Hierarchical ZnO nanorods/Ni(OH)₂ nanoflakes for

room-temperature, cheap fabrication of non-

enzymatic glucose sensor

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SUPPORTING INFORMATION



Figure S1. EDX spectrum of the sample 50-60.



Figure S2. SEM plan view of the sample 50-40 (ZnO NRs grown with 50mM HMTA; 40 electrodeposition cycles for $Ni(OH)_2$). The insets are higher magnification SEM image.



Figure S3. Cyclic voltammograms of bare ITO and ZnO NRs/ITO samples in 0.1M NaOH (scan rate 50 mV s⁻¹).



Figure S4. (a) Cyclic voltammograms of the sample 50-40 in 0.1M NaOH (scan rate 50 mV s⁻¹). (b) Calibration curve of sample 50-40 for several addition of glucose. The sensitivity extracted is 1.42 mA/mM cm^2 .



Figure S5. CVs of 25-20 in 0.1M NaOH solution (scan rate 50mV s⁻¹) acquired with successive addition of 0.2 mM glucose, 0.01 mM AC, 0.01 mM AA, 0.02 mM UA, and, finally, 0.2 mM glucose.

Material; method	Sensitivity	Linear	Selectivity	Chloride	Long-	Ref.
	[mA/mM	range	test	test	term	
	cm²]	[mM]			stability	
Ni/Cu;	1.59	0.01-3.2	AA; UA	n/a	49 days	1
electrodeposition						
Ni/ITO;	0.61	0.03-3	UA;	n/a	50 days	2
electrodeposition			AA;dopamine			
Ni(OH)2/ZnO;	1.57	0.002-	UA;AA;	n/a	30 days	3
CBD/electrodeposition		3.8	aspartic acid;			
			dopamine			
Ni(OH)2/ZnO;	1.85	0.04-	UA; AA;AC	0.2M	40 days	This
electrodeposition/CBD		2.10		KCI		work

Table S1. Comparison of selected non-enzymatic glucose sensors based on Ni nanostructures.

References

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