Supporting Information

Tellurium-Containing Nanoparticles for Controlled Delivery of

Cisplatin Based on Coordination Interaction

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1 EXPERIMENTAL SECTION.

1.1 Materials.

N,N'-Carbonyldiimidazole (CDI), 8-bromooctanoic acid was the products of TCI. Tellurium powder was obtained from Aladdin chemical company. Sodium borohydride was purchased from Alfa-aesar. Cisplatin and triethylene glycol monomethyl ether were from Sigma-Aldrich. All the other chemicals and solvents were used as received unless otherwise stated.

1.2 Methods.

The ¹H NMR spectra and ¹³C NMR spectra were measured on a Bruker AVANCE III HD 400 (400 MHz). While ¹²⁵Te NMR spectra were recorded on a JEOL JNM-ECA 600 (600 MHz) spectrometer.

The TEM images were obtained by a JEM-2010 Microscope with an accelerating voltage of 80 kV. Samples were prepared by drop-coating the aqueous solution on the carbon-coated copper grid for 10 min and then observed without staining. DLS measurements were performed by a Malvern ZEN3690 Zetasizer at 25 $^{\circ}$ C. ESI-mass was carried out on a LTQ LC/MS apparatus.

Synthesis of TeCOOEG. The telluride containing compound was synthesized according to the synthetic procedure in Figure S 1. Briefly, 8-bromooctanoic acid was dissolved in anhydrous DCM and stirred with 1.2 equiv CDI for 30 min. Then triethylene glycol monomethyl ether of 1.3 equiv was added via a syringe and stirred for 3 h under room temperature. Then the reaction was concentrated under reduced pressure and then washed in sequence with 3 M HCl, 1 M NaHCO₃, and 0.5 M NaCl aqueous solutions. Colorless viscous liquid was obtained as Compound 1 after rotary evaporation. Yield: 60%.

Disodium telluride was prepared by the reaction of Te powder and excess amount of sodium borohydride in water under the protection of nitrogen at 50 °C. Compound 1 in THF was added under N_2 flow at 50 °C for 6 h. Products were purified by filtration and extraction. Light yellow liquid was generated at a yield

of 55%.

¹H NMR (400 MHz, CDCl₃, 298 K) δ (ppm): 4.25(4H, t, *CH*₂OCO), 3.72-3.58(20H, m, CH₃O*CH*₂*CH*₂O*CH*₂*CH*₂O*CH*₂CH₂OCO), 3.41(4H, s, O*CH*₃), 2.64(4H, t, Te*CH*₂), 2.35(4H, t, *CH*₂COOH), 1.75-1.25(20H, m, OOCCH₂(*CH*₂)₅CH₂Te). ¹³C NMR (400MHz, CDCl₃, 298 K), δ (ppm): 173.8 (1C, s, *C*OOCH₂), 71.9-63.4(6C, m, CH₃O*CH*₂*CH*₂O*CH*₂*CH*₂O*CH*₂*CH*₂OCO), 59.1(1C, s, O*CH*₃), 34.2-24.8 (6C, m, OOC(*CH*₂)₆CH₂Te), 2.7 (1C, s, Te*CH*₂).



Figure S 1 . The synthetic routes of the TeCOOEG.

In vitro cell cytotoxicity experiments.

HepG2 cells were cultured in Dulbecco's modified Eagle's medium (DMEM) (SH30022.01B; Thermo Inc., Bremen, Germany), supplemented with 10 % fetal bovine serum at 37 °C in a humidified atmosphere containing 5% CO₂. Cells were seeded into 96-well plates at a density of 5000 cells per well. 12 h after seeding, the cells were treated with various drug formulations for up to a further 24 h. Cell viability was evaluated using a CCK-8 assay according to the manufacturer's instructions (Dojindo, Japan).

Imaging of Tumor Cells *in Vitro*. HepG2 cells were seeded onto a 35 mm borosilicate chambered cover glasses (Nunc, USA.) at a density of 2×10^5 cells/well and grown at 37 °C for 12 h. After treatment with cisplatin or cisplatin-loaded nanoparticles for 3 h, the HepG2 cells were washed three times with PBS and incubated with DCFH-DA (10 mm; Sigma, USA) at 37 °C for 20 min. The fluorescence intensities were measured by using a LSM 710 confocal microscope (Carl Zeiss, USA) at 40× magnification.

In vivo antitumor activity assay.

Tumor xenografts were formed in female BALB/C nude mice by injecting 5×10^6 HepG2 cells into the right flank (4 mice/group). When the tumor volume reached 60-70 mm³, various drug formulations (Saline, cisplatin, cisplatin-loaded nanoparticles) were injected intravenously every three or four days. The

antitumor activity was evaluated by tumor size (V), as estimated by the following equation:

$$V = a \times b^2 / 2$$

where a and b are the major and minor axes of the tumor measured by a caliper, respectively. Mice with tumor implants were euthanized after 26 d, and the tumor xenografts were excised and weighed.

Plasma clearance experiments.

Samples were administered by tail vein injection to healthy BALB/c mice with normal immunity. Blood was collected post-injection via retro-orbital bleed (at t = 15 min, 24 h and 48 h); n = 3 was assigned for each time-point. Heparin coated capillaries and collection tubes were used; and collected blood was stored on ice. The Pt concentrations in the collected blood were then measured by ICP-MS.

2 DETAILED CHARACTERIZATION.



Figure S 2 ¹H NMR spectrum of compound 1. Solvent: CDCl₃.



Figure S 3 ¹H NMR spectrum of TeCOOEG. Solvent: CDCl₃.



Figure S 4 ¹H NMR spectrum of TeCOOEG. Solvent: CDCl₃.



Figure S 5 1 H NMR spectrum of TeCOOEG after coordination. Solvent: CDCl₃. After coordination, the chemical shift of α proton of tellurium atom in TeCOOEG shifted from 2.64 to 2.83 ppm.



Figure S 6 ESI MS of TeCOOEG/cisplatin coordination complex.



Figure S 7 DLS result of TeCOOEG nanoparticles. The hydrodynamic diameter was about 120 nm.



Figure S 8 DLS measurement of cisplatin-loaded nanoparticles showed the hydrodynamic diameter was about 140 nm.



Figure S 9 Fluorescent micrograph study based on dichlorofluorescein diacetate on HepG2 cells.



Figure S 10 Serum biochemical parameters of healthy BALB/c mice. UA, Uric Acid. CR, creatinine after i.v. injection of saline, cisplatin and the cisplatin-loaded nanoparticles.



Figure S 11 The side effects of cisplatin and the cisplatin-loaded nanoparticles on PTL.