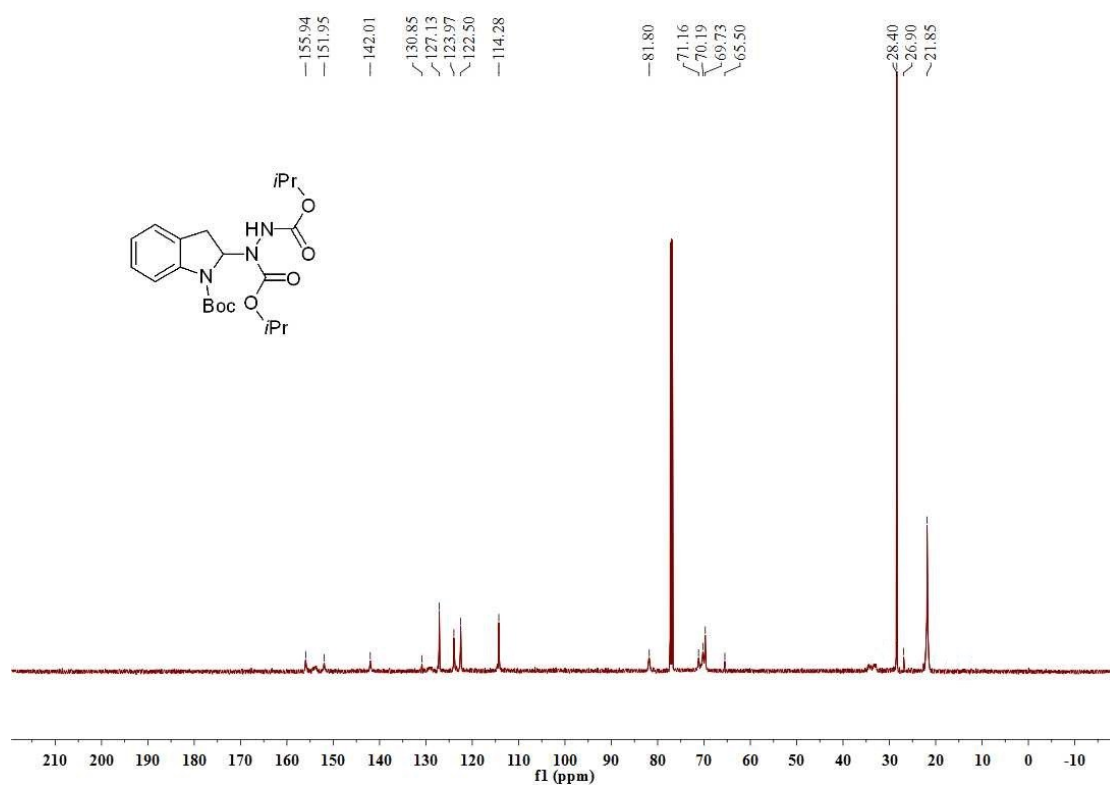
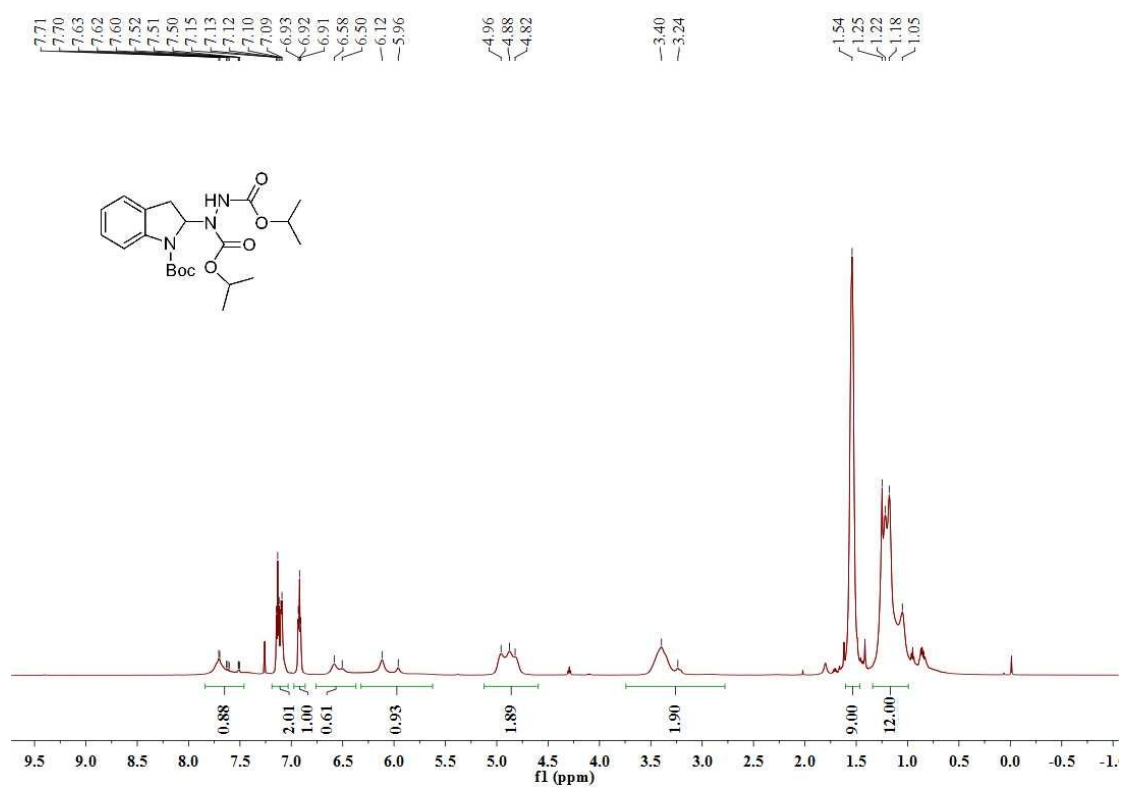
Colorless oil; ^1H NMR (600 MHz, CDCl_3) δ = 7.57 (s, 1H), 7.17 (m, 3H), 6.22 (dd, J =416.6, 316.2, 2H), 4.84 (d, J =201.0, 2H), 4.25 (d, J =110.9, 1H), 3.33 (d, J =130.6, 1H), 2.77 (d, J =30.9, 2H), 1.42 (m, 21H); ^{13}C NMR (151 MHz, CDCl_3) δ = 154.9, 134.4, 128.6, 127.8, 126.6, 80.8, 70.0, 28.9, 28.4, 21.9. HRMS(ESI) m/z 458.2269 ($\text{M}+\text{Na}^+$), Cal. $\text{C}_{22}\text{H}_{33}\text{N}_3\text{O}_6\text{Na}$, 458.2262.

5. ^1H NMR and ^{13}C NMR spectra of the products

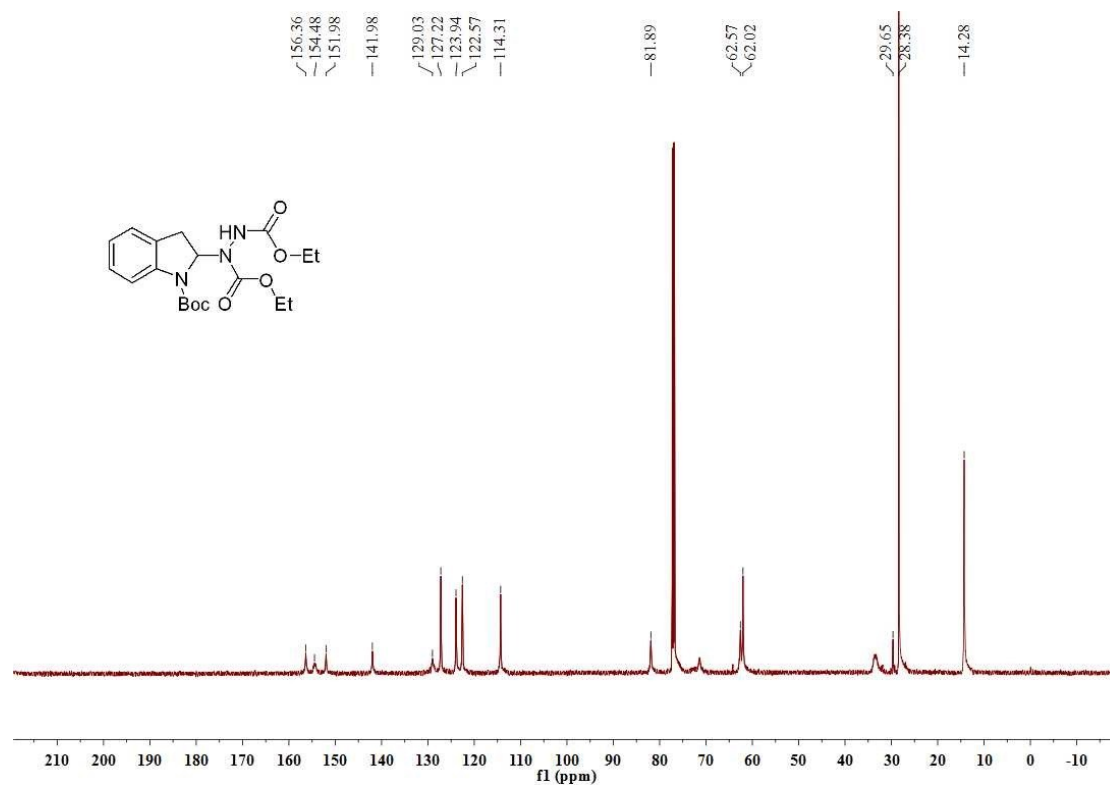
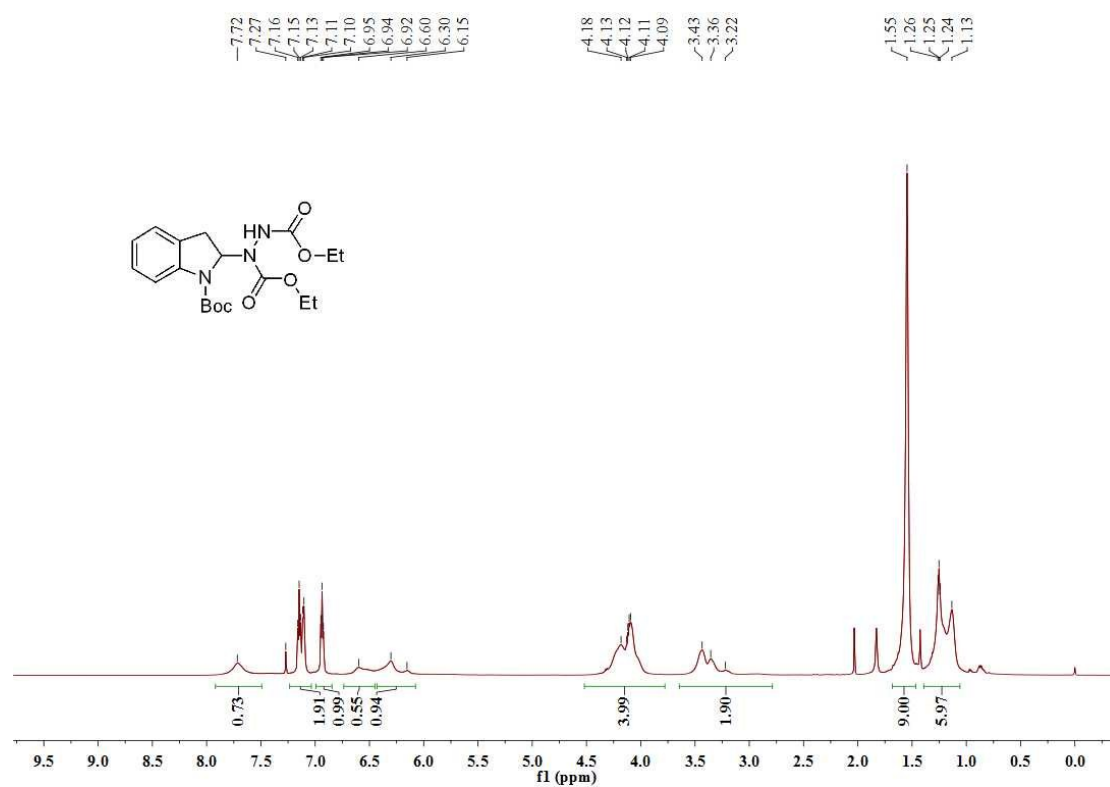
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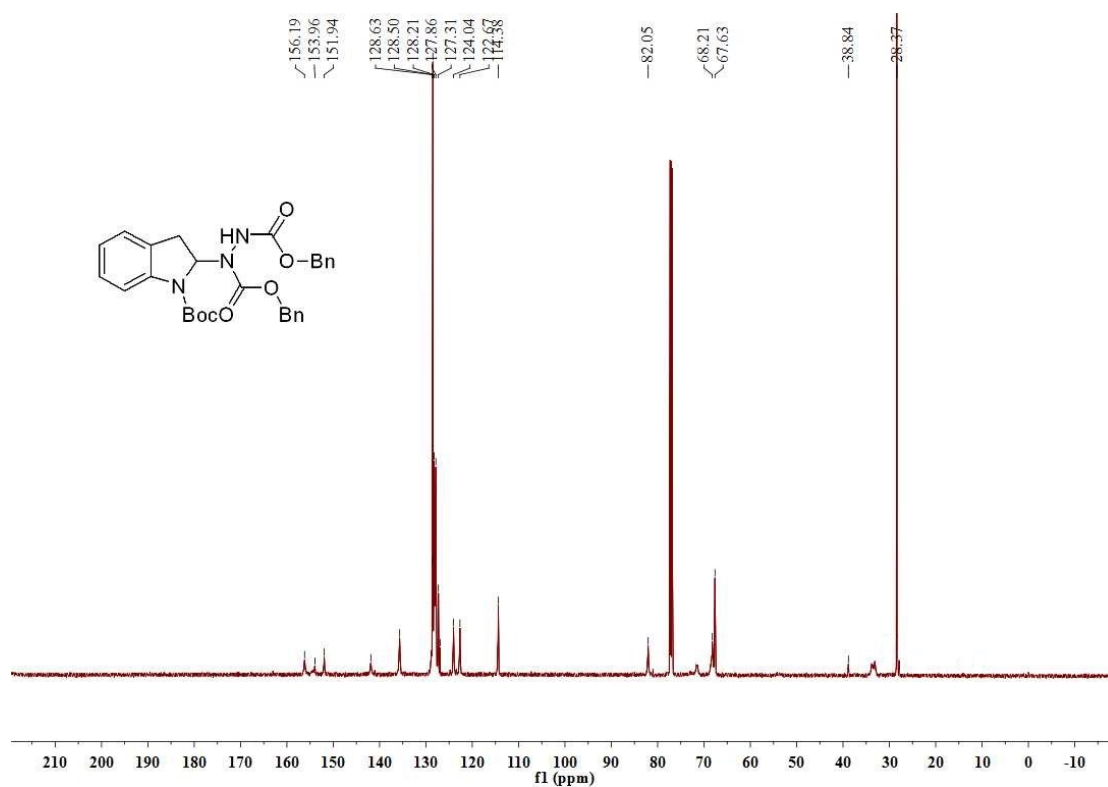
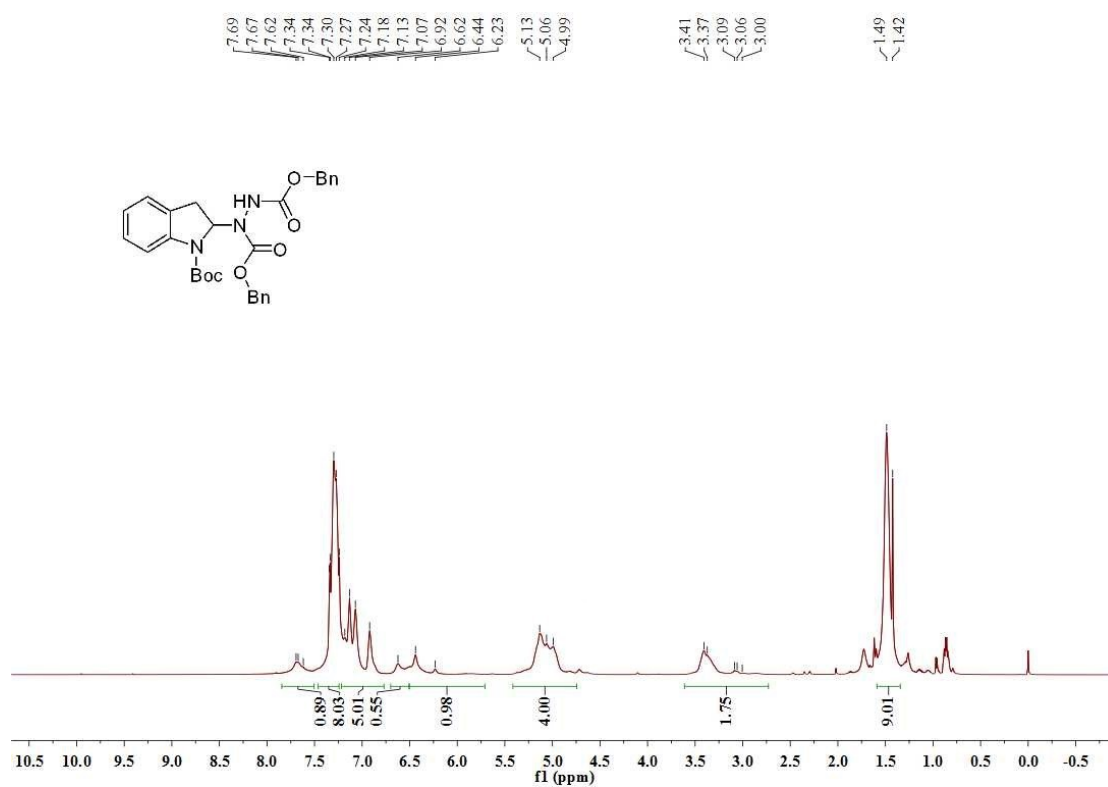
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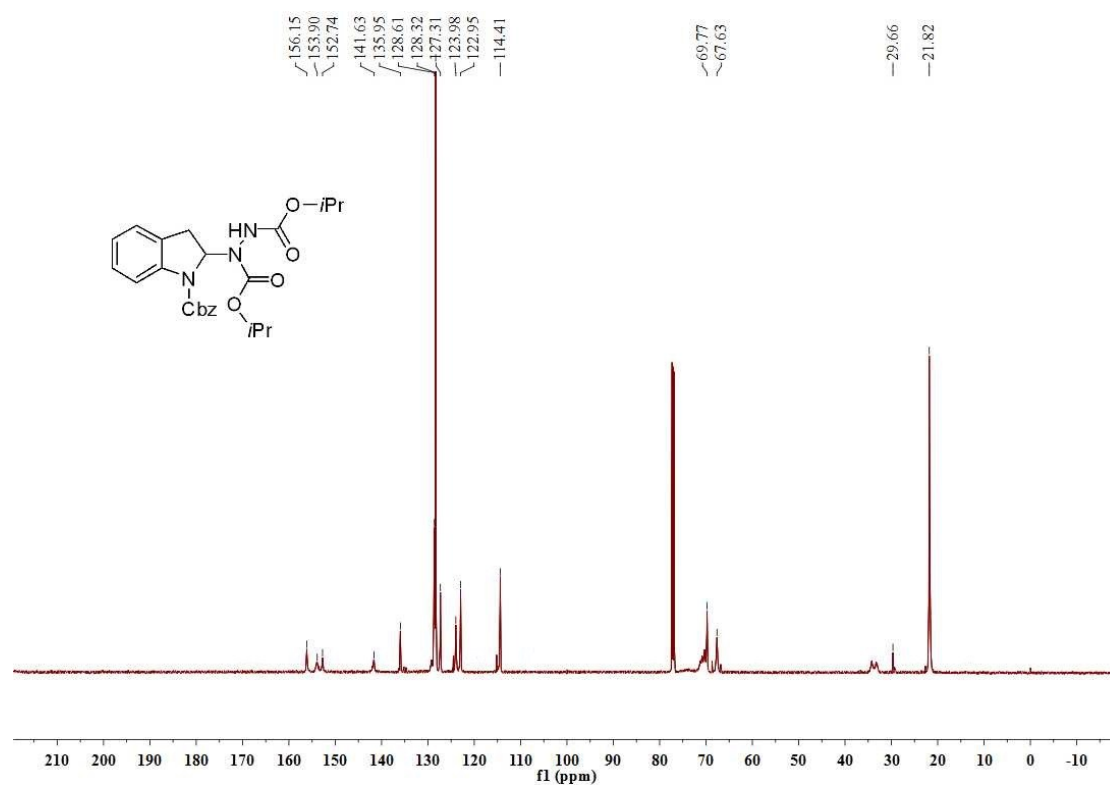
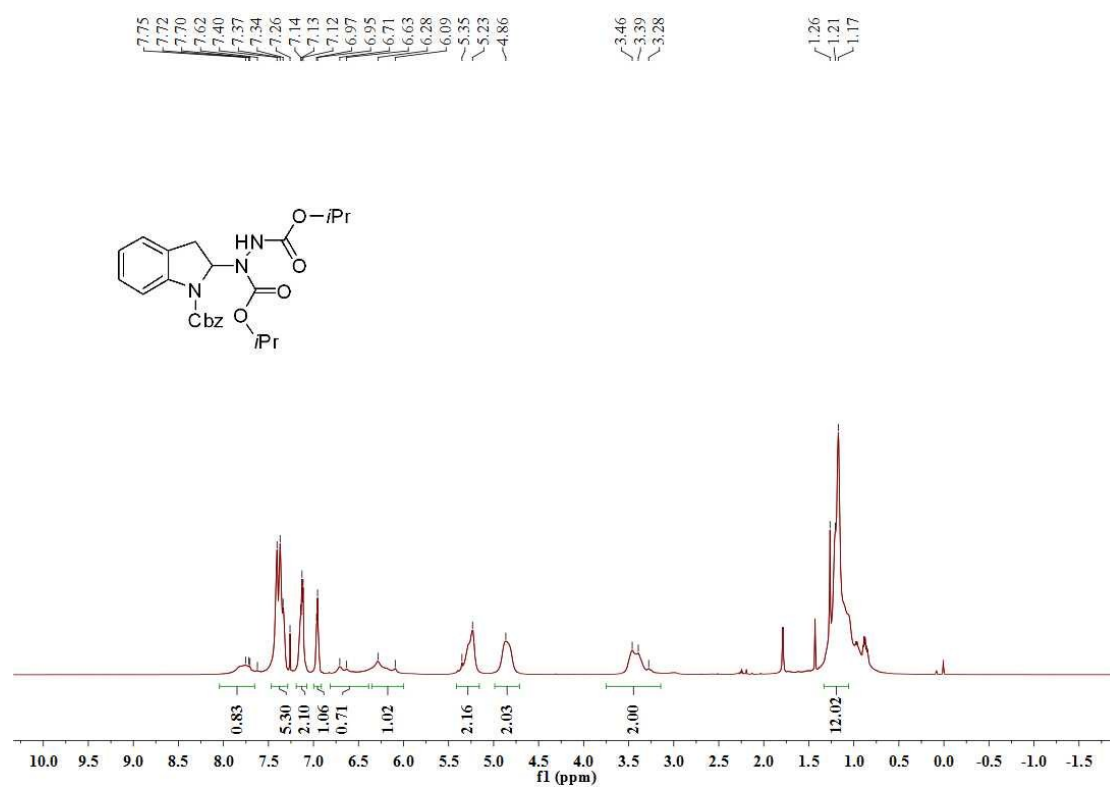
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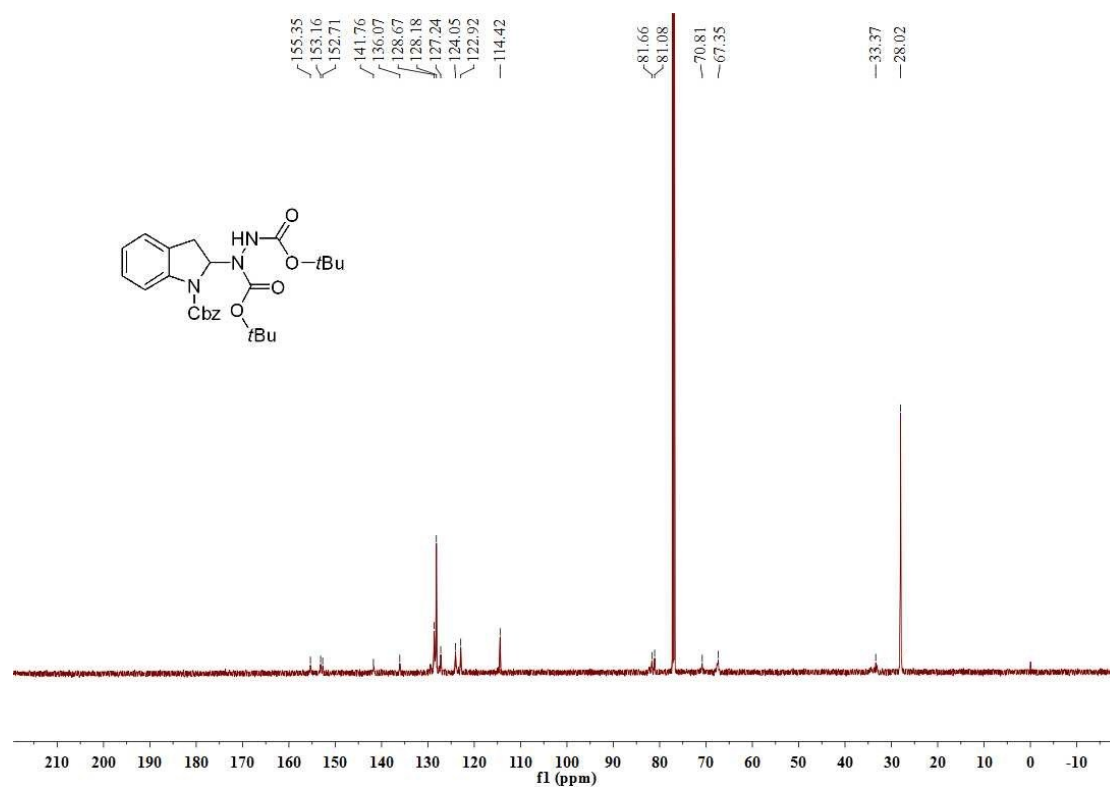
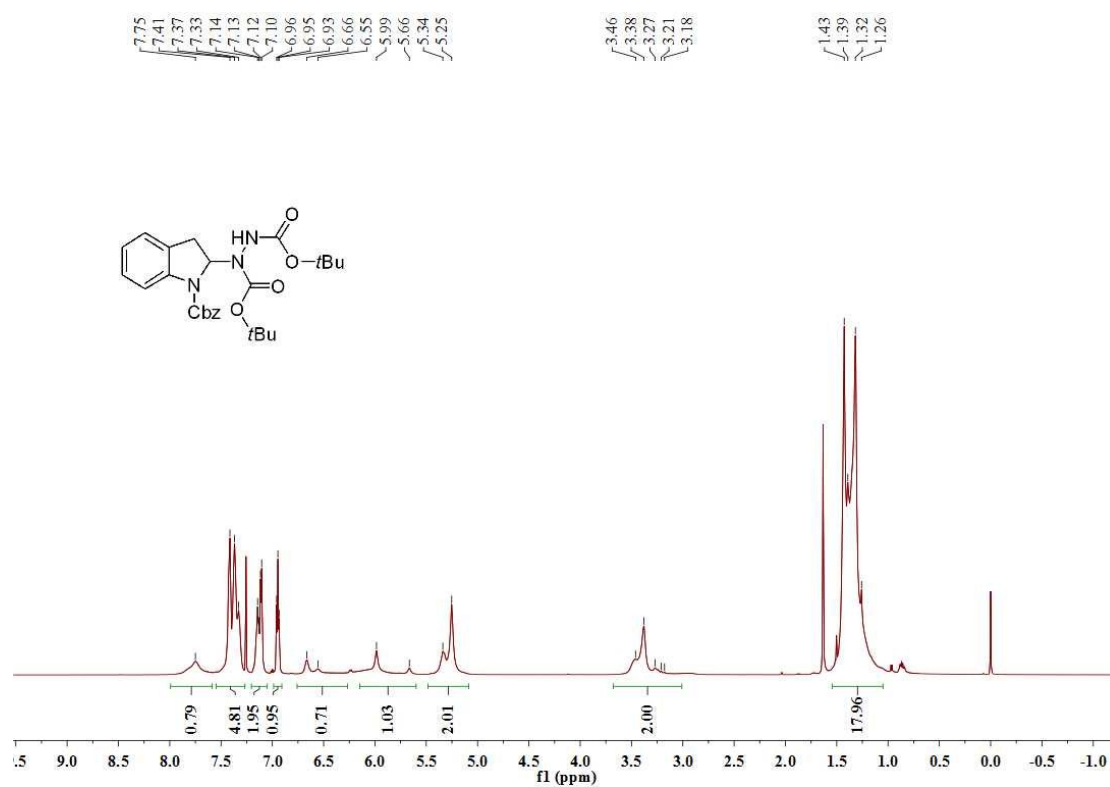
3d



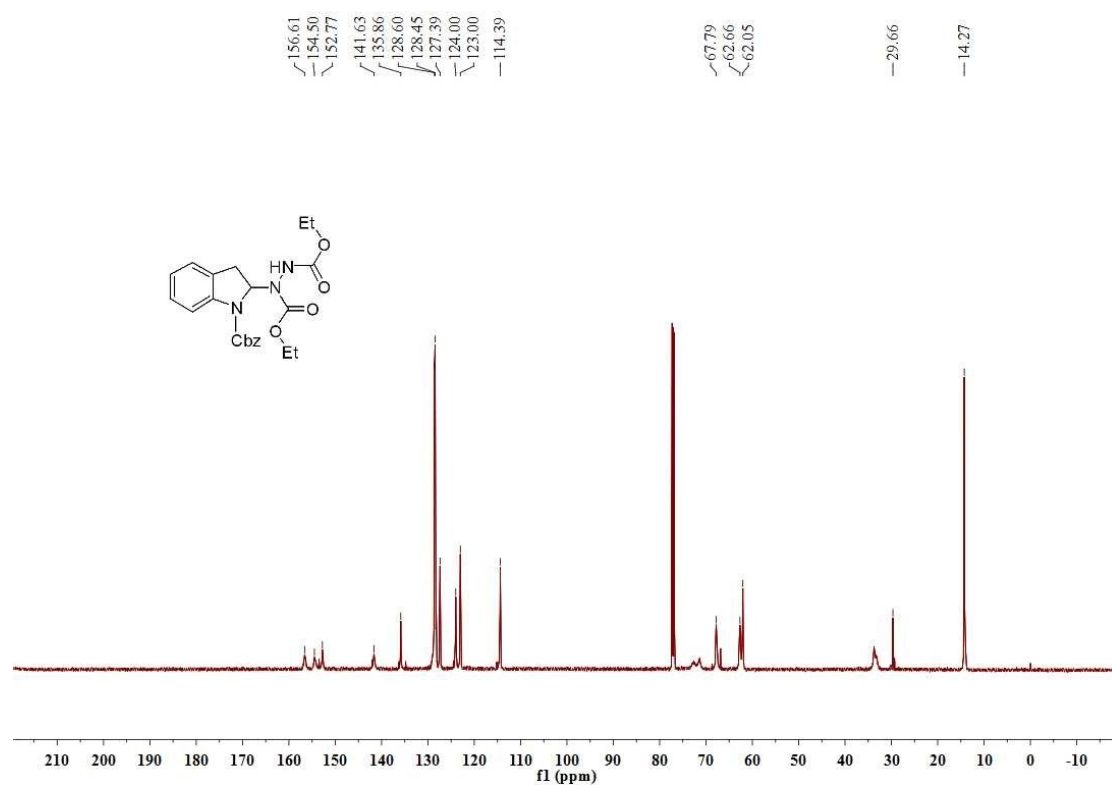
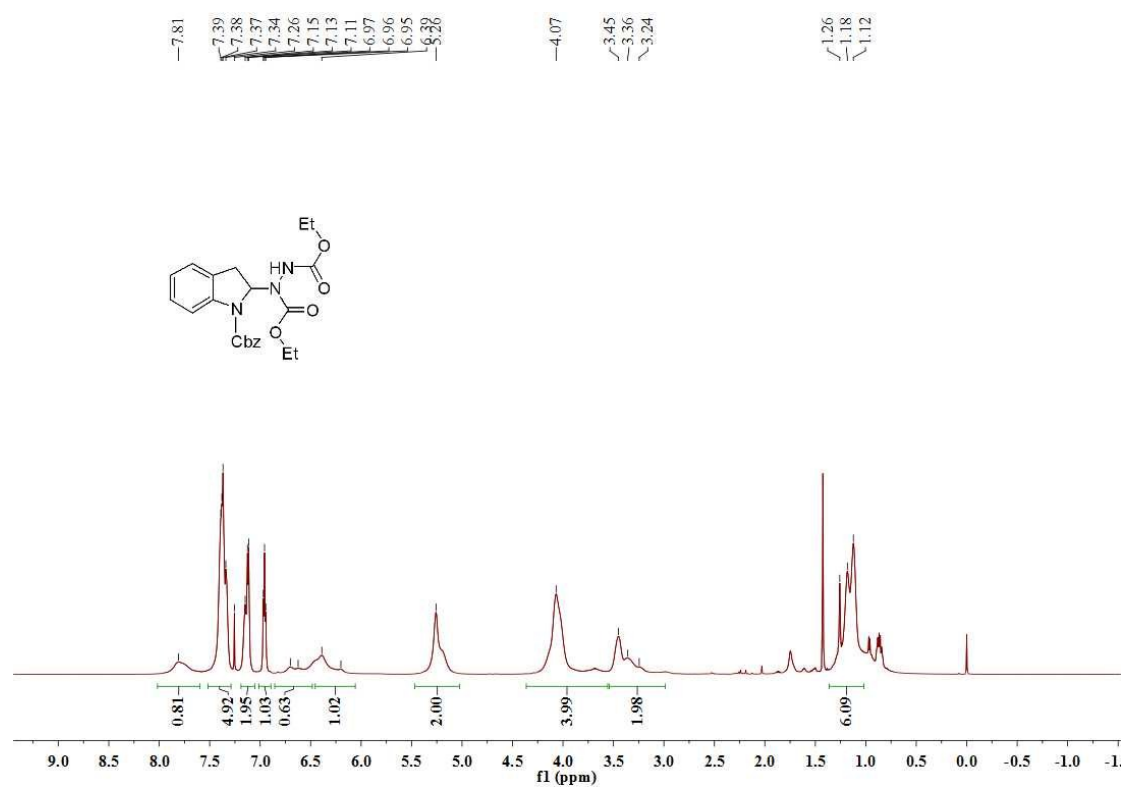
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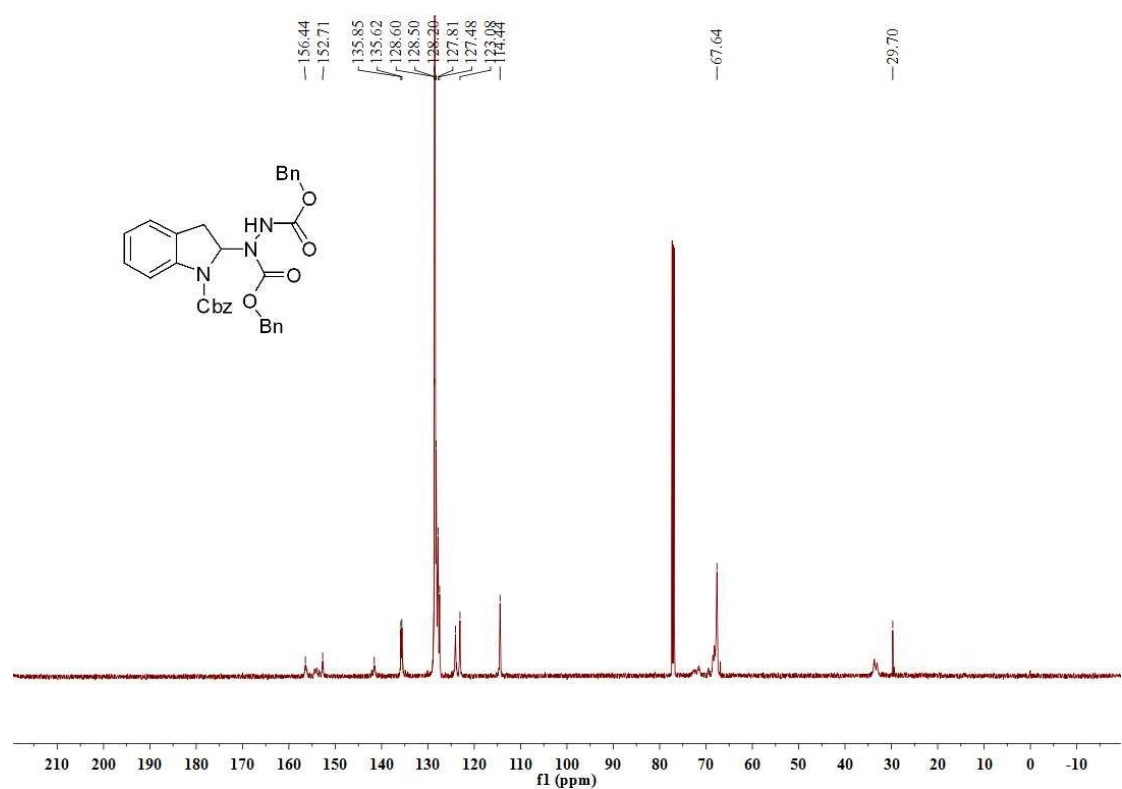
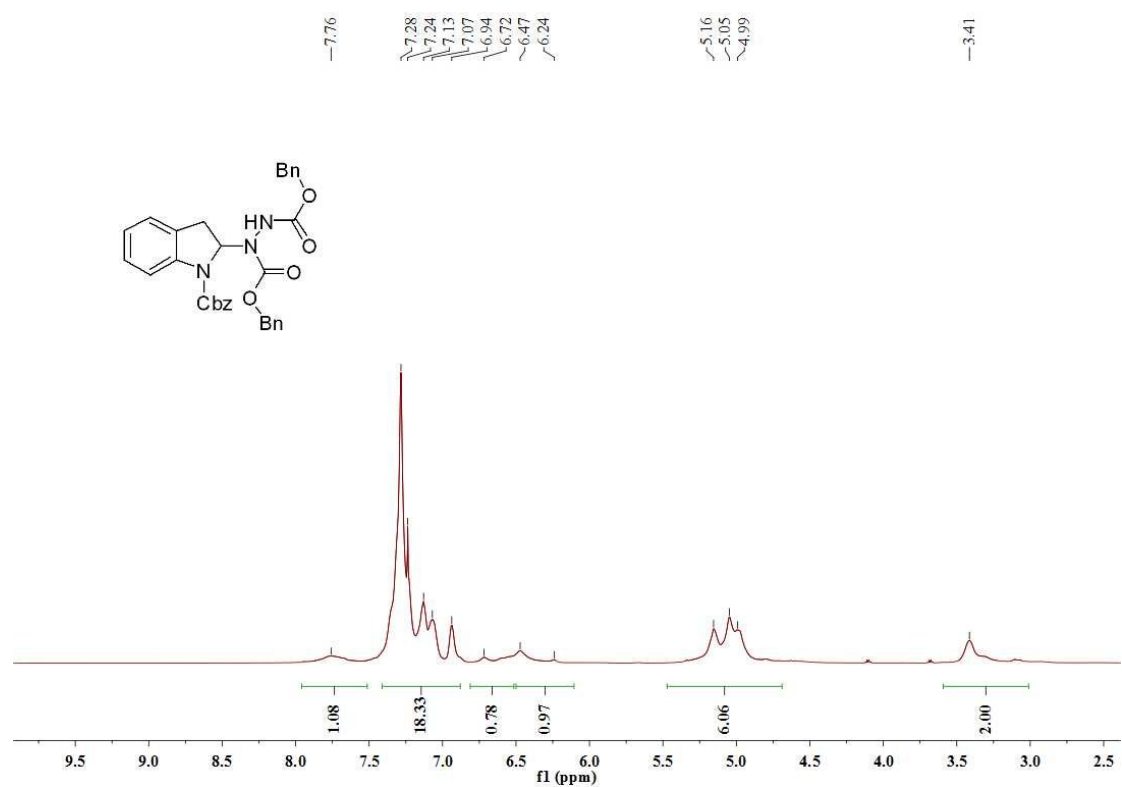
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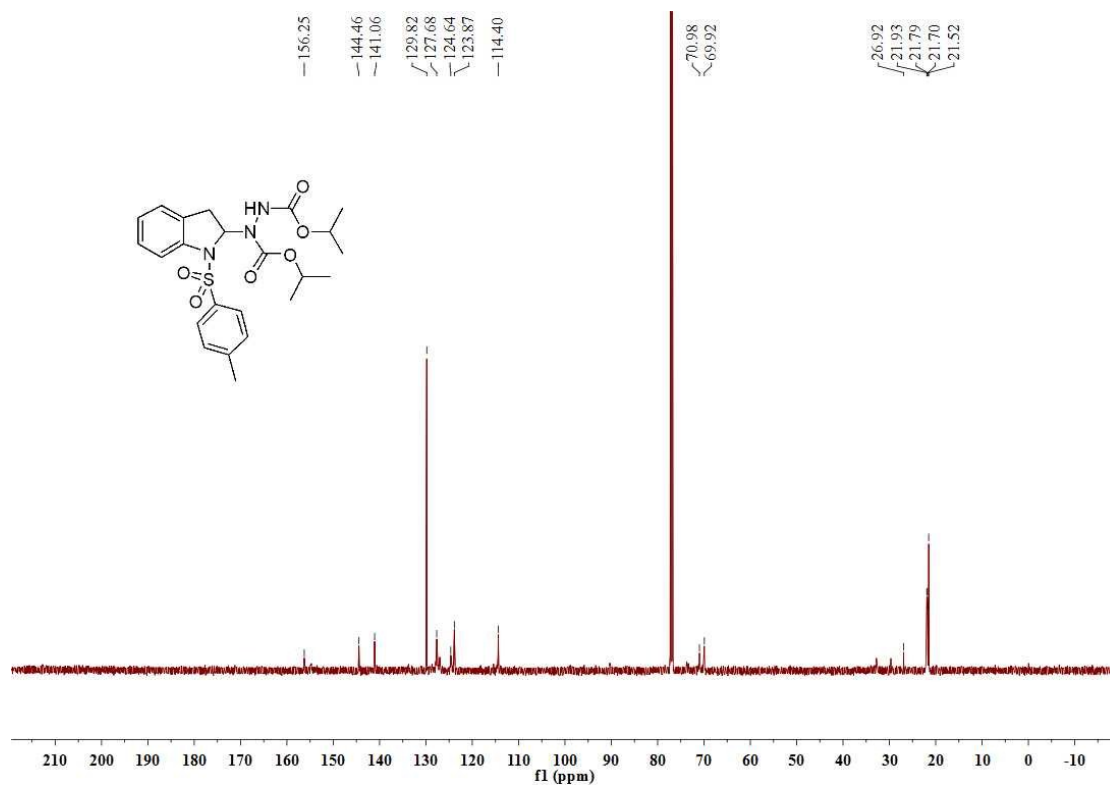
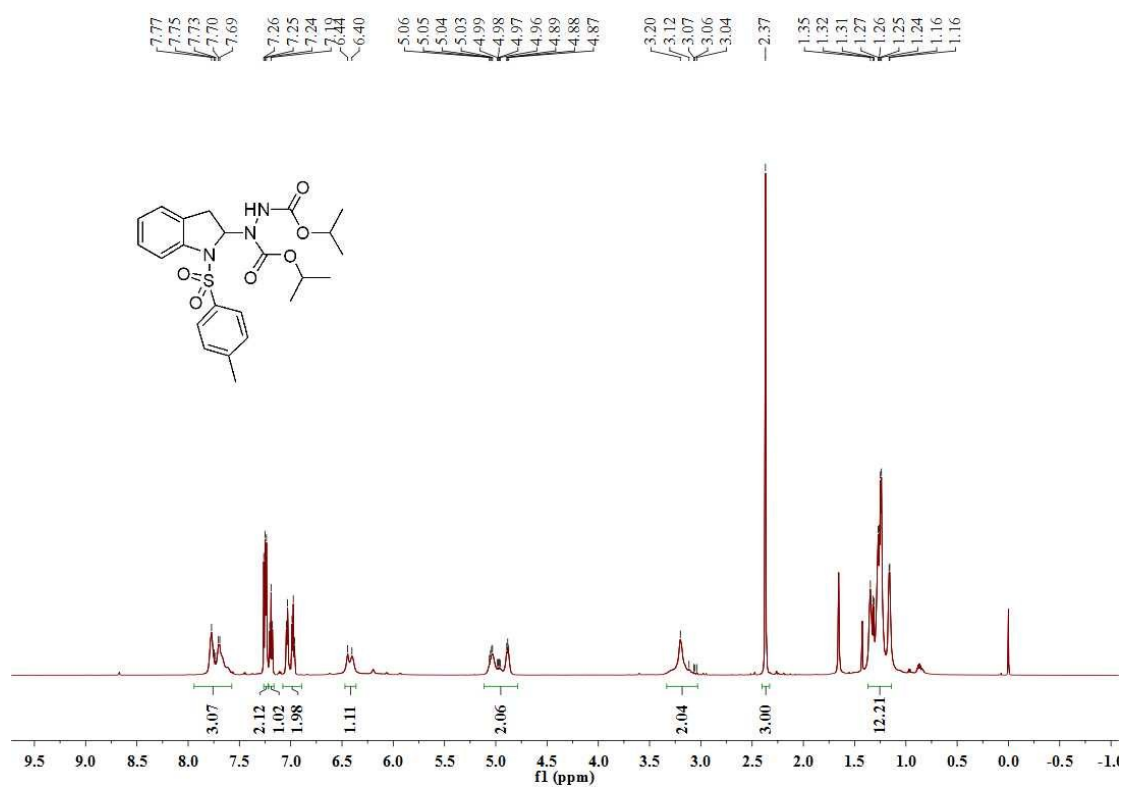
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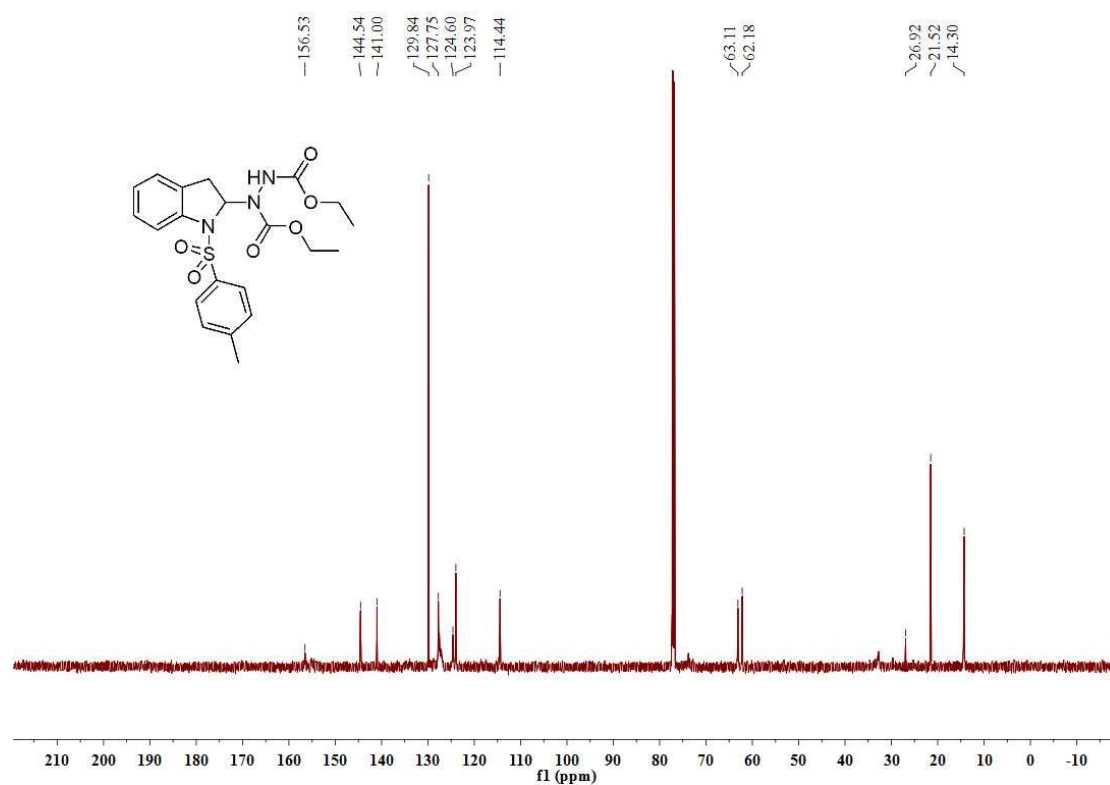
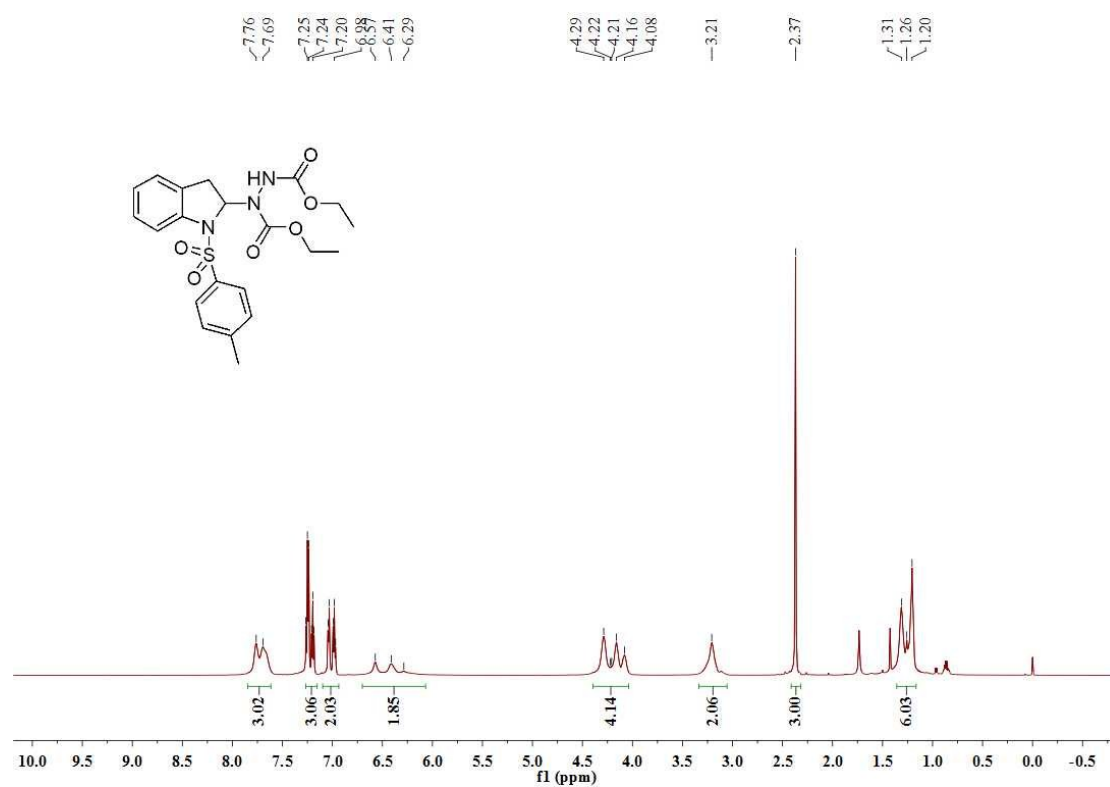
3h



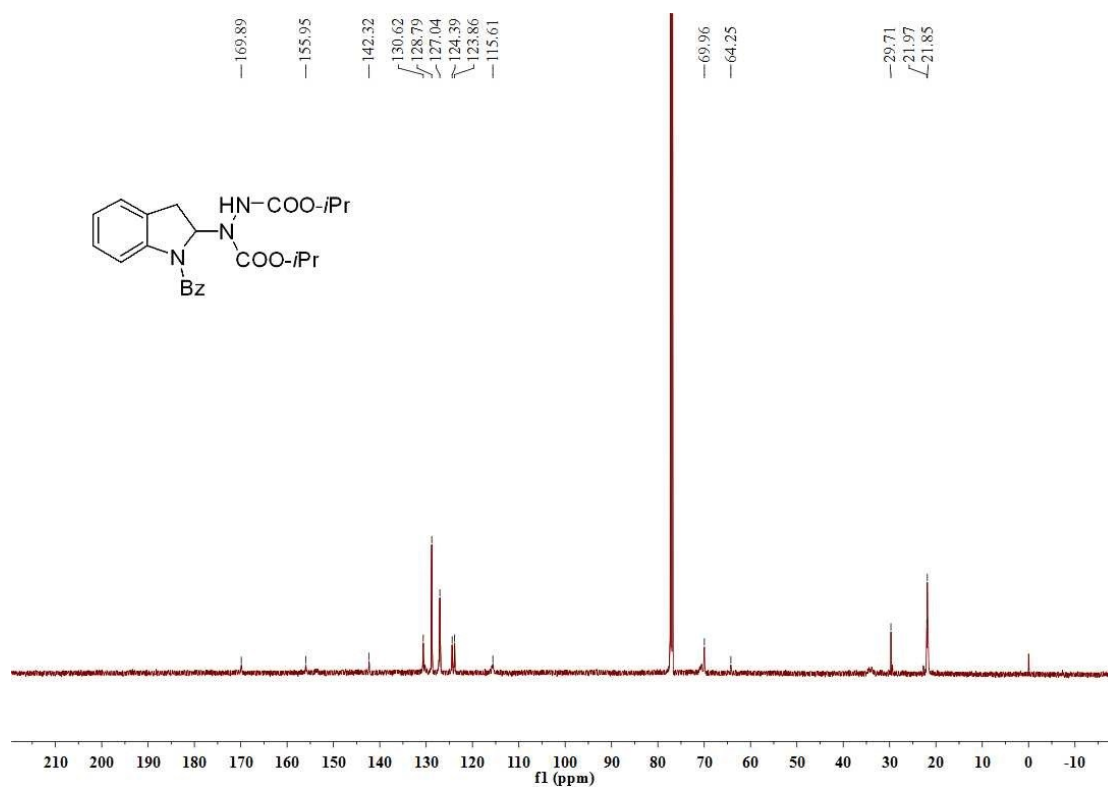
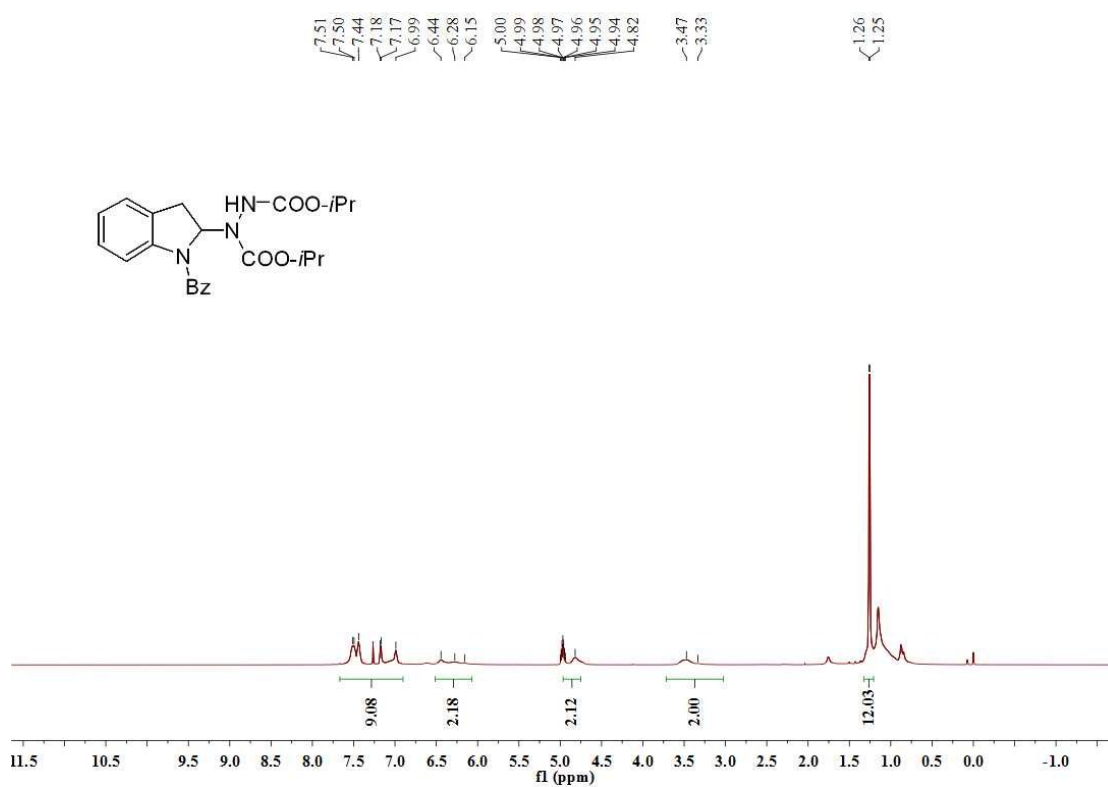
3i



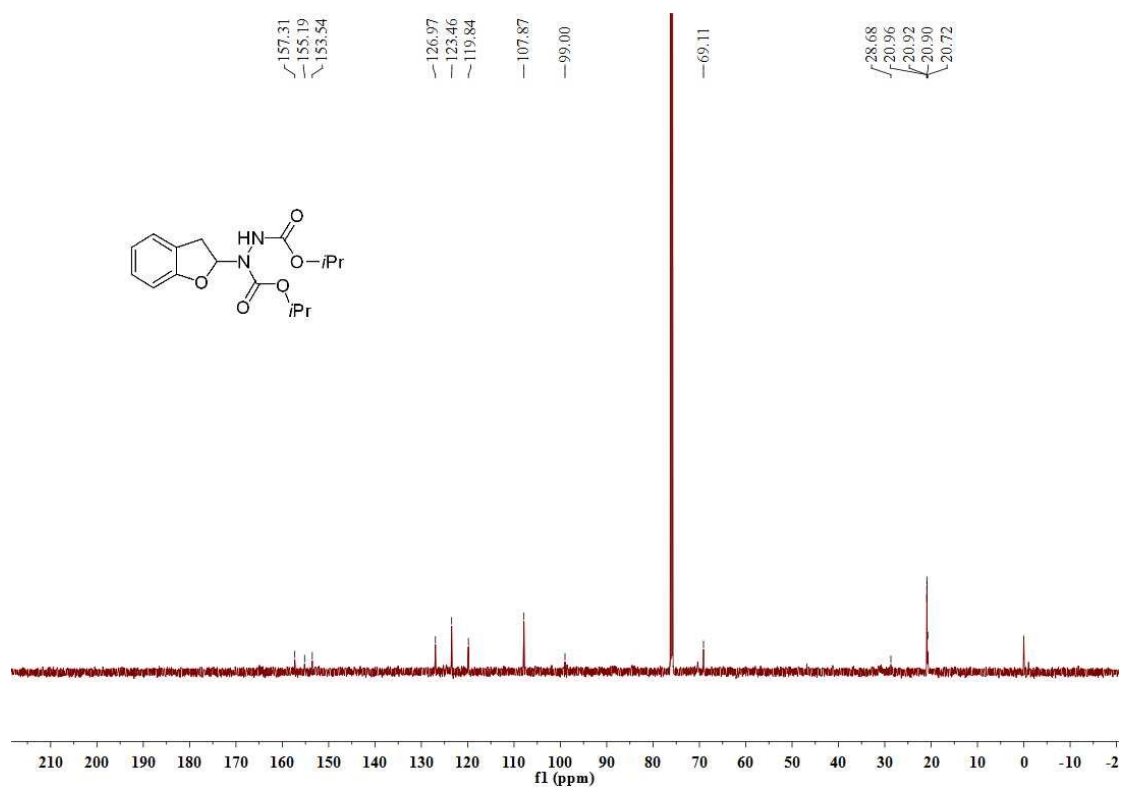
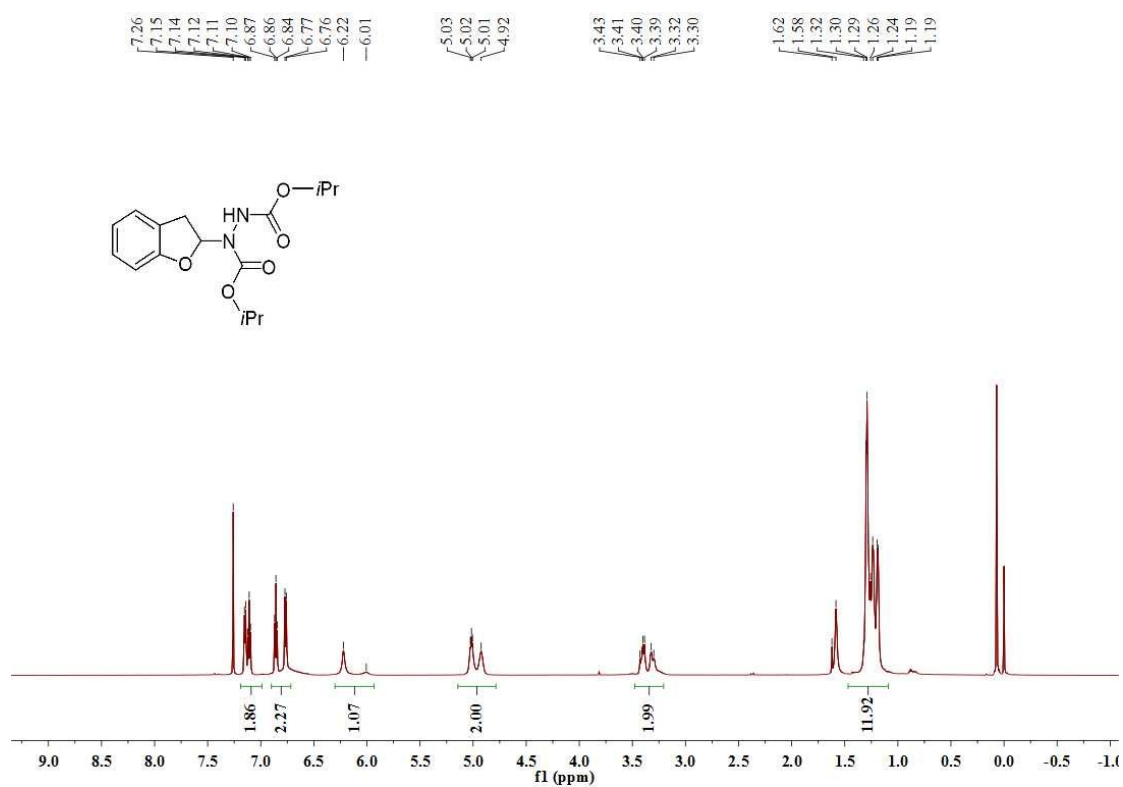
3j



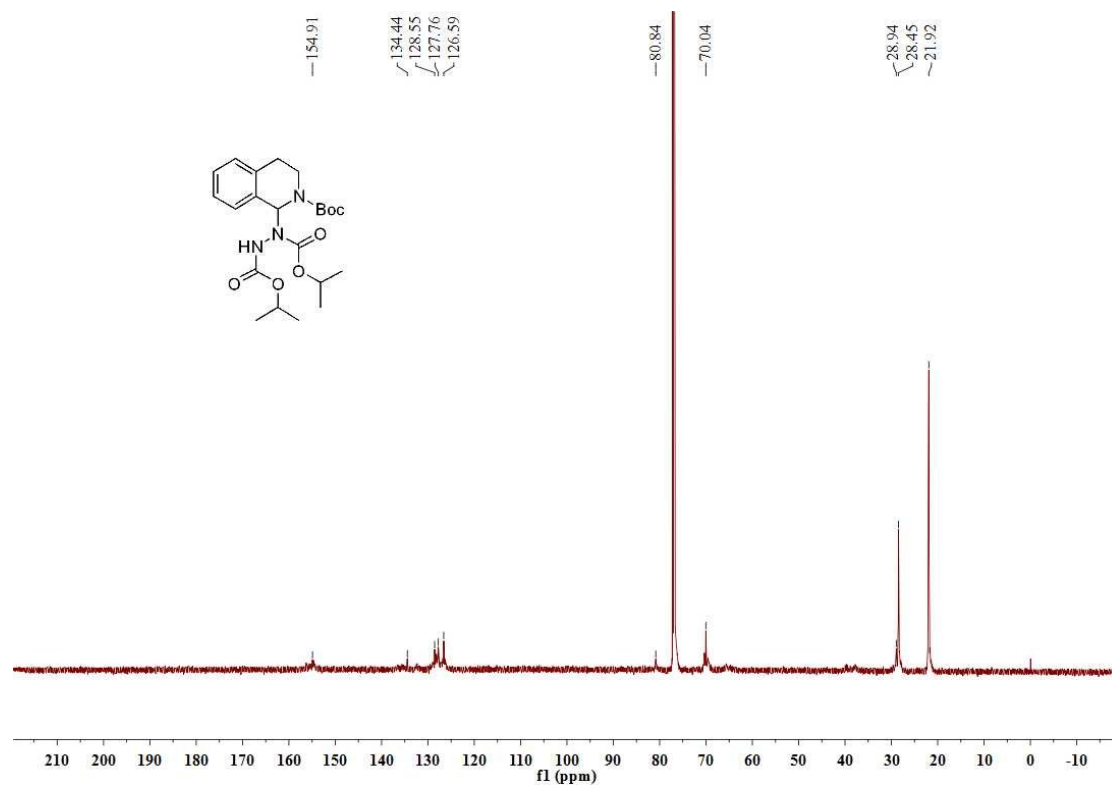
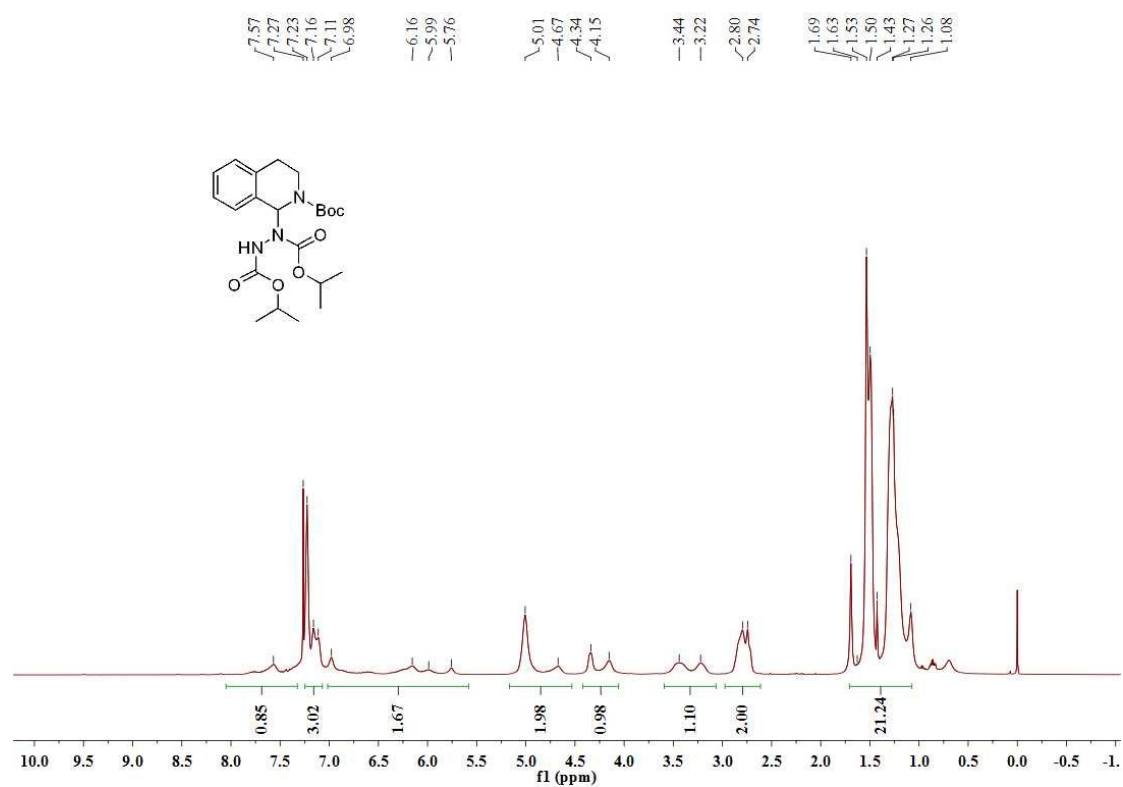
3k



31



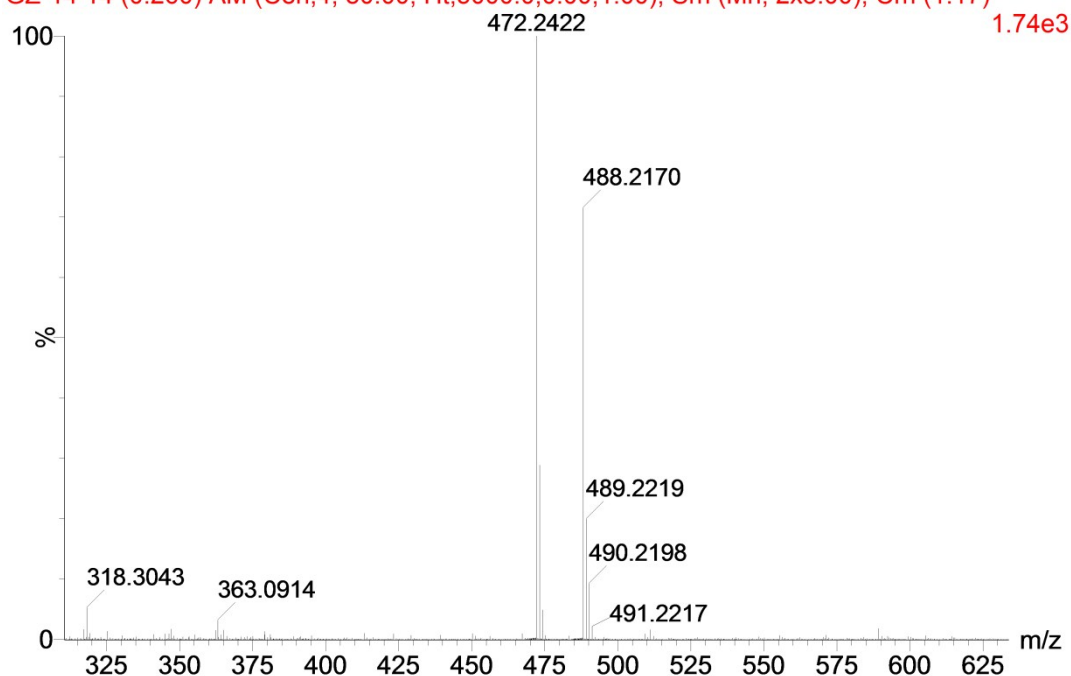
3m



6. HRMS spectra

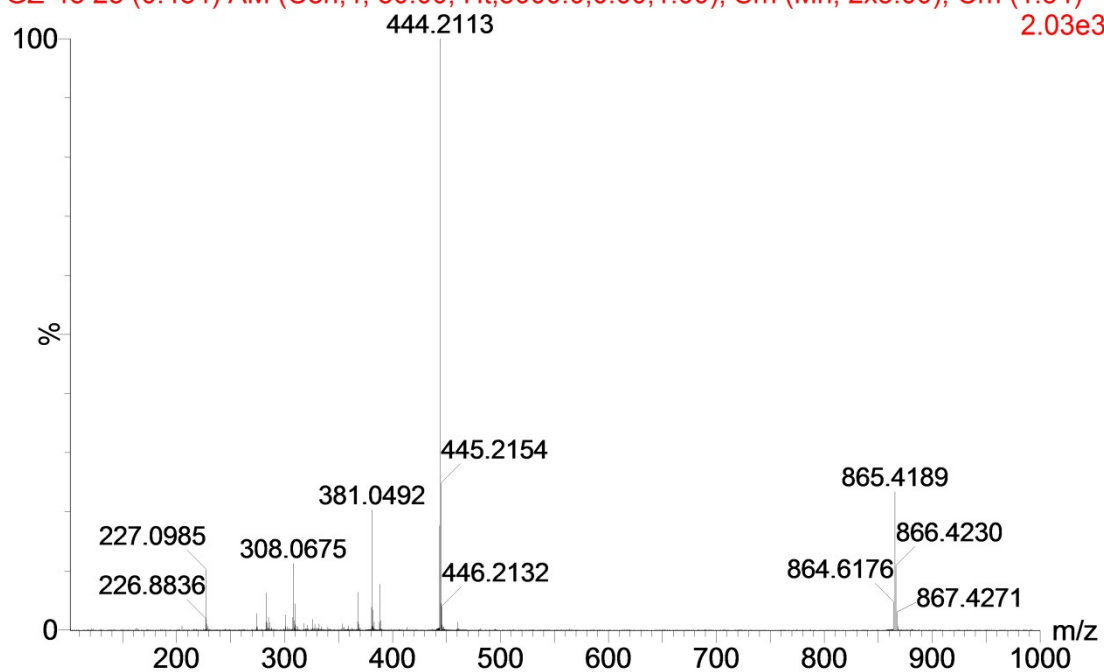
3a

GZ-14 14 (0.260) AM (Cen,4, 80.00, Ht,5000.0,0.00,1.00); Sm (Mn, 2x3.00); Cm (1:17)



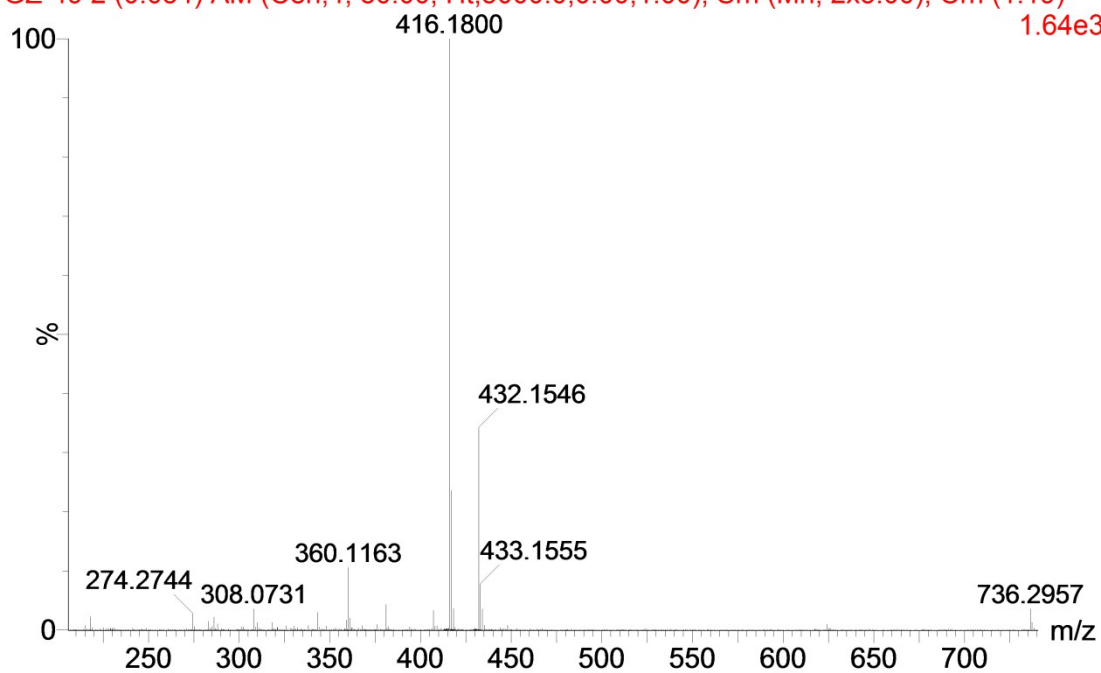
3b

GZ-48 28 (0.484) AM (Cen,4, 80.00, Ht,5000.0,0.00,1.00); Sm (Mn, 2x3.00); Cm (1:34)



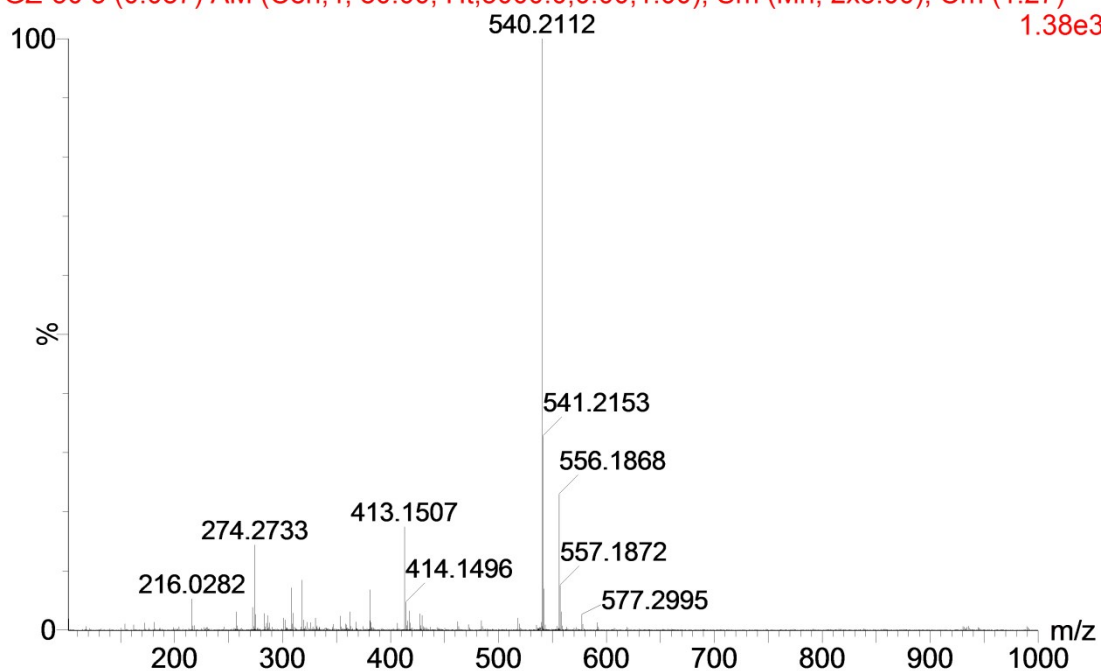
3c

GZ-49 2 (0.034) AM (Cen,4, 80.00, Ht,5000.0,0.00,1.00); Sm (Mn, 2x3.00); Cm (1:19)
1.64e3



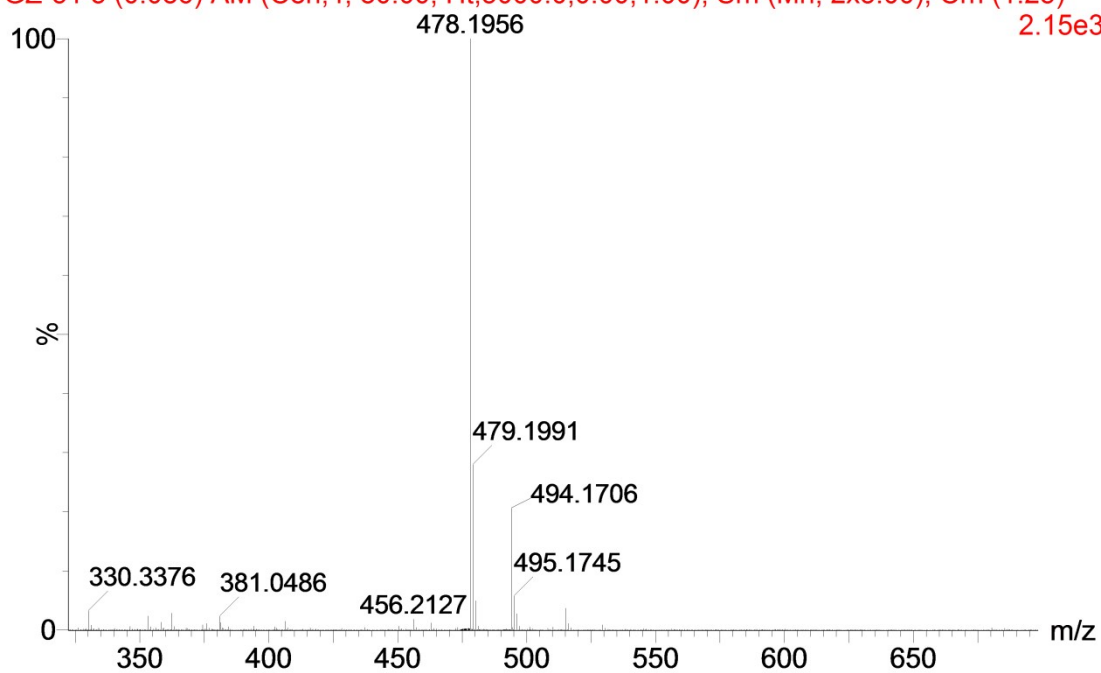
3d

GZ-50 5 (0.087) AM (Cen,4, 80.00, Ht,5000.0,0.00,1.00); Sm (Mn, 2x3.00); Cm (1:27)
1.38e3



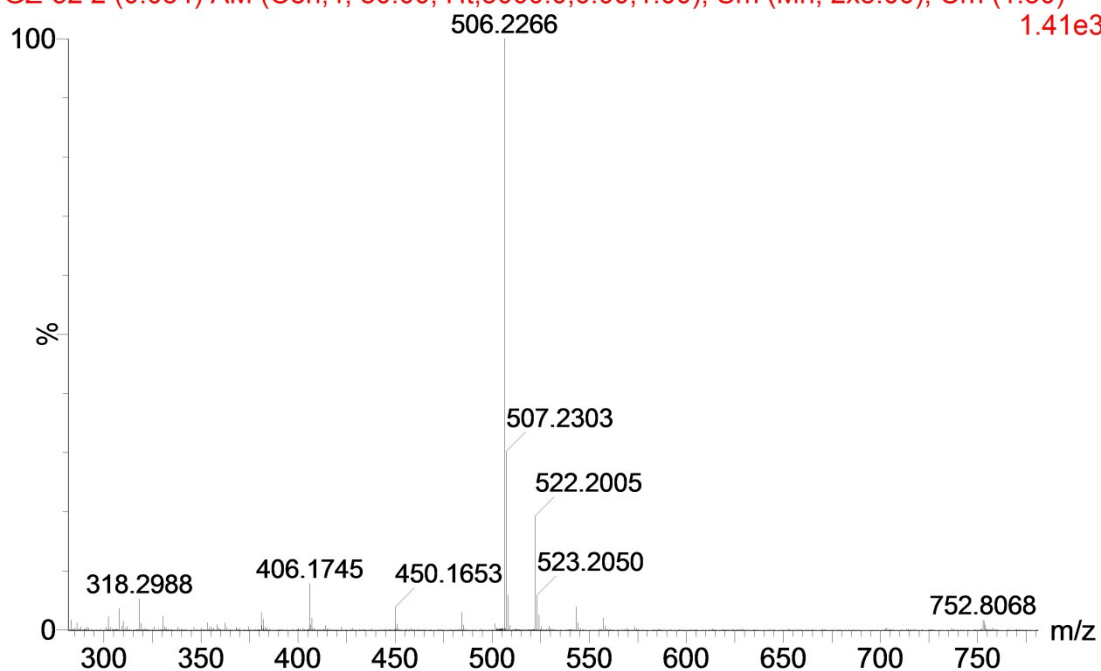
3e

GZ-51 5 (0.086) AM (Cen,4, 80.00, Ht,5000.0,0.00,1.00); Sm (Mn, 2x3.00); Cm (1:25)
2.15e3



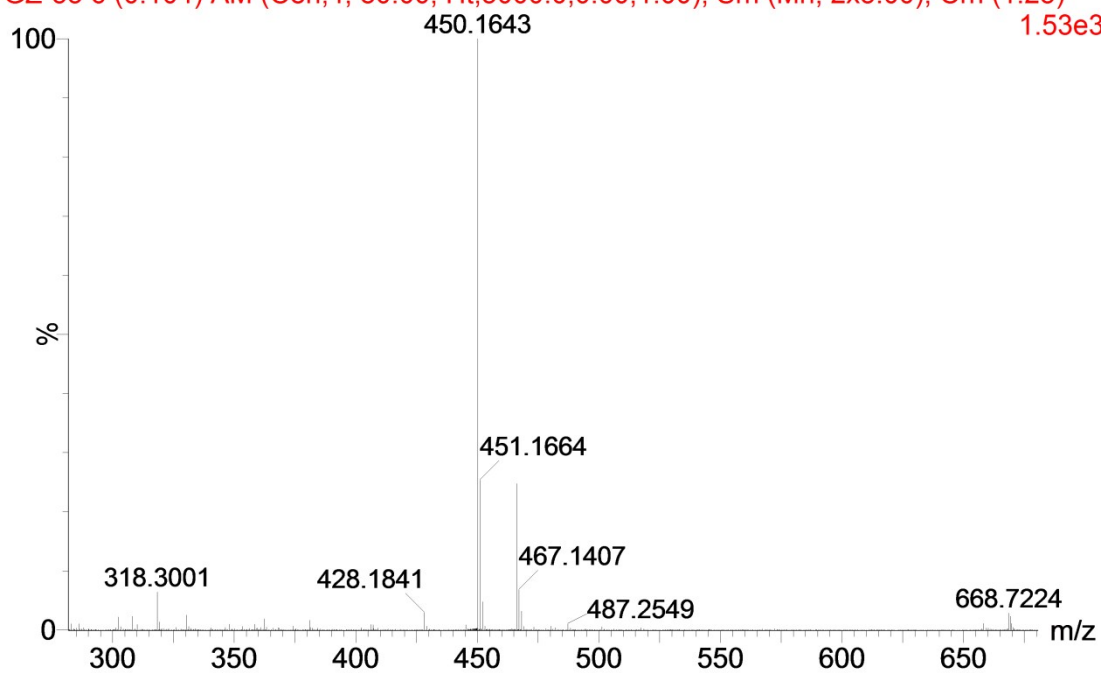
3f

GZ-52 2 (0.034) AM (Cen,4, 80.00, Ht,5000.0,0.00,1.00); Sm (Mn, 2x3.00); Cm (1:30)
1.41e3



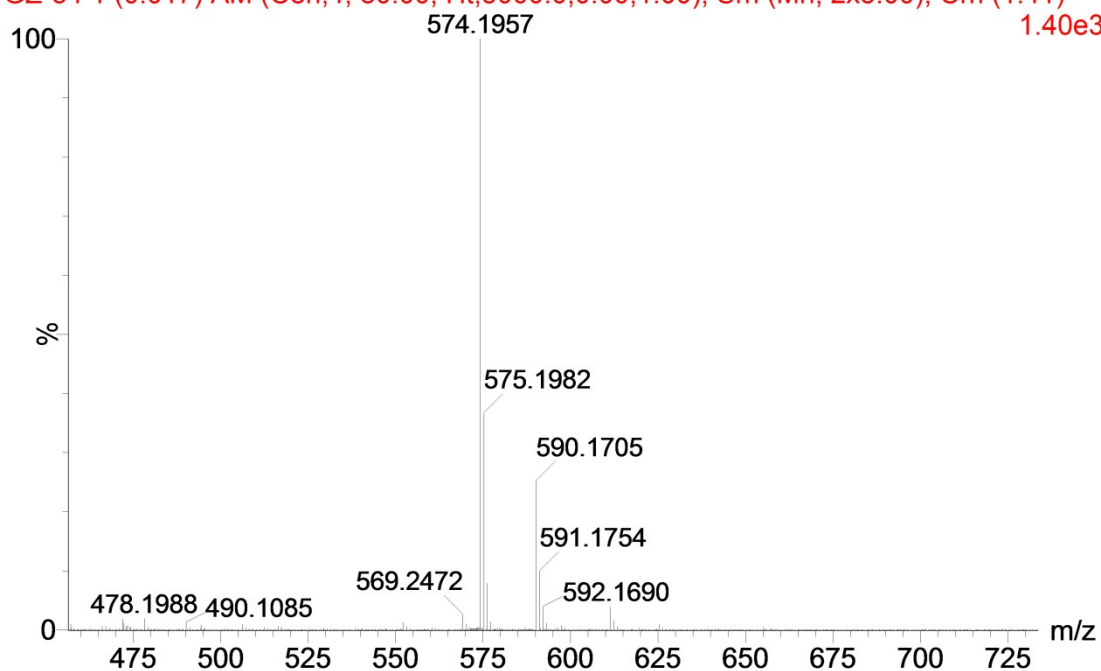
3g

GZ-53 6 (0.104) AM (Cen,4, 80.00, Ht,5000.0,0.00,1.00); Sm (Mn, 2x3.00); Cm (1:23)
1.53e3



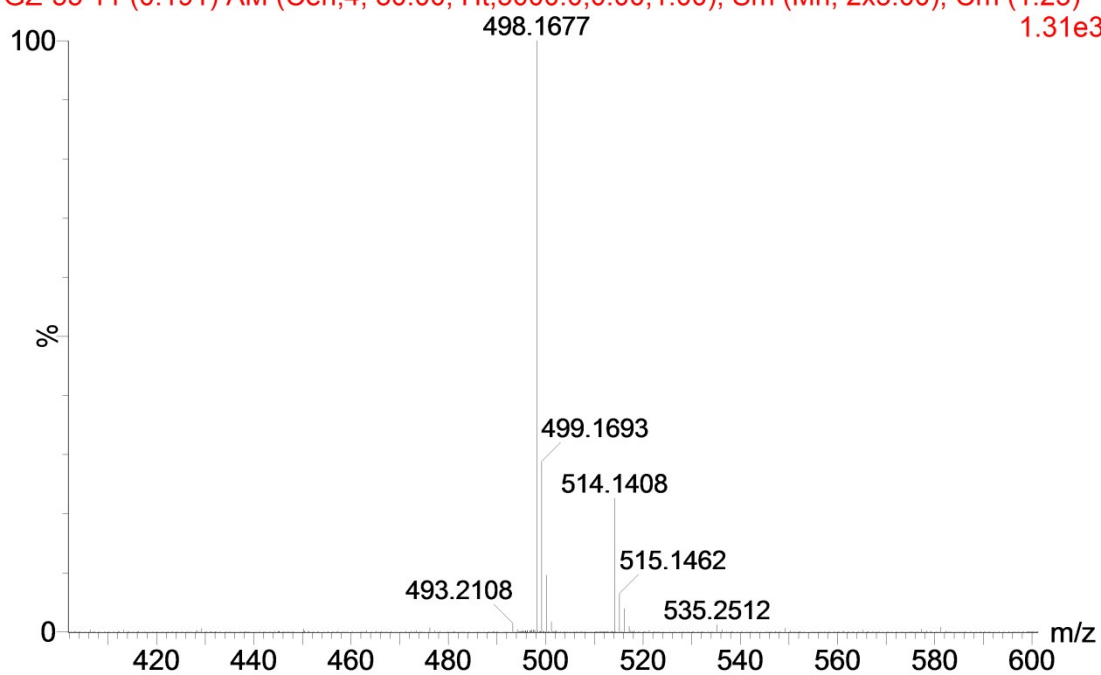
3h

GZ-54 1 (0.017) AM (Cen,4, 80.00, Ht,5000.0,0.00,1.00); Sm (Mn, 2x3.00); Cm (1:41)
1.40e3



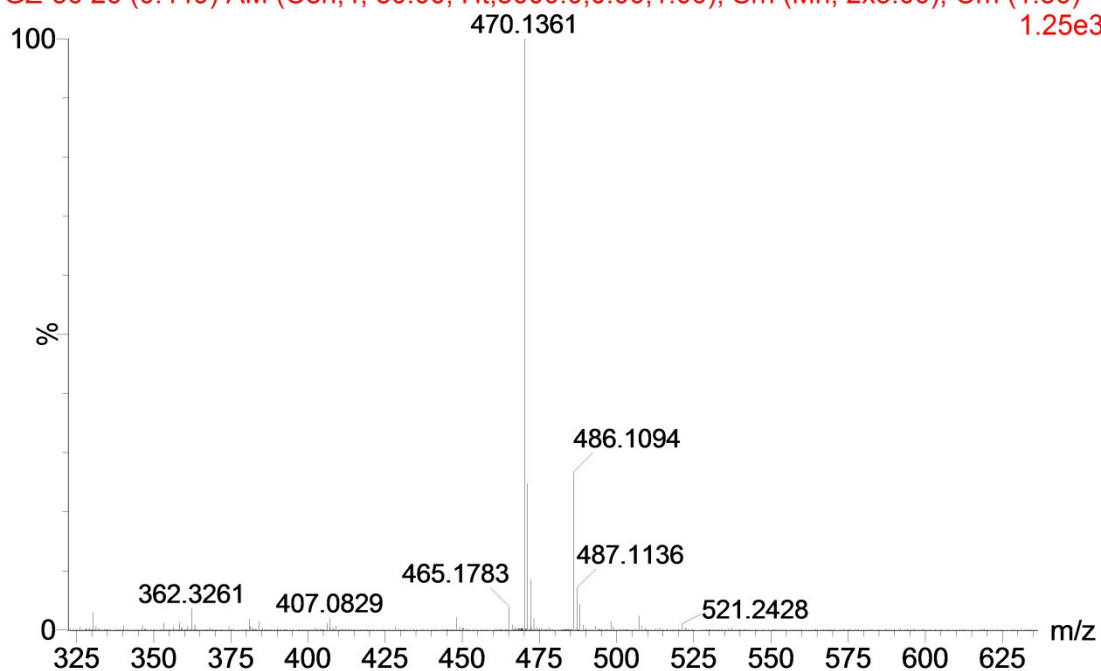
3i

GZ-55 11 (0.191) AM (Cen,4, 80.00, Ht,5000.0,0.00,1.00); Sm (Mn, 2x3.00); Cm (1:23)
1.31e3



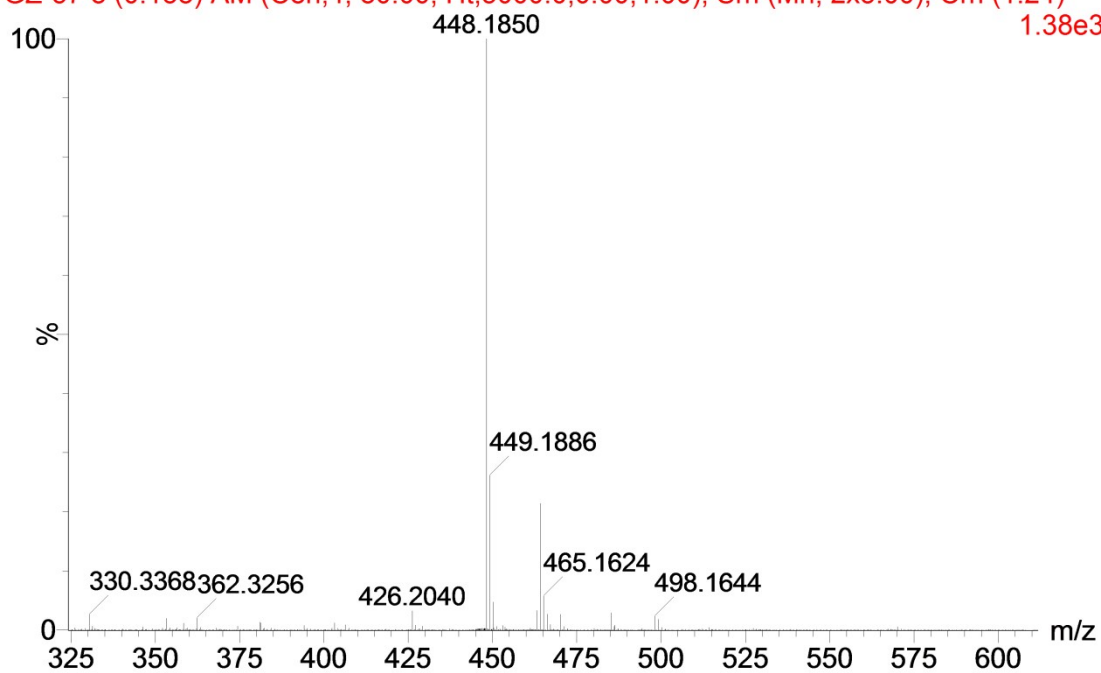
3j

GZ-56 26 (0.449) AM (Cen,4, 80.00, Ht,5000.0,0.00,1.00); Sm (Mn, 2x3.00); Cm (1:36)
1.25e3



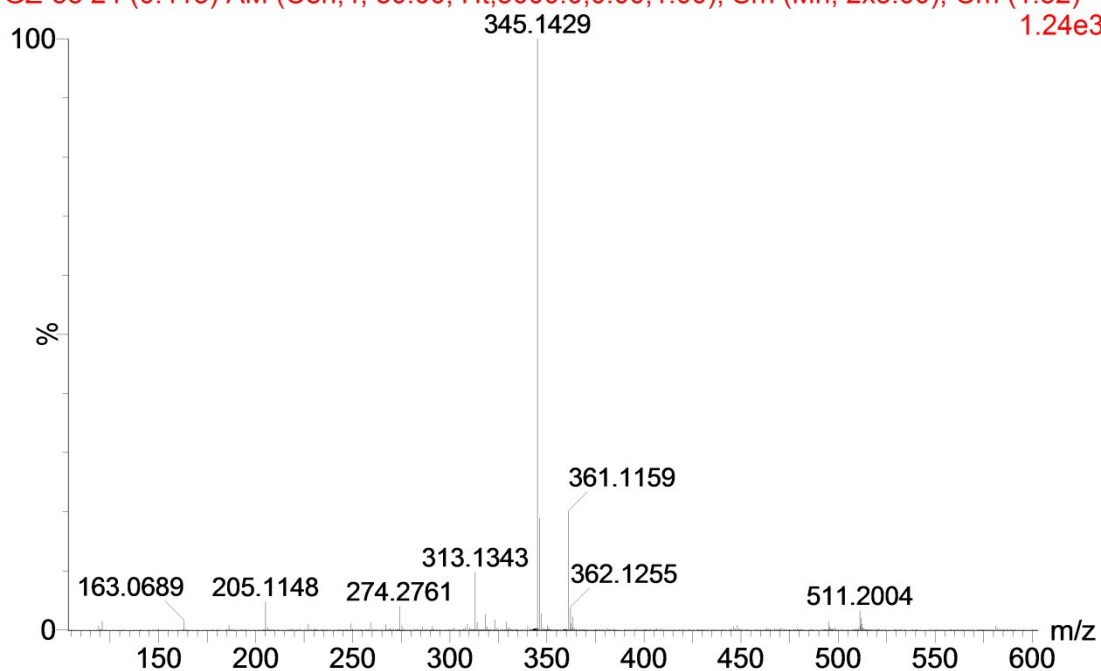
3k

GZ-57 8 (0.138) AM (Cen,4, 80.00, Ht,5000.0,0.00,1.00); Sm (Mn, 2x3.00); Cm (1:24)
1.38e3



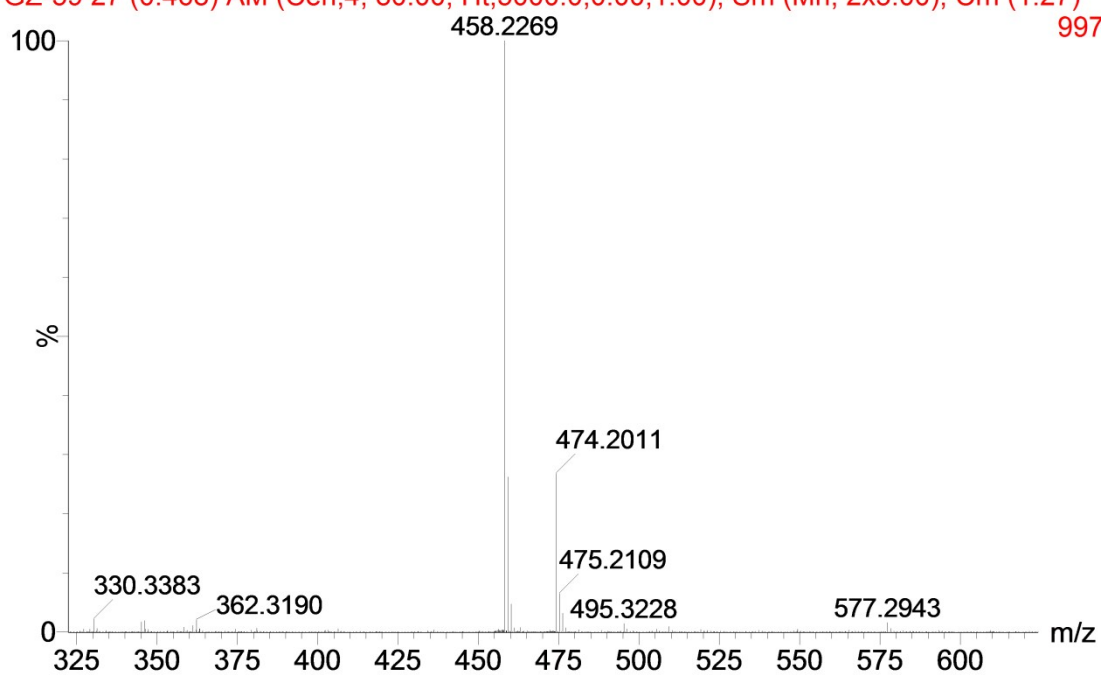
3l

GZ-58 24 (0.413) AM (Cen,4, 80.00, Ht,5000.0,0.00,1.00); Sm (Mn, 2x3.00); Cm (1:32)
1.24e3



3m

GZ-59 27 (0.468) AM (Cen,4, 80.00, Ht,5000.0,0.00,1.00); Sm (Mn, 2x3.00); Cm (1:27)



7. References

1. a) M. Kurokawa and T. Sugai, *Bull. Chem. Soc. Jpn.*, 2004, **77**, 1021–1025. b) J. Charton, A. C. Gassiot, S. G. Mizzi, M. A. D. Fontaine, P. Melnyk and C. Sergheraert, *Bioorg. Med. Chem. Lett.*, 2005, **15**, 4833–4837. c) A. Hartikka, A. T. S' lo'sarczyka and P. I. Arvidsson, *Tetrahedron: Asymmetry*, 2007, **18**, 1403–1409. d) S.-J. Zhang, W.-W. Sun, P. Cao, X.-P. Dong, J.-K. Liu, B. Wu, *J. Org. Chem.*, 2016, **81**, 956–968. e) Y. Sasano, S. Nagasawa, M. Yamazaki, M. Shibuya, J. Park and Y. Iwabuchi, *Angew. Chem. Int. Ed.*, 2014, **53**, 3236–3240.